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Additional Information

**Functional, thermal and rheological properties of high fibre fresh pasta:  
effect of tiger nut flour and xanthan gum addition**

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## **ABSTRACT**

Tiger nut flour (TNF) is a rich source of dietary fibre with potential to be used in cereal based products. However, research on improving the rheological properties of tiger nut based doughs is limited. In this paper, the significance of TNF and xanthan gum (X) incorporation into fresh egg pasta, in terms of its thermal and dynamic rheological properties, has been investigated. Plain semolina pasta (DWS) was used as control. High fibre doughs (20 and 40% TNF) with or without X (0 and 1 %) were assessed. Both fundamental (dynamic oscillatory and creep tests) and empirical (texture profile analysis) tests were performed to assess the viscoelasticity of TNF-DWS composite blends. Raw solids (TNF, DWS) were characterised in terms of their chemical composition, particle size distribution and functional properties. For both fresh and cooked pasta, water activity, water content and gelatinisation temperatures were estimated. The results from the rheological tests revealed that partial replacement of DWS by TNF lead to less cohesive and weaker structures due to the lower presence of a gluten network. X significantly improved the rheological response of the TNF based doughs. Thermal analysis showed a single endothermic peak in the temperature range between 60 and 78°C during heating, which corresponds to the amylopectin gelatinisation. However, when replacing 40% of DWS by TNF, two-phase transitions were observed, probably associated to the starch tiger nut gelatinisation or the formation of amylose-lipid complexes. The optimum cooking time for the tiger nut pasta was two minutes as determined by calorimetric analysis.

Keywords: fresh pasta; tiger nut flour; fibre; xanthan gum; dynamic rheology; DSC

## 1. Introduction

Nowadays, it is a fact that consumers increasingly demand healthier foods obtained by using raw ingredients that are naturally high in components with health-promoting effects. Epidemiological studies show a correlation between diets which are moderate to high in fibre and a lower incidence of some chronic disorders such as cardiovascular diseases (Kendall et al. 2010), diabetes (Chandalia et al. 2000; Schulze et al. 2004), colon cancer (WHO 2003), gastrointestinal disorders (Kaczmarczyk et al. 2012) and obesity (Buttriss and Stokes 2008). It is also a fact that the daily fibre intake per capita is still today lower than that recommended by the WHO (25 g / day, Romo et al. 2008). Tiger nut flour is a rich source of dietary fibre (8-15g/100g), minerals such as phosphorus, potassium, iron and calcium, and vitamins E and C (Sánchez-Zapata et al. 2012). It also has a high lipid content (23-31 g/100 g) with a fatty acid profile similar to olive and hazelnut oil (predominance of oleic, linoleic and linolenic acids) (Aguilar et al. 2015; Demirkesen et al. 2013). Tiger nut flour has a moderate amount of proteins with a demonstrated higher amount of essential amino acids (such as lysine, cystine, arginine and histidine) than the protein standard proposed by the FAO/WHO (1985) for satisfying adult needs (Ade-Omowaye et al. 2008). Therefore, tiger nut flour incorporation into pasta products -a staple cereal-based food that is widely produced and consumed across the world, easy to prepare and with a low price for consumers- could help to overcome this deficit whilst providing health benefits (Aravind et al. 2012; Yokoyama et al. 1997). Other recent studies have been assessed the possibility of turning pasta into a functional food with health benefits by adding ingredients rich in dietary fibre, such as teff (Giuberti et al. 2016), kimchi by-product (Kim et al. 2017), bran (la Gatta et al. 2017; Kaur et al. 2012), brewer's spent grains

27 (Cappa and Alamprese, 2017) or tomato by-product (Padalino et al. 2017).  
28 Considering the nutritional value of tiger nut flour, it may be a potential and  
29 interesting alternative to improve not only the fibre content of wheat formulations  
30 but also to provide a better nutritional quality. Pasta products made from wheat  
31 are characterised by proteins that form a viscoelastic gluten network, providing a  
32 certain cohesiveness to the dough and contributing to the cooked pasta's water  
33 holding capacity (Wieser 2007). Cooked pasta made from durum wheat semolina  
34 maintains good texture and resists surface disintegration (Mastromatteo et al.  
35 2011). Replacement of wheat semolina by significant amounts of non-gluten  
36 forming flours such as tiger nut flour can seriously constrain dough viscoelasticity  
37 and the cooking behaviour of dough matrices that have a weakened mixed  
38 protein network. In a previous work, we investigated the effect of incorporating  
39 tiger nut flour in up to 30% w/w semolina basis into fresh egg pasta (Albors et al.  
40 2016). The obtained results revealed a need to strengthen the protein network  
41 and thus reduce cooking losses and increase the pasta's firmness (Albors et al.  
42 2016). One way to overcome this problem is to use hydrocolloids with high water  
43 binding capacity, such as xanthan gum, to limit the swelling of the tiger nut fibre.  
44 Xanthan gum has strong viscoelastic properties, adopting in aqueous solutions a  
45 double-stranded helix rigidly ordered conformation. It can be used to mimic the  
46 properties of gluten to form the elastic texture of pasta (Larrosa et al. 2013).  
47 Previous trials (paper under publication) have demonstrated that the use of  
48 xanthan gum over 0.8% (DWS basis) may lead to better mechanical properties  
49 and cooking behaviour of tiger nut pasta. Cai et al. (2016) found positive  
50 correlations between xanthan gum content and tensile strength and texture  
51 properties of noodles. And Romero et al. (2017) found an improvement in the  
52 network strength of proso millet pasta with xanthan gum. The rheological and

53 mechanical characterisation of tiger nut dough pasta is important in predicting  
54 the processing behaviour and controlling the quality of fresh pasta products. The  
55 interactions between dough components (starch, protein, fibre, water and  
56 additives) as well as the flour's functional properties and particle size play an  
57 important role in the structure development and thus the rheological and thermal  
58 properties of fresh pasta (Chillo et al. 2009). To the author's knowledge, no data  
59 has been published on fresh pasta affected by tiger nut and xanthan gum  
60 incorporation. Therefore, this paper is aimed at: (i) assessing the functional  
61 properties, particle size distribution and chemical composition of wheat semolina  
62 and tiger nut flour, and their mixtures; (ii) assessing the effect of these  
63 ingredients (tiger nut flour and xanthan gum) on the rheological and thermal  
64 properties of "high fibre" fresh pasta (>6% w/w, Official Journal of European  
65 Commission, 2006). The fundamental (dynamic tests) and empirical (texture  
66 profile analysis) rheological properties, as well as the starch-water interactions  
67 (differential scanning calorimetry) were assessed in tiger nut wheat doughs and  
68 compared to wheat semolina counterparts.

## 69 **2. Material and methods**

### 70 2.1 Experimental design

71 Wheat semolina was replaced with TNF up to 40% (w/w) to obtain pasta with  
72 more than 6% (w/w) of fibre content ("high fibre"), according to the Nutritional  
73 Claims for Dietary Fiber Foods (Official Journal of European Commission, 2006).  
74 The fibre content was estimated considering the chemical composition of the raw  
75 materials (Table 1). The obtained values for tiger nut pasta ranged from 7.6 to  
76 8.6 g/100 g pasta. Six formulations were evaluated in total, considering three  
77 levels for wheat semolina replacement (0%, 20% and 40% w/w, named  
78 henceforth as S, TNF20 and TNF40 respectively) and two for xanthan gum (0%

79 and 1% w/w, this last one being noted with an X at the end of the code, that is,  
80 TNF20X, TNF40X and SX). The percentage of X was chosen taking into account  
81 the obtained results in previous trials with different hydrocolloids (paper under  
82 publication). TNF and DWS were evaluated in terms of their chemical  
83 composition, functional properties and particle size distribution. For fresh pasta  
84 doughs, the following analyses were assessed: water content, water activity,  
85 fundamental (oscillatory and creep tests) and empirical (textural profile analysis,  
86 TPA) rheological properties, and differential scanning calorimetry (DSC). All  
87 measurements were carried out in triplicate.

## 88 2.2 Raw materials and characterisation

89 Commercial durum wheat semolina –abbreviated as DWS– (Harinas Villamayor,  
90 S.A., Huesca, Spain), tiger nut flour –abbreviated as TNF–(Tigernuts Traders  
91 S.L., Valencia, Spain) and xanthan gum 1400 cps –abbreviated as X– (E-415)  
92 (Shandong Fufeng Fermentation Co. Ltd., China) were used. Fresh eggs and  
93 mineral water were purchased in a local market. 2.2.1 Proximate composition

94 DWS and TNF were analysed for their water, protein, fat and ash content  
95 according to the American Association of Cereal Chemists' approved methods  
96 (AACC, 2000) and for their total, soluble and insoluble fibre according to the  
97 Megazyme method K-TDFR (Megazyme Ltd., Ireland). Digestible carbohydrates  
98 were determined by difference (100 – percentage of estimated proximate  
99 chemical composition). Three replicates were carried out for each analysis.

### 100 2.2.2 Particle size analysis

101 The particle size distribution (PSD) of both DWS and TNF was determined by  
102 applying the laser diffraction method and Mie theory, following the ISO13320  
103 normative (AENOR 2009). A Laser diffractometer (Mastersizer 2000, Malvern  
104 Instruments Ltd., Worcestershire, U.K.), equipped with a PS 65 (dry sample) or a

105 wet sample dispersion unit (Malvern Instruments Ltd., Hydro 2000 MU, U.K.) was  
106 employed to perform, respectively, dry and wet analyses. The particle refraction  
107 and absorption indexes were 1.52 and 0.1, respectively, and the water refraction  
108 index was 1.03. Distributions were made in triplicate and for each sample, and  
109 10–20 g of flour was used. Size distribution was quantified as the relative volume  
110 of particles in size bands, presented as size distribution curves (Malvern  
111 MasterSizer Micro software v 5.40). The PSD parameters recorded included  
112 largest particle size  $d(0.9)$ , mean particle volume  $d(0.5)$ , smallest particle size  
113  $d(0.1)$ , and mean particle diameter/volume mean diameter ( $D[4,3]$ ). The Span  
114 value or measurement of the width of the size distribution, calculated from the  
115 values of standard percentiles, was complementarily reported. The wider the  
116 particle size distribution, the bigger the Span becomes. The fundamental size  
117 distribution by laser diffraction method is expressed in terms of equivalent  
118 spheres (the results are volume based), so the number distributions should only  
119 be considered as a guide of the volume distribution.

### 120 2.2.3 Functional properties

121 The functional properties of both DWS and TNF, and their blends with or without  
122 X, were determined as follows. Solvent retention capacity (SRC) was determined  
123 according to the AACC method 56-11 to quantify potential contributions to water-  
124 holding capacity by flour components having water-uptake capabilities (Paul and  
125 Minn 2005). The used solvents were sucrose (50% w/v), sodium bicarbonate  
126 (5% w/v) and lactic acid (5% v/v). The water-holding capacity (WHC) was  
127 determined by using the modified methods from Heywood et al. (2002) and Lin  
128 and Zayas (1987). The fat adsorption capacity (FAC) was determined according  
129 to Ahn et al. (2005). Foam capacity (FC) and foam stability (FS) were determined  
130 as described by Narayana and Narasinga Rao (1982) and modified by Alu'datt et



131 al. (2012).

### 132 2.3 Pasta preparation

133 The S formulation, used as the control sample, was obtained by mixing durum  
134 wheat semolina (71% w/w), fresh egg (13% w/w) and water (16% w/w). For the  
135 other formulations, durum wheat semolina was replaced by TNF and X at  
136 different levels, as described in section 2.1. All raw materials were mixed and  
137 kneaded in an electric cooking device (Thermomix TM-31, Vorwerk Spain M.S.L.,  
138 S.C., Madrid). The dried (semolina/tiger nut flour/xanthan gum) and liquid  
139 (egg/water) components were separately mixed at low speed (set 2) for 45 s.  
140 The resulting blends were then kneaded in two steps for 5 min each (with a rest  
141 period of 5 min between them). The resulting doughs were rested for 20 min at 4  
142 °C inside a plastic bag in order to enable sample relaxation. Afterwards, the fresh  
143 pasta sheets ( $1.0 \pm 0.03$  mm thick) were formed by using a domestic pasta making  
144 machine (Simplex SP150, Imperia, Italy) coupled with a specific motor (A2500,  
145 Imperia, Italy).

### 146 2.4 Water content and water activity of fresh pasta doughs

147 The water content ( $x_w$ , g/g) was determined according to the AACC-approved  
148 gravimetric method 44-40 (AACC 2000). The AquaLab Series 4 TEV equipment  
149 (Decagon, CX-1, sensitivity 0.001) was used to measure water activity ( $a_w$ ).

### 150 2.5 Small deformation mechanical test: oscillatory and creep tests

151 A RheoStress rheometer (RS1-Thermo Haake, Karlsruhe, Germany) with  
152 serrated parallel plate geometry surfaces (60 mm diameter, 1 mm gap) was used  
153 to perform the oscillatory and creep tests on the pasta doughs. Dough excess  
154 was carefully removed and a thin layer of mineral oil was applied to cover the  
155 exposed sample surfaces during the measurements. Before the measurements,  
156 the dough was rested for 5 min to allow relaxation. Temperature analysis was 20

157 °C. The linear viscoelastic region (LVR) was identified and established for each  
158 dough by means of stress sweep tests from 5 to 900 Pa at 1Hz. Frequency  
159 sweeps were carried out from 10 to 0.1 Hz in the LVR (between 45 y 68 Pa,  
160 depending on the pasta sample).

161 Creep tests were performed by applying an instantaneous and constant shear  
162 stress in the LVR for 300 s. Creep data are presented in terms of creep  
163 compliance, J, which is defined as the strain divided by the stress applied. Each  
164 test was performed in triplicate.

## 165 2.6 Empirical rheological test: TPA

166 A TA.XT2 Texture Analyser (Stable Micro Systems, Godalming, Surrey, UK) was  
167 used to perform the texture profile analysis (TPA) of the pasta dough. The TPA  
168 analysis provides a significant measurement of the textural characteristics of the  
169 product (Szczesniak 2002; Olivera and Salvadori 2006) and the dough  
170 machinability (Angioloni and Dalla Rosa 2007). The settings of the experiments  
171 were the following: 50% compression, 30 mm diameter probe (flat-end aluminium  
172 compression disc), test speed 1 mm/s, 75 s gap between compressions, 5 kg  
173 load cell, 5 mm dough thickness. The data was processed using Texture  
174 Exponent 6.1.7 (Stable Micro Systems Software).

## 175 2.7 Thermal properties of fresh pasta doughs

176 Starch gelatinisation temperature ( $T_p$ ) was measured in duplicate for each pasta  
177 dough by differential scanning calorimetry (DSC 1, Mettler Toledo, España). The  
178 freeze-dried (LyoAlfa 10-85, Telstar, Spain) pasta samples (about 5 mg) were  
179 placed into 40  $\mu$ l aluminium hermetic pans (Mettler Toledo, Spain) and 15  $\mu$ l of  
180 water were added. The pans were placed at 4 °C for 24 h to equilibrate the water  
181 content. The heating rate of the samples in the calorimeter was 5 °C/min and the  
182 analysis was carried out between 20 to 80 °C. Air was used as reference. Onset

183 ( $T_o$ ), peak ( $T_p$ ) and end ( $T_e$ ) temperatures, and gelatinisation enthalpy ( $\Delta H_n$ ) were  
184 recorded. The same procedure was used for the cooked pasta samples at  
185 different cooking times to determine the optimal cooking time (complete starch  
186 gelatinisation). The cooking process was carried out as described on Albors et al.  
187 (2016).

## 188 2.8 Field emission scanning electron microscope (FESEM)

189 FESEM (ULTRA 55, Carl Zeiss AG, Oberkochen, Germany) was used in order to  
190 observe the starch granules of tiger nut flour. The samples (cross-section) were  
191 fixed on copper stubs, platinum coated and observed using an accelerating  
192 voltage of 2 kV.

## 193 2.9 Statistical analysis

194 Analysis of variance (ANOVA) was carried out by using Statgraphics Centurion  
195 software version 16.1.17 (StatPoint Technologies, Inc., Warrenton, VA) in order  
196 to evaluate the effects of partial semolina replacement by tiger nut flour and  
197 xanthan gum on the measured parameters. The significance level was  $p= 0.05$  in  
198 all cases.

## 199 **3. Results and discussion**

### 200 3.1 DWS and TNF characterisation: chemical composition, functional properties 201 and particle size distribution

202 The chemical composition and the particle size distribution (PSD) of the solid raw  
203 materials (TNF and DWS) are shown in Table 1. As expected, the TNF fibre and  
204 fat contents were much higher than those of DWS, while the protein content was  
205 much lower. On the other hand, the amount of digestible carbohydrates in tiger  
206 nut flour was lower. All this may affect the various formulations obtained after the  
207 partial substitution of DWS by TNF, meaning that pasta with different nutritional

208 properties were attained. As commented on the material and methods' section,  
209 tiger nut pasta may be labelled as a "high fibre" product taking into account the  
210 obtained proximate composition for TNF and DWS. The particle size distribution  
211 of the solids (DWS and TNF) provides information about their water absorption  
212 capacity (velocity) and uniformity (dissolution into solid systems or suspensions).  
213 Particle size is a factor that will therefore affect the behaviour of the formulations  
214 during their processing and the uniformity of the final product. In water (DWS<sub>h</sub>  
215 and TNF<sub>h</sub> in Table 1), the results correspond with the particle size distribution of  
216 the insoluble fibre. It can be observed that the average particle size (expressed  
217 as the mean diameter of the equivalent volume) was lower for TNF (203 µm  
218 versus 319 µm in DWS) in the dry analysis, but significantly higher in the wet  
219 analysis (253 µm versus 150 µm). Furthermore, the uniformity in the particle size  
220 distribution (span) was higher in dehydrated semolina, while the opposite  
221 occurred in the wet analysis. The greater presence of fibre in tiger nut flour  
222 (15.85% compared to 10% in wheat semolina) -with a much higher insoluble /  
223 soluble ratio- (13.74/2.10% in TNF compared to 5.25/4.75% in DWS) could be  
224 responsible for the greater heterogeneity in TNF (span 2.28). The capacity to  
225 absorb and bind water by this fibre (as shown in section 3.1.3, the presence of  
226 TNF supposes a higher WHC index) causes –therefore- an increase in the mean  
227 particle size (wet analysis), with 10% of particles under 667 µm (d (0.9), Table 3)  
228 and having a much lower span (4.8). The amount of starch granules and their  
229 morphology also influences water absorption and, therefore, the particle size  
230 obtained by wet analysis. The starch granules of tiger nut flour are rounded,  
231 smaller than those of wheat, with greater solubility (Fig. 1; Abo-El-Fetoh et al.  
232 2010).

233 Table 2 shows the results obtained after analysing the functional properties of

234 DWS and the mixtures of wheat semolina and TNF. Water holding capacity  
235 (WHC) reports the ability of a protein matrix to absorb and retain bound,  
236 hydrodynamic, capillary and physically entrapped water against gravity. The  
237 results show a significant increase ( $p < 0.05$ ) when increasing the percentage of  
238 TNF. This increase can be attributed to higher fibre content, by increasing the  
239 percentage of tiger nut in the mix (Ade-Omowaye et al. 2008), fibre with high  
240 water retention capacity due to the high proportion of hemicellulose and lignin  
241 (Sanchez -Zapata et al. 2009). In addition, as discussed above, the smaller  
242 particle size of tiger nut flour implies a larger surface area available for water  
243 absorption (Albors et al. 2016). The solvent retention capacity reports the water  
244 retention capacity of pentosans (SRCS), damaged starch (SRCB) and glutenin  
245 (SRCAL). The values obtained for SRCS were not significantly different, but a  
246 decrease in this parameter can be observed when increasing the percentage of  
247 TNF. As for SRCB, a significant increase ( $p < 0.05$ ) was registered with the rise in  
248 the percentage of TNF in the mixture, which could be due to a greater presence  
249 of damaged starch in the tiger nut flour. The same trend was observed between  
250 SRCB and WHC, with a positive correlation between both variables ( $r = 0.944$ ).  
251 Finally, the value of SRCAL decreased when adding TNF (20% w/w), yet without  
252 significant differences ( $p < 0.05$ ) when the amount increased further. This  
253 decrease is due to the lower presence of gluten and, therefore, of glutenin when  
254 using less amount of wheat in the sample. The fat absorption capacity (FAC) is  
255 attributed to the ability of proteins to bind lipids. The greater presence of fat in  
256 TNF could be responsible for the significant differences found between DWS and  
257 the other analysed samples. Finally, there was a clear decrease in both foam  
258 capacity and foam stability with the substitution of wheat semolina by tiger nut  
259 flour. The presence of xanthan gum only significantly increased the WHC value,

260 due to its high water binding capacity.

### 261 3.2 Water content and water activity of fresh pasta

262 The water content and water activity values obtained for each of the resulting  
263 dough formulations are shown in Table 3. The analysis of variance (ANOVA)  
264 showed significantly higher values ( $p < 0.05$ ) for the 100% wheat semolina  
265 formulation. This seems logical if we consider that the water content of wheat  
266 semolina (13.67%) is higher than that of tiger nut flour (8.83%). The presence of  
267 xanthan gum did not affect the water activity of the doughs.

### 268 3.3 Rheological assessment: fundamental and empirical properties

#### 269 3.3.1 Dynamic oscillatory properties: frequency sweep and creep tests

270 Table 3 shows that the storage modulus values ( $G'$ ) are significantly higher ( $p$   
271  $< 0.05$ ) -in all cases- than those of the loss modulus ( $G''$ ) ( $\tan \delta < 1$ ), which reveals  
272 the more elastic and less viscous nature (greater solid nature) of the studied  
273 doughs. Similar results were obtained previously using various hydrocolloids  
274 (Lazaridou et al. 2007) and modified starches (Witczak et al. 2012). It is also  
275 possible to see that the partial substitution of wheat semolina by TNF resulted -  
276 as expected- in a significant decrease in the  $G'$  and  $G''$  values, exposing the  
277 formation of a less cohesive structure in these cases, due to the lower presence  
278 of gluten. Similar results were observed in pasta made with rice (Sozer 2009)  
279 and maize (Larrosa et al. 2013), where the values of the loss modulus ( $G''$ ) were  
280 between 5 and 10 times lower than the values of the storage modulus ( $G'$ ), and  
281 as the frequency increased  $G''$  was lower, showing a system that acted more  
282 elastically (Sozer 2009). This elastic behaviour is characteristic of a highly  
283 structured material, where the storage modulus is always greater than the loss  
284 modulus over the entire frequency range studied (Larrosa et al. 2013). As for the

285 hydrocolloid xanthan gum, it can be observed that the DWSX formulation has the  
286 highest  $G'$ , significantly higher ( $p < 0.05$ ) than the DWS control formulation,  
287 reflecting the clear influence of this gum. This hydrocolloid, in the tested  
288 concentration, therefore appears to influence the rheological behaviour of the  
289 control sample and that of TNF40, bringing the values of the latter closer to those  
290 obtained by the 20%. Only for TNF20 no significant differences ( $p > 0.05$ ) were  
291 found with the use of X. The dependence of both moduli ( $G'$  and  $G''$ ) on the  
292 angular frequency ( $\omega$ ) can be explained by the parameters (a and b exponents)  
293 obtained by equations 1 and 2 (Georgopoulos et al. 2004; Sivaramakrishnan et  
294 al. 2004) (Table 4).

$$295 \quad G'(\omega) = K' \cdot \omega^a \quad \text{Eq. (1)}$$

$$296 \quad G''(\omega) = K'' \cdot \omega^b \quad \text{Eq. (2)}$$

297 The presence of xanthan gum caused a significant decrease in parameter a,  
298 which shows a greater independence of  $G'$  and, therefore, a more cohesive  
299 structure. However, contrary to expectations, there was no greater dependence  
300 of  $G'$  when substituting the wheat semolina by TNF. Possibly, the greater  
301 presence of fat in these formulations "mitigates" the lower structure cohesion. As  
302 for the loss modulus ( $G''$ ), it is possible to see again how the values are clearly  
303 lower than those of the storage modulus ( $G'$ ) in the whole spectrum of  
304 frequencies. In addition, the values of exponent b are close to 0 in all cases,  
305 which demonstrates the gel-type structure (semi-solid nature) of the studied  
306 doughs.

307 In creep tests, a constant stress is applied and the sample is rapidly deformed,  
308 inflicting a deformation on the material that increases continuously at a  
309 decreasing rate over time (Rao and Skinner 1986). Fig. 2 shows an example of  
310 the creep curves of the control dough (DWS) and samples with the addition of

311 tiger nut flour and xanthan gum (DWSX, TNF20, TNF20X, TNF40 and TNF40X).  
 312 It can be observed that all the curves obtained showed a behaviour that is typical  
 313 of viscoelastic materials. The creep phenomenon is caused by a reorientation of  
 314 the bonds in this type of materials (Mariusz et al. 2012). It has been observed  
 315 that the viscoelastic properties of different wheat (Edwards et al. 2001) and rice  
 316 doughs (Sozer 2009) analysed in trials of this kind can be adequately described  
 317 with the Burger's model (Burgers 1935), which corresponds with the serial  
 318 combination of the Maxwell and Kelvin models. The experimental data from  
 319 creep tests were fitted by the 8-parameter Burger's model (equation 3), where  $J_0$   
 320 is the instantaneous creep compliance,  $J_1$ ,  $J_2$  y  $J_3$  are the retardation creep  
 321 compliances (the sum of  $J_0$ ,  $J_1$ ,  $J_2$  and  $J_3$  is the steady state compliance),  $\lambda_1$ ,  $\lambda_2$   
 322 and  $\lambda_3$  are the retardation times of the elastic response (Kelvin component),  $\mu_0$   
 323 provides information about the steady-state viscosity and  $t$  shows time (Van  
 324 Bockstaele et al. 2011).

$$325 \quad J(t) = J_0 + J_1 \left(1 - e^{-\frac{t}{\lambda_1}}\right) + J_2 \left(1 - e^{-\frac{t}{\lambda_2}}\right) + J_3 \left(1 - e^{-\frac{t}{\lambda_3}}\right) + \frac{t}{\mu_0} \quad (\text{eq.3})$$

326 Instantaneous compliance is related to the energy of elastic stretching of the  
 327 bonds when stress is applied, while retardation compliances are related to the  
 328 disruption and conversion of the bonds (Witczak et al. 2012). Table 5  
 329 summarises the values obtained for the model's coefficients. It is possible to  
 330 observe that when comparing separately the doughs without or with xanthan  
 331 gum, the addition of TNF caused an increase of instantaneous compliance  
 332 values respect to control samples (83.8-105.4% higher in TNF20 and TNF40,  
 333 respectively, and 55.6% higher in the samples with X). This is consistent as the  
 334 instantaneous compliance is greater in materials with weak structures, whereas  
 335 small values are characteristic of strong (solid) or hard structures (Sozer 2009).



336 The incorporation of xanthan gum in the formulation increases the resistance of  
337 the dough to deformation, which is shown in Fig. 2 by the reduction of the  
338 maximum percentage of creep deformation (deformation at the end of the creep  
339 phase) (Lazaridou et al. 2007). This hydrocolloid, once hydrated, may fill up free  
340 space in the system, which strengthens dough structure. The elastic retardation  
341 or viscoelastic compliances ( $J_1$ ,  $J_2$  and  $J_3$ ) increase with the tiger nut substitution  
342 percentage, which means that the greater the presence of tiger nut flour, the less  
343 the deformation resistance of the doughs. As for the retardation time ( $\lambda_1$ ) and  
344 steady-state viscosity ( $\mu_0$ ), no significant differences were observed between the  
345 different formulations except for the TX formulation, which is also the one with  
346 the highest value. Longer retardation times ( $\lambda_1$ ,  $\lambda_2$  and  $\lambda_3$ ) indicate a slower  
347 retarded elastic response. Therefore, the TX sample was the one with the  
348 greatest solid nature (the furthest one down), where the energy absorbed by the  
349 material -due to the deformation suffered- is dissipated to a lesser extent.  
350 Lazaridou et al. (2007) found that, in general,  $\lambda_1$  values were higher in gluten-  
351 free doughs with hydrocolloids.

### 352 3.3.2 Texture profile analysis (TPA)

353 The results in Table 6 show a significant decrease ( $p < 0.05$ ) in the hardness of  
354 the doughs as the percentage of TNF increased in the formulation, with a much  
355 more pronounced decrease between the control formulation and that with a 20%  
356 substitution level. It was also observed that by incorporating the hydrocolloid  
357 xanthan gum in the formulation, the decrease in hardness was less pronounced  
358 (27.67% versus 52.27% when using 20% of TNF), reflecting the clear influence  
359 of this hydrocolloid in the analysed parameter, since it reinforces the structure by  
360 increasing the viscosity and imitating viscoelastic gluten properties. Concerning  
361 adhesiveness, the most adhesive formulation was TNF20 and the least adhesive

362 was TNF40X. It can also be seen how the partial substitution of durum wheat  
363 semolina and incorporation of xanthan gum influence this parameter -although  
364 without showing a visible trend, contrary to what occurs with the hardness. The  
365 same is true for the cohesiveness. Finally, the elasticity parameter shows that  
366 the most elastic formulation is TNF40X and the least elastic is T.

### 367 3.4 Thermal properties: DSC results

368 It is known that DSC thermal analysis allows studying the gelatinisation,  
369 retrogradation and crystallisation phenomena of starch polymers. It is possible to  
370 measure the enthalpies (heat fluxes) associated with the amylose and  
371 amylopectin fusion (with melting temperatures between 60-100 °C, depending on  
372 the type of starch (Cano et al. 2015)) as well as the dissociation and fusion  
373 enthalpy of the amylose-lipid complexes. When the starch is heated in excess of  
374 water (water/starch ratio of 2:1) in the DSC analysis, an endothermic peak is  
375 obtained. The peak's onset (where it deviates from the baseline,  $T_o$ , °C)  
376 corresponds with the starch granule's birefringence loss onset. The area below  
377 the peak is a measurement of the energy (enthalpy,  $\Delta H$ ) required for the  
378 transition from an orderly (crystalline) state to a disordered or amorphous state  
379 (fusion) to take place (Biliaderis 2009). The temperature range between the  
380 onset and the end of the peak ( $T_c - T_o$ ) represents the so-called gelatinisation  
381 period, and the peak temperature ( $T_p$ ) is the gelatinisation temperature. This  
382 gelatinisation in excess water is believed to involve the primary hydration of the  
383 amorphous regions around and above the glass transition temperature, with an  
384 associated glassy-gum transition. This in turn facilitates molecular mobility in  
385 amorphous regions (with reversible swelling), which causes an irreversible  
386 molecular transition. This irreversible step involves the dissociation of double  
387 helices (most of which are in crystalline regions) and the expansion of the

388 granules and polymers (and interstices of the granule). After  $T_c$ , all the  
389 amylopectin double helices have dissociated, although the swollen granular  
390 structures have remained (Tester and Debón 2000). Table 7 summarises the  
391 values obtained for the thermal properties of the fresh pasta formulations. All the  
392 samples, except for those in which wheat semolina was replaced by 40% tiger  
393 nut flour (TNF40 and TNF40X), had a single endothermic peak in a temperature  
394 range between 61 and 78 °C during the heating process, which corresponds with  
395 the gelatinisation of the amylopectin. However, for the TNF40 and TNF40X  
396 samples, two phase transitions were observed. This could be due to the different  
397 gelatinisation temperature of the tiger nut starch -which may gelatinise at higher  
398 temperatures-, or to the fusion of amylose-lipid complexes, due to the  
399 incorporation of lipid-rich tiger nut flour. It seems therefore that the incorporation  
400 of TNF -from a certain value of substitution onwards (40% in the case of this  
401 study) produces the emergence of these complexes, resulting in a dough of fresh  
402 pasta with 7.62% fat content and a fat/starch ratio of 0.2).. Consequently, there is  
403 a smaller amount of starch available to gelatinise and, therefore, a lower amount  
404 of energy is needed for such transition to occur. In these formulations, no  
405 significant differences ( $p < 0.05$ ) were found in terms of the gelatinisation  
406 temperature ( $T_p$  value), either with the incorporation of xanthan gum or with 20%  
407 of tiger nut flour.

408 When performing the DSC thermal analysis on the samples cooked at different  
409 times, it can be observed that as the cooking time increased, the gelatinisation  
410 temperature increased as well, while the size of the endotherm area decreased  
411 until a point when no transition was observed (completely gelatinised starch).  
412 The fraction of starch which remained ungelatinised was calculated -at each  
413 cooking time- on the basis of the area involved in the phase transition

414 ( $\Delta H/\Delta H_0, \%$ ) and taking as a reference the corresponding value of the uncooked  
415 pasta sample ( $\Delta H_0, \text{J/g}$ ). The results obtained are summarised in Table 8. In  
416 short, the optimal cooking time for the control pasta (DWS formulation) was four  
417 minutes. For the rest of the formulations tested, the absence of transition was  
418 already observed at two minutes, which may signal a greater ease of hydration of  
419 the starch granules during cooking, as a result of the lower cohesiveness of the  
420 dough's structure and therefore the greater accessibility of the water molecules  
421 to the amorphous regions of the granule.

#### 422 **4. Conclusions**

423 Basing on the obtained results it could be concluded that the presence of TNF  
424 and X significantly modifies rheological properties of fresh pasta dough. Changes  
425 included: (i) lower values of storage, loss moduli and hardness, and higher  
426 values of instantaneous and retardation compliances with TNF addition,  
427 revealing the formation of a less cohesive structure with a lower resistance to  
428 deformation (less rigid) due to the lower presence of a gluten network. (ii) a  
429 greater independence of storage modulus on frequency, a less pronounced  
430 hardness and a reduction of the deformation at the end of the creep test when  
431 using xanthan gum at 1% (DWS basis), showing the structure strengthen of the  
432 doughs. The thermal analysis showed a single endothermic peak in all the  
433 samples -in a temperature range between 60 and 78 °C -, corresponding with the  
434 gelatinisation of amylopectin. However, in the samples with a higher percentage  
435 of tiger nut flour (TNF40 and TNF40X), two phase transitions were observed,  
436 associated either to the gelatinisation of the tiger nut starch or the emergence of  
437 amylose-lipid complexes. As the cooking time increased, the gelatinisation  
438 temperature increased as well and the size of the endotherm area decreased,  
439 showing an optimal cooking time of four minutes for the control pasta (DWS),

440 while for the rest of the formulations the optimal time was found to be two  
441 minutes. This indicates a greater ease of hydration of the starch granules during  
442 cooking, as a result of the lower cohesiveness of the dough's structure and  
443 therefore the greater accessibility of the water molecules to the amorphous  
444 regions of the granule.

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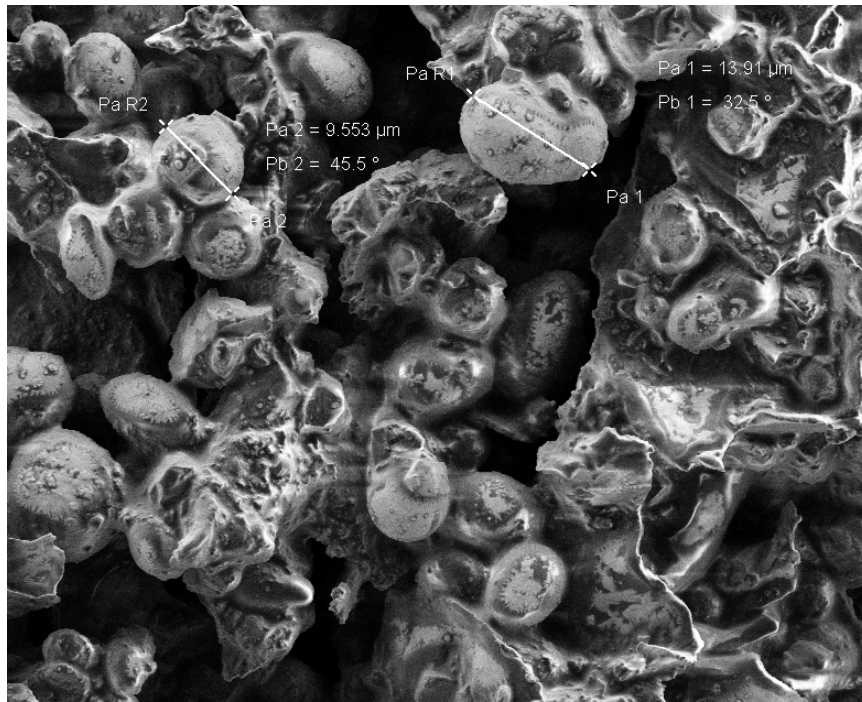
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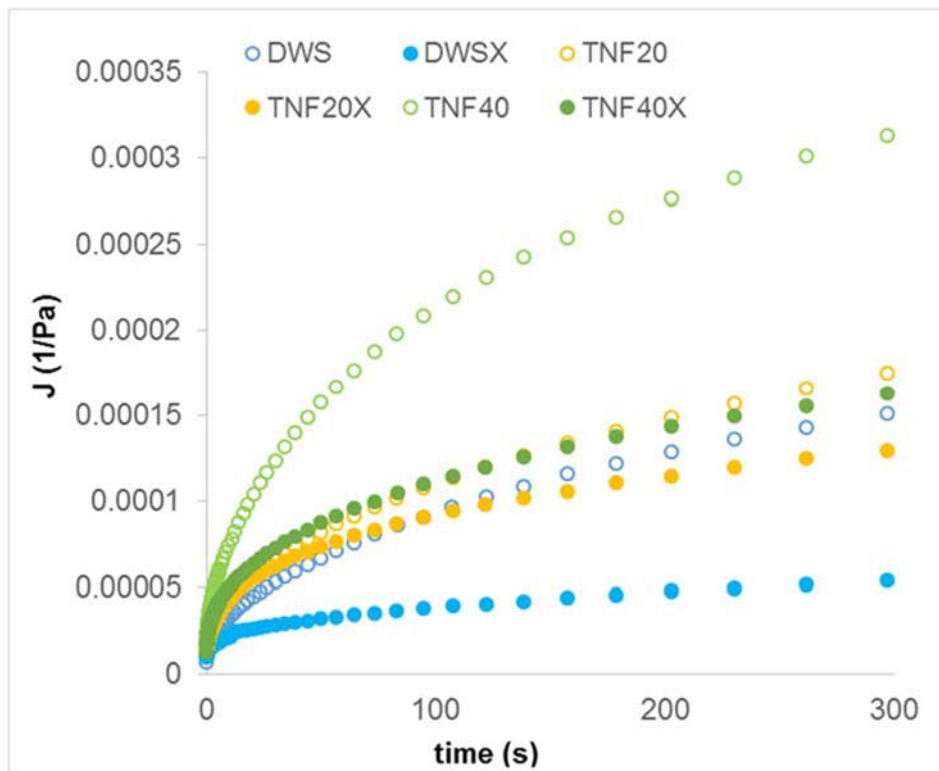


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**Fig. 1.** Scanning electron micrograph of cross section of tiger nut flour (1000x)



**Fig. 2.** Effect of tiger nut flour and xanthan gum incorporation on creep test curves of fresh pasta doughs.

**Table 1.** Proximate chemical composition (PCC) and particle size distribution (PSD) of durum wheat semolina (DWS) and tiger nut flour (TNF) (g/100 g). Mean values of three replicates (standard deviation).

		<b>DWS</b>	<b>TNF</b>	<b>DWS<sub>h</sub></b>	<b>TNF<sub>h</sub></b>
<b>PCC</b>	<b>Water</b>	14.03(0.03) <sup>a</sup>	8.03(0.02) <sup>b</sup>		
	<b>Protein</b>	13.81(0.40) <sup>a</sup>	4.80(0.05) <sup>b</sup>		
	<b>Fat</b>	0.92(0.06) <sup>b</sup>	25.30(0.02) <sup>a</sup>		
	<b>Ash</b>	1.67(0.07) <sup>b</sup>	2.02(0.04) <sup>a</sup>		
	<b>Dietary Fibre</b>			-	-
	<b>-Soluble</b>	3.75(0.02) <sup>a</sup>	2.05(0.03) <sup>b</sup>		
	<b>-Insoluble</b>	4.75(0.02) <sup>b</sup>	13.80(0.03) <sup>a</sup>		
	<b>-Total</b>	9.50(0.02) <sup>b</sup>	15.85(0.03) <sup>a</sup>		
	<b>DC<sup>a</sup></b>	60.08(0.02) <sup>a</sup>	44.00(0.03) <sup>b</sup>		
<b>PSD<sup>b</sup></b>	<b>D[4, 3] (µm)</b>	319(3) <sup>a</sup>	203(4) <sup>c</sup>	150(12) <sup>d</sup>	253(26) <sup>b</sup>
	<b>d<sub>(0.1)</sub> (µm)</b>	133(4) <sup>a</sup>	50(1) <sup>b</sup>	8.5(0.3) <sup>d</sup>	11.6(0.8) <sup>c</sup>
	<b>d<sub>(0.5)</sub> (µm)</b>	305(3) <sup>a</sup>	162(4) <sup>b</sup>	50(4) <sup>d</sup>	135(12) <sup>c</sup>
	<b>d<sub>(0.9)</sub> (µm)</b>	530(4) <sup>b</sup>	419(4) <sup>d</sup>	457(33) <sup>c</sup>	667(68) <sup>a</sup>
	<b>Span</b>	1.30(0.02) <sup>d</sup>	2.28(0.06) <sup>c</sup>	9.00(0.30) <sup>a</sup>	4.81(0.20) <sup>b</sup>

Within rows, values with the same following letter do not differ significantly from each other ( $p > 0.05$ )

<sup>a</sup>Digestible carbohydrates calculated by difference.

<sup>b</sup>D[3,2] and D[4,3], d<sub>(0.1)</sub>, d<sub>(0.5)</sub>, d<sub>(0.9)</sub>, represent, respectively, Sauter mean diameter, mean particle diameter, 10%, 50% and 90% of all particles finer than this size.

**Table 2.** Functional properties of durum wheat semolina (DWS), tiger nut flour (TNF) and DWS/TNF blends (TNF20: 80/20, TNF40: 60/40, w/w semolina basis) with or without xanthan gum (X, 1% semolina basis). Mean values of three replicates (standard deviation).

<b>Sample</b>	<b>SRC<sup>S</sup> (%)</b>	<b>SRC<sup>B</sup> (%)</b>	<b>SRC<sup>AL</sup> (%)</b>	<b>WHC (g/g semolina)</b>	<b>FAC (g/g)</b>	<b>FC (mL)</b>	<b>FS (%)</b>
<b>DWS</b>	93(10) <sup>a</sup>	64.02(0.98) <sup>c</sup>	108(4) <sup>a</sup>	0.925(0.007) <sup>f</sup>	1.210(0.202) <sup>c</sup>	12.5(0.2) <sup>a</sup>	74.0(3.0) <sup>a</sup>
<b>TNF</b>	91(8) <sup>a</sup>	99(9) <sup>c</sup>	-	2.050(0.191) <sup>a</sup>	1.749(0.104) <sup>a</sup>	-	-
<b>TNF20</b>	87(5) <sup>a</sup>	88(6) <sup>b</sup>	83(11) <sup>b</sup>	1.094(0.011) <sup>e</sup>	1.300(0.201) <sup>bc</sup>	1.6(0.1) <sup>b</sup>	16.0(0.2) <sup>b</sup>
<b>TNF40</b>	83(2) <sup>a</sup>	129(9) <sup>a</sup>	79(5) <sup>b</sup>	1.125(0.005) <sup>d</sup>	1.560(0.090) <sup>b</sup>	1.5(0.1) <sup>b</sup>	17.0(0.2) <sup>b</sup>
<b>TNF20X</b>	80(12) <sup>a</sup>	89(7) <sup>b</sup>	96(3) <sup>b</sup>	1.270(0.060) <sup>c</sup>	1.490(0.040) <sup>b</sup>	0.7(0.3) <sup>c</sup>	15.6(0.2) <sup>b</sup>
<b>TNF40X</b>	89(7) <sup>a</sup>	124(2) <sup>a</sup>	82(2) <sup>b</sup>	1.360(0.050) <sup>b</sup>	1.600(0.151) <sup>b</sup>	0.8(0.3) <sup>c</sup>	17.3(0.2) <sup>b</sup>

Within columns, values with the same following letter do not differ significantly from each other ( $p > 0.05$ )

SRC<sup>S</sup>: water retention capacity by pentosans; SRC<sup>B</sup>: water retention capacity by damaged starch; SRC<sup>AL</sup>: water retention capacity by glutenin; WHC: water holding capacity; FAC: fat adsorption capacity; FC: foam capacity; FS: foam stability

**Table 4.** Mean values (standard deviation) of the obtained parameters for the fitted equation for elastic- $G'$ - and viscous- $G''$ - moduli as affected by frequency ( $\omega$ )

Sample	$G'(\omega) = K' \cdot \omega^a$		$G''(\omega) = K'' \cdot \omega^b$	
	K' [Pa s <sup>n</sup> ]	a	K'' [Pa s <sup>n</sup> ]	b
<b>DWS</b>	124049(20712) <sup>b</sup>	0.234(0.009) <sup>a</sup>	32552(2770) <sup>a</sup>	0.14(0.02) <sup>bc</sup>
<b>DWSX</b>	175362(4708) <sup>a</sup>	0.205(0.003) <sup>cd</sup>	33139(523) <sup>a</sup>	0.123(0.010) <sup>c</sup>
<b>TNF20</b>	81494(6736) <sup>c</sup>	0.226(0.007) <sup>ab</sup>	23255(1576) <sup>b</sup>	0.130(0.004) <sup>c</sup>
<b>TNF20X</b>	84365(4895) <sup>c</sup>	0.214(0.009) <sup>c</sup>	21792(1706) <sup>bc</sup>	0.157(0.009) <sup>ab</sup>
<b>TNF40</b>	47246(8849) <sup>d</sup>	0.221(0.006) <sup>b</sup>	14839(1979) <sup>d</sup>	0.161(0.018) <sup>a</sup>
<b>TNF40X</b>	82507(4258) <sup>c</sup>	0.199(0.002) <sup>d</sup>	20266(1028) <sup>c</sup>	0.141(0.007) <sup>abc</sup>

Within columns, values with the same following letter do not differ significantly from each other (p > 0.05)



**Table 5.** Parameters of Burgers model for creep data. Mean values of three replicates (standard deviation)

<b>Sample</b>	<b><math>J_0 \cdot 10^{-6}</math> (Pa<sup>-1</sup>)</b>	<b><math>\mu_0 \cdot 10^6</math> (Pa·s)</b>	<b><math>J_1 \cdot 10^{-5}</math> (Pa<sup>-1</sup>)</b>	<b><math>\lambda_1</math> (s)</b>	<b><math>J_2 \cdot 10^{-6}</math> (Pa<sup>-1</sup>)</b>	<b><math>\lambda_2</math> (s)</b>	<b><math>J_3 \cdot 10^{-6}</math> (Pa<sup>-1</sup>)</b>	<b><math>\lambda_3</math> (s)</b>	<b>R<sup>2</sup></b>
DWS	9(1) <sup>d</sup>	8(2) <sup>b</sup>	4(3) <sup>c</sup>	88(19) <sup>b</sup>	14(1) <sup>c</sup>	10.0(1.2) <sup>ab</sup>	6(8) <sup>bc</sup>	0.94(0.10) <sup>b</sup>	0.9979
DWSX	9(4) <sup>cd</sup>	40(30) <sup>a</sup>	2(1) <sup>c</sup>	233(86) <sup>a</sup>	8(5) <sup>d</sup>	11.9(1.9) <sup>a</sup>	4(2) <sup>c</sup>	1.11(0.07) <sup>a</sup>	0.9989
TNF20	17(3) <sup>ab</sup>	4(26) <sup>b</sup>	9(3) <sup>ab</sup>	113(23) <sup>b</sup>	19(2) <sup>b</sup>	10.3(1.4) <sup>ab</sup>	8(2) <sup>ab</sup>	0.94(0.07) <sup>abc</sup>	0.9980
TNF20X	14(3) <sup>bc</sup>	11(4) <sup>b</sup>	3(1) <sup>c</sup>	70(4) <sup>b</sup>	15(2) <sup>c</sup>	8.5(1.5) <sup>bc</sup>	8(1) <sup>ab</sup>	0.79(0.07) <sup>cd</sup>	0.9956
TNF40	19(2) <sup>a</sup>	3(7) <sup>b</sup>	13(4) <sup>a</sup>	66.2(0.5) <sup>b</sup>	27(1) <sup>a</sup>	6.5(1.2) <sup>c</sup>	10(4) <sup>a</sup>	0.64(0.14) <sup>d</sup>	0.9960
TNF40X	14(2) <sup>bc</sup>	7(1) <sup>b</sup>	5(1) <sup>bc</sup>	79(2) <sup>b</sup>	16(4) <sup>bc</sup>	8.3(0.3) <sup>bc</sup>	9(1) <sup>ab</sup>	0.83(0.03) <sup>bc</sup>	0.9964

Within columns, values with the same following letter do not differ significantly from each other (p > 0.05)

**Table 6.** TPA results. Mean values of three replicates (standard deviation).

<b>Sample</b>	<b>Hardness (N)</b>	<b>Adhesiveness (N·s)</b>	<b>Cohesiveness</b>	<b>Springiness</b>
<b>DWS</b>	572(18) <sup>a</sup>	-4086(276) <sup>c</sup>	0.53(0.01) <sup>c</sup>	0.55(0.02) <sup>d</sup>
<b>DWSX</b>	553(63) <sup>a</sup>	-1303(637) <sup>a</sup>	0.76(0.04) <sup>a</sup>	0.73(0.09) <sup>bc</sup>
<b>TNF20</b>	273(8) <sup>c</sup>	-2850(278) <sup>b</sup>	0.62(0.03) <sup>b</sup>	0.61(0.05) <sup>cd</sup>
<b>TNF20X</b>	400(21) <sup>b</sup>	-6924(951) <sup>d</sup>	0.65(0.06) <sup>b</sup>	0.82(0.14) <sup>b</sup>
<b>TNF40</b>	220(12) <sup>d</sup>	-6273(244) <sup>d</sup>	0.46(0.02) <sup>c</sup>	0.66(0.07) <sup>cd</sup>
<b>TNF40X</b>	231(3) <sup>cd</sup>	-10505(369) <sup>e</sup>	0.68(0.07) <sup>b</sup>	0.97(0.01) <sup>a</sup>

Within columns. values with the same following letter do not differ significantly from each other (p > 0.05)

**Table 7.** Thermal properties of doughs (gelatinisation enthalpy ( $\Delta H_n$ ), gelatinisation range ( $T_o$ - $T_e$ ), and onset ( $T_o$ ), peak ( $T_p$ ) and conclusion ( $T_e$ ) temperatures.

	<b>Sample</b>	$\Delta H_n$ (J/g)	$T_o$ (°C)	$T_p$ (°C)	$T_e$ (°C)	$T_o$ - $T_e$ (°C)
<b>A</b>	<b>DWS</b>	1.11(0.21) <sup>a</sup>	54.6(0.9) <sup>c</sup>	61.1(0.1) <sup>c</sup>	66.8(0.5) <sup>b</sup>	12.2(1.3) <sup>a</sup>
	<b>DWSX</b>	0.88(0.12) <sup>a</sup>	55.0(0.2) <sup>c</sup>	61.2(0.4) <sup>c</sup>	67.1(0.4) <sup>b</sup>	12.1(0.1) <sup>a</sup>
	<b>TNF20</b>	1.02(0.06) <sup>a</sup>	55.6(0.3) <sup>bc</sup>	62.4(0.3) <sup>c</sup>	68.0(0.2) <sup>b</sup>	12.4(0.2) <sup>a</sup>
	<b>TNF20X</b>	0.96(0.12) <sup>a</sup>	55.3(0.3) <sup>c</sup>	62.1(0.3) <sup>c</sup>	67.9(0.2) <sup>b</sup>	12.6(0.3) <sup>a</sup>
	<b>TNF40</b>	0.72(0.08) <sup>b</sup>	57.3(0.2) <sup>bc</sup>	66.0(3.0) <sup>bc</sup>	68.9(0.2) <sup>b</sup>	11.6(0.4) <sup>a</sup>
	<b>TNF40X</b>	0.84(0.13) <sup>ab</sup>	57.1(0.5) <sup>ab</sup>	64.0(1.0) <sup>ab</sup>	69.5(0.8) <sup>a</sup>	12.4(0.3) <sup>a</sup>
<b>B</b>	<b>TNF40</b>	0.33(0.02)	74.1(0.3)	78.1(0.2)	82.7(0.1)	8.6(0.2) <sup>b</sup>
	<b>TNF40X</b>	0.30(0.03)	74.55(1.12)	78.6(1.0)	83.1(1.2)	8.5(0.1) <sup>b</sup>

Within columns, values with the same following letter do not differ significantly from each other ( $p > 0.05$ ). **A:** wheat amylopectin gelatinisation; **B:** associated transition to amylose-lipid complexes or tiger nut starch gelatinisation

**Table 8.** Gelatinisation temperature ( $T_p$ ) and percentage of non gelatinised starch (NGS) at different cooking times

	Time (min)	$T_p$ (°C)				NGS (%)			
		t=0	t=2	t=3	t=4	t=0	t=2	t=3	t=4
<b>Sample</b>	<b>DWS</b>	61.1(0.1)	71.0(1.2)	74.5(0.1)	-	100	15	6	0
	<b>DWSX</b>	61.2(0.4)	-	-	-	100	0	0	0
	<b>TNF20</b>	62.4(0.3)	-	-	-	100	0	0	0
	<b>TNF20X</b>	62.1(0.3)	-	-	-	100	0	0	0
	<b>TNF40</b>	66.0(3.0)	-	-	-	100	0	0	0
	<b>TNF40X</b>	64.0(1.0)	-	-	-	100	0	0	0

Within columns, values with the same following letter do not differ significantly from each other ( $p > 0.05$ )