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Additional Information

1 **A robust rabbit line increases leukocytes counts at weaning and reduces mortality**  
2 **by digestive disorder during fattening**

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12

13 **Abstract**

14 The aim of this study was to determine how genetic type could affect the physiological  
15 and immune status of commercial rabbits at weaning, as well as their performance and  
16 health during the growing period. The study was conducted on a total of 2904 young  
17 rabbits weaned at 30 days, belonging to three different genetic types (line H, founded for  
18 litter size at birth and selected for litter size at weaning during 17 generations; line LP,  
19 founded for reproductive longevity criteria and selected for litter size at weaning for 7  
20 generations; and line R, founded and selected during 25 generations for average daily  
21 gain from the 4<sup>th</sup> to the 9<sup>th</sup> week of life). Two different diets were used during lactation.  
22 The two diets were both isoenergetic and isoproteic but their main energy source differed,  
23 being either animal fat (AF) or cereal starch (CS). Leukocyte subsets were characterised  
24 at weaning, and growing performance was studied until 58 days of age (feed intake, live  
25 weight, mortality by digestive disorders and morbidity) for both medicated and non-  
26 medicated dietary versions. At weaning, young rabbits fed an AF lactating diet evidenced  
27 greater B lymphocyte count (+ 46%) than those fed a CS diet. Blood from LP rabbits had  
28 higher counts for total B, T CD5<sup>+</sup> and CD8<sup>+</sup> lymphocytes with respect to H and R (on av.  
29 +40, +57, +28, and +27%, respectively; P<0.05), and CD4<sup>+</sup> lymphocytes, monocytes and  
30 granulocytes with respect to R (on av. +24, +32 and +44%, respectively; P<0.05) at  
31 weaning. LP line rabbits also showed lower mortality by digestive disorders (on av. -8  
32 points of percentage) and morbidity (on av. -4 points) than those from H and R lines  
33 during the growing period (P<0.05). R animals presented higher feed intake and daily  
34 weight gain, and a lower feed conversion ratio than H and LP animals (on av. +16.7±2.7  
35 g dry matter/day, +10.3±0.4 g/day and -0.22±0.04 g dry matter/g, respectively). In  
36 conclusion, the foundation of a line for reproductive longevity, which has been previously  
37 reported to give greater robustness (low environmental sensitivity) to their reproductive

38 stock, could have also conferred a greater immunological development at weaning to their  
39 offspring, as well as a better ability to confront digestive disorders as compared to other  
40 lines founded or selected exclusively for productive criteria.

41 **Keywords:** Litter size; Growth rate; Robustness; Leukocytes subsets; Mortality; Rabbit.

42

## 43 **Introduction**

44 Maintaining the right balance between health and production rates during the growing  
45 period is one of the main targets in rabbit farming. The rise in digestive disorders during  
46 this period, such as Epizootic Rabbit Enteropathy (ERE), has increased the dependence  
47 on the use of antimicrobials and has reduced farm profitability in recent decades. Several  
48 studies have been conducted to define the aetiology of this illness (Licois et al., 2006) and  
49 to develop adequate feeding systems to minimize sanitary risk during the fattening period  
50 (Falção-e-Cunha et al., 2007; Gidenne et al., 2010). Genetic type could also condition the  
51 body condition and health of females, as well as the predisposition of the litter to suffer  
52 digestive disorders. In fact, a maternal effect on the development of digestive microbiota  
53 (Abecia et al., 2007) and on the rate of digestive disorders in growing rabbits (Quevedo  
54 et al., 2003; Carabaño et al., 2006) has previously been described.

55 Several studies have indicated a line founded by productive longevity has increased  
56 ability to overcome productive (Theilgaard et al., 2009), environmental (Savietto et al.,  
57 2013) and immunological challenges with LPS (Ferrian et al., 2013). The concept of  
58 robustness in farm animals was defined by Knap (2005) as ‘the ability to combine a high  
59 production potential with resilience to stressors, allowing for unproblematic expression  
60 of a high production potential in a wide variety of environmental conditions’. Therefore,  
61 this line founded by productive longevity could be considered as robust. This robust line  
62 was characterized by greater modulation of their body resources under heat stress  
63 conditions, when the immune system is widely affected (Ferrian et al., 2012), and  
64 improved ability of the immune system to respond when faced with a specific  
65 immunological challenge (Ferrian et al. 2013). Therefore, the use of genetic types  
66 characterized by a greater robustness could contribute to improving the general health  
67 status of the farm.

68 Weaning happens when the immune system is still being developed, and the transition to  
69 solid feeding frequently leads to increased digestive disorders and mortality (Rosell and  
70 De la Fuente, 2009). The adequate performance of young rabbits during the growing  
71 period could be also affected by their degree of maturity at this time. The immune system  
72 in young rabbits is subjected to several changes during the first weeks of life. Increased  
73 levels of red (RBC) and white blood cells (WBC) are related to greater maturity (Jeklova  
74 et al., 2009), as well as changes in leukocyte populations in peripheral blood. On the other  
75 hand, the type of feed given to rabbits during the lactation period affects the amount of  
76 milk ingested, the promotion of solid feed intake and probably the degree of maturity at  
77 weaning. Fat enriched diets usually increase milk yield of females and live weight (LW)  
78 of kits at weaning (Pascual et al., 2003), but could lead to a less progressive weaning  
79 process.

80

81 The aim of the present study was to evaluate how three genetic types founded and selected  
82 for different criteria (reproduction, robustness or growth rate) and the dietary energy  
83 source of the lactation feed (fat or starch) could have affected the maturity and leukocyte  
84 populations in peripheral blood at weaning, as well as their subsequent performance and  
85 health status during the growing period.

86

87 **Material and methods**

88 *Animals*

89 The experimental procedure was approved by the animal welfare ethics committee of the  
90 Universitat Politècnica de València (UPV) and carried out following the European Union (2003)  
91 recommendations on care and protection of animals used for experimental purposes and the  
92 advice for applied nutrition research in rabbits according to the European Group on Rabbit  
93 Nutrition (Fernández-Carmona et al., 2005). The experiment involved a total of 325 litters from  
94 196 female rabbits studied at the most until third parity (from January to August of 2012). A total  
95 of 2904 young rabbits weaned at 30 d were used. Weaned rabbits belonged to three different  
96 genetic types from the Institute for Animal Science and Technology of the Universitat Politècnica  
97 de Valencia: line H, founded for litter size at birth and selected for litter size at weaning during  
98 17 generations (n=807); line LP, founded for reproductive longevity criteria by selecting females  
99 from commercial farms that had a minimum of 25 parturitions with more than 7.5 kits born alive  
100 per parity (more details of LP line constitution are given in Sánchez et al., 2008) and then selected  
101 for litter size at weaning for 7 generations (n=1311), characterized by a high robustness  
102 (Theilgaard et al., 2009; Ferrián et al., 2012, 2013); and line R, founded and selected during 25  
103 generations for average daily gain from the 4<sup>th</sup> to the 9<sup>th</sup> week of life (n=786).

104 *Diets*

105 Two experimental diets were formulated according to the recommendations of De Blas and  
106 Mateos (2010) for reproductive rabbit does. The two diets were meant to be both isoenergetic and  
107 isoproteic [approx. 11.6 MJ of digestible energy (DE) and 117 g of digestible protein per kg of  
108 dry matter (DM)], but were to have a differing main energy source. Diet CS was prepared using  
109 cereal starch [247 g of starch and 21 g of ether extract (EE) per kg DM], whereas in diet AF, part  
110 of starch was replaced by animal fat (104 g of starch and 85 g of EE per kg DM). After weaning,  
111 two versions of the same commercial diet were used: non-medicated (NM) and medicated [M; 40  
112 ppm of tiamulin fumarate A (Caliermutin 2%, Laboratorios Calier S.A., Barcelona, Spain), 120  
113 ppm of neomycin sulphate (Hipramix 14%, Hipra S.A., Girona, Spain), 29 ppm of lincomycin

114 hydrochloride and 29 ppm of spectinomycine sulphate (Linco-spectin 880, Pfizer, Madrid,  
115 Spain)]. Both diets included 66 ppm of robenidine as coccidiostat.

116 Chemical analyses of diets were performed according to the methods of the Association of  
117 Official Analytical Chemists (2000): 934.01 for DM, 942.05 for ash, 976.06 for crude protein and  
118 920.39 for EE, with acid-hydrolysis of samples prior to the extraction. Starch content was  
119 determined according to Batey (1982), by means of a two-step enzymatic procedure with  
120 solubilisation and hydrolysis to maltodextrins with thermostable  $\alpha$ -amylase followed by complete  
121 hydrolysis with amyloglucosidase (both enzymes from Sigma-Aldrich, Steinheim, Germany), and  
122 the resulting glucose being measured using the hexokinase/glucose-6 phosphate dehydrogenase/  
123 NADP system (R-Biopharm, Darmstadt, Germany). Neutral detergent fibre (NDF), ADF and acid  
124 detergent lignin (ADL) fractions were analysed sequentially (Van Soest et al., 1991) with a  
125 thermo-stable  $\alpha$ -amylase pre-treatment and expressed exclusive of residual ash, using a nylon  
126 filter bag system (Ankom, Macedon, NY, USA). The ingredients of the reproduction diets and  
127 chemical composition of all the diets are shown in the Table 1.

### 128 *Experimental design*

129 At first parturition, females of each genetic type were randomly allocated to both reproduction  
130 diets, which were provided ad libitum throughout the whole experiment. Females and their litters  
131 received the same reproduction diet. After weaning at 30 days of age, young rabbits from the  
132 same litter were identified by tattoo and housed together in collective cages until 58 days of age.  
133 During this growing period, two thirds of the litters received the M diet and one third the NM  
134 diet, provided ad libitum and randomly allocated from within genetic type and reproduction diet.  
135 Blood samples of 3 kits per litter from 136 females in the first reproductive cycle were extracted,  
136 mixed and processed as a unique sample at weaning for subsequent flow cytometry analysis  
137 (n=408). Samples were drawn from the median artery of the ear using vacuum EDTA tubes.  
138 Diurnal variations in haematological parameters were minimized by collecting blood at  
139 approximately the same time (8:00-9:00 h). Mortality by digestive disorders was studied daily  
140 (n=2904). Individual LW and litter feed intake were studied at 30, 44 and 58 days of age for 1231  
141 growing rabbits. An animal was considered as morbid when there were sings of illness and/or



142 abnormally low growth compared with animals to the same group based on a subgroup analysis  
143 (SAS, 2002).

#### 144 *Flow cytometry analysis*

145 Blood was processed 1 h after sampling. Before performing flow cytometry, white blood cell  
146 (WBC) count and percentage of total lymphocytes were determined using a haematology analyzer  
147 (MEK-6410, Nihon Kohden, Japan). A millilitre of whole blood was pipetted into a 50 mL tube.  
148 WBC were isolated lysing erythrocytes by adding 40 mL of ammonium chloride lysing solution  
149 (8.02 g NH<sub>4</sub> Cl, 0.84 g NaHCO<sub>3</sub>, and 0.37 g EDTA per liter of Millipore water) at 4°C. After 6  
150 min incubation in the dark, samples were centrifuged at 400g for 5 min at room temperature. The  
151 supernatant was carefully discarded and the pellet was resuspended in 1 mL of phosphate-buffered  
152 saline (PBS). The cells number of the suspension was adjusted by counting with Neubauer  
153 Chamber to 10<sup>6</sup> cells per mL. Primary monoclonal antibodies were added (Table 2), and incubated  
154 for 25 min at room temperature in the dark. Then, the pellet was washed with 1 mL of PBS, and  
155 centrifuged again under the same conditions mentioned above. Afterwards, secondary antibodies  
156 (Rat anti-mouse IgG2a+b Phycoerythrin [VMRD, Inc.  $\alpha$ -exalpha] and Goat anti-mouse IgM: R-  
157 Phycoerythrin [AbD Serotec]) were added, and incubated for 20 min at room temperature in the  
158 dark. Finally, 1 mL of PBS was added before running the flow cytometer. The outcome WBC  
159 suspensions were analysed in a Cytomics FC500 flow cytometer (Beckman Coulter, Brea, CA).  
160 The common leukocyte antigen CD14 and CD45 expression was used for the “lymphogate” setup  
161 as previously described (Jeklova et al., 2007; Guerrero et al., 2010). Total lymphocyte count was  
162 calculated from WBC count and lymphocyte percentage, and lymphocyte subset counts as  
163 described by Guerrero et al. (2010).

#### 164 *Statistical analysis*

165 Maturity Index (MI) at weaning was calculated as the ratio between LW at weaning and mature  
166 weight. Mature weight for each line (4082, 4105 and 5645 g for H, LP and R line, respectively)  
167 was obtained from the Gompertz modelling of the LW of reproductive rabbit does until 12 months  
168 of age (Arnau, personnel communication). Data from leukocyte subset counts averaged per litter  
169 and from individual MI at weaning were analysed using a general linear model (SAS, 2002),

170 including the genetic line (H, LP, R), the reproduction diet (CS, AF) and their interaction as fixed  
171 effects, as well as the parity order (primiparous, multiparous) for only MI at weaning.

172 Data on growing performance were analysed using a mixed procedure (SAS, 2002), in a repeated  
173 measure design which allows variance among animals and the intra-animal covariance to be  
174 considered, modelled with a compound symmetry function. Random terms included the  
175 permanent effect of each animal or litter ( $p$ ) and the error term ( $e$ ), both assumed to have an  
176 average of zero and a variance of  $\sigma_p^2$  and  $\sigma_e^2$ , respectively. Analysis of LW and daily gain data  
177 was performed excluding morbid animals (to separate the effect of feed and genetic type from the  
178 effect of illness), with a model including as fixed effects the genetic line (H, LP, R), the  
179 reproduction diet (CS, AF), the parity order (primiparous, multiparous), the growing diet (NM,  
180 M), the control day (30, 44, 58 for LW and 44, 58 for daily gain) and their interactions. Feed  
181 intake and feed conversion ratio (FCR) were only analysed from 186 litters receiving M diet  
182 excluding morbid or dead animals, with a model similar to that used for daily gain data excluding  
183 the growing diet as a fixed effect.

184 Orthogonal contrasts were computed to test the differences between lines (H–LP, H–R and LP–R)  
185 and reproduction diets (AF–CS).

186 Mortality by digestive disorders and morbidity data were analysed using the genmod procedure  
187 (SAS, 2002), with a binomial probability distribution and a logit transformation  $[\ln(\mu/1-\mu)]$  as  
188 link function. The model included the genetic line (H, LP, R), the reproduction diet (CS, AF), the  
189 growing diet (M, NM), the control day (35, 42, 49, 58) and their interactions.

190

191 **Results**

192 Table 3 shows the LW, MI and blood leukocyte counts of young weaned rabbits at 30  
193 days of age. Young rabbits in the AF dietary group had greater LW and MI (+8%;  
194  $P<0.001$ ) at weaning than those in the CS group, especially in the case of animals from  
195 H and LP lines. Dietary group did not affect main blood leukocyte counts at weaning, but  
196 AF animals showed a greater count of B lymphocytes (+46%;  $P<0.05$ ). Regarding genetic  
197 line, H weaning rabbits had lower LW than those from the LP and R lines (on av. -12%;  
198  $P<0.001$ ). The highest MI at weaning corresponded to LP animals (15.3 % of the adult  
199 weight), followed by H (13.6%;  $P<0.001$ ), R showing the lowest MI (11.1%;  $P<0.001$ ).  
200 Blood from LP weaning rabbits showed higher counts for total, B, T CD5<sup>+</sup> and CD8<sup>+</sup>  
201 lymphocytes in comparison to H and R rabbits (on av. +40, +57, +28, and +27%,  
202 respectively;  $P<0.05$ ), and higher CD4<sup>+</sup> lymphocytes, monocytes and granulocytes with  
203 respect to R (on av. +24, +32 and +44%, respectively;  $P<0.05$ ). In addition, blood from  
204 H weaning rabbits had a higher CD4<sup>+</sup>/CD8<sup>+</sup> ratio than the LP and R lines (on av.  
205  $+0.40\pm 0.12$ ;  $P<0.01$ ), and higher CD25<sup>+</sup> lymphocyte and monocyte counts than the R line  
206 (on av. +93 and +33%, respectively;  $P<0.05$ ).

207 Table 4 shows the performance of young rabbits during the growing period. The type of  
208 diet offered during lactation did not affect main growth performance traits. Only daily  
209 weight gain from 30 to 44 days of age was higher for animals from the CS group (+6%;  
210  $P<0.001$ ). As expected, R rabbits showed higher feed intake, daily weight gain and lower  
211 FCR than H and LP throughout the whole growing period (on av.  $+16.7\pm 2.7$  g DM/day,  
212  $+10.3\pm 0.4$  g/day and  $-0.22\pm 0.04$  g DM/g, respectively;  $P<0.001$ ). The difference in LW  
213 at weaning between LP and H lines was maintained at slaughter time ( $+81\pm 12$  g in favour  
214 of LP animals;  $P<0.05$ ).

215 All dead and morbid animals presented digestive disorders compatible with ERE. As can  
216 be seen in Table 5, animals from the LP line showed lower mortality (on av. -8 points of  
217 percentage) and morbidity (on av. -4 points) than those from H and R lines during the  
218 growing period ( $P<0.05$ ). Type of diet offered during lactation did not affect subsequent  
219 mortality and morbidity, but R animals from the AF group had higher mortality (+7 points  
220 of percentage;  $P<0.05$ ) than those from CS (Figure 1). The inclusion of antimicrobials in  
221 the growing diet significantly reduced the mortality registered during the growing period  
222 (-19 points of percentage;  $P<0.05$ ). However, this effect was different depending on the  
223 genetic line (Figure 2). Although the LP line showed the lowest cumulated mortality  
224 regardless of the diet, the highest cumulated mortality was registered in H animals when  
225 fed the M diet (3.6, 7.9 and 14.6 for LP, R and H, respectively;  $P<0.05$ ), but on R animals  
226 when fed the NM diet (20.8, 27.8 and 37.1 for LP, H and R, respectively;  $P<0.05$ ).

227

## 228 **Discussion**

229 As was expected, an animal fat enriched diet (AF) led to a higher milk yield in females,  
230 resulting in better and greater development of kits during the lactation period (Pascual et  
231 al., 2002). In fact, these young rabbits reached weaning with higher LW and increased B  
232 lymphocyte counts. Milk could be exerting a protecting effect, as several compounds in  
233 the milk could promote the proliferation of B lymphocytes in lactating animals (Juto,  
234 1985; Orlando, 1995; Tuailon et al., 2009). In this regard, Bienertova-Vasku et al. (2012)  
235 described a direct link between the B-cell activating factor (BAFF) secretion and leptin  
236 plasma level, concluding that BAFF expression was tightly related to adipose tissue. In  
237 fact, BAFF plasma level was also significantly correlated with the energy derived from

238 the diet, therefore the higher milk energy output could be responsible for the greater B  
239 lymphocytes counts in AF weaned rabbits.

240 However, higher LW and increased B lymphocyte counts at weaning do not seem to give  
241 any additional advantage in the subsequent performance during the fattening period. Due  
242 to the compensatory growth observed in the CS group there was no difference in LW  
243 between diets at slaughter time, mortality by digestive disorders and morbidity rates being  
244 similar in both dietary groups. In this respect, Quevedo et al. (2006) suggested that  
245 lactating kits with higher milk availability, which usually lead to lower feed intake at late  
246 lactation, could result in more sudden weaning (less progressive transition from milk to  
247 solid feeding). This fact might be related to the gut maturity of each animal, as the lower  
248 the milk and the more the feed intake before weaning, the more the digestive tract  
249 develops. Frequently, lactating rabbits showing earlier solid intake present a higher food  
250 intake and growth during fattening, as well as lower digestive incidences (Maertens and  
251 De Groote, 1990; Fortun-Lamothe et al., 2001; Pascual et al., 2001). In fact, animals from  
252 the R line, which showed the lowest MI, had higher mortality by digestive disorders  
253 during the fattening period when they had access to a lactation diet promoting milk yield.

254 As expected, and previously reported by Gómez et al. (1992), animals from the genetic  
255 lines selected for daily gain showed better growth performance traits than those from the  
256 lines founded and/or selected for reproductive criteria. Otherwise, a line characterized by  
257 high robustness (LP), but also reproductive performance, showed traits which were  
258 similar to the other maternal line (H). These results concur with those obtained in a large  
259 scale trial (n=344,608; Mínguez et al., 2011), where similar or even better growing  
260 performance in LP rabbits was reported in comparison to other maternal lines (A, V and  
261 H).

262 On the other hand, foundation criterion seems to affect the maturity and leukocyte  
263 peripheral blood counts, being both higher at weaning for the animals from the line  
264 characterized by greater robustness (LP). Higher MI was due to their heavier weight at  
265 weaning as compared to the other maternal line (H) and to their lighter mature weight  
266 with respect to the paternal line (R). In young rabbit females, Theilgaard et al. (2005,  
267 2009) already described a greater LW for LP line animals than in the case of other  
268 maternal lines at the beginning of their reproductive life (approx. +5%), and better  
269 maturity at first mating has frequently been related to higher life span and robustness in  
270 sows (Tarrés et al., 2006) and rabbit females (Sánchez et al., 2008).

271 The higher blood leukocyte counts observed for LP young rabbits at weaning are  
272 consistent with those previously reported for female rabbits by Ferrian et al. (2012), where  
273 total lymphocytes were significantly higher for LP animals when compared to another  
274 maternal line. Based on the results of this study it can be concluded that the use of a line  
275 characterized by higher robustness could improve the ability of the animals to cope with  
276 usual farm diseases. In fact, females of this robust line showed more efficient resource  
277 allocation and immunological response, allowing them to maintain reproductive  
278 performance in the long term even when subjected to different eventual environmental  
279 challenges, such as high productive level (Theilgaard et al., 2009), heat stress (Ferrian et  
280 al., 2012; Savietto et al., 2014), feed restriction (Savietto et al., 2014), or  
281 lipopolysaccharide inoculation (Ferrian et al., 2013).

282 On the other hand, several studies have hypothesised that some selection criteria based  
283 solely on production traits, such as reproduction or growth rate, could have affected the  
284 ability of their offspring to face immunological challenges or stress conditions  
285 (Clapperton et al., 2005; Pascual et al., 2012). Weaned rabbits of the line selected for  
286 growth rate (R) had lower levels of innate immune cells (monocytes) than maternal lines,

287 which have been reported to be more heritable than adaptive immune cells counts (Edfors-  
288 Lilja et al., 1994 and 1998). Moreover, apparent negative genetic correlation between  
289 growth performance traits and innate immune cell counts has been reported in pigs  
290 (Clapperton et al., 2008). These results might suggest that the possible reduction in innate  
291 immunity associated with selection for growth performance might negatively affect the  
292 ability of the animals to confront pathogens.

293 In addition, weaned rabbits from lines founded and selected for productive criteria (H and  
294 R) presented lower adaptive immune cells counts than those from the robust line. In other  
295 species, a reduction in lymphocyte peripheral blood counts was reported in lines  
296 characterized by high growth rate (in turkeys, Huff et al., 2005), hyper-prolificacy (in  
297 Chinese sow lines, Clapperton et al., 2005) or higher milk energy output (in cows, Banos  
298 et al., 2013). Higher lymphocytes counts in LP animals could be a characteristic of the  
299 robust line, but heritability of adaptive immune cells counts have been described as  
300 moderate (Clapperton et al., 2008; Thompson-Crispi et al., 2012). These higher counts  
301 could be also related to the previously mentioned greater maturity of LP animals, which  
302 might have a more mature immune system than animals from the other lines at the same  
303 age. In fact, Jeklova et al. (2009) described how the number of total lymphocytes in the  
304 peripheral blood of SPF rabbits rises during the first 10 weeks of life until reaching adult  
305 values, while monocytes and neutrophils counts were not affected by age.

306 The inclusion of four antibiotics in the feed leads to a decrease in mortality by digestive  
307 disorders of 19 points of percentage in the present study. These results concur with those  
308 presented by Chamorro et al. (2007), who evaluated some of the main factors affecting  
309 performance and health in growing rabbits, reporting the use of antibiotics as the best  
310 option for reducing mortality by digestive disorders (-21 points of percentage).  
311 Therefore, these results evidence the dependence on antimicrobials in environments

312 characterized by high rates of digestive disorders, being the most efficient way to reduce  
313 the incidence of these disorders that highly affect farm profitability. However, the effect  
314 of genetic type on mortality by digestive disorders and morbidity of growing rabbits in  
315 this trial should be highlighted. Using animals from a robust line alone led to an average  
316 reduction of 13 points of percentage in the sanitary risk index (mortality by digestive  
317 disorders plus morbidity). Quevedo et al. (2006) described a higher sanitary risk index  
318 (+4 percentage points;  $P < 0.05$ ) for the three-way crossbred young rabbits (A×V×R) from  
319 current generations with respect to those from previous generations (on av. 12 generations  
320 of difference). Thus, selection for exclusively production criteria could have increased  
321 the susceptibility of young rabbits to common pathogens (as described in other species,  
322 i.e. in pigs, Frank et al., 1997), and the introduction of genetic resources characterized by  
323 a greater robustness could contribute to reducing the dependence on the use of  
324 antimicrobials against infectious diseases.

325 However, although LP rabbits always had the lowest rates, the mortality by digestive  
326 disorders of the other two lines was different depending on antimicrobial use, being  
327 higher for the R animals which were not medicated and for H rabbits when feed was  
328 medicated. It would be expected that animals of the R line with a higher feed intake had  
329 a higher sanitary risk when fed with a non-medicated diet, as weaning could be less  
330 progressive and the amount of nutrients reaching cecum would be increased (including N  
331 which has been related with higher ERE incidence; Villamide et al., 2010). In fact, feed  
332 restriction is a common strategy to reduce the ERE incidence (Gidenne et al., 2009). On  
333 the contrary, higher feed intake of R animals when the diet contained antimicrobials led  
334 to a higher dosage per animal as compared to the H line (+21%), which could explain the  
335 inversion in the mortality rate between both lines.



336 In conclusion, the results of the present study show how the foundation of a rabbit line  
337 for reproductive longevity criteria, which has been previously reported to give a greater  
338 robustness to their reproductive stock, might have also conferred positive attributes to  
339 their offspring. These young rabbits would have greater maturity and immune status at  
340 weaning, showing a better ability to confront the digestive disorders associated with an  
341 ERE context as compared to other lines founded or selected exclusively for productive  
342 criteria. The implementation of these lines at the commercial farms might be considered  
343 of interest for a possible improvement of general farm health and a decreased dependence  
344 on antimicrobials.

345

346

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355

356 **References**

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**Table 2.** Monoclonal antibodies used in this study.

Monoclonal antibodies	Isotype	Specificity	Cell labeling	Clone	References	Company
Mouse anti-rabbit T lymphocytes: FITC <sup>a</sup>	IgG1	CD5	T cell	KEN-5	Kotani et al., 1993	Abd Serotec
Mouse anti-rabbit $\alpha$ -pan B	IgM	IgM	B cell	MRB143A	Davis and Hamilton, 2008	VMRD, Inc.
Mouse anti-rabbit CD4	IgG2a	CD4	T cell subset	KEN-4	Kotani et al., 1993	Abd Serotec
Mouse anti-rabbit $\alpha$ - CD8	IgG2a	CD8	T cell subset	ISC27A	Davis and Hamilton, 2008	VMRD, Inc.
Mouse anti-rabbit CD25	IgG2b	CD25	Activated T cells	KEI-ALPHA1	Kotani et al., 1993	Abd Serotec
Mouse anti-human CD14: FITC	IgG2a	CD14	Monocytes and granulocytes	TÜK4	Jacobsen et al., 1993	Abd Serotec
Mouse anti-rabbit $\alpha$ - CD45	IgM	CD45	All leukocytes	ISC76A	Davis and Hamilton, 2008	VMRD, Inc.

<sup>a</sup> Clon KEN-5 recognises rabbit T lymphocytes and immunoprecipitates. This antibody recognises rabbit CD5, but does not bind to rabbit CD5 transfectants. Known rabbit CD5 antibodies also show binding to most B lymphocytes, which are not labelled by this clone (information obtained from datasheet).



**Table 3.** Effect of genetic line and reproductive diet on the status of young rabbits at weaning (30 days of age)

Lines <sup>1</sup>	H			LP		R		Contrasts <sup>3</sup>			
	No. <sup>5</sup>	AF	CS	AF	CS	AF	CS	AF-CS	H-LP	H-R	LP-R
Reproduction diet <sup>2</sup>											
Live weight (g)	1231	593 <sup>b</sup>	515 <sup>a</sup>	661 <sup>d</sup>	606 <sup>bc</sup>	636 <sup>cd</sup>	622 <sup>c</sup>	48 ± 10 <sup>***</sup>	-79 ± 12 <sup>***</sup>	-75 ± 14 <sup>***</sup>	4 ± 12
Maturity index <sup>4</sup>	1231	0.144 <sup>c</sup>	0.127 <sup>b</sup>	0.159 <sup>d</sup>	0.147 <sup>c</sup>	0.114 <sup>a</sup>	0.109 <sup>a</sup>	0.011 ± 0.002 <sup>***</sup>	-0.017 ± 0.002 <sup>***</sup>	0.024 ± 0.002 <sup>***</sup>	0.041 ± 0.002 <sup>**</sup>
Leukocytes counts (10 <sup>6</sup> /L):											
Total lymphocytes	129	1820 <sup>bc</sup>	1368 <sup>ab</sup>	1944 <sup>c</sup>	2175 <sup>c</sup>	1342 <sup>a</sup>	1344 <sup>ab</sup>	73 ± 139	-465 ± 168 <sup>**</sup>	251 ± 176	716 ± 166 <sup>***</sup>
Lymphocytes B	129	25.3 <sup>ab</sup>	15.8 <sup>a</sup>	35.3 <sup>b</sup>	25.3 <sup>ab</sup>	21.1 <sup>a</sup>	14.9 <sup>a</sup>	8.6 ± 3.5 <sup>*</sup>	-9.8 ± 4.2 <sup>*</sup>	2.5 ± 4.4	12.3 ± 4.1 <sup>**</sup>
Lymphocytes T CD5 <sup>+</sup>	129	910 <sup>a</sup>	958 <sup>a</sup>	1048 <sup>a</sup>	1293 <sup>b</sup>	922 <sup>a</sup>	881 <sup>a</sup>	-83 ± 69	-236 ± 84 <sup>**</sup>	33 ± 88	296 ± 83 <sup>**</sup>
CD4 <sup>+</sup>	129	519 <sup>a</sup>	579 <sup>ab</sup>	575 <sup>ab</sup>	667 <sup>b</sup>	490 <sup>a</sup>	510 <sup>a</sup>	-57 ± 39	-72 ± 48	49 ± 50	121 ± 47 <sup>*</sup>
CD8 <sup>+</sup>	129	246 <sup>a</sup>	273 <sup>ab</sup>	313 <sup>bc</sup>	377 <sup>c</sup>	297 <sup>ab</sup>	274 <sup>ab</sup>	-23 ± 23	-86 ± 28 <sup>**</sup>	-26 ± 29	59 ± 28 <sup>*</sup>
CD25 <sup>+</sup>	129	15.7	13.6	12.6	12.2	8.0	7.2	1.1 ± 2.4	2.3 ± 3.0	7.1 ± 3.1 <sup>**</sup>	4.9 ± 3.0
CD4 <sup>+</sup> /CD8 <sup>+</sup>	129	2.23 <sup>c</sup>	2.33 <sup>c</sup>	1.87 <sup>ab</sup>	1.81 <sup>ab</sup>	1.75 <sup>a</sup>	2.09 <sup>bc</sup>	-0.13 ± 0.10	0.44 ± 0.12 <sup>***</sup>	0.36 ± 0.12 <sup>**</sup>	-0.08 ± 0.11
Monocytes	129	278 <sup>ab</sup>	275 <sup>ab</sup>	244 <sup>ab</sup>	304 <sup>b</sup>	188 <sup>a</sup>	228 <sup>ab</sup>	-33 ± 27	23 ± 33	69 ± 34 <sup>*</sup>	66 ± 32 <sup>*</sup>
Granulocytes	129	1640 <sup>abc</sup>	1552 <sup>abc</sup>	1705 <sup>bc</sup>	2015 <sup>c</sup>	1209 <sup>a</sup>	1376 <sup>ab</sup>	-129 ± 152	-264 ± 184	303 ± 194	567 ± 182 <sup>**</sup>

<sup>1</sup> Genetic line: line H, founded by litter size at birth and selected by litter size at weaning during 17 generations; line LP, founded by reproductive longevity criteria by selecting females from commercial farms that had a minimum of 25 parturitions with more than 7.5 kits born alive per parity and then selected by litter size at weaning for 7 generations; line R, founded and selected during 25 generations by average daily gain from the 4th to the 9th week of life.

<sup>2</sup> Reproduction diet: CS, mainly based on cereal starch (247 g of starch and 21 g of ether extract (EE) per kg dry matter (DM)); AF, mainly based on animal fat (104 g of starch and 85 g of EE per kg DM).

<sup>3</sup> Orthogonal contrasts to test the differences between lines [H-LP, H-R and LP-R] and reproduction diets [AF-CS].

<sup>4</sup> Maturity index at weaning as the quotient between live weight at weaning and mature weight for each line.

<sup>5</sup> No. for leukocytes counts refers to single samples. Each sample was obtained by pulling together blood from three kits from the same litter.

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**Table 4.** Effect of genetic line and reproductive diet on the performance of growing rabbits

Lines <sup>1</sup>		H		LP		R		Contrasts <sup>3</sup>			
Reproduction diet <sup>2</sup>	No.	AF	CS	AF	CS	AF	CS	AF-CS	H-LP	H-R	LP-R
30 to 44 days:											
Daily weight gain (g)	1231	34.2 <sup>a</sup>	36.1 <sup>b</sup>	34.0 <sup>a</sup>	37.5 <sup>c</sup>	43.6 <sup>d</sup>	45.1 <sup>e</sup>	-2.3 ± 0.4 <sup>***</sup>	-0.6 ± 0.4	-9.2 ± 0.5 <sup>***</sup>	-8.6 ± 0.5 <sup>***</sup>
Daily feed intake (g DM)	186	65.5 <sup>a</sup>	64.4 <sup>a</sup>	67.7 <sup>a</sup>	71.2 <sup>ab</sup>	74.5 <sup>ab</sup>	79.6 <sup>b</sup>	-2.5 ± 3	-4.5 ± 3.5	-12.1 ± 4 <sup>**</sup>	-7.6 ± 3.5 <sup>*</sup>
Feed conversion ratio (g DM/g)	186	1.95 <sup>b</sup>	1.84 <sup>ab</sup>	1.97 <sup>b</sup>	1.93 <sup>b</sup>	1.73 <sup>a</sup>	1.72 <sup>a</sup>	0.06 ± 0.05	-0.05 ± 0.06	0.17 ± 0.07 <sup>*</sup>	0.22 ± 0.06 <sup>***</sup>
44 to 58 days:											
Daily weight gain (g)	1231	34.6 <sup>a</sup>	35.6 <sup>a</sup>	35.2 <sup>a</sup>	35.7 <sup>a</sup>	47.8 <sup>b</sup>	47.4 <sup>b</sup>	-0.4 ± 0.4	-0.3 ± 0.4	-12.5 ± 0.5 <sup>***</sup>	-12.2 ± 0.5 <sup>***</sup>
Daily feed intake (g DM)	186	99.5 <sup>a</sup>	109.6 <sup>a</sup>	105.7 <sup>a</sup>	107.0 <sup>a</sup>	129.9 <sup>b</sup>	128.2 <sup>b</sup>	-3.3 ± 3	-1.8 ± 3.5	-24.5 ± 4 <sup>***</sup>	-22.7 ± 3.5 <sup>***</sup>
Feed conversion ratio (g DM/g)	186	2.94 <sup>b</sup>	2.93 <sup>b</sup>	3.01 <sup>b</sup>	3.00 <sup>b</sup>	2.72 <sup>a</sup>	2.62 <sup>a</sup>	0.04 ± 0.05	-0.07 ± 0.06	0.27 ± 0.07 <sup>***</sup>	0.34 ± 0.06 <sup>***</sup>
Live weight at 58 days (g)	1231	1550 <sup>a</sup>	1524 <sup>a</sup>	1611 <sup>b</sup>	1626 <sup>b</sup>	1931 <sup>c</sup>	1900 <sup>c</sup>	14 ± 10	-81 ± 12 <sup>***</sup>	-378 ± 14 <sup>***</sup>	-297 ± 12 <sup>***</sup>

<sup>1</sup> Genetic line: line H, founded by litter size at birth and selected by litter size at weaning during 17 generations; line LP, founded by reproductive longevity criteria by selecting females from commercial farms that had a minimum of 25 parturitions with more than 7.5 kits born alive per parity and then selected by litter size at weaning for 7 generations; line R, founded and selected during 25 generations by average daily gain from the 4th to the 9th week of life.

<sup>2</sup> Reproduction diet: CS, mainly based on cereal starch (247 g of starch and 21 g of ether extract (EE) per kg dry matter (DM)); AF, mainly based on animal fat (104 g of starch and 85 g of EE per kg DM).

<sup>3</sup> Orthogonal contrasts to test the differences between lines [H-LP, H-R and LP-R] and reproduction diets [AF-CS].

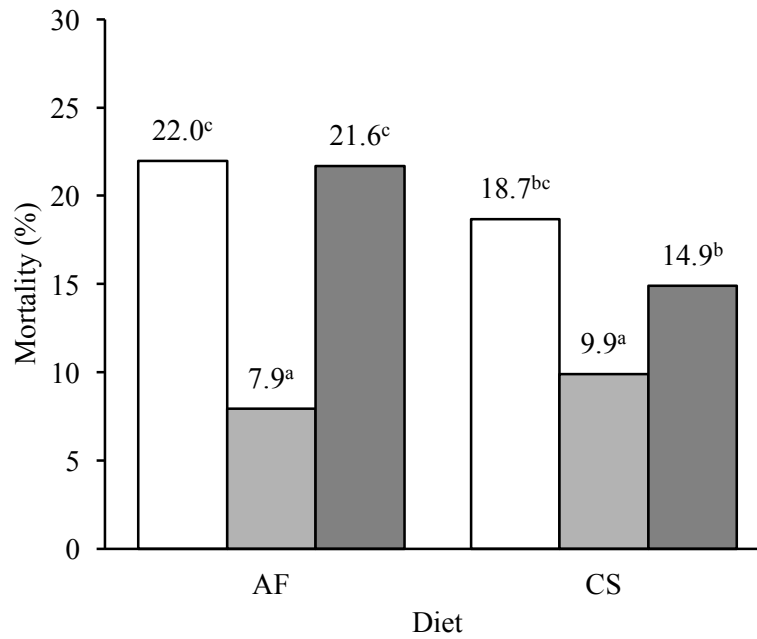
**Table 5.** Effect of genetic line, reproductive diet and growing diet on the mortality by digestive disorders and morbidity during the growing period.

		Mortality (%)	Morbidity (%)
Genetic line <sup>1</sup> :	H	19.0 <sup>b</sup>	6.6 <sup>b</sup>
	LP	9.5 <sup>a</sup>	3.9 <sup>a</sup>
	R	16.5 <sup>b</sup>	9.9 <sup>b</sup>
Reproductive diet <sup>2</sup> :	AF	13.7	7.2
	CS	14.4	5.3
Growing diet <sup>3</sup> :	M	7.9 <sup>a</sup>	6.7
	NM	26.8 <sup>b</sup>	5.4

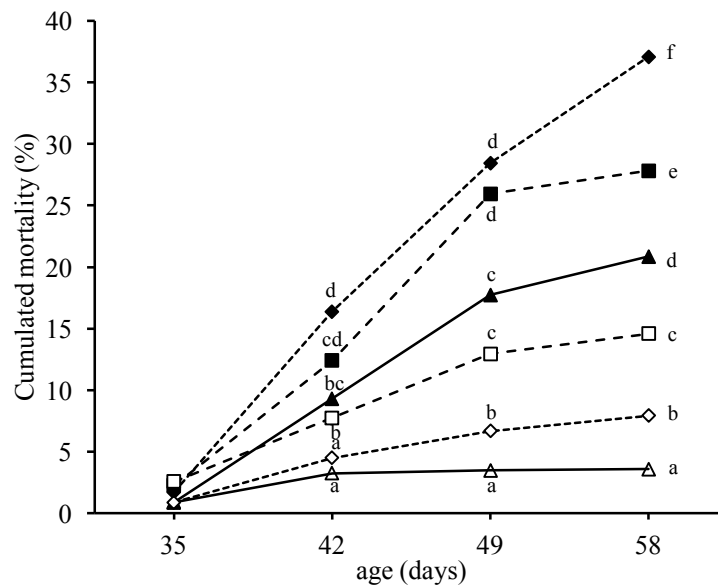
<sup>1</sup> Genetic line: line H, founded by litter size at birth and selected by litter size at weaning during 17 generations; line LP, founded by reproductive longevity criteria by selecting females from commercial farms that had a minimum of 25 parturitions with more than 7.5 kits born alive per parity and then selected by litter size at weaning for 7 generations; line R, founded and selected during 25 generations by average daily gain from the 4th to the 9th week of life.

<sup>2</sup> Reproduction diet: AF, mainly based on animal fat (104 g of starch and 85 g of EE per kg DM); CS, mainly based on cereal starch (247 g of starch and 21 g of ether extract (EE) per kg dry matter (DM));

<sup>3</sup> Growing diet: M, medicated (40 ppm of tiamulin fumarate A, 120 ppm of neomycin sulfate, 29 ppm of lincomycin hydrochloride and 29 ppm of spectinomycine sulphate); and NM, non-medicated.



**Figure 1.** Effect of genetic line (□H, ■LP and ■R) and reproduction diet (AF and CS) on mortality by digestive disorders registered during the growing period (30 to 58 days of age).



**Figure 2.** Evolution of cumulated mortality by digestive disorders depending on genetic line [ - -■- - H, —▲— LP and ---◆--- R] and the use of antimicrobials in the feed [medicated, (□△◇); non-medicated, (■▲◆)].<sup>abcdef</sup> Means at a same age not sharing superscript differ significantly at P<0.05.

