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Modeling and optimization of sensory changes and shelf-life in vacuum packaged cooked ham treated by E-beam irradiation

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27 **ABSTRACT**

28 The E-beam irradiation of vacuum-packaged RTE cooked ham was carried out to
29 establish the dose required to achieve the food safety objective (FSO) and to minimize
30 changes in selected sensory attributes. Cooked ham was irradiated with doses ranging
31 from 1 to 4 kGy. After the treatment the microbial inactivation of *Listeria*
32 *monocytogenes*, the shelf-life of the product and some sensory attributes (appearance,
33 odor and flavor) were determined. The inactivation of *Listeria monocytogenes* was
34 satisfactorily described by a first-order kinetics equation ($R^2=0.99$). The influence of the
35 irradiation dose on appearance, odor and flavor was modeled through the Gompertz
36 ($R^2=0.99$, for appearance) and activation/inactivation ($R^2=0.99$, for odor and flavor)
37 equations. A model was also developed to determine the shelf-life of irradiated cooked
38 ham depending on the irradiation dose ($R^2>0.91$). The dose that maximized the scores
39 of the sensory attributes was 0.96 kGy resulting in an acceptable sensory quality for 80
40 days. It is possible to apply up to 2 kGy to ensure microbial safety while provoking no
41 significant changes in the above mentioned sensory attributes.

42 **Key words:** microbial safety, modeling, quality attributes, shelf-life, E-beam irradiation.

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56 **1. Introduction**

57 Nowadays, people's dietary habits are undergoing a transformation. Traditional food is
58 being replaced by ready-to-eat (RTE) products (IAEA, 2003; Jacxsens et al., 2002; Hoz
59 et al., 2006). Although consumers are demanding foods with short preparation times,
60 there is also great concern about the need for a healthy diet which drives consumers to
61 demand more and more natural foods, free of chemical additives (Jacxsens et
62 al., 2002). On the other hand, the meat industry is focused on manufacturing long shelf-
63 life RTE products in domestic portions from processed blocks (Cabeza et al., 2009; Gil-
64 Díaz et al., 2009). The rising number and severity of food poisoning outbreaks world-
65 wide has increased public awareness of the microbial safety of foods, including meats
66 (Maurice, 1994). Therefore, it is necessary to apply some preservation technique to
67 these products to both reduce the spoilage microorganisms and to guarantee the
68 microbial safety (Zhu et al., 2005).

69 Although thermal treatments have been the most commonly used technique for
70 reducing the microbial load of foods, they can destroy heat-sensitive nutrients and
71 affect properties such as flavor, odor, appearance or texture. Non-thermal methods
72 allow foods to be processed at lower temperatures than when they are pasteurized,
73 and, therefore, flavors, essential nutrients, and vitamins undergo minimal or no
74 changes. Foods can be non-thermally processed by irradiation (Patterson et al., 1993;
75 Zhu et al., 2005), high hydrostatic pressure (Lakshmanan and Dalgaard, 2004), the use
76 of antimicrobials (Vogel et al., 2006), ultrasound (Knorr et al., 2004), filtration, and
77 electrical methods such as pulsed electric fields (Barbosa-cánovas et al., 1998a), light
78 pulses (Wang et al., 2005), and oscillating magnetic fields (Barbosa-Cánovas et al.,
79 1998b). Due to recent technological developments over the last decade, E-beam
80 irradiation processing has been receiving special attention. Additionally, it is a very
81 useful method for sanitizing RTE foods, since it is not feasible to apply the conventional
82 technologies to these products with that goal in mind (Hoz et al., 2006; Sommers et al.,

83 2003; Thayer et al., 1990; Zhu et al., 2005).

84 Irradiation is an effective way to eliminate pathogens present in foods, including *Listeria*
85 *monocytogenes*, *Salmonella* spp., *Yersinia enterocolitica*, *Escherichia coli* O157:H7,
86 and others (Burgess et al., 2010; Cabeza et al., 2009; Schilling et al., 2009). However,
87 some reports (Arthur et al., 2005; Lee and Ahn, 2005; Rababah et al., 2010) indicate
88 that its application to meat is limited since irradiation can produce changes in the
89 aroma, color and flavor that significantly affect consumer acceptance. The odor of
90 irradiated meat has been described as being like rotten egg, sweet, cooked meat,
91 barbecued corn, burnt, sulphur, metallic, alcohol, acetic acid, liver-like serummy, and
92 bloody (Brewer, 2009; Hampson et al., 1996; Lee et al., 1996). It is, therefore, critical to
93 carefully adjust the irradiation doses to achieve an adequate level of microbial
94 inactivation to produce only negligible changes in sensory properties, thereby avoiding
95 consumer rejection of the irradiated product.

96 Since food quality and food safety normally require opposing process conditions, the
97 modeling and optimization of food preservation processes can lead to an equilibrium
98 between both factors allowing the optimal process conditions to be found. In this
99 regard, several studies have been carried out to optimize the processing and storage of
100 different meat products in order to prevent oxidative damage, the growth of pathogens
101 or the loss of antioxidant components (Álvarez et al., 2007; Marselles-Fontanet and
102 Martin-Belloso, 2007; Shi and Le Maguer, 2000). To formulate an optimization problem,
103 mathematical models that describe the effect of the process variables not only on the
104 inactivation of microorganisms, but also on the quality attributes and the shelf-life of the
105 products, must be obtained.

106 Examples of models to describe the inactivation of microorganisms or other
107 compounds (quality factors, enzymes, etc.) are the Gompertz function (Ding et al.,
108 2010), the Weibull distribution (Bermúdez-Aguirre et al., 2009), the Fermi Distribution

109 (Elez-Martinez et al., 2006) or the activation/inactivation model (Soysal, 2008).

110 On the other hand, models for describing the microbial growth include the modified
111 Gompertz equation (Huang, 2010) or the Hill's Model (Wang, 2010).

112 Due to the great potential of E-beam irradiation as a method of sanitizing RTE meat
113 products (Cabeza et al., 2007; Hoz et al., 2008), it is necessary to develop
114 mathematical models which describe the response of microorganisms and the quality
115 factors of meat products treated using this technology. Moreover, it is very interesting
116 to develop optimization procedures to find the optimum process conditions. The main
117 objective of this work was to optimize the irradiation treatment of vacuum packaged
118 cooked ham.

119 **2. Materials and methods**

120 **2.1. Estimation of the food safety objective (FSO)**

121 Assuming a contamination of 10 cells/g, the performance criterion values (Gorris, 2005)
122 of 1.7D and 5.09D reductions of the load of *L. monocytogenes*, for EC and USDA
123 statements respectively, have been previously determined (Cabeza et al., 2007) in
124 order to reach the FSO in cooked ham.

125 **2.2. Organism**

126 The *L. monocytogenes* Scott A (CIP 103575, serotype 4b) strain was used. The strain
127 was maintained by freezing (-40 °C) in trypticase soy broth (TSB; Difco, BD, Sparks,
128 MD) adding 10% glycerol as the cryogenic agent. Fresh culture was prepared for each
129 experiment by removing a piece of frozen culture from vials and inoculating it into 9 ml
130 of TSB, then incubating at 32 °C for 24 h. The culture was then centrifuged (at 4 °C)
131 and the pellet suspended in a beaker with 50 ml sterile saline solution, which yielded a
132 bacterial load of approximately 10⁸ cells/ml. The slices were contaminated by

133 immersion in the beaker for a few seconds. In experiments, a large number of cells
134 were used to calculate the radioresistant parameters precisely.

135

136 **2.3. Sample preparation and irradiation treatment**

137 Heat processed cooked ham blocks (1 kg weight) packed into cylindrical (diameter 7
138 cm, length 25 cm) thermoplastic bags were purchased in a local supermarket. Slices (2
139 mm thick) were cut in an electric machine, whose knife and contact surfaces were
140 previously thoroughly cleaned and then washed in sterile distilled water. For
141 microbiological purposes, slices were contaminated as described above. The
142 contaminated (for microbial analysis) and uncontaminated (for shelf-life determination)
143 slices (1 for microbial and 3–6 per bag for shelf-life analysis) were vacuum packaged to
144 reach about 20 kPa in 10×10 cm laminated film bags of low gas permeability (diffusion
145 coefficient of 35 cm³/24 h m² bar to oxygen and 150 cm³/24 h m² bar to carbon
146 dioxide). The samples were transferred (less than 1 h) in insulated polystyrene boxes
147 to the irradiation plant (IONISIOS sterilization SA, Tarancón, Cuenca, Spain) and
148 irradiated under an electron beam radiation source, which operates at 10 MeV. The
149 radiation doses employed were between 1 and 4 kGy and the dose absorbed by the
150 samples was checked by determining the absorbance of cellulose triacetate dosimeters
151 (ASTM, 2000) simultaneously irradiated with samples. The experiments were carried
152 out in triplicate and at room temperature (18 – 20 °C). During treatment, the product
153 temperature went up by less than 2 °C. After E-beam treatment, samples were
154 transferred to the laboratory and stored at 4 °C until use.

155 **2.4. Microbial analyses**

156 To count the *L. monocytogenes* survivors, about 10 g of the material was weighed and
157 homogenized with 90 ml of a sterile saline solution in a Stomacher bag. Counts were
158 determined on the surface of plates with trypticase soy agar (Difco) and the use of a

159 spiral plate system (model Eddy Jet, IUL Instrument, Barcelona, Spain). Plates were
160 incubated at 32 °C for 24 h. Colonies were enumerated with an automatic counter
161 (Counterstat Flash model, IUL Instrument, Barcelona, Spain).

162 **2.5. Shelf-life determination**

163 The shelf-life of irradiated RTE cooked ham slices was determined by periodically
164 counting the bacterial number and by means of a sensory analysis (odor and visual
165 appearance) of samples stored at 4 °C. Non-irradiated vacuum-packaged slices were
166 used as controls. From a bacterial point of view, the shelf-life was considered to end
167 when the bacterial count was higher than 10⁷ cfu./g.

168 **2.6. Sensory analysis**

169 To determine the possible sensory differences between the non-treated (0 kGy) and
170 irradiated samples (1, 2, 3 and 4 kGy) stored at 4 °C, a triangular test, a rank order
171 test, and a descriptive test were performed. Samples were evaluated by a panel of
172 twenty tasters (ten females and ten males) selected from among the members of the
173 “Departamento de Nutrición, Bromatología y Tecnología de los Alimentos”. The
174 panelists were previously trained in the sensory assessment of meat products. In this
175 training, several models (“fresh cooked ham”, “concentrated meat broth”, “hot culture
176 medium”, “burnt beef broth”, “scalded feather”, “burnt feather”, “pungent pepper”,
177 “cooked cabbage”, “spoiled milk” and “spoiled vacuum cooked ham”) were prepared as
178 reference in order to familiarize the testers with the flavors expected to be produced by
179 the E-beam treatment. The analyses were performed as described by Cabeza et al.
180 (2008). The evaluation was carried out between meals (after breakfast but before the
181 midday meal). The samples were allowed to sit for 20 min to reach room temperature.
182 The evaluations were performed in individual booths built according to the International
183 Standards Organization DP 66.58 criteria (ISO, 1981). The tasters received unsalted
184 crackers and water at room temperature to cleanse the palate between samples. Three

185 independent tests were performed to evaluate appearance, odor, and flavor. White
186 fluorescent light was used during appearance analysis. The odor and flavor of samples
187 were evaluated under red light conditions just after opening the bags.

188 The forced-choice option of the triangle test (ISO, 1981) was chosen, in which the
189 tasters must select the sample that, in their opinion, is different. All the possible
190 combinations of untreated and irradiated samples were tested. These sensory
191 analyses were carried out on the 0 and the 1st day after treatment and on the 16 and
192 17th days of storage at 4 °C. To complement the triangle test, tasters were asked to
193 indicate their reasons for selecting one particular sample of the three used in the
194 analysis.

195 For the rank order test, the tasters were instructed to rank samples in order of
196 preference, according to the proximity of the sensory characteristic (appearance, odor,
197 or flavor) of the sample analyzed to the optimal sensory quality of the cooked ham
198 (appearance: bright red color, high marbling degree; odor: richness and intensity,
199 absence of off-odors; flavor: typical cooked ham flavor intensity, richness of taste
200 notes, cured and rancid intensity, absence of off-flavors, and intensity of aftertaste). For
201 this test, a 5-point scale (1, lowest preference; 5, highest preference) was used. No
202 repetitions were allowed. Results of the rank order test were used to obtain the sum of
203 ranks, which corresponds to the sum of scores of cooked ham preference for a specific
204 sensory characteristic (calculating the sum of the products of values given to each
205 sample on a 5-point scale multiplied by the number of times that each sample was
206 allocated to a specific score). The significance level of data obtained in these tests was
207 determined by Friedman's rank addition according to the model proposed by Joanes
208 (1985) and the tables for multiple comparison procedures for the analysis of ranked
209 data (Christensen et al., 2006). The sum of the ranks, as quantitative values of the
210 sensory evaluation, was used in the modeling, statistical analysis and optimization of

211 the irradiation process. The rank order test was performed on the 2nd and 18th days,
212 after sample treatment and storage at 4 °C.

213 Panelists were also asked to provide information about the cooked ham characteristics
214 (appearance, odor, and flavor and any off-sensory aspect) following a profile
215 descriptive analysis. This procedure was carried out on the 3rd and 19th days, after
216 irradiation treatment and storage at 4 °C.

217 **2.7. Modeling inactivation of *Listeria monocytogenes* Scott A**

218 The kinetic data of the organism inactivation was analyzed using first-order kinetics
219 (Eagerman, 1976), with the variable time replaced by the treatment dose (d_{irr}) (Eq. 1
220 and 2). $N(d_{irr})$ and $N(d_0)$ are the number of microorganisms after an irradiation dose
221 (d_{irr}) and without treatment (d_0), respectively and k is the rate constant (kGy^{-1}) for
222 given treatment conditions. k values were obtained from the linear regression analysis
223 of $\log [N(d_{irr}) / N(d_0)]$ versus dose.

$$224 \log [N(d_{irr}) / N(d_0)] = -k d_{irr} \quad (1)$$

225 **2.8. Modeling of quality factors**

226 Appearance, odor and flavor were described using adapted inactivation models, such
227 as the Gompertz Function (Eq. 2), the Weibull Distribution (Eq. 3) and the
228 Activation/Inactivation model (Eq. 4). Normally, these equations relate the response
229 factor to the variable time, however, in this work time was replaced by the irradiation
230 dose.

$$231 \log [Q(d_{irr}) / Q(d_0)] = C \exp [-\exp(A + B d_{irr})] - C \exp [-\exp(A)] \quad (2)$$

$$232 \log [Q(d_{irr}) / Q(d_0)] = -b d_{irr}^n \quad (3)$$

$$233 \log [Q(d_{irr}) / Q(d_0)] = [d_{irr} / (k_1 + k_2 d_{irr})] \left[1 - \log(1 + \exp(k_3 d_{irr}))^m \right] \quad (4)$$

234 Where $Q(d_{irr})$ and $Q(d_0)$ are the scores assigned by tasters for the quality attributes
235 (appearance, odor or flavor) after 2 and 18 days of different irradiation treatments (d_{irr})
236 and without treatment (d_0), respectively. A, B and C; b and n; m and k_i ($i=1, 2$ and 3),
237 are the kinetic constants for the Gompertz Function, the Weibull Distribution and the
238 Activation/Inactivation model, respectively. The models were fitted to the scores
239 obtained after 2 and 18 days of treatment.

240 **2.9. Modeling of the bacterial growth. Shelf-life of cooked ham.**

241 In the present study, the growth of the surviving spoilage microorganisms during the
242 storage period was fitted using Hills's model (Eq. 5). This model was developed by Hills
243 and coworkers and it is based on two concepts: synthesis and cell division and
244 biomass availability (Hills and Mackey, 1995; Hills and Wright, 1994).

$$245 \quad N(t) = N_0 - \left[\frac{k - \exp(-Pt + P \exp(-kt))}{P + k} \right] \quad (5)$$

246 Where $N(t)$ and N_0 is the microbial population after time t and the initial time,
247 respectively, after a specific irradiation dose. This model includes two different kinetic
248 parameters, P and k , which depend on the environmental factors. The shelf-life was
249 obtained from Eq. 5, taking the limit of cfu/g to be 10^7 . Eq. 5 was adapted to
250 incorporate the irradiation dose and to be able to describe the microorganism growth
251 during the storage time after different doses of treatment.

252 **2.10. Statistical analysis and optimization of the irradiation process**

253 The kinetic constants of the models were calculated by minimizing the sum of the
254 square differences between experimental and predicted data using the Excel Solver
255 tool. The root mean square error (RMSE, Eq. 6) and the coefficient of determination
256 (R^2 , Eq. 7) were used to evaluate the goodness of the fit and the accuracy of the
257 estimation. RMSE is a measure of the standard error in the estimation, whereas R^2 is a

258 statistical parameter that defines the variability explained by the model (Schemper,
259 2003).

$$260 \quad \text{RMSE} = \sqrt{\frac{\sum_{k=1}^n (y_k - y_k^*)^2}{N}} \quad (6)$$

$$261 \quad R^2 = 1 - (S_{yx}^2 / S_y^2) \quad (7)$$

262 Where y is the experimental data; y^* is the estimated value; N is the number of
263 experimental values and S_y and S_{yx} are the total standard deviation and the standard
264 deviation of the estimation, respectively.

265 The main objective of this work was to optimize the irradiation process, which involves
266 determining the dose that best preserves the quality characteristics (defined by the
267 objective function) of the product while ensuring the food safety and a minimum shelf-
268 life (restraints).

269 In the optimization problem, the models for the microbial inactivation and the changes
270 in the sensory attributes were used. In the case of the sensory properties, the models
271 selected were those describing food quality (odor, flavor and appearance) after 18 days
272 of treatment. This was because interest was focused on a shelf-life of over 2 days and
273 18 days was a more realistic storage period.

274 The optimization was based on a mathematical problem that contained the objective
275 function, the restraints and the decision variable. The objective function (to be
276 maximized) was defined as the sum of the scores for odor, flavor and appearance.

277 The prior modeling of experimental scores was necessary and this was done by using
278 different models (previously described) and selecting the best model for each attribute
279 according to the defined statistical parameters. Therefore, the sum of the selected
280 models for odor, flavor and appearance represented the objective function.

281 Two restraints were considered, one related to food safety, regarding the number of log
282 reductions that must be reached according to the European Commission's or United
283 States Department of Agriculture's safety criterion, which was calculated through Eq. 1.
284 On the other hand, the second restraint was related with the shelf-life of the product,
285 given by Eq. 5, with the minimum period before the count of 10^7 cfu/g was reached
286 being 60 days

287 Finally, the decision variable was the irradiation dose, which must be sought to
288 maximize the objective function while coping with the restraints. The Solver tool
289 (Microsoft Excel TM) was used to solve this optimization problem.

290

291 **3. Results and discussion**

292 Mathematical models were used to describe the effect of the irradiation dose on the
293 microbial inactivation and sensory characteristics of vacuum packaged cooked ham
294 slices. These equations were used in the optimization of the process and the dose
295 which minimized the effect on the quality properties (objective function) thereby
296 ensuring the safety of the product and the required shelf-life was chosen (restraints).
297 Finally, once the optimum irradiation dose was calculated for each FSO (EC, USDA),
298 the shelf-life of the product was determined through the corresponding model.

299 **3.1. Modeling inactivation of *Listeria monocytogenes* Scott A**

300 Treatments at 1, 2, 3 and 4 kGy were considered to obtain the survival curve and the
301 D-value. The experimental data obtained for the inactivation of *L. monocytogenes* in
302 cooked ham slices after E-beam irradiation was satisfactorily described by Eq. 8
303 ($R^2=0.997$, $RMSE=0.133$). The inactivation kinetics of this strain of *L. Monocytogenes*
304 is extensively reported previously (Cabeza et al., 2007).

$$305 \log N(d_{irr}) = -2.7802 d_{irr} + 7.5882 \quad (8)$$

306 The response of *L. monocytogenes* to the irradiation treatment fits first-order
307 inactivation kinetics, showing about 2.78 log reductions as the dose increases by 1
308 kGy. The strain of *L. monocytogenes* used here presented the lowest D-value (0.36
309 kGy), compared with other listeria strains (Cabeza et al, 2007). This means that, to
310 avoid the risk that comes from the routine use of *L. monocytogenes* Scott A in
311 experiments, it is possible to work normally with more resistant strains (Dion et al.,
312 1994; Grant and Patterson, 1992; Mendonca et al., 2004). Other authors have used
313 non-linear models (Gompertz) to describe the inactivation kinetics of *L. monocytogenes*
314 (Linton et al., 1995).

315 **3.2. Modeling of quality factors**

316 The panel of tasters assessed cooked ham slices after E-beam irradiation in order to
317 evaluate the sensory properties of the samples after both a short storage period (2
318 days) and also one of 18 days. The data obtained for the quality properties (odor, flavor
319 and appearance) after irradiation was fitted to three mathematical models (the
320 Gompertz Function, Weibull's Distribution and the Activation/Inactivation model). For
321 each quality property, a model was selected according to the statistical parameters, R²
322 and RMSE, and used for the optimization procedure. Table 1 shows the mean scores
323 obtained via the rank order test for the three attributes assessed, including the
324 significant differences between the doses applied for each attribute.

325 **3.2.1. Appearance**

326 The intensity of the treatment had a similar effect on the samples analyzed after both 2
327 and 18 days of storage (Figure 1). Non-treated samples and samples treated with 1
328 kGy obtained a similar preference, which indicates that this dose is not high enough to
329 affect this feature significantly. When analyzing samples stored for both 2 and 18 days,
330 it was possible to observe a clear decrease in consumer acceptance for doses of over
331 2 kGy, which shows that once this threshold is exceeded, the irradiation negatively

332 affects the appearance. In the triangular analysis, only significant differences ($p < 0.05$)
333 between non-irradiated and irradiated samples at doses of over 2 kGy were found. In
334 the color descriptive analysis, samples with 3 and 4 kGy were judged to be darker and
335 brown-gray. These sensory results coincide completely with those obtained by means
336 of the instrumental color analysis of several irradiated meat products, since a
337 progressive decrease in redness (a^* values of tristimulus colorimeter) is observed as
338 the irradiation treatment becomes more intense (Cabeza et al., 2007; Nam and Ahn,
339 2003).

340 The fact that, after 18 days, the preference of non-treated samples obtained the
341 highest score indicates that the growth of the surviving spoilage microorganism did not
342 significantly affect the appearance of cooked ham. Therefore, this attribute seems to be
343 mainly affected by the intensity of the irradiation treatment.

344 Table 2 shows the values for the different parameters obtained when the appearance
345 was fitted to the three previously described models. As regards how the dose affects
346 the appearance, the calculated statistical parameters show that, although all the
347 models provided a good fit, the Gompertz Function offered the best result ($R^2 = 0.99$;
348 RMSE = 0.001 and 0.002 for 2 and 18 days, respectively). The goodness of the model
349 fitting can also be observed in Figure 1.

350 **3.2.2. Odor and flavor**

351 Flavor is a sensory property which is defined as the combined chemical sensations of
352 taste and smell. Consequently, odor and flavor behaved in a similar way (Table 1,
353 Figure 2), after both 2 and 18 days of storage. After 2 days of irradiation, non-treated
354 samples were better accepted than treated ones. When 1 kGy was applied, a decrease
355 in the score was observed, although this dose was not high enough to provoke any
356 significant differences ($p > 0.05$) in odor and flavor, compared to non-treated samples. A
357 significant ($p < 0.05$) decrease in the acceptance of odor and flavor was observed for a
358 dose of 2 kGy, compared to non-treated samples, although the difference was not

359 significantly different ($p>0.05$) from 1 kGy treated samples. Overall, the samples were
360 poorly rated as the dose increased. In the triangular analysis, significant differences
361 ($p<0.05$) were found at doses ≥ 3 kGy as compared with 0 and 1 kGy. In the irradiated
362 samples, doses ≥ 2 kGy gave rise to weak off-odors and off-flavors defined as “hot
363 culture medium”, “burnt beef broth”, “scalded feather” and a negligible, astringent-feel
364 aftertaste. The higher the irradiation dose, the more intense the odor. These changes
365 have been observed by other authors and they have been associated with the
366 irradiation treatment (Brewer, 2009; Hampson et al., 1996; Jo et al., 1999; Nam and
367 Ahn, 2003). Brewer (2009) reported that irradiation can induce the formation of iso-
368 octane-soluble carbonyl compounds in the lipid fraction and low molecular weight, acid-
369 soluble carbonyls in the protein fraction of meat. Raising the irradiation dose increases
370 these compounds.

371 As for odor, the most widely accepted samples after 18 days of storage were those
372 treated with 1 kGy (Figure 2). The high score reached by these, compared to non-
373 treated ones, could be due to the growth of spoilage organisms which generated off-
374 odors and off-flavors, reducing the cooked ham preference. In samples treated with
375 doses of over 1 kGy, the preference began to decrease (Table 1, Figure 2). This fact
376 was not attributed to the spoilage microorganisms, which were more affected than
377 when using 1 kGy, but rather, as it happened after 2 storage days, was a consequence
378 of the dose. In the descriptive analysis, the odor of the non-irradiated samples was
379 defined as moderately lactic acid, and sour. A similar slight off-flavor was detected in
380 these samples, but not in irradiated samples. These changes may be explained by the
381 fact that the presence of a high number of spoilage organisms, presumably lactic acid
382 bacteria (De Pablo et al., 1989). In irradiated samples, however, the original spoilage
383 microbiota was effectively reduced by the E-beam in such a way that the off-odor and
384 off-flavor in the samples treated at doses ≥ 3 kGy were associated to the irradiation

385 treatment. However, in the samples irradiated at 2 kGy, the formerly perceived off-odor
386 and off-flavor were minimized during the storage time.

387 Several authors (Houser et al., 2005; Zhu et al., 2003) have reported that irradiation
388 has negative effects on the odor of some RTE meat products, such as ham and pork
389 frankfurters, while in others, like irradiated turkey ham, it increases dimethyl disulfide
390 concentration and sulphur odor and flavor as the dose rises from 0 to 2 kGy. It has also
391 been shown to induce the production of hydrocarbons, 1,7-hexadecadiene, 1,7,10-
392 hexadecatriene, and 6,9-heptade-cadiene in hams and sausages (Kwon et al., 2007).

393 Table 3 shows the values of the parameters obtained when the odor and flavor were
394 fitted to the three models (the Gompertz Function, Weibull's Distribution and the
395 Activation/Inactivation model). After 2 days of treatment, the three models satisfactorily
396 fitted the acceptance of odor and flavor, the Gompertz Function being the one that
397 provided the best result for odor (Figure 2; $R^2 = 0.998$; RMSE = 0.008) and the Weibull
398 Distribution for flavor ($R^2 = 0.998$; RMSE = 0.007). After 18 days of treatment, the
399 Activation/Inactivation model satisfactorily described odor (Figure 2) and flavor ($R^2 =$
400 0.999 ; RMSE = 0.007 and 0.003 for odor and flavor, respectively).

401 **3.3. Modeling of the bacterial growth. Shelf-life of cooked ham.**

402 The growth of surviving spoilage microorganisms in the E-beam treated vacuum
403 packaged cooked ham was progressively slower and the lag-phase longer as the dose
404 of irradiation increased (Figure 3). The growing behavior was properly described using
405 the Hills model (Eq. 5). Table 4 shows the values for the kinetic constants of the model
406 and the statistics of the fit. As can be observed, the high values of the explained
407 variance and the values of RMSE demonstrate how accurately the model fits the
408 experimental data.

409 Table 4 shows that similar values of P parameter were obtained, while the values of
410 the k parameter decreased as the dose increased. To obtain a single equation that

411 predicted the growth of spoilage microorganisms, for any irradiation dose, P was
 412 assigned a constant value of 0.21, and a relationship between k and the dose was
 413 determined using the Weibull model (Eq. 9).

$$414 \quad k(d_{irr}) = k(d_0) \cdot 10^{\left[\frac{1}{2.303} (d/\alpha)^\beta \right]} \quad (9)$$

415 Where $k(d_{irr})$ and $k(d_0)$ represent the value for a dose with treatment (d_{irr}) and without
 416 (d_0); α and β are the model constants ($\alpha=0.037$; $\beta=0.38$; $R^2=0.955$; $RMSE=0.257$). By
 417 substituting Eq. 9 in Eq. 5, Eq. 10 was obtained, which satisfactorily described the
 418 growth of spoilage microorganisms in the vacuum packaged cooked ham for the
 419 different irradiation doses applied. Although the fit for the different doses was slightly
 420 poorer (Table 4), the model (Eq. 10) permitted a good estimation of the population
 421 growth (Figure 3) and, therefore, it can be used to interpolate the growth for doses in
 422 the range of 0 to 4 kGy. Equation 10 will be used to calculate the shelf-life in the
 423 optimization procedure considering the limit to be 10^7 cfu/g.

$$424 \quad N(d_{irr}, t) = N_0 - \frac{\left[\left[\left[k(d_0) \cdot 10^{\left[\frac{1}{2.303} \left[\frac{d_{irr}}{\alpha} \right]^\beta \right]} \right] - \exp \left[P \cdot t + P \exp \left(t \cdot k(d_0) \cdot 10^{\left[\frac{1}{2.303} \left[\frac{d_{irr}}{\alpha} \right]^\beta \right]} \right) \right] \right] \right]}{P + k(d_0) \cdot 10^{\left[\frac{1}{2.303} \left[\frac{d_{irr}}{\alpha} \right]^\beta \right]}} \quad (10)$$

425

426 **3.4. Optimization of the irradiation process**

427 The optimization strategy was based on the maximization of the objective function,
 428 defined as the sum of the scores of odor, flavor and appearance. The restraints were
 429 related to the safety conditions (Eq. 1) and the shelf-life of the product (Eq. 10).

430 Initially, it was necessary to find equations which included the objective function and
 431 the restraints. These equations have been previously described and selected and they
 432 depend on the dose of treatment, which will be the decision variable.

433 The preference of the appearance and odor after 18 days was described by means of
434 the Gompertz Function, using the parameters from Tables 2 and 3. The acceptance of
435 flavor after 18 days was described through the Activation/Inactivation model, using
436 parameters from Table 3. The restraint of the minimum shelf-life (60 days) was
437 described using Eq. 10 and the safety restraint of the number of log reductions for the
438 *L. monocytogenes* population was described using Eq. 8 and it was different for EC
439 compared to USDA. Therefore, two different criteria will be considered in the
440 optimization.

441 Using the Solver tool, the dose was sought which maximized the sum of odor, flavor
442 and appearance scores (objective function) and allowed the specific number of log
443 reductions in the microbial population (safety criterion) and a minimum shelf-life of 60
444 days to be achieved.

445

446 **3.4.1. European Commission's safety criterion**

447 The safety condition established by EC required reductions of the initial microbial
448 concentration of 1.7 log cfu/g. According to the inactivation kinetics of *L.*
449 *monocytogenes*, the minimum dose of irradiation required was 0.81 kGy (Figure 4,
450 Limit EC). The optimum dose will be the one that maximizes the sum of the
451 appearance, odor and flavor scores. The best appearance score was in the range of 0
452 kGy up to 1.8 kGy (in the irradiation range where the appearance is not affected).
453 However, it can be observed that the flavor ($\log [F(d_{irr})/F(d_0)] = 0.1178$) and odor (\log
454 $[O(d_{irr})/O(d_0)] = 0.1798$) values calculated at 0.81 kGy are close but not at the maximum
455 of their curves. The intensity of 0.81 kGy reduced the initial microbial concentration by
456 1.7 log cfu/g and allowed a shelf-life of 78.5 days to be obtained (Figure 5, calculated
457 from Eq. 10).

458 When the optimization procedure was applied using the tool Solver, the optimum dose
459 was identified as 0.96 kGy. Moreover, this dose improved the safety restraint obtaining
460 2.02 log cfu/g reductions. Therefore, the EC safety criterion was met and the shelf-life
461 was extended to 79.46 days (Figure 5), longer than the imposed limit of 60 days.

462

463

464 **3.4.2. United States Department of Agriculture's safety criterion**

465 The safety criterion established by USDA required reductions of the initial microbial
466 concentration of 5.2 log cfu/g. This restraint imposes the dose be higher than 2.49 kGy.
467 The appearance, odor and flavor ($\log [A(d_{irr})/A(d_0)] = -0.0656$, $\log [O(d_{irr})/O(d_0)] = -0.0342$
468 and $\log [F(d_{irr})/F(d_0)] = -0.0900$) values calculated (Figure 4), are not close to the
469 maximum of the curves and if the dose increases, all the sensory attributes will
470 decrease (Figure 4). This dose allowed a shelf-life of 86.23 days to be obtained.

471 According to the safety levels required by USDA (5.2 log cfu/g reductions), using a
472 dose of 2.49 kGy leads to slight changes being observed in the sensory quality after 18
473 days, compared to non-treated samples. Nevertheless, once the threshold of 2.49 kGy
474 is exceeded, the irradiation will rapidly affect the ham quality, diminishing the
475 acceptability of this product. According to the safety levels required by USDA (5.2 log
476 cfu/g reductions) the optimum dose calculated through Solver tool is coincident with the
477 limit of 2.49 kGy, necessary to meet the safety restraint.

478

479 **4. Conclusions**

480 Models were formulated to describe the effect of the irradiation dose on the quality
481 attributes (odor, flavor and appearance) of vacuum packaged RTE cooked ham. The
482 spoilage microorganisms affected the odor and flavor of non-treated samples leaving
483 the appearance unaffected. In treated samples, doses of up to 2 kGy did not affect

484 appearance, while doses of around 1 kGy gave the highest scores for odor and flavor
485 after 18 storage days. The EC and USDA safety criterion was followed and the
486 optimum irradiation dose was calculated, reaching an equilibrium between the safety
487 and the quality of the meat products. Mathematical modeling and the use of
488 optimization procedures can lead to a better understanding of the irradiation process
489 and they allow the optimum operational conditions to be determined.

490

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495 **6. References**

496 Albert, I., Mafartb, P., 2005. A modified Weibull model for bacterial inactivation. Int.
497 J. Food Microbiol. 100 (1-3), 197-211.

498 Álvarez, I., Sommers, C.H., Fan, X., Niemira, B.A., 2007. Modeling the Irradiation
499 Followed by Heat Inactivation of Salmonella Inoculated in Liquid Whole Egg.
500 Journal of food science an official publication of the Institute of Food
501 Technologists, 72 (5), 145-152.

502 Arthur, T.M., Wheeler, T.L., Shackelford, S.D., Bosilevac, J.M., Nou, X.W.,
503 Koohmaraie, M., 2005. Effects of low-dose, low-penetration electron beam
504 irradiation of chilled beef carcass surface cuts on Escherichia coli O157: H7
505 and meat quality. J. Food Protect. 68 (4), 666-672.

506 ASTM. American Society for Testing and Materials. 2000. E1650-97e1 Standard
507 practice for use of cellulose acetate dosimetry systems (Vol. 12, pp. 2).

508 Barbosa-Cánovas, GV, Pothakamury, UR, Palou, E, Swanson, BG., 1998b.
509 Oscillating Magnetic fields for food processing. In: Nonthermal preservation
510 of foods. Marcel Dekker, Inc., p. 113-138.

511 Barbosa-Cánovas, GV, Pothakamury, UR., Palou, E, Swanson, BG., 1998a. High
512 intensity pulsed electric fields: Processing equipment and design. In:
513 Nonthermal preservation of foods. Marcel Dekker, Inc., p. 53-112.

514 Bermúdez-Aguirre, D., Corradini, M.G., Mawson, R., Barbosa-Cánovas, G.V., 2009.
515 Modeling the inactivation of *Listeria innocua* in raw whole milk treated under
516 thermo-sonication. *Innov. Food Sci. Emerg.* 10 (2), 172-178.

517 Brewer, M.S., 2009. Irradiation effects on meat flavour: A review. *Meat Sci.* 81, 1-
518 14.

519 Burgess, H.W., MacKrell, J., Toms, D., Karunanidhi, A., Vaidya, S., Hollinger, J.O.,
520 Grieb, T.A., Bertenshaw, G.P., 2010. Response of Bone Subjected to
521 Optimized High Dose Irradiation. *J. Biomater. Appl.* 24, 387-400.

522 Cabeza, M.C., Cambero, I., de la Hoz, L., Ordoñez, J.A., 2007. Optimization of E-
523 beam irradiation treatment to eliminate *Listeria monocytogenes* from ready-
524 to-eat (RTE) cooked ham. *Innov. Food Sci. Emerg.* 8 (2), 299-305.

525 Cabeza, M.C., Cambero, M.I., Ordoñez, J.A., de la Hoz, L., Velasco, R., 2009.
526 Safety and quality of ready-to-eat dry fermented sausages subjected to E-
527 beam radiation. *Meat Sci.* 83 (2), 320-327.

528 Cabeza, M.C., Hoz, L de la, Velasco, R., Cambero, M.I., Ordóñez, J.A., 2009.
529 Safety and quality of ready-to-eat dry fermented sausages subjected to E-
530 beam radiation. *Meat Sci.* 83, 320–327

531 Christensen, Z.T., Eggett, D.L., Dunn, M.L., Ogden, L.V., 2006. Multiple
532 comparison procedures for analysis of ranked data. *J. Food Sci.* 71 (2), 132-
533 143.

534 De Pablo, B., Asensio, M.A., Sanz, B., Ordoñez, J.A., 1989. The D(-) lactic acid and
535 acetoin/diacetyl as potential indicators of the microbial quality of vacuum-
536 packed pork and meat products. *J. Appl. Bacteriol.* 66 (3), 185-190.

537 Ding, T., Oh, D., Jin, Y., 2010. Predictive model for growth of *Listeria*
538 *monocytogenes* in untreated and treated lettuce with alkaline electrolyzed
539 water. *Word J. Microb. Biot.* 26 (5), 863-869.

540 Dion, P., Charbonneau, R., Thibault, C., 1994. Effect of ionizing dose rate on the
541 radioresistance of some food pathogenic bacteria. *Can. J. Microbiol.* 40 (5),
542 369-374.

543 Eagerman, B. A., Rouse, A. H., 1976. Heat inactivation temperature time
544 relationships for pectinesterase inactivation in citrus juices. *J. Food Sci.* 41
545 (6), 1396-1397.

546 Elez-Martinez, P., Aguilo-Aguayo, I., Martin-Belloso, O., 2006. Irradiation of orange
547 juice peroxidase by high-intensity pulsed electric fields as influenced by
548 process parameters. *J. Sci. Food Agr.* 86, 71-81.

549 Gibson, A.M., Bratchell, N., Roberts, T.A., 1987. The effect of sodium chloride and
550 temperature on the rate and extent of growth of *Clostridium botulinum* type A
551 in pasteurized pork slurry. *J. Appl. Bacteriol.* 62 (6),479-490.

552 Gil-Díaz, M., Santos-Delgado, M. J., Rubio-Barroso, S., Polo-Díez, L. M., 2009.
553 Free D-amino acids determination in ready-to-eat cooked ham irradiated with
554 electron-beam by indirect chiral HPLC. *Meat Sci.* 82, 24–29.

555 Grant, I. R., Patterson, M. F., 1992. Sensitivity of foodborne pathogens to irradiation
556 in the components of a chilled ready meal. *Food Microbiol.* 9 (2), 95–103.

557 Hampson, J. W., Fox, J. B., Lakritz, L., Thayer, D. W., 1996. Effect of low dose
558 gamma radiation on lipids in five different meats. *Meat Sci.* 42 (3), 271–276.

559 Hills, B.P., Mackey, B.M., 1995. Multi-compartment kinetic models for injury,
560 resuscitation, induced lag and growth in bacterial cell populations. *Food*
561 *Microbiol.* 12 (4), 333-346.

562 Hills, B.P., Wright, K.M., 1994. A new model for bacterial growth in heterogeneous
563 systems. *J. Theor. Biol.* 168 (1), 31-41.

564 Houser, T. A., Sebranek, J. G., Maisonet, W. N., Cordray, J. C., Wiegand, B. R.,
565 Ahn, D. U., 2005. The effects of irradiation at 1.6 kGy on quality
566 characteristics of commercially produced ham and pork frankfurters over
567 extended storage. *J. Food Sci.* 70 (4), S262-S266.

568 Hoz, L., Cambero, M. I., Cabeza, M. C., Herrero, A. M., Ordoñez. J. A., 2008.
569 Elimination of *Listeria monocytogenes* from Vacuum-Packed Dry-Cured Ham
570 by E-Beam Radiation. *J. Food Protect.* 71 (10), 2001-2006.

571 Hoz, L., Cambero, M.I., Cabeza, C., Herrero, A. M., Ordóñez, J. A., 2008.
572 Elimination of *Listeria monocytogenes* from Vacuum-Packed Dry-Cured Ham
573 by E-Beam Radiation. *J. Food Protect.* 71, 2001–2006.

574 Huang, L. 2010. Growth kinetics of *Escherichia coli* O157:H7 in mechanically-
575 tenderized beef. *Int. J. Food Microbiol.* 30 (140), 40-48.

576 IAEA (International Atomic Energy Agency). 2003. Radiation processing for safe
577 shelf-stable and ready-to-eat food. IAEA-TECDOC-1337. Proceedings of a
578 final research Co-ordination meeting held in Montreal, Canada, 10-14 July
579 2000. Food and Environmental Protection Section. International Atomic
580 Energy Agency. Vienna. Austria.

581 International Commission on Microbiological Specifications for Foods., 1996.
582 *Salmonellae*, In: *Microorganisms in foods*, Volume 5. Microbiological
583 specifications of food pathogens. New York: Academic Press, pp. 217–264.

584 ISO. International Standards Organization, 1981. Analyse sensorielle guide pour
585 l'implantation d'un local destiné aux analyses sensorielles (ISO-DP 6658).
586 Genève, Switzerland: International Organization for Standardization.

587 Jacxsens, L, Devlieghere, F., Debevere, J., 2002. Temperature dependence of
588 shelf-life as affected by microbial proliferation and sensory quality of
589 equilibrium modified atmosphere packaged fresh produce. *Postharvest Biol*
590 *Technol.* 26, 59-73.

591 Jo, C., Lee, J. I., Ahn, D. U., 1999. Lipid oxidation, color changes and volatile
592 production in irradiated pork sausage with different fat content and packaging
593 during storage. *Meat Sci.* 51 (4), 355-361.

594 Joanes, D. N., 1985. On a rank sum test due to Kramer. *J. Food Sci.* 50 (5), 1442-
595 1444.

596 Knorr, D., Zenker, M., Heinz, V., Lee, DU., 2004. Applications and potential of
597 ultrasonics in food processing. *Trends in Food Sci. & Techn.* 15, 261-266.

598 Kwon, J. H., Kausar, T., Lee, J., Kim, H. K., Ahn, D. U., 2007. The microwave-
599 assisted extraction of fats from irradiated meat products for the detection of
600 radiationinduced hydrocarbons. *Food Sci. Biotechnol.* 16 (1), 150-153.

601 Lakshmanan, R., Dalgaard, P., 2004. Effect of high-pressure processing on *Listeria*
602 *monocytogenes*, spoilage microflora and multiple compound quality indices
603 in chilled cold-smoked salmon. *J. Appl. Microbiol.* 96, 398–408.

604 Lee, E. J., Ahn, D. U., 2005. Quality characteristics of irradiated turkey breast rolls
605 formulated with plum extract. *Meat Sci.* 71 (2), 300-305.

606 Lee, M., Sebranek, J. G., Olson, D. G., Dickson, J. S., 1996. Irradiation and
607 packaging of fresh meat and poultry. *J. Food Protect.* 59 (1), 62-72.

608 Linton, R.H., Carter, W.H., Pierson, M.D., Hackney, C.R., Eifert, J.D., 1995. Use of
609 a modified Gompertz equation to predict the effects of temperature, pH and
610 NaCl on the inactivation of *Listeria Monocytogenes* Scott A. *J. Food Protect.*
611 59 (9), 16-23.

612 Marselles-Fontanet, A.R., Martin-Belloso, O., 2007. Optimization and validation of
613 PEF processing conditions to inactivate oxidative enzymes of grape juice. *J.*
614 *Food Eng.* 83 (3), 452-462.

615 Maurice, J., 1994. The rise and rise of food poisoning. *New Sci.* 144, 28-33.

616 McKellar, R.C., X.W. Lu., 2004. In: CRC Press Boca Ratón (Eds), *Modeling*
617 *Microbial Response in Food.* Florida, EE.UU.

618 Mendonca, A. F., Romero, M. G., Lihono, M. A., Nannapaneni, R., Jonson, M. G.,
619 2004. Radiation resistance and virulence of *Listeria monocytogenes* Scott A
620 following starvation in physiological saline. *J. Food Protect.* 67 (3), 470-474.

621 Nam, K. C., Ahn, D. U., 2003. Use of antioxidants to reduce lipid oxidation and off-
622 odor volatiles of irradiated pork homogenates and patties. *Meat Sci.* 63 (1),
623 1-8.

624 Patterson, M.F., Damoglou, A.P., Buick, R.K., 1993. Effects of irradiation dose and
625 storage temperature on the growth of *Listeria monocytogenes* on poultry
626 meat. *Food Microbiol.* 10, 197–203.

627 Rababah, T., Over, K., Hettiarachchy, N.S., 2010. Infusion of plant extracts during
628 processing to preserve quality attributes of irradiated chicken breasts over 9
629 months storage at -20 °C. *J. Food Process Pres.* 34 (1), 287-307.

630 Schemper, M., 2003. Predictive accuracy and explained variation. *Stat Med.* 22,
631 2299-2308.

632 Schilling, M.W., Yoon, Y., Tokarskyy, O., Pham, A.J., Williams, R.C., Marshall, D.L.,
633 2009. Effects of ionizing irradiation and hydrostatic pressure on *Escherichia*
634 *coli* O157:H7 inactivation, chemical composition, and sensory acceptability of
635 ground beef patties. *Meat Sci.* 81 (4), 705-710.

636 Shi, J., Le Maguer, M., 2000. Lycopene in tomatoes chemical and physical
637 properties affected by food processing. *CRC. Cr. Rev. Food. Sci.* 40 (1), 1-
638 42.

639 Sommers, C., Fan, X., Niemira, B.A., Sokorai, K., 2003. Radiation (gamma)
640 resistance and postirradiation growth of *Listeria monocytogenes* suspended
641 in beef bologna containing sodium diacetate and potassium lactate. *J Food*
642 *Prot*, 66, 2051-2056.

643 Soysal, Ç. 2008. Kinetics and thermal activation/inactivation of starking apple
644 polyphenol oxidase. *J. Food Process Pres.* 32 (6), 1034-1046.

645 Thayer, D.W., Boyd, G., Muller, W.S., Lipson, C. A., Hayne, W.C., Baer, S.H.,
646 1990. Radiation resistance of *Salmonella*. *J Ind Microbiol.* 5, 383-390.

647 Vogel, B.F., Ng, Y.Y., Hyldig, G., Mohr, M., Gram, L., 2006. Potassium lactate
648 combined with sodium diacetate can inhibit growth of *Listeria*
649 *monocytogenes* in vacuum packed cold smoked salmon and has no adverse
650 sensory effects. *J. Food Prot.* 69, 2134–2142.

651 Wang, P. 2010. Robust Growth of *Escherichia coli*. *Curr. Biol.* 20 (12), 1099-1103.

652 Wang, T., MacGregor, S.J., Anderson, J.G., Woolsey, G.A., 2005. Pulsed ultra-
653 violet inactivation spectrum of *Escherichia coli*. *Water Res.* 39, 2921–2925.

654 Zhu, M. J., Lee, E. J., Mendonca, A., Ahn, D. U., 2003. Effect of irradiation on the
655 quality of turkey ham during storage. *Meat Sci.* 6 (1), 63-68.

656 Zhu, M., Du, M., Cordray, J., Ahn, D. U. 2005. Control of *Listeria monocytogenes*
657 contamination in ready-to-eat meat products. *Comprehensive Reviews in*
658 *Food Science and Food Safety.* 4, 34–42.

1 **FIGURE CAPTIONS**

2 **Figure 1.** Effect of E-beam irradiation on the appearance of vacuum packaged RTE cooked
3 ham after 2 and 18 storage days. Continuous line: Gompertz Function.

4 **Figure 2.** Effect of E-beam irradiation on the odor of vacuum packaged RTE cooked ham after
5 2 and 18 days. Continuous line: Gompertz function; dashed line: Activation/Inactivation model.

6 **Figure 3.** Growth of spoilage microorganisms on vacuum packaged RTE cooked ham after
7 different doses of E-beam irradiation. Continuous line: modified Hill's model.

8 **Figure 4.** Effect of E-beam irradiation on the sensory properties (Q: appearance, odor or flavor;
9 A: appearance; F: flavor; O: odor). A, F and O → modeled curves. EC limit: minimum dose of
10 0.81 kGy to reach reductions of the initial microbial concentration of 1.7 log cfu/g; Optimum
11 dose for EC criterion: 0.96 kGy; Limit and optimum dose for USDA criterion: 2.49 kGy to reach
12 reductions of initial microbial concentration of 5.2 log cfu/g.

13 **Figure 5.** Modeling of the spoilage microorganism's growth after 0.814 kGy (dotted line), 0.96
14 kGy (continuous line) and 2.49 kGy (dashed line) of E-beam irradiation treatment.

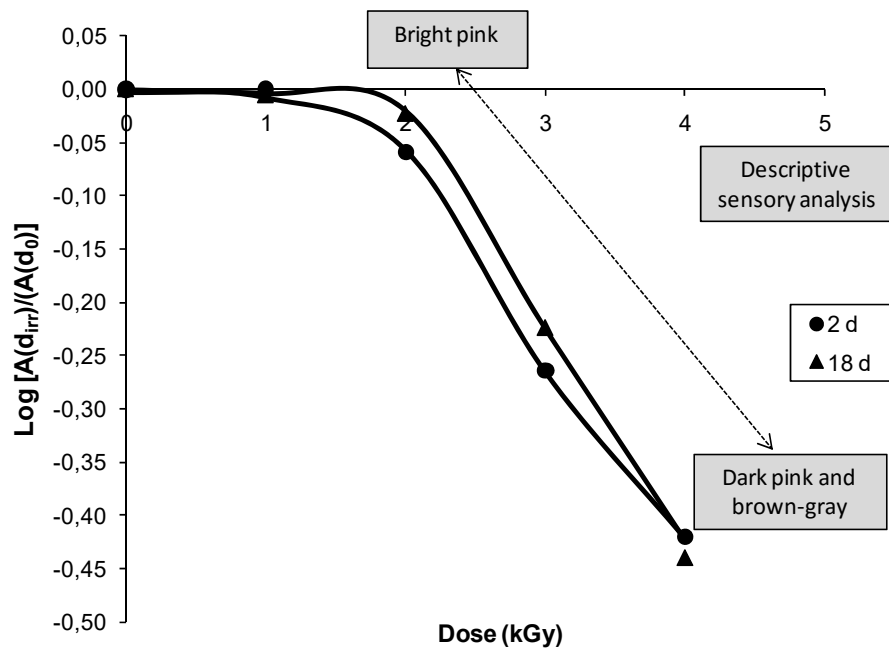


Figure 1.

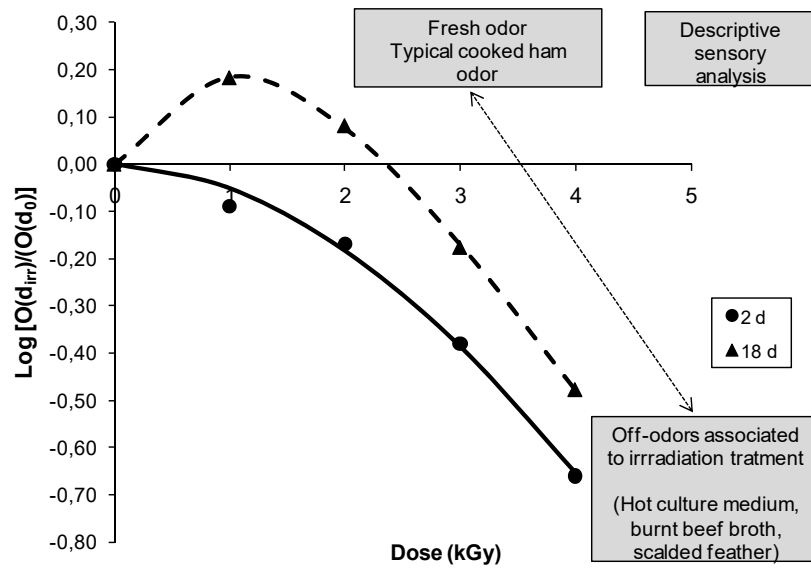


Figure 2.

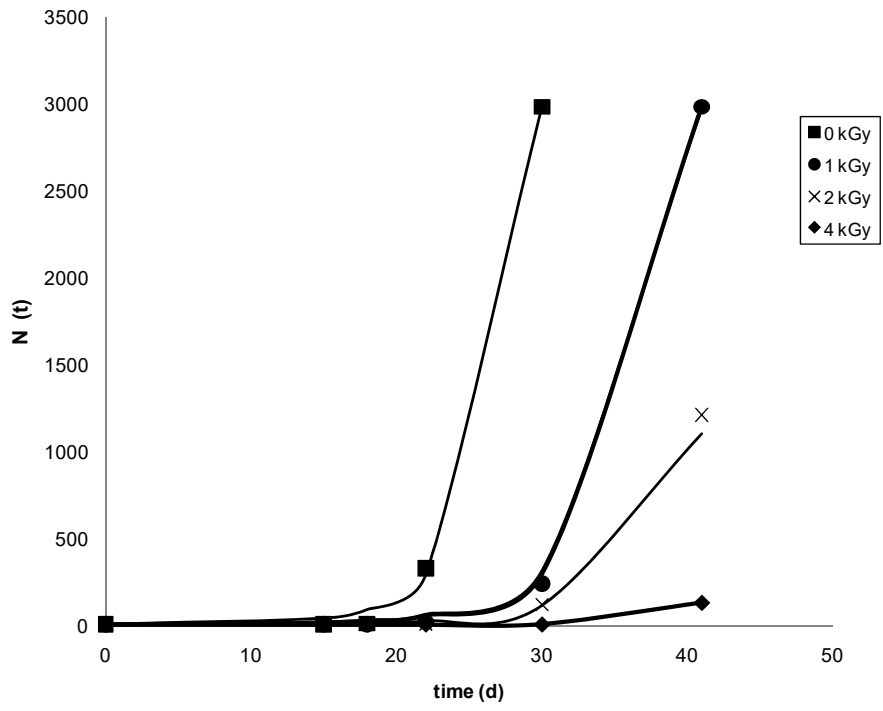


Figure 3.

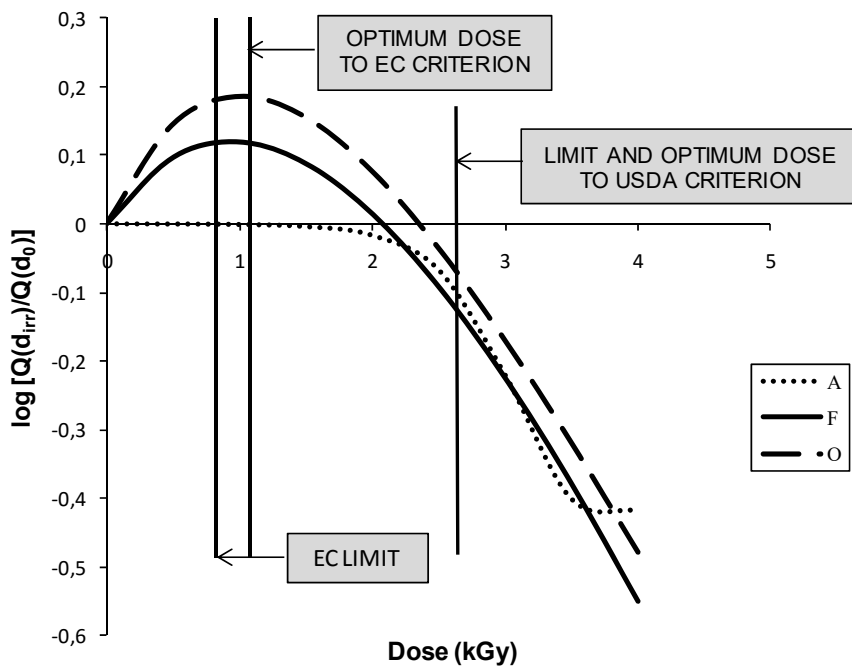


Figure 4.

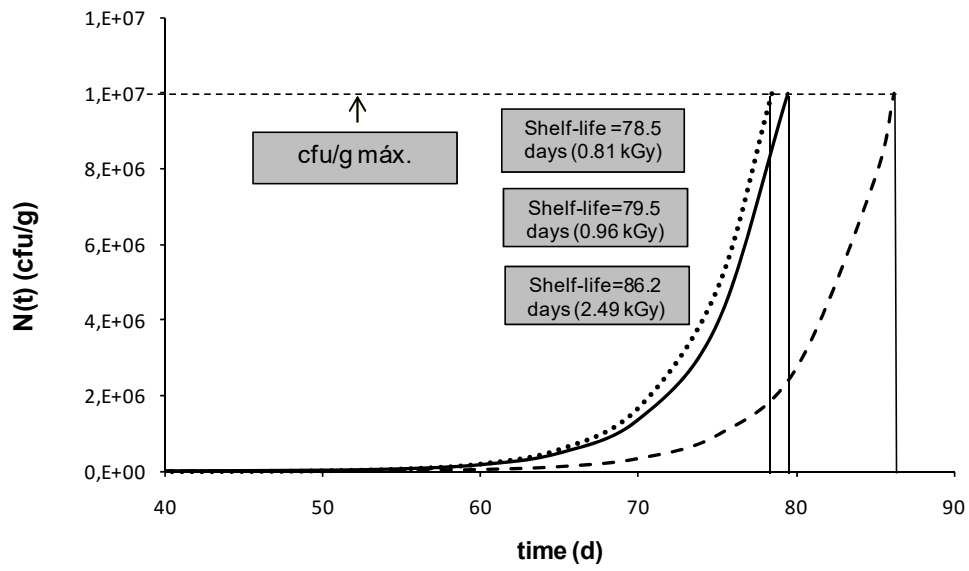


Figure 5.

Table 1. Sensory evaluation by rank order test of vacuum packaged RTE cooked ham after 2 and 18 days of irradiation treatment.

Irradiation treatment (kGy)	Appearance		Odor		Flavor	
	2 days	18 days	2 days	18 days	2 days	18 days
0	79 _a	77 _a	96 _a	63 _{b,c}	91 _a	71 _a
1	79 _a	76 _a	78 _{a,b}	96 _a	81 _{a,b}	93 _a
2	69 _{a,b}	73 _{a,b}	65 _{b,c}	76 _{a,b}	60 _{b,c}	74 _a
3	43 _{b,c}	46 _{b,c}	40 _{c,d}	42 _c	41 _{c,d}	42 _b
4	30 _c	28 _c	21 _d	21 _c	27 _d	20 _b

Final score= (N1 X 1) + (N2 X 2) + (N3 X 3) + (N4 X 4) + (N5 X 5), where N1, N2, N3, N4, and N5 are the number of panellists that ranked the sample in position 1 (minimal preference), 2, 3, 4, or 5 (maximum preference) in the rank order test. Within a column, values with different letters are significantly different (P < 0.05).

Table 2. Estimated model parameters for modeling the effect of the irradiation dose on the appearance of vacuum packaged RTE cooked ham after 2 and 18 storage days.

	Storage time	Parameters	R ²	RMSE	
Gompertz Function	2 days	A	-5.725	0.999	0.001
		B	1.906		
		C	0.422		
	18 days	A	-7.972		
		B	2.544		
		C	0.440		
Weibull Distribution	2 days	b	0.019	0.973	0.030
		n	2.259		
	18 days	b	0.008		
		n	2.880		
Inactivation / Activation model	2 days	k ₁	4.201	0.932	0.048
		k ₂	99.852		
		k ₃	27.758		
		m	1.594		
	18 days	k ₁	2.500		
		k ₂	99.888		
		k ₃	27.743		
		m	2.179		

Table 3. Estimated model parameters for modeling the effect of the irradiation dose on the odor and flavor of vacuum packaged RTE cooked ham after 2 and 18 storage days.

	Storage time	ODOR				FLAVOR				
		Parameters	R ²	RMSE	Parameters	R ²	RMSE			
Gompertz Function	2 days	A	-3.426	0.998	0.008	A	-2.477	0.995	0.019	
		B	0.522			B	0.804			
		C	3.329			C	0.662			
	18 days	A	-52.403	0.701	0.142	A	7.382	0.901	0.083	
		B	17.209			B	-2.474			
		C	0.477			C	-0.596			
Weibull Distribution	2 days	b	0.052	0.992	0.024	b	0.061	0.998	0.007	
		n	1.825			n	1.564			
	18 days	b	0.001	0.770	0.124	b	0.003	0.908	0.080	
		n	4.549			n	3.779			
	Inactivation / Activation model	2 days	k ₁	52.564	0.996	0.013	k ₁	52.624	0.996	0.013
			k ₂	37.859			k ₂	38.085		
k ₃			-3.574	k ₃			-0.769			
m			0.352	m			0.383			
18 days		k ₁	0.935	0.999	0.007	k ₁	1.054	0.999	0.003	
		k ₂	0.942			k ₂	1.249			
		k ₃	0.225			k ₃	0.081			
		m	0.405			m	0.322			

Table 4. Estimated kinetic constants and statistical parameters (R^2 ; RMSE) obtained from the fit of Hill's model to the growth of remaining spoilage microorganisms in irradiated vacuum packaged RTE cooked ham.

Dose (kGy)	Hill's model				Hill's model (k depending on dose)		
	P	k	R^2	RMSE (cfu/g)	k	R^2	RMSE (cfu/g)
0	0.205	0.940	0.998	54	0.554	0.978	189
1	0.178	0.063	0.998	41	0.016	0.999	36
2	0.218	0.004	0.999	13	0.005	0.999	12
4	0.222	0.0004	0.998	2	0.001	0.914	116