

CONJUGATED LINOLEIC ACID CONTENT IN CECOTROPHES, SUPRARENAL AND INTRAMUSCULAR FAT IN RABBITS FED COMMERCIAL DIETS

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Abstract: An experiment was conducted to measure recycling through cecotrophes and retention of conjugated linoleic acid (CLA) in the body fat of rabbits. A commercial diet was formulated and given *ad libitum* to six New Zealand × Californian rabbits. Animals were weaned at 25 days of age and were reared in individual cages for five weeks, reaching an average body weight of 2.5 kg. Excretion of soft faeces was individually determined for 24 h after putting a wooden collar around the animals' neck. Ether extract content and fatty acid profile were determined in both soft and hard faeces, and also in the suprarenal and intramuscular fat. CLA cis-9, trans-11 isomer was detected in faeces and tissues. The CLA concentration was higher in soft than in the hard faeces (6.4 vs. 3.6 g/kg total fatty acids). No difference in CLA concentration was found between tissues (muscle vs. adipose). However, CLA incorporation in muscle was only detected in the neutral lipid fraction. It was concluded that the amount of CLA cis-9, trans-11 isomer recycled through cecotrophy in rabbits fed a commercial diet is retained in similar proportions in the suprarenal and intramuscular fat, but differs in the lipid classes.

Key words: cecotrophes, fat, dietary fibre, conjugated linoleic acid

INTRODUCTION

Conjugated linoleic acid (CLA) is a mixture of linoleic acid isomers that is produced in biohydrogenation processes, such as those occurring in the rumen. CLA is considered to have health benefits (anticarcinogenic, antiatherogenic, antioxidant and antidiabetic) at relatively low doses in laboratory animals and *in vitro* human models (Parodi, 1999; Williams, 2000; Pariza *et al.*, 2001; Azain, 2003). However, average CLA consumption in human diets (Ritzenthaler *et al.*, 2001) is below the recommended intake (Williams, 2000). As a result, there has been considerable interest in increasing CLA concentrations in animal feeds in order to provide healthful products for human consumption. Previous works (Corino *et al.*, 2002 and 2003) have shown that CLA supplementation of rabbit diets led to significant effects on lipid metabolism that improved carcass quality.

Concentrations of CLA in ruminant milk and meat fats is appreciable (from 3 to 6 g/kg; Chin *et al.*, 1992) and can be increased (up to 22 g/kg) through the use of appropriate dietary supplementation (Dhiman *et al.*, 1999). In contrast, CLA levels in products obtained from single-stomached animals such as pigs and poultry are much lower (Azain, 2003; Alvarez *et al.*, 2004). This difference is related to the lower amount of CLA precursors reaching the fermentative area and to the low capacity for long-chain fatty acids absorption in the hindgut. However, rabbits are able to recycle part of the end

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microbial fermentation products through cecotrophy, so that the CLA retained in their meat might be higher than in other non ruminant species.

The objective of this study was to measure the amount of CLA recycled through the soft faeces, and also the concentration of CLA in hard faeces, suprarenal and intramuscular fat of rabbits fed a commercial diet.

MATERIAL AND METHODS

A diet was formulated to meet nutrient recommendations from De Blas and Mateos (1998) and De Blas *et al.* (1999) by using typical ingredients from rabbit diets in Spain. The ingredients and chemical composition are shown in Table 1. Six New Zealand × Californian rabbits were weaned at 25 days of age and fed *ad libitum* the experimental diet in individual cages. Five weeks later, soft faeces excretion was individually determined throughout a 24 h period, after fitting the animals with a wooden collar (25-cm diameter) on the neck to prevent cecotrophy. The collar was put on at 8 am, before the usual

Table 1: Ingredient and chemical composition of the diet (g/kg as fed basis).

<i>Ingredients</i>	Barley grain	323
	Wheat bran	84
	Sunflower meal, 36%	71
	Soybean meal, 47%	111
	Alfalfa hay	288
	Sunflower hulls	60
	Beet pulp	23
	Lard	23
	L-Lysine HCl, 78% pure	4
	DL-methionine, 99% pure	1
	L-threonine	1
	Sodium chloride	6
	Vitamin-mineral premix ¹	5
<i>Chemical analysis</i>	Digestible Energy (MJ/kg) ²	11.3
	Crude protein	180
	Lysine ²	10.6
	Methionine ²	3.7
	Threonine ²	7.1
	Ether extract	50
	Starch	190
	Neutral detergent fibre	301
	Acid detergent fibre	164
	Acid detergent lignin	42
	Calcium ²	6.2
	Phosphorus ²	4.3

¹ Provided by Trouw Nutrition España, S.A. (Madrid, Spain). Mineral and vitamin composition (per kg of food): vitamin A, 8,375 IU/kg; vitamin D3, 750 IU/kg; vitamin E, 20 IU/kg; vitamin K3, 1 mg/kg; vitamin B2, 2 mg/kg; vitamin B6, 1 mg/kg; vitamin B1, 1 mg/kg; vitamin B3, 20 mg/kg; Fe: (FeSO₄), 76 mg/kg; Zn: (ZnO), 59 mg/kg; Mn: (MnO), 20 mg/kg; Cu: (CuSO₄), 10 mg/kg; Co: (CoCO₃), 0.7 mg/kg; I: (KI), 1.3 mg/kg; Choline, 250 mg/kg; S, 275 mg/kg; BHA-etoxiquin, 54 mg/kg; Flavofosfolipol, 2.5 mg/kg; Robenidine, 60 mg/kg.

² Calculated according to FEDNA (2003).

time rabbits begin to practise cecotrophy (Carabaño *et al.*, 1998). One day after finishing the cecotrophy trial, animals (weighing 2.44 ± 0.03 kg, SE) were slaughtered by cervical dislocation at 12 am. Samples of suprarenal fat and loin were taken for analysis.

Procedures of the AOAC (1995) were used to determine dietary contents of dry matter (930.15), N (954.01), ether extract, and starch according to the alpha-amyloglucosidase method (996.11). Neutral-detergent fibre, acid-detergent fibre and acid-detergent lignin were determined according to the sequential method of Van Soest *et al.* (1991). Fatty acids of diets were extracted and quantified by the one-step procedure described by Sukhija and Palmquist (1988) in freeze-dried samples with pentadecanoic acid (C15:0) (Sigma, Alcobendas, Madrid) as internal standard. Ether extract content in faeces was determined using Soxhlet extraction with diethyl ether. Intramuscular neutral and polar lipids were extracted by consecutive solvent elution with dichloromethane/methanol (90/10 v/v) according to the method developed by Marmer and Maxwell (1981). Fatty acid methyl esters (FAMEs) from muscle and faeces total lipids were prepared by acidic-trans-esterification in the presence of sulphuric acid (5% sulphuric acid in methanol) as described elsewhere (López-Bote *et al.*, 1997). All FAMEs were then analysed using a Hewlett Packard HP-6890 gas chromatograph equipped with flame ionisation detection and a $30\text{ m} \times 0.32\text{ mm} \times 0.25\text{ mm}$ cross linked polyethylene glycol capillary column (HP-Innowax). Analyses were performed with a temperature program from 170 to 240°C at a rate of $1^\circ\text{C}/\text{min}$. Injector and FID detector were maintained at 250°C . Carrier gas was helium at a flow rate of 3 ml/min. Individual methyl esters were identified by comparison with retention times of the corresponding standards (Qualimix Fich S. Ref: 89-5550, LARODAN, Malmö, Sweden).

General linear model procedures of SAS version 6 (Statistical Analysis System Inst., 1990) were used to test, by non-orthogonal contrasts, differences between fatty acid profile of fat extracted from i) hard vs. soft feces, ii) suprarenal vs. intramuscular fat, and iii) polar vs. neutral intramuscular lipids.

RESULTS AND DISCUSSION

Feed intake during the three days before cecotrophy control averaged $149 (\pm 3.0, \text{SE})$ g dry matter per day. Daily dry matter and ether extract recycling with cecotrophes were $17.7 (\pm 0.39)$ and $0.441 (\pm 0.101)$ g, which accounted for 10.7 and 5.0% of total intake (feed + cecotrophes), respectively.

Cis-9, trans-11 CLA isomer was detected in both soft and hard faeces (Table 2), the concentration being significantly higher in the former (6.4 vs. 3.6 g/kg total fatty acids). Previous works (García *et al.*, 1995, 2000) have shown that microbial nitrogen content in hard faeces was about half that in soft faeces, which indicates that a significant proportion of microorganisms are not recycled with cecotrophes. Also, ether extract concentration of total odd-numbered (C15:0 and C17:0) fatty acids, which are characteristics of microbial fat (Bauchart *et al.*, 1990) were higher (more than twice) in soft than in hard faeces (Table 2).

Ether extract recycled with cecotrophes might be higher when increasing the amount of energy substrate reaching the fermentative area, as observed by Gómez-Conde *et al.* (2004). Accordingly, CLA recycling could hypothetically be increased by using diets supplemented with highly digestible fibre.

CLA recycled with soft faeces was retained in a similar proportion (around 0.5 g/kg total fatty acids) both in suprarenal and total intramuscular fat. These values were close to those reported recently (Lo Fiego *et al.*, 2005) for cis-9, trans-11 concentration (0.4 g/kg) in *longissimus dorsi* samples from rabbits fed control diets with no CLA added. They were higher than those observed in meat from pigs (Azain, 2003) or eggs from poultry (Alvarez *et al.*, 2004), but lower than those found in meat or milk from ruminants (Chin *et al.*, 1992).

Table 2: Fatty acid profile (g/100 g fatty acids) of the diet (mean), soft and hard faeces, suprarenal and intramuscular fats (mean \pm SE).

Fatty acids	Diet	Hard faeces	Soft faeces	Suprarenal fatty acids	Intramuscular fatty acids			<i>P</i> -value Contrasts ^a
					Neutral	Polar	1 2 3	
C _{14:0}	0.80	1.21 \pm 0.06	1.85 \pm 0.13	2.60 \pm 0.06	2.84 \pm 0.09	0.90 \pm 0.09	0.001	0.02
C _{16:0}	20.9	25.7 \pm 0.18	23.5 \pm 0.38	27.8 \pm 0.14	28.2 \pm 0.51	25.2 \pm 0.22	0.001	0.29
C _{16:1 n-7}	1.40	1.12 \pm 0.03	0.86 \pm 0.04	5.03 \pm 0.31	8.47 \pm 0.72	3.10 \pm 0.27	0.66	0.02
C _{18:0}	8.20	16.1 \pm 1.57	17.9 \pm 0.78	6.71 \pm 0.20	6.01 \pm 0.27	6.32 \pm 0.09	0.08	0.60
C _{18:1 n-7}	2.01	5.01 \pm 1.16	4.88 \pm 0.58	1.93 \pm 0.04	2.02 \pm 0.07	2.57 \pm 0.10	0.85	0.71
C _{18:1 n-9}	28.9	24.4 \pm 0.38	17.6 \pm 0.50	32.9 \pm 0.41	33.5 \pm 0.67	23.9 \pm 0.71	0.001	0.013
C _{18:2 n-6}	31.9	14.8 \pm 1.78	12.5 \pm 0.60	18.4 \pm 0.59	15.3 \pm 0.62	20.8 \pm 0.79	0.06	0.29
C _{18:3 n-3}	3.50	1.61 \pm 0.23	1.06 \pm 0.09	1.72 \pm 0.05	1.40 \pm 0.05	0.56 \pm 0.04	0.001	0.001
C _{20:1 n-9}	ND ^b	ND	ND	ND	0.47 \pm 0.011	0.26 \pm 0.016	-	-
C _{20:3 n-9}	ND	ND	ND	ND	0.28 \pm 0.009	0.31 \pm 0.014	-	-
C _{20:4 n-6}	ND	ND	ND	ND	0.30 \pm 0.030	6.39 \pm 0.36	-	-
C _{22:6 n-3}	ND	ND	ND	ND	ND	0.82 \pm 0.09	-	-
Total odd-numbered	ND	1.41 \pm 0.04	3.31 \pm 0.20	0.98 \pm 0.005	0.87 \pm 0.04	0.69 \pm 0.02	0.001	0.47
cis-9,trans-11 CLA ^c	ND	0.36 \pm 0.07	0.64 \pm 0.05	0.053 \pm 0.003	0.082 \pm 0.006	ND	0.001	0.98
Total NSFA ^d	30.5	44.4 \pm 1.79	46.6 \pm 0.98	38.1 \pm 0.15	37.9 \pm 0.61	33.1 \pm 0.28	0.12	0.20
Total BSFA ^e	ND	3.14 \pm 0.02	8.72 \pm 0.88	0.39 \pm 0.03	ND	ND	0.002	-
Total MUFA ^f	34.1	31.4 \pm 1.15	24.0 \pm 0.92	40.8 \pm 0.71	44.4 \pm 1.04	29.8 \pm 0.85	0.001	0.40
Total PUFA ^g	35.4	16.6 \pm 2.01	14.4 \pm 0.67	20.5 \pm 0.64	17.3 \pm 0.64	28.8 \pm 0.82	0.10	0.56

^aContrast: 1 = hard vs. soft faeces; 2 = suprarenal vs. weighted average intramuscular fat; 3 = polar vs. neutral intramuscular fat. ^bND = not detected. ^cConjugated linoleic acid. ^dNormal saturated fatty acids. ^eBranched saturated fatty acids. ^fMonounsaturated fatty acids. ^gPolyunsaturated fatty acids.

Incorporation of CLA in muscle was only detected in the neutral lipid fraction. This may suggest that the amount of CLA cis-9, trans-11 concentration found in the soft faeces is not high enough to be detected in the intramuscular polar lipid fraction. Demaree *et al.* (2002) were unable to find CLA in the muscle polar lipid fraction of pigs fed corn oil or tallow, although feeding the same diets supplemented with a mixture of CLA isomers resulted in higher amounts of CLA in the polar than in the neutral lipid fraction of longissimus muscle.

From the results of this study it can be concluded that soft faeces, and hence meat from rabbits fed control diets with no CLA added, contain detectable amounts of cis-9 trans-11 CLA. Further work is necessary to establish the possibility of improving CLA concentration in rabbit meat through increasing caecal microbial growth.

REFERENCES

- Álvarez C., Cachaldora P., Méndez J., García-Rebollar P. 2004. Effects of conjugated linoleic acid addition on its deposition in eggs of laying hens, fed with no other fat source. *Span. J. Agric. Res.*, 2, 203-209.
- AOAC. 1995. Official Methods of Analysis (16th edn.). Association of Official Analytical Chemists, Washington, DC.
- Azain M.J. 2003. Conjugated linoleic acid and its effects on animal products and health in single-stomached animals. *P. Nutr. Soc.*, 62, 319-328.
- Bauchart D., Legay-Carmier F., Doreau M., Gaillard, B. 1990. Lipid metabolism of liquid-associated and solid-adherent bacteria in rumen contents of dairy cows offered lipid-supplemented diets. *Brit. J. Nutr.*, 63, 563-578.
- Carabaño R., Fraga M.J., Santoma G., De Blas J.C. 1988. Effect of diet on composition of cecal contents and on excretion and composition of soft and hard feces of rabbits. *J. Anim. Sci.*, 66, 901-910.
- Chin S.F., Liu W., Storkson J.M., Ha Y.L., Pariza M.W. 1992. Dietary sources of conjugated linoleic acid, a newly recognised class of anticarcinogens. *J. Food Compos. Anal.*, 5, 185-197.
- Corino C., Mourot J., Magni S., Pastorelli G., Rosi F. 2002. Influence of dietary conjugated linoleic acid on growth, meat quality, lipogenesis, plasma leptin and physiological variables of lipid metabolism in rabbits. *J. Anim. Sci.*, 80, 1020-1028.
- Corino C., Filetti F., Gambacorta M., Manchisi A., Magni S., Pastorelli G., Rossi R., Maiorano G. 2003. Influence of dietary conjugated linoleic acids (CLA) and age at slaughtering on meat quality and intramuscular collagen in rabbits. *Meat Sci.*, 66, 97-103.
- Demaree S.R., Gilbert C.D., Mersmann H.J., Smith S.B. 2002. Conjugated linoleic acid differentially modifies fatty acid composition in subcellular fractions of muscle and adipose tissue but not adiposity of postweanling pigs. *J. Nutr.*, 132, 3272-3279.
- De Blas J.C., Mateos G.G. 1998. Feed formulation. in: *De Blas J.C., Wiseman J. (eds.). The Nutrition of the Rabbit. Commonwealth Agricultural Bureau, Wallingford, UK*, 241-253.
- De Blas J.C., Gutiérrez I., Carabaño R. 1999. Destete precoz en gazapos, situación actual y perspectivas. in: *García-Rebollar P., De Blas J.C., Mateos G.G. (eds.). Avances en Nutrición y Alimentación Animal. FEDNA, Madrid, Spain*, 67-81.
- Dhiman T.R., Anand G.R., Satter L.D., Pariza M.W. 1999. Conjugated linoleic acid content of milk from cows fed different diets. *J. Dairy Sci.*, 82, 2146-2156.
- FEDNA (2003). Normas FEDNA para la formulación de piensos compuestos, in: *De Blas J.C., Mateos G.G., García-Rebollar P., (Eds) (Madrid, Fundación Española para el Desarrollo de la Nutrición Animal)*.
- García J., De Blas J.C., Carabaño R., García-Rebollar P. 1995. Effect of type of lucerne hay on caecal fermentation and nitrogen contribution through caecotrophy in rabbits. *Reprod. Nutr. Dev.*, 35, 267-275.
- García J., Carabaño R., Pérez-Alba L., De Blas J.C. 2000. Effect of fiber source on cecal fermentation and nitrogen recycled through caecotrophy in rabbits. *J. Anim. Sci.*, 78, 638-646.
- Gómez-Conde M.S., Chamorro S., Menoyo D., García-Rebollar P., De Blas J.C. 2004. Effects of source of fibre on fat composition and fat recycling with caecotrophes. In Proc.: *8th Worl Rabbit Congress, Puebla, Mexico*, 131.
- Lo Fiego D.P., Maccioni P., Santoro P., Rossi R., Pastorelli G., Corino C. 2005. Influence of conjugated linoleic acid (CLA) on intramuscular fatty acid composition in the rabbit. *Ital.J. Anim. Sci.*, 4, 553-555.
- López-Bote C.J., Rey A., Isabel B., Arias R.S. 1997. Dietary fat reduce odd-numbered and branched-chain fatty acids in depot lipids of rabbits. *J. Sci. Food Agr.*, 73, 517-524.
- Marmer W.N., Maxwell R.J. 1981. Dry column method for the quantitative extraction and simultaneous class separation of lipids from muscle tissue. *Lipids*, 16, 365-371.
- Pariza M.W., Park Y., Cook M.E. 2001. The biologically active isomers of conjugated linoleic acid. *Prog. Lipid Res.*, 40, 283-298.
- Parodi P.W. 1999. Conjugated linoleic acid and other anticarcinogenic agents of bovine milk fat. *J. Dairy Sci.*, 82, 1339-1349.
- Ritzenthaler K.L., McGuire M.K., Falen R., Shultz T.D., Dasgupta N., McGuire M.A. 2001. Estimation of conjugated linoleic acid intake by written dietary assessment methodologies underestimates actual intake evaluated by food duplicate methodology. *J. Nutr.*, 131, 1548-1554.
- SAS. 1990. SAS/STATÒ User's Guide, Version 6. 4th ed. SAS Inst. Inc., Cary NC, USA.
- Sukhija P.S., Palmquist D.L. 1988. Rapid method for determination of total fatty acid content and composition of feedstuffs and feces. *J. Agr. Food Chem.*, 36, 1202-1206.
- Van Soest J.P., Robertson J.B., Lewis B.A. 1991. Methods for dietary fiber, neutral detergent fiber and nonstarch polysaccharides in relation to animal nutrition. *J. Dairy Sci.*, 74, 3583-3597.
- Williams C.M. 2000. Dietary fatty acids and human health. *Ann. Zootech.* 49, 165-180.

