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1 Running head: feeding programmes for young rabbit does

2

3 **Effects of rearing feeding programme on the performance and energy balance**
4 **of young rabbit does**

5

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20 ABSTRACT

21

22 A total of 228 young rabbit females aged 9 weeks were used to evaluate five rearing
23 feeding programmes: CAL, fed *ad libitum* with a control diet [C: 11.0 MJ digestible
24 energy (DE) and 114 g digestible protein (DP) per kg dry matter (DM)] until first
25 parturition; CR, receiving the C diet restricted (140 g/d) from 12 weeks of age to first
26 partum; F, fed *ad libitum* with a moderate energy fibrous diet [F: 8.7 MJ DE and 88 g
27 DP per kg DM] until first partum; and finally, FC and FCF, fed with F diet *ad libitum*
28 until 16 weeks of age, whereupon FC group received the C diet *ad libitum* until first
29 partum, while FCF group received the C diet *ad libitum* until 20 weeks of age and
30 then the F diet *ad libitum* until first partum. CAL group had a higher mortality rate
31 compared to the other groups between 9 and 12 weeks of age (34 vs. 3%; $P<0.05$)
32 and during the last 3 weeks of first pregnancy (14 vs. 3%; $P<0.05$). CAL and FC
33 females presented higher BW and perirenal fat thickness (PFT) than CR females at
34 week 20 (+0.41 kg and +0.6 mm; $P<0.05$), with F females showing medium values.
35 The type of feeding procedure did not affect the fertility rate of young females at first
36 AI. Differences in BW disappeared at parturition, when only CAL females presented a
37 greater PFT than CR and FC females (+0.3 mm; $P<0.05$). In comparison to FCF,
38 CAL females had smaller and thinner live born litters (-2.5 kits and -139 g,
39 respectively; $P<0.05$), with CR, F and FC females showing medium values. The low
40 number of kits born alive for CAL females was due to their lesser total number of kits
41 born (-1.7 kits; $P<0.05$) and the greater mortality of their litters at birth (+13.9 %;
42 $P<0.05$) compared to FCF. NEFA was higher in the blood of females fed C diet (CAL
43 and CR) than in others at partum day (on average +0.15 mmol/L; $P<0.05$). In
44 conclusion, the *ad libitum* use of diets for lactating rabbit does throughout the rearing

45 period could lead young rabbit females to present a higher risk of early death and
46 smaller litter size at first parturition. Feed restriction or earlier use of suitably fibrous
47 diets led females to achieve the critical BW and fat mass at first mating to ensure
48 reproduction.

49

50 **Key words:** rabbit females, rearing, pubertal development, body condition, metabolic
51 status.

52

53

54 **IMPLICATIONS**

55

56 Obtaining well-developed rabbit females that produce a large number of healthy,
57 marketable litters per mating over multiple parities is still one of the main priorities for
58 rabbit production. This objective not only involves the use of adequate management
59 programmes during reproduction, but also appropriate management of nutrition
60 during pre- and post-pubertal growth to ensure better development of future
61 reproductive females. The correct design of rearing programmes that take into
62 account the young rabbit female's requirements and priorities, while ensuring both
63 adequate pubertal development and future reproductive performance, is a pressing
64 need.

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79 **INTRODUCTION**

80

81 The negative effects of underfeeding on pubertal maturation have long been known
82 in numerous species (Frisch, 1984). However, overfeeding during rearing period has
83 been also related with lower reproductive performance in dairy heifers (Sejrsen *et al.*,
84 1982), pullets (Whitehead, 1988) and gilts (Klindt *et al.*, 1999). Young rabbit females
85 fed *ad libitum* until first parturition usually suffer similar problems to those mentioned
86 for other species.

87 For this reason, in the last decade some works assessed the possible impact of
88 different management and feeding plans for rearing period on female development
89 and reproduction: feed restriction (Rommers *et al.*, 2004), BW at weaning or at the
90 first artificial insemination (AI) (Rommers *et al.*, 2001a, 2001b, 2002), or the use of
91 fibrous diets (Xiccato *et al.*, 1999; Pascual *et al.*, 2002; Quevedo *et al.*, 2005).
92 However, some of these works have shown an antagonism between proper
93 development and improvement of reproductive response. The earlier the introduction
94 of the restriction programme and the lower the energy supply, the higher the
95 voluntary feed intake of primiparous does with improved milk yield or reduced body
96 reserves mobilisation during first lactation (Nizza *et al.*, 1997; Xiccato *et al.*, 1999;
97 Pascual *et al.*, 2002), but the later or the lower their pubertal maturation (Pascual *et*
98 *al.*, 2002; Rommers *et al.*, 2004).

99 On the basis of this previous information, in this work we evaluated the effects on
100 nulliparous rabbit does development of a diet for reproductive rabbit does provided
101 during the rearing period both *ad libitum* and restricted, compared with three different
102 feeding programmes based on the use of a moderate-energy fibrous diet designed

103 for young rabbit females and provided: i) until first parturition, ii) until first mating and
104 iii) until first parturition applying a flushing around first mating.

105

106

MATERIAL AND METHODS

The experimental procedure was approved by the animal welfare ethics committee of the Universitat Politècnica de València (UPV) and carried out following the European Union recommendations on care and protection of animals used for experimental purposes (2003) and the advice for applied nutrition research in rabbits according to the European Group on Rabbit Nutrition (Fernández-Carmona *et al.*, 2005).

Diets

Ingredients and chemical composition of the experimental pelleted diets used in this trial are summarised in Table 1. A control diet (C), similar to a commercial diet for reproductive rabbit does [11.0 MJ digestible energy (DE) and 114 g digestible protein (DP) per kg dry matter (DM)], was formulated following the recommendations of De Blas and Mateos (2010). In addition, a moderate energy diet with a high fibre content (F) was also formulated [8.7 MJ DE and 88 g DP per kg DM], including some minor ingredients and supplements to partially correct obvious deficiencies in amino acids and minerals.

(Table1)

Apparent digestibility coefficients of energy and CP were determined for each diet, using a total of 30 three-way crossbred rabbits, aged 42 days with an average BW of 1.32 (s.d. 0.07) kg according to Perez *et al.* (1995).

Chemical analysis of diets and faeces were performed following the AOAC (1999) methods for DM, ash, ether extract, CP, and crude fibre (934.01, 942.05, 920.39, 976.06 and 978.10, respectively). Ether extract was determined after acid hydrolysis.

132 NDF, ADF and ADL were analysed sequentially (Van Soest *et al.*, 1991) using a
133 thermo-stable amylase (Thermamyl L120, Novo Nordisk, Gentofte, Denmark) pre-
134 treatment and expressed exclusive of residual ash. Gross energy was determined by
135 adiabatic bomb calorimetry (Gallenkamp Autobomb, Loughborough, UK) following
136 the recommendation of EGRAN (2001).

137

138 *Animals and experimental procedure*

139

140 A total of 228 young rabbit does (line A from UPV, selected over 36 generations for
141 litter size at weaning) were used from 9 weeks of age to first parturition. The animals
142 were housed in a traditional building under controlled environmental conditions, with
143 light alternating on a cycle of 16 h light and 8 h dark. The experiment was carried out
144 from January to June 2007.

145 Until 9 weeks of age, young rabbit females were caged collectively, receiving the
146 same commercial diet *ad libitum* (185 g crude fibre and 175 g CP per kg DM), and
147 subsequently housed in individual cages with access to one of the experimental
148 diets. Combining two diets and three different feeding schemes, five feeding
149 programmes were formed (Figure 1). The CAL group included females which
150 received the C diet *ad libitum* until first parturition. The CR group included females
151 which received the C diet *ad libitum* until 12 weeks of age and then 140 g per day
152 until first parturition, with a 7-day flushing period (C diet *ad libitum*) around AI. The F
153 group included females which received the F diet *ad libitum* until first parturition.
154 Finally, FC and FCF groups were females that received F diet *ad libitum* until 16
155 weeks of age, whereupon the FC group received the C diet *ad libitum* until first

156 parturition, while FCF group received the C diet *ad libitum* until 20 weeks of age and
157 then the F diet *ad libitum* until first parturition.

158 (Figure 1)

159 While animals from different experimental groups kept the same feeding programme,
160 data were analysed and presented as a whole (CAL and CR until 12 weeks of age, F,
161 FC and FCF until 16 weeks of age, and then FC and FCF until 20 weeks of age).

162 Does were artificially inseminated at the end of the 18th week of age. As of this date,
163 successive AI were carried out every 21 days, as necessary. After the 28th day of
164 pregnancy, maternal cages were provided with a nest equipped for the litter.

165 The traits measured for all does were BW and food intake at 9, 12, 16, 18 (AI), 20
166 and 23 (parturition) weeks of age, as well as perirenal fat thickness (PFT) by
167 ultrasounds at 9, 12, 18, 20 and 23 weeks of age. Total and live litter size and weight
168 at partum were also recorded. From 12 rabbit does per group, blood samples were
169 collected at 9, 12, 18 and 23 weeks of age. On sampling day, feeders were closed at
170 07:00 h and blood samples were taken from the central ear artery into EDTA-
171 containing tubes from 11:00 to 13:00 h. Blood samples were centrifuged immediately
172 after sampling (3000×g, 4°C and 10 minutes) and plasma was stored at –20°C before
173 being assayed for insulin, glucose, non-esterified fatty acids (NEFA), leptin, cortisol
174 and tri-iodothyroxine (T3) concentrations. Controls at 9, 12, 16 and 20 weeks of age
175 were done on Mondays, and those at 18 weeks of age (AI) on Friday.

177 *Ultrasound measurements*

178
179 The PFT of does was measured by ultrasound to evaluate body condition, as
180 described by Pascual *et al.* (2000 and 2004). Images were obtained with an

181 ultrasound unit (JustVision 200 'SSA-320A' real-time machine; Toshiba) equipped
182 with image analyser software to determine distances. Estimated body energy
183 content (EBE; MJ/kg) was determined at AI and parturition from BW and PFT data as
184 described by Pascual *et al.* (2004).

185

186 *Hormone and metabolite assays*

187

188 Plasma insulin concentrations were determined by the double antibody/PEG
189 technique using porcine insulin radioimmunoassay (RIA) kit (Linco Research Inc., St
190 Charles, MO, USA). The antiserum was guinea pig anti-porcine insulin, while both
191 labelled antigen and standards used purified recombinant human insulin. Leptin
192 concentrations were determined by double antibody RIA using the multi-species
193 leptin kit (Linco Research Inc.) as previously reported (Brecchia *et al.* 2006). Total
194 T3 was assayed by RIA according to the procedure provided by the manufacturer
195 (Immunotech, Marseille, France). The assay sensitivity was 0.13 ng/mL, and the
196 major analogues of T3 did not interfere with the assay. Plasma cortisol was assayed
197 by RIA, using the CORT kit (ICN Biomedicals Inc., Costa Mesa, CA, USA). CORT
198 assay sensitivity was 0.15 ng/mL. Dilution and recovery tests done on insulin, leptin,
199 T3 and corticosterone using five different samples of rabbit plasma showed linearity.
200 Glucose was analysed by the glucose oxidase method using the Glucose Infinity kit
201 from Sigma (Sigma Diagnostic Inc., St. Louis, MO, USA). NEFA concentrations were
202 analysed using enzymatic colorimetric assay from Wako (Wako Chemicals GmbH,
203 Neuss, Germany) as previously reported (Brecchia *et al.*, 2006).

204

205 *Statistical Analysis*

206

207 The model used to analyse performance, hormonal and metabolic data of young
208 rabbit does during rearing and first gestation was a mixed model (PROC MIXED by
209 SAS, Statistical Analysis System, 2002), in a repeated measure design that took into
210 account the variation between animals and covariation within them. Covariance
211 structures were objectively compared using the most severe criteria (Schwarz
212 Bayesian criterion), as suggested by Littell *et al.* (1998). The model included the
213 feeding programme (CAL, CR F, FC and FCF), the week of age (9, 12, 18, 20 and 23
214 weeks; data for week 16 was also included for consumption and BW), and their
215 interaction as fixed effects. Random terms in the model included a permanent effect
216 of each animal (p) and the error term (e), both assumed to have an average of zero,
217 and variance σ_p^2 and σ_e^2 .

218 Different contrasts were computed to test the significance of the differences between
219 treatments while animals of different experimental groups received the same feeding
220 programme at 12 weeks [(CAL+CR)/2 vs. (F+FC+FCF)/3], at 16 weeks [CAL vs. CR
221 vs. (F+FC+FCF)/3], at 18 and 20 weeks [CAL vs. CR vs. F vs. (FC+FCF)/2] and
222 parturition [CAL vs. CR vs. F vs. FC vs. FCF].

223 To analyse the litter data at first parturition, a fixed effects model (PROC GLM of
224 SAS, 2002) was used that included only the feeding programme (CAL, CR F, FC and
225 FCF). Data concerning mortality of females during the rearing and first pregnancy
226 were analysed according to a nonparametric procedure (PROC NPAR1WAY of SAS,
227 2002), using a chi-square test for mean separation.

228

229 RESULTS

230

231 *Animal Performance*

232

233 A high mortality rate was observed in the CAL group (34%) between 9 and 12 weeks
234 of age (Figure 2) compared to the F group (3%; $P<0.05$), probably due to an outbreak
235 of epizootic rabbit enteropathy (ERE). Mortality was low and similar in groups under
236 different feeding programmes from 12 to 20 weeks of age. However, the CAL group
237 again presented a significantly higher mortality (14%; $P<0.05$) compared to the other
238 groups (on average 3%) during the last 3 weeks of pregnancy.

239

(Figure 2)

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Daily intake, BW and PFT of young rabbit does during rearing and pregnancy are
presented in Table 2 and Figures 3 and 4. The BW and PFT at 9 weeks of age was
1.97±0.03 (standard error) kg and 6.9±0.1 mm, respectively. Females *ad libitum* fed
with C diet (CAL) showed significantly higher DE and DP intake between 9 and 12
weeks of age (+89 kJ and +11 g per day, respectively; $P<0.05$) and BW at week 12
(+0.11 kg; $P<0.05$) than those with F diet. From 12 to 16 weeks, DE and DP intake
were similar for CAL and F females (on average 841 kJ and 86 g per day,
respectively), but significantly lower for those restricted (CR; 699 kJ and 72 g per
day; $P<0.05$). Thus, BW at week 16 was significantly higher for CAL than for F group
(3.69 and 3.47 kg, respectively; $P<0.05$), and higher for both than for CR (3.24 kg;
 $P<0.05$).

251

(Table 2)

252

253

254

From 16 to 20 weeks of age, DE and DP intake of F group was even higher (on
average 792 kJ and 80 g per day, respectively) than that observed for CAL group
(742 kJ and 76 g per day; $P<0.05$) and higher for both than for CR group (690 kJ and

255 71 g per kg per day; $P<0.05$). In fact, F females going on to *ad libitum* C diet at 16
256 weeks (FC) showed the highest intake values (on average 883 kJ and 91 g per day;
257 $P<0.05$). In consequence, CAL and FC females at week 20 presented higher BW
258 and PFT (4.34 kg and 7.3 mm, respectively) than CR females (3.93 and 6.7 mm;
259 $P<0.05$), with F females showing medium values (4.14 kg and 7.1 mm).

260 The type of feeding programme did not affect the fertility rate of young females at first
261 AI (85.2, 84.0, 89.7 and 85.0% for CAL, CR, F and FC females, respectively).

262 During the last 3 weeks of pregnancy, F and FC females presented higher DE and
263 DP intake (on av. 630 kJ and 65 g per day, respectively) than CR and FCF females
264 (on av. 586 kJ and 60 g per day; $P<0.05$), with CAL females showing the lowest
265 intake values (537 kJ and 55 g per day; $P<0.05$). Thus, differences in BW between
266 feeding programmes disappeared at parturition (Figure 3), while only CAL females
267 presented a greater PFT than CR and FC females (6.4 vs. 6.1 mm, respectively;
268 $P<0.05$).

269 (Figures 3 and 4)

270 Table 3 shows the effect of the feeding programme adopted during doe rearing on
271 litter traits at the first parturition. In comparison to the CAL group, FCF females had
272 larger (7.7 vs. 5.2 kits; $P<0.05$) and heavier live born litters (419 vs. 280 g; $P<0.05$),
273 with CR, F and FC females showing medium values (on av. 6.1 kits and 349 g). The
274 small number of kits born alive at first parturition to CAL females was due to their
275 lower number of total kits born (6.6 vs. 8.3 kits $P<0.05$) and the greater mortality of
276 their litters at birth (20.6 vs. 6.7 %; $P<0.05$) compared to FCF.

277 (Table 3)

278

279 *Metabolic and hormonal parameters*

280
281 The plasma profiles of insulin, glucose, NEFA, leptin, cortisol and T3 during rearing
282 and first pregnancy in the different feeding programmes are shown in Figure 5. An
283 increase in circulating insulin concentrations was observed with advancing age in all
284 the groups, although it decreased at parturition (Figure 5a). CAL group animals
285 presented lower mean plasma insulin concentration than F females at 18 weeks of
286 age ($-19.5 \mu\text{UI/mL}$; $P<0.05$).

287 Both glucose and NEFA plasma concentrations showed the highest values at 9
288 weeks of age and dropped thereafter. Glucose concentration in plasma was
289 opposite to insulin, being lower for CAL (-31.4 mg/dL ; $P<0.05$) than for F females at
290 12 weeks of age (Figure 5b). At partition day, glucose was lower in CAL, CR and
291 FCF than F and FC females (on av. -20.7 mg/dL ; $P<0.05$). Although NEFA levels
292 were similar for all the groups at 18 weeks of age (Figure 5c), females receiving the
293 C diet (CAL, CR and FC) presented the highest NEFA values in plasma at parturition,
294 only being significantly higher in CAL and CR compared to F females (on av. $+0.18$
295 mmol/L ; $P<0.05$).

296 Leptin levels were similar for all groups at 12 weeks of age and at partum day (Figure
297 5d). An increase in plasma leptin concentration was observed at 18 weeks,
298 especially in CR females (6.6 ng/mL ; $P<0.05$), where plasma had higher leptin levels
299 than CAL (5.1 ng/mL), and both F and FC females (on average 3.3 ng/mL ; $P<0.05$).

300 Plasma cortisol increased from 9 to 12 and 18 weeks of age, although it decreased at
301 parturition (Figure 5e). No significant differences between feeding programmes on
302 cortisol in plasma were observed throughout the experiment. Plasma concentrations
303 of T3 at 12 weeks of age were similar for all the groups (Figure 5f). Females given C
304 diet *ad libitum* at 18 weeks (CAL and FC) had higher levels of plasma T3 than CR

305 females (on av. 0.75 mmol/L; $P<0.05$). However, CAL females showed higher T3
306 levels than FC females at parturition (-0.96 mmol/L; $P<0.05$).

307 (Figure 5)

308

309

310 **DISCUSSION**

311
312 No previous work evaluating the use of rearing diets described the high mortality rate
313 observed in the present work from 9 to 12 weeks of age when young females were
314 fed the control diet. This fact seems to be related to ERE incidence when no
315 medicated diets are used. Under these conditions, insufficient level or inadequate
316 quality of dietary fibre can increase the risk of digestive disorders in young rabbits
317 (Gidenne, 1997; Gidenne and Garcia, 2006). In the current work, although higher
318 soluble fibre was expected for the F diet (from alfalfa and beet pulp), both diets were
319 designed to meet fibre recommendations to prevent digestive problems from 9 to 12
320 weeks of age (ADL>50, ADF>190 and NDF-ADF>80 g/kg). However, a recent review
321 (Blas and Gidenne, 2010) highlighted that, even if requirements proposed to prevent
322 digestive disorders are met, replacing starch with low or high digestible fibre reduces
323 mortality rate, especially in the context of ERE.

324 Young rabbit female needs from 9 to 12 weeks of age (approx. 1.52 MJ per day,
325 considering their mean live weight and daily gain; Xiccato and Trocino, 2010) were
326 met with both C and F diets (1.74 and 1.54 MJ per day, respectively). Although the
327 lower DE intake led females receiving the F diet to reach 12 weeks with a smaller
328 BW, as in a previous work (Pascual *et al.*, 2002), the main metabolic and hormonal
329 parameters here examined were not greatly affected. Rebollar *et al.* (2011) also
330 found similar concentrations of leptin (2.8 ng/mL) and NEFA (0.22 mmol/L) in the
331 blood of young females at 11 weeks of age when comparing *ad libitum* supplying of
332 control and fibre-rich diets. However, when higher feed restriction is asserted (even
333 below animal needs 1.03 MJ per day; Rommers *et al.*, 2004), the blood levels of
334 glucose, leptin, insulin, and T3 of young females (from 6 to 12 weeks of age) were
335 clearly reduced.

336 As a consequence of feed intake restriction from 12 to 18 weeks of age, CR females
337 reached 18 weeks of age with a delay in their development, showing lower BW and
338 PFT than those fed *ad libitum*. These results agree with those reported in previous
339 works where feed restriction reduced BW as well as body fat and protein content of
340 young rabbit females at first AI (Rommers *et al.*, 2001, 2004), and even caused a
341 delay in the effectiveness of this AI (Rebollar *et al.*, 2011). In the present work, CR
342 females presented a slight reduction of T3 blood levels at 18 weeks of age together
343 with an unexpected higher concentration of leptin compared to those with free access
344 to the control diet. Several studies have shown that fasting reduces leptin, mainly
345 synthesised and secreted by adipocytes, circulating in blood at levels proportional to
346 body fat stores in humans (Weigle *et al.*, 1997), gilts (Barb *et al.*, 2001), ruminants
347 (Chilliard *et al.*, 2000), and also in rabbits (Rommers *et al.*, 2004; Brecchia *et al.*,
348 2006; Rebollar *et al.*, 2011). However, the mechanisms whereby feeding restriction
349 affects circulating leptin levels are still unclear, and different responses were
350 observed depending on type and length of fasting and blood sampling protocols. In
351 this respect, Brecchia *et al.* (2006) described higher leptin levels in the plasma of 48-
352 h fasted than in 24-h fasted does. In any case, it might be considered that CR
353 females were subject to a 4-day flushing period prior to AI, where animals had free
354 access to the C diet, which could have conditioned the plasma metabolic profile for
355 these days.

356 On the other hand, females with free access to the F diet were able to compensate
357 for the lower nutritive dietary concentration with a greater feed intake from 12 to 18
358 weeks of age. Thus, they achieved DE and DP intakes similar to those of rabbits
359 receiving the C diet *ad libitum* and, consequently, reduced their gaps in BW and PFT
360 at 18 weeks of age. In fact, these differences disappeared when females of F group

361 had free access to C diet as of 16 weeks of age. Pascual *et al.* (2002) described
362 how young rabbit females fed with a low-energy diet (8 MJ DE/kg DM) as of 10
363 weeks of age presented, during late rearing, a greater DE intake than those fed 150
364 g per day of a standard diet (11 MJ DE/kg DM). The greater feed intake, however,
365 did not compensate earlier differences in BW and these rabbits achieved first AI 10
366 days later. However, a later introduction of a low-energy diet at 13 weeks of age
367 (Quevedo *et al.*, 2005) or the use of moderate-energy diets (9.5 MJ DE/kg DM;
368 Xiccato *et al.*, 1999) enabled young rabbit females to achieve first mating at an
369 adequate age and BW.

370 In this sense, although the use of a moderate low-energy diet during the rearing
371 period led females to reach first mating with lower energy body reserves (Figure 6)
372 and lower blood leptin levels than those fed with a conventional diet for reproductive
373 does, no consequence on fertility at first AI was reported. It is well-known that
374 nutrient restriction may delay the onset of puberty, leading to the hypothesis that a
375 critical soma must be achieved before puberty can occur (Frisch, 1980).
376 Furthermore, Arias-Álvarez *et al.* (2009) recently proposed that reaching the
377 permissive leptin threshold should be necessary for pubertal reproductive activity,
378 and may be associated with inhibition of reproduction if the critical soma is
379 insufficient to trigger gestation (Moschos *et al.*, 2002). In fact, when the relationship
380 between fertility and blood leptin levels of young rabbit females around first
381 insemination is drawn (Figure 7), the hypothesis of a leptin threshold for initiation of
382 puberty and reproductive success which is not improved by additional provision of
383 this hormone seems to be confirmed. Consequently, these results reveal that in
384 terms of *ad libitum* feeding during rearing, both feed restriction and earlier use of a
385 moderate low-energy diet (8.7 MJ/kg DM) led females to achieve the critical BW and

386 fat mass at first AI to ensure reproduction, in spite of their lower fatness and leptin
387 content in blood.

388 (Figures 6 and 7)

389 After the first AI, although young females receiving the C diet *ad libitum* maintained a
390 greater consumption than those restricted until 20 weeks of age, the fatness
391 accumulated by CAL females throughout rearing allowed them to reduce their feed
392 intake as pregnancy progressed, allowing CR females to diminish the differences in
393 BW, PFT, and EBE observed up to this point with the CAL group during late
394 pregnancy. In a previous work (Rommers *et al.*, 2004), where development between
395 young females fed *ad libitum* and early restricted (restriction: from 5 to 10 weeks of
396 age; recovery: 10 to 17.5 weeks of age) was compared, although compensatory
397 growth of the restricted group was also observed during pregnancy, the early
398 differences achieved in BW of females were maintained throughout the 3
399 reproductive cycles controlled by the authors. In gilts, where feed restriction of young
400 females has been studied extensively, most works (Sørensen *et al.*, 1998; Klindt *et*
401 *al.*, 1999 and 2001b) show that moderate feed restriction during the rearing period
402 helps females avoid excessive fatness, while more intense restriction (earlier and/or
403 stronger) leads to smaller development and sometimes even to lower reproductive
404 performance. Therefore, these results seem to confirm the effectiveness of
405 moderate restrictive feeding in preventing excessive fatness in young females,
406 although the starting age and restriction level should be controlled to avoid an
407 inadequate pre-pubertal body development.

408 A practical alternative to restriction could be the use of fibrous diets. Several works
409 found in the literature showed that the use of fibrous diets during rearing led
410 nulliparous rabbit females to a greater DE intake after first mating, independently of

411 their previous growth rate during development. Even so, when low-energy fibrous
412 diets are used (<8.5 MJ DE/kg DM), females are not able to compensate the
413 previous developmental delay (Pascual *et al.*, 2002; Quevedo *et al.*, 2005).
414 However, when females have the chance of receiving a moderate-energy fibrous diet
415 (approx. 9 MJ DE/kg DM), they reach the first parturition with a development and BW
416 similar to those of rabbit does fed *ad libitum* a diet for reproductive does (>10.5 MJ
417 DE/kg DM), but with a lower fatness (Xiccato *et al.*, 1999; Rebollar *et al.*, 2011). In
418 the present work, and independently of the fibrous feeding systems used (F, FC or
419 FCF), females reached first parturition in an intermediate developmental situation to
420 that observed in females fed with the C diet *ad libitum* or restricted. Similar results
421 were also obtained by Rebollar *et al.* (2011), where the use of a fibrous diet (9.4 MJ
422 DE/kg DM) from 11 weeks of age to first parturition led young rabbit females to reach
423 the end of first pregnancy with body energy and protein content halfway between *ad*
424 *libitum* and restricted administration of a control diet (11.6 MJ DE/kg DM). The use of
425 a fibrous diet with 8.5 to 9.5 MJ DE/kg DM should therefore allow young rabbit
426 females to reach first parturition in an adequate state of development, avoiding
427 excessive fatness without the need for feeding restriction.

428 In fact, the possible negative effects of excessive fatness could be behind the
429 problems detected around first parturition in the CAL group. Compared to the other
430 feeding systems evaluated here, females fed the C diet *ad libitum* during rearing
431 showed the lowest DE intake and the highest body energy mobilisation recorded
432 during late pregnancy. In fact, the plasma of these females at partum day was
433 characterised by higher NEFA and lower glucose levels. The aforementioned profile
434 is frequently related to pregnancy toxemia risk (Martenink and Herdt, 1988; Bezille,
435 1995; Rosell, 2000), and could explain the higher mortality in late pregnancy for the

436 females of this group and the smaller size of their litters at first birth caused by both
437 lower total litter size and higher mortality at birth. Rommers *et al.* (2002) also
438 observed that heavier young females at first AI (more than 4 kg BW) had a higher
439 percentage of stillborn at first parturition (13.4%) than smaller females (5%). In gilts,
440 Klindt *et al.* (2001a and 2001b) related an excessive energy intake during rearing
441 with a lower number of corpora lutea and live embryos per gilt, and also observed a
442 tendency towards the reduction of litter size at first birth (−0.8 piglets born) and the
443 increase of gilts removed until this time (+13%). In this sense, the highest prolificacy
444 and the lowest mortality at birth were recorded for females given the F diet with a
445 flushing of 4 weeks with the C diet applied around first mating. In a recent revision,
446 Theau-Clement (2007) concluded that feed flushing after nutritive restriction could
447 improve the reproduction performance, at least at the beginning of the reproductive
448 career.

449 From the results of the present work it could be concluded that the *ad libitum* use of
450 diets formulated to cover the needs of lactating rabbit does for the whole rearing
451 period could lead young rabbit females to present a higher risk of early death and
452 smaller litter size at first parturition. As an alternative, either feed restriction or earlier
453 use of an adequate fibrous diet could lead females to achieve the critical BW and fat
454 mass at first AI to ensure reproduction. However, under these feeding programmes
455 for young females, the starting age and nutritive level of the fibrous diet should be
456 controlled to avoid an inadequate pre-pubertal development.

457

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462 **REFERENCES**

- 463 Association of Official Analytical Chemist 1999. Official methods of analysis, 18th edition.
464 AOAC, Gaithersburg, MD, USA (5th revision).
- 465 Arias-Álvarez M, García-García RM, Rebollar PG, Nicodemus N, Revuelta L, Millán P,
466 Lorenzo PL 2009. Effects of a lignin-rich fibre diet on productive, reproductive and
467 endocrine parameters in nulliparous rabbit does. *Livestock Science* 123, 107-115.
- 468 Barb CR, Barrett JB, Kraeling RR, Rampacek GB 2001. Serum leptin concentrations,
469 luteinizing hormone and growth hormone secretion during feed and metabolic fuel
470 restriction in the prepuberal gilt. *Domestic Animal Endocrinology* 20, 47-63.
- 471 Bezille P 1995. Toxémie de gestation et hypocalcémie chez la brebis. In: *Maladies*
472 *métaboliques des ruminants*. *Le Point Vétérinaire* 27, special number, 101-104.
- 473 Blas E., Gidenne T. 2010. Digestion of sugars and starch. In: *Nutrition of the Rabbit* (eds C
474 De Blas and Wiseman), pp.19-38. CABI Publishing, Wallingford, UK.
- 475 Brecchia G, Bonanno A, Galeati G, Federici C, Maranesi M, Gobbetti A, Zerani M, Boiti C
476 2006. Hormonal and metabolic adaptation to casting: Effects on the hypothalamic-
477 pituitary-ovarian axis and reproductive performance of rabbit does. *Domestic Animal*
478 *Endocrinology* 31, 105-122.
- 479 Chilliard Y, Ferlay A, Faulconnier Y, Bonnet M, Rouel J, Bocquier F 2000. Adipose tissue
480 metabolism and its role in adaptations to undernutrition in ruminants. *Proceedings of*
481 *the Nutrition Society* 59, 127–134.
- 482 De Blas C, Mateos GG 2010. Feed formulation. In: *Nutrition of the Rabbit* (eds C De Blas
483 and Wiseman), pp. 222-232. CABI Publishing, Wallingford, UK.
- 484 EGRAN 2001. Attempts to harmonize chemical analyses of feeds and faeces for rabbit feed
485 evaluation. *World Rabbit Science* 9, 57–64.
- 486 European Union 2003. Protection of animals used for experimental purposes. Directive
487 86/609/EEC of 24th November 1986, amended 16th September 2003.

488 Fernández-Carmona J, Blas E, Pascual JJ, Maertens L, Gidenne T, Xiccato G and García J
489 2005. Recommendations and guidelines for applied nutrition experiments in rabbits.
490 World Rabbit Science 13, 209–228.

491 Frisch RE 1980. Pubertal adipose tissue: is it necessary for normal sexual maturation?
492 Evidence from the rat and human female. Federation Proceedings 39, 2395-2400.

493 Frisch RE 1984. Body fat, puberty and fertility. Biological Reviews 59, 161-188.

494 Gidenne T 1997. Caeco-colic digestion in the growing rabbit: impact of nutritional factors and
495 related disturbances. Livestock Production Science 51, 73-88.

496 Gidenne T, García J 2006. Nutritional strategies improving the digestive health of the
497 weaned rabbit. In: Recent Advances in Rabbit Science (eds L Maertens and P
498 Coudert), pp. 229-238. COST. ILVO, Merelbeke, Belgium.

499 Klindt J, Yen JT, Christenson RK 1999. Effect of prepubertal feeding regimen on
500 reproductive development of gilts. Journal of Animal Science 77, 1968-1976.

501 Klindt J, Yen JT, Christenson RK 2001a. Effect of prepubertal feeding regimen on
502 reproductive development and performance of gilts through the first pregnancy.
503 Journal of Animal Science 79, 787-795.

504 Klindt J, Yen JT, Christenson RK 2001b. Level of dietary energy during prepubertal growth
505 and reproductive development of gilts. Journal of Animal Science 79, 2513-2523.

506 Littell RC, Henry PR and Ammerman CB 1998. Statistical analysis of repeated measures
507 data using SAS procedures. Journal of Animal Science 76, 1216–1231.

508 Martenink JV, Herdt T.H. 1988. Pregnancy toxemia and ketosis of ewes and does. Veterinary
509 Clinics of North America: Food Animal Practice 4 , 307-315.

510 Moschos S, Chan JL, Mantzoros CS 2002. Leptin and reproduction: a review. Fertility and
511 Sterility 77, 433–444.

512 Pascual JJ, Castella F, Cervera C, Blas E, Fernández-Carmona J 2000. The use of
513 ultrasound measurement of perirenal fat thickness to estimate changes in body
514 condition of young female rabbits. Animal Science 70, 435-442.

515 Pascual JJ, Cervera C, Fernández-Carmona J 2002. A feeding program for young rabbit
516 does based on all lucerne diets. *World Rabbit Science* 10, 7-13.

517 Pascual JJ, Blanco J, Piquer O, Quevedo F, Cervera C 2004. Ultrasound measurements of
518 perirenal fat thickness to estimate the body condition of reproducing rabbit does in
519 different physiological states. *World Rabbit Science* 12, 7-31.

520 Pérez JM, Lebas F, Gidenne T, Maertens L, Xiccato G, Parigi-Bini R, Dalle Zotte A, Cossu
521 ME, Carazzolo A, Villamide MJ, Carabaño R, Fraga MJ, Ramos MA, Cervera C, Blas
522 E, Fernández-Carmona J, Falcao e Cunha L, Bengala Ferre J 1995. European
523 reference method for in vivo determination of diet digestibility in rabbits. *World Rabbit*
524 *Science* 3, 41-43.

525 Quevedo F, Cervera C, Blas E, Baselga C, Costa C, Pascual JJ 2005. Effect of selection for
526 litter size and feeding programme on the performance of young rabbit females during
527 rearing and first pregnancy. *Animal Science* 2005, 80, 161-168.

528 Rebollar PG, Pereda N, Schwarz BF, Millán P, Lorenzo PL, Nicodemus N 2011. Effect of
529 feed restriction or feeding high-fibre diet during the rearing period on body
530 composition, serum parameters and productive performance of rabbit does. *Animal*
531 *Feed Science and Technology* 163, 67-76.

532 Rommers JM, Kemp B, Meijerhof R, Noordhuizen JPTM 2001a. The effect of litter size
533 before weaning on subsequent body development, feed intake, and reproductive
534 performance of young rabbit does. *Journal of Animal Science* 79, 1973-1982.

535 Rommers JM, Meijerhof R, Noordhuizen JPTM, Kemp B 2001b. Effect of different feeding
536 levels during rearing and age at first insemination on body development, body
537 composition and puberty characteristics of rabbit does. *World Rabbit Science* 9, 101-
538 108.

539 Rommers JM, Meijerhof R, Noordhuizen JPTM, Kemp B 2002. Relationships between body
540 weight at first mating and subsequent body development, feed intake, and
541 reproductive performance of rabbit does. *Journal of Animal Science* 80, 2036-2042.

542 Rommers JM, Meijerhof R, Noorhuizen JPTM, Kemp B 2004. Effect of feeding program
543 during rearing and age at first insemination on performances during subsequent
544 reproduction in young rabbit does. *Reproduction Nutrition Development* 44, 321-332.

545 Rosell JM 2000. Enfermedades de menor presentación. Enfermedades metabólicas. In:
546 Enfermedades del conejo. Tomo II. (eds JM Rosell), pp.399-454. Mundiprensa,
547 Madrid, Barcelona, México.

548 Sejrsen K, Hubert JT, Tucker HA, Akers RM 1982. Influence of nutrition on mammary
549 development in pre- and postpubertal heifers. *Journal of Dairy Science* 65, 793-800.

550 SAS 2002. SAS/SAT User's Guide (Release 9.1). SAS Inst. Inc. Cary NC, USA.

551 Sørensen MT, Danielsen V, Busk H 1998. Different rearing intensities of gilts: I. Effects on
552 subsequent milk yield and reproduction. *Livestock Production Science* 54, 159-165.

553 Theau-Clement M 2007. Preparation of the rabbit doe to insemination: a review. *World*
554 *Rabbit Science* 15, 61-80.

555 Van Soest, PJ, Robertson JB, Lewis BA 1991. Methods for dietary fiber, neutral detergent
556 fiber and non starch polysaccharides in relation to animal nutrition. *Journal of Dairy*
557 *Science* 74, 3583-3597.

558 Weigle D, Duell P, Connor W, Steiner R, Soules M, Kuijper J 1997. Effect of fasting,
559 refeeding, and dietary fat restriction on plasma leptin levels. *Journal of Clinical*
560 *Endocrinology and Metabolism* 82, 561–565.

561 Whitehead CC 1988. Selection for leanness in broilers using lipoprotein concentrations as
562 selection criterion. In: *Leanness in Domestic Birds: Genetic, Metabolic and Hormonal*
563 *Aspects* (eds B Leclercq and CC Whitehead), pp. 41-57. Butter-worhs and INRA,
564 London and Paris.

565 Xiccato G, Trocino A 2010. Energy and protein metabolism and requirements. In: *Nutrition of*
566 *the Rabbit* (eds C De Blas and Wiseman.), pp. 83-118. CABI Publishing, Wallingford,
567 UK.

568 Xiccato G, Bernardini M, Castellini C, Dalle Zotte A, Queaque PI, Trocino A 1999. Effect of
569 postweaning feeding on the performance and energy balance of female rabbits at
570 different physiological states. *Journal of Animal Science* 77, 416-426.

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Table 1 *Ingredients and chemical composition of experimental diets*

<i>Ingredient (g/kg)</i>	Diets	
	C	F
Barley	312	78
Alfalfa hay	450	570
Sunflower meal	94	51
Soya meal	85	-
Sugar beet pulp	-	152
Cereal straw	-	100
Soya oil	30	10
HCl L-lysine, 780	2	3.9
DL-methionine, 990	-	0.85
L-threonine, 980	-	1.45
L-tryptophan, 980	1	1.5
L-Arginine, 990	-	4
Dicalcium phosphate	17	1.8
Monosodium phosphate	-	16.5
Salt	5	5
Vitamin-mineral mixture ¹	4	4
<i>Chemical composition (g/kg DM)</i>		
Dry Matter (DM, g/kg)	899	900
Ash	90	103
Starch	205	63
Ether Extract	52	29
Crude Protein	179	146
Neutral Detergent Fibre	358	476
Acid Detergent Fibre	277	394
Acid Detergent Lignin	59	88
Gross Energy (MJ/kg DM)	18.24	18.67
Digestible energy (DE; MJ/kg DM)	11.03	8.72
Digestible protein (DP; g/kg DM)	114	88
DP/DE (g/MJ)	10.3	10.1

¹ Per Kg of feed: Vitamin A: 8,375 IU; Vitamin D3: 750 IU; Vitamin E: 20 mg; Vitamin K3: 1 mg; Vitamin B1: 1 mg; Vitamin B2: 2 mg; Vitamin B6: 1 mg; Nicotinic acid: 20 mg; Choline chloride: 250 mg; Mg: 290 mg; Mn: 20 mg; Zn: 60 mg; I: 1.25 mg; Fe: 26 mg; Cu: 10 mg; Co: 0.7; Butyl hydroxylanisole+ethoxyquin: 4 mg.

Table 2 Daily dry matter (g DM per kg metabolic weight ($BW^{0.75}$)), digestible energy (kJ DE per kg $BW^{0.75}$) and digestible protein (g DP per kg $BW^{0.75}$) intake of young rabbit does during rearing and first pregnancy (mean \pm standard error)

	Feeding programme ¹				
	CAL		F		
9-12 wk					
DM intake	82.44 ^a \pm 1.21		94.05 ^b \pm 0.94		
DE intake	909.3 ^b \pm 12.3		819.7 ^a \pm 9.5		
DP intake	93.86 ^b \pm 1.28		83.00 ^a \pm 0.99		
12-16 wk					
	CAL	CR	F		
DM intake	75.46 ^b \pm 1.62	63.34 ^a \pm 1.72	97.43 ^c \pm 0.91		
DE intake	832.3 ^b \pm 16.3	698.6 ^a \pm 12.3	849.0 ^b \pm 9.2		
DP intake	85.91 ^b \pm 1.7	72.11 ^a \pm 1.8	85.96 ^b \pm 0.96		
16-18 wk					
	CAL	CR	F	FC	
DM intake	68.58 ^b \pm 1.50	63.17 ^a \pm 1.59	90.54 ^d \pm 1.5	84.28 ^c \pm 1.03	
DE intake	756.4 ^b \pm 15.6	696.7 ^a \pm 16.5	789.0 ^b \pm 15.5	929.5 ^c \pm 10.6	
DP intake	78.07 ^b \pm 1.60	71.91 ^a \pm 1.69	79.88 ^b \pm 1.6	95.94 ^c \pm 1.09	
Early pregnancy; 18-20 wk					
DM intake	66.14 ^b \pm 1.5	61.78 ^a \pm 1.59	91.09 ^d \pm 1.48	75.61 ^c \pm 1.03	
DE intake	729.5 ^b \pm 15.6	681.3 ^a \pm 16.5	793.9 ^c \pm 15.3	833.9 ^d \pm 10.6	
DP intake	75.29 ^b \pm 1.6	70.33 ^a \pm 1.69	80.38 ^c \pm 1.57	86.08 ^d \pm 1.09	
Late pregnancy; 20-23 wk					
	CAL	CR	F	FC	FCF
DM intake	48.68 ^a \pm 1.68	53.13 ^b \pm 1.61	73.18 ^d \pm 1.5	56.42 ^b \pm 1.47	61.35 ^c \pm 1.55
DE intake	536.8 ^a \pm 17.2	585.9 ^b \pm 16.5	637.8 ^c \pm 15.3	622.3 ^{bc} \pm 15.0	586.5 ^b \pm 15.9
DP intake	55.41 ^a \pm 1.76	60.48 ^{bc} \pm 1.69	64.57 ^c \pm 1.57	64.23 ^c \pm 1.55	59.79 ^{ab} \pm 1.63

¹ Feeding programme: CAL group received the C diet *ad libitum* until 1st partum; CR group received the C diet *ad libitum* until 12 wk and then, 140 g per day until 1st partum; F group received the F diet *ad libitum* until 1st partum; FC and FCF group received F diet *ad libitum* until 16 wk and then, FC group received the C diet *ad libitum* until 1st partum and FCF group the C diet *ad libitum* until 20 wk and then the F diet *ad libitum* until 1st partum.

^{a,b,c,d} Means within a row not sharing any superscript are significantly different at $P < 0.05$.

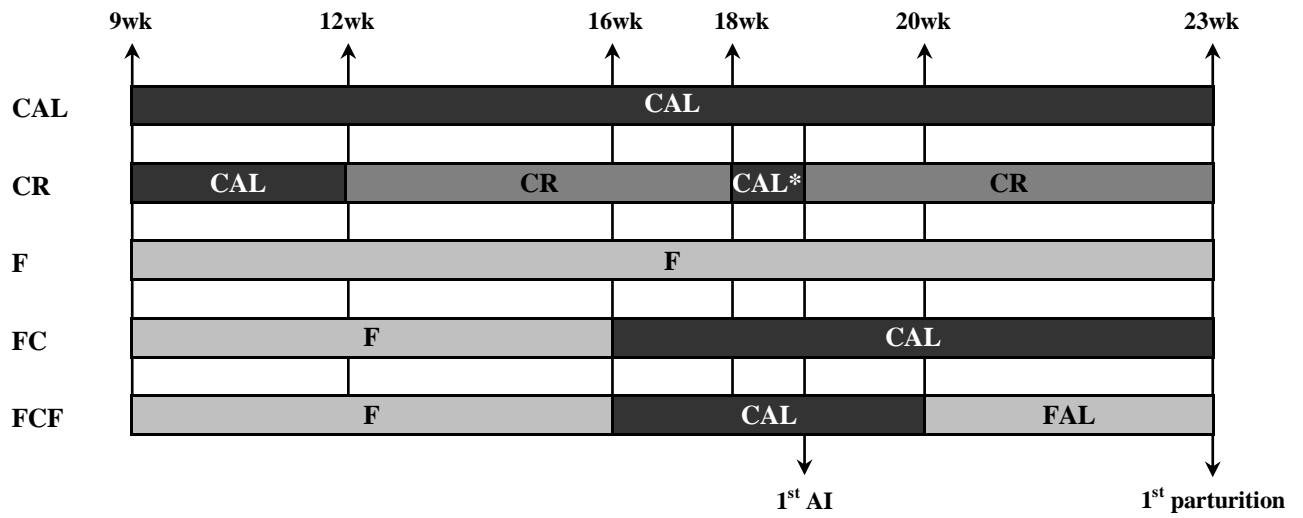
Table 3 Litter size and weight at first partum (mean \pm standard error)

	Feeding programme ¹				
	CALminusc ula?	CR	F	FC	FCF
<i>Litter size at partum</i>					
Total	6.6 ^a \pm 0.6	6.8 ^{ab} \pm 0.6	6.9 ^{ab} \pm 0.06	7.0 ^{ab} \pm 0.6	8.3 ^b \pm 0.6
Alive	5.2 ^a \pm 0.7	6.2 ^{ab} \pm 0.7	6.0 ^{ab} \pm 0.6	6.1 ^{ab} \pm 0.6	7.7 ^b \pm 0.6
<i>Mortality at birth</i>					
	20.6 ^c	8.3 ^{ab}	12.1 ^{ab}	12.4 ^b	6.7 ^a
<i>Litter weight at partum (g)</i>					
Total	340 ^a \pm 25.4	391 ^{ab} \pm 25.4	406 ^{ab} \pm 23.6	368 ^a \pm 23.2	448 ^b \pm 24.5
Alive	280 ^a \pm 31.6	355 ^{abc} \pm 31.6	369 ^{bc} \pm 29.3	323 ^{ab} \pm 28.8	419 ^b \pm 30.4

¹ Feeding programme: Abbreviations as in Table 2.

^{a,b} Means within a row not sharing any superscript are significantly different at $P < 0.05$.

Figure 1 Diagram of the different feeding programmes carried out during rearing and first pregnancy for the 5 experimental groups (CAL, CR, F, FC and FCF)



*Flushing 4 days before insemination

CAL: C diet *ad libitum*; CR: C diet restricted at 140 g per day; F: F diet *ad libitum*

AI: Artificial Insemination

Figure 2 Percentage of does dead during the rearing and first pregnancy (from 9 to 23 week of age) with the different feeding programmes (abbreviations as in Table 2). Bars within a period not sharing any superscript are significantly different at $P < 0.05$. AI: Artificial Insemination

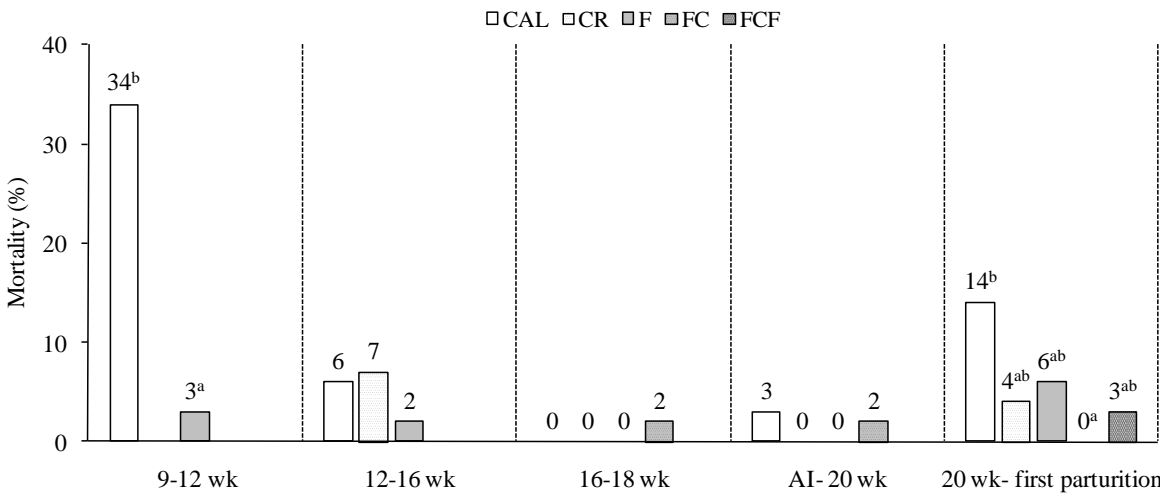


Figure 3 Live weight evolution of young rabbit does during rearing and first pregnancy (9 to 23 wk of age) with the different feeding programmes (abbreviations as in Table 2). Data at 9 week of age are presented as a whole. Bars not sharing any superscript are significantly different at $P < 0.05$

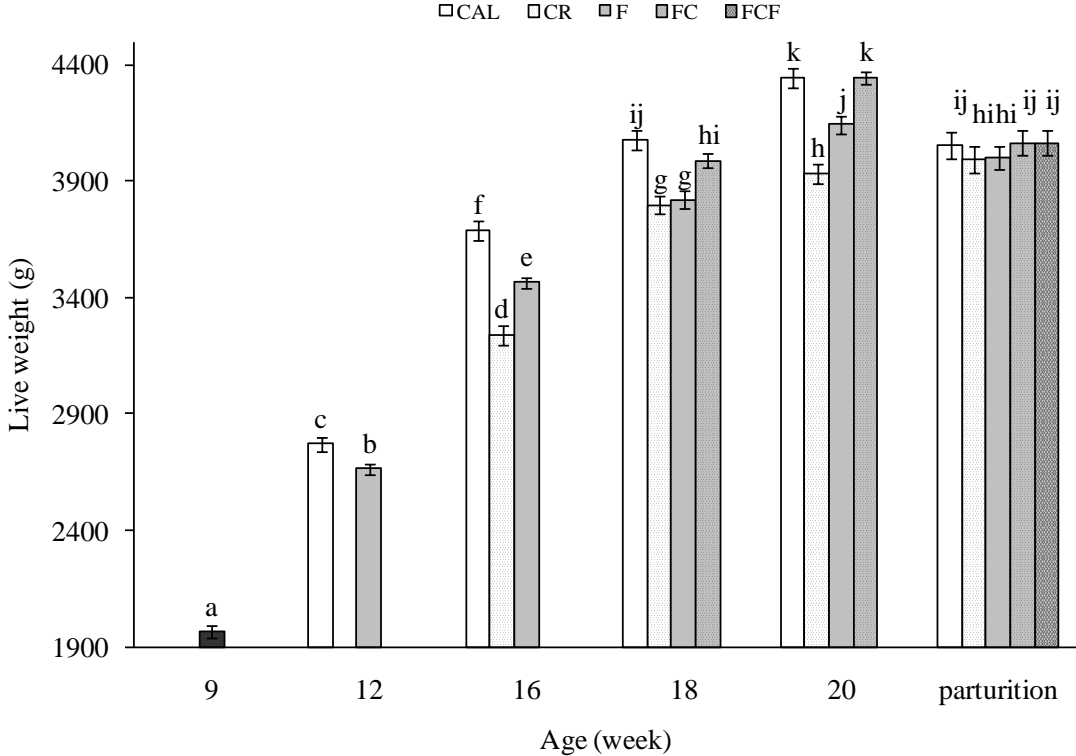


Figure 4 Perirenal fat thickness evolution of young rabbit does during rearing and first pregnancy (9 to 23 wk of age) with the different feeding programmes (abbreviations as in Table 2). Data at 9 week of age are presented as a whole. Bars not sharing any superscript are significantly different at $P < 0.05$

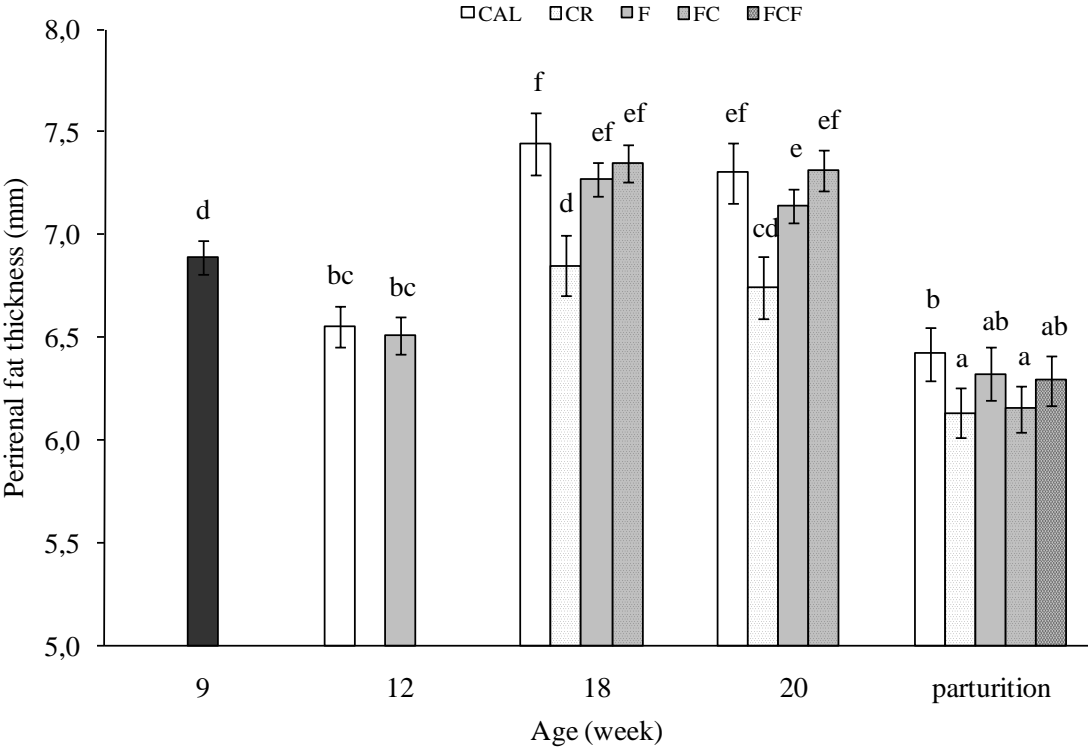


Figure 5 Evolution of blood plasma (a) insulin, (b) glucose, (c) non esterified fatty acids (NEFA), (d) leptin, (e) cortisol and (f) tri-iodothyroxine (T3) concentrations in young rabbit does during rearing and first pregnancy (9 to 23 wk of age) with the different feeding programmes (□ CAL □ CR □ F □ FC □ FCF; abbreviations as in Table 2). Data at 9 week of age are presented as a whole. Bars not sharing any superscript are significantly different at $P < 0.05$

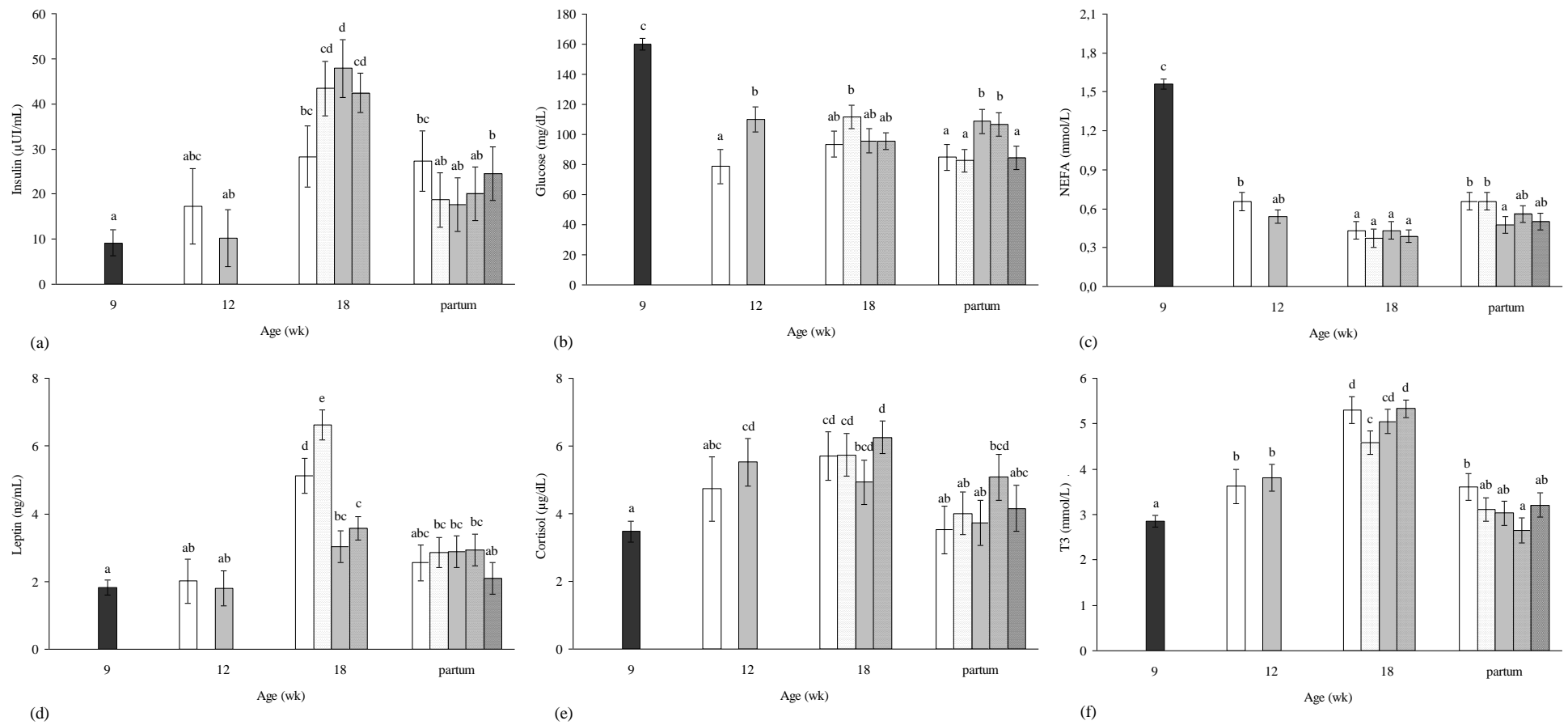


Figure 6 Estimated energy content of young rabbit does at effective artificial insemination (AI) and parturition days with the different feeding programmes (abbreviations as in Table 2). Bars not sharing any superscript are significantly different at $P < 0.05$

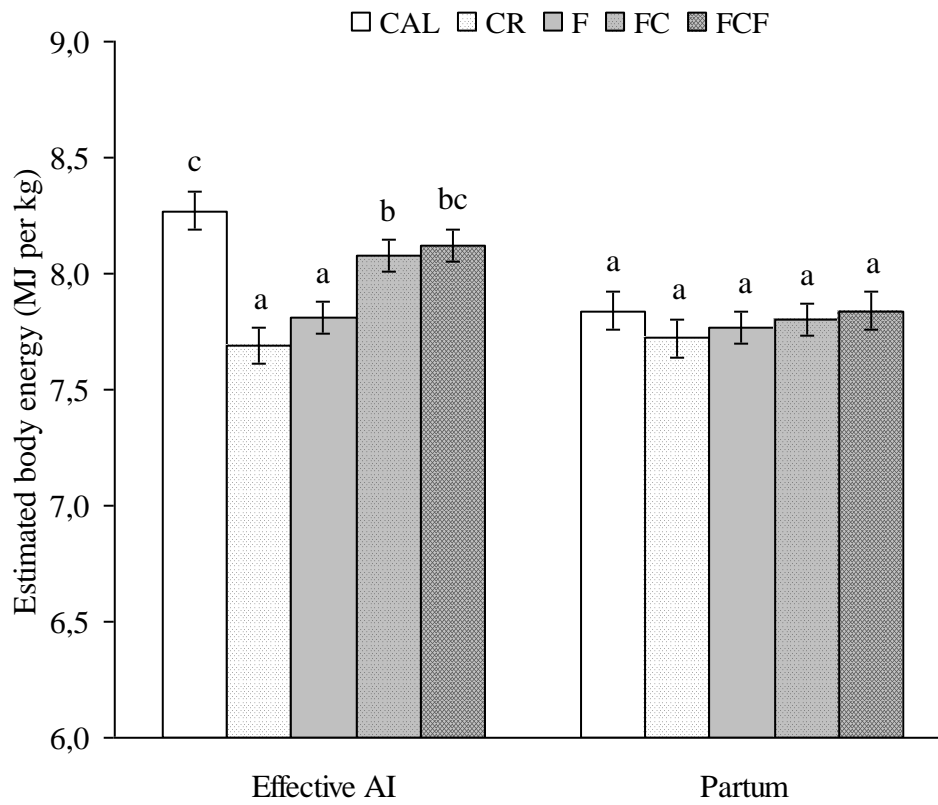


Figure 7 Relationship between leptin levels in the blood of young rabbit does at first mating (16-18 wks of age) and the fertility observed during the first reproductive cycle. Data obtained from the present results and three previous works of the literature

