

Document downloaded from:

<http://hdl.handle.net/10251/52430>

This paper must be cited as:

Bernabeu García, A.; Palacios Guillem, S.; Vicente Candela, R.; Vercher Pérez, RF.; Arqués Sanz, A.; Amat Payá, AM. (2012). Solar photo-Fenton at mild conditions to treat a mixture of six emerging pollutants. *Chemical Engineering Journal*. 198-199:65-72. doi:10.1016/j.cej.2012.05.056.



The final publication is available at

<http://dx.doi.org/10.1016/j.cej.2012.05.056>

Copyright Elsevier

Highlights

- Solar photo-Fenton has been able to remove a mixture of six emerging pollutants.
- The reaction was performed at acidic and circumneutral media.
- Reaction rate was highly influenced by the sample matrix.
- Intermediates were detected and related with changes in toxicity.

1 Solar photo-Fenton at mild conditions to treat a mixture of six 2 emerging pollutants

3 A. Bernabeu¹, S. Palacios¹, R. Vicente¹, R. Vercher¹, S. Malato², A. Arques*¹, A.M.
4 Amat¹

5 ¹ Grupo Procesos Oxidación Avanzada, Departamento de Ingeniería Textil y Papelera.
6 Universidad Politécnica de Valencia. Plaza Ferrándiz y Carbonell s/n, E-03801, Alcoy,
7 Spain. aarques@txp.upv.es

8 ² Plataforma Solar de Almería-Ciemat, Carretera de Senés km4, 04200 Tabernas, Spain

9 Abstract

10 The applicability of photo-Fenton to degrade a mixture of emerging pollutants (EPs)
11 namely amoxicillin, acetaminophen, acetemiprid, caffeine, clofibric acid and
12 carbamazepine has been studied at different scenarios. At high concentrations, acidic
13 photo-Fenton was able to achieve a fast removal of the EPs. Although, complete
14 mineralization was not reached, the toxicity of the solution was decreased according to
15 the respiration of activated sludge and luminescence of *V.fischeri* assays, although
16 according to this last assay a transitory enhancement of the toxicity was found,
17 attributable to the formation of toxic by-products such as phenols, chlorophenols and
18 chlorinated pyridines. Experiments carried out with 5 mg/l of each EP showed that at
19 neutral media the process was two orders of magnitude less efficient than at acidic pH,
20 although it was still able to remove the EPs. The aqueous matrix has a remarkable effect
21 on the process as the presence of humic acids increased the reaction rate and inorganic
22 salts played an inhibitory role. Finally, experiments performed with 10 µg/l of each EP
23 showed that under those experimental conditions nearly complete removal of the EPs

24 was reached with neutral photo-Fenton after 60 min of irradiation; in this case, humic
25 substances played a disfavourable role.

26

27 **Keywords:** photo-Fenton, neutral media, emerging pollutants, humic acids,
28 detoxification

29

30 **1. Introduction**

31 In recent years, an important number of synthetic compounds have been found in
32 surface waters at trace amounts, as a consequence of human activity. They are
33 commonly referred to as emerging pollutants (EPs), because they have been recently
34 detected and hence their potential effects on humans and natural ecosystems have not
35 yet been elucidated [1]. Among the substances that can be considered within this group
36 are perfluorinated compounds, pharmaceuticals, hormones, endocrine disruptors,
37 sunscreens, algal toxins, dioxane or nanomaterials [2,3].

38

39 Emerging pollutants have been found in the effluent of wastewater treatment plants,
40 which constitutes a major limitation for their potential reuse. Hence, development of
41 new strategies to deal with this concern are needed. In particular, coagulation-
42 flocculation, adsorption onto activated carbon, biological processes or nanofiltration have
43 been employed, as recently reviewed [4,5]. Oxidative processes are promising methods
44 to deal with this environmental concern [6]: electrochemical methods have been
45 demonstrated to be efficient to remove pharmaceuticals and personal care products, as
46 reviewed by Brillas et al. [7] and, on the other hand, photochemical methods employing

47 sunlight as irradiation source might also be of special interest because of the enhanced
48 sustainability they involve [8]. In this context, the Fenton reagent could be useful. It
49 consists in a mixture of iron salts and hydrogen peroxide at acidic pH; under these
50 conditions, iron catalyzes decomposition of hydrogen peroxide into highly oxidizing
51 species, such as hydroxyl radicals. Although this process occurs in the dark, it is greatly
52 enhanced upon UV-vis irradiation ($\lambda < 500$ nm) and hence, sunlight can be used as
53 irradiation source [9].

54

55 There is some information on the treatment of emerging pollutants by the photo-Fenton
56 process [10]; however, the use of highly acidic pH might be an important drawback for
57 the real applicability of this method. Processes based in the Fenton reagent, including
58 photo-Fenton, are less efficient at circumneutral media than at the optimum acidic pH
59 because of the low solubility of iron at neutral media, but also to a modified mechanism
60 which favours formation of highly oxidized iron species (e.g. ferryl ions) instead of the
61 more reactive hydroxyl radical [11-14]. However this process might be a useful
62 alternative to remove chemicals at $\mu\text{g-ng L}^{-1}$, as it is the case of EPs in effluent of
63 wastewater treatment plants. In fact, some recent work has been devoted to explore the
64 applicability of this methodology for this purpose [15, 16].

65

66 However, further research on this topic is meaningful. As EPs show diverse chemical
67 structures, their behaviour vs. photo-Fenton might be different. In addition, primary
68 removal of the pollutant might not represent detoxification of the effluent and a careful
69 identification of major intermediates and assessment of the toxicity of treated solutions

70 is required. Finally, the presence of natural organic matter (e.g. humic acids) can play
71 an important role in the process that deserves to be investigated.

72

73 With this background, the aim of this work is to gain further insight into the use of
74 circumneutral photo-Fenton as a tertiary process to remove EPs at the effluent of a
75 wastewater treatment plant. Experiments have been performed with a mixture of six
76 EPs, namely amoxicillin (antibiotic), acetaminophen (analgesic), acetamiprid
77 (insecticide), caffeine (stimulating agent), clofibric acid (metabolite of clofibrate, also
78 employed as herbicide) and carbamazepine (psychiatric drug); chemical structures can
79 be found at Scheme 1. Preliminary experiments have been performed at high initial
80 concentrations of the EPs in order to allow a reliable monitoring of the process, then
81 concentrations have been decreased down to 10 µg/l to better reproduce EPs in the
82 effluents of wastewater treatment plants. Changes in the toxicity, major by products
83 formed in the process and the role of humic acids have been studied

84

85 **2. Experimental**

86 **2.1. Reagents**

87 Acetaminophen, caffeine, amoxicillin, clofibric acid, carbamazepine, acetamiprid and
88 humic acid sodium salt (AHA) were purchased from Sigma-Aldrich. Hydrogen
89 peroxide (30% v/v) and ferrous sulphate, used in the photo-Fenton reactions, were
90 obtained from Panreac.

91

92 Water employed in most reactions was Milli-Q grade. In other experiments, EPs were
93 spiked in samples taken from the effluent of the secondary settling tank of a wastewater
94 treatment plant, which consisted in a physical-chemical primary treatment followed by
95 an aerobic activated sludge biological reactor. The main parameters of these samples
96 were systematically inside the following ranges: pH, 7.5; conductivity, 2.4-2.8 mS/cm;
97 dissolved organic carbon (DOC), 15-50 mg/l; inorganic carbon, IC, 70-90 mg/l (which
98 results in a HCO_3^- concentration of ca. 350-450 mg/l) and chemical oxygen demand
99 (COD) 60-120 mg/l.

100

101 2.2. Photo-Fenton experiments

102 Photo-Fenton processes were carried out with the mixture of six EPs; the initial
103 concentration of each EP was between 10 $\mu\text{g/l}$ and 50 mg/l. The amount of iron was 5
104 mg/l, added as iron(II) sulphate. The stoichiometric amount of hydrogen peroxide
105 required to mineralize all the organics was added at the beginning of the experiment.
106 The reaction was carried out at the optimal pH value (2.8), and at pH = 7; sulphuric acid
107 or sodium hydroxide were employed to adjust the pH. Eventually, humic acids (10
108 mg/l) were added to the reaction mixture.

109

110 Samples were irradiated with a solar simulator (Oriel Instruments, Model 81160
111 equipped with a 300 W xenon lamp) in cylindrical open glass vessels. Magnetic stirring
112 was kept all along the reaction time and water was periodically added in order to
113 compensate for the evaporation loss. Temperature was ca. 30°C throughout the
114 experiment. Before analysis all samples were filtered through polypropylene (0.45 μm)
115 and, eventually, preconcentrated following the procedure described below, or

116 neutralized when submitted to bioassays. Those samples used for biological analysis or
117 for COD measurements were free of hydrogen peroxide.

118

119 **2.3. Analysis**

120 The concentration of each EP was determined by UHPLC (Perkin Elmer model Flexar
121 UPLC FX-10) equipped with a Brownlee Analytical column (DB-C18). A mixture of
122 acetonitrile (A) and a 0.1% formic acid aqueous solution (B) was used as eluent. The
123 relative amount of each solvent was changed in a linear gradient, from 3% A to 70% A
124 in 8 min; the flow rate was 0.3 mL/min. Retention times were 1.7 for acetaminophen 2.1
125 for amoxicillin, 3.3 for caffeine, 4.9 for acetamiprid, 5.6 for carbamazepine and 6.2 for
126 clofibric acid. The wavelengths used for the EPs quantitations were: 205 nm
127 (acetaminophen, amoxicillin, caffeine and carbamazepine), 225 nm (clofibric acid) and
128 245 nm (acetamiprid). For the experiments containing 10 µg/L of each EP, the samples
129 were preconcentrated by solid phase extraction (SPE); briefly, 250 ml of sample were
130 flown through a LiChrolut EN 200 mg (Merck) cartridge; then EPs were recovered with
131 3 mL of methanol.

132

133 Eventually, the reaction products were analyzed by GC-MS; a gas chromatograph
134 GCMS-QP2010S (Shimadzu) equipped with a quadrupole mass analyzer was
135 employed. 10 µl of sample obtained after SPE concentration (see above) were injected;
136 the temperature of the injector was 250°C and the split ratio was 1:30. A Meta X5
137 Teknokroma column was used and helium was employed as carrier gas, with a flow rate
138 of 35 cm/s. The column temperature was kept at 60°C for 3 min, then it was increased
139 with a 8 °C/min rate from 60 °C to 280 °C and finally it was kept at 280 °C for 3 min.

140

141 DOC was determined with a Shimadzu model TOC-V CSH apparatus. COD was
142 determined photometrically according to the dichromate method [17]; digestions were
143 performed at 148°C in a Thermoreaktor TR300 (Merck) and a Spectroquant NOVA 60
144 (Merck) was used for the photometric determination. The concentration of iron was
145 monitored according to the o-phenantroline standardized procedure [16]. Briefly, iron
146 is reduced to iron (II) by addition of hydroquinone at a pH = 3.5; then o-phenantroline is
147 added and the red complex formed is determined photometrically at 508 nm. Hydrogen
148 peroxide was estimated by a colorimetric method with test stripes (Merckoquant),
149 purchased to Merck.

150

151 Inhibition of the luminescence of *V. fischeri* assays were performed according to the
152 standardized ISO 11348-3 norm, using lyophilized bacteria (*V. fischeri*, NRRL B-
153 11177, Macherey-Nagel). Reconstitution of the bacteria was performed according to the
154 standard procedure, using a salty solution. The luminescence was determined with a
155 Luminometer Lumifix-Bio-10 (Macherey-Nagel). Toxicity was determined after 15 min
156 incubation. Distilled water and zinc sulphate were used in control experiments. Toxicity
157 of slightly coloured samples, due to the presence of humic substances, was corrected by
158 subtracting their absorbances at 490 nm. Inhibition of the respiration of activated sludge
159 respiration was performed according to an adaptation of the OECD 209 test, as
160 described in detail elsewhere [18], using a BM3-LAB (Neurtek) respirometer equipped
161 with an oxygen sensor (WTW-Cell Ox); Briefly, activated sludge was brought to its
162 maximum oxygen uptake rate (OUR) by addition of highly biodegradable sodium

163 acetate; Then, sample containing the EPs was added and the decrease in the OUR was
164 associated to the toxicity .

165

166 **3. Results and discussion**

167 3.1. Photo-Fenton process at acidic pH

168 The effect of a photo-Fenton process was first checked at acidic medium (pH = 2.8)
169 using an initial high concentration of the target pollutants (50 mg/l). Figure 1 shows that
170 complete abatement of the parent pollutants was achieved after 50 min of irradiation. In
171 fact, more than 90% removal of the pollutants was reached after 15 min, except for
172 acetamiprid, which was more reluctant to this treatment.

173

174 However, the elimination of the pollutants did not result in a complete mineralization of
175 the sample, due to the formation of organic intermediates during the process. Figure 2
176 shows that the elimination of dissolved organic carbon after 60 minutes of irradiation
177 was only moderate (ca. 30%). A more remarkable decrease of chemical oxygen demand
178 was observed during the reaction (ca. 50% in 60 min) indicating that noticeable
179 oxidation occurred. This is more easily appreciated by the carbon oxidation state (COS),
180 which is an intensive parameter that can be considered as a good indicator of the
181 efficiency of the oxidative process [19]. COS can be calculated by equation 1,

182

$$183 \quad COS = 4 - 1.5 \frac{COD}{DOC_0} \quad \text{Equation 1}$$

184

185 where DOC_0 is the initial dissolved organic carbon of the solution and COD is the
186 chemical oxygen demand at the sampling time (both parameters expressed as mg/l).
187 Before $t = 30$ min, there was a fast increase in this parameter, while it remained more
188 stable beyond this point, showing that the photo-Fenton was more efficient at the early
189 stages of the process.

190

191 Another parameter relating DOC and COD is average oxidation state (AOS). It can be
192 calculated by equation 2, which is similar to that employed for COS, but with DOC
193 determined at the sampling time.

194

$$195 \quad AOS = 4 - 1.5 \frac{COD}{DOC} \quad \text{Equation 2}$$

196

197 Also in this case, a noticeable variation in this parameter was observed until $t = 30$ min
198 (see Figure 2), which can be associated to changes in the composition of dissolved
199 organics, being more oxidized as the process proceeded [20] (note that AOS only refers
200 to dissolved organics, as DOC at the sampling time is used for calculations, while CO_2
201 formed by mineralization is also included with an oxidation state +4 in the COS, as
202 initial DOC is used in this case).

203

204 Changes in the dissolved organic matter might result in a concomitant variation in the
205 toxicity of the sample. This was studied using two different methods, namely inhibition
206 of the respiration of activated sludge and inhibition of the luminescence of *V. fischeri*.

207 Results in Figure 3 show that according to the activated sludge assay, the initial toxicity
208 was ca. 50% and the sample was detoxified after 10 min of irradiation. In sharp contrast
209 with this, luminescence assays showed an increase in the toxicity in the first stages of
210 the process, reaching a maximum after 10 minutes of reaction. Then, toxicity decreased
211 to reach values below 10% after 40 min. This quantitative divergence in the toxicity
212 values is not surprising, as this parameter strongly depends on the bioassay that has
213 been employed [21]. In particular, differences between activated sludge and *V. fischeri*
214 assays have been previously observed [22] and they are attributable to the sensibility of
215 the microorganisms that are employed: activated sludge consists in a consortium of
216 microorganisms present in biological reactors of wastewater treatments plants and
217 hence, they have developed some adaptation and resistance to chemical contaminants;
218 on the other hand, *V. fischeri* bacteria show high sensibility towards pollutants [23].
219 Using 50 mg/l as the initial concentration of each EP is far above those typically found
220 in the effluent of wastewater treatment plants; hence, detoxification observed at these
221 high concentrations might indicate that applying photo-Fenton at $\mu\text{g/L}$ level to this
222 mixture of EPs will not produce any worrying effect on toxicity.

223

224 It seems interesting to gain further insight into the changes in toxicity determined
225 according to the *V. fischeri* assay. First, the EC_{50} was calculated for each pollutant, as
226 well as their individual toxicity at the concentration employed in the experiment (see
227 Table 1). It is interesting to note that the toxicity of the untreated sample (ca. 60%) was
228 well above the cumulative effect of the individual contaminants (below 25%), indicating
229 that there is some synergetic effect due to the mixture of all six emerging pollutants.
230 The trends observed in the *V. fischeri* experiments along the photo-Fenton process can
231 be explained by the formation of toxic by-products in the oxidation of the parent EPs.

232 This behaviour has been already observed by other authors in advanced oxidation
233 processes; for instance, Oturan et al. associated an initial increase in the toxicity of a
234 solution containing diuron submitted to an electro-Fenton treatment to the formation of
235 more toxic oxidation by-products [24]. In order to investigate this point, GC-MS
236 analyses were performed with samples taken after different periods of reaction (see
237 table 2). Although it is difficult to associate the enhancement of the toxicity in such a
238 complex mixture to the formation of a single compound, it is remarkable that significant
239 amounts of phenols, chlorophenols and chlorinated pyridines, were detected in those
240 samples showing higher toxicity (t = 5-15 min of treatment). In fact, the toxicity of all
241 six EPs used in the work (EC₅₀ of hundreds or even thousands mg/l, see Table 1) is
242 well below that found in literature for chlorinated aromatics, whose EC₅₀ are typically
243 below some tens of mg/l [25, 26]. Formation of these chlorinated by-products can be
244 attributed to the photo-oxidation of clofibric acid and acetamiprid. In this context, it has
245 been reported that no significant detoxification or even some increase in the toxicity has
246 been observed along the reaction of acetamiprid with •OH radicals or singlet oxygen
247 [27, 28].

248

249 3.2. Photo-Fenton at neutral media

250

251 Although using 50 mg/l as the initial concentration of each EP is useful to obtain a
252 better monitoring of the reaction, these values are far above those typically found in the
253 effluents of wastewater treatment plants. Lower EP concentrations might allow using
254 milder reaction conditions and hence, testing other strategies might be meaningful,
255 namely, the use of the Fenton reagent at circumneutral media. In a first step, a more

256 diluted mixture of the six emerging pollutants (5 mg/l) was treated at the pH value of
257 2.8 and at circumneutral pH (7.2). Figures 4A and 4H show that at the optimal acidic
258 pH all the pollutants were removed after 15 min of treatment, while under neutral
259 conditions irradiation times were noticeably higher, as some EPs remained in the
260 solution after 6 hours. In order to obtain more quantitative data, the kinetic profiles
261 obtained for each pollutant were fitted to a semilogarithmic plot and the pseudo-first
262 order rate constants were calculated from the slope of this plot. Rate constants obtained
263 at neutral pH (ranging from 0.002 min^{-1} for acetamiprid to 0.015 min^{-1} for clofibric
264 acid) were two orders of magnitude lower than at the optimal pH (which were between
265 0.17 min^{-1} for acetamiprid and 0.72 for amoxicillin); the worse performance of the
266 photochemical process at neutral medium is attributable to the low solubility of iron
267 salts at this pH, changes in the speciation of dissolved iron and to the existence of an
268 alternative reaction mechanism, which favors formation of less reactive iron(IV) species
269 instead of hydroxyl radicals [11]. However, the photochemical elimination of the EPs in
270 the neutral media in the presence of the Fenton reagent is faster than in the case of
271 photolysis (Figure 4F). Only in the case of clofibric acid, which suffers very fast
272 photolysis under the studied conditions, and acetamiprid, similar reaction rates were
273 measured in neutral photo-Fenton and photolysis.

274

275 In order to gain further insight into the mechanism at neutral medium, experiments were
276 carried in the presence of t-butanol, a well known hydroxyl radical scavenger. In order
277 to simplify the system, experiments were carried out only with caffeine; this compound
278 was chosen because suffers negligible photolysis and reaction is slow enough to allow
279 the measurement of accurate kinetic data. Pseudo first order rate constants were
280 determined with and without t-butanol at neutral pH; for the sake of comparison these

281 experiments were also driven at the optimal pH = 2.8 (Figure 5). In both cases, a
282 decrease in the reaction rate was observed in the presence of t-butanol, however, it was
283 more remarkable at acidic medium as k was 0.45 min⁻¹ without t-butanol and 0.018 min⁻¹
284 ¹ in the presence of the radical scavenger, what means a 96% decrease, in sharp contrast,
285 only 59% decrease was measured at neutral medium (0.027 min⁻¹ and 0.011 min⁻¹
286 respectively). This is in agreement with a higher formation rate of ·OH at acidic pH
287 according to a classical photo-Fenton process, while a modified mechanism occurs at
288 circumneutral pH favoring the formation of other oxidizing agents, such as ferryl
289 species.

290

291 Other species found in the aqueous matrix (e.g. inorganic ions or dissolved organic
292 matter) might show some influence on neutral photo-Fenton. Regarding to the effect of
293 organic matter, studying the effect of humic substances appeared interesting, as this
294 group of compounds are the most widely distributed organic materials [29] and they are
295 photochemically active; for this purpose, humic acids (10 mg/l) were added to mixture
296 of the EPs. Comparison between Figures 4 A and B indicates that faster removal of the
297 pollutants was reached in the presence of the humic substances. This effect has been
298 previously observed in the removal of other chemicals by Fenton process [30, 31] and a
299 complex formed between iron and the humic substances was proposed as the key
300 species in the process. Recently, a significant increase in the photo-Fenton reaction rate
301 has been observed in the presence of humic acids [32]; however, according to literature,
302 ·OH radical have been found not to play a major role in the process and hence, other
303 oxidizing species might be responsible for the pollutants oxidation [12]. In fact,
304 experiments carried out in the presence of t-butanol with caffeine as model compound
305 showed that in the presence of humic acids, the decrease in the rate constant was only

306 37% (data not shown), even lower than the 59% recorded at neutral media without
307 humic acids; this is in agreement with an alternative mechanism to $\cdot\text{OH}$ playing a more
308 relevant role under these conditions. Despite the possible modification of the
309 mechanism at neutral pH and in the presence of humic substances, the Fenton reagent
310 plays the major role in the process and hydrogen peroxide is the actual oxidizing agent,
311 as irradiation of a solution containing humic acids and iron (but without H_2O_2) gave
312 similar reaction profiles than photolysis of the EPs (compare Figures 4F and 4G).

313

314 Experiments were carried out using tap water, with and without humic substances
315 (Figure 4C and 4D, respectively). In this case, lower reaction rates were obtained,
316 showing that the sample matrix played an inhibitory role. This can be attributed to the
317 presence of inorganic salts in the medium, which have been described to inhibit the
318 (photo)-Fenton reaction, probably because of their scavenging role towards the reactive
319 species involved in the process. In this context, the presence of carbonates (mainly
320 HCO_3^-) at neutral pH, have been reported to inhibit the photo-Fenton process [33] as it
321 is well known that CO_3^{2-} and HCO_3^- compete with organic contaminants for hydroxyl
322 radical reactions, and significantly decrease the pollutants degradation [34]; hence, the
323 remaining reactivity detected under these conditions might be attributed to the existence
324 of alternative oxidizing species, as indicated by the t-butanol experiments. In addition to
325 this, carbonates have been reported to change iron speciation, which might also have a
326 remarkable influence on the performance of the Fenton system [35].

327

328 Finally, an effluent from a wastewater treatment plant (see experimental section for
329 main parameters) was spiked with 5 mg/l of each EP and submitted to a neutral photo-

330 Fenton process (Figure 4E). In this complex sample, different effects are expected to
331 occur: a) the presence of inorganic salts might inhibit the photo-Fenton process, b)
332 organics able to form photo-active complexes with iron might increase the reaction rate
333 and c) other organics could compete with the EPs for the reactive species resulting in a
334 lower reaction rate. Comparison of Figures 4A and 4E indicates that the removal of the
335 EPs was slower than in distilled water, meaning that the inhibitory effects are
336 predominating. In fact, results are only slightly better than those achieved with tap
337 water. However, the EPs removal is still faster than for photolysis, showing that reactive
338 species generated by neutral photo-Fenton are responsible for some EPs oxidation.

339

340 Toxicity was measured in all cases according to the *V. fischeri* assay. The initial values
341 were in this case ca. 5%, and some increase was determined at the beginning of the
342 process, attributable to the formed by-products; nonetheless toxicities were
343 systematically below 20 %. As the concentration of EPs found in the effluents of
344 wastewater treatment plants are several orders of magnitude below those employed in
345 these experiments, a hypothetical increase in the toxicity along the photo-Fenton
346 process should not be considered a major threat.

347

348 3.3. Neutral photo-Fenton at low EPs concentrations

349

350 Experiments above reported have been conducted at concentrations well above those
351 commonly found at the effluents of wastewater treatment plants. Hence, it is convenient
352 to explore the behavior of those compounds at lower concentrations. For this purpose, a

353 series of experiments were performed in distilled water with the mixture of EPs, at an
354 initial concentration of 10 µg/l of each contaminant, in the presence and absence of
355 humic acids. The percentages of removal for the reaction with and without humic acids,
356 together with the photolysis of the EPs after 30 and 120 min of irradiation are given in
357 Figure 6. Neutral photo-Fenton enhanced significantly the elimination of
358 acetaminophen, carbamazepine and caffeine; this was not the behavior of clofibric acid,
359 which suffered the fastest photolysis among the studied EPs, and acetamiprid, which
360 was the most reluctant to photo-Fenton. Finally, removal of amoxicillin was very fast in
361 all cases, as its concentration was below the quantitation limit after 30 min.

362

363 Regarding to humic substances, the presence of these organics did not result in an
364 improved performance of the photo-Fenton, in sharp contrast with observations with the
365 mixture containing 5 mg/l of each EP. This could be due to a competitive role of humic
366 acids for the reactive species formed in the process which compensates for the potential
367 enhanced formation of reactive species: as the employed amount of humics was the
368 same in both situations (10 mg/l), in the more diluted solution, the ratio EP/humic
369 substances decreases and hence, reaction of the generated oxidative species becomes
370 more likely with the humic substances than with the EPs.

371

372 **4. Conclusions**

373 Photo-Fenton process at mild conditions has been able to treat a mixture of six
374 emerging pollutants at moderate reaction times when they are at low concentrations.
375 The aqueous matrix had a remarkable influence in the process; in particular, humic
376 substances played a favorable role in the process at high EPs concentrations, although

377 the opposite behavior was observed as the EPs/humics ratio decreased, most probably
378 due to a scavenging effect of these substances. Despite some toxic by-products have
379 been detected when experiments were conducted at high concentration of the EPs, this
380 is not a major **drawback**, as they are eliminated after longer reaction times and, more
381 important, toxicities are very low at the concentration ranges commonly reported for
382 EPs at the effluents of wastewater treatment plants.

383

384 **Acknowledgements**

385 We want to thank the financial support of Spanish Ministerio de Ciencia e Innovación
386 (CTQ 2009-13459-C05-03) and (CTQ 2009-13459-C05-01)

387

388 **References:**

389

390 [1] K.E. Murray, S.B. Thomas, A.A. Badour, Prioritizing research for trace pollutants
391 and emerging contaminants in the freshwater environment, *Environ. Pollut.* 158 (2010)
392 3462-3471.

393 [2] S.D. Richardson, Environmental mass spectrometry: emerging contaminants and
394 current issues. *Anal. Chem.* 80 (2008) 4373-4402.

395 [3] M. Petrovic, D. Barceló, Liquid chromatography-mass spectrometry in the analysis
396 of emerging environmental contaminants, *Anal. Bioanal. Chem.* 385 (2006) 422-424.

397 [4] N. Bolong, A.F. Ismail, M.R. Salim, T. Matsuura, A review of the effects of
398 emerging contaminants in wastewater and options for their removal, *Desalination*, 239
399 (2009) 229-246.

- 400 [5] Z. Liu, Y. Kanjo, S. Mizutani, Removal mechanisms for endocrine disrupting
401 compounds (EDCs) in wastewater treatment – physical means, biodegradation, and
402 chemical advanced oxidation: a review, *Sci. Total Environ.* 407 (2009) 731-748.
- 403 [6] S.K. Khetan, T.J. Collins, Human pharmaceuticals in the aquatic environment: a
404 challenge to green chemistry, *Chem. Rev.* 107 (2007) 2319-2364.
- 405 [7] E. Brillas, I. Sirés, M. A. Oturan, Electro-Fenton process and related electrochemical
406 technologies based on Fenton's reaction chemistry, *Chem. Rev.*, 109 (2009) 6570-6631.
- 407 [8] S. Malato, P. Fernández-Ibáñez, M.I. Maldonado, J. Blanco, W. Gernjak,
408 Decontamination and disinfection of water by solar photocatalysis: Recent overview
409 and trends, *Catal. Today* 147 (2009) 1-59.
- 410 [9] J.J. Pignatello, E. Oliveros, A. MacKay, Advanced oxidation processes for organic
411 contaminant destruction based on the Fenton reaction and related chemistry, *Critical*
412 *Rev. Environ. Sci. Technol.* 36 (2006) 1-84.
- 413 [10] M. Klavarioti, D. Mantzavinos, D. Kassinos, Removal of residual pharmaceuticals
414 from aqueous systems by advanced oxidation processes, *Environ. Int.* 35 (2009) 402-
415 412.
- 416 [11] S.J. Hug, O. Leupin, Iron-catalyzed oxidation of arsenic(III) by oxygen and by
417 hydrogen peroxide: pH dependent formation of oxidants in the Fenton reaction.
418 *Environ. Sci. Technol.* 37 (2003) 2734-2742.
- 419 [12] A.W. Vermilyea, B.M. Voelker, Photo-Fenton reaction at near neutral pH.
420 *Environ. Sci. Technol.* 43 (2009) 6927-6933.

- 421 [13] A.L. Pham, C. Lee, F.M. Doyle, D. Sedlak, A silica-supported iron oxidative
422 catalyst capable of activating hydrogen peroxide at neutral pH values. *Environ. Sci.*
423 *Technol.* 43 (2009) 8930-8935.
- 424 [14] S. Pang, J. Jiang, J. Ma, Oxidation of sulfoxides and arsenic(III) in corrosion of
425 nanoscale zero valent iron by oxygen: evidence against ferryl ions (Fe(IV)) as active
426 intermediates in Fenton reaction, *Environ. Sci. Technol.* 45 (2011) 307-312.
- 427 [15] N. Klammerth, L. Rizzo, S. Malato, M.I. Maldonado, A. Agüera, A.R. Fernández-
428 Alba, Degradation of fifteen emerging contaminants at $1\mu\text{g L}^{-1}$ initial concentrations by
429 mild solar photo-Fenton in MWTP effluents. *Water Res.* 44 (2010) 545-554.
- 430 [16] A. Bernabeu, R.F. Vercher, L. Santos-Juanes, P.J. Simón, C. Lardín, M.A.
431 Martínez, J.A. Vicente, R. González, C. Llosá, A. Arques, A.M. Amat, Solar
432 photocatalysis as a tertiary treatment to remove emerging pollutants from wastewater
433 treatment plant effluents, *Catal. Today* 161 (2011) 235-240.
- 434 [17] APHA AWWA WPCF Standard methods for the examination of water and
435 wastewater (20th ed.). American Public Health Association. Washington, DC, 1998
- 436 [18] A.M. Amat, A. Arques, A. García-Ripoll, L. Santos-Juanes, R. Vicente, I. Oller, M.
437 I. Maldonado, S. Malato, A reliable monitoring of the biocompatibility of an effluent
438 along an oxidative pre-treatment by sequential bioassays and chemical analyses, *Water*
439 *Res.* 43 (2009) 784-792.
- 440 [19] A.M. Amat, A. Arques, F. Galindo, M.A. Miranda, L. Santos-Juanes, R. Vercher,
441 R. Vicente, Acridine yellow as solar photocatalyst for enhancing biodegradability and
442 eliminating ferulic acid as model pollutant. *Appl. Catal. B: Environ.* 73 (2007) 220-226.

- 443 [20] V. Sarria, S. Parra, N. Adler, P. Peringer, N. Benitez, C. Pulgarin, Recent
444 developments in the coupling of photoassisted and aerobic biological processes for the
445 treatment of biorecalcitrant compounds. *Catal. Today* 76 (2002) 301-315.
- 446 [21] L. Rizzo, Bioassays as a tool for evaluating advanced oxidation processes in water
447 and wastewater treatment. *Water Res.* 45 (2011) 4311-4340.
- 448 [22] A.M. Amat, A. Arques, A. García-Ripoll, L. Santos-Juanes, R. Vicente, I. Oller,
449 M.I. Maldonado, S. Malato, A reliable monitoring of the biocompatibility of an effluent
450 along an oxidative pre-treatment by sequential bioassays and chemical analyses. *Water*
451 *Res.* 43 (2009) 784-792.
- 452 [23] M. Gutiérrez, J. Etxebarria, L. de las Fuentes, Evaluation of wastewater toxicity:
453 comparative study between microtox® and activated sludge oxygen uptake inhibition,
454 *Water Res.* 36 (2002) 919-924.
- 455 [24] N. Oturan, S. Trajkovska, M.A. Oturan, M. Couderchet, J.J. Aaron, Study of the
456 toxicity of diuron and its metabolites formed in aqueous medium during application of
457 the electrochemical advanced oxidation process "electro-Fenton", *Chemosphere*, 73
458 (2008) 1550-1556.
- 459 [25] M. Ma, Z. Tong, Z. Wang, W. Zhu, Acute toxicity bioassay using the freshwater
460 luminescent bacterium *Vibrio-qinghaiensis* sp. Nov.-Q67. *Bull. Environ. Contam.*
461 *Toxicol.* 62 (1999) 247-253.
- 462 [26] M. Krebel, H. Kusic, N. Koprivanac, J. Meixner, A.L. Bozic, Treatment of
463 chlorophenols by UV-based processes: correlation of oxidation by-products, wastewater
464 parameters and toxicity, *J. Environ. Eng.* 137 (2011) 639-649.
- 465 [27] M.L. Dell'Arciprete, L. Santos-Juanes, A. Arques, R. Vicente, A.M. Amat, J.P.
466 Furlong, D.O. Mártire, M.C. Gonzalez, Reactivity of hydroxyl radicals with

467 neonicotinoid insecticides: mechanism and changes in toxicity. *Photochem. Photobiol.*
468 *Sci.* 8 (2009) 1016-1023.

469 [28] M.L. Dell'Arciprete, L. Santos-Juanes, A. Arques, R.F. Vercher, A.M. Amat, J.P.
470 Furlong, D.O. Mártire, M.C. Gonzalez, Reactivity of neonicotinoid insecticides with
471 singlet oxygen. *Catal. Today* 151 (2010) 137-142.

472 [29] R. Sutton, G. Sposito, Molecular structure in soil humic substances: the new view,
473 *Environ. Sci. Technol.* 39 (2005) 9009-9015.

474 [30] E. Lipczynska-Kochany, J. Kochany, Effect of humic substances on the Fenton
475 treatment of wastewater at acidic and neutral pH, *Chemosphere* 73 (2008) 745-750.

476 [31] Georgi, A., Schierz, A., Trommler, U., Horwitz, C.P., Collins, T.J., Kopinke, F.D.
477 (2007) Humic modified Fenton reagent for enhancement of the working pH range.
478 *Appl. Catal. B: Environ.* 72, 26-36.

479 [32] N. Klamerth, S. Malato, M.I. Maldonado, A. Agüera, A.R Fernández-Alba,
480 Modified photo-Fenton for degradation of emerging contaminants in municipal
481 wastewater effluents. *Catal. Today*, 161 (2011) 241-246.

482 [33] N. Klamerth, N. Miranda, S. Malato, A. Agüera, A.R. Fernández-Alba, M.I.
483 Maldonado, J.M. Coronado, Degradation of emerging contaminants at low
484 concentrations in MWTPs effluents with mild photo-Fenton and TiO₂, *Catal. Today*,
485 144 (2009) 124-130.

486 [34] G. V. Buxton, A. J. Elliot, Rate constant for reaction of hydroxyl radicals with
487 bicarbonate ions, *Radiat. Phys. Chem.* 27 (1986) 241 - 243.

488 [35] D.W. King, R. Farlow, Role of carbonate speciation on the oxidation of Fe(II) by
489 H₂O₂. *Mar. Chem.* 70 (2000) 201-209.

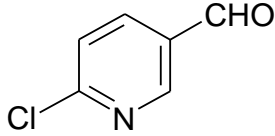
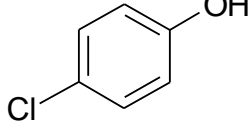
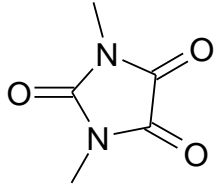
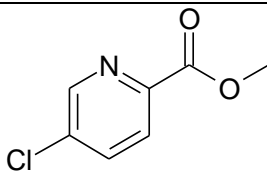
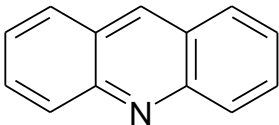
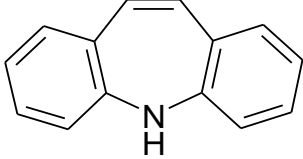
490 Table 1. EC₅₀ and inhibition caused by 50 mg/l of each EP, determined according to the
491 inhibition of luminescence of *V. fischeri* bacteria.

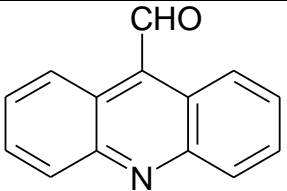
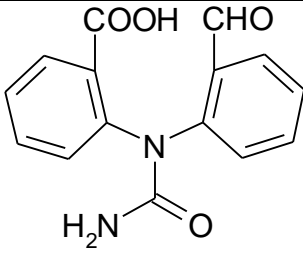
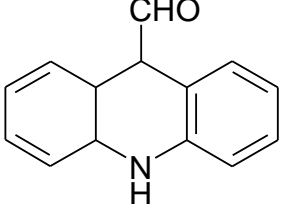
Compound	EC ₅₀ (mg/l)	% Inhibition caused by 50 mg/l
Acetaminophene	2600	>1
caffeine	3700	>1
Acetamidiprid	110	20
Amoxicillin	1350	>1
Carbamazepine	280	>1
Clofibric acid	280	>1

492

493

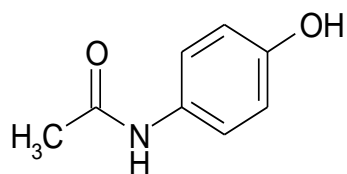
494 Table 2. Organic by products detected in sampes taken at diferent periods of a photo-
 495 Fenton process at acidic pH under simulated sunlight (initial concentration of each EP =
 496 50 mg/l. Compounds are qualitatively devided into major (M), noticeable or tare (T)
 497 amounts depending on the peak area. When possible, a parent EC has been assigned to
 498 each intermediate.

tr	Compound	t= 5min	t= 10 min	t=15 min	t=30 min	Parent compound
11.4		T	T	T	T	Acetamiprid
12.0		M	M	N	-	Clofibric acid
12.1		T	N	T	N	Caffeine
13.5	Unidentified chlorophenol	-	T	-	-	
13.8		-	-	-	T	
16.5	Unidentified chlorophenol	N	N	N	-	Acetamiprid
19.0	Clofibric acid	T	-	T	-	-
20.8	Acetaminophene	M	M	M	-	
21.6		T	T	-	-	Carbamazepine
22.3	Caffeine	M	M	N	-	-
24.0		N	N	T	-	Carbamazepine

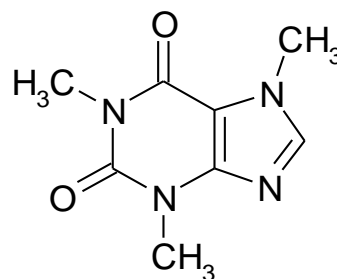
25.0		N	N	T	-	Carbamazepine
27.2		T	-	-	-	Carbamazepine
27.6		N	N	T	-	Carbamazepine
28.4	Carbamazepine	M	N	N	-	-
29.6	Acetamidiprid	M	M	M	N	-

499

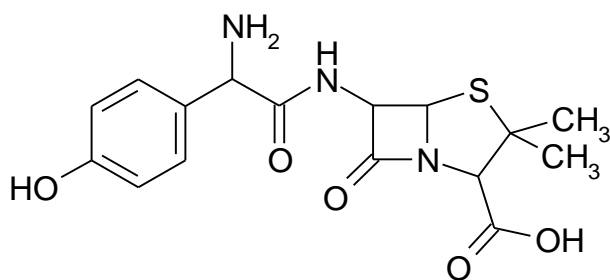
500



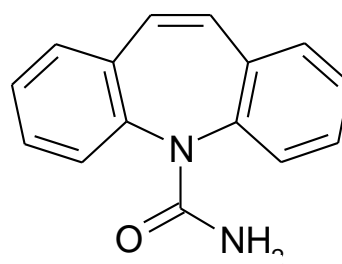
Acetaminophen



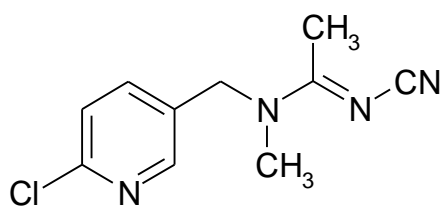
Caffeine



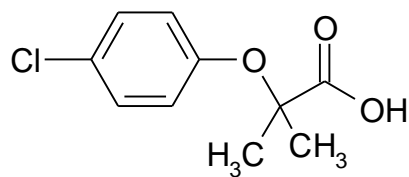
Amoxicillin



Carbamazepine

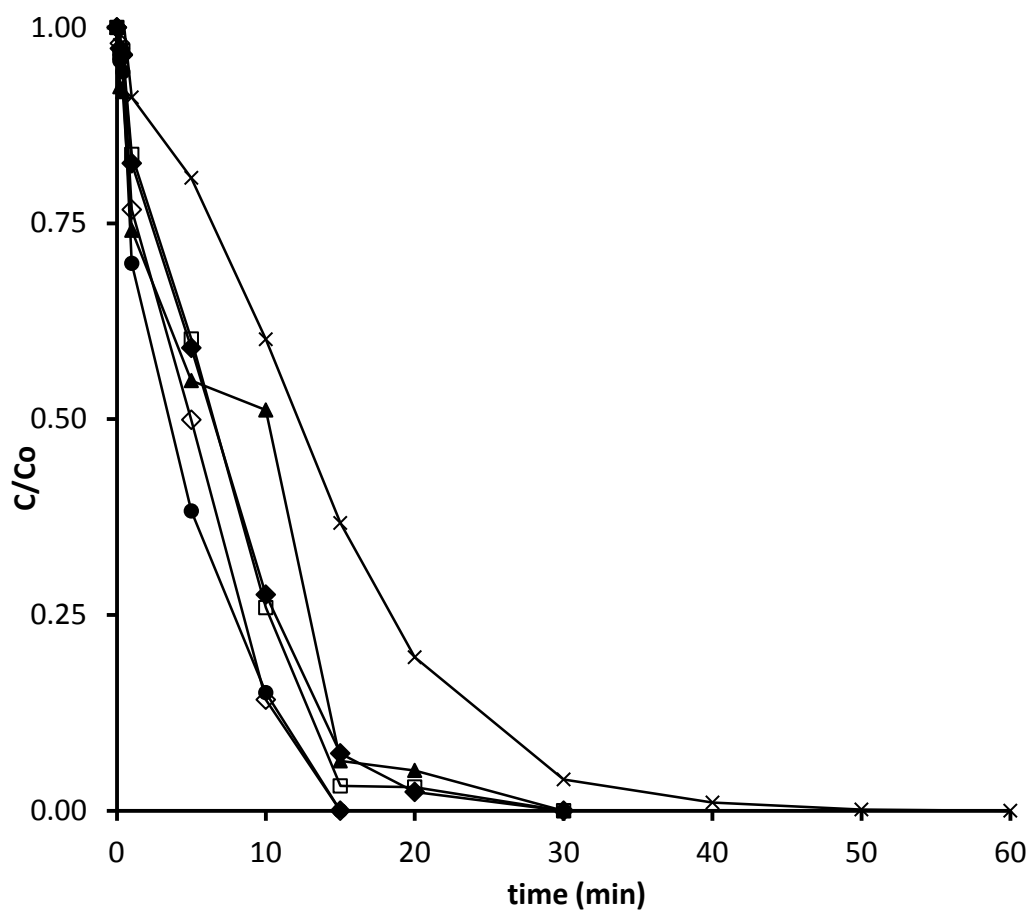


Acetamiprid



Clofibric acid

502



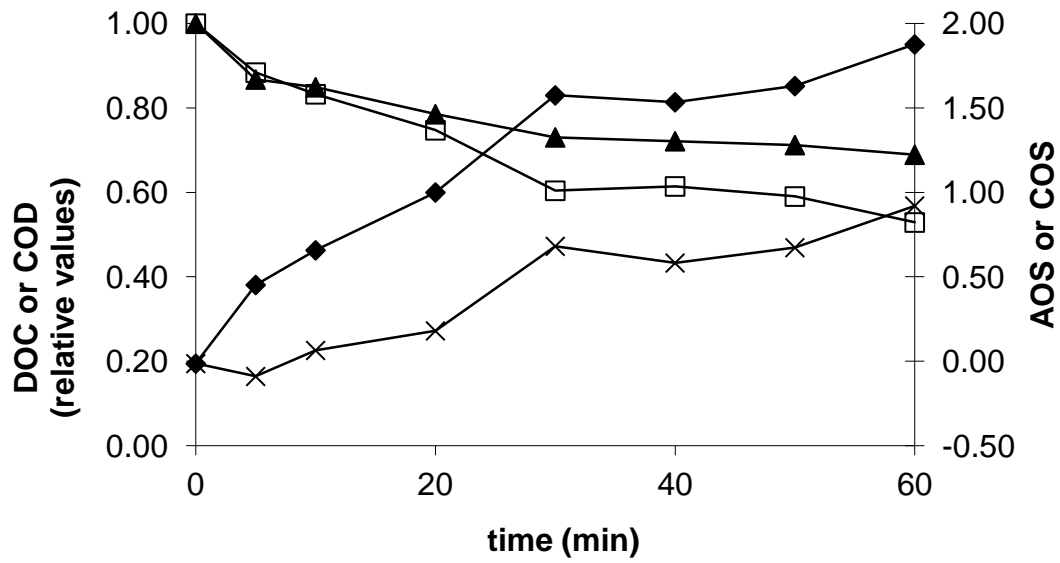
503

504 Figure 1. Photodegradation of a mixture of 6 EPs with an initial concentration of 50
505 mg/l by means of a photo-Fenton process at acidic medium under simulated sunlight.

506 Plot of the relative concentration vs time: amoxicillin (▲), acetaminophen (●),

507 **acetamiprid** (×), caffeine (□), clofibric acid (◆) and carbamazepine (◇).

508

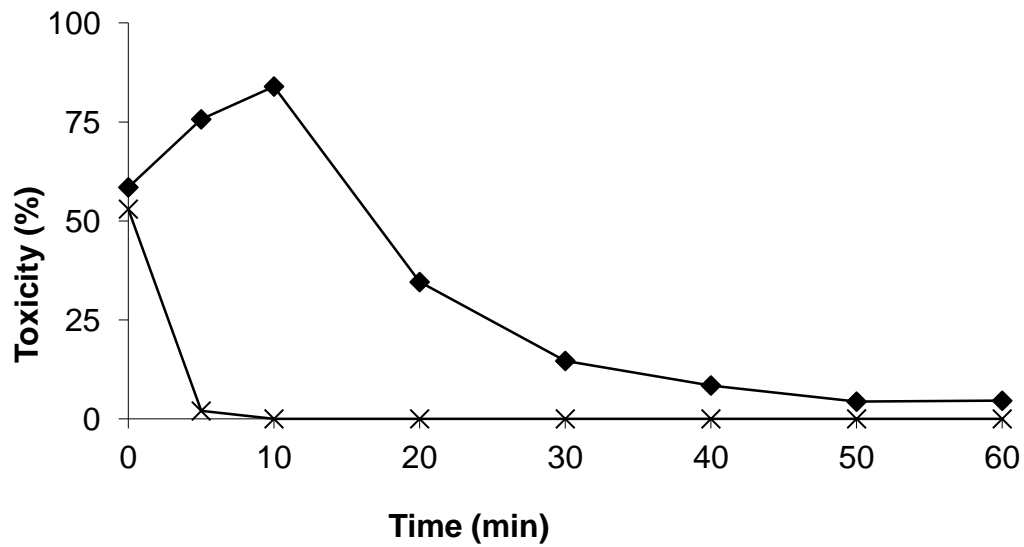


509

510

511 Figure 2. Photodegradation of a mixture of 6 EPs with an initial concentration of 50
 512 mg/l by means of a photo-Fenton process at acidic medium under simulated sunlight.
 513 Plot of changes in COD (□), DOC (▲), AOS (×) and COS (◆) vs. irradiation time.

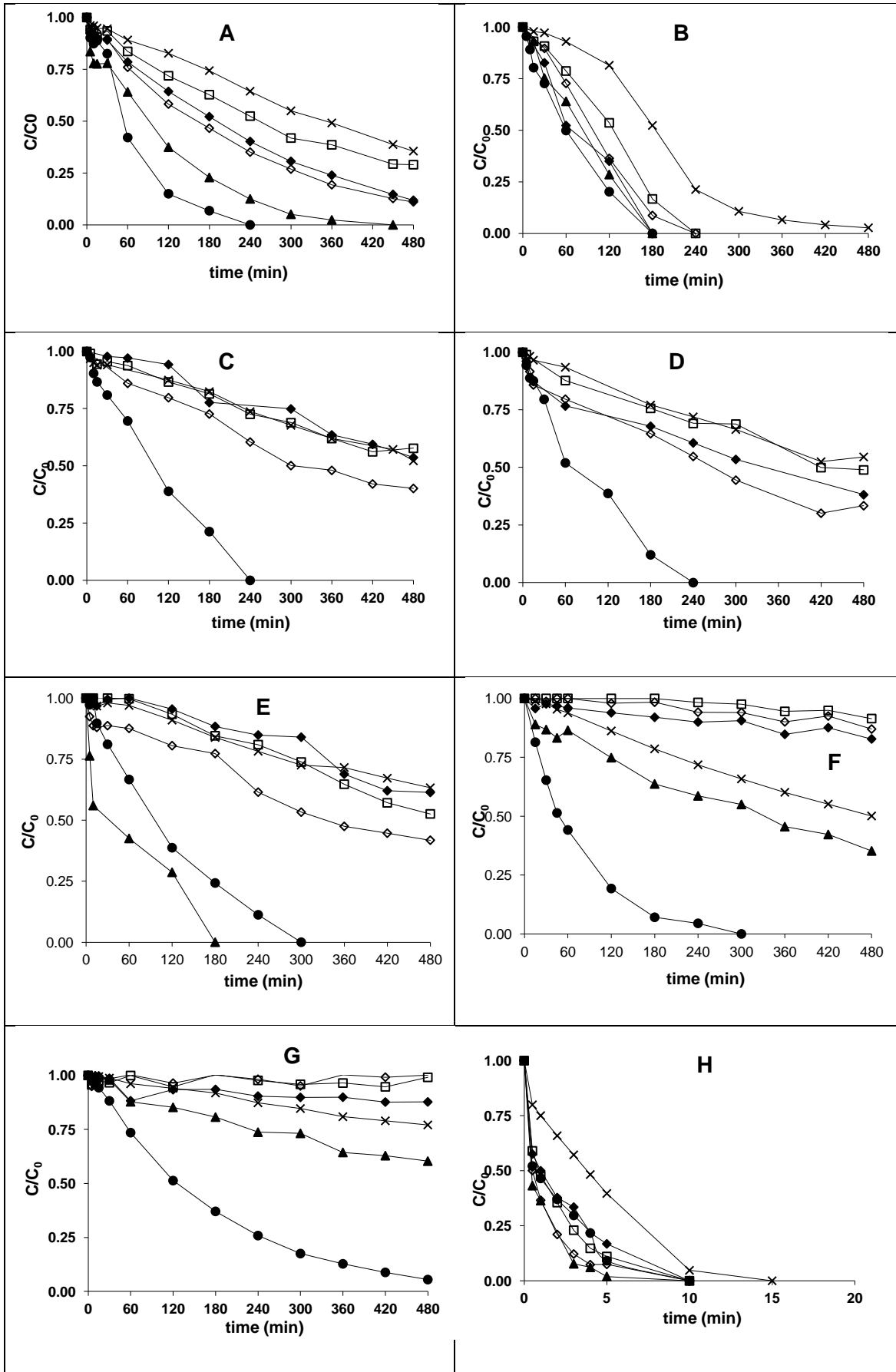
514



515

516 Figure 3. Changes in the toxicity according to the *V. fischeri* assay (◆) and inhibition of
 517 the respiration of activated sludge (×) of a mixture of 6 EPs with an initial
 518 concentration of 50 mg/l of each EP submitted to a photo-Fenton process at acidic
 519 medium under simulated sunlight.

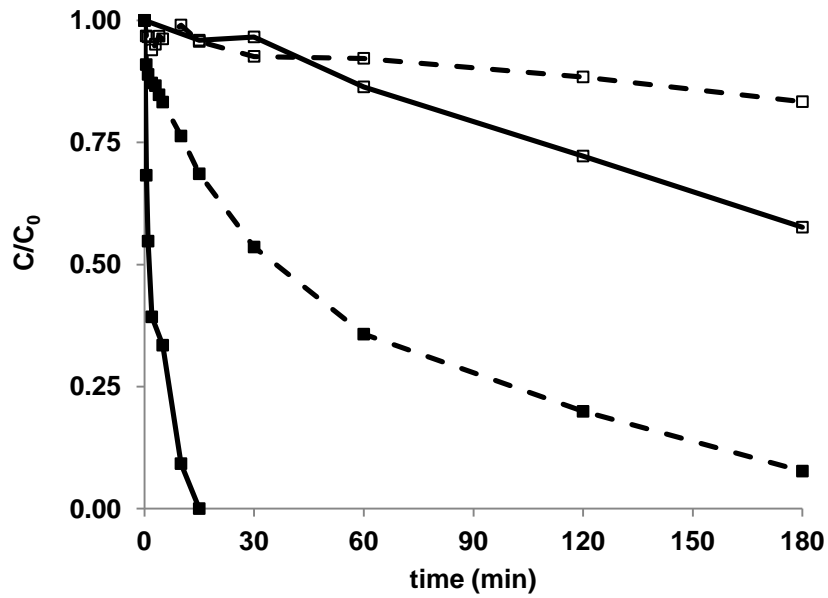
520



521 Figure 4: Photodegradation of a mixture of 6 EPs with an initial concentration of 5 mg/l
522 under simulated sunlight. Plot of the relative amounts of amoxicillin (▲),
523 acetaminophen (◆), acetemiprid (×), caffeine (□), clofibric acid (●) and carbamazepine
524 (◇). Experimental condition were: neutral photo-Fenton in distilled water (A), neutral
525 photo-Fenton in distilled water and humic acids (B), neutral photo-Fenton in tap water
526 (C), neutral photo-Fenton in tap water and humic acids (D), neutral photo-Fenton
527 spiking the EPs in a wastewater treatment plant effluent (E), photolysis in distilled
528 water (F), photolysis in the presence of humic acids and iron (G), and photo-Fenton at
529 pH = 2.8 in distilled water (H, note the different scale).

530

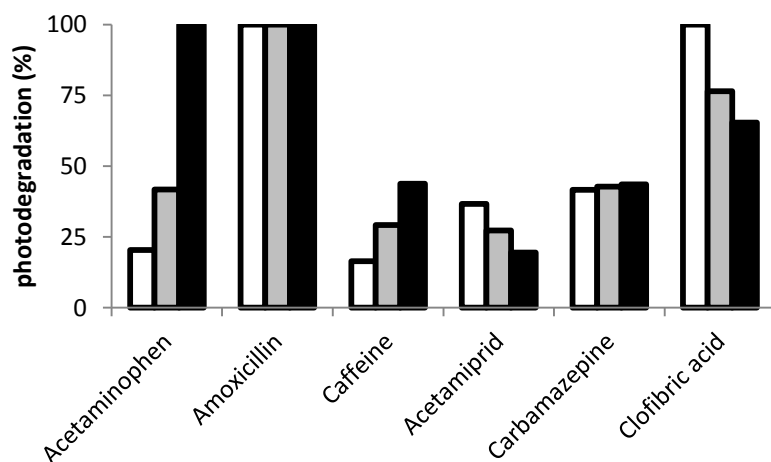
531



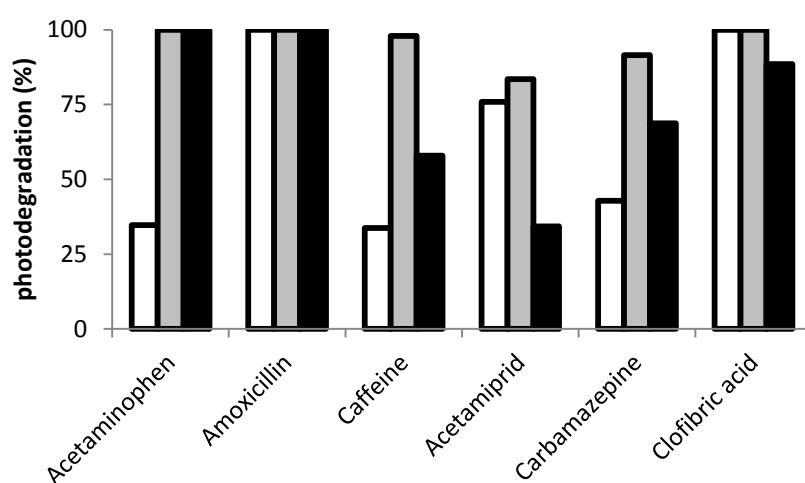
532

533 Figure 5: plots of the concentration of caffeine during a photo-Fenton process with
534 (discontinuous line) and without (continuous line) t-butanol at acidic (black symbols) or
535 neutral (empty symbols) media.

536



537



538

539 Figure 6: Photodegradation of a mixture of 6 EPs with an initial concentration of 10 $\mu\text{g/l}$
 540 by means of a photo-Fenton process at neutral pH under simulated sunlight. Percentage
 541 of photodegradation achieved after 30 min (above) and 120 min (below) of irradiation.
 542 Reactions were carried out without (grey bars) and with 10 mg/l of humic acids, (black
 543 bars). Photolysis of the EPs is also shown (white bars)

544

