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Additional Information

# Biocleaning of Cultural Heritage stone surfaces and frescoes: which delivery system can be the most appropriate?

Pilar Bosch-Roig · Giuseppe Lustrato ·  
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**Abstract** The use of the advanced biotechnology of micro-biological systems for the biological cleaning of Cultural Heritage (CH) has been recently improved and optimized taking into account different factors. Biocleaning systems have been indeed applied to historic buildings, statues and frescoes. Such application has developed new techniques and optimised and refined the existing systems. These systems remove altered forms like sulfate and nitrate crusts and organic substances like animal glue in a more effective, less invasive way than the traditional cleaning techniques. This review focuses on several delivery systems (sepiolite, hydrobiogel-97, cotton wool, carbogel, mortar and alginate beads, agar, and arbocel) used for the biocleaning of Cultural Heritage, comparing their main properties and characteristics, making a critical evaluation on how easy they can be applied, and on their future potentiality as ready-to-use and risk-free formulations. Therefore, this review will help conservation scientists, conservator-restorers, and researchers in the field to choose the most appropriate delivery system for any specific applications.

**Keywords** Biocleaning · Cultural Heritage stone surfaces ·  
Frescoes · Delivery systems · Microorganisms

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## Introduction

35

Amongst the damage found on works of art in both outdoor and indoor environments, those relative to lithoid-type matrices (stone, frescoes) are well documented. Both natural and artificial (obtained from transformation) stones have been widely studied because they are the key element in terms of archaeological and monumental heritage (Pinna and Salvadori 2008). Physical, chemical, and biological agents act to deteriorate stone (Warscheid and Braams 2000). Deterioration of stones depends largely on their physical and chemical properties such as strength, porosity, hardness, durability, texture, absorption, etc. Therefore, stones with high porosity, high rates of swelling, and low strength tend to be poor building materials (Doehne and Price 2010). Before starting to clean any type of stone surface, conservator-restorers and researchers must define the mineralogical properties of the stones involved, characterize the decay, measure its extent, severity, rate of progress and try to understand causes and mechanisms of that specific type of decay. Stone decay is a complex phenomenon produced by several factors that can have rapid or slow effects. There are many factors that can contribute to stone decay, but the main ones are air pollution, presence and concentration of salts, and biodeterioration (Camuffo 1998; Doehne and Price 2010).

Exposed stonework in polluted urban areas shows “black crust”, “sulfation” and “nitratation” decay mainly caused by atmospheric pollution. Sulphur oxides, nitrogen oxides (NO<sub>x</sub>), and carbon oxides present in the atmosphere create acidic solutions in the presence of water, which are capable of reacting with calcareous materials. Air pollutants can therefore cause the transformation of calcium carbonate in the calcareous matrix into calcium sulfate dihydrate and calcium nitrate with consequent loss of structure and increased susceptibility to corrosion phenomena (Gauri et al. 1989; Rivadeneyra et al. 1991, 1994; Oriol et al. 1993; Salvadori

70 and Realini 1996; Saiz-Jimenez 2004; Doehne and Price  
71 2010). These transformations cause deterioration processes  
72 of the structural, morphological, chemical, and aesthetic prop-  
73 erties of the original material and, by definition, lead to what  
74 are termed “decay” (Saiz-Jimenez 2003). The effect of acidic  
75 pollutants on stone depends on the environment where mon-  
76 uments are located. When stone surfaces are in an exposed  
77 location with frequent rainfall, products from the reaction with  
78 air pollutants are wetted, and the stone surface is progressively  
79 deteriorated. If instead, when the stone surface is in a sheltered  
80 location, products from reactions accumulate and can form  
81 black crusts on stone surfaces. The blackness of the crust is  
82 mainly caused by combustion of fossil fuels. The rate of air  
83 pollutant decay mostly depends on the level of pollution, the  
84 amount of rain, and its acidity (Doehne and Price 2010).  
85 Indirect effects of atmospheric pollution like climate change  
86 may also alter stone monuments in different ways, such as by  
87 increasing the biodeterioration of the stone due to higher tem-  
88 perature and rainfall (Duthie et al. 2008), or by increasing salt  
89 crystallization on the stone due to greater fluctuations in humidity  
90 (Brimblecombe and Grossi 2007; Grossi et al. 2008).

91 Salts can also cause serious damage to Cultural Heritage  
92 (CH) stone surfaces and frescoes due to the stresses generated  
93 by crystal growth in the pores in the stone. Crystallization  
94 pressure and, therefore, damage result when a growing crystal  
95 encounters a pore wall (Scherer 2000). Salt damage may occur  
96 in both indoor and outdoor environments, and salts can be  
97 present from various sources (air pollution, soil, wind from the  
98 sea or the desert, deicing salt, cleaning materials, garden  
99 fertilizers, etc.) (Doehne and Price 2010).

100 Biodeterioration of historical and artistic stone objects and  
101 frescoes is another important cause of stone decay (Dornieden  
102 et al. 2000; Saiz-Jimenez 2001). Different organisms (includ-  
103 ing bacteria, fungi, algae, and plants) can grow on CH  
104 stoneworks by using the mineral components and surface de-  
105 posits (Bhatnagar et al. 2010). Biological colonization can  
106 accelerate deterioration of the mineral matrix due to the met-  
107 abolic activity producing physical, chemical, and aesthetic  
108 damage (Warscheid and Braams 2000). The degree of the  
109 biodeterioration processes is related to the type of organisms  
110 present, the kind of material, and its state of conservation, as  
111 well as the environmental conditions where the works are  
112 located (indoors, outdoors, semi-confined areas), etc.  
113 (Ranalli et al. 2009). In the presence of biofilm extracellular  
114 polymeric substances (EPS) can result in mechanical stresses  
115 to the mineral structure leading to alterations in the size of the  
116 pores in the stone and in changes in the moisture circulation  
117 patterns and temperature response. In addition the presence of  
118 biofilms on stone surfaces can accelerate the accumulation of  
119 atmospheric pollutants and, therefore, act as a preliminary  
120 precursor for the formation of damaging crusts (Warscheid  
121 and Braams 2000). The best preventive method has proven to  
122 be the control of water, nutrients, and light levels around the

123 artwork. The use of biocides must only be considered as a last  
124 resort due to their toxicity (Blazquez et al. 2000; Martin-  
125 Sanchez et al. 2012; Gómez-Alarcón and Sáiz-Jiménez  
126 2013; Sanmartín et al. 2014). Other studies show that not all  
127 biological organisms produce stone decay, but biological pa-  
128 tinas in some situations may help to protect fragile stone  
129 surfaces (Caneva et al. 2005; De Muynck et al. 2010). Also,  
130 specific microorganisms, because of their metabolic activities,  
131 can be positively used to clean stone surfaces (Atlas et al.  
132 1988; Ranalli et al. 2003).

133 The conservation of stone surfaces and frescoes needs a  
134 technical and scientific approach. The problem of conserving  
135 CH is not simple, decay is a natural process, and, therefore, we  
136 can only slow it down (Fassina 2000). Stones can accumulate  
137 different kinds of materials: atmospheric deposits, salts from  
138 efflorescence phenomena, and residues from past CH inter-  
139 ventions. Deposits are due to different deposition mechanisms  
140 of materials from the environment; the main mechanism is  
141 inertia, so materials deposit as to their weight. On the contrary,  
142 black crusts are formed with an important contribution of  
143 material coming from the substrate. The main mechanism of  
144 black crust is sulphation reaction involving calcium carbonate.  
145 Therefore, black crusts and deposits are due to different mech-  
146 anisms. These materials present on the surface may present a  
147 significant hazard for CH damaging their aesthetic, chemical  
148 and physical aspects, and properties producing fissures, exfo-  
149 liation, disintegration, loss of original material, and so on. The  
150 removal of these deposits is, therefore, an important concern  
151 for conservator-restorers. Cleaning of CH is a complex issue  
152 and a critical part of conservation and attention must be paid to  
153 avoid altering the original surfaces during the treatments. To  
154 select the most appropriate cleaning method, the substrate  
155 characteristics and the chemical characterisation of the mate-  
156 rials to be removed should be firstly studied (Gulotta et al.  
157 2014). The basic evaluation criteria that the cleaning method-  
158 ology should take into account are: physical and chemical  
159 harmfulness, homogeneity of the deposits removal, efficiency,  
160 selective cleaning, absence of aesthetic alteration, and dura-  
161 bility (Vergès-Belmin 1996; Gulotta et al. 2014). The main  
162 cleaning methods usually utilized are mechanical (brushing  
163 and rubbing, washing and steaming, wet and dry  
164 abrasives, etc.) and chemical (alkaline treatments, acidic treat-  
165 ments or organic solvents, etc.) methods. Due to the risk of  
166 damage (like loss of original material) using some of these  
167 conventional cleaning techniques, most of the researches has  
168 been focused on alternative, more selective, and less aggres-  
169 sive techniques like laser and biological cleaning (Doehne and  
170 Price 2010). Laser cleaning allows high selectivity (lasers can  
171 discriminate between the soiling and the substrate) being a  
172 less intrusive, more easily controlled, method than the tradi-  
173 tional ones. Laser technique can be sometimes unsatisfactory  
174 because it does not allow the complete removal of the de-  
175 posits, and it can cause color changes. It can also cause

176 problems if applied to polychrome sculptures and the costs are  
177 significantly high when applied to large superficial areas  
178 (Salimbeni et al. 2003).

179 Biological methods that use microorganisms and enzymes  
180 as biological cleaning agents in the “biorestitution” of art-  
181 works are becoming attractive alternatives to the mechanical  
182 and chemical methods. They offer significant advantages in  
183 terms of soft intervention on the works themselves, lack of  
184 health risks for conservator-restorers, and also guaranteeing  
185 environmental safety (Saiz-Jimenez 1997; Cremonesi 2002;  
186 Ranalli et al. 2005; Valentini et al. 2010). Under optimal  
187 controlled conditions, biological methods reproduce the same  
188 processes that occur in nature (Boquet et al. 1973; Atlas et al.  
189 1988; Ferrer et al. 1988; Heselmeyer et al. 1991; Tiano et al.  
190 1996; Castanier et al. 2000; Maier et al. 2000; Rodriguez-  
191 Navarro et al. 2000; Zanardini et al. 2000; Ranalli et al. 2003;  
192 Biavati and Sorlini 2008).

193 The key idea of using living cells in the conservation and  
194 preservation of works of art is supported by the fact that  
195 microorganisms (mainly bacteria) are the most versatile and  
196 ubiquitous organisms found on earth, and they appear to be  
197 capable of colonizing almost any environment (Maier et al.  
198 2000). Even if we know that some microorganisms have a  
199 negative effect, many of them are responsible for “positive  
200 processes” such as the degradation of unwanted organic sub-  
201 stances (Sorlini et al. 2010a). Recently various Cultural Her-  
202 itage stone surfaces have been cleaned of organic and inor-  
203 ganic unwanted materials. Biocleaning techniques have been  
204 performed on stone (marble, tuff, sandstone, limestone, etc.),  
205 on ceramic material (brick-work), on paper materials, and on  
206 concrete using specific bioformulations containing  
207 *Desulfovibrio* sp. and *Pseudomonas* sp. cells. (Ranalli et al.  
208 2005; De Graef et al. 2005; De Belie et al. 2005; Cappitelli  
209 et al. 2006, 2007; Barbabietola et al. 2012). Until now, posi-  
210 tive results have been obtained from experiments conducted  
211 on significant historical monuments like the frescoes at  
212 Camposanto Monumentale, Pisa, Italy (removal of a cloth  
213 firmly glued to the painted layer) (Antonioli et al. 2005;  
214 Ranalli et al. 2005; Lustrato et al. 2012), Milan Cathedral  
215 facade (removal of black crusts) (Cappitelli et al. 2006,  
216 2007), and Matera Cathedral - both in Italy (removal of  
217 nitrates) (May et al. 2008; Alfano et al. 2011). Other positive  
218 results involve the colored lithotypes of Florence Cathedral  
219 (removal of black crust) (Gioventù et al. 2011), the frescoes on  
220 the Santos Juanes Church in Valencia, Spain (removal of  
221 animal glue residues and salt efflorescence) (Bosch-Bosch-  
222 Roig et al. 2013a, 2013b), and original paper specimens from  
223 the *Istituto Nazionale per la Grafica*, Rome (Removal of  
224 animal glue) (Barbabietola et al. 2012). Recently Gioventù  
225 et al. 2011 in a specific case study compared the biocleaning  
226 treatments on stone materials to chemical and laser  
227 treatments. They concluded that the most satisfactory cleaning  
228 treatment for sulfate removal was the biocleaning process. In

another recent study, combined chemical and biological 229  
methods allowed efficient cleaning with a notable reduction 230  
in the treatment duration (Troiano et al. 2013). 231

Although considerable studies have been devoted to 232  
biocleaning strategies, there is still some work to do to obtain 233  
ready-to-use products for the biocleaning of organic and inor- 234  
ganic residues from works of art. In order to introduce a ready- 235  
to-use biocleaning product to the restoration market, two main 236  
basic aspects must be taken into account: the microorganism 237  
itself and the delivery system to be used. 238

Microorganism biodiversity includes Bacteria, Archaea 239  
and Eukaria, which can live in every habitat of the biosphere 240  
(soil, rocks, hot springs, oceans, etc.), since they are extremely 241  
adaptable to environmental conditions. Microorganisms are 242  
extraordinarily diverse in their requirements for growth, and 243  
their growth is greatly affected by the nutrients that are avail- 244  
able in environment. However, they have common living 245  
requirements: energy (from light or from organic or inorganic 246  
compounds), macronutrients (carbon, nitrogen, hydrogen, 247  
oxygen, etc.), trace elements (Co, Zn, Cu, Mn, etc.), and water. 248  
A careful selection of the appropriate microorganisms that 249  
perform well in the removal of the desired substances (nitrates, 250  
sulfates, and organic matter) is one of the first steps to be made 251  
in the biorestitution strategies. Microorganisms can be isolat- 252  
ed from the environment, like from the soil, that is one of the 253  
most abundant sources of microorganisms with an estimation 254  
up to  $4 \times 10^6$  different taxa in a ton of soil (Curtis et al. 2002) 255  
ad These microorganism display a wide diversity in enzymatic 256  
activities (lipases, proteases, oxido-reductases, etc.) as described 257  
by metagenomic studies (Neelakanta and Sultana 2013). 258

In order to use microorganisms for the CH biorestitution, 259  
the viability and efficiency of the selected microorganisms 260  
must be guaranteed, and it must be verified that they do not 261  
cause deterioration to the CH surfaces to be cleaned. There- 262  
fore, an appropriate “delivery system” for the application of 263  
microorganisms, providing them an adequate microenviron- 264  
ment to optimise their activity, is essential. The delivery 265  
system is, therefore, one of the most important aspects of 266  
biocleaning technology. To guarantee the best conditions, 267  
the ideal delivery system should have the following 268  
characteristics: 269

- i) be able to retain the microorganisms and provide them 270  
the right conditions (aerobic or anaerobic) and the water that 271  
they need in order to remove the cause of decay, but without 272  
any damage to the art work itself and any undesirable changes 273  
in the color of the surface; 274
- ii) be applicable to all types of surfaces (horizontal, vertical, 275  
oblique, rough, smooth, etc.). 276
- and iii) be quick and easy to prepare, but also easy to apply 277  
and to eliminate at the end of the treatment, and using as far as 278  
possible a few cheap materials. 279

Among the recent studies carried out on CH biocleaning 280  
over the last few decades (see Table 1), eight delivery systems 281



282 have been utilized and reported: immersion (Gauri et al. 1989  
283 and 1992; Heselmeyer et al. 1991), sepiolite (Ranalli et al.  
284 1996a, 1996b, 1997, 2000; Cappitelli et al. 2006),  
285 hydrobiogel-97 (Ranalli et al. 2000; Cappitelli et al. 2006),  
286 cotton wool (Ranalli et al. 2005; Antonioli et al. 2005; Bosch-  
287 Roig et al. 2010), carbogel (Cappitelli et al. 2005, 2006, 2007;  
288 May et al. 2008; Polo et al. 2010; Alfano et al. 2011), mortar  
289 and alginate beads (May et al. 2008), agar (Bosch-Roig et al.  
290 2012, 2013a, b; Barbabietola et al. 2012), and arbocel  
291 (Troiano et al. 2013). Table 1 reports a summary of the  
292 delivery systems used in the biocleaning (on stone surfaces  
293 and frescoes) literature, the decay agents, the used microor-  
294 ganisms, their type of metabolism, the application times, and  
295 the removal efficiency evaluation. The first study performed  
296 in this field used the immersion technique. Gauri et al. and  
297 Heselmeyer et al., immersed marble statues and sandstone  
298 blocks affected by sulfur dioxide crusts in liquid containing  
299 *Desulfovibrio desulfuricans*, under laboratory conditions, for  
300 about 60 h (Gauri et al. 1989 and 1992; Heselmeyer et al.  
301 1991). After these studies, it has been recognized that the  
302 immersion technique in liquid culture can have many limita-  
303 tions for large, fragile artworks and that a portable application  
304 delivery system is needed to provide a suitable biocleaning  
305 strategy. Therefore, subsequent research strategies were based  
306 on the use of delivery systems (contact compresses), and new  
307 techniques were developed.

308 However, with the existing diversity in delivery sys-  
309 tems found in the literature, often it remains unclear  
310 which one should be selected. Therefore, this critical  
311 review aims to describe and compare the main character-  
312 istics and properties of the delivery systems adopted until  
313 now in order to help conservator-restorers and researchers  
314 to choose the most appropriate system according to spe-  
315 cific biocleaning application requirements.

## 316 Immersion

317 The immersion system consists of introducing the works of art  
318 into a liquid culture with the appropriate microorganism for a  
319 determined period of time (Gauri et al. 1989, 1992;  
320 Heselmeyer et al. 1991). The immersion system has been  
321 shown to have three main drawbacks: firstly, this kind of  
322 treatment cannot be applied to large objects, such as buildings,  
323 as it necessitates the complete immersion of the object in a  
324 recipient with a liquid medium; secondly, consolidation of the  
325 artwork prior to the treatment becomes obligatory in many  
326 stone types to prevent severe damages due to the immersion;  
327 and, thirdly, treatment efficiency has not been fully proved  
328 since gypsum removal was only evaluated by visual observa-  
329 tion and not by a careful chemical analysis (Gauri et al. 1989,  
330 1992; Heselmeyer et al. 1991). The type of damage produced  
331 to the work of art after immersion treatment and during the

drying of the artwork can be due to salt migration that can  
produce efflorescence on the surface; the presence of diverse  
materials on the work of art can lead to water retention  
variation and unequal material expansion, causing exfoliation,  
fissures or fractures, for example, to the internal structure of  
the work of art. If the artwork is made up from different pieces  
linked together by metallic elements, these elements could be  
oxidized by the immersion and could produce fractures in the  
work of art itself. The laboratory works from De Belie et al.,  
and De Graef et al., performed in 2005 compared the immer-  
sion and the sprinkling strategies to bioclean concrete (blast  
furnace slag cement and ordinary Portland cement) fouled by  
lichens using a mixture of bacteria of the genus *Thiobacillus*  
sp. with an appropriate nutrient broth for nine days. This study  
showed that the sprinkling treatments had about 50 % of the  
effectiveness of the immersion treatments. But both treatments  
had the drawback of the formation of a white gypsum layer on  
some of the cement specimens (De Graef et al. 2005; De Belie  
et al. 2005).

## 351 Sepiolite

352 The sepiolite mineral matrix is a clay mineral, which is a  
353 complex magnesium silicate that increases the total useful  
354 water fraction of a substrate, making the implementation of  
355 active liquids easier by transforming them into semisolids,  
356 permitting the gradual release of active ingredients.

357 Sepiolite is normally used in oil drilling, for cat litter,  
358 and in a solid form for carving. It is also used, due to its  
359 high water retention capacity, in construction lime mortars  
360 and in agriculture. It is used to control water and fertilizer  
361 loss in sandy soils.

362 Sepiolite was introduced for biocleaning strategies in 1996.  
363 Good results were obtained for the removal of nitrates using  
364 *Pseudomonas stutzeri* on brickwords and calcareous stones  
365 (marble, Vicenza-stone, etc.) in laboratory conditions (Ranalli  
366 et al. 1996b); sulfates were removed from an old marble  
367 sculpture and an old marble column using *Desulfovibrio* sp.  
368 under anaerobic conditions (Ranalli et al. 1996a, 1997); and  
369 black crusts on stone and marble materials were efficiently  
370 removed after 30 h to 72 h (Cappitelli et al. 2006).

371 Its preparation for application for biocleaning consists of  
372 mixing the sepiolite powder (50-70 %) with a suspension of  
373 microorganisms and water under anaerobic conditions,  
374 allowing the microorganisms (bacteria) to colonize the sepio-  
375 lite for 10 - 14 days until it is ready to be applied over Japanese  
376 paper to the work of art surface to be treated (Cappitelli et al.  
377 2006) (Fig. 1). The Japanese paper is used to facilitate the  
378 removal of the delivery system and to reduce its penetration  
379 into the pores and cavities of the original surface reducing the  
380 possibility of residues on the cleaned artwork. Despite the  
381 high efficiency of the biocleaning treatments using sepiolite

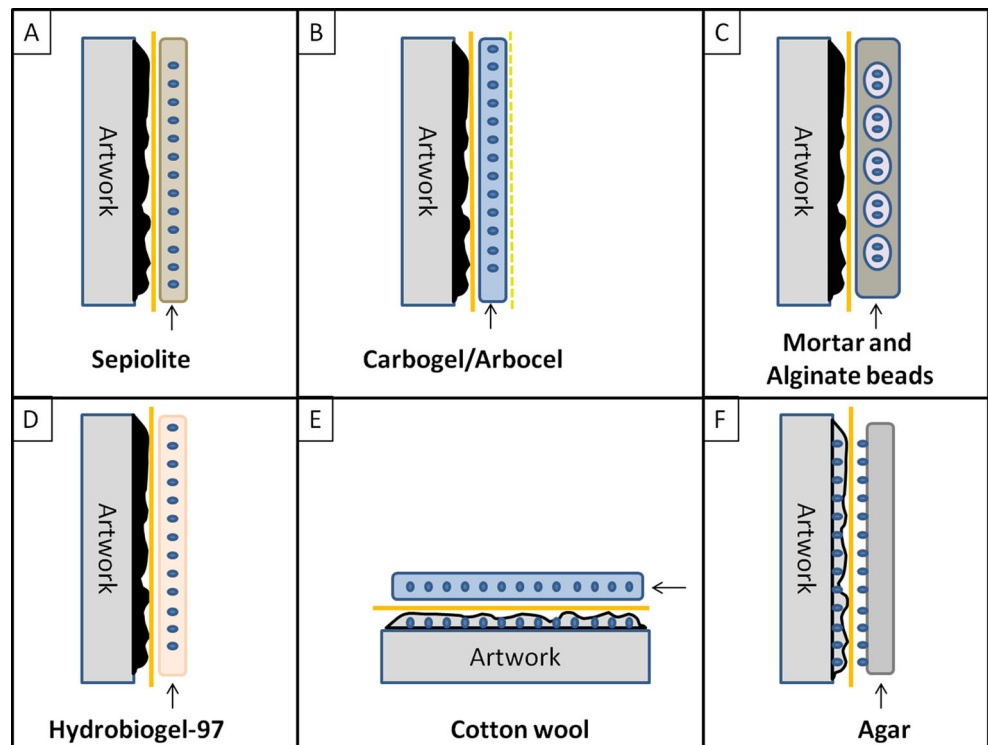
**Table 1** Summary of the delivery systems described in the stone surfaces and frescoes biocleaning literature

| Delivery system | Decay agents                       | Type of materials   | Used microorganisms                      | Type of metabolism | Time (h)                | Removal efficiency evaluation (%) | References                                       |
|-----------------|------------------------------------|---|--|--------------------|-------------------------|-----------------------------------|--|
| t1.1            |                                    |   |  |                    |                         |                                   |  |
| t1.2            |                                    |   |  |                    |                         |                                   |  |
| t1.3            | Immersion                          | Marble (Georgia) stone and statue (U.S.)                          | <i>D. desulfuricans</i>                  | Anaerobic          | 60–84                   | 80                                | Gauri et al. (1989), 1992)                       |
| t1.4            |                                    | Marble and sandstone, (D) Concrete, (B)                           | <i>D. vulgaris Thiobacillus sp.</i>      | Aerobic            | ns <sup>a)</sup> 9 days | 100, 40 ns                        | Heselmeyer et al. (1991)                         |
| t1.5            |                                    |   |  |                    |                         |                                   | De Graef et al. (2005)<br>De Belie et al. (2005) |
| t1.6            | Septolite                          | Marble, (I)   | <i>D. desulfuricans, D. vulgaris</i>     | Anaerobic          | 36                      | 81                                | Ranalli et al. (1996a), 1997)                    |
| t1.7            | Black crusts                       | Marble (Candoglia stone), (I)                                     | <i>D. vulgaris subsp. vulgaris</i>       | Anaerobic          | 45                      | 98                                | Cappitelli et al. (2006);                        |
| t1.8            | Nitrates                           | Brick-works and calcareous stones (I) (marble and Vicenza-stones) | <i>P. stutzeri</i>                       | Aerobic            | 30                      | ns                                | Ranalli et al. (1996b)                           |
| t1.9            | Hydrobiogel-97                     | Marble (Candoglia stone), (I)                                     | <i>D. vulgaris subsp. vulgaris</i>       | Anaerobic          | 7 days                  | 28                                | Cappitelli et al. (2006);                        |
| t1.10           |                                    |   |  |                    |                         |                                   | Ranalli et al. (2000)                            |
| t1.11           | Cotton wool                        | Frescoes, Pisa (I)  | <i>P. stutzeri</i> A29+Protease          | Aerobic            | 2–12                    | 80–100                            | Ranalli et al. (2005);                           |
| t1.12           |                                    |   |  |                    |                         |                                   | Antonoli et al. (2005)                           |
| t1.13           |                                    |   |  |                    |                         |                                   | Lustrato et al. (2012)                           |
| t1.14           | Animal glue and salt efflorescence | Frescoes, Valencia (E)  | <i>P. stutzeri</i>                       | Aerobic            | 1.5–3                   | 60                                | Bosch-Roig et al. (2010)                         |
| t1.15           | Carbogel                           | Marble (Candoglia stone) Milan (I)                                | <i>D. vulgaris subsp. vulgaris</i>       | Anaerobic          | 45                      | 98                                | Cappitelli et al. (2006), 2007)                  |
| t1.16           |                                    | Marble sculpture, Milan, (I)                                      |  |                    | 12                      | ns                                | Cappitelli et al. (2005)                         |
| t1.17           |                                    | Limestone sculpture, Trento (I) Coloured lithotypes*, Firenze (I) |  |                    | 36 30–40                | ns ns                             | Polo et al. (2010)                               |
| t1.18           |                                    |   |  |                    |                         |                                   | Gioventù et al. (2011)                           |
| t1.19           | Nitrates and sulphates             | Sandstone walls, Matera (I)                                       | <i>P. pseudoalcaligenes, D. vulgaris</i> | Aerobic/Anaerobic  | 24–72                   | 55, 85                            | Alfano et al. (2011)                             |
| t1.20           |                                    |   |  |                    |                         |                                   | May et al. (2008)                                |
| t1.21           | Mortar and alghate beads           | Sandstone walls, Matera (I)                                       | <i>P. pseudoalcaligenes</i>              | Aerobic/Anaerobic  | 1 month                 | ns                                | May et al. (2008)                                |
| t1.22           | Agar                               | Frescoes, Valencia (E)  | <i>P. stutzeri</i>                       | Aerobic            | 1.5–2                   | 92                                | Bosch-Roig et al. (2012), 2013a, b)              |
| t1.23           | Arbocel                            | Marble column and statue, Cemetery of Milan (I)                   | <i>D. vulgaris subsp. vulgaris</i>       | Anaerobic          | 68–110                  | ns                                | Troiano et al. (2013)                            |

<sup>a)</sup> (ns: not specified) \* Green serpentine, red marlstone, and white Carrara marble

|     |  |   |     |
|-----|--|---|-----|
| 382 | and the advantage represented by the high level of sepiolite                         | animal glue from frescoes on the Santos Juanes church in                    | 429 |
| 383 | specific surface ( $\text{cm}^2/\text{cm}^3$ ), it has also the ability to offer the | Valencia (Bosch-Roig et al. 2010).  | 430 |
| 384 | anaerobic conditions required during treatment by the micro-                         | For the use of this delivery system in a biocleaning process,               | 431 |
| 385 | organisms like <i>Desulfovibrio</i> sp. involved in the process                      | the live bacterial cell suspension at a high cell density                   | 432 |
| 386 | (Ranalli et al. 1997). Nevertheless, there are some notable                          | ( $5 \times 10^8 \text{CFU/ml}$ ) must be brushed on to the work of art and | 433 |
| 387 | disadvantages when using it (Ranalli et al. 1996a, 1996b,                            | then entirely covered with a thin layer of hydrophilic, sterile,            | 434 |
| 388 | 1997, 2000). The drawbacks in the use of sepiolite for                               | white cotton wool impregnated with the same bacterial cell                  | 435 |
| 389 | biocleaning are: firstly, that it requires a long time (about                        | suspension (Fig. 1). Although good biocleaning results were                 | 436 |
| 390 | two weeks) for the bacterial colonization before application,                        | obtained, this delivery system has some drawbacks. It is not                | 437 |
| 391 | secondly, there can be a rapid loss of water after application                       | recommended for use on vertical surfaces because, due to the                | 438 |
| 392 | that can lead to bacterial inactivity and, thirdly, there is a risk                  | effect of gravity on the water and bacteria suspension within               | 439 |
| 393 | of hydrogen sulfide reaction with the iron in the sepiolite                          | the cotton, there can be seepage through the lower part,                    | 440 |
| 394 | forming iron sulfide precipitates (Cappitelli et al. 2006).                          | leading to water loss through leakage and heterogeneous                     | 441 |
|     |  | biocleaned areas (with a more intense degree of cleaning on                 | 442 |
|     |  | the lower part). This is due to the high water retention and                | 443 |
| 395 | <b>Hydrobiogel-97</b>  | water release of the cotton wool, and long treatments (periods              | 444 |
|     |  | of longer than 15 - 17 h) can produce some risk of damage to                | 445 |
| 396 | The hydrobiogel-97 delivery system is a polymer formed by                            | the fresco, such as the swelling and detachment of paint                    | 446 |
| 397 | an acrylic resin hydrogel (Ranalli et al. 2000; Cappitelli et al.                    | fragments (Bosch-Roig et al. 2010, 2013b).                                  | 447 |
| 398 | 2006). This biogel displayed optimal hydration during a 9-day                        |   |     |
| 399 | biocleaning treatment of stone material to remove black                              |   |     |
| 400 | crusts. There were no chromatic changes and no physical or                           |   |     |
| 401 | chemical modifications of the composition of the stone                               |   |     |
| 402 | (Ranalli et al. 2000). Preparation for biocleaning system con-                       |   |     |
| 403 | sists of mixing for ten min at room temperature by mechanical                        |   |     |
| 404 | agitation of the two physic-chemical liquid components of the                        |   |     |
| 405 | hydrobiogel-97. One of the components is the main agent, and                         |   |     |
| 406 | the second is a catalyst that allows for obtaining the final gel                     |   |     |
| 407 | (the composition of the gel is confidential, know-how of                             |   |     |
| 408 | EniTecnologie, San Donato Milanese, Italy). Then, just before                        |   |     |
| 409 | use, the suspension of microorganisms in water solution is                           |   |     |
| 410 | added, and the biocleaning agent is applied over Japanese                            |   |     |
| 411 | paper onto the work of art to be treated (Fig. 1) (Cappitelli                        |   |     |
| 412 | et al. 2006). The disadvantages of this delivery system are: the                     |   |     |
| 413 | difficulty in eliminating it after the treatment due to the intrinsic                |   |     |
| 414 | aspects of its high adhesiveness, and excessive fluidity that                        |   |     |
| 415 | complicates its application to nonhorizontal surfaces.                               |   |     |
|     |  | <b>Carbogel</b>   | 448 |
| 416 | <b>Cotton wool</b>   |   |     |
|     |  | Carbogel is composed of a neutralized polyacrylic acid, which               | 449 |
| 417 | Cotton wool is a cellulose material obtained from the                                | permits the gel to be prepared simply by adding water. Vis-                 | 450 |
| 418 | <i>Gossypium</i> sp. seeds with a characteristic microstructure that                 | cosity can be varied as desired. It has been used in restoration            | 451 |
| 419 | makes it very resistant, malleable, soft, and very absorbent.                        | for the cleaning of wall paintings (Borgioli et al. 2001).                  | 452 |
| 420 | Cotton wool has been used in biocleaning since 2000. It was                          | Several biocleaning treatments using carbogel as a delivery                 | 453 |
| 421 | first used, with <i>Pseudomonas stutzeri</i> A29 strain and applied                  | system have shown that it can be used successfully to clean                 | 454 |
| 422 | for 2 - 12 h, to efficiently bioclean (80-100 % animal glue                          | black crusts and calcite and gypsum deposits (from Matera                   | 455 |
| 423 | removal) a fresco <i>Conversione di S. Efsio e battaglia</i> (The                    | Cathedral, Milan Cathedral, Florence Cathedral, from sculp-                 | 456 |
| 424 | conversion of St. Efsio and the battle), which had been                              | tures like the Rondanini Pietà base by Michelangelo                         | 457 |
| 425 | detached from the walls of the Camposanto Monumentale,                               | Buonarroti in Sforzesco Castle in Milan, and from sculptures                | 458 |
| 426 | Pisa, Italy (Ranalli et al. 2005; Antonioli et al. 2005). Inter-                     | in the Courtyard of Buonconsiglio Castle in Trento)                         | 459 |
| 427 | esting results were also obtained with cotton wool as a deliv-                       | (Cappitelli et al. 2005, 2006, 2007; May et al. 2008; Polo                  | 460 |
| 428 | ery system and <i>P. stutzeri</i> to clean nitrate salt efflorescence and            | et al. 2010; Alfano et al. 2011; Gioventù et al. 2011). It has              | 461 |
|     |  | also been successfully used to remove nitrates from stonework               | 462 |
|     |  | (at Matera Cathedral) (May et al. 2008; Alfano et al. 2011).                | 463 |
|     |  | Carbogel preparation for biocleaning consists of mechanical                 | 464 |
|     |  | mixing, preferably using an electrical device (automatic                    | 465 |
|     |  | mixer) adding the carbogel powder (0.5-4 %) with a suspen-                  | 466 |
|     |  | sion of microorganisms (in water/P-buffer/DNT medium) at                    | 467 |
|     |  | room temperature. The biocleaning agent obtained is then                    | 468 |
|     |  | applied over a Japanese paper to the artwork to be treated.                 | 469 |
|     |  | When a vertical surface has to be biocleaned with carbogel,                 | 470 |
|     |  | the multilayer biosystem should be applied. This system con-                | 471 |
|     |  | sists of a Japanese paper in contact with the wall, a 0.8-1.9 cm            | 472 |
|     |  | thick layer of carbogel powder mixed with a bacterial suspen-               | 473 |
|     |  | sion, a plastic reticulated net (PET with 25 empty space/ $\text{cm}^2$ )   | 474 |
|     |  | to facilitate adhesion and the distribution of the carbogel over            | 475 |
|     |  | the surface area, and, lastly, a plastic film to reduce undesired           | 476 |
|     |  | water evaporation (see Fig. 1) (Alfano et al. 2011). To avoid               | 477 |
|     |  | carbogel handling difficulties, some researchers added                      | 478 |

**Fig. 1** Delivery systems for stone surface and fresco decay recovery. **a:** Sepiolite; **b:** Carbogel/arbocel; **c:** Mortar and alginate beads; **d:** Hydrobiogel-97; **e:** Cotton wool; **f:** Agar. Figure notes: delivery systems are indicated with a black arrow; alterations are indicated with a black area over the artwork; bacteria are indicated as blue spheres/circles; Japanese paper is indicated as a yellow line; and plastic net is indicated with yellow discontinuous line



479 micronized silica to the carbogel, creating a more homoge- 505  
 480 neous, compact delivery system, which is, therefore, easier to 506  
 481 handle and contributes to more anaerobic conditions when 507  
 482 adopted to remove black crusts with *Desulfovibrio vulgaris* 508  
 483 (Sorlini et al. 2010b). 509

484 The advantages of the carbogel biocleaning delivery sys- 510  
 485 tem are that it has a high water retention capacity with conse- 511  
 486 quent evaporation over longer periods, that no color change 512  
 487 appeared on the stone, and no visual residue was found 513  
 488 through SEM (Scanning Electron Microscopy) analyses after 514  
 489 treatment. The drawbacks of this delivery system are that it 515  
 490 can liquefy with high salt content making its handling very 516  
 491 difficult, and it induced a few changes in the structure that can 517  
 492 increase porosity and water uptake of the treated materials On 518  
 493 long-term treatments it dries out, so it can only be used for 519  
 494 brief applications (May et al. 2008); it has limited adhesive 520  
 495 properties and; therefore, it can be difficult to use for cleaning 521  
 496 vertical and hydrophobic surfaces, ceiling and vaults. Finally, 522  
 497 carbogel, given its less compact structure, has been shown to 523  
 498 be slightly difficult to handle (Beltrami et al. 2012). 524

### 499 Mortar and alginate beads

500 This delivery system is made up of a base of mortar that is 525  
 501 mixed with alginate beads containing the selected micro- 526  
 502 organisms. Its preparation consists of mixing 100 ml gel 527  
 503 (containing 90 ml mineral medium nutrient solution, 10 ml 528  
 504 bacteria-tween-solution containing  $10^6$  cells/ml, 529

505 and 3 % alginate beads) with 700 g of mortar. The 506  
 507 alginate beads are formed as follows: mix the solution 508  
 509 for 10 min on a magnetic stirrer, pick up the gel mix in a 510  
 511 syringe and drip the gel mix into a 2 % calcium chloride 512  
 513 solution, leave the alginate beads for 10 min in the solu- 514  
 515 tion, and, finally, wash them in purified water. The mortar 516  
 517 is formed by 205 g of CEM III/C Portland cement, 301 g 518  
 519 of standard sand (0 - 2 mm particle size), 91 g of pumice 520  
 521 (0.3 - 1.5 mm particle size), 103 ml H<sub>2</sub>O, and 0.5 g of air 522  
 523 entraining agent. 524

525 This delivery system was used by May et al. (2008) in the 526  
 527 European BIOBRUSH project (Bioremediation for building 528  
 529 Restoration of the Urban Stone Heritage in European States, 530  
 531 no. EVK4-2001-00055) to clean the external walls of Matera 532  
 533 Cathedral in Italy. 534

535 The mortar gel system used for bioremediation of 536  
 537 monuments consists of either applying it directly, or over 538  
 539 Japanese paper, to the work of art surface to be treated 539  
 540 (see Fig. 1). This delivery system has been shown not to 541  
 542 be suitable for biocleaning strategies. This is due to the 542  
 543 fact that the powerful adhesive properties of the mortar 543  
 544 make its removal after treatment very problematic. Me- 544  
 545 chanical strategies have to be used, producing damage to 545  
 546 the original matrix (May et al. 2008). Mortar performance 546  
 547 is also reduced due to its long preparation procedure (May 547  
 548 et al. 2008). May et al. (2008) in comparative studies 548  
 549 between the carbogel and the mortar and alginate bead 549  
 550 delivery systems showed faster capabilities of the 550  
 551 carbogel over mortar in removing nitrates at Matera. 551  
 552 553



Agar is a complex polysaccharide extracted from a group of red purple algae of Rhodophyceae, usually from the *Gelidium* and *Gracilaria* genera. It is composed of two types of polysaccharides, agarose and agaropectine. It is water soluble, neutral, nontoxic, and able to produce gels with no liquid water release (Praiboon et al. 2006; Gulotta et al. 2014). It is commonly used in the food industry as a jelling and thickening agent; in microbiology and botanic fields as a support for the growth of certain organisms (Hesse and Hesse 1992); in pharmaceuticals for drug delivery (Santoro et al. 2011); and in chemical field as an electrode binder for electrolyte cells (An et al. 2013). For a few years, agar has also been used in restoration, for the cleaning of different types of works of art (mural painting, wood, stone, plaster, paper, and textiles), for its ability to control and limit water release onto artistic materials, and for its respect for the substrate (Iannucelli and Sotgiu 2009; Gorel 2010; Sansonetti et al. 2012; Casoli et al. 2013; Nualart-Torroja et al. 2013; Shaeffer and Gardiner 2013; Baglioni et al. 2014; Gulotta et al. 2014). Recent studies evidenced that after agar applications on porous art works no residue remains inside the porous materials (measured by FTIR spectroscopy and mass spectroscopy) (Cremonesi 2013; Tortajada Hernando and Blanco Domínguez 2013). Agar gel has also the advantage that it retains water-soluble dissolved substances like a sponge, reducing post treatment cleaning of the surface, and suggesting its ability to be used not only to clean surfaces but also to extract salts (Anzani et al. 2008; Cremonesi 2013; Gulotta et al. 2014).

Recently agar 2 % has been used as biocleaning delivery system, performing effective removal of nitrate salt efflorescence and animal glue residues during in situ treatments, with *Pseudomonas stutzeri*, on the frescoes of the central vault of the Santos Juanes Church in Valencia, Spain (Bosch-Bosch-Roig et al. 2012, 2013a, b). Animal glues have been also successfully removed from paper materials after 4 h of biocleaning treatment with *Ochrobacterium* sp. bacteria immobilized in agar 1 % (Barbabetola et al. 2012). The preparation protocol of this delivery system consists of diluting the agar powder in distilled water to a final 1–2 % concentration, then heated to above 85 °C or autoclaved and placed under sterile conditions in plastic molds of the desired size producing agar layers. Once the solution is cooled, the polymer molecules assemble forming a thermoreversible rigid gel (Medina-Esquivel et al. 2008; Cremonesi 2013). The agar microstructure has a high number of homogeneous size pores enhancing its water retention (Pernodet et al. 1997). Some authors advised against the use of 4 % agar over 15 min and to avoid leaving the agar completely dry (at any concentration) (Tortajada Hernando and Blanco Domínguez 2013; Gulotta et al. 2014). The protocol consists of an initial application of Japanese paper to protect the altered paint surface; the

bacterial suspensions are then applied with a sterile brush both onto the Japanese paper and onto one side of the agar surface and, finally, the side of the agar with the bacteria is placed onto the surface of the work of art (see Fig. 1). After treatment, the agar and Japanese paper have to be removed and the treated surface cleaned using sterile water and a sponge to avoid undesirable bacterial metabolic processes and damage to the original material. The painted surface must then be dried (Bosch-Roig et al. 2013b). Recent applications show that agar has good adhesive properties (it can be applied onto horizontal, vertical, and oblique surfaces); it gives a significant level of water retention and bacteria retention and produces homogeneous cleaning without leaving stain marks or residues on the surface (Bosch-Roig et al. 2013a). Another advantage of agar is that it is semitransparent, due to its lack of inherent color, which facilitates the control of the biocleaning process (Iannucelli and Sotgiu. 2010; Cremonesi 2013; Gulotta et al. 2014). In using agar the fact that capillary forces tend to draw material from the substrate into the gel, may also be of interest (Wolbers 2000).

The rigid agar delivery system has shown two drawbacks, poor adhesion when used for the biocleaning of rough surfaces and the difficulty in finding plastic molds in which to prepare agar of the desired shape when the artwork shapes are complex. However, recent studies in restoration have solved these problems by using semisolid agar that consists of applying the warm gelling agar (40 °C–45 °C) just before it becomes cold and solid, thus, allowing it to cover rough and irregular surfaces and objects with complex shapes by brushing without damaging the surface nor releasing residues (Anzani et al. 2008; Cremonesi 2013; Tortajada Hernando and Blanco Domínguez 2013; Gulotta et al. 2014).

## Arbocel

Arbocel is a natural cellulose fiber with a wide range of uses including: complementary food for animals (for regulation and support of the function of the gastrointestinal tract), paper and board production in the pharmaceutical industry (used as an economic, inert additive in tablets and capsule filling, improving tablet hardness, and disintegration time), in the production of modern detergents, for the coating of flower and vegetable seeds, in chemical construction products, and so on. In art restoration and conservation, it is used as a support for compresses for the cleaning of natural and artificial stone materials, frescoes, plaster, and stucco. It can also be used as inert filler for the preparation of mortars and plasters.

Arbocel preparation for biocleaning consists of mixing the arbocel powder (CTS, Vicenza, Italy) with a biomass suspension (in water/P-buffer/DNT medium) until obtaining a homogeneous mixture with the desired density. The system is applied to the work of art over Japanese paper previously

636 moistened with a phosphate buffer. When working with sul- 686  
637 fate reduction bacterium like *D. vulgaris*, all the manipula- 687  
638 tions should be done under anaerobic conditions in an anaer- 688  
639 obic box for a guarantee of the opportune anoxic-anaerobic 689  
640 metabolism (Troiano et al. 2013). The biological cleaning 690  
641 system is then covered with a plastic film to reduce undesired 691  
642 evaporation of water (see Fig. 1) (Troiano et al. 2013). This 692  
643 procedure is based on the one previously reported for carbogel 693  
644 (Alfano et al. 2011). The arbocel delivery system with 694  
645 *Desulfovibrio vulgaris* subsp. *vulgaris* entrapped biomass 695  
646 has shown good results for the biocleaning of black crusts 696  
647 present on stone artwork (columns and marble statues) 697  
648 (Troiano et al. 2013), but it has not yet been used for cleaning 698  
649 frescoes. Due to its similarities in composition to cotton wool, 699  
650 it is probable that the drawbacks of this delivery system will 700  
651 be similar to those described for cotton wool. Further studies 701  
652 should be performed with this delivery system to confirm its 702  
653 suitability for the biocleaning of polychromic works of art, 703  
654 such as frescoes. 704

#### 655 **Critical comparison of delivery systems main** 656 **characteristics**

657 A discussion and comparison of the main characteristics of the 710  
658 various delivery systems used for biocleaning of stone CH and 711  
659 frescoes is reported according to all published works (Fig. 2). 712  
660 The delivery system to use for restoration purposes must, 713  
661 therefore, be chosen depending on the water retention, water 714  
662 release, bacterial compatibility, adhesion capabilities, risks 715  
663 including CH undesired color changes, performance charac- 716  
664 teristics and difficulties, economical sustainability, and, of 717  
665 course, the state of conservation of the artwork to be cleaned. 718

666 In Table 2 the relationship between the main decay agents, 719  
667 the suggested biocleaning microorganisms, and the delivery 720  
668 systems benefits and drawbacks are shown. The benefits 721  
669 (including water retention and bacterial compatibility) and 722  
670 drawbacks (including adhesion capabilities, risks, and perfor- 723  
671 mance difficulties) are indicated in detail for each delivery 724  
672 system. To facilitate an easy reading and a correct understand- 725  
673 ing for conservator-restorers, on the basis of our past experi- 726  
674 ence and of the data present on the literature, the following 727  
675 notes both for benefits and drawbacks have been attributed: 728  
676 absence; low; high; very high; and not applicable. Finally, an 729  
677 economical evaluation shows the costs of the delivery system 730  
678 alone (€/kg) and the total costs (€/m<sup>2</sup>) including full 731  
679 biocleaning systems (microorganisms and delivery systems). 732

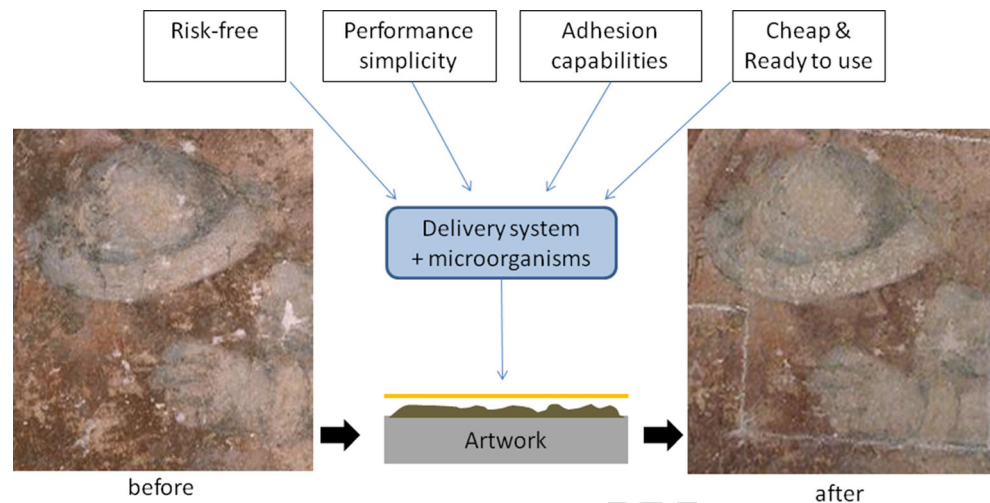
680 An ideal delivery system must have high water retention 733  
681 (water is necessary for the microorganism) while water release 734  
682 should be relatively low, in order not to damage the works of 735  
683 art themselves (salt migration, exfoliation, fissures, fractures, 736  
684 oxidation, etc.). The fact that the water release and water 737  
685 retention is related to the ratio raw powder/water must be 738

686 taken into account when using cleaning gels. Gels, for exam- 687  
688 ple made with a high content of agar (4–6 % w/v), have a 688  
689 much slower water release and a lower water retention com- 689  
690 pared to gels made with lower agar content (1–3 % w/v) 690  
691 (Wolbers 2000; Iannuccelli and Sotgiu 2010). When 3 % 691  
692 semisolid agar gel is used for 1 h in the biocleaning of 692  
693 Candoglia marble sculpted stone, the contact of the water 693  
694 liquid and the treated surface is more limited compared with 694  
695 the use of sepiolite and cellulose pulp based delivery systems 695  
696 (Beltrami et al. 2011; Gulotta et al. 2014). These results are in 696  
697 accordance with a study showing that a thin layer of cotton 697  
698 wool soaked in water and applied to gypsum models for only 698  
699 3 min show water penetration of up to 5–6 mm in depth, while 699  
700 agar water penetration varies from 1 to 4 mm depending on 700  
701 application times (3–20 min) and the percentage of agar 701  
702 (2–4 %) (Anzani et al. 2008). Given its water-related 702  
703 characteristics, cotton wool in contact with nonhorizontal 703  
704 porous surfaces or in the presence of surface alterations 704  
705 (and more probably also arbocel due to its cellulosic 705  
706 nature) can produce water marks on the treated areas 706  
707 (Bosch-Roig et al. 2010, 2013b). This undesired effect is 707  
708 reduced or entirely absent when using the other delivery 708  
709 systems tested (Bosch-Roig et al. 2013b). 709

710 The compatibility between the delivery system and the 710  
711 microorganism is another characteristic of the delivery sys- 711  
712 tem, which is vital for an efficient biocleaning process, be- 712  
713 cause incorrect bacterial compatibility will lead to poor or 713  
714 inappropriate cleaning. All the delivery systems reported have 714  
715 shown high or very high bacterial compatibility. Sepiolite 715  
716 displays the disadvantage, among others, that the living cells 716  
717 have to colonize the delivery systems, and; therefore, addi- 717  
718 tional time consumption is required before application 718

719 Delivery system adhesion capabilities are very important 719  
720 when cleaning vertical or oblique surfaces, but very high 720  
721 adhesion can be negative or problematic. The ideal delivery 721  
722 system adhesion capabilities must be enough to guarantee the 722  
723 effectiveness of the biocleaning treatment (limited adhesion 723  
724 can reduce biocleaning efficacy), and assure the complete 724  
725 removal of the delivery system after treatment with the ab- 725  
726 sence of the original material's modification. Excessive adhe- 726  
727 sion can make the removal of the delivery system very com- 727  
728 plicated (as with the mortar system) and may cause decay to 728  
729 the works of art or make impossible the complete removal of 729  
730 the delivery system from the surface. Cotton wool's low 730  
731 adhesion capabilities cannot guarantee long treatments on 731  
732 vertical surfaces. Various strategies may be adopted to im- 732  
733 prove the limited adhesive capabilities of some delivery sys- 733  
734 tems. Jelling polyacrylic acid materials like carbopol have 734  
735 recently been introduced for restoration cleaning strategies 735  
736 after their use in the cosmetics industry in last few decades 736  
737 (Beltrami et al. 2012; Baglioni et al. 2014). This interesting 737  
738 material seems to avoid the adhesive drawbacks of carbogel 738

**Fig. 2** Main characteristics of the most commonly used delivery systems as key components in biocleaning strategies of Cultural Heritage artwork adopted as a useful tool for researchers and conservator-restorers. Pictures show the effects of the biocleaning process on a fragment of the Conversione di S. Eufisio e battaglia, fresco (XIV century) at Pisa Camposanto Monumentale, (Italy)



739 (low adhesion capabilities), and may, therefore, also be stud- 775  
 740 ided for biocleaning strategies. Generally the agar adhesion 776  
 741 capabilities are high but when applied to rough surfaces 777  
 742 adhesion can be low. To avoid these adhesion problems of 778  
 743 solid agar, studies in restoration have solved this problem by 779  
 744 using it as semisolid agar. Various restoration researches per- 780  
 745 formed in 2013 (Tortajada Hernando and Blanco Domínguez 781  
 746 2013) have also shown the suitability of semisolid agar for 782  
 747 cleaning fragile plaster materials (partially soluble in water) by 783  
 748 using low concentrations of agar, short treatment times (2–30 784  
 749 min), and very thick layers of agar gel (to facilitate the remov- 785  
 750 al). Several semisolid agar treatments did not show any remov- 786  
 751 al). Several semisolid agar treatments did not show any remov- 787  
 752 al). Several semisolid agar treatments did not show any remov- 788  
 753 al). Several semisolid agar treatments did not show any remov- 789  
 754 al). Several semisolid agar treatments did not show any remov- 790  
 755 al). Several semisolid agar treatments did not show any remov- 791  
 756 al). Several semisolid agar treatments did not show any remov- 792  
 757 al). Several semisolid agar treatments did not show any remov- 793  
 758 al). Several semisolid agar treatments did not show any remov- 794  
 759 al). Several semisolid agar treatments did not show any remov- 795  
 760 al). Several semisolid agar treatments did not show any remov- 796  
 761 al). Several semisolid agar treatments did not show any remov- 797  
 762 al). Several semisolid agar treatments did not show any remov- 798  
 763 al). Several semisolid agar treatments did not show any remov- 799  
 764 al). Several semisolid agar treatments did not show any remov- 800

765 Among the different drawbacks, important factors to consid- 801  
 766 er are the risks to CH works themselves (including undes- 802  
 767 ired color changes and excessive water release), to conservator- 803  
 768 restorers, and to the environment. When working with artwork 804  
 769 materials, no unwanted color changes must be caused by the 805  
 770 biocleaning treatment. Unfortunately, the liquid culture used 806  
 771 in the immersion technique can dilute and dye artwork material. 807  
 772 When using sepiolite, there is a risk of the presence of iron ions 808  
 773 leading to discoloration. This disadvantage for sepiolite can be 809  
 774 avoided by a pretreatment that

eliminates the iron ions present (Ranalli et al. 1996a, 775  
 1996b). An absence of color changes is reported for the other 776  
 delivery systems (Ranalli et al. 2005; Cappitelli et al. 2006; 777  
 May et al. 2008; Alfano et al. 2011; Bosch-Roig et al. 2013a; 778  
 Troiano et al. 2013). Previous studies have demonstrated that, 779  
 among the techniques used for biocleaning, the immersion 780  
 technique and the mortar system are the delivery systems with 781  
 the highest risks for CH works, especially when preconsolidation 782  
 treatments are not applied (Gauri et al. 1989, 1992; May et al. 2008). 783  
 As discussed before, sepiolite that has not been pretreated has a 784  
 risk of color change and cotton wool has a risk of leaving water 785  
 marks when applied to nonhorizontal surfaces. Until now, when 786  
 microorganisms associated with a delivery system have been used, 787  
 no risks for the operators or for the environment have been reported 788  
 for any biocleaning technologies. This is due to the fact that, on 789  
 the one hand, the microorganisms used are adequately isolated 790  
 from the environment (non-GMO) and are always nonpathogenic; 791  
 and on the other hand, the cleaning strategies are nontoxic both 792  
 for the environment and for conservator-restorers as they are 793  
 based on water avoiding the use of toxic organic solvents (Wolbers 794  
 2000; Pietropolli 2001; Cremonesi 2004; Lustrato et al. 2012; 795  
 Bosch-Roig et al. 2013a, 2013b). 796  
 797

798 Furthermore, it is important to take into account the perfor- 800  
 799 mance characteristics and time consuming of each delivery 801  
 802 system including preparation, application, elimination, and 803  
 804 material needs. Cappitelli et al. (2006) reported a comparative 805  
 806 study on sepiolite, carbogel, and hydrobiogel-97 for cleaning 807  
 808 black crusts from stone materials showing that carbogel was 809  
 810 better than sepiolite and hydrobiogel-97 because carbogel was 811  
 the easiest to apply and remove, while maintaining also the 812  
 best bacterial activity. But carbogel, due to its less compact 813  
 structure, has been shown high performance difficulties because 814  
 it is difficult to handle compared to agar and cotton wool, and 815  
 it easily fragments leaving more residues on the artwork surface. 816  
 However, arboce, agar, and cotton have been 817

**Table 2** Benefits drawbacks and economical evaluation of the used biocleaning delivery systems for Cultural Heritage stone surfaces and frescoes

|       | Delivery system   | Benefits                 |                         |                       | Drawbacks |                            |                               | Economical evaluation           |  |
|-------|---|--------------------------|-------------------------|-----------------------|-----------|----------------------------|-------------------------------|---------------------------------|--|
|       |   | Water retention          | Bacterial compatibility | Adhesion capabilities | Risks*    | Performance difficulties** | Delivery system costs*** €/Kg | Total cost**** €/m <sup>2</sup> |  |
| t2.1  |   |                          |                         |                       |           |                            |                               |                                 |  |
| t2.2  | Main decay agents and used biocleaning bacteria                                 |                          |                         |                       |           |                            |                               |                                 |  |
| t2.3  |   |                          |                         |                       |           |                            |                               |                                 |  |
| t2.4  | Black crusts, nitrates, sulphates; <i>D. vulgaris</i> / <i>D. desulfuricans</i> | Immersion                | Very high               | na                    | Very high | Very high                  | 100–300                       | 300–800 <sup>#</sup>            |  |
| t2.5  | <i>P. pseudoalcaligenes</i>   | Sepiolite                | Low                     | High                  | High      | High                       | 30–100                        | 70–80                           |  |
| t2.6  |   | Hydrobiogel-97           | high                    | High                  | Absence   | Very high                  | 200–250                       | 80–90                           |  |
| t2.7  |   | Arbocel                  | Low                     | High                  | Absence   | Low                        | 10–50                         | 70–80                           |  |
| t2.8  |   | Carbogel                 | High                    | Low                   | Absence   | High                       | 30–50                         | 70–80                           |  |
| t2.9  |   | Mortar and alginat beads | Low                     | Very high             | Very high | Very high                  | 400–500                       | 90–100                          |  |
| t2.10 | Salt efflorescence, organic matters; <i>P. stutzeri</i>                         | Agar                     | Very high               | High-low              | Absence   | Low                        | 50–300                        | 10–40                           |  |
| t2.11 |   | Cotton wool              | Low                     | Low                   | Low       | Low                        | 10–20                         | 40–50                           |  |

Legend: The evaluation criteria: absence; low; high; very high; na (not applicable) are related to benefits (water retention and bacterial compatibility) and drawbacks (adhesion capabilities, risks, and performance difficulties) of the delivery systems

\* Risks refers to the hazard to art works (including undesired color changes and excessive water release)

\*\* Performance difficulties include time consumption and handling complexity related to: material needs, preparation, application and elimination of each delivery system

\*\*\* Delivery system costs are referred only to the materials (do not include bacterial cells)

\*\*\*\* Average total costs are calculated including delivery systems and microorganisms according to the application modality reported on in Fig. 1 and Table 1; microorganisms costs are calculated assuming the use of bacterial cells (i.e., *D. vulgaris* and *P. stutzeri*) in relation to the delivery systems reported on the second column of this Table 2

<sup>#</sup>: Average total costs in €/m<sup>3</sup>



811 shown to be very simple to apply due to their consistent  
812 and compact structure, therefore showing, low perfor-  
813 mance difficulties.

814 On the basis of the benefits and drawbacks evaluation  
815 criteria selected and given the notes previously described,  
816 we can do a classification to help conservator-restorers. In  
817 particular, the best evaluated delivery systems are: arbo-  
818 cel, carbogel, cotton wool, and agar. Among them, all have good  
819 evaluation on all the criteria except for low water retention of  
820 arbo-  
821 cel; carbogel has low adhesion capabilities and high  
822 performance difficulties; cotton wool has low water retention  
823 and low adhesion capabilities on vertical surfaces; and, finally,  
824 agar has low adhesion capabilities on rough surfaces.  
825 Immersion and mortar delivery systems must be avoided due  
826 to their high potential risk for damage to the CH works.

826 The economic aspects of the delivery systems are also very  
827 important, but few studies have been done. Ranalli et al.  
828 (2005) reported a cost study comparing the costs of cleaning  
829 1 m<sup>2</sup> fresco surface with biological cleaning using viable  
830 bacterial cells (with cotton wool as a delivery system) and  
831 using enzymes. The costs of the biological cleaning using  
832 viable bacterial cells, comparing a *P. stutzeri* bacterial culture,  
833 protease enzyme, and a collagenase enzyme were determined.  
834 This analysis of the biocleaning costs showed a cost ratio of  
835 1:3:10, respectively, demonstrating, therefore, that the use of  
836 bacterial cultures instead of enzymes was significantly  
837 cheaper (Ranalli et al. 2005). Lustrato et al. (2012) did another  
838 economic analysis, determining that, under real conditions  
839 and using cotton wool as a delivery system, the bacterial  
840 suspension (*P. stutzeri*) required to clean 1 m<sup>2</sup> of fresco  
841 surface, cost less than 200 euros (including the personal costs  
842 of the bioapplication). Recent studies have shown that, when  
843 using agar as a delivery system, a ten times smaller volume  
844 cell suspension is required per unit of wall painting area,  
845 compared to the cell volume needed for a cotton wool delivery  
846 system (Bosch-Roig et al. 2013a, 2013b). However, the higher  
847 cost of high purity agar, which can be even ten times more  
848 expensive than cotton wool, must be also considered. Another  
849 interesting study shows a cost evaluation of the use of bacteria  
850 for consolidation of stone CH with prices between 23–40€/m<sup>2</sup>  
851 considering the product and the application costs (De Muynck  
852 et al. 2010). An overview of the costs related to the different  
853 biocleaning systems has been summarized in Table 2 accord-  
854 ing to all the existing data. The costs show a wide range of  
855 prices for each delivery system due to the diverse market  
856 price. The total costs of the biocleaning treatments compre-  
857 hend the price of the delivery system including the microor-  
858 ganisms' biomass, according to the application modality  
859 showed in Fig. 1 and the bacteria selection reported in the  
860 literature (see Table 1). Two microorganisms have been taking  
861 into account: *D. vulgaris* (cell suspension of 5x10<sup>9</sup> CFU/ml  
862 and 60 €/m<sup>2</sup>, price of Micro4you) and *P. stutzeri* (cell suspen-  
863 sion of 5x10<sup>8</sup> CFU/ml and 3–30 €/m<sup>2</sup> for agar and cotton

wool, respectively). The microorganism *D. vulgaris* has been  
864 considered to calculate the prices of all delivery systems  
865 except for agar and cotton wool where *P. stutzeri* are adopted.  
866 The price of biocleaning systems based on *D. vulgaris* is  
867 affected by its intrinsic slow anaerobic metabolism compared  
868 to the price of the systems based on *P. stutzeri* (whose prev-  
869 alently aerobic metabolisms permit an easy and fast growth  
870 determining a cost reduction). In addition, the application  
871 modality affects the biocleaning system total price because  
872 the quantity of needed microorganisms can vary substantially  
873 (as is the case of the agar which needs ten times less micro-  
874 organisms in suspensions than does cotton wool). The final  
875 price reported in Table 2 doesn't consider the bioapplication  
876 time consumption and personal costs because they change  
877 depending on the type of artwork and country.

879 In comparing all the delivery system costs, the lower in cost  
880 are arbo-  
881 cel, carbogel, and cotton wool, and the lowest total  
882 cost is for agar, followed by cotton wool. The average of the  
883 total costs is under 100€, except by immersion whose price is  
884 ten times higher than the others.

884 To conclude, on the basis of the selected criteria and in  
885 order to help conservator-restorers to choose the appropriate  
886 system according to specific biocleaning application require-  
887 ments, a useful classification is given: a) when black crust,  
888 nitrate, and sulfate decay agents must be bioremoved,  
889 *D. vulgaris* should be used, and the suggested delivery sys-  
890 tems are arbo-  
891 cel and carbogel; b) when salt efflorescence, and  
892 organic matter decay agents have to be bio-removed, *P. stutzeri*  
893 and agar or cotton wool should be used, on vertical or hori-  
894 zontal surfaces, respectively.

894 Future studies must be conducted to verify and confirm the  
895 medium and long-term safety of these biocleaning methodol-  
896 ogies (Bosch-Roig and Ranalli 2014). Attention must be paid  
897 to strategies for controlling and monitoring any new, posttreat-  
898 ment microbial interaction on biocleaned artwork. This mon-  
899 itoring should include advanced on-site technologies based on  
900 noninvasive tools to understand the potential risks for  
901 biocleaned tangible heritage (Rampazzi et al. 2011;  
902 Raimondi et al. 2013).

903 To conclude, intense research has shown that innovative  
904 microbiological systems based on microorganisms for remov-  
905 ing decay on CH to be good alternatives for restoration. This  
906 review has reported and compared the characteristics of the  
907 existing delivery systems for biocleaning of CH giving infor-  
908 mation that will help conservator-restorers and researchers in  
909 the choice of the most appropriate methods to use in future  
910 applications.

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