Document downloaded from:

http://hdl.handle.net/10251/43090

This paper must be cited as:

Alba Martínez, J.; Trujillo Guillen, M.; Berjano, E. (2012). Relationship between roll-off occurrence and spatial distribution of dehydrated tissue during RF ablation with cooled electrodes. International Journal of Hyperthermia. 28(1):62-68. doi:10.3109/02656736.2011.631076.



The final publication is available at

http://dx.doi.org/10.3109/02656736.2011.631076

Copyright

Informa Healthcare

Relation between roll-off occurrence and spatial distribution of dehydrated

tissue during RF ablation with cooled electrodes

MACARENA TRUJILLO¹, JOSÉ ALBA², & ENRIQUE BERJANO²

¹Instituto Universitario de Matemática Pura y Aplicada, Universitat Politècnica de València,

Spain

² Biomedical Synergy, Electronic Engineering Department, Universitat Politècnica de

València, Spain

*To whom all correspondence should be addressed:

Dr. Enrique Berjano

Biomedical Synergy, Electronic Engineering Department (7F)

Universitat Politècnica de València, Spain

Camino de Vera, 46022 Valencia, Spain

Phone: 34–963877607 Fax: 34–963877609

Email: eberjano@eln.upv.es

1

Abstract

Purpose: To study the relationship between roll-off (sudden increase in impedance) and

spatial distribution of dehydrated tissue during RF ablation using a cooled electrode

(temperatures around 100°C).

Methods: We used a double approach: 1) theoretical modeling based on the Finite Element

Method; and 2) 20 ablations using an experimental study on ex-vivo excised bovine liver in

which we measured impedance progress and temperature at three points close to the electrode

surface: 0.5 (T1), 1.5 (T2) and 2.5 (T3) mm from the tip. T2 was located exactly at the center

of the 30 mm long electrode.

Results: Temperatures at T1 and T3 quickly rose to 100° C (at ≈ 20 and 40 s, respectively),

while at the rise at T2 was somewhat slower, stabilized around 50 s and reached a maximum

value of 99°C at about 60 s. Impedance reached a minimum of 65 Ω (plateau), began

increasing at 50 s and continued rising throughout the procedure, reaching a value equal to the

initial value at 70 s. Likewise, computed impedance dropped to \approx 73 Ω (plateau), began

increasing at 50 s and reached an impedance value equal to the initial value at ≈78 s, which

approximately coincided with the time when the entire zone surrounding the electrode was

within the 100°C isotherm.

Conclusion: There is a close relationship between the moment at which roll-off occurs and the

time when the entire electrode is completely encircled by the dehydrated tissue. The mid-

electrode zone is the last in which tissue dessication occurs.

Keywords: Cooled electrode, ex vivo model, radiofrequency ablation, roll-off, theoretical

model

2

Introduction

Radiofrequency (RF) ablation (RFA) is a minimally invasive ablative technique that allows the treatment of tumors [1,2]. It uses RF current (≈500 kHz) to produce a coagulation zone characterized by a thermal lesion which destroys the tissue by a necrosis process. The coagulation zone is defined as that zone including irreversibly damaged tissue, i.e. with temperatures above 50°C [3]. The electrical current is delivered to the tissue through a small active electrode placed in the center of the tumor and a large dispersive electrode located on the patient's back. One of the most widely used electrodes is the cooled electrode [4], which consists of a single metal needle typically 30 mm long, inside which cooled water (5–10°C) is circulated by means of a hydraulic circuit. This cooling prevents tissue on the electrode surface becoming totally dehydrated and therefore charred [5]. A special feature is that it shifts the most highly desiccated zone 1–2 mm away from the electrode surface. During the operation it is usual to observe a sudden increase in the electrical impedance measured between the active and dispersive electrodes, with a consequent drop in electrical power. This phenomenon is known as roll-off and has been traditionally associated with the high temperatures (≈100°C), vaporization phenomenon and hence dehydration of the tissue. Standard operation with a cooled electrode uses a constant-impedance mode for delivering RF power, which is based on a pulsed protocol: roll-off causes the cessation of the RF pulse, which returns after a short pause. Roll-off impedes the lesion growing, and makes longer the treatment. It would be important to know more about the phenomena associated with roll-off to minimize its appearance.

It has been reported that the addition of hypertonic saline perfusion at 2 mm from the electrode surface improves the performance of the cooled electrode by increasing the volume of the coagulation zone obtained from the cooled electrode only [6,7]. This improvement is also associated with a delay in the emergence of roll-off [8]. We think that the possible

physical explanation of these results is that the infused saline impedes the dehydration of the zone of tissue responsible for roll-off. As far as we know, pronounced dehydration and vaporization processes occur at tissue temperatures around 100°C [3]. The resulting loss of water content consequently involves an abrupt drop in electrical conductivity (σ). Since the σ value at different points on the tissue determines total impedance, this could indicate a relationship between the zones reaching 100°C (dehydrated tissue) and the evolution of impedance. It is now known that the coagulation zone begins in the tissue close to the electrode, i.e. at the tip and the proximal edge (see Fig. 1). This is due to the edge effect, i.e. the electrical power (Specific Absorption Rate, W/m³) is mainly deposited in these zones [9]. In addition, in the first few seconds tissue temperature can reach up to 100°C in these zones, which is associated with the gas bubbles observed by means of ultrasound guidance [10]. From computer modeling studies [9] we know that the coagulation zone is then enlarged by thermal conduction, in particular around the central electrode zone. It is therefore reasonable to conclude that the highly desiccated tissue zone (≈100°C) also extends in the same direction (see Fig. 1). Although many theoretical [9,11,12] and experimental [11-13] studies have assessed the thermal performance of the cooled electrode, the exact relationship between spatial-temporal distribution of the dehydrated tissue and the occurrence of roll-off has yet to be characterized in detail. In this respect, we hypothesized that the roll-off episode coincides approximately with the time when the tissue around the center of the active electrode becomes highly desiccated (around 100°C), i.e. when the entire electrode is completely encircled by the dehydrated tissue. Our aim was to demonstrate this hypothesis by means of a double theoretical modeling and experimental approach.

Methods

Experimental study

The experimental study was conducted on ex vivo tissue. A piece of fresh bovine liver (3.5) kg) was positioned in a 600 cm² container on a dispersive electrode (aluminum foil). A 3 cm long Cool-tip was used connected to a CC-1 RF generator (Radionics, Burlington, MA, USA). Three lengthwise points were chosen close to the electrode surface to measure temperature evolution: 0.5, 1.5 and 2.5 mm from the tip (T1, T2 and T3 in Fig. 2, respectively). T2 coincided exactly with the center of the electrode, so that a custom-made multi-thermocouple Series T-type probe (Physitemp Instruments, Crifton, NY, USA) was employed. It consisted of three ultra-fine thermocouples embedded in a flexible Teflon-coated probe of 17-gauge diameter. We used a narrow gage (<1 mm) nylon thread to tie the multithermocouples probe to the electrode. This knot was located away from the electrode tip to avoid its influencing the heating process (see Fig. 2). Thermocouple temperatures were recorded by a custom-made data acquisition card Thermes USB-RF (Physitemp Instruments, Crifton, NY, USA) and acquired by DASYLab software 9.0 (Measurement Computing, Norton, MA, USA). Tissue impedance was monitored by a separate data acquisition system and processed by Agilent VEE software (Agilent Technologies, Santa Clara, CA, USA). The sampling frequency used in both systems was 10 Hz. Temperature and impedance were recorded up to the first roll-off. Fig. 3 shows the experimental setup. The peristaltic pump model 323 by Watson-Marlow (Wilmington, MA, USA) was used for internal cooling of the electrode with cooled water (5°C) at a rate of 45 mL/min. Twenty four ablations were conducted at a constant voltage of 80 V. The experiments were conducted at room temperature (26°C).

Theoretical model

The theoretical model was based on a coupled electric-thermal problem and was solved numerically using the FEM by means of COMSOL Multiphysics software (COMSOL Inc.

Burlington, MA, USA). The geometry and dimensions of the theoretical model are shown in Fig. 4. The problem presented axial symmetry and a two-dimensional analysis could be conducted. The model included three different materials: plastic partially covering the electrode, metallic electrode, and hepatic tissue. The values of the characteristics of these materials are shown in Table I [14,15]. We considered temperature-dependent functions for the electrical and thermal conductivity [16] (see Fig. 5). For the thermal conductivity (k) we considered linear growth of 0.0015/°C up to 100°C, after which k was kept constant [17]. The electrical conductivity (σ) grew exponentially +1.5%/°C up to 100°C [18], between 99 and 100°C was kept constant and then σ decreased linearly 4 orders for five degrees (i.e. up to 105°C) [12,17]. The governing equation for the thermal problem is the Bioheat Equation [16]:

$$\rho c \frac{\partial T}{\partial t} = \nabla (k \nabla T) + q + Q_p + Q_m \tag{1}$$

where T is temperature, t is time, ρ is density, c is specific heat, k is thermal conductivity, q is heat source produced by RF power, Q_p is heat loss from blood perfusion and Q_m is metabolic heat generation. The term Q_m was not considered since this effect is insignificant in the RFA. The blood perfusion term Q_p was not considered since the theoretical results were later compared to the experimental results obtained from an ex vivo model. The heat source q (Joule losses) is given by q=JE, where J is current density and E is electric field strength. Both were obtained from the electrical problem, which used the Laplace equation as governing equation $\nabla^2 V$ =0, where V is the voltage. The electric field was calculated by means of E = $-\nabla V$ and J using Ohm's law (J= σ E). We used a quasi-static approach, i.e. the tissues were considered as purely resistive [16], due to the value of the frequencies used in RF (\approx 500 kHz) and for the geometric area of interest (electrical power is deposited in a very small zone close the electrode). Tissue vaporization was considered in the case of the hepatic tissue and we varied the formulation proposed in (1) according to the enthalpy method [17].

The product density×specific heat of the liver at the gas phase was 798×10^3 J/K·m³ [15]. The latent heat was 1.54×10^6 J/Kg, which corresponds to a water fraction of 68% inside the liver tissue. The liver density in the change of phase was 179 kg/m^3 .

The electrical boundary conditions were zero current density in the transversal direction to the symmetry axis and inside the electrode. The dispersive electrode was modeled as an electrical condition on boundaries far from the active electrode. Zero voltage was set in the dispersive electrode. A voltage value of 80 V (identical to that used in the experiments) was applied in the active electrode. The thermal boundary conditions were: Null thermal flux in the transversal direction to the symmetry axis and constant temperature of 17° C in the dispersive electrode. Initial temperature of the tissue was considered to be 17° C. These two values were chosen as identical to those measured in the experiments. The circulating liquid inside the electrode produced a cooling effect modeled by means of a thermal convection coefficient h with a value of 3,366 W/K·m² and a coolant temperature of 10° C. The value of h was calculated by considering a length of 30 mm and a flow rate of 45 mL/min through an area of 1.57×10^6 m², which is equivalent to half of the cross section of the inner diameter of the electrode (see Fig. 4).

Results

Experimental results

Initial temperature was $16.9\pm2.0^{\circ}$ C (mean of T1, T2 and T3), and initial impedance was $98.8\pm5.6~\Omega$. Fig. 6 shows mean temperature progress recorded at T1, T2 and T3, along with mean impedance evolution for the 20 ablations. We observed that temperatures at T1 and T3 increased quickly up to 100° C (≈ 20 and 40 s, respectively), i.e. these zones of tissue became dehydrated, in spite of which impedance continued decreasing. On the other hand, the temperature at T2 increased more slowly, became approximately steady around 50 s and

reached a maximum value of 99°C at about 60 s. Impedance reached a minimum of 65 Ω (plateau) around 40 s, began increasing at 50 s and continued rising throughout the procedure, reaching a value equal to the initial value at 70 s. Fig. 7 shows mean values and standard deviations of T1, T2, T3 and impedance.

Theoretical modeling results

The theoretical results were obtained with the double aim of showing the validity of our hypothesis from a theoretical point of view and to compare them with the experimental results. Fig. 5 shows the evolution of the theoretical impedance measured between the active (cooled) electrode and dispersive electrode. Fig. 8 shows the theoretical temperature distributions in the tissue around the electrode (above) and the progress of the 100°C isotherm in the same zone (below) at four specific times. We observed that, as in the experimental study, impedance was initially $\approx 100~\Omega$, dropped to $\approx 73~\Omega$ (plateau), began increasing at 50 s and reached the an impedance value equal to the initial value at $\approx 78~s$, which approximately coincided with the time when the 100°C isotherm completely surrounded the electrode (see Fig. 8).

Discussion

The aim of this study was to demonstrate that the appearance of the roll-off is closely associated with the time when the tissue in the electrode middle zone reaches temperatures $\approx 100^{\circ}$ C, i.e. when dehydrated tissue completely surrounds the electrode. There are different ways of defining roll-off, from the time when impedance begins increasing to when it exceeds the initial value by 30 Ω . In this context, we considered roll-off as the phenomenon when impedance starts to rise after remaining more or less constant (plateau). We therefore employed this criterion and focused on the time relation between the occurrence of roll-off

and the spatial distribution of the severely desiccated tissue, i.e. reaching temperatures $\approx 100^{\circ}\text{C}$. This was done by means of a double approach: theoretical modeling and experiments.

Even though the results from the two approaches are slightly different, they both suggest that the hypothesis could be true. The experimental results showed that impedance began increasing when temperature in the mid-electrode zone (T2) attained a more or less steady value (i.e. at 50 s). The theoretical results showed that impedance began increasing at 50 s, the experimental temperature then reached a maximum value of 99°C around 60 s and impedance reached a value equal to the initial value at 70 s. Likewise, the impedance computed from the theoretical results reached a value equal to the initial value at ≈ 78 s, which approximately coincided with the time when the electrode was entirely within the 100°C isotherm.

The theoretical model used here to check the hypothesis considered the sudden drop in tissue electrical conductivity and latent heat, as used by Haemmerich [9], including tissue vaporization by the enthalpy method. This author also conducted an ex vivo experiment in order to validate his theoretical model by measuring temperatures and impedance at distances of 10, 15 and 20 mm from the middle zone of a cooled electrode. As these distances are too far from the points where tissue reaches ≈100°C, no information about the relationship between roll-off and 100°C isotherm zones could be obtained from his study. In our study we focused on this zone, and for this reason we used an ultra-fine custom-made multi-thermocouple probe, which allowed us to measure the temperature at three different locations close to the electrode surface along its longitudinal axis. With this setup our experimental results also allowed us to confirm the hypothesis.

This study also had other limitations. Although the temperature sensors were very thin, they could possibly have altered the true temperature distributions during the experiments

and, from an electrical point of view, they could have modified the true conditions. To check this possible effect, we conducted five measurements of impedance in the hepatic tissue with the electrode alone, i.e. without the multi-thermocouples probe. The initial impedance measured with this set-up was $98.2\pm4.2~\Omega$, without any significant differences (p>0.05) with the combined electrode and multi-thermocouples probe ($98.8\pm5.6~\Omega$). This suggests that the presence of the multi-thermocouple probe next to the electrode does not significantly perturb RF heating from an electrical point of view. However, the difference between the precise position of the temperature sensor (T2) in the experimental study and the location of the 100° C isotherm in the theoretical model, and the possible self-heating of the thermocouples, could have caused the small differences found between the theoretical and experimental studies.

Both the experimental and theoretical findings suggest that the instant when roll-off occurs (when impedance starts to increase) is closely related to the time when dehydrated tissue reaches the middle zone of the electrode, i.e. when the area close to the electrode reaches a temperature around 100°C. This conclusion could provide an explanation for the previous experimental results obtained with a hybrid applicator based on a cool-tip electrode combined with a low-rate interstitial infusion of hypertonic saline at 2 mm from the electrode surface and exactly in the middle zone [6]. To be more precise, the saline infusion at that point could significantly delay roll-off, which could be due to the saline impeding severe tissue desiccation at this point.

Acknowledgements

This work received financial support from the Spanish "Plan Nacional de I+D+I del Ministerio de Ciencia e Innovación" Grant No. TEC2008-01369/TEC and FEDER Project MTM2010-14909. The translation of this paper was funded by the Universitat Politècnica de València, Spain

References

- 1. Poon RT, Fan ST, Tsang FH, Wong J. Locoregional therapies for hepatocellular carcinoma: a critical review from the surgeon's perspective. Ann Surg 2002; 235: 466-486.
- Solbiati L, Livraghi T, Goldberg SN, Ierace T, Meloni F, Dellanoce M, et al. Percutaneous radio-frequency ablation of hepatic metastases from colorectal cancer: Long-term results in 117 patients. Radiology 2001; 221: 159-166.
- 3. Ahmed M, Brace CL, Lee Jr FT, Goldberg SN. Principles of and advances in percutaneous ablation. Radiology 2011; 258: 351-369.
- Goldberg SN, Grassi CJ, Cardella JF, Charboneau JW, Dodd 3rd GD, Dupuy DE, et al. Society of Interventional Radiology Technology Assessment Committee; International Working Group on Image-Guided Tumor Ablation. Image-guided tumor ablation: Standardization of terminology and reporting criteria. Radiology 2005: 235: 728-739.
- 5. Pereira PL, Trübenbach J, Schenk M, Subke J, Kroeber S, Shaefer I, et al. Radiofrequency ablation: In vivo comparison of four commercially available devices in pig livers. Radiology 2004; 232:482-490.
- 6. Burdío F, Tobajas P, Quesada-Diez R, Berjano E, Navarro A, Poves I, et al. Distant infusion of saline may enlarge coagulation volume during radiofrequency ablation of liver tissue using cool-tip electrodes without impairing predictability. AJR Am J Roentgenol 2011;196:W837-843.
- 7. Burdío F, Navarro A, Berjano EJ, Burdío JM, Gonzalez A, Güemes A, et al. Radiofrequency hepatic ablation with internally cooled electrodes and hybrid applicators with distant saline infusion using an in vivo porcine model. Eur J Surgical Oncology 2008; 34:822-830.
- 8. Burdío F, Berjano EJ, Navarro A, Burdío JM, Güemes A, Grande L, et al. RF tumor ablation with internally cooled electrodes and saline infusion: what is the optimal location of the saline infusion? Biomedical Engineering Online 6:1-9, 2007.
- Haemmerich D, Mathematical Modeling of Impedance Controlled Radiofrequency Tumor Ablation and Ex-Vivo Validation. Proc. 32nd Annual International Conference of the IEEE EMBS, Buenos Aires, Argentina, 2010, 1605-1608.
- 10. Arata MA, Nisenbaum HL, Clark TW, Soulen MC, "Percutaneous radiofrequency ablation of liver tumors with the LeVeen probe: is roll-off predictive of response? J Vasc Interv Radiol 2001:12;455-458.
- 11. Quaranta V, Manenti G, Bolacchi F, Cossu E, Pistolese CA, Buonomo OC, et al. FEM analysis of RF breast ablation: Multiprobe *versus* cool-tip electrode. Anticancer Research 2007; *27*: 775-784.

- 12. Haemmerich D, Chachati L, Wright AS, Mahvi DM, Lee Jr FT, Webster JG. Hepatic radiofrequency ablation with internally cooled probes: effect of coolant temperature on lesion size. IEEE Trans Biomed Eng 2003;50:493-500.
- 13. McGahan JP, Loh S, Boschini FJ, Paoli EE, Brock JM, Monsky WL, et al.Maximizing parameters for tissue ablation by using an internally cooled electrode. Radiology 2010;. 256:397-405.
- 14. Berjano EJ, Burdío F, Navarro AC, Burdío JM, Güemes A, Aldana O, et al. Improved perfusion system for bipolar radiofrequency ablation of liver. Physiol Meas 2006;27:55-66.
- Pätz T, Körger T, Preusser T. Simulation of Radiofrequency Ablation Including Water Evaporation. IFMBE Proceedings 2009;25:1287-1290.
- 16. Berjano EJ. Theoretical modeling for radiofrequency ablation: State of-the-art and Challenges for the future. Biomed Eng Online 2006; 18;5:24.
- 17. Byeongman J, Aksan A. Prediction of the extent of thermal damage in the cornea during conductive thermokeratoplasty. J Therm Biol 2010; 35:167-174.
- 18. Pearce J, Panescu D, Thomsen S. Simulation of dioptre changes in radio frequency conductive keratoplasty in the cornea. WIT Trans Biomed Health 2005; 8:469-477.
- Abraham JP, Sparrow EM. A thermal-ablation bioheat model including liquid-to-vapor phase change, pressure- and necrosis-dependent perfusion, and moisture-dependent properties. Int J Heat Mass Tran 2007;50:2537-2544.

Table I. Characteristics of the materials used in the model [14,15].

Tissue/material	σ(S/m)	$k \left(W/m \cdot K \right)$	$\rho (\text{kg/m}^3)$	c (J/kg·K)
Electrode	7.4×10^6	15	8×10 ³	480
Plastic	10 ⁻⁵	0.026	70	1045
Liver	$0.128^{(1)}$	$0.472^{(2)}$	1080	3455

 $[\]sigma$, electric conductivity; k, thermal conductivity; ρ , density; c, specific heat.

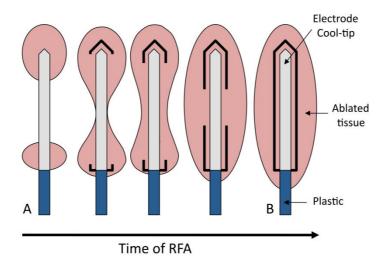


Figure 1 Schematic diagram of the progress of the coagulation zone in the tissue during radiofrequency ablation. At the beginning, the coagulated tissue is mainly located at the edges (distal and proximal), while at the end of heating the coagulated lesion has broadly extended and forms an ellipsoidal shape with the longest axis being the electrode axis. The bold lines represent the tissue heated over 100°C, i.e. strongly dehydrated zones.

⁽¹⁾ Assessed at 17°C (initial temperature). This allows theoretical initial impedance (\approx 100 Ω) to match the experimental mean value.

⁽²⁾ Assessed at 17°C [14].

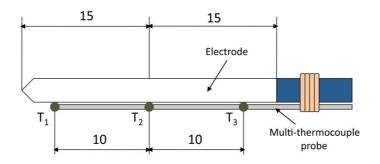


Figure 2 Placement of the multi-thermocouple probe tied to the electrode. T1, T2 and T3 are the locations of the thermocouples (dimensions in mm).

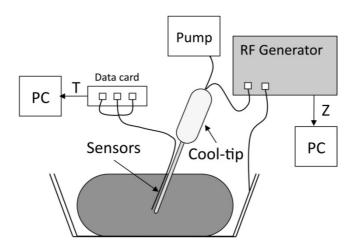


Figure 3 Experimental setup to record temperatures and impedance evolution. The multi-thermocouple probe and electrode were jointly inserted into the tissue.

The electrode was internally cooled by a peristaltic pump.

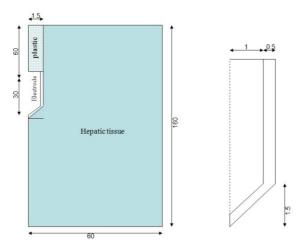


Figure 4 Geometry of the two-dimensional theoretical model (out of scale, and dimensions in mm). The domain is divided into three zones: plastic portion of the electrode, metallic electrode, and hepatic tissue.

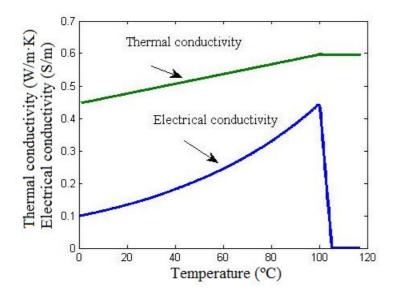


Figure 5 Change in the tissue thermal and electrical conductivity with temperature.

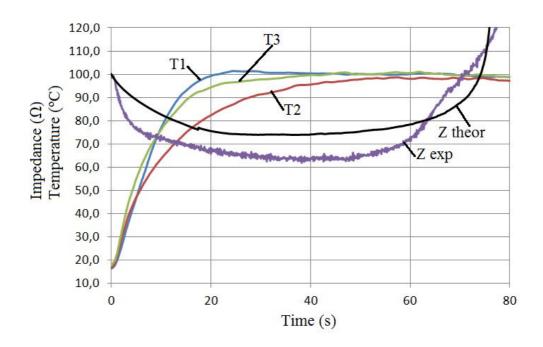
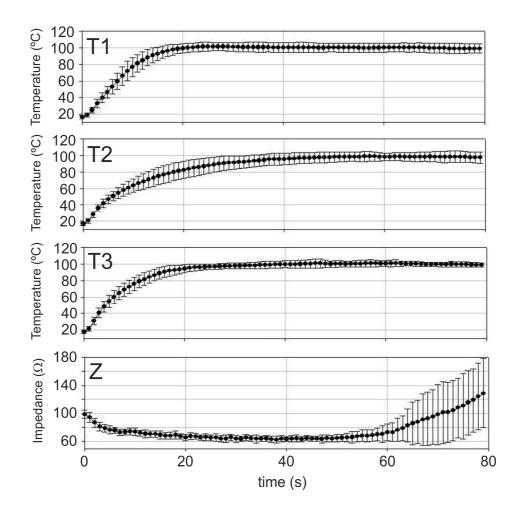


Figure 6 Results from ex vivo experiments and theoretical model. Mean temperature evolution of the sensors placed in the tissue surrounding the surface of the electrode tip: T1, T2 and T3 were the thermocouples placed at 0.5, 1.5 and 2.5 mm, respectively, from the electrode tip. Mean progress of the impedance Zexp. Theoretical progress of the impedance Ztheor. Note that the impedance began increasing when temperature at the middle of the electrode (T2) became approximately steady at around 50 s.



Results from ex vivo experiments. Progress of the impedance (Z) and temperature evolution of the sensors placed in the tissue surrounding the surface of the electrode tip: T1, T2 and T3 were the thermocouples placed at 0.5, 1.5 and 2.5 mm, respectively, from the electrode tip. Dots are mean values and error bars are standard deviations.

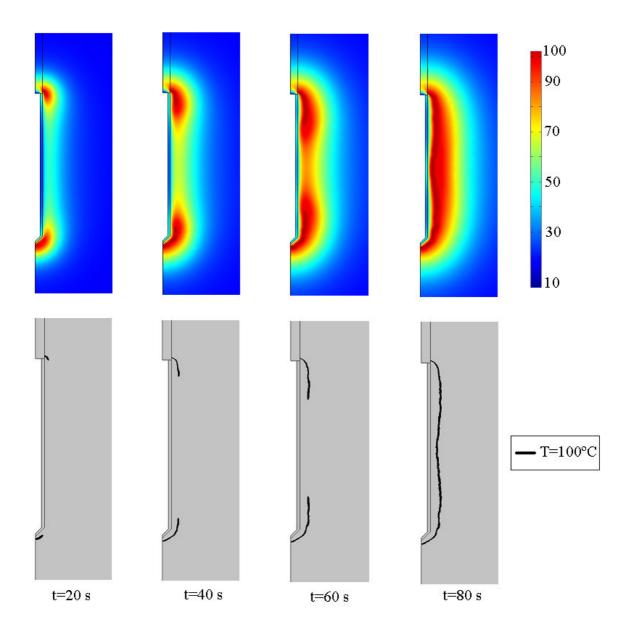


Figure 8 Temperature distributions computed from the theoretical model (top, scale in $^{\circ}$ C) and progress of the 100 $^{\circ}$ C isotherm in the tissue (bottom). Note that the 100 $^{\circ}$ C isotherm completely encloses the electrode at \approx 80 s.