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Dr. Youling Xiong
Journal of Food Science

Dear Editor,

Please find enclosed a copy of the manuscript entitled “**Designing a clean label sponge cake with reduced fat content**” which we would like to be considered for publication in the **Journal of Food Science**.

In this study a natural functional ingredient derived from flaxseed, OptiSol™5300, is used as fat replacer. This ingredient is high in fiber and has hydrocolloid functionality. In this research, functional cakes with reduced fat content and good acceptability by consumers are obtained avoiding E-numbers on their labels.

We hope that the paper will be suitable for publication.

Thank you in advance,

Yours sincerely

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1 **Designing a Clean Label Sponge Cake with Reduced Fat Content.**

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3

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10 **5,060 words**

11 Low fat cake with functional ingredient...

12 Food Chemistry

13

14

15 **ABSTRACT**

16 The fat in a sponge cake formulation was partially replaced (0, 30, 50, 70%) with
17 OptiSol™5300. This natural functional ingredient derived from flax seeds, rich in fiber
18 and alpha-linoleic acid, provides a natural substitute for guar and xanthan gums,
19 avoiding E-numbers on labels. The structure and some physicochemical properties
20 of the formulations were examined, sensory analysis was conducted and changes in
21 starch digestibility due to adding this ingredient were determined. Increasing
22 quantities of OptiSol™5300 gave harder cakes, with less weight loss during baking,
23 without affecting the final cake height. There were no significant differences ($p >$
24 0.05) in texture, flavor and overall acceptance between the control and the 30%
25 substitution cake, nor in the rapidly digestible starch values. Consequently, replacing
26 up to 30% of the fat with OptiSol™5300 gives a new product with health benefits
27 and a clean label that resembles the full-fat sponge cake.

28
29 **Keywords:** cake; flaxseed; fat replacer; structure; clean-label.

30
31 **Practical Application:** Sponge cakes with a functional flaxseed ingredient as fat
32 replacer were obtained in this work. This ingredient, like oil, is distributed uniformly
33 through the batter matrix. Cakes with 30% fat replacement were very well accepted
34 by consumers and could be prepared and sold by food industry as clean-label
35 reduced-fat cakes.

36

37 1. INTRODUCTION

38 Bakery products are very high consumption foods throughout the world. Sponge
39 cakes are particularly popular and consumers consider them delicious products with
40 certain particular organoleptic characteristics (Matsakidou and others 2010). Cakes
41 contain approximately 15-25% fat (Matsakidou and others 2010). Fat influences the
42 sensory and texture characteristics of the food product and, consequently, its
43 acceptability to consumers. In sponge cakes, fat not only provides flavor but also
44 makes it easier to incorporate air into the batter, contributing to its increase in
45 volume, and interferes with the continuity of the gluten structure, favoring the
46 formation of a softer cake (Psimouli and Oreopoulou 2013).

47 In developed countries, food-related diseases are increasing because, among
48 other factors, energy ingestion is too high and the consumption of dietary fiber is
49 below the recommended levels (Zahn and others 2010). Fat is the food constituent
50 with the highest energy value (Zahn and others 2010). Excessive fat consumption is
51 associated with obesity, cardiovascular problems and different types of cancer
52 (Román and others 2015). A promising way for the food industry to supply healthier
53 foods could therefore be to replace fat with dietary fiber in food formulations (Zahn
54 and others 2010). Various studies have been published on reducing the fat content
55 of cakes and muffins by adding different carbohydrates and fibers, such as inulin
56 (Zahn and others 2010; Rodríguez-García and others 2012; Psimouli and
57 Oreopoulou 2013; Rodríguez-García and others 2014b), β -glucan (Lee and others
58 2005; Kalinga and Mishra 2009), oat bran and flaxseed flour (Lee and others 2004),
59 cocoa fiber (Martínez-Cervera and others 2011), polydextrose (Kocer and others
60 2007), maltodextrin (Lakshminarayan and others 2006; Psimouli and Oreopoulou
61 2013) and citrus pectin (Psimouli and Oreopoulou 2013). Few references to fat
62 replacement by gums appear to have been published. Zambrano and others (2004)

63 used guar and xanthan gums to obtain low-fat sponge cakes, Khouryieh and others
64 (2005) prepared muffins in which they replaced fat with a mixture of maltodextrin
65 and xanthan gum and Kaur and others (2000) used guar gum and carboxymethyl
66 cellulose to reduce the fat content of muffins.

67 The importance of gums lies in their ability to control the rheological
68 characteristics of an aqueous system. They can also influence the stabilization of
69 emulsions and help to suspend particles, control crystallization and inhibit syneresis
70 (Zambrano and others 2004).

71 In the present study, a functional ingredient with hydrocolloid capacity derived
72 from flax seeds was used as a fat substitute. This ingredient can be used to replace
73 guar and xanthan gums and, being a natural product, allows clean labeling of the
74 food, avoiding E-numbers.

75 Flax seeds are a potential source for functional food formulations (Marpalle and
76 others 2014). They are rich in health beneficial nutrients (Shearer and Davies 2005)
77 and present a unique nutritional profile with a high concentration of polyunsaturated
78 fatty acids (73% of the total fatty acids), a moderate concentration of
79 monounsaturated fatty acids (18%) and a low concentration of saturated fatty acids
80 (9%). Linoleic acid comprises 16% and alpha-linolenic acid (ALA) 57% of their total
81 fatty acids (Marpalle and others 2014). ALA is an omega-3 fatty acid which is
82 involved in reducing inflammatory processes and plays an important role in
83 reducing/preventing cardiovascular diseases, osteoporosis, diabetes and digestive
84 system diseases (Mercier and others 2014). Flax seeds contain approximately 28%
85 fiber, of which a third is soluble. Soluble fiber is associated with the ability to reduce
86 cholesterol and regulate blood sugar levels. The rest of the dietary fiber is insoluble.
87 Insoluble fiber favors increased stool bulk and reduces bowel transit time, so it helps
88 to prevent constipation and may provide protection against colon cancer (Moraes
89 and others 2010).

90 The aim of this study was to assess the effect on the structure and the
91 physicochemical and sensory properties of sponge cakes of using a natural
92 functional ingredient derived from flaxseed to replace part of the fat in their
93 formulation. A further aim was to study changes in starch digestibility as a result of
94 adding this ingredient to the sponge cake formulation.

95

96 **2. MATERIALS AND METHODS**

97 **2.1 Ingredients**

98 The ingredients used in the preparation of the cake batters were: wheat flour
99 (Harinas Segura S.L, Torrente, Spain, composition provided by the supplier: 13.50-
100 15.50 g/100 g moisture, 9-11 g/100 g proteins); sugar (AB Azucarera Iberica S.L.U.,
101 Madrid, Spain); liquid pasteurized egg white and yolk (Ovocity, Llombay, Spain);
102 skim milk (Corporación Alimentaria Peñasanta, S.A., Siero, Spain); refined
103 sunflower oil (Aceites del Sur-Coosur, S.A., Vilches, Spain); OptiSol™5300 (natural
104 ingredient derived from flaxseed; Glambia Nutritionals Ltd, Kilkenny, Ireland;
105 specifications provided by the supplier: 28-36 g/100 g protein, 8-12 g/100 g
106 moisture, 15-20 g/100 g fat, 30-40 g/100 g carbohydrate, 25-37 g/100 g dietary fiber)
107 sodium bicarbonate and citric acid (Sodas y Gaseosas A. Martínez, S.L., Cheste,
108 Spain); salt and distilled water. **Oil was used instead of shortening as usually done
109 in Mediterranean countries (Rodriguez-Garcia and others, 2012; Martinez-Cervera
110 and others 2011)**

111

112 **2.2 Batter and cake preparation**

113 Four formulations were prepared: the control formulation (R0) and three
114 formulations (R30, R50 and R70) where fat was replaced at 30%, 50% and 70%
115 respectively, by increasing amounts of OptiSol™5300 (Table 1). The fat content of
116 OptiSol™5300 was taken into account to calculate the level of fat replacement in the

5

117 different formulations. In order to achieve the proper dispersion for OptiSol™5300 to
118 act as a fat mimetic, the appropriate amounts of water were added for an
119 OptiSol™5300-to-water ratio of 1:5.

120 The batters were prepared according to the all-in mixing procedure proposed by
121 Rodríguez-García and others (2014a), with some modifications. The liquid eggs,
122 milk and water were placed in a Kenwood Major Classic mixer (Havant, England,
123 UK). The dry ingredients (wheat flour, sugar, OptiSol™5300, sodium bicarbonate
124 and citric acid and salt) were added to the liquids, and the oil was finally placed on
125 the top. The mixing proceeded using a wire whisk at speed 1 for 30 sec, followed by
126 1 min at speed 2 and 3 min at speed 3.

127 For cake preparation, a conventional oven (Electrolux, EOC3430DOX,
128 Stockholm, Sweden) was preheated to 180 °C for 30 min, and the batters (700g)
129 were placed in a Pyrex baking pan (diameter = 20 cm) and baked at 180 °C for 43
130 min. The cakes were then kept at room temperature for at least 1 h 30 min before
131 being analyzed. All the batters and cakes were prepared in triplicate and the
132 analyses were performed within 24 hours of their preparation.

133

134 2.3 Confocal laser scanning microscopy (CLSM)

135 **Equipment and dyes.** A Nikon confocal microscope C1 unit fitted on a Nikon
136 Eclipse E800 microscope (Nikon, Tokyo, Japan) was used. An Ar laser line (488nm)
137 was employed as the light source to excite the fluorescent dyes: Rhodamine B and
138 Nile Red. Rhodamine B (Fluka, Sigma-Aldrich, St. Louis, Mo., U.S.A), with $\lambda_{ex\ max}$ 488
139 nm and $\lambda_{em\ max}$ 580 nm, was solubilized in distilled water at 0.2 g/100 mL. This dye
140 was used to stain proteins and carbohydrates. Nile Red (Fluka, Sigma-Aldrich), with
141 $\lambda_{ex\ max}$ 488 nm and $\lambda_{em\ max}$ 515 nm, was solubilized in PEG 200 at 0.1 g/L and used to
142 stain fat. The objective lenses were 40x/1.0 and 60x/1.40 NA/Oil/ Plan Apo VC
143 Nikon.

144 **Sample viewing.** A microscopy slide was prepared with 2 razor blades stuck to
145 the glass (Alava and others 1999; Sahi and Alava 2003). A drop of batter was
146 placed on the slide, in the central gap between the blades. Rhodamine B solution
147 and Nile Red solution were added and the cover slide was carefully positioned to
148 exclude air pockets. The images were observed and stored with a 1024 × 1024 pixel
149 resolution using the microscope software (EZ-C1 v.3.40, Nikon).

150

151 **2.4 Weight loss during baking**

152 The weight loss during baking (WL) was calculated, taking into account the initial
153 water content of each formulation (Rodríguez-García and others 2014b). The weight
154 loss during baking was calculated as follows:

155

$$156 \text{ WL (g/100 g) = } [(B-C/IW) \times 100]$$

157

158 where:

159 WL= weight loss during baking

160 B = weight (in grams) of batter before baking.

161 C = weight (in grams) of cake after baking

162 IW = initial water content (in grams).

163

164 The initial water content was the sum of the initial water content of each
165 ingredient in the formulation being measured. Measurements were performed in
166 triplicate.

167

168 **2.5 Cake height**

169 The maximum cake height was measured on the vertical cross section of the
170 product, using *ImageJ* software (National Institutes of Health, Bethesda, MD, USA).

7

171 The baked product was cut and photographed with a digital camera (E-510
172 Olympus, Hamburg, Germany). The images were stored in a 3648 × 2736 pixel
173 format. The measurements were performed in triplicate.

174

175 **2.6 Macroscopic structure of the crumb**

176 The cakes were cut vertically in the central area and scanned using a HP
177 Scanjet G2710 scanner (Hewlett-Packard, Palo Alto, CA, USA). The scanned
178 images were analyzed using ImageJ software (National Institutes of Health,
179 Bethesda, MD, USA). The image was cropped to a 5 × 5 cm section, on which the
180 analysis was performed. Firstly, the image was split into color channels, then the
181 contrast was enhanced and finally the image was binarized after applying a
182 grayscale threshold. For each formulation, three different images were analyzed.

183

184 **2.7 Color measurements**

185 The instrumental measurements of the cake crust and crumb color were made
186 with a Chroma meter CR-400 (Konica Minolta Sensing Americas, Inc., Ramsey, NJ,
187 USA). The results were expressed in accordance with the CIELAB system, with
188 reference to illuminant C and a visual angle of 2°. The parameters determined were
189 L^* ($L^* = 0$ [black], $L^* = 100$ [white]), a^* ($-a^*$ = greenness, $+a^*$ = redness), b^* ($-b^*$ =
190 blueness, $+b^*$ = yellowness), C_{ab}^* (chroma [$C_{ab}^* = (a^{*2} + b^{*2})^{1/2}$]) and h_{ab} (hue [$h_{ab} =$
191 $\arctan (b^*/a^*)$]).

192 The total color difference (ΔE^*) was calculated as follows (Francis and
193 Clydesdale 1975):

194

$$195 \quad \Delta E^* = [(\Delta L^*)^2 + (\Delta a^*)^2 + (\Delta b^*)^2]^{1/2}$$

196

197 The values used to determine whether the total color difference was appreciable
198 by the human eye were (Baixauli and others 2008):

199

200 $\Delta E^* < 1$ color differences are not obvious to the human eye.

201 $1 < \Delta E^* < 3$ color differences are not appreciated by the human eye.

202 $\Delta E^* > 3$ color differences are obvious to the human eye.

203 The measurements were performed in quadruplicate.

204

205 **2.8 Cake texture**

206 Texture profile analysis (TPA) was carried out using a TA-TXT plus Texture
207 Analyzer (Stable Microsystems, Godalming, U.K.) with Texture Exponent Lite 32
208 software (version 6.1.4.0, Stable Microsystems). The test was performed on 6 cubes
209 (3 cm each side) taken from the central crumb of each cake. TPA was performed
210 under the conditions described by Rodríguez-García and others (2012). The
211 parameters studied were hardness, springiness, cohesiveness and chewiness.
212 Measurements were performed in duplicate.

213

214 **2.9 Sensory analysis**

215 140 consumers (aged 18 to 64) were recruited among employees and students
216 of the Universitat Politècnica de València. The samples were assessed in a
217 standardized tasting room equipped with individual booths. Each consumer received
218 4 pieces of cake (1 piece each of the control with no OptiSol™5300 and the 3 fat-
219 replacement cakes). 3-digit random numbers were used to code the pieces of
220 cakes. They were served at room temperature in random order. Water was supplied
221 to clean the consumers' mouths between each sample. Consumer acceptance
222 testing was performed with a successive category scale to score the "appearance",
223 "texture", "taste", and "overall acceptance" of the product. The scale was a 5-point

9

224 hedonic scale labeled 5 = like very much, 4 = like moderately, 3 = neither like nor
225 dislike, 2 = dislike moderately and 1 = dislike very much.

226

227 **2.10 Field Emission Scanning Electron Microscopy (FESEM)**

228 Samples of each formulation (0.5 cm each side) were frozen at -80°C and
229 lyophilized (Telstar, Lyoquest 55, Terrassa, Spain). The samples were coated with
230 platinum in a vacuum, then observed in a field emission scanning electron
231 microscope (FESEM) (Zeiss, Ultra 55, Oberkochen, Germany).

232

233 **2.11 *In vitro* digestion**

234 The simulation of intestinal digestion consisted of a jacketed glass reactor (1 L
235 capacity) maintained at 37 °C with continuous magnetic stirring at 120 rpm in a
236 temperature-controlled circulating water bath throughout the test.

237 The *in vitro* digestions were carried out according to the method of Sozer and others
238 (2014), with modifications. 4 g of ground sample, 100 mL 0.05 mol/L of sodium
239 potassium phosphate buffer (pH 6.9) and 5 mL of 2.5g/100 mL pancreatin (P3292,
240 pancreatin from porcine pancreas containing trypsin, amylase, lipase, ribonuclease
241 and protease, Sigma-Aldrich, St. Louis, Mo., U.S.A) in 0.1 mol/L of pH 6 maleate
242 buffer were placed in the jacketed glass reactor and maintained there for 120 min.
243 Aliquots (7.5 mL) were removed at 20, 60, 90 and 120 min, placed in boiling water
244 for 5 min, and cooled on ice. They were then centrifuged at 6600 rpm for 5 min and
245 the supernatant was analyzed for reducing sugar content using the dinitrosalicylic
246 acid (DNS) colorimetric method. The amount of digested starch was determined by
247 multiplying the reducing sugar values by a 0.9 stoichiometric conversion constant for
248 glucose to starch (Hardacre and others 2015). For each formulation, two different
249 sponge cakes were analyzed.

250

251

252 2.12 Statistical analysis

253 Analysis of variance (ANOVA) was performed on the data using the Statgraphics
254 Centurion XVI.I software package (StatPoint Technologies, Inc., Warrenton, VA,
255 USA). Fisher's least significant difference (LSD) test was used to evaluate mean
256 difference values ($p < 0.05$).

257

258 3. RESULTS AND DISCUSSION**259 3.1 Confocal laser scanning microscopy (CLSM) of the batters**

260 Figure 1 shows CLSM images of the sponge cake batters prepared with different
261 levels of fat substitution (R0, R30, R50 and R70).

262 In the control batter (R0), a continuous phase with a homogeneous appearance
263 can be seen, stained red with rhodamine. It is mainly formed by interaction between
264 the proteins of the sponge cake ingredients: milk, egg and gluten. Associated with
265 this protein network, intact starch granules in black and fat globules stained green
266 with Nile Red can also be identified. The fat globules are of different sizes, and
267 many are clustered together, but hardly any coalescence can be seen.

268 In the R30 sponge cake batters, the continuous phase seems to be formed by
269 interaction between the proteins of the different ingredients in the formulation (milk,
270 egg, gluten and OptiSol™5300), and probably also the fat phase of the functional
271 ingredient (OptiSol™5300). Unlike the sunflower oil, this fat phase does not appear
272 in the form of globules but interacts with the protein fraction of the formulation.
273 Another extensive dark or blackish network can be observed superimposed on the
274 continuous phase. It is probably composed of the carbohydrate components of the
275 OptiSol™5300 ingredient added to the formulation. The starch granules and fat
276 globules can be seen associated with this network. Coalescence between the
277 globules is slightly higher than in the control batter.

11

278 **At the highest substitution level (R70)**, the extent, compaction and intensity of
279 the dark/blackish network associated with the addition of OptiSol™5300 have
280 increased. Also, as the levels of fat replacement in the sponge batter rise, fewer fat
281 globules can be seen. Coalescence between the globules appears not to be
282 significant in the high-substitution batters. The decreased fat content of these
283 formulations appears to minimize coalescence.

284

285 **3.2 Weight loss during baking and cake height**

286 In general, adding OptiSol™5300 to the formulation led to significant low weight
287 loss during baking ($p < 0.05$) from 21.48 ± 0.51 g/100 g in the control (R0) to
288 20.09 ± 0.36 g/100 g in R70 (**Table 2**). Hydrocolloids have the ability to increase
289 moisture retention, although this depends on their chemical structure and interaction
290 with the other ingredients (Gomez and others 2007), and consequently to reduce
291 weight loss in the food. From this point of view, OptiSol™5300 presented
292 hydrocolloid behavior in the sponge cake. The present results agree with those
293 obtained by Khouryieh and others (2005), who observed lower moisture loss in
294 muffins prepared with xanthan gum and maltodextrin as fat replacers than in those
295 prepared with fat or with maltodextrin alone.

296 The addition of OptiSol™5300 to the formulation did not influence the maximum
297 height of the sponge cakes. No significant differences ($p > 0.05$) between
298 formulations were found in the values for this parameter (**Table 2**).

299

300 **3.3 Macroscopic structure of the crumb**

301 Figure 2 shows scanned, contrasted and binarized images of the different
302 sponge cakes (R0, R30, R50 and R70).

303 The crumb macrostructure of the control cake (R0) was practically homogenous
304 (Figure 2). In contrast, a series of diffusion pathways were observed in the crumb of

12

305 the sponge cakes formulated with OptiSol™5300. Furthermore, these pathways
306 appeared to a greater extent in the formulations with higher levels of substitution
307 (R50 and R70) than in the R30 sponge cake.

308

309 **3.4 Color measurement**

310 The color data (L^* , a^* , b^* , C_{ab}^* , h_{ab}^* , ΔE^*) for the crust and crumb of the different
311 sponge cakes studied are shown in Table 3.

312 In the crust, the values for parameters L^* , a^* , b^* and C_{ab}^* were significantly lower
313 ($p < 0.05$) in the control formulation (R0) than in the fat substitution formulations
314 studied (R30, R50 and R70), and no significant differences ($p > 0.05$) were
315 observed between the formulations with the different levels of substitution. Lee and
316 others (2004) also found significant differences in L^* for the crust of cakes in which
317 fat had been replaced by oat bran and powdered flaxseed: the greater the
318 replacement of fat by these ingredients, the higher the L^* values obtained, indicating
319 greater luminosity. As regards hue (h_{ab}^*), significant differences ($p < 0.05$) were
320 found between the control cake and the formulations with higher levels of
321 substitution. Lastly, differences in overall color ($\Delta E^* > 3$), visible to the human eye,
322 were found between the control cake and the other formulations. The reason for
323 these differences was that the surface color became less dark as more
324 OptiSol™5300 was included in the formulation. Shearer and Davies (2005) also
325 found overall color differences in muffins prepared with 5 g flaxseed flour/100g
326 batter, but not in those prepared with 2 g of this flour/100g batter.

327 No significant differences in the crumb of the four types of sponge cake studied
328 were found for L^* , b^* or C_{ab}^* (Table 3). However, the a^* value rose significantly ($p <$
329 0.05) as greater quantities of OptiSol™5300 were added to the formulation. Lee and
330 others (2004) also encountered higher a^* values on increasing the level of flaxseed
331 powder used to replace fat in cakes. In contrast, the h_{ab}^* value fell significantly as

13

332 more OptiSol™5300 was added, as shown in Table 3. Lastly, no significant
333 differences in overall color were encountered, as all the ΔE^* values were below 3.

334

335 **3.5 Sponge cake texture**

336 Table 4 shows the parameters obtained from the texture profile analysis (TPA)
337 curves.

338 The hardness values showed no significant differences ($p > 0.05$) between the
339 control cake and R30. However, the addition of OptiSol™5300 in the formulations
340 with higher fat replacement (R50 and R70) gave place to significant ($p < 0.05$)
341 higher hardness values. Zambrano and others (2004) obtained a significantly higher
342 firmness value in cake prepared with xanthan gum and 50% less fat than the
343 control. Khouryieh and others (2005) also found significantly higher hardness values
344 in muffins prepared with maltodextrin and xanthan gum to replace fat, when
345 compared with the control.

346 The springiness and cohesiveness values indicate that there were no significant
347 differences between the formulations studied. This could be because all the sponge
348 cakes contained the same flour and, therefore, the same proportion of gluten
349 proteins, which are responsible for the elasticity and strength of the batter.

350 The chewiness values followed a similar trend to those for hardness. No
351 significant differences ($p > 0.05$) were observed between control cake and R30. In
352 contrast significant differences were observed between R0 and cakes with higher
353 quantity of OptiSol™5300 in their formulations (R50 and R70). R70 was the
354 chewiest sponge cake – which could be related to a more compact cake – which is
355 why it was necessary to apply more force compared to the other cakes to ready it for
356 swallowing. Khouryieh and others (2005) also found significantly higher chewiness
357 values than those of the control in muffins prepared with less fat and the addition of
358 maltodextrin and xanthan gum.

359 **3.6 Sensory analysis**

360 The mean sensory acceptability values for the appearance, texture, taste and
361 overall acceptance of the control cake and the sponge cakes prepared with different
362 levels of fat replacement are shown in Figure 3.

363 The statistical analysis showed that there were no significant differences ($p >$
364 0.05) between the control cake (R0) and R30 for the attributes of texture, taste and
365 overall acceptance. In the case of taste, R0 and R50 did not exhibit any significant
366 differences either. Nevertheless, the appearance value was significantly lower ($p <$
367 0.05) for R30 than for the control.

368 For all the attributes studied, it was found that the sensory acceptability fell
369 significantly as the quantity of OptiSol™5300 in the formulation rose, since
370 significantly lower values were obtained on comparing R50 with R0 (except for the
371 attribute of taste, as commented above) and R70 with R0. Of all the fat replacement
372 formulations studied, R30 seemed to have the highest sensory acceptability.

373

374 **3.7 Field emission scanning electron microscopy (FESEM)**

375 Figure 4 shows the images obtained through field emission scanning electron
376 microscopy (FESEM), allowing observation of the sponge cake microstructure.

377 The structure of the control cake (R0) can be seen to be made up of a well-
378 developed hydrated protein network, mainly formed by the gluten from the flour, into
379 which the rest of the ingredients have been incorporated. The gluten network
380 contains the partially gelatinized starch granules and the oil has acted as a lubricant,
381 creating a flexible continuous structure. The fat globules coalesced during baking
382 and liberated fat, which was incorporated into the matrix, creating a film coating, as
383 can be seen in Figure 4.

384 The R30 cake shows a more irregular microstructure, as the oil coating is less
385 extensively distributed compared to R0. With the higher fat replacement levels (R50

15

386 and R70), it becomes more difficult it to find the oil coating in the images. It may also
387 be seen that as greater quantities of OptiSol™5300 were added to in the
388 formulation, this ingredient acted as a filler in the spaces within the protein network,
389 making the sponge cake more compact. This information was correlated with the
390 textural values, since the hardness and chewiness increased significantly, compared
391 with the R0 and R30 cakes, as the OptiSol™5300 content rose.

392

393 **3.8 *In vitro* digestion**

394 Table 5 shows the proportion of starch digested *in vitro* for each of the sponge
395 cakes studied.

396 The gelatinized starch fraction digested during the first 20 min of the digestion
397 process is considered rapidly digestible starch (RDS) (Hardacre and others 2015).
398 After subjecting the different cakes (R0, R30, R50 and R70) to 20 min *in vitro*
399 digestion, no significant differences ($p > 0.05$) in RDS values were found between
400 R0, R30 and R50 cakes; being RDS values in R70 significantly lower ($p < 0.05$). The
401 digestibility of starch values significantly decreased ($p < 0.05$) in cakes with higher
402 levels of substitution (R70) after 60, 90 and 120 min too. Adding fiber when
403 processing foods is known to limit starch gelatinisation and mask the starch
404 granules, increasing the proportion of resistant starch as observed by Hardacre et
405 al., (2015). This could be also related to the field emission scanning electron
406 microscope (FESEM) images, where the starch granules in the R70 cake matrix
407 were observed to be very compact within the matrix, which would limit their physical
408 accessibility to the digestive enzymes.

409

410 **4. CONCLUSIONS**

411 OptiSol™5300, a functional ingredient derived from flax seeds, can be used to
412 replace fat in sponge cakes when formulating low fat products; as a natural product,

16

413 it allows clean labeling avoiding E-numbers. When preparing fat reduced cakes
414 using this ingredient it is found that sponge cakes with 30% fat replacement present
415 good sensory acceptability to consumers. The hydrocolloid properties of
416 OptiSol™5300 avoid moisture loss without affecting batter rise, and the hardness
417 and crumb color of the cakes are similar than in control. Sponge cakes with 30% fat
418 replacement by this functional ingredient could be elaborated by food industry as an
419 appropriate strategy to reduce fat and calories in this bakery product.

420

421 **ACKNOWLEDGMENTS**

422 The authors are grateful to INIA for financial support through the *BERRYPOM-*
423 *Adding value to fruit processing waste: innovative ways to incorporate fibers from*
424 *berry pomace in baked and extruded cereal-based foods* project included in the
425 ERA-NET - SUSFOOD program.

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427 editing assistance, and to the Electron Microscopy Service of the Universitat
428 Politècnica de Valencia.

429

430 **AUTHOR CONTRIBUTIONS**

431 Cristina Eslava-Zomeño elaborated the cakes and carried out experimental work
432 on physicochemical analysis and *in vitro* digestion determinations. Amparo Quiles
433 collaborated in the *in vitro* digestion determinations and carried out the CLSM
434 studies. Isabel Hernando supervised the work and carried out the FESEM studies.

435

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- 510
- 511

512 **TABLES**513 **Table 1.** Composition of the formulations studied (g/100 g, wheat flour basis)

514

Ingredients	R0	R30	R50	R70
Wheat flour	100	100	100	100
Sugar	100	100	100	100
Egg yolk	27	27	27	27
Egg white	54	54	54	54
Skim milk	50	50	50	50
Water	0	15	25	35
OptiSol™ 5300	0	3	5	7
Sunflower oil	46	31.6	22	12.4
Sodium bicarbonate	4	4	4	4
Citric acid	3	3	3	3
Salt	1.5	1.5	1.5	1.5

515 R0: control sponge cake, R30: sponge cake with 30 g/100 g of fat replacement; R50:

516 sponge cake with 50 g/100 g of fat replacement; R70: sponge cake with 70 g/100 g

517 of fat replacement.

518

519 **Table 2.** Mean weight loss during baking and maximum height of the sponge cakes,
 520 by formulation.

Parameter	R0	R30	R50	R70
Weight loss (%)	21.48 ^a (0.51)	20.68 ^b (0.27)	19.78 ^c (0.23)	20.09 ^{bc} (0.36)
Height (cm)	10.06 ^a (0.79)	9.72 ^a (0.10)	9.43 ^a (0.71)	9.32 ^a (0.41)

Values in parentheses are the standard deviations. Means in the same row without a common letter are significantly different ($p < 0.05$) according to the LSD multiple range test ($n=3$); R0: control sponge cake, R30: sponge cake with 30 g/100 g of fat replacement; R50: sponge cake with 50 g/100 g of fat replacement; R70: sponge cake with 70 g/100 g of fat replacement.

521

522 **Table 3.** Mean values of sponge cake color parameters, by formulation

Parameter	Crust color				Crumb color			
	R0	R30	R50	R70	R0	R30	R50	R70
L*	40.05 ^a (1.12)	42.57 ^b (1.81)	43.51 ^b (1.90)	43.06 ^b (2.22)	66.86 ^a (1.45)	66.39 ^a (1.99)	66.47 ^a (0.83)	65.40 ^a (2.02)
a*	14.91 ^a (0.5)	15.35 ^b (0.37)	15.31 ^b (0.55)	15.59 ^b (0.52)	-2.23 ^a (0.20)	-1.11 ^b (0.24)	-0.60 ^c (0.16)	-0.20 ^d (0.12)
b*	24.95 ^a (2.18)	27.30 ^b (2.26)	28.00 ^b (2.29)	28.69 ^b (1.67)	22.97 ^a (1.61)	22.48 ^a (1.46)	23.06 ^a (1.25)	22.28 ^a (0.60)
C*_{ab}	29.08 ^a (2.00)	31.34 ^b (1.92)	31.94 ^b (1.88)	32.66 ^b (1.42)	23.08 ^a (1.61)	22.50 ^a (1.46)	23.06 ^a (1.25)	22.29 ^a (0.60)
h*_{ab}	59.02 ^a (2.01)	60.54 ^{ab} (2.26)	61.20 ^b (2.59)	61.42 ^b (1.79)	95.56 ^a (0.56)	92.85 ^b (0.63)	91.48 ^c (0.38)	90.52 ^d (0.30)
ΔE*		3.47	4.62	4.84		1.31	1.68	2.59

523 Values in parentheses are the standard deviations. Means in the same row without a common letter are significantly different ($p < 0.05$) according to the524 LSD multiple range test ($n=12$); R0: control sponge cake, R30: sponge cake with 30 g/100 g of fat replacement; R50: sponge cake with 50 g/100 g of fat

525 replacement; R70: sponge cake with 70 g/100 g of fat replacement.

526

527 **Table 4.** Mean values of sponge cake texture properties, by formulation

Formulation	Hardness (N)	Springiness	Cohesiveness	Chewiness (N)
R0	5.35 ^a (0.71)	0.88 ^a (0.01)	0.73 ^a (0.00)	3.45 ^a (0.43)
R30	5.25 ^a (0.66)	0.89 ^a (0.01)	0.73 ^a (0.01)	3.42 ^a (0.39)
R50	7.84 ^b (1.39)	0.89 ^a (0.01)	0.72 ^a (0.01)	5.01 ^b (0.79)
R70	9.39 ^c (1.81)	0.89 ^a (0.01)	0.73 ^a (0.02)	6.07 ^c (0.99)

528 Values in parentheses are the standard deviations. Means in the same column without a common
 529 letter are significantly different ($p < 0.05$) according to the LSD multiple range test ($n=12$); R0:
 530 control sponge cake, R30: sponge cake with 30 g/100 g of fat replacement; R50: sponge cake
 531 with 50 g/100 g of fat replacement; R70: sponge cake with 70 g/100 g of fat replacement.

532

533 **Table 5.** Proportion of starch (g starch/100 g sponge cake) digested during *in vitro*
 534 digestion of sponge cakes.

Formulation	g starch/100g sponge cake			
	20 min	60 min	90 min	120 min
R0	46.06 ^a (0.36)	54.16 ^a (0.00)	56.46 ^a (1.09)	58.51 ^a (1.33)
R30	47.94 ^a (1.45)	57.32 ^{ab} (0.00)	58.17 ^a (0.72)	58.94 ^a (2.53)
R50	48.79 ^a (1.45)	58.43 ^b (2.05)	61.50 ^b (0.84)	61.92 ^a (0.48)
R70	40.51 ^b (1.81)	52.11 ^c (1.33)	54.16 ^c (1.09)	52.20 ^b (1.21)

535 Values in parentheses are the standard deviations. Means in the same column without a common
 536 letter are significantly different ($p < 0.05$) according to the LSD multiple range test ($n=2$); R0:
 537 control sponge cake, R30: sponge cake with 30 g/100 g of fat replacement; R50: sponge cake
 538 with 50 g/100 g of fat replacement; R70: sponge cake with 70 g/100 g of fat replacement.
 539

540 **FIGURE CAPTIONS**

541 **Figure 1.** Confocal laser scanning microscopy (CLSM) images of sponge cake batters
542 R0, R30, R50 and R70. R0: control sponge cake, R30: sponge cake with 30 g/100g of
543 fat replacement; R50: sponge cake with 50 g/100g of fat replacement; R70: sponge
544 cake with 70 g/100 g of fat replacement. sg: starch granules, fg: fat globules.
545 Magnification 40X.

546

547 **Figure 2.** Scanned images of whole R0, R30, R50 and R70 sponge cakes, 5 x 5 cm
548 field, and respective binarized images. R0: control sponge cake, R30: sponge cake
549 with 30 g/100g of fat replacement; R50: sponge cake with 50 g/100g of fat
550 replacement; R70: sponge cake with 70 g/100 g of fat replacement. Arrows: diffusion
551 pathways

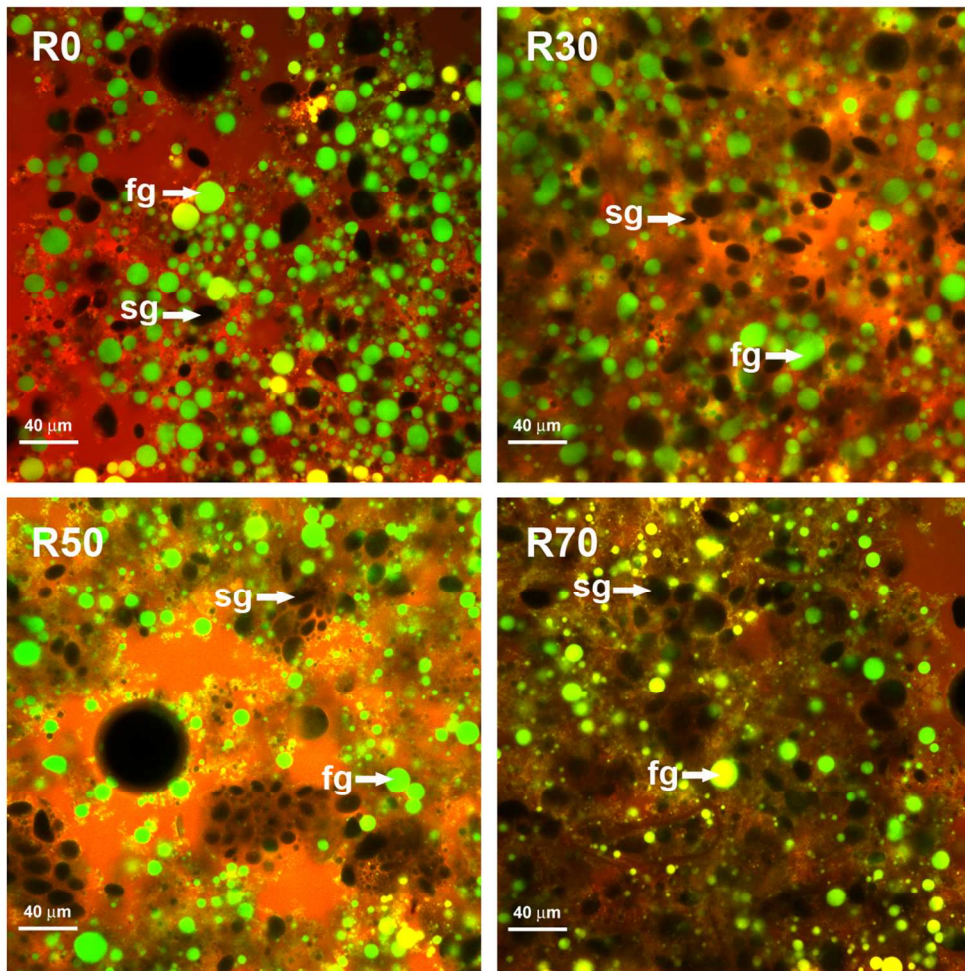
552

553 **Figure 3.** Mean consumer acceptability values of the different sponge cakes. Means in
554 the same attribute without a common letter are significantly different ($p < 0.05$) according
555 to the LSD multiple range test ($n=140$); R0: control sponge cake, R30: sponge cake
556 with 30 g/100g of fat replacement; R50: sponge cake with 50 g/100g of fat
557 replacement; R70: sponge cake with 70 g/100 g of fat replacement.

558

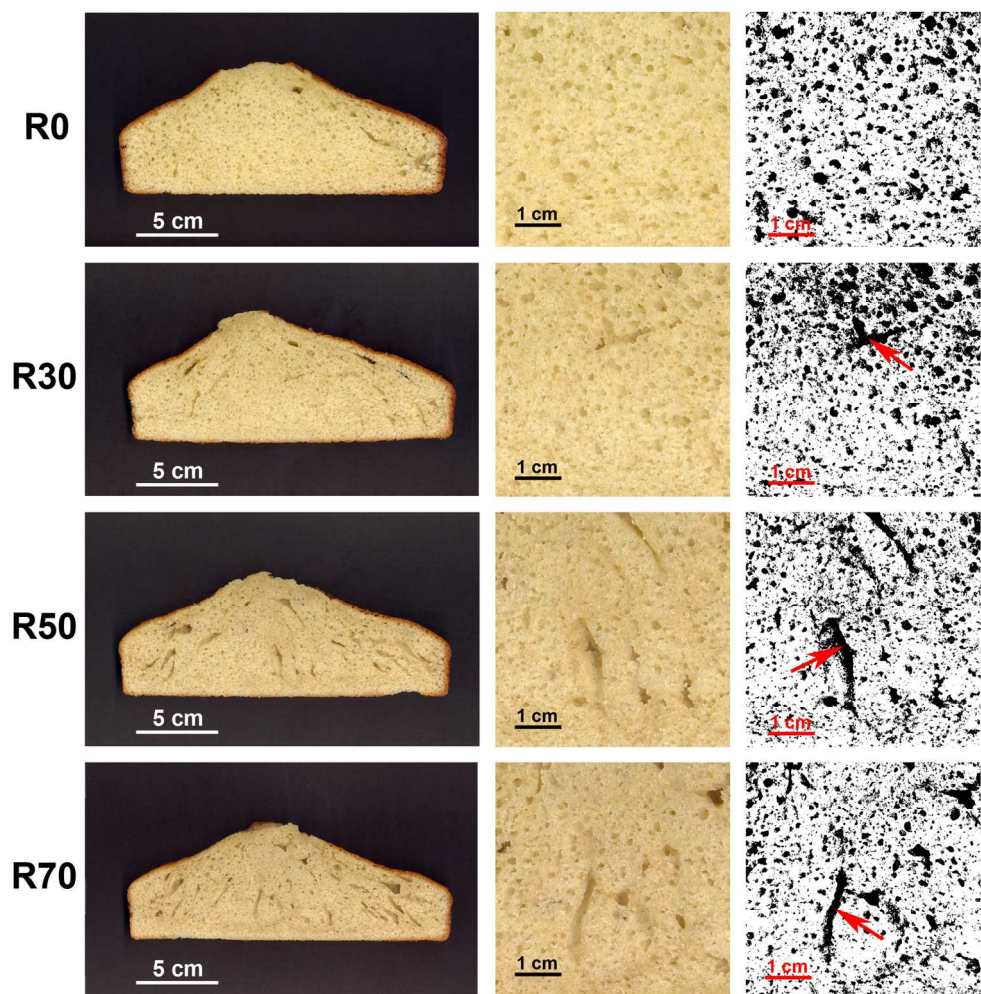
559 **Figure 4.** Field emission scanning electron microscopy (FESEM) images of R0, R30,
560 R50 and R70 sponge cakes. R0: control sponge cake, R30: sponge cake with 30
561 g/100g of fat replacement; R50: sponge cake with 50 g/100g of fat replacement; R70:
562 sponge cake with 70 g/100 g of fat replacement. ~~sg: starch granules~~, o: oil.
563 Magnification 200x. Bar = 20 μ m.

564

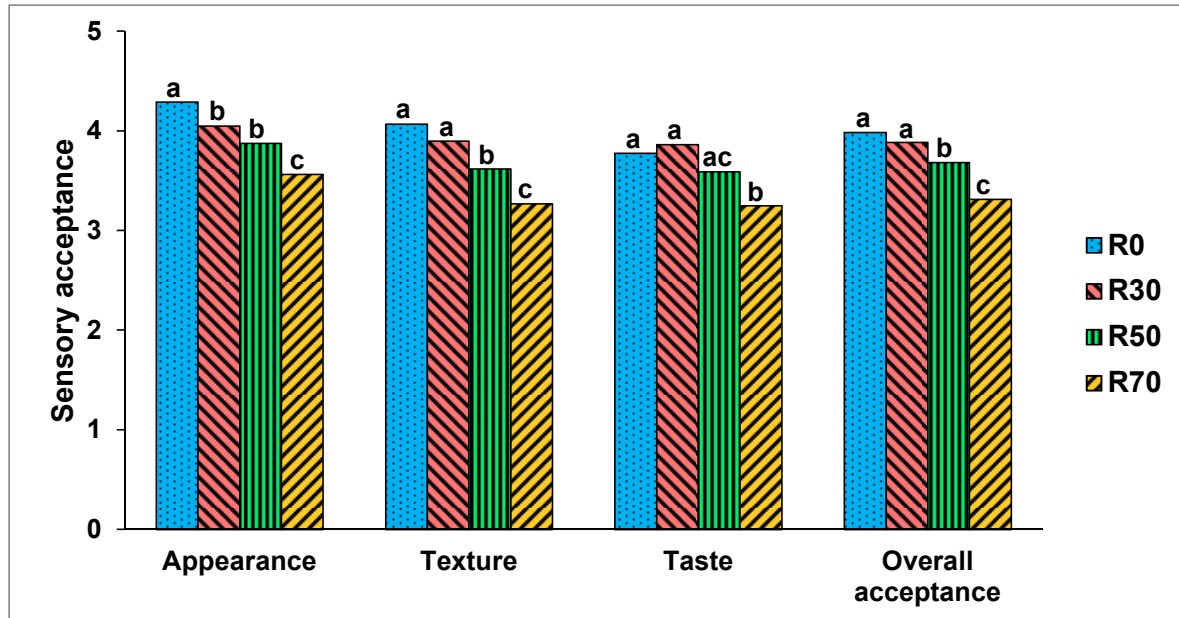


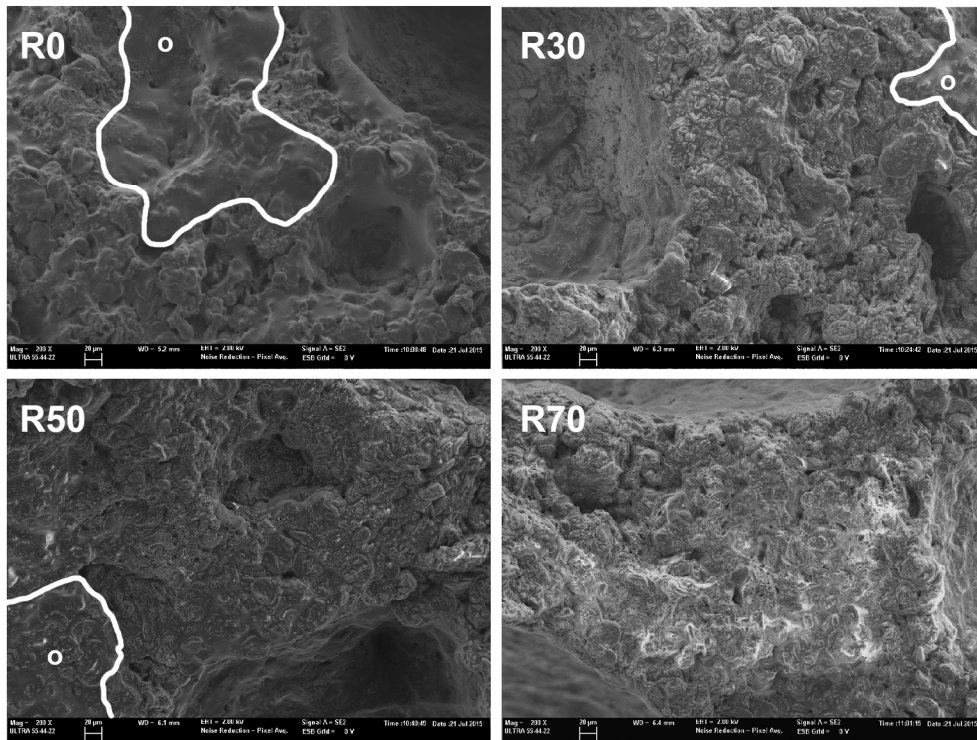
105x105mm (300 x 300 DPI)





165x167mm (300 x 300 DPI)





155x117mm (300 x 300 DPI)

Review

Answer to editor and reviewer

Comments to the Author:

Please address the comments of the reviewer and mine below in a separate cover letter and make all corrections in red font in the revised manuscript.

- Line 154 to 160 Why do you need to include IW in the equation? Why not just use the difference in weight before and after baking?

We decided to include IW in the equation because the initial content of water is very different as we need to add water depending of the amounts of Optisol used in the formulation. We have done it in this way in previous works as that published in Journal of Food Science:

Rodriguez-Garcia J, Puig A, Salvador A, Hernando I. 485 2012. Optimization of a sponge cake formulation with inulin as fat replacer: structure, physicochemical, and sensory properties. J Food Sci 77(2):C189-C97.

- Line 237 Why was amylase not included in the procedure?

The pancreatin used in this study includes amylase. The P3292 pancreatin (Sigma-Aldrich) contains: trypsin, amylase, lipase, ribonuclease and protease produced by exocrine cells of the porcine pancreas. The composition of the pancreatin has been added to the manuscript (lines 240-241)

- Line 275 to 276 R50 seems to have less darkness than the other samples and the control. Please clarify statement.

It was a mistake. This sentence has been changed into "At the highest substitution level (R70) (Line 278)"

- Line 325 The values decreased not rose for a* as more fiber was added. Please revise statements in this section as the reference found the opposite to what you found.

In the crumb, a values increased from -2,23 in R0 to -0,20 in R70, as observed by Lee et al. (2004). In fact they also found negative values for a* when using low levels of substitution (e.g. 20% of flaxseed powder instead of shortening)*

- line 359 to 364 The error bars on figure 3 overlap which means the samples and control are not different at all. Please carefully review the results and revise your statements.

It is true that the error bars in the figure overlap, which can lead to misunderstandings. In fact, in acceptability studies conducted with consumers the results are usually presented without error bars because acceptability can vary a lot among consumers. So, the statistics are often given only with the letters indicating the significant differences (or their absence). Some papers where the acceptability results are presented in this way are:

- *Rodríguez-García J. et al. 2012. Optimization of a sponge cake formulation with inulin as fat replacer: structure, physicochemical, and sensory properties. J Food Sci 77(2):C189-C97.*
- *Corral, S. et al. 2014 Effect of fat and salt reduction on the sensory quality of slow fermented sausages inoculated with Debaryomyces Hansenii yeast. Food Control 45: 1-7*
- *Tarancón, A. et al. 2015. Use of healthier fats in biscuits (olive and sunflower oil): changing sensory features and their relation with consumers' liking. Food Research International 69 (2015) 91-96.*
- *Laura Laguna et al. 2012. Balancing texture and other sensory features in reduced fat short-dough biscuits. Journal of Texture Studies 43: 235-245.*

- Figure 4 I cannot see the starch granules that are identified with arrows. Please clarify the figures or remove the identification of starch granules.

We agree to the editor. The starch granules are hard to distinguish. We have removed their identification in the figure (line 562).

- lines 395 to 399 Please explain why digestibility changed.

The explanation has been added to the manuscript (lines 402-408). "Adding fibre when processing foods is known to limit starch gelatinisation and mask the starch granules, increasing the proportion of resistant starch as observed by Hardacre et al., (2015). This could be also related to the field emission scanning electron microscope (FESEM) images, where the starch granules in

the R70 cake matrix were observed to be very compact within the matrix, which would limit their physical accessibility to the digestive enzymes.”

Reviewers' Comments to the Author:

Reviewer: 1

Comments to the Author

- L98-108 L113, Table 1: A typical sponge cake is made based on the flour, sugar, and eggs. However, a oil was used in your cake preparation. Citation(s) should be mentioned with brief explanation for the oil usage in a sponge cake preparation.

The use of oil instead of shortening in products as sponge cakes and muffins is typical in Mediterranean countries. Some references where oil is used in these products have been added in the manuscript (lines 108-110). “Oil was used instead of shortening as usually done in Mediterranean countries (Rodriguez-Garcia and others, 2012; Martinez-Cervera and others 2011).” The explanation about how to use oil in sponge cakes preparation is provided in line 124-125 (the oil was finally placed on the top)

- L126: batter quantities for being placed in a pan should be mentioned. Total amounts of the prepared batters were different, due to the different amounts of the fat replacers. It may affect the optimum cake texture.

Yes, it is true. It may affect texture and other characteristics as height. We always use the same amount of batter (700 g) for each cake. It has been added to the manuscript (line 128) as we forgot to mention it in the previous version.

- L282-295: Data of weight loss and cake height should be presented.

Data have been included in Table 2

-L391-399 L408-410 Table 4.: The digestibility significantly increased or the same with the increased substitution levels of R30 or R50, but significantly decreased with the highest substitution levels of R70, in all the digestion periods. More explanation is needed for the sudden decrease with the samples with R70 citing more references.

An explanation had been included following the reviewer's suggestion (lines 402-408)

- L401-413: More discussions seem to be needed. Most of the results are jumping to the conclusions such as "clean-labeled ...". To me, sunflower oil is more natural than the processed OptiSol™5300

This section has been rewritten trying not to list again the results and avoiding the jump to the last conclusion. The idea of preparing a low fat/low calorie product has been reinforced in this new version as sunflower oil is natural but more caloric than its replacer- Optisol+water (Lines 411-419).

For Peer Review