

Document downloaded from:

<http://hdl.handle.net/10251/145409>

This paper must be cited as:

Barbarroja-Ortiz, P.; Zornoza-Zornoza, AM.; Aguado García, D.; Borrás Falomir, L.; Alonso Molina, JL. (09-2). A multivariate approach of changes in filamentous, nitrifying and protist communities and nitrogen removal efficiencies during ozone dosage in a full-scale wastewater treatment plant. *Environmental Pollution*. 252-B:1500-1508.
<https://doi.org/10.1016/j.envpol.2019.06.068>



The final publication is available at

<https://doi.org/10.1016/j.envpol.2019.06.068>

Copyright Elsevier

Additional Information

1 **A multivariate approach of changes in filamentous, nitrifying and protist**
2 **communities and nitrogen removal efficiencies during ozone dosage in a full-scale**
3 **wastewater treatment plant.**

4 **Authors**

5 Paula Barbarroja^{a*}, Andrés Zornoza^a, Daniel Aguado^a, Luis Borrás^b, José Luis Alonso^a.

6 **Affiliations**

7 ^aInstituto de Ingeniería del Agua y Medio Ambiente, Universitat Politècnica de València.
8 Camino de vera s/n, 46022, Valencia, Spain

9 ^bDepartamento de Ingeniería Química, Universitat de València, Avda de la Universidad
10 s/n, 46100, Burjassot, Valencia.

11 *Corresponding author. Tel. +34 963877090; Fax +34 963877090. E-mail address:
12 pbarbarroja@gmail.com, paubaror@iiama.upv.es

13 **Abstract:**

14 The application of low ozone dosage to minimize the problems caused by filamentous
15 foaming was evaluated in two bioreactors of an urban wastewater treatment plant.
16 Filamentous and nitrifying bacteria, as well as protist and metazoa, were monitored
17 throughout a one-year period by FISH and conventional microscopy to examine the
18 effects of ozone application on these specific groups of microorganisms. Multivariate
19 data analysis was used to determine if the ozone dosage was a key factor determining the
20 low carbon and nitrogen removal efficiencies observed throughout the study period, as
21 well as to evaluate its impact on the biological communities monitored. The results of
22 this study suggested that ozonation did not significantly affect the COD removal
23 efficiency, although it had a moderate effect on ammonia removal efficiency.
24 Filamentous bacteria were the community most influenced by ozone (24.9% of the

25 variance explained by ozone loading rate), whilst protist and metazoa were less affected
26 (11.9% of the variance explained). Conversely, ozone loading rate was not a factor in
27 determining the nitrifying bacterial community abundance and composition, although this
28 environmental variable was correlated with ammonia removal efficiency. The results of
29 this study suggest that different filamentous morphotypes were selectively affected by
30 ozone.

31 **Keywords:**

32 Activated sludge, Ozonation, Nitrification, Filamentous bacteria, Multivariate
33 analysis.

The results of our study suggest that the structure of microbial communities significantly changed with ozone addition and different filamentous morphotypes are selectively affected.

34 **1. Introduction**

35 Bulking and foaming problems are frequent causes of effluent quality loss in
36 wastewater treatment plants (WWTPs) with activated sludge (AS) systems. These
37 episodes, caused by the proliferation of filamentous bacteria in the aerated tank, affect
38 the sludge settling properties (Jenkins et al., 2004), producing the occurrence of residual
39 solids in the secondary effluent. When problems became recurrent or chronic the
40 excessive growth of filamentous bacteria can be controlled by non-specific methods such
41 as continuous low ozone addition (Van Leeuwen and Pretorius, 1988). However, it has
42 been suggested that the effectiveness of oxidative treatments is regarded to cell membrane
43 composition and its permeability, particularly in bulking systems where hydrophobic
44 filaments are abundant (Seka et al., 2001).

45 Ozonation is a particularly interesting technology for wastewater treatment. The
46 strong oxidation potential of ozone contributes to solubilisation of suspended solids (SS)
47 and extracellular polymeric substances (EPS), cell lysis, oxidation and mineralization of
48 organic matter when it is applied to the AS process (Chu et al., 2009). Ozone also reacts
49 with inert solids, oxidizing and transforming them into biodegradable forms. During the
50 sequential decomposition processes produced by the ozonation in AS, organic matter and
51 debris are released from the microbial cells (Komanapalli and Lau, 1996). The leakage
52 of these soluble substrates results in an increase of soluble chemical oxygen demand
53 (sCOD), nitrogen and phosphorous loading rates in the reactor (Demir and Filibeli, 2012;
54 Sui et al., 2011), producing indirect impacts on microorganisms during ozonation
55 (Yapsakli et al., 2010). These potential impacts are in addition to direct effect of
56 ozonation on microorganism activity and mortality rates, which depend on each species
57 and their location in the floc (Böhlher and Siegrist, 2004; Fall et al., 2017). Previous
58 research (Fall et al., 2017; Isazadeh et al., 2014; Yan et al., 2009a) highlights the potential
59 of ozonation to transform the microbial community, thus impacting the overall WWTP
60 performance.

61 Among these issues, there has been a major concern about the effect that ozone
62 treatment can cause in nitrification processes. Nitrification is the oxidation of ammonia
63 and nitrite to nitrate by chemolithotrophic bacteria, mainly ammonia-oxidizing bacteria
64 (AOB) and nitrite-oxidizing bacteria (NOB). These bacterial groups are well known for
65 their slow growth rate and high sensitivity to inhibitory compounds and environmental
66 factors. Negative effects of ozonation on nitrification have been reported by several
67 authors (Gardoni et al., 2011; Naso et al., 2008; Richardson et al., 2009; Romero et al.,
68 2015), but only a few studies provide information about the impact of ozonation on the

69 structure of nitrifying bacterial communities (Chen et al., 2017; Izahadeh et al., 2014;
70 Levén et al., 2016; Yan et al., 2009a). In most of these studies, despite negative effects,
71 the nitrogen elimination ratios reached were enough to ensure good effluent quality,
72 indicating that ozonation and biological nitrogen removal are still compatible
73 technologies (Sui et al., 2014).

74 To date, most of the studies related with ozone treatment, have focused on the
75 optimization of operational conditions and have been conducted on pilot plants over short
76 experimental periods. This was because ozonation in full-scale WWTPs is both a
77 challenging operational process and energy intensive (Semblante et al., 2017). These
78 studies have provided a better understanding of the effects that ozonation treatment has
79 on microbial communities and the reactions that take place. Nevertheless, longer ozone
80 applications will likely have a greater influence on AS processes (Dytczak et al., 2008),
81 as the performance of WWTP is the result of a complex interaction between
82 environmental factors, influent conditions and operational variables. Therefore, it is
83 necessary to elucidate which combination of factors contributes to the stability of the
84 nitrification process when combined with ozonation.

85 In this study we propose a new approach to evaluate the impact of long-term ozone dosage
86 on microbial community composition (filamentous bacteria, nitrifying bacteria, protist
87 and metazoa), as well as on biological process efficiency in a full-scale AS system. In
88 order to assess the contribution of environmental variables to the variability observed in
89 microbial community structure and nitrifying performance, distance-based linear models
90 (DISTLM) were used.

91 **2. Materials and methods**

92 2.1. WWTP and ozone system description

93 AS samples were collected from aerated reactors of a full-scale urban WWTP in
94 Spain. The WWTP consists of two independent wastewater treatment lines (CT1, CT2)
95 with A/O configuration, which together treat up to 34157m³ wastewater per day. Each
96 reactor contains a floating ozonation unit installed with the aim to control filamentous
97 foaming, caused mainly by *Gordonia* sp. The sludge ozonation system consisted of an
98 ozone generator (Turboxal, Air Liquide S.A., France) using pure oxygen as the feed gas.
99 This generator distributes ozone through an ejector downward into the aerated basin,
100 producing a gas/liquid emulsion in the mixed liquor. Ozone dosage was regulated in the
101 aerated basins by controlling the oxygen input flow. Both treatment lines were
102 monitored throughout one-year period. The ozone was applied continuously during 180
103 days in CT1. In CT2 an ozonation period of 90 days was followed by non-ozonised period
104 of 45 days, after that ozone was applied 45 more days. During the study period the system
105 was operated with different ozone concentrations, varying dosages from 0 to 47.8 kgO₃/d,
106 which corresponds from 0 to 0.15 gO₃/kgMLSS·h.

107 2.2. Physico-chemical parameters and operational variables

108 Samples from aerated tank, influent and treated effluent were collected every fifteen
109 days during a year. The average characteristics of biological process operational
110 variables, analysed physico-chemical parameters, and treatment efficiencies are shown in
111 the supplementary material. Samples for determining soluble components were
112 immediately filtered after sampling using 0.45 µm glass-fibre filters. All the physico-
113 chemical parameters were examined in accordance with Standard Methods for the
114 Examination of Water and Wastewater (APHA, 2005). BOD samples were determined
115 by respirometry using Oxitop® system. Sludge volumetric index (SVI) was determined
116 according to Jenkins et al. (2004).

117 2.3. Biological variables

118 For fluorescence *in situ* hybridization (FISH) analysis samples of mixed liquor
119 collected every fifteen days throughout a one-year, were fixed within 24 h of sampling in
120 4% paraformaldehyde for 3 h at 4°C, washed with PBS and stored in PBS/Ethanol (50%)
121 at -20 °C until analysis.

122 FISH was used to identify and quantify the nitrifying bacterial community and
123 filamentous bacteria present in AS from both reactors monitored in this study.
124 Hybridization was performed at 46°C according to Daims et al. (2004) with hybridization
125 times of 2-4 h for all oligonucleotide probes used in this study (Supplementary data S2).
126 Hybridized samples were examined with an Olympus BX50 microscope (Olympus, Paris)
127 equipped with 100W mercury high-pressure bulb and set filters U-MWB, U-MWIB and
128 U-MWIG.

129 In order to determine the relative abundance of AOB and NOB thirty images,
130 randomly selected, were captured per sample with camera Olympus DP70 for probe-
131 conferred fluorescence. MATLAB routine developed by Borrás, (2008) was used for
132 image processing and data analysis.

133 The identification and estimation of filamentous bacteria abundance was performed
134 by conventional microscopy before FISH, within 48 h of AS sampling. The identification
135 of morphological and structural characteristics where conducted according to Eikelboom
136 (2000, 2006) and Jenkins et al. (2004). Filamentous bacteria were quantified according
137 to a subjective scoring of filament abundance, ranging from 0 (none) to 5 (abundant)
138 (Eikelboom, 2000).

139 Sludge samples for determining protist and metazoan abundance were analysed
140 within 24 hours sampling. Two replicates of 25 µl were analysed to survey the density of

141 protist and metazoa, by direct counting, using phase contrast microscopy. Four additional
142 replicates were performed for the following organisms: *Epistylis* sp., *Opercularia* sp. and
143 *Carchesium* sp. Different staining procedures, dry silver nitrate (Klein, 1926), silver
144 carbonate impregnations (Fernández-Galiano, 1976) and Flutax-2 staining (Arregui et al.,
145 2003), were carried out to confirm several ciliate species. Two groups of naked amoebae
146 were considered based on their cell size ($> 50 \mu\text{m}$ and $< 50 \mu\text{m}$) (Zornoza, 2017).
147 Organisms were identified by using identification guides (Foissner et al., 1991, 1992,
148 1994, 1995).

149 2.4. Multivariate analysis

150 In order to elucidate which possible factors correlate with nitrifying populations,
151 filamentous bacteria, protist and metazoan dynamics (biological variables), and
152 biological process performance (explained variables) multivariate data analyses was
153 used. Explanatory matrix was constructed with 51 environmental variables (physico-
154 chemical variables of influent, effluent and mixed liquor) and 6 operational parameters
155 of the process. Distance-based linear modelling (DISTLM) procedure was used; this
156 multivariate multiple regression routine uses a resemblance matrix of biological process
157 performance multivariate results and species abundance data to regress against to the
158 explanatory matrix (Anderson et al., 2008). Statistical analyses were performed for both
159 reactors together (CT1 and CT2) as they have the same process and similar operational
160 conditions but different ozone dosage.

161 To avoid background noise, those biological variables with occurrence higher than
162 20% (protists and metazoa) were selected for the multivariate analysis (Pérez-Uz et al.,
163 2010). Prior to analysis, count data were transformed. Square root (nitrifying and
164 filamentous bacteria) and logarithmic (protists and metazoa) transformation were used to

165 down-weight the effect of high occurrence of certain species, whilst still enabling
166 sufficient signal from rarer taxa to be observed (Clark et al., 2014). Environmental
167 variables were log-transformed and normalized prior to multivariate data analyses
168 (euclidean similarity). A Bray-Curtis resemblance matrix was created from the
169 transformed data (Bray and Curtis, 1957). Draftsman plots and correlation matrix were
170 produced to assess the distribution of each variable and to identify multicollinearity. In
171 the pairs of variables with a Pearson's correlation coefficient of 0.85 or larger, one of the
172 variables was excluded from the analysis (Clarke et al., 2014).

173 DISTLM was constructed by using stepwise forward selection procedure. Akaike's
174 information criterion corrected (AIC_c) and Bayesian information criterion (BIC) were
175 selected in order to obtain the most parsimonious model, identifying the simplest
176 reproductions with the greatest explanatory power (Anderson et al., 2008). We use
177 distance-based redundancy (dbRDA) routine to visualize a constrained ordination on the
178 tailored values from the best model (Anderson et al., 2008). Analysed variables were
179 represented as bubble where variable value is represented by bubble size. The strength
180 and direction of the relationship between the explanatory variables and biological
181 variables (Pearson values >0.45) were presented as vectors to visualize the ordination.

182 Six statistical models were employed to evaluate the correlation between
183 environmental variables and the considered explained variables (carbon removal
184 performance, nitrogen removal performance, filamentous bacteria, nitrifying bacteria and
185 protist and metazoan communities). Three of them were constructed to reach a better
186 understanding of possible correlations influencing nitrogen removal efficiencies and
187 nitrifying bacteria population dynamics. The first was used to investigate which
188 explanatory variables influence the observed changes in nitrogen removal efficiencies;

189 the second was used to find the correlations between the nitrifying bacteria populations
190 and the environmental variables; the third examines the relations between the relative
191 abundance of nitrifying bacteria and nitrogen removal efficiency.

192 All multivariate analyses were performed with the statistical software package
193 PRIMER 7 (Clarke and Gorley, 2015) and PERMANOVA+ (Anderson et al., 2008).

194 **3. Results and discussion**

195 3.1. Carbon removal performance

196 The effect of ozonation on effluent quality has been an issue of major concern since
197 ozone affects microbial activity and viability, solubilizes the SS and enhances the
198 biodegradability of inert solids, which results in increased sCOD (Chu et al., 2009). With
199 the aim of observe the biological process removal efficiency separately from the
200 settlement process efficiency, values of sCOD removal efficiency (sCODre) were
201 determined. sCODre ranged from 45% to 90% and 38 to 90% for CT1 and CT2,
202 respectively, and remained above 80% in both reactors during ozonated period.
203 Examination of the relationships between environmental variables and organic matter
204 removal efficiencies did not show any significant correlation between ozone dosage and
205 carbon removal performance (Table 1). The results of our study coincide with those
206 reported by other studies (Lee et al., 2005; Dytczak et al., 2007; Nie et al., 2014), which
207 found that ozonation did not significantly affect the COD removal efficiency (CODre),
208 despite of the wide ranges of ozone concentrations tested. DISTLM showed that 19% of
209 the variation in CODre could be attributed to dissolved oxygen concentration (DO) in the
210 reactor and sludge retention time (SRT). Therefore ozonation did not appreciably affects
211 the COD removal capacity. This might be mainly attributed to the high biodegradability

212 of the organic material released during SS solubilisation that is promptly used as substrate
213 by heterotrophic bacteria (Dytczak et al., 2007; Nagare et al., 2008).

214 3.2. Nitrogen removal performance and nitrifying bacterial community

215 The effluent total nitrogen (TN) and NH_4^+ -N concentrations were similar in
216 treatment lines, ranging from 20 to 54 mg/L (mean value = 35 mg/L) and from 4.4 to 45
217 mg NH_4^+ -N /L (mean value= 27 mg NH_4^+ -N /L) respectively. These values were the
218 result of low nitrogen removal efficiencies (TNre) observed through the studied period.
219 The TNre and NH_4^+ -N removal efficiency (NH_4^+ -Nre) varied from 16 to 57% (mean value
220 = 40%) and 10 to 90% (mean value = 44%) respectively for CT1, whereas these values
221 for CT2 ranging from 20 to 65% (mean value = 44%) and 15 to 63% respectively (mean
222 value = 41%).

223 AOB and NOB were detected during all the studied period despite of the low NH_4^+
224 -Nre achieved. The average relative abundance of these species was 3% for AOB and 1%
225 for NOB (Fig. 2). *Nitrosomonas oligotropha* lineage and members of the genus *Nitrotoga*
226 were found by FISH as the dominant nitrifiers responsible for ammonia and nitrite
227 oxidation, respectively. Halophilic and halotolerant *Nitrosomonas* spp., as *N. eutropha*
228 and *N. halophila*, and members of the genus *Nitrospira* were present at lower relative
229 abundance. All of them are common nitrifying species founded in AS nitrogen removal
230 systems. FISH probe signals were not detected for *N. europea*, *Nitrosococcus mobilis*,
231 *Nitrobacter*, *Nitrolancetus hollandicus*, Subcluster Thaumarchaeota group I.1b and
232 *Candidatus Nitrosopumilus maritimus*. In Fig. 2, where the relative abundance of
233 nitrifying populations together with O₃LR is illustrated, non-obvious patterns are found
234 between the two parameters. The model (Table 1) used to investigate which explanatory
235 variables influence the observed changes in nitrogen removal efficiencies showed that

236 41% of the variation in nitrogen removal performances was explained by using four
237 environmental variables. Ozone loading rate (O_3LR) was the best explanatory variable
238 that predicted its behaviour (18.6 % of variance explained), followed by MLSS, PO_4^+ -P
239 and organic loading rate (OLR) expressed as sCOD. The model results were visualized
240 using a dbRDA (Fig. 1a). In order to detect the variables showing best correlations to the
241 set of environmental variables, nitrogen removal efficiencies and nitrogen species on
242 effluent were superimposed as vectors. Total of 32.5% variation was explained by the
243 axis 1, being soluble total nitrogen (sTN) and NH_4^+ -N the variables with highest
244 correlation coefficients ($r = 0.83$ and $r = 0.73$ respectively). These results indicate that
245 sTN and NH_4^+ -N in the effluent were positively correlated with the increasing O_3LR (Fig.
246 1a). NH_4^+ -Nre and dissolved Kjeldahl nitrogen removal efficiency (sTKNre) also present
247 relevant correlations with axis 1 ($r = -0.60$ and $r = -0.56$, respectively) indicating that best
248 removal efficiencies were correlated with O_3LR lower values. The results of the
249 sequential test (Table 1) provide information about the environmental variables (MLSS,
250 PO_4^+ -P and OLR) that can contribute to balance the negative effects of ozone in nitrifying
251 removal performance. DISTLM used to evaluate the correlations between nitrifying
252 bacteria populations and environmental variables, showed that ozone is not the variable
253 that contributes to the highest percentage variance explained (4.8%) (Table 1). Oils and
254 fats loading rate (OFLR) (6.9%) and chemical oxygen demand in mixed liquor (CODML)
255 (6.8%) where the variables mostly correlated with the environmental variables. OFLR
256 were correlated with the axis 1, explaining 14.4% of the variation, however genera
257 *Nitrosomonas* was the only taxa that correlates significantly with the axis 1 ($r=-0.55$). The
258 model shows that O_3LR was explaining the 4.8% of the variation of nitrifying bacterial

259 communities, but no relevant correlation was found by dbRDA with AOB or NOB
260 species.

261 Multivariate data analysis for environmental interpretation of relations between
262 nitrogen removal performances and nitrifying bacteria shows that 22% of nitrifying
263 bacterial communities variation can be related to sTN removal efficiency (sTNre) (13.1%
264 explained variance) and the percentage of NO₂-N in effluent (8.8% explained variance).
265 Percentage of NO₂-N was introduced as a variable in order to detect if there were any
266 factors determining the presence of NO₂ on effluent. It was noticed by the dbRDA (data
267 not shown), that the sTNre correlates with the relative abundance of *N. oligotropha* and
268 *Nitrotoga*, corresponding to dominant nitrifiers in the studied period.

269 The results obtained in our study using multivariate analysis indicate that ozone had
270 a limited impact on NH₄⁺-Nre, and no strong correlations were found between ozone
271 dosage and the nitrifying bacterial community dynamics. Therefore, the impact of ozone
272 on the nitrifying bacterial structure does not explain at all the low ammonia removal
273 efficiency observed, although relationship between dominant species and removal
274 efficiencies were found. Levén et al. (2016) evaluated the effect of low ozone dosage to
275 control *M. parvicella* on nitrifying bacterial community structure in a full-scale plant. In
276 this study, no relevant variations on nitrifying bacterial community composition or
277 abundance were found for the different ozone dosages tested (4.4 – 6.6 gO₃/kgSS).
278 Conversely, in the study by Levén et al. (2016) nitrogen removal performances were also
279 unaffected by the ozone application, probably because other factors balancing the
280 potential effects of ozone. Discrepancies between different studies are possibly due to
281 variations in sludge properties and experimental conditions (Semblante et al., 2017; Yan

282 et al., 2009a), due to the fact that the ozone effect is dependent on the ozonisation
283 conditions (Gardoni et al., 2001; Richardson et al., 2009; Meng et al., 2013).

284 It is well known that nitrifiers form compact aggregates as stress protection (de
285 Boer et al., 1991). *Nitrospira* has been found as dominant NOB in other ozonised AS
286 systems (Isazadeh et al., 2014; Levén et al., 2016). Previous reports noticed that this NOB
287 present higher resistance to ampicillin than *Nitrobacter* and other heterotrophs (Spieck et
288 al., 2006), probably due to its tendency to aggregate in dense flocs embedded in an EPS
289 matrix, that offers them protection against the inhibitors. Yan et al. (2009b) observed that
290 at high ozone dosage (80 mg O₃/g SS) some Gram negative bacteria, typically found in
291 WWTP structured in tetrads or clusters, resisted to the oxidative attack. These findings
292 imply that the presence of an EPS matrix might protect the nitrifying bacteria against the
293 oxidative effect. Other authors (Böhler and Siegrist 2004) suggested that nitrifiers are
294 normally overgrown by faster-growing heterotrophs and may therefore be partly
295 protected in the sludge floc and not exposed to ozone as much as the heterotrophs.

296 3.3. Filamentous bacteria

297 Conventional microscopy and FISH analysis were conducted in order to identify
298 and quantify 11 filamentous bacteria morphotypes, present in AS samples of both reactors
299 (Fig. 3). FISH probes used for the study are presented in the supplementary material.
300 *Gordonia* sp. *Haliscomenobacter hydrosis*, Type 0092, Type 0803-I (industrial
301 phylotype), Type 0803-D (domestic phylotype) and Type 1851 were the most dominant
302 filamentous bacteria observed during the studied period. As showed in Table 1, the degree
303 of variation explained for filamentous bacteria by the set of environmental variables was
304 60.6%. Highest correlations with environmental variables and the set of explanatory
305 variables were found for O₃LR and temperature in the reactor (Tr), indicating that ozone

306 plays an important role structuring the filamentous bacterial community. Results from the
307 multivariate data analysis revealed significant differences in the response of filamentous
308 bacteria to ozone and Tr. Type 0803-I, Type 0041, Type 1851 ($r > 0.45$) declined in
309 abundance as ozone increased and temperatures decreased. The opposite pattern between
310 variables was associated with the occurrence of *Gordonia sp.* and T0803-D. Type 0803-
311 I was the most strongly associated with ozone ($r = -0.71$), indicating that this type was
312 most affected by ozonation, in contrast to *Gordonia sp.* which seemed to be less affected
313 when ozone was applied at concentrations up to 47.8 kgO₃/d, as reflected in the dbRDA
314 (Fig. 1b), which corresponds with a dosage of 0.15 gO₃/kgSSVLM·h. *Gordonia sp.* as
315 well as 0803-I type are both bacteria that grow into the floc matrix, but their occurrence
316 has an opposite trend, indicating there are other factors determining ozone resistance. In
317 this study, the oxidizing agent did not further affect the filaments that grow outside flocs
318 more than other filaments present in the floc matrix, as observed in other studies
319 (Caravelli et al., 2006).

320 In previous studies lower ozone doses were effectively used to control filamentous
321 bulking (1.4 to 2.8 gO₃/kg SS) (Lyko et al., 2012; Nilsson et al., 2014; Ried et al., 2014;
322 Saayman et al., 1996), than ozone dosage applied in this study. Different factors as
323 process configuration, floc characteristics and ozone dosage point, probably influenced
324 the observed results. The study of Guo et al. (2012), where the influence on chlorination
325 or cetyltrimethyl ammonium bromide (CTAB) was evaluated on 021N type, suggests that
326 the most important factor influencing the ozone treatment success seems to be the cell
327 wall penetration capacity of the components. Previous studies have demonstrated that
328 different morphotypes are selectively affected by the chemical treatment applied (Nilsson
329 et al., 2018; Seka et al., 2001). In our study, the observed resistance of *Gordonia sp.* to

330 ozone treatment can be the result of mycolic acids presented in the cell wall of these
331 organisms, since these acids are strongly hydrophobic. Goi et al. (1998) considered that
332 the *Gordonia* foaming suppression mechanism of ozone, within a range of ozone below
333 3 mg/l, was due to the decomposition of mycolic acid and not because the number of
334 *Gordonia* was reduced by oxidative action of ozone. These findings are similar to those
335 reported where *M. parvicella* and *Nostocoida limicola* presented resistance to chemical
336 treatments due to their hydrophobic cell walls (Seka et al., 2001). Therefore it would be
337 of particular relevance to determine the penetration capacity before to implementation of
338 non-specific filamentous control treatment.

339 3.4. Protist and metazoan dynamics.

340 It is well known that protist and metazoa play an important role in maintaining a
341 good balance in AS treatment systems. These populations consume excess free bacteria
342 decreasing turbidity and stimulating their growth, as well as they functioning as a
343 bioindicators in the AS process. Twenty-six species of protist and three species of
344 metazoa were identified. The most common protists quantified were small flagellates and
345 amoebae (Fig. 4).

346 According to the DISTLM results (Table 1), the most important structural factors
347 for the protist and metazoan community dynamics were O₃LR (11.9% of variance
348 explained), OFLR (2.2 % of variance explained) and SRT (2.9% of variance explained).
349 dbRDA on Fig. 1c shows that almost all the protist and metazoan species presented a
350 negative correlation with ozone concentration, indicating that ozonation significantly
351 impacted on this community. Only *Tokophyra infusionum* and *Aspidisca cicada* presented
352 positive correlations, although they were not statistically significant. Protists less tolerant

353 to ozone addition were *Acinertia uninata* ($r = -0.62$), *Amoebae* ($r = -0.50$), *Opercularia*
354 *articulata* ($r = -0.48$) and *Vorticella microstoma* ($r = -0.48$).

355 Only few studies (Nilsson et al., 2018; van Leeuwen et al., 2009; Yan et al., 2009a),
356 have evaluated the effects of ozone addition on protist and metazoan communities. Van
357 Leeuwen et al. (2009) did not find evidence of harmful effects of ozone on protist
358 communities, meanwhile Yan et al. (2009a) found an increase of 1.57 in the number of
359 protist and metazoa in a lab-scale ozonised reactor in contrast with the non-ozonated
360 reactor. They attributed this increase to the direct composition of cell debris from cellular
361 lysis, which probably leads to propagation of microfauna. Nevertheless, in this study it
362 has been concluded that full-scale studies are necessary to evaluate the degree of ozone
363 impact in these populations, since the differences found between reactors may be the
364 consequence of other factors or changes in the influent characteristics. The results of our
365 study are in agreement with those from Nilsson et al. (2018), as they observed certain
366 sensitivity of these communities when ozone concentrations were above 3.0 to 4.8
367 gO_3/kgTSS .

368 **4. Conclusions**

369 Ozone treatment did not significantly affect the structure of nitrifying communities,
370 although it was linked to its functional attributes, being ozone the most significant
371 environmental correlated variable with NH_4^+ -N_{re}.

372 This study suggested that different filamentous morphotypes are selectively
373 affected by ozone, and highlights the importance of determining the factors influencing
374 the ozone treatment resistance. From an implementation perspective, the concentration
375 applied should be investigated prior to full-scale application

376 Multivariate data analysis is an interesting tool to evaluate, as particular reference
377 for an specific process, which environmental variables can contribute to balance the
378 negative effects of ozone in nitrifying removal performance.

379

380 E-supplementary data of this work can be found in online version of the paper.

381 **Acknowledgements**

382 This work was supported by grant from the Entitat de Sanejament d'Aigües
383 (EPSAR). P. Barbarroja acknowledges support from MINECO grant PTA2014-09555-I.
384 We thank the operator of the full-scale plant for the donation of activated sludge samples
385 and operational features.

386 **References**

387 APHA, AWWA, WEF, 2005. Standard Methods for the Examination of Water and
388 Wastewater, 21st ed. American Public Health Association/American Water Works
389 Association/Water Environment Federation, Washington, D.C.

390 Anderson, M.J., Gorley R.N., y Clarke, K.R., 2008. PRIMER + for PERMANOVA:
391 Guide to Software and Statistical Methods. PRIMER-E. Ltd, Plymouth. United Kingdom.

392 Arregui, L., Muñoz, C., Guinea, A., y Serrano, S., 2003. FLUTAX employment
393 simplifies the visualization of the ciliature of oxytrichid hypotrichs. Eur. J. Protistol. 39,
394 169-172.

395 Bray, J.R., and Curtis., J.T., 1957 Ordination of the upland forest community of
396 Southern Wisconsin. Ecol. Monogr. 27, 325-349.

397 Böhler, M., Siegrist, H., 2004. Partial ozonation of activated sludge to reduce
398 excess sludge, improve denitrification and control scumming and bulking. In: Water Sci.
399 Technol. 49, 41-49.

400 Borrás, L., 2008. Técnicas microbiológicas aplicadas a la identificación y
401 cuantificación de organismos presentes en sistemas EBPR. Tesis. Valencia: Universitat
402 Politècnica de València.^[1]

403 Caravelli, A., Giannuzzi, L., & Zaritzky, N., 2006. Effect of ozone on filamentous
404 bulking in a laboratory scale activated sludge reactor using respirometry and INT-
405 dehydrogenase activity. *J. Environ. Eng.* 132, 1001-1010.

406 Chen, J., Liu, S., Yan, J., Wen, J., Hu, Y., & Zhang, W., 2017. Intensive removal
407 efficiency and mechanisms of carbon and ammonium in municipal wastewater treatment
408 plant tail water by ozone oyster shells fix-bed bioreactor– membrane bioreactor combined
409 system. *Ecol. Eng.* 101, 75-83.

410 Chu, L., Yan, S., Xing, X.-H., Sun, X., Jurcik, B., 2009. Progress and perspectives
411 of sludge ozonation as a powerful pretreatment method for minimization of excess sludge
412 production. *Water Res.* 43, 1811-1822.

413 Clarke, K.R., Gorley, R.N, Somerfield, P.J., y Warwick, R.M., 2014. Change in
414 Marine Communities: an Approach to Statistical Analysis and Interpretation, 3rd edition.
415 PRIMER-E, Plymouth, pp. 260.

416 Clarke, K.R, y Gorley, R.N., 2015. PRIMER v7: User Manual/Tutorial. PRIMER-
417 E, Plymouth, pp. 296.^[1]

418 Daims, H., Stoecker, K., & Wagner, M., 2004. Fluorescence in situ hybridization
419 for the detection of prokaryotes, in *Molecular Microbial Ecology*. Taylor & Francis, pp.
420 208-228.

421 De Boer W., Gunnewiek PGAK., Veenhuijs M., Bock E., Laanbroek HJ., 1991.
422 Nitrification at low pH by aggregated chemolithotrophic bacteria. *Appl. Environ.*
423 *Microbiol.* 57, 3600–3604

424 Demir, O., Filibeli, A., 2012. Fate of return activated sludge after ozonation: an
425 optimization study for sludge disintegration. *Environ. Technol.* 33, 1869-1878.

426 Dytczak, M. A., Londry, K. L., Siegrist, H., & Oleszkiewicz, J. A., 2007. Ozonation
427 reduces sludge production and improves denitrification. *Water res.* 41, 543-550.

428 Dytczak, M.A., Oleszkiewicz, J.A., 2008. Performance change during long-term
429 ozonation aimed at augmenting denitrification and decreasing waste activated sludge.
430 *Chemosphere* 73, 1529-1532.

431 Eikelboom, D., 2000. *Process Control of Activated Sludge Plant by Microscopic*
432 *Investigations.* London: IWA Publishing.

433 Eikelboom, D., 2006. *Identification and Control of Filamentous Microorganisms in*
434 *Industrial Wastewater Treatment Plants (CD).* London: IWA Publishing.^[1]

435 Fall, C., & Silva-Hernández, B. C., 2017. Bacterial inactivation and regrowth in
436 ozonated activated sludges. *Chemosphere*, 189, 357-364.

437 Fernández-Galiano, D. 1976. Silver impregnation of ciliated protozoa: procedure
438 yielding good results with the pyridinated silver carbonate method. *Trans.Am. Micros.*
439 *Soc.* 557-560.

440 Foissner, W., Blatterer, H., Berger, H., y Kohmann, F., 1991. *Cyrtophoria,*
441 *Oligotrichi, Hypotrichia, Colpodea en Taxonomische und Ökologische Revision der*
442 *Ciliaten des Saprobiensystems: Informationsberichte des Bayer. Landesamtes für*
443 *Wasserwirtschaft* 1/91, 471.

444 Foissner, W., Berger, H. y Kohmann, F., 1992. *Peritrichia, Heterotrichida,*
445 *Odontostomatida en Taxonomische und Ökologische Revision der Ciliaten des*
446 *Saprobiensystems: Informationsberichte des Bayer. Landesamtes für Wasserwirtschaft*
447 *1/92, 502.*

448 Foissner, W., Berger, H., y Kohmann, F., 1994. Hymenostomata, Protomatida,
449 Nassulida en Taxonomische und Ökologische Revision der Ciliaten des
450 Saprobiensystems: Informationsberichte des Bayer. Landesamtes für Wasserwirtschaft
451 1/94, 548.

452 Foissner, W., Berger, H., Blatterer, H., y Kohmann, F., 1995. Gymnostomatea,
453 Loxodes, Suctoria en Taxonomische und Ökologische Revision der Ciliaten des
454 Saprobiensystems: Informationsberichte des Bayer, Landesamtes für Wasserwirtschaft
455 1/95, 540.

456 Gardoni, D., Ficara, E., Fornarelli, R., Parolini, M., Canziani, R., 2011. Long-term
457 effects of the ozonation of the sludge recycling stream on excess sludge reduction and
458 biomass activity at full-scale. *Water Sci. Technol.* 63, 2032-2038.

459 Goi, M., Odagawa, K., Nishimura, T., Okoch, T., & Yuzawa, H. 1998. Gordona
460 scum suppression mechanism of ozone added in wastewater treatment plants. *Water Sci.*
461 *Technol.* 38, 87-94.

462 Guo, J., Peng, Y., Wang, Z., Yuan, Z., Yang, X., & Wang, S. , 2012. Control
463 filamentous bulking caused by chlorine-resistant Type 021N bacteria through adding a
464 biocide CTAB. *Water res.* 46, 6531-6542.

465 Isazadeh, S., Ozcer, P. O., & Frigon, D., 2014. Microbial community structure of
466 wastewater treatment subjected to high mortality rate due to ozonation of return activated
467 sludge. *J. Appl. Microbiol.* 117, 587-596.

468 Jenkins, D., Richard, M.G., Daigger, G.T., 2004. Manual on the causes and control
469 of activated sludge bulking and other solids separation problems, 3rd ed. IWA Publishing,
470 London, UK.

471 Klein, B. M., 1926. Über eine neue Eigentümlichkeit der Pellicula von *Chilodon*
472 *uncinatus* Ehrbg. Zool. Anz, 67, 1-2.

473 Komanapalli, I.R., Lau, B.H.S., 1996. Ozone-induced damage of *Escherichia coli*
474 K-12. Appl. Microbiol. Biotechnol. 46, 610-614.

475 Lee, J.W., Cha, H.Y., Park, K.Y., Song, K.G., Ahn, K.H., 2005. Operational
476 strategies for an activated sludge process in conjunction with ozone oxidation for zero
477 excess sludge production during winter season. Water Res. 39, 1199–1204.

478 Levén L, Wijnbladh E, Tuveesson M., 2016. Control of *Microthrix parvicella* and
479 sludge bulking by ozone in a full-scale WWTP. Water Sci. Technol. 73, 866-872.

480 Lyko, S., Teichgräber, B. & Kraft, A., 2012 Bulking control by low- dose ozonation
481 of returned activated sludge in a full-scale wastewater treatment plant. Water Sci.
482 Technol. 65, 1654–1659.

483 Nagare, H., Tsuno, H., Saktaywin, W., & Soyama, T., 2008. Sludge ozonation and
484 its application to a new advanced wastewater treatment process with sludge
485 disintegration. Ozone Sci. Eng. 30, 136-144.

486 Naso, M., Chiavola, A., Rolle, E., 2008. Application of excess activated sludge
487 ozon- ation in an SBR Plant. Effects on substrate fractioning and solids production. Water
488 Sci Technol, vol. 58, 239-245.

489 Nie, Y., Qiang, Z., Ben, W., & Liu, J., 2014. Removal of endocrine-disrupting
490 chemicals and conventional pollutants in a continuous-operating activated sludge process
491 integrated with ozonation for excess sludge reduction. Chemosphere, 105, 133-138.

492 Nilsson, F., Hagman, M., Mielczarek, A. T., Nielsen, P. H. & Jönsson, K., 2014
493 Application of ozone in full-scale to reduce filamentous bulking sludge at Öresundsverket
494 WWTP. Ozone Sci. Eng. 36, 238–243.

495 Nilsson, F., Davidsson, Å., Falås, P., Bengtsson, S., Bester, K., & Jönsson, K.,
496 2018. Impact of activated sludge ozonation on filamentous bacteria viability and possible
497 added benefits. *Environ. Technol.* 1-7.

498 Pérez-Uz, B., Arregui, L., Calvo, P., Salvadó, H., Fernández, N., Rodríguez, E.,
499 Zornoza, A., y Serrano, S., 2010. Assesment of advanced wastewater treatments for
500 nitrogen removal searching for plausible efficiency bioindicators. *Water Res.*, 44, 5059-
501 5069.

502 Richardson, E.E., Hanson, A., Hernandez, J., 2009. Ozonation of continuous-flow
503 activated sludge for reduction of waste solids. *Ozone Sci. Eng.* 31, 247-256.

504 Ried, A., Wang, J., Rand, W. & Fabiyi, M., 2014. Ozone to control bulking and
505 foaming in municipal waste water treatment plant. In: DSD International Conference,
506 Hong Kong, B1–B3.

507 Romero, P., Coello, M.D., Arago_n, C.A., Battistoni, P., Eusebi, A.L., 2015. Sludge
508 reduction through ozonation: effects of different specific dosages and operative
509 management aspects in a full-scale study. *J. Environ. Eng.*, 141.

510 Saayman, G. B., Schutte, C. F., and van Leeuwen, J., 1996. The effect of chemical
511 bulking control on biological nutrient removal in a full scale activated sludge plant. *Water*
512 *Sci. Technol.*, 34, 275–282.

513 Seka, M. A., Kalogo, Y., Hammes, F., Kielemoes, J., & Verstraete, W. 2001.
514 Chlorine-susceptible and chlorine-resistant type 021N bacteria occurring in bulking
515 activated sludges. *Appl. Environ. Microbiol.*, 67, 5303-5307.

516 Semblante, G. U., Hai, F. I., Dionysiou, D. D., Fukushi, K., Price, W. E., & Nghiem,
517 L. D., 2017. Holistic sludge management through ozonation: A critical review. *J. Environ.*
518 *Manage.* 185, 79-95.

519 Sui, P., Nishimura, F., Nagare, H., Hidaka, T., Nakagawa, Y., Tsuno, H., 2011.
520 Behavior of inorganic elements during sludge ozonation and their effects on sludge
521 solubilization. *Water Res.* 45, 2029-2037.

522 Sui, P., Nishimura, F., Tsuno, H., 2014. Nitrogen behavior during sludge ozonation:
523 a long-term observation by pilot experiments. *Water Sci. Technol.* 70, 289-296.

524 Spieck, E., Hartwig, C., McCormack, I., Maixner, F., Wagner, M., Lipski, A., &
525 Daims, H., 2006. Selective enrichment and molecular characterization of a previously
526 uncultured *Nitrospira* - like bacterium from activated sludge. *Environ. Microbiol.* 8,
527 405-415.

528 van Leeuwen, J., & Pretorius, W. A., 1988. Sludge bulking control with
529 ozone. *Water Environ. J.* 2, 223-227.

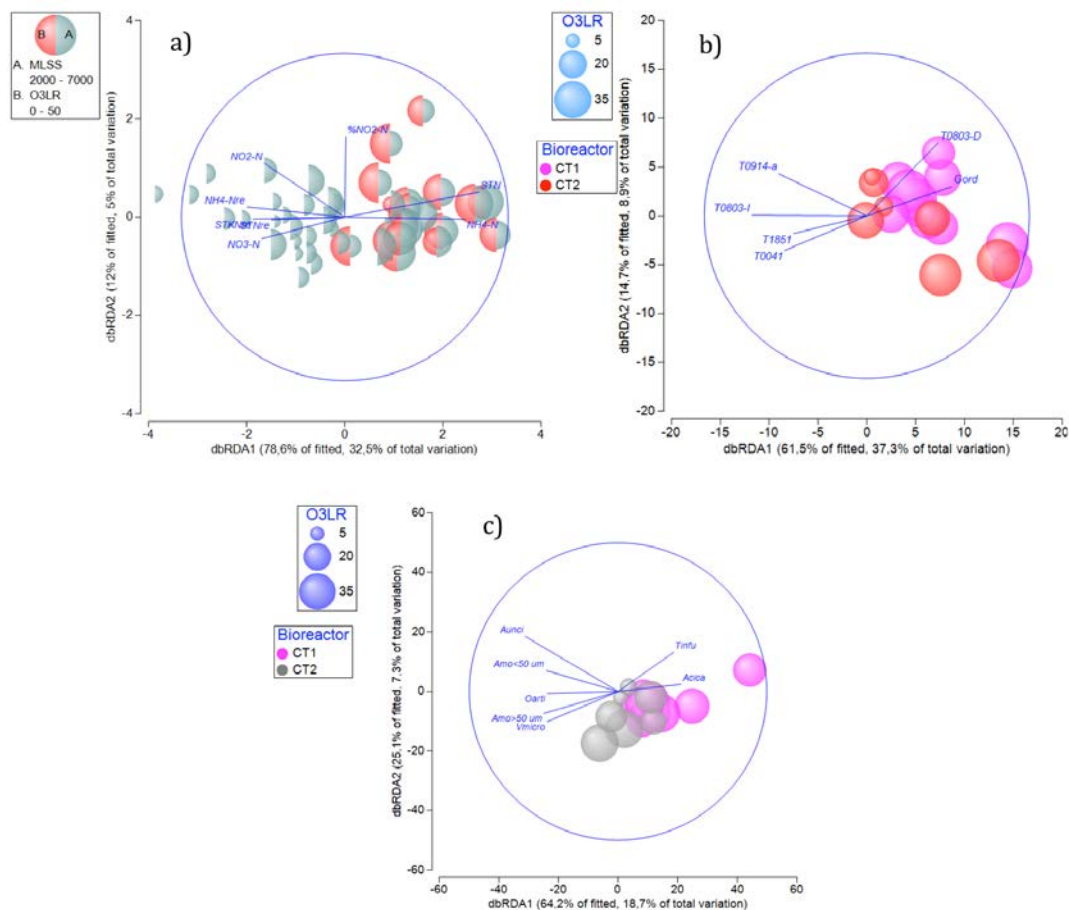
530 van Leeuwen J.H., Sridhar A., Harrata A.K., Esplugas M., Onuki S., Cai L., Koziel
531 J.A., 2009. Improving the biodegradation of organic pollutants with ozonation during
532 biological wastewater treatment. *Ozone Sci. Eng.* 31, 63-70.

533 Yan, S.-T., Zheng, H., Li, A., Zhang, X., Xing, X.-H., Chu, L.-B., Ding, G., Sun,
534 X.-L., Jurcik, B., 2009a. Systematic analysis of biochemical performance and the
535 microbial community of an activated sludge process using ozone-treated sludge for
536 sludge reduction. *Bioresour. Technol.* 100, 5002-5009.

537 Yan, S. T., Chu, L. B., Xing, X. H., Yu, A. F., Sun, X. L., & Jurcik, B., 2009b.
538 Analysis of the mechanism of sludge ozonation by a combination of biological and
539 chemical approaches. *Water res.* 43, 195-203.

540 Yapsakli, K., Mertoglu, B., Cecen, F., 2010. Identification of nitrifiers and
541 nitrification performance in drinking water biological activated carbon (BAC) filtration.
542 *Proce. Biochem.* 45, 1543–1549.

543 Zornoza, A., 2017. Estudio de la dinámica poblacional de protistas, metazoos y
544 bacterias filamentosas y su interpretación ecológica en fangos activos Tesis. Valencia:
545 Universitat Politècnica de València.



546

547

Fig. 1. dbRDA plots representing: (a) model of temporal variation on nitrogen

548

species on effluent and nitrogen removal efficiencies in relationship to environmental

549

variables, (b) model of temporal variation on filamentous bacterial community in

550

relationship to environmental variables and (c) model of temporal variation on protist

551

and metazoan community in relationship to environmental variables. Bubbles are scaled

552

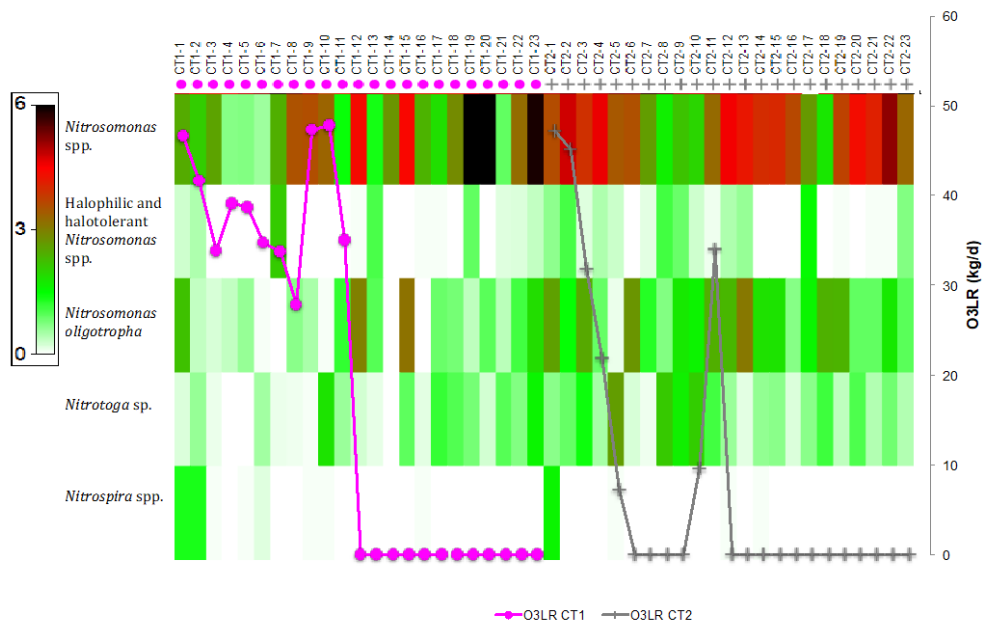
to represent the value of the explanatory variables. Vectors show the direction and

553

strength of the correlation between species and explanatory variables. MLSS, mixed

554

liquor suspended solids (mg/L); O3LR, ozone loading rate (kgO₃/d).

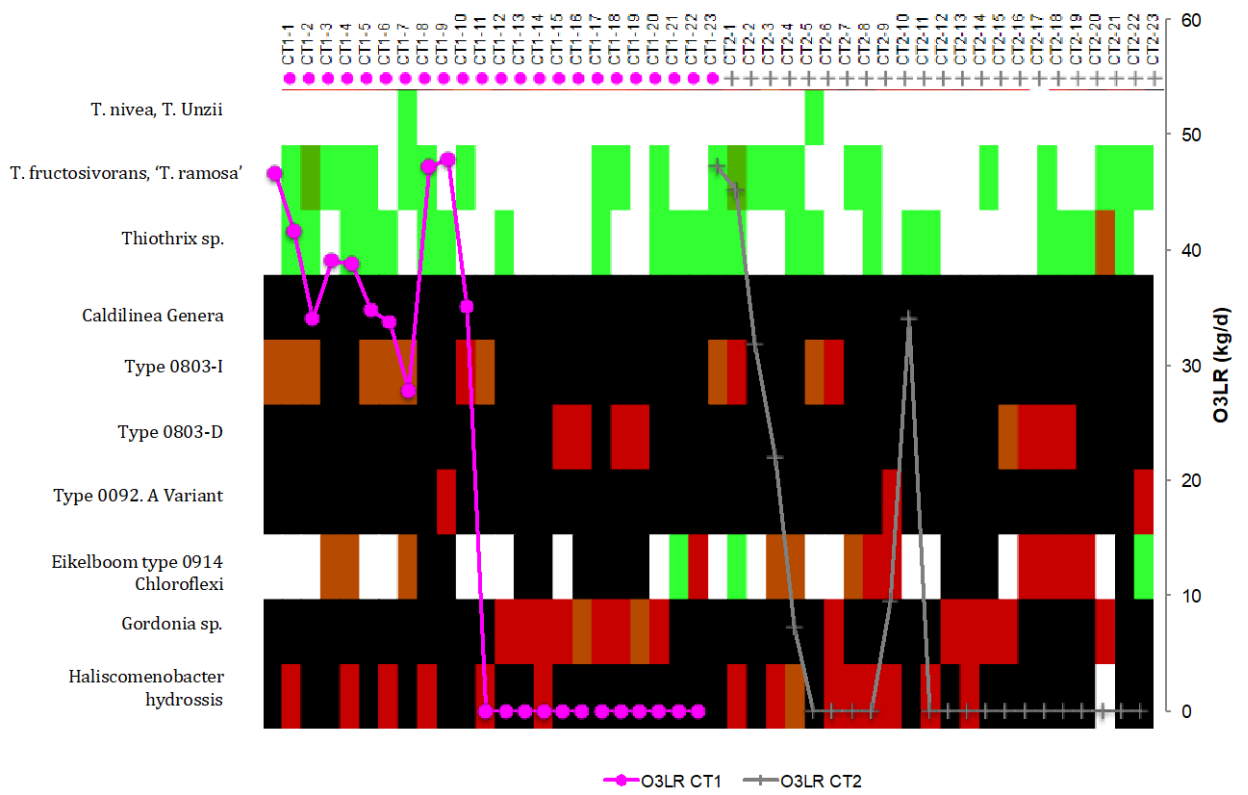


555

556 Fig. 2. Relative abundance of nitrifying bacterial community and ozone loading rate

557 (O3LR) in the reactors 1 (CT1) and 2 (CT2).

558



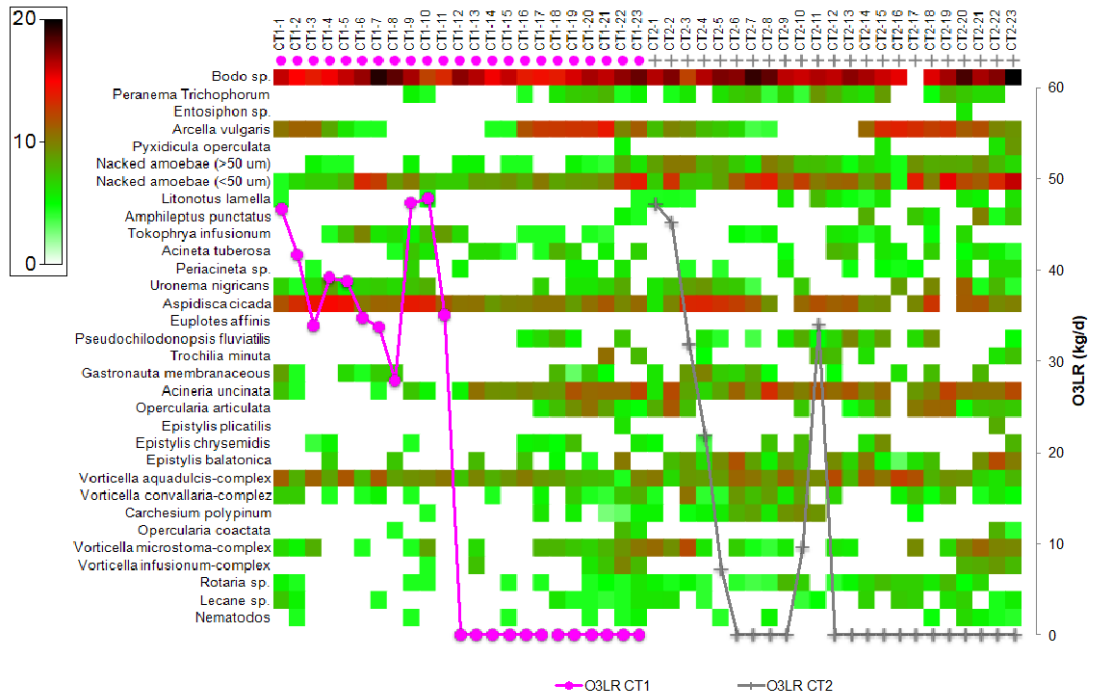
559

560

Fig. 3. Filamentous bacteria abundance and ozone loading rate (O3LR) in the

561

reactors 1 (CT1) and 2 (CT2).



563

564 Fig. 4. Protist and metazoan abundance and ozone loading rate (O3LR) in the
 565 reactors 1 (CT1) and 2 (CT2).

Table 1.
Results of multivariate multiple regression analyses (DistLM)

1) Operational variables and carbon removal performance

Environmental variables	Sequentially			
	AICc	Pseudo-F	p	% Prop.
HO	99.692	4.6785	0.021	9.611
SRT	96.499	5.4459	0.015	10.161

2) Environmental variables and nitrogen removal performance

Environmental variables	Sequentially			
	BIC	Pseudo-F	p	% Prop.
O ₃ LR	92.809	10.084	0.001	18.645
MLSS	92.446	4.1027	0.012	7.0861
PO4-PLR	90.502	5.6149	0.004	8.7581
OLR (COD)	89.304	4.7344	0.004	6.7817

3) Environmental variables and relative abundance of nitrifying bacteria

Environmental variables	Sequentially			
	AICc	Pseudo-F	p	% Prop.
OFLR	283.74	3.2839	0.016	6.9452
MLCOD	282.54	3.3867	0.011	6.7939
O ₃ LR	282.3	2.4818	0.05	4.8127
PO4-PLR	281.95	2.6492	0.027	4.9433

4) Nitrifying bacteria and nitrogen removal performance

Environmental variables	Sequentially			
	AICc	Pseudo-F	p	% Prop.
STNre	280.56	6.6631	0.001	13.152
%NO ₂ -N	277.91	4.8746	0.001	8.8429

5) Environmental variables and protist and metazoa

Environmental variables	Sequentially			
	BIC	Pseudo-F	p	% Prop.
O ₃ LR	307.19	5.9873	0.0001	11.978
OFLR	305.74	5.2366	0.0001	2.1533
SRT	304.88	4.5025	0.0001	2.9131

6) Environmental variables and filamentous bacteria

Environmental variables	Sequentially			
	BIC	Pseudo-F	p	% Prop.
O ₃ LR	1271.8	14.565	0.001	24.869
NTLM	406.71	5.0908	0.002	7.9532
T ^a r	334.49	4.5305	0.002	6.5408
F/M (COD)	349.55	5.2089	0.001	6.8353
MLSS	270.26	4.3572	0.003	5.2849
CPT	286.16	5.0846	0.001	5.5957
CarbLR	180.66	3.4082	0.013	3.5327

Percentage proportion of variance in species or biological removal efficiencies data explained by that variable. Significant values (<0.05) are indicated in bold.
