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Additional Information

1 **STUDY OF THE CHEESE SALTING PROCESS BY DIELECTRIC**
2 **PROPERTIES AT MICROWAVE FREQUENCIES**

3 **J. Velázquez-Varela¹, M. Castro-Giraldez^{*1}, L. Cuibus², J.A. Tomas-Egea¹, C.**
4 **Socaciu², P.J. Fito¹**

5 ¹Instituto Universitario de Ingeniería de Alimentos para el Desarrollo, Universitat
6 Politècnica de València, Camino de Vera s/n, 46022 Valencia, Spain

7 ² University of Agricultural Sciences and Veterinary Medicine, Mănăştur Street, 3-5,
8 400372, Cluj-Napoca, Romania

9 *Author for correspondence: marcasgi@upv.es

10

11 **Abstract**

12 Salting process involves complex phenomena that affect the overall quality of cheese
13 because of its effect on water activity and induced biochemical changes. Permittivity of
14 cheese was analysed throughout cheese salting treatment in order to relate it with water
15 and salt transports. Salting treatment was carried out by using 25% (w/w) sodium chloride
16 brine at 4°C. Samples were immersed in a vessel containing the osmotic solution with
17 continuous stirring, for 0, 10, 20, 30, 40, 50, 60, 90, 120, 180, 240, 360, 480, 720, 900
18 and 1440 min. Samples were subsequently equilibrated in an isothermal chamber at 4°C
19 for 24 hours. Mass, volume, surface water activity, moisture, ion content and permittivity
20 were determined in fresh and salted samples. Permittivity was measured from 500 MHz
21 to 20 GHz, by using an open-ended coaxial probe connected to a Vector Network
22 Analyzer. Results showed that measurements at 20 GHz explain the water loss and water
23 flux in the overall product. The state of the electrolytes in cheese can be followed using
24 the ionic conductivity at 500 MHz. A coupled measurement of permittivity at 20 GHz
25 and 500 MHz can predict the chemical species involved in cheese salting process, and its

26 structural changes. In conclusion, measures of permittivity in microwaves range can be
27 used to monitor the salting cheese process.

28

29 **Keywords:** cheese, brining, permittivity, sodium chloride

30

31 1. Introduction

32 Cheese production is a complex process, which involves different units operations and
33 biochemical transformations (Walstra et al., 2006). The salting treatment is an important
34 operation in the manufacture of cheese; the salt normally used is sodium chloride (NaCl),
35 which serves to many purposes: to improve the flavour, texture and colour of cheese, but
36 it is also meant to kill off the starter cultures used in the cheese-making process,
37 preventing further growth and the development of acid. In addition, it helps to reduce the
38 water activity that prevents the growth of undesirable microorganisms (Aly & Galal,
39 2002; Abdalla and El-Zubeir, 2006).

40 Cheese consists on a matrix of protein, with fat and water components as an emulsion
41 (Dickinson & Miller, 2001; Green, 1997). Essentially the matrix is composed of
42 overlapping and cross-linked strands, mainly composed of casein aggregates; the integrity
43 of the matrix is maintained by various intra- and inter-aggregate hydrophobic and
44 electrostatic attractions (Fox et al., 1996). The addition of salt in cheese affects its
45 functional characteristics (Floury et al., 2009), principally in the protein hydration and in
46 the modification of the water binding capacity of casein within the cheese matrix,
47 affecting the stability properties (Mamdouh, 2012).

48 The salting process is commonly performed using brines, where the control of NaCl
49 content is very important to determine the physicochemical characteristics of cheese. It is

50 necessary to obtain a technique that allows determining, in a rapid and easy way, the salt
51 content in cheese; this still remains as an unsolved problem in the food industry
52 (Chevalier et al., 2006). In this sense, microwave dielectric spectroscopy could be an
53 interesting technique to monitor salting process. This technique has been already used for
54 monitoring the salting meat process (Castro-Giráldez et al., 2010). Some authors
55 determined the dielectric properties of salted and unsalted butter over the MW frequency
56 range (Ahmed et al., 2007; Shiinoki et al., 1998). Moreover, this technique have been
57 already used in the analysis of cheese composition and cheese maturity (Ball et al., 1996;
58 Cevoli et al., 2012; Everard et al., 2006; Green, 1997; Herve et al., 1998; Kubiš et al.,
59 2001; Smith et al., 2011).

60 Complex permittivity (ϵ^*) describes the behaviour of a food when subjected to an external
61 electric field (Nelson & Datta, 2001). The two components of complex permittivity (ϵ'
62 and ϵ'') are called dielectric constant and loss factor, respectively. The dielectric constant
63 represents the real part of complex permittivity, and is related to the material ability to
64 store energy; the loss factor represents the imaginary part, which is related to the
65 absorption and dissipation of the electric energy in other kinds of energy, such as thermal
66 or mechanical (Castro-Giráldez et al., 2010; Schwan, 1988).

67 In the range of microwaves, the interaction of the electric field with foods produces two
68 main effects, γ -dispersion and ionic conductivity. The first one is caused by dipolar
69 molecules orientation and induction, and it is produced at GHz frequencies (Mohiri et al.,
70 2011). The other important effect in microwave range is ionic conductivity; the
71 application of an electric field causes the vibration of food ions (Metaxas and Meredith,
72 1983) increasing the internal energy of the molecules.

73 The aim of the present study is to obtain and analyse the dielectric properties of cheese
74 during the salting process and to assess the potential use of dielectric spectroscopy to
75 monitor this process.

76 2. Material and Methods

77 2.1 Raw material

78 18 cheeses were proportionated by Formatgeria Granja Rinya S.L. factory (Valencia,
79 Spain). All the cheeses were of the fresh Tronchon type and they were conducted under
80 the same pressing conditions and with identical compositional characteristics. The
81 cheeses were provided unsalted and just after the pressing operation. Samples were
82 obtained from the center of each cheese. Brine was prepared with sodium chloride (PRS-
83 codex, Panreac® Química SA, Barcelona, Spain) and distilled water.

84 2.2 Salting Operation

85 Salting treatment was carried out by using 25 % (w/w) sodium chloride brine at 4°C. 72
86 cheese cylinders (30 mm diameter and 10 mm thick) were cut. Samples were immersed
87 in a vessel containing the osmotic solution with continuous stirring, for 0, 10, 20, 30, 40,
88 50, 60, 90, 120, 180, 240, 360, 480, 720, 900 and 1440 min; later they were equilibrated
89 in an isothermal chamber at 4°C for 24 hours (equilibrated samples), in order to remove
90 the water and salt profiles and estimate the sorption isotherm (relation between moisture
91 and water activity in an isothermal and equilibrated system), following the methodology
92 of Castro-Giraldez et al. (2010).

93 Mass was determined by using a Mettler Toledo Balance (± 0.0001 g) (Mettler-Toledo,
94 Inc., USA). The surface water activity was determined by a dew point hygrometer
95 Decagon (Aqualab® series 3TE) with precision ± 0.003 . Also the water activity of the
96 osmotic solution was measured at each osmotic time. Measurements were done in
97 structured samples (not minced), thus the obtained a_w is considered to be the surface a_w

98 (Velázquez-Varela et al., 2014). The analysis of water content was adapted from the
99 method of AOAC (Method 934.06, 2000) where moisture of equilibrated samples were
100 obtained by using a vacuum oven, 24 hours after the treatment, and moisture of non-
101 equilibrated samples were estimated using weight balances. Two to three grams of grated
102 cheese were accurately weighed into a tared tempered glass dish and the cheese spread
103 into a uniform layer. The sample was placed in a vacuum oven at room temperature and
104 the pressure in the oven reduced to 100 mbar. The temperature was then slowly increased
105 to 65°C, and the samples were maintained at this temperature for 14 d. The dried samples
106 were then removed from the oven, cooled to room temperature in a desiccator, and
107 weighed. The volume was determined by image analysis (Canon® Power Shot SX210
108 IS), using Adobe Photoshop® software (Adobe Systems Inc., San Jose, CA, USA) in
109 order to determine the diameter and the thickness of the samples by a non-destructive
110 method. Ion content was carried out by means of an ion chromatograph (Methrom Ion
111 Analysis, Herisau, Switzerland), using an universal standard column (Metrosep C2–150,
112 4.0 x 150 mm) along with an eluent composed of tartaric acid (4.0 mmol/L) and
113 dipicolinic acid (0.75 mmol/L), equipped with electronic detectors. In every case, the
114 cheese samples were previously homogenized at 4200 rpm in an Ultraturrax T25 for 5
115 min and centrifuged (J.P. Selecta S.A., Medifriger-BL, Barcelona, Spain) at 8000 rpm for
116 20 min at 4°C. Afterwards, 1 mL of supernatant was diluted with Milli®-Q water in a 250
117 mL Erlenmeyer flask. The clarified extract was filtered through a 0.45 µm Millipore filter;
118 15 mL was used to analyse the anion content. Measurements were made in duplicate.

119 2.3 Permittivity measurement

120 Permittivity was measured by using an Agilent 85070E Open-ended coaxial probe
121 connected to an Agilent E8362B vector network analyser. The software of the network
122 analyser calculates the dielectric constant and loss factor. The system was calibrated by

123 using three different types of loads: open circuit (air), short-circuit and 4°C distilled
124 water. The dielectric properties were measured by attaching the probe to the surface of
125 the samples at 4°C from 500 MHz to 20 GHz. The mean values of at least three replicates
126 of the cheese samples are reported in this article.

127 2.4 Statistical analysis

128 Statistical analysis was carried out with the Statgraphic Centurion ®, version 16.2.04
129 (Statpoint Technologies, Inc, U.S.A.). Multiple regression factorial analyses were
130 performed in order to find significant interaction among dielectric constant at 20 GHz
131 with the number of water molecules and the water fluxes. Moreover, multiple regression
132 factorial analyses were carried out in order to find significant interaction among
133 conductivity at 0.5 GHz with the number of Calcium, Chloride, and Sodium molecules
134 and the Sodium chloride fluxes.

135 3. Results and Discussion

136 Cheese samples were measured just after the salting treatment (non-equilibrated samples)
137 and after 24 hours of repose (equilibrated samples). 24 hours of repose is considered
138 enough time to reach the equilibrium in the whole sample, demonstrated by the salt
139 profiles disappearance (Velázquez-Varela et al., 2014). During the repose time, samples
140 were on disposable sample cups, closed with parafilm®, and preserved in an isothermal
141 chamber in order to isolate the samples and to reach equilibrium. The salting process in
142 brine produces a movement of ions (Na^+ and Cl^-) from the brine into the cheese, and
143 simultaneously there is an outflow of water and calcium from the cheese to the solution,
144 promoted by gradients of chemical potentials. In cheese, the mass water loss is higher
145 than the amount of salt incorporated; as a result, there is a loss of weight in the cheese
146 (Velázquez-Varela et al., 2014).

147 The desorption profiles of non-equilibrated samples and the desorption isotherms of
148 equilibrated samples were obtained in order to estimate the moisture of the surface (x_w^{surf})
149 following the procedure described by Traffano-Schiffo et al., 2014 (see Figure 1) .
150 Surface water activity is a superficial measurement and moisture is an average value of
151 the whole sample, as was explained in the Material and Methods section. Equilibrated
152 samples did not have moisture profiles; therefore, equilibrated isotherm represents the
153 real relation between moisture and water activity. Thus, the equilibrated isotherm is an
154 excellent tool to obtain, with the surface water activity of non-equilibrated samples, the
155 surface moisture of those samples (samples with moisture profiles). In figure 1, it is also
156 possible to observe an increase in surface water activity caused by the internal fluxes that
157 occur in cheese during the repose time.

158 The water and salt fluxes were calculated as follows:

$$159 \quad J_w^{\text{oh}} = \frac{-\Delta M_w^{\text{oh}} \cdot M_0}{\Delta t \cdot S \cdot Mr_w} \quad (\text{Equation 1})$$

$$160 \quad J_{\text{NaCl}}^{\text{oh}} = \frac{-\Delta M_{\text{NaCl}}^{\text{oh}} \cdot M_0}{\Delta t \cdot S \cdot Mr_{\text{NaCl}}} \quad (\text{Equation 2})$$

161

162 Where, J is the flux ($\text{mol s}^{-1} \text{m}^{-2}$), ΔM represents the mass variation (dimensionless), M_0
163 is the initial mass of the sample (g), Δt is the process time, S is the area of the sample
164 during treatment (m^2), Mr is the molecular weight of the compound (18 g mol^{-1} for water
165 and 58.4 g mol^{-1} for sodium chloride), and the subscripts “w” and “NaCl” represent water
166 and sodium chloride, respectively. The superscript “Oh” represents the non-equilibrated
167 samples.

168 Figure 2 shows the water and NaCl molar fluxes. These fluxes decrease with time, caused
169 by a decrease of the engine of the transport (chemical potential) throughout the treatment
170 (Velázquez-Varela et al., 2014).

171 In order to understand the electric behaviours affected by water and ions molecules in the
172 sample surface, it is necessary to calculate its content in the surface. The surface water
173 mass fraction was calculated by using the surface water activity of non-equilibrated
174 samples, from the equilibrated isotherm. The surface sodium chloride mass fraction was
175 estimated by using the surface water activity following the model of Fito et al. (2011).
176 Figure 3 shows the evolution of the overall value of water mass fraction and the water
177 mass fraction in the surface, both expressed in liquid phase (z_w), throughout the salting
178 treatment (non-equilibrated samples).

179 In order to understand the permittivity, measured by a coaxial probe in contact with the
180 surface of the sample, it is necessary to know its penetration depth (D_p), which was
181 calculated as follows (Metaxas & Meredith, 1983):

$$182 \quad D_p = \frac{\lambda_0}{2\pi\sqrt{2\varepsilon'}} \left[(1 + \tan^2 \delta)^{1/2} - 1 \right]^{-1/2} \quad (\text{Equation 3})$$

183 where λ_0 is the wavelength, being 60 cm at 0.5 GHz and 1.5 cm at 20 GHz; $\tan \delta$ is the
184 loss tangent ($\tan \delta = \frac{\varepsilon''}{\varepsilon'}$).

185 Figure 4 shows a scheme of cheese sample in contact with the microwave probe,
186 indicating the penetration depth of the electric field at 20 GHz and 0.5 GHz. At frequency
187 of 20 GHz the predominant effect is the γ -dispersion, explained as the induction and
188 orientation of the dipolar molecules. The most important dipolar molecule is water,
189 therefore water status can be studied in this spectral range. The two properties that
190 influence this dispersion are the number of water molecules and their mobility (Talens et
191 al., 2016, Traffano-Schiffo et al 2015). During the salting process, the microwave
192 penetration volume (MPV) reaches a minimum quantity of water molecules before the
193 rest of the sample volume; at this point, the dielectric signal will be affected by the loss
194 of water molecules. When the steady state is reached at the surface, the sample will

195 continue loosing water molecules from the rest of the sample volume, generating a water
196 flux across the MPV, changing the mobility of the water molecules and, therefore, the
197 water mobility has the major effect of the dispersion.

198 Figure 4 shows, at the bottom of the scheme, the effect at 500 MHz. Ionic conductivity is
199 the major contribution to the dielectric losses. Ionic conductivity does not affect the
200 electric storage because no molecular orientation occurs by the electric field. The
201 conductivity value changes with the quantity and mobility of the electrolytes (chloride,
202 sodium and calcium). The penetration index reaches the whole sample, indicating an
203 average value of the state of the ions. The saturation value of the liquid phase is rapidly
204 reached, taking a constant value of the ions molecules. Nevertheless, the fluxes of water
205 and calcium maintain the fluxes of sodium and chloride (Velázquez-Varela et al., 2014),
206 changing the value of conductivity.

207 Figure 5 shows the dielectric constant and the loss factor spectra of the samples just after
208 the salting treatment. It is also important to analyse the water molecules in the sample
209 surface by dielectric spectroscopy. For this purpose, there were calculated the number of
210 water and ions molecules by using the surface mass fraction in the case of water, and the
211 ions mass fraction of the sample in the case of ions (Equation 4).

$$212 \quad N_j = \frac{X_j N_A}{M_{rj}} \quad (\text{Equation 4})$$

213 where: N represents the molecules ($\text{molecules} \cdot \text{g}_{\text{dm}}^{-1}$), X_j is the surface moisture
214 expressed by solid matrix (neither water nor ions) in the case of water ($\text{g}_w \cdot \text{g}_{\text{SM}}^{-1}$) and ion
215 mass fraction (sodium, calcium and chloride) expressed by solid matrix (neither water
216 nor ions) in the case of ions ($\text{g}_i \cdot \text{g}_{\text{SM}}^{-1}$), N_A is the Avogadro constant ($6.022 \cdot 10^{23} \text{ mol}^{-1}$)
217 and M_{rj} is the molecular weight, being $18 \text{ g} \cdot \text{mol}^{-1}$ for water, $22.99 \text{ g} \cdot \text{mol}^{-1}$ for sodium,
218 $40.08 \text{ g} \cdot \text{mol}^{-1}$ for calcium and $35.45 \text{ g} \cdot \text{mol}^{-1}$ for chloride.

219 The interaction of the electric field with cheese in γ -dispersion is affected by the amount
220 of water molecules and its motion. Multiple regression factorial analyses were made in
221 order to find significant interaction amongst the dielectric constant at 20 GHz with the
222 number of water molecules and the water fluxes throughout the salting treatment. This
223 analysis showed that the relation between dielectric constant at 20 GHz and the number
224 of water molecules is significant ($p<0.05$) until the surface saturation phenomena of
225 cheese liquid phase, i.e. during the first 180 minutes of treatment. The analysis also
226 showed that there exists a very significant ($p<0.01$) relation between the dielectric
227 constant at 20 GHz and the water flux from 90 minutes until the end of the treatment.
228 Figure 6 presents the relation between the dielectric constant at 20 GHz and the number
229 of the surface water molecules in non-equilibrated samples. It is possible to observe that
230 there exists a linear relationship between both variables at the beginning of the salting
231 treatment as was explained before; thus, the surface water molecules can be obtained by
232 measuring the dielectric constant at 20 GHz when the surface is in an unsteady state.
233 When the system reaches the steady state on the surface, the amount of water molecules
234 is constant but the water is still flowing throughout this volume, coming from the middle
235 of the samples and changing the motion of the water molecules in the microwave
236 penetration volume. Therefore, the capability of water molecules to be induced and
237 orientated changes with this water flux. In figure 7, it is possible to observe that the
238 dielectric constant at 20 GHz is related very significantly ($p<0.01$) with the water flux
239 after 90 minutes of treatment, at which the steady state is reached by the system.
240 Therefore, the dielectric model to predict water losses has to be non-linear, explaining not
241 only the water losses in the microwave penetration volume, but also in the overall product,
242 because when the penetration microwave volume reaches the steady state, the model
243 predicts the water flux.

244 The conductivity of the sample (σ) was calculated at 0.5 GHz as follows:

$$245 \quad \sigma = \varepsilon'' \varepsilon_0 \omega \quad (\text{Equation 5})$$

246 where: ω is the angular frequency and the ε_0 is the dielectric constant in vacuum
247 ($\varepsilon_0=8.8542 \cdot 10^{-12} \text{ F}\cdot\text{m}^{-1}$).

248 The conductivity at 0.5 GHz was represented with regard to the amount of ions molecules
249 of the sample estimated by equation 4 (Figure 8). It is important to highlight that, at this
250 frequency, the microwave penetration depth covers most of the sample; thus, the
251 conductivity can be considered an overall value of the whole sample.

252 In figure 8, at first 60 minutes of salting, a linear relation between the accumulation of
253 the electrolytic molecules and ionic conductivity can be appreciated, being very
254 significant ($p<0.01$). When saturation is achieved in the liquid phase, the conductivity
255 decreases. So, in the first period (the first 60 minutes) conductivity changes very
256 significantly with the accumulation of electrolyte molecules, but from this time it is no
257 longer significant.

258 After 60 minutes, when the relation between the conductivity and the accumulation of
259 electrolyte molecules is non significant, the conductivity begins to decrease. This
260 decrease is promoted by a decrease in the mobility of the electrolytic molecules in the
261 liquid phase, sodium, chloride and calcium. The cause of mobility in this period of salting
262 is the motion produced by the ion fluxes, where the fluxes are induced by the absorption
263 of sodium and chloride ions in the casein matrix and by the loss of calcium from the
264 casein structure (Velázquez-Varela et al., 2014). In Figure 9 it can be observed, after 60
265 minutes, a very significant ($p<0.01$) linear relationship between the sodium chloride flux
266 and the ionic conductivity until the end of the process. Therefore, it is possible to predict

267 the level of salting and thus the structural changes in the casein with the measurement of
268 the conductivity at 500 MHz

269 4. **Conclusions**

270 The non-linear model proposed by using the dielectric constant at 20 GHz explains not
271 only the water losses in the MPV, but also in the overall product, because when the MPV
272 reaches the steady state, the model predicts the water flux. In case of salting prediction,
273 the ionic conductivity estimated at 500 MHz explains the level of salting, and for bigger
274 samples, the variable can also be explained, because it is related not only to the
275 accumulation of the electrolyte molecules, but also to the fluxes.

276 Therefore, coupled measurements of dielectric properties at 20 GHz and 500 MHz can
277 predict the chemical species involved in cheese salting process, and its structural changes.

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285 6. **References**

286 Ahmed, J., Ramaswamy, H. S., & Raghavan, V. G. S. (2007). Dielectric properties of
287 butter in the MW frequency range as affected by salt and temperature. *Journal of Food*
288 *Engineering*, 82, 351-358.

289 AOAC, 2000. AOAC, Official Methods of Analysis (17th ed.) Association of Official
290 Analytical Chemists, Arlington, VA (2000).

291 Ball, J. A. R., Horsfield, B., Holdem, J. R., Keam, R. B., Holmes, W. S., & Green, A.
292 (1996). Cheese curd permittivity and moisture measurement using a 6-port reflectometer.
293 In Asia Pacific Microwave Conference, New Delhi.

294 Castro-Giráldez, M., Fito, P.J., & Fito, P. (2010). Application of microwaves dielectric
295 spectroscopy for controlling pork meat (*Longissimus dorsi*) salting process. *Journal of*
296 *Food Engineering*, 97, 484-490.

297 Cevoli, C., Ragni, L., Gori A., Berardinelli A., & Fiorenza C. M. (2012). Quality
298 parameter assessment of grated Parmigiano-Reggiano cheese by waveguide
299 spectroscopy. *Journal of Food Engineering*, 113, 201-209.

300 Chevalier, D., Ossart, F., & Ghommidh, C. (2006). Development of a nondestructive salt
301 and moisture measurement method in salmon (*Salmo salar*) fillets using impedance
302 technology. *Food Control*, 17, 342–347.

303 Dickinson, E., & Miller, R. (2001). Food Colloids: Fundamentals of Formulation. The
304 Royal Society of Chemistry, UK.

305 Everard, C.D., Fagan, C.C., O'Donnell, C.P., O'Callaghan, D.J., & Lyng, J.G. (2006)
306 Dielectric properties of process cheese from 0,3 to 3 GHz. *Journal of Food Engineering*,
307 75, 415-422

308 Fito, P, Fito, P.J, Betoret, N., Argüelles A. & Chenoll, C. (2011) Thermodynamic
309 approach to equilibrium isotherms in salted structured food. *Journal of Food Process*
310 *Engineering*, 34, 623–638

311 Flourey, J., Camier, B., Rousseau, F., Lopez, C., Tissier, J. P., & Famelart, M. H. (2009).
312 Reducing salt level in food: Part 1. Controlled manufacture of model cheese systems and
313 their structure-texture relationships. *LWT – Food Science and Technology*, 42, 1611–
314 1620.

315 Fox, P.F., O'Connor, T.P., McSweeney, P.L.H., Guinee, T.P., & O'Brien, N.M. (1996).
316 Cheese: physical, biochemical, and nutritional aspects. *Advances. Food Nutrition*
317 *Research*, 39, 163-328.

318 Green, A. D. (1997). Measurements of the dielectric properties of Cheddar cheese.
319 *Journal of Microwave Power and Electromagnetic Energy*, 32, 16–27.

320 Herve, A. G., Tang, J., Luedecke, L., & Feng, H. (1998). Dielectric properties of cottage
321 cheese and surface treatment using microwaves. *Journal of Food Engineering*, 37, 309–
322 410.

323 Kubiš, I., Krivanek, I., & Gajdusek, S. (2001). The relationships between the chemical,
324 dielectric and sensory properties of Edam cheese during ripening. *Czech Journal of Food*
325 *Science*, 19, 85–89.

326 Mamdouh, E.B. (2012). Salt in Cheese: A Review. *Current Research in Dairy Sciences*,
327 4, 1-5.

328 Metaxas A. C. & Meredith R. J. (1993). *Industrial Microwave Heating*. UK: IEE Power
329 Engineering Series 4. Peter Peregrinus Ltd. (Chapter 2 and 4)

330 Mohiri, A., Burhanudin, A. Z., & Ismail, I. (2011). Dielectric Properties of Slaughtered
331 and Non-slaughtered Goat Meat. In IEEE International RF and Microwave Conference.
332 Malaysia.

333 Nelson, S.O. & Datta, A.K. (2001). Dielectric properties of food materials and electric
334 field interactions. In A.K. Datta & R.C. Anantheswaran (Eds), *Handbook of Microwave*
335 *Technology for Food Applications*. New York: Marcel Dekker Inc.

336 Payne, M.R., & Morison, K.R. (1999). A multi-component approach to salt and water
337 diffusion in cheese. *International Dairy Journal*, 9, 887–894.

338 Schwan, H. P. 1988. Dielectric spectroscopy of biological cells, *Special Issue on*
339 *Electricity and Biophysics, Ferroelectrics*, 86, 205-223.

340 Shiinoki, Y., Motouri, Y. & Ito. K. (1998). On-line Monitoring of Moisture and Salt
341 Contents by the Microwave transmission Method in a Continuous Salted Butter-making
342 Process. *Journal of Food Engineering*, 38, 153-167.

343 Smith, J., Carr, A., Golding, M., Reid, D., & Zhang, L. (2011) Assessing the use of
344 dielectric spectroscopy to analyse calcium induced compositional and structural changes
345 in a model cheese. Selection of 11th International Congress on Engineering and Food
346 (ICEF11). *Proceedings Food Science*, 1, 1833 – 1840

347 Talens, C., Castro-Giraldez, M., Fito, P.J. (2016). Study of the effect of microwave power
348 coupled with hot air drying on orange peel by dielectric spectroscopy. *LWT-Food Science*
349 *and Technology*, 66, 622-628

350 Traffano-Schiffo, M.V., Castro-Giraldez, M., Fito P.J., Balaguer, N. (2014).
351 Thermodynamic model of meat drying by infrared thermography. *Journal of Food*
352 *Engineering*, 128, 103-110

353 Traffano-Schiffo, M.V., Castro-Giraldez, M., Colom, R.J., Fito P.J. (2015). Study of the
354 application of dielectric spectroscopy to predict the water activity of meat during drying
355 process. *Journal of Food Engineering*, 166, 285-290

356 Velázquez-Varela, J., Fito, P.J. & Castro-Giráldez, M. (2014). Thermodynamic analysis
357 of salting cheese process. *Journal of Food Engineering*, 130, 36-44.

358 Venkatesh, M.S. & Raghavan, G.S.V. (2004). An Overview of Microwave Processing
359 and Dielectric Properties of Agri-food Materials. *Biosystems Engineering*, 88, 1-18

360 Walstra, P., Wouters, J. T. M. & Geurts, T. J. (2006). Dairy science and technology. (2nd
361 ed.). Florida: Taylor & Francis Group, LLC.

362 Wang, S., Tang, J., Johnson, J. A., Mitcham, E., Hansen J. D., Hallman, G., Drake, S.R,
363 & Wang, Y. (2003). Dielectric Properties of Fruits and Insect Pests as related to Radio
364 Frequency and Microwave Treatments. *Biosystems Engineering*, 85, 201–212.

365

366

367

368 **FIGURE CAPTION**

369 Figure 1. Desorption profile of non-equilibrated (●) and desorption isotherm of
370 equilibrated (◆) samples.

371 Figure 2. Evolution of water (■) and NaCl (◆) fluxes throughout the osmotic treatment.

372 Figure 3. Evolution of the surface water mass fraction in the liquid phase with regard to
373 the treatment time, where: (▲) represents the value of the sample surface and (●)
374 represents the overall value.

375 Figure 4. Scheme of the dielectric properties measurements of cheese samples throughout
376 the salting treatment.

377 Figure 5. Evolution of the dielectric constant and loss factor spectra of the samples
378 throughout the treatment (non-equilibrated samples), where: (—) 0 min; (—) 10 min; (- -
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380 (—) 3 h; (- - -) 4 h; (- · -) 6 h; (···) 8 h; (- · - · -) 12 h; (- · - · -) 15 h; (- · - · -) 24 h.

381 Figure 6. Relation of the dielectric constant at 20 GHz with regard to the number of
382 surface water molecules in solid matrix throughout the salting treatment.

383 Figure 7. Evolution of the water flux with regard to the dielectric constant at 20 GHz.

384 Figure 8. Conductivity at 0.5 GHz with regard to the amount of ions molecules of the
385 sample.

386 Figure 9. Flux of sodium chloride with regard to conductivity.

387

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360 Wang, S., Tang, J., Johnson, J. A., Mitcham, E., Hansen J. D., Hallman, G., Drake, S.R,
361 & Wang, Y. (2003). Dielectric Properties of Fruits and Insect Pests as related to Radio
362 Frequency and Microwave Treatments. *Biosystems Engineering*, 85, 201–212.

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365 **FIGURE CAPTION**

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367 Figure 2. Evolution of water (■) and NaCl (◆) fluxes throughout the osmotic treatment.

368 Figure 3. Evolution of the surface water mass fraction in the liquid phase with regard to

369 the treatment time, where: (▲) represents the value of the sample surface and (●)

370 represents the overall value.

371 Figure 4. Scheme of the dielectric properties measurements of cheese samples

372 throughout the salting treatment.

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Figure 1

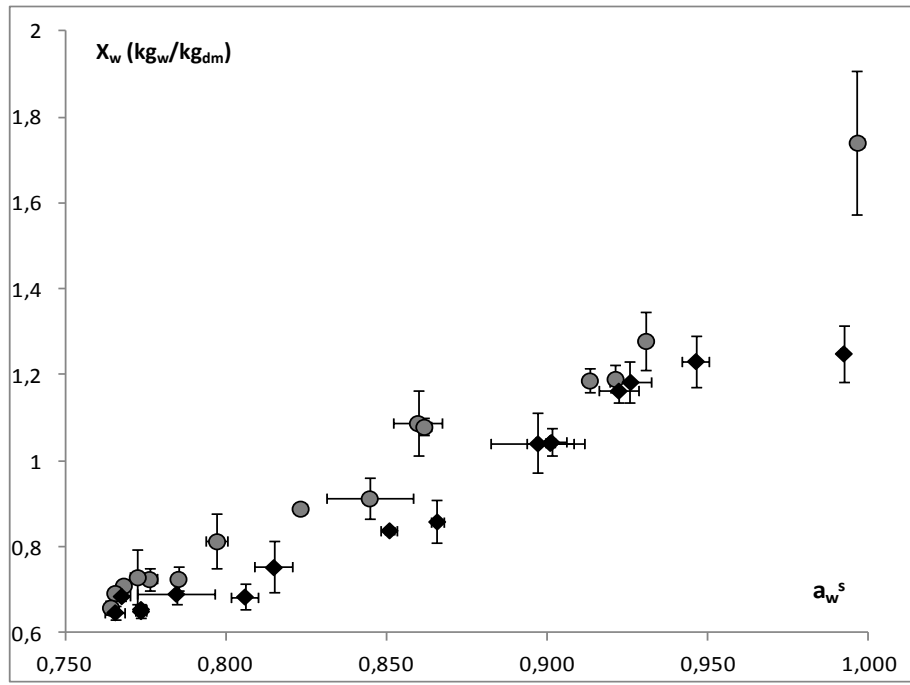


Figure 1.

Figure 2

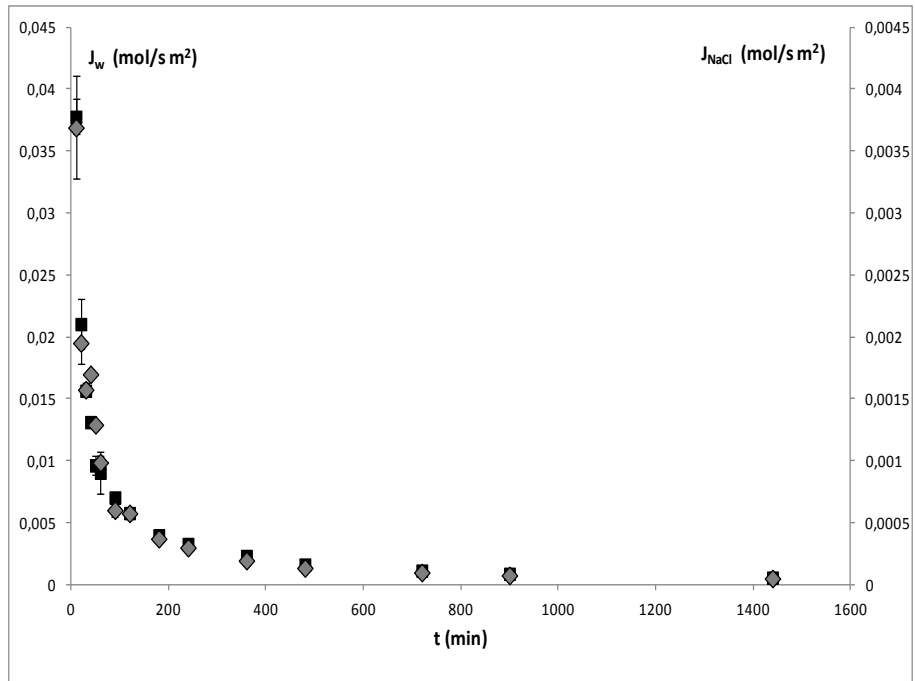


Figure 2.

Figure 3

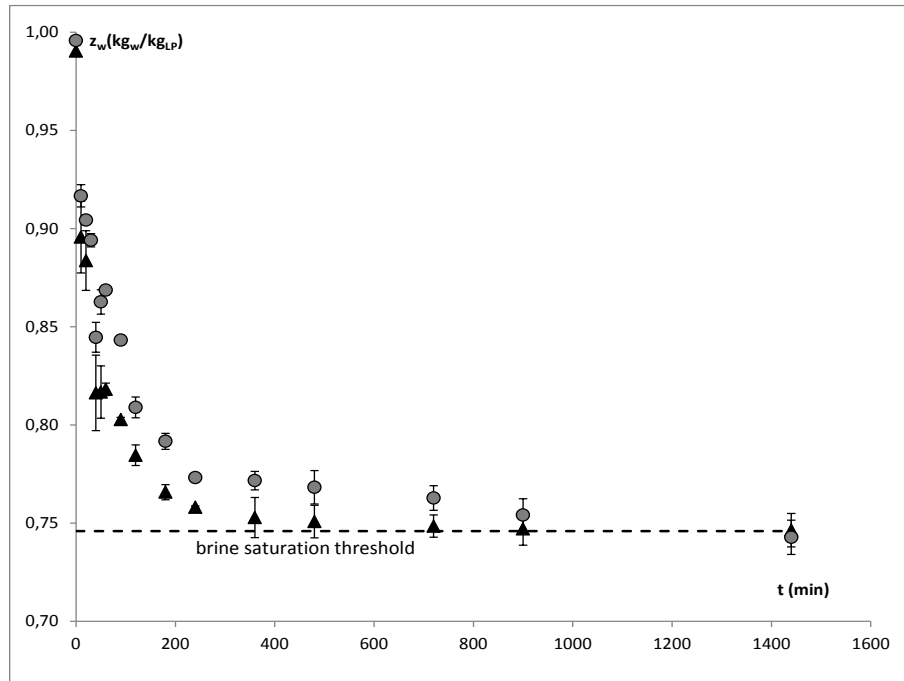


Figure 3.

Figure 4

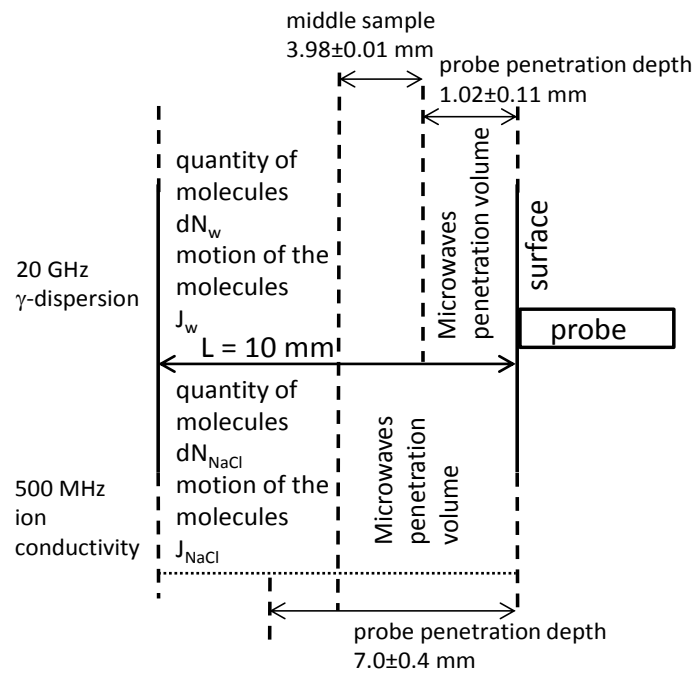


Figure 4.

Figure 5

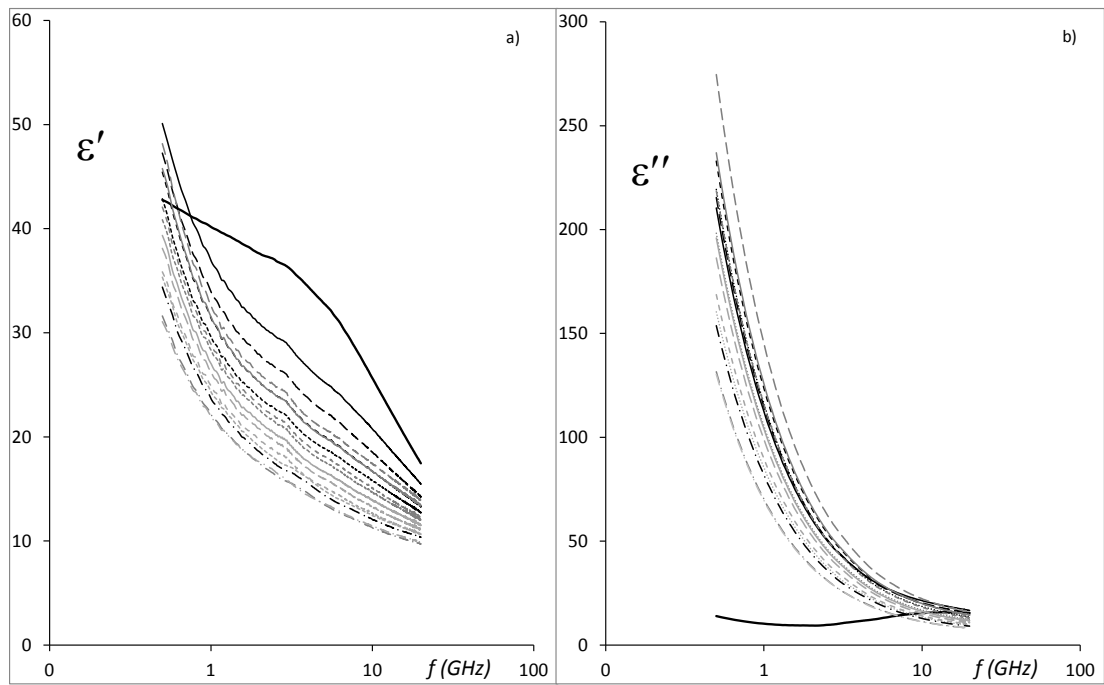


Figure 5.

Figure 6

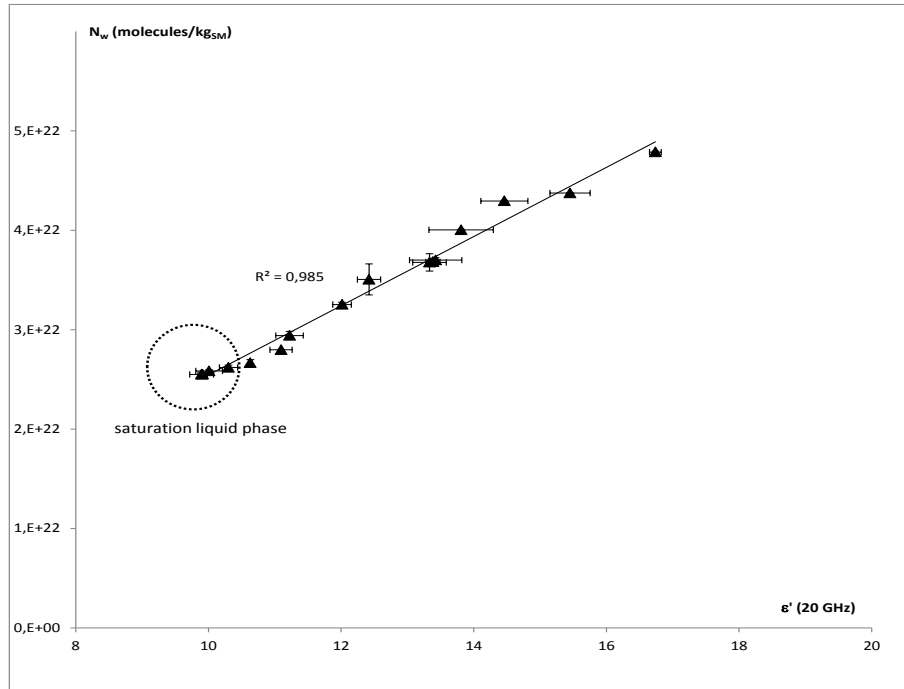


Figure 6.

Figure 7

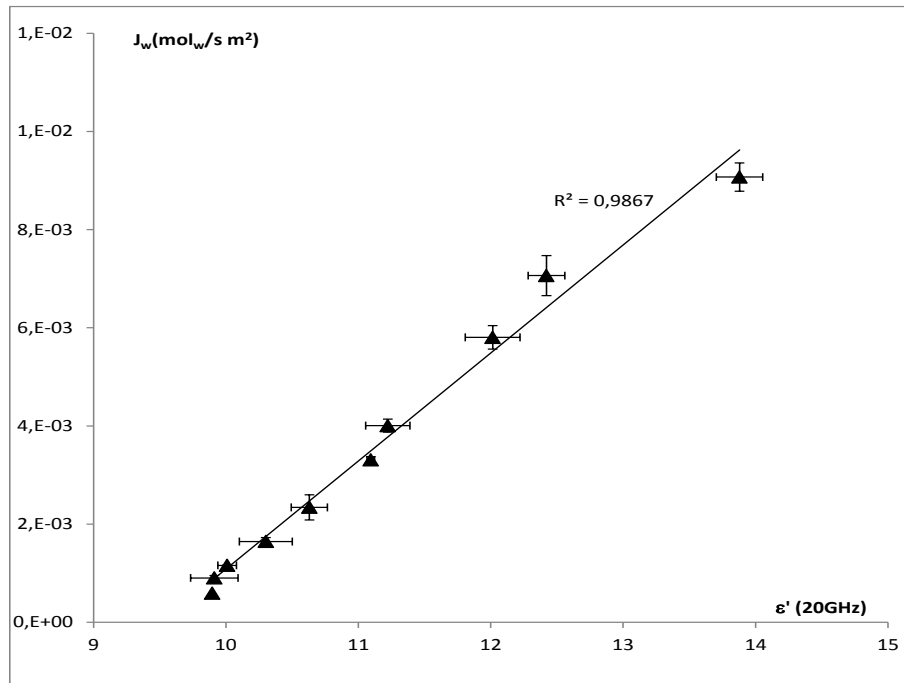


Figure 7.

Figure 8

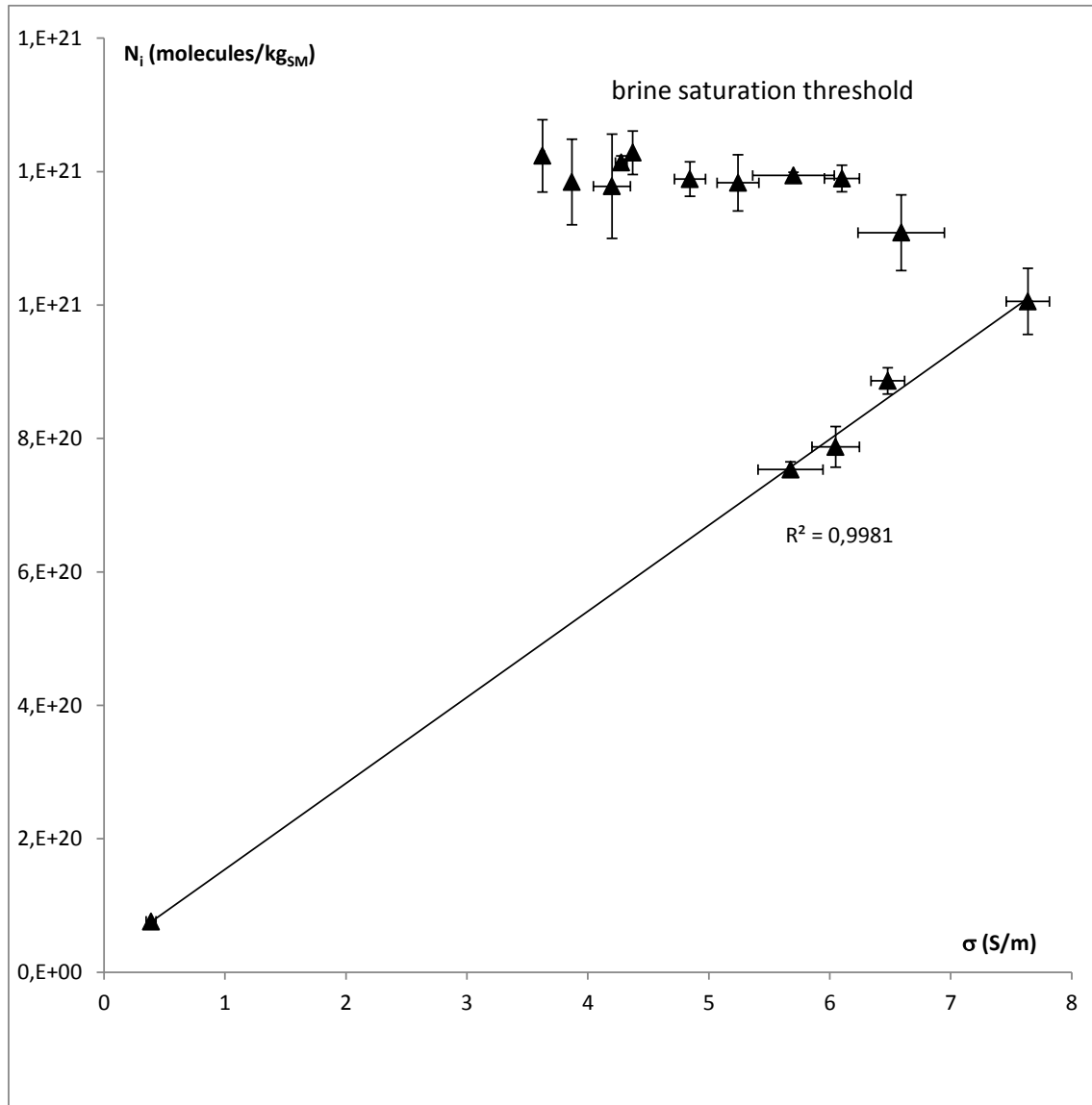


Figure 8.

Figure 9

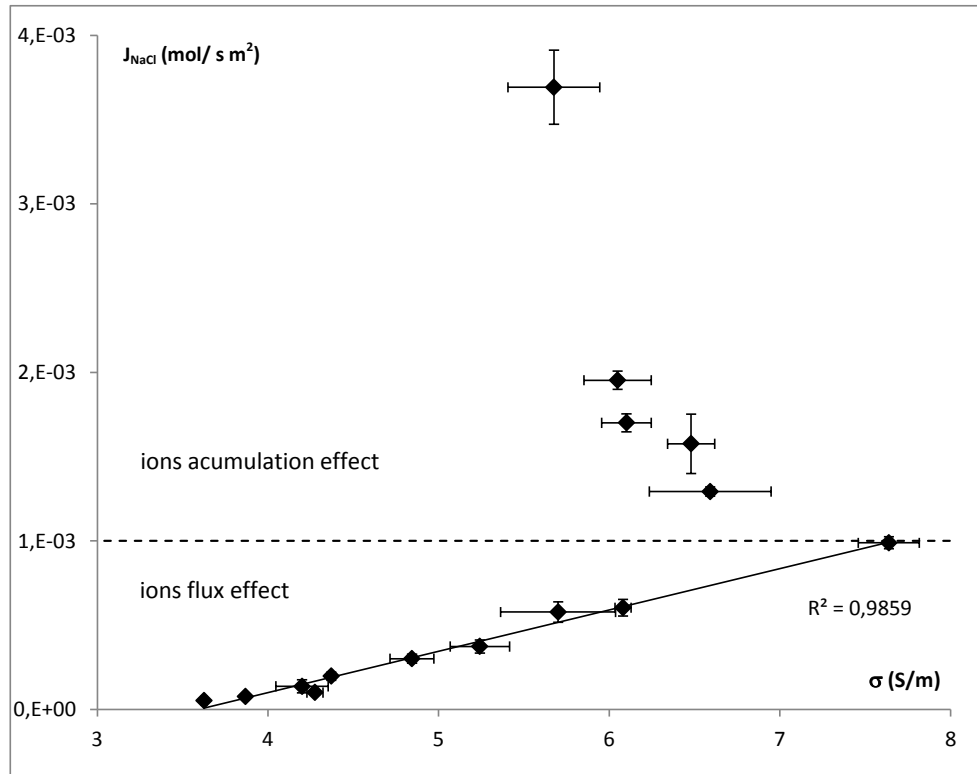


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