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Additional Information

1 Non-destructive assessment of the internal quality of intact persimmon using colour and VIS/NIR hyperspectral imaging 2

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18 Abstract

19 The internal quality of intact persimmon cv. 'Rojo Brillante' was assessed trough 20 visible and near infrared hyperspectral imaging. Fruits at three stages of commercial 21 maturity were exposed to different treatments with CO₂ to obtain fruit with different ripeness and level of astringency (soluble tannin content). Spectral and spatial 22 23 information were used for building classification models to predict ripeness and 24 astringency trough multivariate analysis techniques like linear and quadratic 25 discriminant analysis (LDA and QDA) and support vector machine (SVM). Additionally, flesh firmness was predicted by partial least square regression (PLSR). 26 27 The full spectrum was used to determine the internal properties and later principal 28 component analysis (PCA) was used to select optimal wavelengths (580, 680 and 1050 29 nm). The correct classification was above 92% for the three classifiers in the case of ripeness and 95% for QDA in the case of astringency. A value of $R^2 = 0.80$ and a ratio 30 31 of prediction deviation (RPD) of 1.86 were obtained with the selected wavelengths for 32 the prediction of firmness which demonstrated the potential of hyperspectral imaging as

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a non-destructive tool in the assessment of the firmness, ripeness state and astringency
level of 'Rojo Brillante' persimmon.

35 *Keywords*: *Diospyros kaki*, internal fruit quality, soluble tannins, astringency,
36 classification, computer vision.

37

38 **1. Introduction**

Spain is one of the major producers of persimmon (*Diospyros kaki* L.) among European countries (Plaza et al., 2012). The principal variety grown in Spain is 'Rojo Brillante', mostly located in the region of Ribera del Xuquer Valley near Valencia (Spain) with more then 100.000 T per year. This cultivar is very appreciated by consumers because its good aspect, high size, flavour and absence of seeds. However, this cultivar is astringent at harvest and the fruit cannot be consumed until a high degree of overripeness when allowed to rest and soften for a long period after harvest.

46 This has been traditionally a handicap for the commercialization of this fruit since once 47 the fruit losses the astringency by overripe, it acquires a soft jelly-like consistency being 48 difficult to handle and eat. Now, some methods have been developed to eliminate 49 quickly the astringency without losing the firmness, as exposing fruit to high CO₂ 50 concentrations (95-100%) during 18 to 24 hours. This method is based on promoting 51 anaerobic respiration in the fruit, giving rise to an accumulation of acetaldehyde which 52 reacts with the soluble tannins that are the responsible for the astringency (Matsuo et al., 53 1991). In figure 1 can be appreciated the differences between a persimmon naturally 54 deastringed by overripeness and another deastringed using a CO₂ treatment. Since the 55 success of the treatment was demonstrated (Salvador et al., 2007; Besada et al., 2010), it 56 has been adopted by industry as the standard deastringency method, and utilized to give 57 the fruit in addition a sweet taste and firm texture similar to the apple, highly appreciated by the consumers. However, the effectiveness depends on the fruit firmness at harvest, since maturation process is accompanied by a gradual decrease of firmness (Salvador et al., 2008). A problem is that the stage of maturity at harvest is currently determined based on the visual inspection of experienced growers or using colorimeters due the relationship between the changes in external colour and the internal changes (Salvador et al., 2006 & 2007).

The current way to know the level of astringency in the fruit after CO_2 -treating is by destructive measurement of soluble tannin content (ST) in random fruits by means of the tannin print method (Matsuo and Ito, 1982) which consists of using a FeCl₃impregnated filter paper to obtain a print of the content and distribution of the tannins trough the reaction with the FeCl₃ in the paper. Then, this print is visually assessed by trained workers being this method subjective and destructive and therefore the development of other new non-destructive and accurate methods is needed.

Computer vision systems have been traditionally used to create tools for the objective estimation of the quality of intact fruit production (Cubero et al., 2011) and have already been explored to assess quality of persimmon. Mohammadi et al., (2015) used colour information to determine the maturity of this fruit through colour analysis and classify the fruit into three commercial maturity stages.

Standard computer vision systems tend to mimic the human eye and hence are based on sensors sensible to visible wavelengths. But to analyse internal composition it is necessary the use of technology sensible to non-visible wavelengths related with chemical compounds. This can be achieved by using hyperspectral imaging (Lorente et al., 2012) that is a powerful non-invasive technology that allows obtaining the spatial distribution of the spectral information and it is being used from recent in the internal quality inspection of food (Cheng et al., 2016a; Cheng et al., 2016b, Gómez-Sanchis et

al., 2013) or to assess some properties of fruits like the ripeness in apples (ElMasry et
al., 2008), citrus fruits (Folch-Fortuny et al., 2016), pepper (Schmilovitch et al., 2014),
or mango (Velez-Rivera et al., 2012).

Hyperspectral imaging in persimmon has been used by Munera et al., (2017) to create 86 87 images showing the distribution of the predicted astringency of each pixel in the fruit, 88 and by Wei et al., (2014) to predict firmness. However, in this work, the authors 89 claimed that more research is needed to include more samples as well as different 90 regions and different postharvest treatments to ascertain the discrimination power of this 91 method and it is therefore necessary to investigate new methods especially to 92 discriminate among fruits with slightly different stages of maturity or levels of 93 astringency as those exposed to a CO_2 treatment, to achieve a demand from both the 94 industry and the consumers. This work proposes a new non-destructive approach based 95 on visible and near infrared (VIS/NIR) hyperspectral imaging and multivariate analysis 96 to determine the firmness, ripeness state and astringency level of intact persimmon 97 'Rojo Brillante' as alternative to the current destructive and/or subjective techniques.

98 2. Materials and methods

99 2.1 Plant material and internal quality assessments

100 A total of 90 persimmon (Diospyros kaki cv. 'Rojo Brillante') fruits were harvested in 101 L'Alcudia (Valencia, Spain) at three different stages of commercial maturity (M1, M2 102 and M3) corresponding to different moments of the season (early November, end 103 November, and mid December). A total of 30 fruits, with apparently similar size and 104 colour were collected for each maturity stage. In order to obtain three different levels of 105 astringency, the fruits in each maturity stage were equally divided into three sets. The 106 first set (control fruits with high astringency, HA) consisted of fruits not treated, the 107 second set (medium astringency fruits, MA) consisted of fruits treated in closed 108 containers at 20 °C with 90% of relative humidity (RH) and 95% of CO_2 for a period of 109 12 h, and the remaining set (non astringent fruits, NA) were fruits treated under the 110 same conditions for 24 h.

111 After each treatment, all the fruits were measured using a colorimeter, a digital camera, 112 and a hyperspectral imaging system. Later, flesh firmness of all fruits was determined 113 by means of a universal testing machine (4301, Instron Engineering Corp., MA, USA) 114 equipped with an 8 mm puncture probe. The crosshead speed during the firmness 115 testing was 10 mm/min. During the test, the force increased smoothly until it drastically 116 decreased when the flesh was broken and the maximum peak force was registered. 117 Results were expressed as the mean of the load (in N) required for breaking the flesh of 118 the fruit on the two sides after peel removal. To analyse the astringency of the fruits, 119 they were sliced and frozen at -20 °C to determine soluble tannins using the Folin-120 Denis method (Taira, 1995), as described by Arnal and Del Río (2004). This method is 121 based on the reduction of the Folin-Ciocalteu reagent by soluble tannins in alkaline 122 solution. Calibration curve was made with gallic acid. Soluble tannins were extracted by 123 homogenization of 5 g of flesh with 25 mL of 80% methanol solution. Thereafter, 124 samples were filtered and centrifuged for 20 minutes and the supernatant was reserved. 125 More supernatant was extracted from the precipitant with methanol 80% and added to 126 the first. The supernatant was diluted in water at 1:7 and then Folin-Ciocalteu reagent 1 127 N was used to conduct the reaction. After 3 minutes 1 ml of saturated Na₂CO₃ was 128 added, and the absorbance of the mixture at 725 nm was measured by colorimetry after 129 stand for 1 h.

130 **2.2. Colour analysis**

At harvest this fruit presents a uniform colour that ranges from bright to dark orangedepending on the maturity being the colour a good indicative of this property (Salvador

133 et al., 2007). The external colour the fruit under study was characterised using two 134 techniques. On the one hand, a colorimeter (CR-300, Konica Minolta Inc, Tokyo, 135 Japan) was used to obtain the colour at three points of the equatorial part of the fruit. 136 Hunter Lab colour coordinates were obtained by the average of three measures. On the 137 other hand, the colour was also evaluated trough images of the two sides of each fruit. 138 The image acquisition system consisted on a digital camera (EOS 550D, Canon Inc, 139 Japan) arranged into a squared inspection chamber that included a calibrated and 140 uniform illumination system composed of eight fluorescent tubes (BIOLUX 18W/965, 141 6500 K, Osram GmbH, Germany). The angle between the axis of the lens and the 142 sources of illumination was approximately 45° to avoid direct reflections to the camera 143 (Diago et al., 2015), but due to the spherical shape of the samples these reflections 144 could not be totally avoided this way and hence cross-polarization was also used 145 (ElMasry et al., 2012).

146 A total of 180 images were obtained with a size of 2592 x 1944 pixels and a resolution 147 of 0.11 mm/pixel. Figure 2 shows examples of images of the fruits in the three maturity 148 stages. For each image, the mean red, green and blue (RGB) colour values of the pixels 149 of the skin were obtained using the application Food ColorInspector (free download at 150 http://www.cofilab.com). RGB values were later converted to Hunter Lab colour space 151 for analysis using the equations described in Mendoza et al., (2006) and HunterLab 152 (1996) for illuminant D65 and standard observer 10°. The Hunter Lab coordinates were 153 finally transformed to the colour attributes Hunter luminosity (L), Hunter hue (h) and 154 Hunter chroma (C) (Hutchings, 1999). In addition, RGB values were transformed into 155 HSI (hue, saturation, intensity) values and other indices were estimated such as the 156 ratios a/b and a/L and the colour index (CI=1000a/Lb) (Salvador et al., 2006).

157 2.3 Hyperspectral imaging

Hyperspectral images of the intact persimmons in the spectral range 450-1020 nm were acquired using a camera (CoolSNAP ES, Photometrics, USA) coupled to two liquid crystal tuneable filters (LCTF) (Varispec VIS-07 and NIR-07, Cambridge Research & Instrumentation, Inc., MA, USA). The illumination system consisted of 12 halogen lights arranged equally into a domo inspection chamber where whole fruits were manually introduced (Figure 3).

164 Hyperspectral images with a spatial resolution of 0.14 mm/pixel and a spectral 165 resolution of 10 nm were captured in both sides of each fruit (Figure 4), which lead to a 166 tagged database of 180 hyperspectral images. In each image, a region of interest (ROI) 167 of 225×225 pixels in the central part of the fruit was selected and analysed as the 168 average of spectrum of all pixels for maturity and firmness analysis since these 169 properties are quite uniformly distributed in the fruit. However, for the case of the 170 astringency, the individual spectrum of each pixel in the ROI was included in the 171 models due the uneven distribution in the fruit of tannins responsible of the astringency. 172 To obtain the relative reflectance of a pixel in the position (x, y) of the monochromatic 173 band λ , the original reflectance was corrected using a dark and white reference 174 (Spectralon 99%, Labsphere, Inc, NH, USA) following the procedure described in Gat 175 (2000).

176 **2.4 Data analysis**

Analysis of variance (ANOVA) and Tukey multiple range test (Statgraphics Centurion XVI - Statpoint Technologies Inc., Virginia, USA) were used to show the effects of ripeness on colour parameters obtained with both, colorimeter and computer vision system. In this analysis, the three maturity stages were the observed values (*Y*) and the Hunter Lab colour coordinates captured by both the colorimeter and the vision systems were the predictive variables.

183 Hyperspectral images consisted of 67 wavelengths and therefore the spectra obtained 184 from these images were distributed in a matrix with 67 columns each corresponding to 185 the reflectance value of each band where the rows represented the fruits. In addition, the 186 pixels were labelled as belonging to any of the maturity stages (M1, M2 and M3) and 187 treatments (HA, LA, NA) to carry out the analysis for firmness and astringency 188 prediction. First step was a preprocessing of data using Standard Normal Variate (SNV) 189 to remove scatter effects from original spectral data (The Unscrambler X 10.1, CAMO 190 Software, Oslo, Norway). Classification models to sort the fruit by ripeness stage and 191 treatment duration (astringency level) were developed using linear and quadratic 192 discriminant analysis (LDA & QDA), and support vector machine (SVM) (Dutta et al., 193 2016). The difference between LDA and QDA classifier is that LDA uses pooled 194 covariance to assign an unknown sample to one of the pre-defined groups while QDA 195 uses the covariance of each group instead of pooling them (Naes et al., 2002). On the 196 other hand, the SVM algorithm was developed based on the concept of hyperplane and 197 support vectors, using a linear function kernel with C value set to 1. In addition, 198 firmness prediction was conducted by partial least square regression (PLSR) (Cheng et 199 al., 2015b) using the ratio of prediction deviation (RPD), that was defined by Williams 200 (1987) as the ratio of standard deviation of reference values in training set to the root 201 mean square error of prediction (RMSEP).

Hyperspectral systems capture a huge amount of information that is redundant and correlated, especially between contiguous wavelengths (Lorente et al., 2012). Therefore, principal component analysis (PCA) was used to know if it was possible to obtain good prediction using a reduced subset of bands. Four different PCA models were built, one of using the spectral data of the ripeness assessment and the other three PCA with data of the astringency assessment for each harvest. The variables (wavelengths) were 208 chosen on the basis of the size of coefficients or loadings in the eigenvectors of the209 principal components.

210 **3. Results and discussion**

211 **3.1. Maturity assessment**

212 Several differences can be observed among the spectra of the fruit in the three maturity 213 stages shown in Figure 5. Fruits of M1 gave higher reflection values than the others in 214 the visible region, which is in agreement with the colour analysis. An absorption peak 215 was found around the bands 670-680 nm only for fruits in M1 stage which could be due 216 the presence of chlorophyll in the more unripe fruit (Lleó et al., 2011). However, the 217 fruits in M2 stage are those which gave a higher reflection in the NIR region that can be 218 due to the chemical differences among fruit at different ripeness. The absorption peak 219 observed around 900-1050 nm could be assigned to water absorption band. This peak 220 was higher in M3, which may be related to water content increases in the flesh during 221 the onset of ripening, which in other fruits has been related to cell breakage and osmotic 222 movement of water from the flesh to the peel.

223 The PCA model generated with the 67 wavelengths was analysed to identify the 224 variable with the highest factor loadings since they reflected the importance of each 225 wavelength in discriminating differences in the fruit (Wang et al., 2012). The loadings 226 of the first two principal components were used for wavelength selection because these 227 were responsible for 96% of the variance in the spectral data. The wavelengths 228 corresponding to higher module values (peaks and valleys) at these particular principal 229 components were selected as candidates for optimum wavelengths (Rodríguez-Pulido et 230 al., 2013) (Figure 6). Four optimum wavelengths (450, 580, 680, and 1050 nm) were 231 thus identified for discrimination purposes of different maturity stages. Wavelengths 232 450 nm and 680 nm are related with the presence of beta-carotene and chlorophyll a

respectively. On the other hand, the importance of the wavelength 580 nm can be due to the colour changes during ripeness since it corresponds to the yellow colour. This would be in accordance with the ranges of h and C values shown in Table 3. The band 1050 nm could be related with an absorption region of water content although the peak is situated below 1000 nm (Lu and Peng, 2006).

Statistical models to classify the fruit into maturity stages were developed using the spectra of the full spectra and only the selected wavelengths. In order to build and validate the model, a 3-fold cross validation procedure was used (Simon, 2007). The data set of pixels was randomly partitioned into three disjoining subsets. The classifier development process was repeated three times using each two different subsets and the resulting classifiers used to classify the remaining test set. Finally the results of the three iterations were averaged.

The four selected bands were used to build the models but also the possible combinations of three bands resulting that using only 580 nm, 680 nm, and 1050 nm, the results were similar to those achieved using the four bands. Using only these three selected wavelengths the success rate of correct classification was slightly lower (mean value of 94.8%) than using the full spectrum (mean value of 98.5%) as shown in Table 1.

251 Comparing the three classification methods, all of them achieved a good classification 252 above 98% using the all wavelengths. Moreover, using only the three selected 253 wavelengths only LDA showed an important reduction in the success rate while the 254 other two classifiers still remain above 95% which is considered as a good result for a 255 non-destructive technique.

256 **3.2. Firmness prediction**

257 Table 2 shows the firmness evolution with the harvesting time (ripeness). A model 258 based on PLSR was built to know if it was possible to predict this property in this 259 cultivar using the wavelengths selected in the previous study for ripeness assessment. 260 For each fruit there was obtained only one global value of the flesh firmness so the 261 prediction model was built using the average values of the pixels selected for each fruit 262 at the determined wavelengths of the hyperspectral images. Cross validation leaving 5% 263 of samples for test was chosen to validate this study. This method splits randomly the 264 data set into the training (95%) and test (5%), repeating the process 20 times. Results 265 were achieved as the mean of the 20 repetitions.

The coefficient of determination for the prediction (R^2_P) was 0.80 and the RPD was 1.86 266 267 \pm 0.26. Viscarra-Rossel et al., (2006) suggested that calibration models will suffice for 268 good quantitative application if RPD is larger than 1.8. The prediction results obtained 269 was something higher than the minimum proposed but not as good as the prediction results of Wei et al., (2014) for 'Fangshi' persimmon who achieved a R² value of 0.91. 270 271 However, in their work the firmness of the fruit ranged from 25 N to 1 N with large 272 differences among the studied classes. In addition, during the ripening process of this 273 cultivar not only drastic changes in firmness happened but also the skin begins to 274 wrinkle and lose shine clearly affecting the reflectance. On the contrary, in the present 275 work, the firmness gave values from 47 N to 21 N which means that these fruits are 276 apparently firm in all maturity stages, which is logical since it is treated to be consumed 277 as firm and crispy fruit. Figure 1 highlights the visual differences between a soft 278 persimmon naturally deastringed and another deastringed a using CO₂ treatment. Hence, 279 for this fruit 80% of prediction capability is considered as a good achievement taking 280 into account the little differences between classes, especially between M2 and M3 281 classes.

282 A study was also carried out to analyse the possible correlation between the colour 283 analysis and the firmness of the samples. The characterisation of the external colour was 284 carried out using the colorimeter and the camera only for the control samples of the 285 three stages to avoid the influence of the treatment in colour changes (Table 3). In 286 general, the L and b, Hunter Lab coordinates, decreased but there were not statistical 287 differences for M2 and M3. On the contrary, the value of a increased along the three 288 stages. As a consequence of the changes observed in a and b, the hue decreased and the 289 chroma slightly increased along the three stages. These differences were observed in the 290 measures given by both, the colorimeter and the camera, and reflect the loss of 291 luminosity of the fruit caused by the maturity process and the changes in the fruit from 292 yellowish-orange to reddish-orange.

293 The values of the colour attributes (L, h and C) of the colorimeter were higher than the 294 ones obtained from the images. The higher differences were observed for the L values 295 since the glossiness leads to a specular reflectance that reduces the contribution to the 296 components a and b. In fact, colorimeter is very dependent on the scattering properties 297 of the sample while the diffuse illumination of the vision system gives less dependency 298 on the lightness of the sample than the simple illumination and filtering employed by 299 the colorimeter (Trinderup et al., 2015). Despite the differences observed, good correlations where found between the values obtained by both methods (R^2 of 0.87, 300 301 0.80, and 0.96 for the L, chroma, and hue respectively).

Linear regressions were performed between the different colour values, obtained with both the colorimeter and the camera, and the firmness. Table 4 and 5 summarise the results achieved for the coefficient of determination R^2 for each colour component using the imaging system and the colorimeter, respectively. In general, better results are achieved with the imaging system which on the other hand makes sense since they integrate the colour of the whole surface of the fruit while colorimeter only measures ina small spot and thus increasing the variability.

Good correlations are found in H (R²=0.83), G (R²=0.82) and h (R²=0.81) or using simple ratios like a/b (R²=0.83), G/R (R²=0.83) or a/L (R²=0.83). It is worthy of interest that using the simple ratios measured with the imaging system could be obtained better correlations (R²=0.83) than using the *CI* (*CI*=1000a/Lb) that was the index used by Salvador et al., (2006) to estimate the firmness trough a colorimeter achieving a R²=0.81.

315 **3.1.** Astringency prediction

316 The results of the measurements of soluble tannins of fresh weight for all maturity 317 stages are shown in Table 6. It can be observed that the tannin content decreased in a 318 similar way for the three maturity stages along with the duration of the treatment. The 319 soluble tannins content decreased to values close to 0.4% in the fruits treated for 12 h to 320 0.03% in the fruits exposed for 24 h to CO₂. Accordingly, Besada et al., (2010) reported 321 that the CO₂-treatment applied for 12 h to fruit with firmness around 40 N led to a 322 reduction of soluble tannins to values close 0.3%. Besides, it has been widely reported 323 that a content of soluble tannins of 0.03% after the CO₂-treatment is associated with a complete effectiveness of the deastringency process in 'Rojo Brillante' cultivar 324 325 (Salvador et al., 2007; Salvador et al., 2008).

Like in ripeness classification, three PCA models were analysed to identify the highest factor loadings in each ripeness stage. However, no wavelength selection could contribute to the astringency classification. This may be because tannins are mainly detected in the ultraviolet (UV) in the range 190 to 400 nm (Boulet et al., 2016), or in the NIR (2200 to 2300 nm) (Cozzolino et al., 2004). For this reason, the whole spectrum in the studied range (450-1020 nm) was necessary to discriminate the astringency. 332 Table 7 shows the results of astringency classification using the three classifiers. In 333 general, QDA obtained the best overall classification but a reduction of the 334 classification rate along with the maturity was observed, especially for the astringent 335 fruits (HA and LA) in the M3 stage. As it was shown in Table 2, a decrease of firmness 336 in M1 between control and non astringent fruits was observed. However, in M2 and M3 337 there was no difference. This could be due because the effect of high CO₂ 338 concentrations on the cell structure could be the cause of the important loss of firmness 339 observed after deastringency treatment. But when the more ripe samples are treated with CO_2 , no effect happens on flesh firmness because the loss of intercellular adhesion is 340 341 already generalised due to the ripeness process (Salvador et al., 2007). Therefore, those 342 changes detected by hyperspectral imaging are assigned to changes in the soluble 343 tannins content and not to changes in texture.

4. Conclusions

In this study, VIS/NIR hyperspectral imaging were evaluated as potential nondestructive methods to determine the flesh firmness, maturity stage and the astringency
level of 'Rojo Brillante' persimmon.

348 The characterisation of the colour showed that the L and b, Hunter Lab coordinates 349 decreased while the value of *a* increased along with the maturity. As a consequence the 350 hue decreased and the chroma slightly increased along the three stages using both 351 colorimeter and image methods. Good correlations were found in some colour parameters like $H(R^2=0.83)$, $G(R^2=0.82)$ and $h(R^2=0.81)$, but also using ratios like a/b352 $(R^2=0.83)$, G/R $(R^2=0.83)$ and a/L $(R^2=0.83)$ with the data obtained by the imaging 353 354 system improving previous results. Moreover, better correlations were obtained using these ratios than using the previously proposed CI ($R^2=0.80$) which indicates the 355

feasibility of images to assess the colour as a valid alternative to traditional andexpensive colorimeters.

358 Using the hyperspectral system, three wavelengths (580, 680 and 1050 nm) were 359 proposed as the optimum wavelengths for the classification of the fruits into three 360 ripeness stages with high accuracy, more than 94% of all samples were well classified 361 for all of the used classifiers (LDA, QDA and SVM). Moreover, these wavelengths 362 were used for flesh firmness prediction and the RPD value indicated that the obtained 363 model is useful for good quantitative application. Regarding the astringency, the whole 364 spectrum of the fruits needed to be used to classify the fruits into three levels of 365 astringency: astringent fruit, fruit with a low-medium level of astringency and non-366 astringent fruit. The overall classification for the three ripeness stages was higher than 367 90% for the three classifiers and higher than 95% for QDA. These results indicate the 368 potential proposed methodology based on hyperspectral imaging as a promising non-369 destructive tool to assess the internal quality of persimmon fruits destined to be 370 deastringed and rapidly marketed as fresh sweet fruit. However, more research is 371 needed, involving more fruits from different regions and collected in different seasons 372 to ascertain the discrimination power of the proposed methodology in other markets and 373 conditions.

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492 Tables

Table 1. Ripeness classification of testing set by LDA, QDA and SVM using all and selected wavelengths (cross validation)

Class	A	All wavelength	S	Selected wavelengths			
Class	LDA	QDA	SVM	LDA	QDA	SVM	
M1	$99.5\pm0.8^{\rm a}$	99.8 ± 0.4^{a}	99.1 ± 1.0^{a}	$98.6\pm0.0^{\rm a}$	99.3 ± 0.7^{a}	98.4 ± 0.4^a	
M2	96.8 ± 3.2^{a}	96.2 ± 2.9^{a}	$96.0\pm2.7^{\rm a}$	95.5 ± 2.5^{a}	$94.1\pm2.2^{\rm a}$	94.7 ± 2.6^{a}	
M3	$99.0\pm0.8^{\rm a}$	100 ± 0.0^{a}	$99.8\pm0.4^{\rm a}$	83.7 ± 2.3^a	$93.9\pm2.9^{\text{b}}$	$94.9 \pm 1.5^{\text{b}}$	
Total	98.5 ± 2.1^{a}	98.8 ± 2.2^{a}	$98.3\pm2.3^{\rm a}$	92.6 ± 7.1^a	95.8 ± 3.3^{a}	96.0 ± 2.4^{a}	
Values a	re the mean of	three models ±	standard deviation	n. Different supe	erscript letters ir	the same row	

indicate significant differences between groups (p-value<0.05), according to Tukey's test.

500	Table 2. Flesh firmness (in N) of 'Rojo Brillante' persimmon fruits before and after
501	treatments in the three ripeness stages

	treatments in the	three ripeness st	uzes
Group	M1	M2	M3
HA	$47.0^{\mathtt{a}}\pm4.3_{\mathtt{a}}$	$29.0^{a}\pm2.6_{b}$	$25.1^{a} \pm 3.4_{c}$
LA	$44.7^{ab}\pm2.6_a$	$30.9^a \pm 3.0_b$	$25.0^{\text{a}}\pm4.7_{\text{c}}$
NA	$40.6^b \!\pm 2.8_a$	$31.9^a \pm 2.1_b$	$21.1^{a}\pm4.8_{c}$

Values are the flesh firmness $(N) \pm$ standard deviation. Different superscript letters in the same column (astringency) and different subscript letters in the same row (ripening) indicate significant differences between groups (p-value<0.05), according to Tukey's test.

Table 3. Colour coordinates and attributes of the samples in the three harvests

Stage	Colorimeter						Imaging				
Stuge	L	а	b	h	С	-	L	а	b	h	С
M1	58.93 ±1.83 ^a	21.71 ±3.29 ^c	34.84 ±1.75 ^a	60.29 ±4.41 ^a	40.67 ±1.61 [°]		43.75 ±1.03 ^a	27.82 ±3.54°	26.03 ±0.53 ^a	49.51 ±4.42 ^a	36.32 ±2.00°
M2	53.49 ±1.94 ^b	34.49 ± 1.80^{b}	31.20 ± 1.41^{b}	46.46 ± 4.60^{b}	45.42 ±1.64 ^b		33.88 ±2.44 ^b	38.30 ± 2.54^{b}	20.51 ±1.35 ^b	34.46 ±3.78 ^b	42.07 ±1.64 ^b
M3	52.64 ± 1.38^{b}	38.38 ± 1.65^{a}	30.77 ±1.09 ^b	39.20 ±1.90 ^c	48.29 ±0.53 ^a		34.71 ±1.95 ^b	41.24 ± 1.40^{a}	21.02 ± 1.05^{b}	28.62 ±1.78 ^c	43.64 ±1.26ª

Values are the mean of control samples in each harvest± standard deviation. Different superscript letters
 in the same column indicate significant differences between groups (p-value<0.05), according to Tukey's
 test.

	R	G	В	I	H	S	Ι	G/R
R^2	0.49	9 0.8	2 0.	46 0	.83	0.48	0.17	0.83
	L	а	b	CI	a/b	a/L	h	С
\mathbf{R}^2	0.79	0.78	0.78	0.80	0.83	0.83	0.81	0.69

								0.09	_
							minosity	, a Hunte	er a value,
b Hunter b vo	alue, CI colou	er index, h	Hunter	hue, C I	<i>Hunter cl</i>	hroma			
Table 5.	Coefficient	of deter	rminati	on for	the firm	nness a	and the	differe	nt colour
	co	mponen	ts meas	ured w	vith the	colori	meter		_
	L	а	b	CI	a/b	a/L	h	С	_
	R^2 0.66	5 0.78	0.63	0.77	0.78	0.78	0.77	0.76	
L Hunter lun chroma	ninosity, a Hi	inter a va	ılue, b H	lunter b	value, (CI coloi	ır index,	, h Hunt	er hue, C Hunter
Table 6. Sol	uble tannin	s conten	ıt (%) iı	n 'Rojo	o Brilla	nte' pe	ersimm	on fruit	ts before and
	aft	er treatr	nents ir	n the th	nree rip	eness s	stages		
	Group		M1		M2		M	3	
	HA	0.61	$a^{a} \pm 0.0$	9 0.	$65^a \pm 0$.06	$0.63^{a} \pm$	0.07	
	LA	0.45	$5^{b} \pm 0.04$	4 0.	$43^{b} \pm 0$.10	$0.39^{b} \pm$	0.06	
	NA	0.03	$r^{c} \perp 0.00$	0	$0.3^{\circ} \pm 0$	00	0.02°	0.00	
	INA	0.0.	± 0.00	J U.	03 ± 0	.00	$0.03 \pm$	0.00	
	b Hunter b vo Table 5. L Hunter lun chroma	R red, G green, B blue, H b Hunter b value, CI colour Table 5. Coefficient $\frac{co}{L}$ R ² 0.66 L Hunter luminosity, a Hu chroma Table 6. Soluble tannin $\frac{aft}{Group}$ HA LA	R red, G green, B blue, H hue, S sat b Hunter b value, CI colour index, h Table 5. Coefficient of deter component L a R^2 0.66 0.78 L Hunter luminosity, a Hunter a val chroma Table 6. Soluble tannins content after treatr Group HA 0.61 LA 0.45	R red, G green, B blue, H hue, S saturation, b Hunter b value, CI colour index, h HunterTable 5. Coefficient of determination components meas L a b R^2 0.66 0.78 0.63 L Hunter luminosity, a Hunter a value, b H chromaTable 6. Soluble tannins content (%) in after treatments in \overline{Group} M1HA $0.61^a \pm 0.09$ LA $0.45^b \pm 0.04$	R red, G green, B blue, H hue, S saturation, I intens b Hunter b value, CI colour index, h Hunter hue, CITable 5. Coefficient of determination for components measured w L a b CIR ² 0.66 0.78 0.63 0.77L Hunter luminosity, a Hunter a value, b Hunter b chromaTable 6. Soluble tannins content (%) in 'Roja after treatments in the th $\overline{Group M1}$ HA0.61 ^a ± 0.09 0. LALA0.61 ^a ± 0.09 0.	R red, G green, B blue, H hue, S saturation, I intensity, L Hu b Hunter b value, CI colour index, h Hunter hue, C Hunter citTable 5. Coefficient of determination for the firm components measured with the L a b CI a/b R^2 0.66 0.78 0.63 0.77 0.78L Hunter luminosity, a Hunter a value, b Hunter b value, CchromaTable 6. Soluble tannins content (%) in 'Rojo Brilla after treatments in the three rip $Group M1 M2$ HA 0.61 ^a ± 0.09 0.65 ^a ± 0LA 0.45 ^b ± 0.04 0.43 ^b ± 0	R red, G green, B blue, H hue, S saturation, I intensity, L Hunter lue b Hunter b value, CI colour index, h Hunter hue, C Hunter chromaTable 5. Coefficient of determination for the firmness a components measured with the colori L a b CI a/b a/L R^2 0.66 0.78 0.63 0.77 0.78 0.78L Hunter luminosity, a Hunter a value, b Hunter b value, CI color chromaTable 6. Soluble tannins content (%) in 'Rojo Brillante' per after treatments in the three ripeness s Group M1 M2HA0.61 ^a ± 0.090.65 ^a ± 0.06LA0.43 ^b ± 0.10	b Hunter b value, CI colour index, h Hunter hue, C Hunter chromaTable 5. Coefficient of determination for the firmness and the components measured with the colorimeter L a b CI a/b a/L h R^2 0.66 0.78 0.63 0.77 0.78 0.78 0.77 L Hunter luminosity, a Hunter a value, b Hunter b value, CI colour index, chromaTable 6. Soluble tannins content (%) in 'Rojo Brillante' persimm after treatments in the three ripeness stagesGroupM1M2MHA $0.61^a \pm 0.09$ $0.65^a \pm 0.06$ $0.63^a \pm$ LA $0.43^b \pm 0.10$ $0.39^b \pm$	R red, G green, B blue, H hue, S saturation, I intensity, L Hunter luminosity, a Hunter b Hunter b value, CI colour index, h Hunter hue, C Hunter chroma Table 5. Coefficient of determination for the firmness and the differe components measured with the colorimeter L a b CI a/L h C R ² 0.66 0.78 0.63 0.77 0.78 0.78 0.77 0.76 L Hunter luminosity, a Hunter a value, b Hunter b value, CI colour index, h Hunter chroma Hunter a value, b Hunter b value, CI colour index, h Hunter chroma Table 6. Soluble tannins content (%) in 'Rojo Brillante' persimmon fruit after treatments in the three ripeness stages Group M1 M2 M3 HA 0.65 ^a ± 0.06 0.63 ^a ± 0.07

Values are the mean of three measures of soluble tannins content (%) \pm standard deviation. Different superscript letters in the same column indicate significant differences between groups (p-value<0.05), according to Tukey's test.

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Table 7. Astringency classification of test set by LDA, QDA and SVM								
	Class	Correct classification (%)						
	Class	LDA	QDA	SVM				
	HA	95.9 ± 0.0^{b}	99.3 ± 0.0^{a}	97.1 ± 0.8^{b}				
M1	LA	$92.4\pm4.0^{\rm a}$	$94.5\pm4.3^{\rm a}$	$94.9\pm2.2^{\rm a}$				
111	NA	$93.9\pm2.6^{\rm a}$	$97.3\pm1.5^{\rm a}$	93.2 ± 2.1^{a}				
	Avg	$94.1\pm2.8^{\rm a}$	97.0 ± 3.1^{a}	95.1 ± 2.3^{a}				
	HA	$93.2\pm2.5^{\rm a}$	$96.2 \pm 2.1^{\mathrm{a}}$	$92.7\pm1.7^{\rm a}$				
M2	LA	$93.0\pm3.4^{\rm a}$	$95.3\pm2.2^{\rm a}$	$91.1\pm3.9^{\rm a}$				
IVIZ	NA	93.9 ± 1.1^{a}	$95.6\pm2.6^{\rm a}$	$95.9\pm1.5^{\rm a}$				
	Avg	$93.4\pm2.2^{\rm a}$	95.7 ± 2.1^{a}	93.2 ± 3.1^{a}				
	HA	$83.7\pm0.7^{\text{b}}$	94.3 ± 1.6^{a}	$90.0\pm3.4^{\rm a}$				
M3	LA	72.0 ± 3.4^{b}	$86.0\pm3.4^{\rm a}$	$64.5\pm2.2^{\rm b}$				
IVIS	NA	$93.4\pm2.2^{\rm a}$	$97.3\pm1.5^{\rm a}$	93.4 ± 0.7^{a}				
	Avg	$83.0\pm9.5^{\rm a}$	$92.5\pm5.5^{\rm a}$	82.7 ± 13.8^{a}				
Overall classification (%)		90.2 ± 7.6^{b}	95.1 ± 4.1^{a}	90.3 ± 9.7^{b}				

Table 7. Astringency classification of test set by LDA, QDA and SVM

Values are the mean of three models \pm standard deviation. Different superscript letters in the same row indicate significant differences between groups (p-value<0.05), according to Tukey's test.

542 543 544 545	Captions of the figures
546 547	Figure 1. Persimmon deastringed using a CO ₂ treatment (left) and persimmon naturally
548	deastringed by overripeness (right). The first shows firm and crisp flesh while the
549	second present a very soft texture.
550	
551	Figure 2. Images of persimmon at maturity stage M1, M2 and M3 from left to right.
552	
553	Figure 3. Hyperspectral acquisition system
554	
555	Figure 4. Images of a persimmon (M1) with selected ROI: (a) colour image;
556	hyperspectral image with (b) the VIS filter centred in 640 nm; and with (c) the NIR
557	filter centred in 900 nm
558	
559	Figure 5. Average spectra of control fruits in three ripeness stages M1 (long dashed
560	line), M2 (medium dashed line) and M3 (short dashed line)
561	
562	Figure 6. PC Loadings of the PC1 (solid line) and PC2 (dashed line) showing the
563	selected wavelengths for ripeness classification of 'Rojo Brillante' persimmon fruits.
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566	