

Document downloaded from:

<http://hdl.handle.net/10251/105472>

This paper must be cited as:

Lujan Facundo, MJ.; Soler Cabezas, JL.; Mendoza Roca, JA.; Vincent Vela, MC.; Bes-Piá, M.; Doñate Hernández, S. (2017). A study of the osmotic membrane bioreactor process using a sodium chloride solution and an industrial effluent as draw solutions. *Chemical Engineering Journal*. 322:603-610. doi:10.1016/j.cej.2017.04.062



The final publication is available at

<http://dx.doi.org/10.1016/j.cej.2017.04.062>

Copyright Elsevier

Additional Information

1 **A study of the osmotic membrane bioreactor process using a sodium chloride**
2 **solution and an industrial effluent as draw solutions**

3

4

5 M.J. Luján-Facundo^{1*}, J.L. Soler-Cabezas¹, J.A. Mendoza-Roca¹, M.C. Vincent-Vela¹,
6 A. Bes-Piá¹, S. Doñate-Hernández².

7

8 ¹Instituto de Seguridad Industrial, Radiofísica y Medioambiental, Universitat
9 Politècnica de València, Camino de Vera, s/n, Valencia 46022 (Spain).

10 ²Depuración de Aguas del Mediterráneo (DAM). Avenida Benjamín Franklin, 21.
11 46980 Parque Tecnológico, Paterna, Valencia (Spain).

12

13 Tel. +34963876386

14 e-mail: malufa@etsii.upv.es

15

16

17 **Keywords:** Forward osmosis, Osmotic membrane bioreactor, Membrane fouling,
18 **Wastewater treatment, Draw solution**

19

20

21

22

23

24 **Abstract**

25

26 Osmotic membrane bioreactor (OMBR) is an emerging membrane process which has
27 gained interest in the recent years because of the low energy consumption and the high
28 effluent quality. The osmotic membrane bioreactor combines a forward osmosis (FO)
29 membrane and a biological treatment. However, salt reverse flux is the main problem
30 because of the negative effect of the salt concentration increase in the reactor on the
31 microbial activity. This is the reason why the study of a suitable draw solution (DS) is
32 very important in the overall performance of the reactor. This study compares the
33 process performance using two draw solutions: a 53 g·L⁻¹ NaCl solution and a real
34 waste water solution (waste water from an absorption column consisting mainly of SO₄⁻
35 ² and NH₄-N with concentrations of 153 g·L⁻¹ and 19 g·L⁻¹, respectively). The
36 comparison is focused on the salt reverse flux during the reactor operation, the mixed
37 liquor characteristics, the membrane fouling and the overall performance. The results
38 indicated that the industrial wastewater showed a higher salt reverse flux, but also a less
39 severe fouling and a higher the osmotic pressure difference in comparison with the
40 NaCl solution. In terms of chemical oxygen demand (COD) removal efficiencies, both
41 draw solutions attained values higher than 80%, though the efficiency was slightly
42 lower when the industrial effluent was used as DS. This was related to the higher
43 conductivity reached in the bioreactor when the industrial effluent was used as draw
44 solution. In spite of it, the use of this industrial effluent as draw solution is strongly
45 recommended because of the high permeate fluxes yielded, the low membrane fouling
46 and the lack of necessity of regenerating the draw solution.

47

48 **1. Introduction**

49

50 Water scarcity is further intensified due to climate change, high population growth, and
51 environmental pollution. Nowadays, it already affects billions of people around the
52 world [1]. Overall, efforts for developing and improving novel wastewater treatments
53 and reclamation processes have been progressively introduced, focusing efforts on
54 reusing water even achieving pa drinking water quality [2]. In the recent years,
55 membrane bioreactors (MBR) have gained importance for the municipal and industrial
56 wastewater treatment [3]. Whereas a conventional MBR uses ultrafiltration (UF) or
57 microfiltration (MF) membranes, an OMBR works with FO membranes.

58 Thus, the OMBR is an emerging wastewater treatment technique that combines FO
59 membranes and a biological reactor [4]. In this way, integrating FO membranes in a
60 biological reactor offers many advantages, such as excellent water quality and very low
61 energy consumption (since OMBR process works without applying mechanical
62 pressure) [5,6]. A common OMBR includes a bioreactor, a FO separation unit and a DS
63 system that makes possible the regeneration of the DS or the provision of fresh DS. FO
64 membranes allow water permeation across the FO membrane from activated sludge feed
65 solution to the DS [4,7,8]. The osmotic pressure difference from one membrane side to
66 the other, due to the low-salinity of activated sludge and the high-salinity of the DS, is
67 the driven force of the OMBR [9]. The selection of the DS is of paramount importance,
68 since the salt concentration difference between feed and draw solutions can cause the
69 salt reverse flux due to the Fick's law.

70 The main advantage of the FO nonporous membranes is the high rejection capacity for
71 trace organic compounds [10], pathogens [11] and ions [12]. However, membrane

72 fouling, salinity build-up in the bioreactor and cellular debris accumulated in the mixed
73 liquor are key issues on the OMBR performance. On the one hand, membrane fouling is
74 due to organic fouling, inorganic fouling and biofouling [13]. Organic fouling is due to
75 the adsorption of organic compounds (such as SMP) on the membrane surface or in the
76 membrane pores [13]. The organic substances accumulation together with reversible
77 and irreversible attachment of bacterial cells and extracellulars polymeric substances
78 (EPS) on the membrane surfaces drives to biofilm formation (biofouling) [9]. Inorganic
79 fouling is caused by salts precipitation onto the membrane surface [14]. Membrane
80 fouling implies a water flux reduction, a membrane life decrease and an increase of the
81 operational costs [15–17].

82 In order to mitigate the membrane fouling, a suitable and periodic membrane cleaning is
83 required. The cleaning of FO membranes installed in an OMBR is more complex than
84 the one of the UF or MF membranes in a MBR. UF or MF membranes are usually
85 cleaned by means of hydraulic or chemical backflushing, whereas FO membrane
86 requires an osmotic backflushing. This type of cleaning needs invert the membrane
87 water flux across the membrane. For it, the hypersaline solution is located in the feed
88 tank, whereas distilled water is placed in the DS tank. In this way, water will flow from
89 the DS to the feed side of the membrane, removing organic and inorganic substances
90 from the membrane active layer [9].

91 One of the most important problems described in the OMBR operation is the high
92 increase of the salt concentration in the bioreactor. This phenomenon is due to the
93 rejection of the feed ions by the FO membrane and, at the same time, by the salt reverse
94 flux (salt passage from the DS to the bioreactor). The control of the salt reverse flux is a
95 key factor for the reactor performance. The salt concentration increase in the bioreactor
96 will reduce the effective driving force for water permeation through the membrane,

97 change the microbial community characteristics, rise the SMP and EPS in the bioreactor
98 [18]. Recently, several authors have investigated these critical issues in OMBR
99 technology to improve its application. Luo et al. [19] studied the salt reverse flux for
100 water reuse in a OMBR using different draw solutions. They concluded that ionic
101 organic draw solutes can mitigate salinity build-up in the mixed liquor. Bell et al. [9]
102 compared the fouling behavior of two different FO membranes, cellulose triacetate
103 membranes and polyamide thin film composite (TFC) membranes. This group of
104 authors published that TFC membranes were more prone to fouling during a long-term
105 OMBR study. However, more studies both in pilot and in full-scale plants are necessary
106 to gain knowledge leading to achieve a better OMBR performance.

107 This work aims to study the performance of an OMBR comparing the use of two DS in
108 terms of water flux, reverse salt flux, biological stability and membrane fouling. For it,
109 two OMBR experiments were carried out at the same experimental conditions but using
110 different draw solutions, a sodium chloride solution and an actual industrial wastewater
111 solution of ammonium sulphate. The use of actual industrial effluents in OMBR has
112 been hardly reported in the bibliography. Until now, previous studies have reported
113 results with synthetic wastewater as draw solution. Particularly, the use of a residual
114 ammonia absorption solution has already been not described.

115

116

117

118

119

120 **2. Materials and methods**

121

122 2.1. Synthetic wastewater

123 The simulated wastewater used for the experiment consisted of bacteriological peptone,
124 meat extract (both supplied by Panreac, Spain) and tri-sodium phosphate 12-hydrate
125 (from Panreac, Spain). The concentrations of these chemicals were selected in order to
126 achieve a COD:N:P relation of 100:5:1 mg·L⁻¹ to ensure the appropriate nutrients
127 amount. Chemicals were mixed and dissolved in tap water. The wastewater solution
128 (influent to the OMBR) was prepared three times per week. COD of the simulated
129 wastewater was 4,000 mg·L⁻¹.

130

131 2.2. Draw solutions

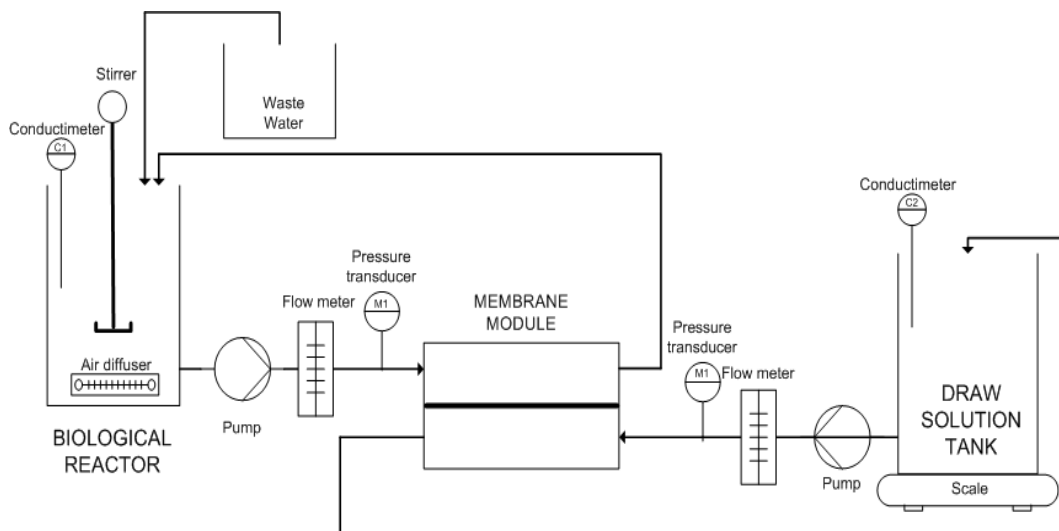
132 In this study, the performance of two different DS was compared. On the one hand, a
133 sodium chloride solution with a concentration of 53 g·L⁻¹ and conductivity of 68 mS/cm
134 was used in test 1. This sodium chloride concentration was selected in order to mimic
135 the saline characteristics of a wastewater from the table olive processing according to
136 Malheiro et al. [20] and Ferrer-Polonio et al. [21]. On the other hand, a liquid effluent
137 from an absorption process for ammonia removal was the DS in the test 2. This
138 wastewater was generated in an industrial wastewater treatment plant and its
139 composition mainly consists of ammonium sulphate (SO₄⁻² and NH₄-N concentrations
140 of 153 g·L⁻¹ and 19 g·L⁻¹, respectively). This waste water had a conductivity of 130
141 mS/cm and pH very low (1.2). Therefore, pH was increased up to 4.0 to ensure that the
142 FO membrane was not chemically damaged.

143 2.3. FO membrane and OMBR plant

144 The FO membrane used in this study was CTA-NW membrane from HTI (USA). The
145 commercial membrane material is cellulose triacetate (CTA) supported by an embedded
146 polyester screen. According to previous research in OMBR (Lay et al. [14] and Wang et
147 al. [18]), active layer was placed in the membrane module facing feed solution (FO
148 mode) to carry out the experiments in order to prevent membrane fouling, especially
149 pore clogging in support layer.

150 Fig. 1 shows a scheme of the OMBR laboratory plant used in this study. The plant was
151 equipped with a separated bioreactor with an effective volume of 1 L. The bioreactor
152 contained a mechanical stirrer (Velp Scientifica, Spain) to agitate the mixed liquor (feed
153 solution to the FO membrane) and a air pump EHEIM 100 (Spain) to provide air in
154 order to keep an oxygen concentration in the bioreactor around $2 \text{ mg} \cdot \text{L}^{-1}$.

155



156

157

158

159

Figure 1: Schematic diagram of the OMBR plant.

160 The membrane module was CF042-FO (Sterlitech, USA) with capacity for a flat sheet
161 membrane with an effective area of 42 cm^2 . The flow rate in both channels of the FO
162 module was $30 \text{ L}\cdot\text{h}^{-1}$. The feed and the draw solutions were pumped through the system
163 by means of two peristaltic pumps (Pumpdrive 5006, Heidolph, Germany). In addition,
164 the conductivity values both in the feed and draw solutions were registered using two
165 conductivity meters model CDH-DS1 from Omega Engineering (United Kingdom). The
166 water mass permeation through the membrane was monitored by the measuring of the
167 evolution of the draw solution weight with the time. Thus, a digital scale PKP (Kern
168 Instruments, Germany) was employed and mass measurements were registered every 15
169 minutes, using the software “Kern Balance Connection SCD-4.0”.

170

171

172 2.4. Experimental sep-up

173 As commented in Section 2.2, two DS were tested under the same experimental
174 conditions. To carry out the start-up of the OMBR, secondary sludge of a municipal
175 wastewater treatment plant (located in Valencia, Spain) was used to seed the bioreactor.
176 The mixed liquor suspended solids (MLSS) concentration was adjusted to $5 \text{ g}\cdot\text{L}^{-1}$ for
177 both tests. The initial pH and conductivity of the mixed liquor were 7.10 and 1.35
178 $\text{mS}\cdot\text{cm}^{-1}$, respectively.

179 The initial hydraulic retention time (HRT) was calculated from the initial membrane
180 water flux (2.65 days and 3.02 days for test 1 and 2, respectively). The food to
181 microorganisms (F/M) ratio tested for the test 1 was $0.12 \text{ g COD}\cdot\text{g SS}^{-1}\cdot\text{d}^{-1}$ (for the
182 three first weeks) and $0.24 \text{ g COD}\cdot\text{g SS}^{-1}\cdot\text{d}^{-1}$ (for the last three weeks). Test 2 was

183 operated at F/M ratio of 0.24 g COD·g SS⁻¹·d⁻¹. This parameter was calculated
184 according to Eq. 1:

185

$$186 \quad \frac{F}{M} (\text{g COD} \cdot \text{g SS}^{-1} \cdot \text{d}^{-1}) = \frac{Q \cdot \text{COD}_e}{\text{SSLM} \cdot V} \quad (1)$$

187

188 In addition, an anoxic phase was introduced in test 2 after each bioreactor feeding to
189 study the overall nutrients removal. With respect to the cleaning procedure, when
190 membrane water flux was lower than 1 LMH, around 1 LMH or every four days
191 instead, a membrane cleaning step was conducted applying a backflushing with
192 deionized water as DS and a sodium chloride solution of 50 g·L⁻¹ (for test 1) and 70 g·L⁻¹
193 (for test 2) as feed solution. After each membrane cleaning step, new fresh DS was
194 supplied for the OMBR operation.

195

196 2.5. Analytical methods

197

198 2.5.1. Measurement of membrane water flux and reverse salt flux

199 The membrane water flux and reverse salt flux were measured in order to characterize
200 the pristine FO membrane tested. For it, deionized water was used as FS and sodium
201 chloride solutions as DS. The experimental water flux J_w (LMH) was calculated at
202 different sodium chloride concentrations in the DS by measuring the weight variation of
203 the draw solution over time as it is described in Eq.2.

204
$$J_w = \frac{\Delta V}{A \cdot \Delta t} \quad (2)$$

205

206 Where, ΔV is the total volume increase in the draw solution tank (L) in a Δt (h) period,
207 and A is the active FO membrane area (m^2). The reverse salt flux J_s ($g \cdot m^{-2} \cdot h^{-1}$) was
208 calculated following Eq. 3.

209

210
$$J_s = \frac{V_t \cdot C_t - V_{t-1} \cdot C_{t-1}}{A \cdot \Delta t} \quad (3)$$

211

212 Where, V_t and C_t are the volume and the concentration of sodium chloride in the feed
213 solution measured at time t .

214

215

216 2.5.2. Wastewater characterization

217 Water quality parameters were analyzed in the bioreactor. These parameters were:
218 COD, total nitrogen (TN), total phosphorous (TP), ammonium nitrogen (NH_4^+ -N),
219 nitrate nitrogen (NO_3 -N) and nitrite nitrogen (NO_2 -N). In addition, when ammonium
220 sulphate was used as DS (test 2), sulphate (SO_4^{2-}) content was also measured in the
221 mixed liquor. Before the analyses, the samples were centrifuged at 10,000 rpm for 15
222 minutes and the supernatant was filtered using a 0.45 μm filter reference 16555 from
223 Sartorius (Spain). In this way, these measurements corresponded to the soluble fraction
224 of the reactor, indicating the quality of the treated water. COD was measured three

225 times per week using kits from Merck (Spain) and SO_4^{-2} , TN, TP, NH_4^+ -N, NO_3 -N and
226 NO_2 -N was evaluated weekly by means of kits from Merck (Spain). In addition, COD
227 was also measured in the prepared wastewater in order to check its properties and
228 calculate the COD removal efficiency of the OMBR.

229

230 2.5.3. Biomass characterization

231 The measurement of MLSS and mixed liquor volatile suspended solids (MLVSS) was
232 carried out following the methodology described in [22]. In addition, capillary suction
233 time of the mixed liquor was measure weekly to study the sludge filterability using the
234 equipment 304M from Triton Electronics Ltd (United Kingdom). Each sample was
235 taken from the bioreactor each 10 days and CST was determined, values were
236 normalized against MLSS concentration and the result was expressed in units of
237 $\text{s}\cdot\text{gMLSS}^{-1}$.

238 Extracted extracellular polymeric substances and soluble microbial products were
239 measured weekly through the experiment by means of analyze proteins and
240 carbohydrates concentrations. Deoxyribonucleic acid (DNA) in SMP was also
241 measured. Thus, eEPS in the sludge were extracted using a cation exchanger resin
242 (Dowex Marathon C, Sigma Aldrich, Spain) following the procedure described by
243 Zuriaga et al. [23]. Proteins content was evaluated using Bicinchoninic acid (BCA)
244 assay test from Novagen. Carbohydrates content was measured with Antrone method
245 [24]. Finally, DNA was determined using Quant-it™ dsDNA HS (0.2–100 ng) kit from
246 Invitrogen (Spain). The three procedures were described extensively in [23].

247

248

249

250 2.6. Membrane microscopy characterization

251 Fouled membrane surfaces morphology were examined for the observation of eventual
252 damages and fouling with a Field Emission Scanning Electron Microscope Ultra 55
253 (Zeiss, Oxford instruments, United Kingdom). In addition, the composition of the
254 fouling layer was characterized by energy diffusive X-ray (EDX) analyzer with the
255 same microscopy. Membrane samples were air-dried in a desiccator before being covered
256 with graphite.

257

258 2.7. Statistical analysis

259 A statistical analysis was carried out by means of the software STATGRAPHICS
260 Centurion XVI to study the relation between the COD removal efficiencies and the feed
261 solution conductivity values. For that, a simple linear regression analysis was
262 conducted. In addition, ANOVA simple analysis was carried out to research if there is
263 any difference between both DS tested in terms of COD removal efficiencies.

264

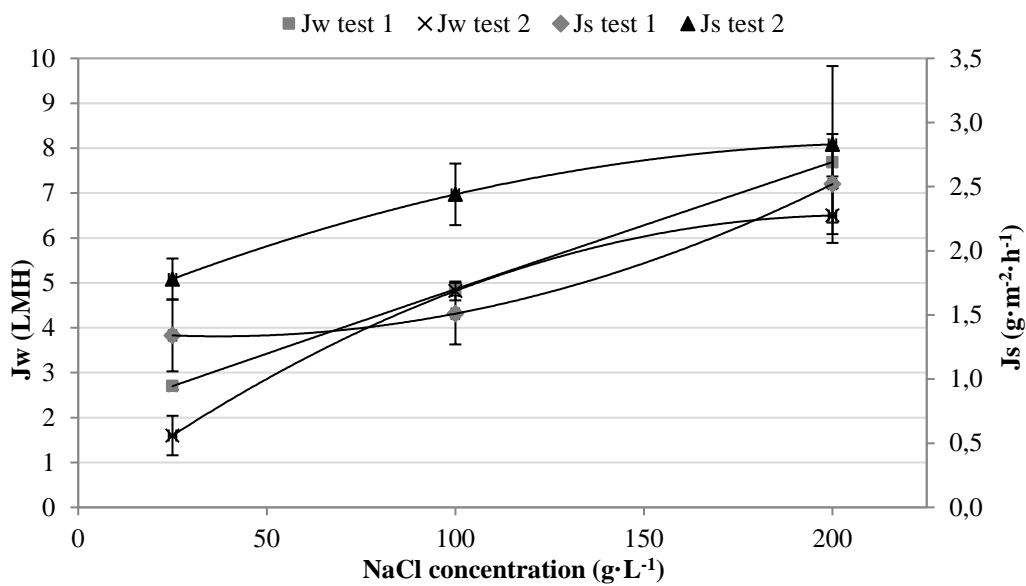
265 **3. Results and discussions**

266

267 3.1. Characterization of the virgin membranes used in the OMBR

268 Fig. 2 shows the J_w and J_s of the CTA-NW virgin membrane samples used in both tests.
 269 The variations in the water flux and reverse salt flux for different NaCl concentrations
 270 in the DS when deionized water was used as feed solution can be observed. As
 271 expected, both membranes samples had practically the same J_w . Slightly higher values
 272 were reported by Yang et al. [25] (4.79 LMH using 58.44 $\text{g}\cdot\text{L}^{-1}$ of NaCl as DS) and
 273 Takahashi et al. [26] (4.8 LMH using 58.44 $\text{g}\cdot\text{L}^{-1}$ of NaCl as DS). Unlike J_w , J_s values
 274 were slightly different for the two tested membranes since they were different
 275 membrane pieces. For example, Takahashi et al. [26] published a higher J_s (about 11
 276 $\text{g}\cdot\text{m}^{-2}\cdot\text{h}^{-1}$), employing the same membrane type.

277



278

279

280 3.2. Water flux and mixed liquor salinity

281 The evolutions of the FO membrane water flux and of the conductivity of the mixed
 282 liquor over the testing period for each test have been represented in Fig. 3. Each vertical
 283 line corresponds with a backflushing cleaning step. The variation of the FO membrane

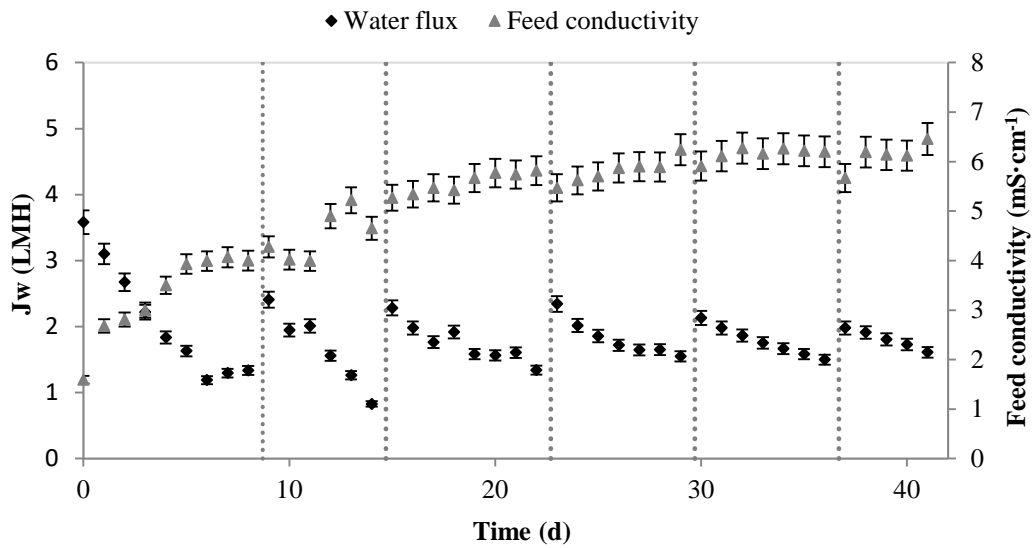
284 water flux over time for test 1 (Fig. 3.a) can be divided into two stages. In the first stage
285 (days 0-10), the water flux significantly decreased from 3.6 LMH to around 1 LMH.
286 This fact could be explained due to the fast deposition of foulants on the membrane
287 surface [27]. After that, when the fouling layer was formed, membrane water flux was
288 more constant varying between 2.2 LMH and 0.9 LMH. In addition, this behavior was
289 very similar to that shown by Wang et al. [28] in their work. Regarding membrane
290 water flux evolution over time for test 2, a similar trend was observed as it is shown in
291 Fig. 3.b. However, fouling layer was formed more slowly since the first backflushing of
292 test 2 achieved a better flux recovery than the first one carried out in test 1.

293 As commented above, the salinity build-up in the bioreactor is a key factor since this
294 phenomenon is an inherent problem associated with OMBR operation. As it can be
295 observed in Fig. 3, the mixed liquor conductivity increased significantly within the first
296 15 days of the OMBR operation for both experiments. For test 1 (Fig. 3.a), thereafter
297 feed conductivity increased slightly until reaching a final value around $6.5 \text{ mS}\cdot\text{cm}^{-1}$.
298 This trend was also observed by Qiu et al. [29]. This fact could be associated with the
299 decrease in the reverse draw solute flux due to the water flux decline. In addition, the
300 fouling layer formed and the daily sampling carried out (sludge withdrawal) helped
301 control salinity build-up. By contrast, for test 2 (Fig. 3.b), a greater increase of the
302 reactor conductivity was observed after the 15th day of operation. More details are given
303 in Section 3.3 in that SO_4^{2-} and NH_4^+ concentrations evolution are showed. In
304 comparison with test 1, more salinity build-up was found in the reactor in test 2 since
305 membrane was less fouled and the osmotic pressure difference was higher, enhancing
306 the salt reverse flux. Thus, the resistance for the reverse salt passage was lower. At the
307 end of the test 2 the feed conductivity slightly decreased due to the sludge withdrawal
308 carried out in this period in order to control the MLSS concentration increase and the

309 salinity build-up. In this way, Luo et al. [30] studied the salinity build-up in a OMBR
310 operated with a DS of 1M of NaCl. They reported a stable feed conductivity value of
311 around $6 \text{ mS}\cdot\text{cm}^{-1}$ after the 8th operation day due to the biofilm formed on the
312 membrane side facing the mixed liquor.

313 a)

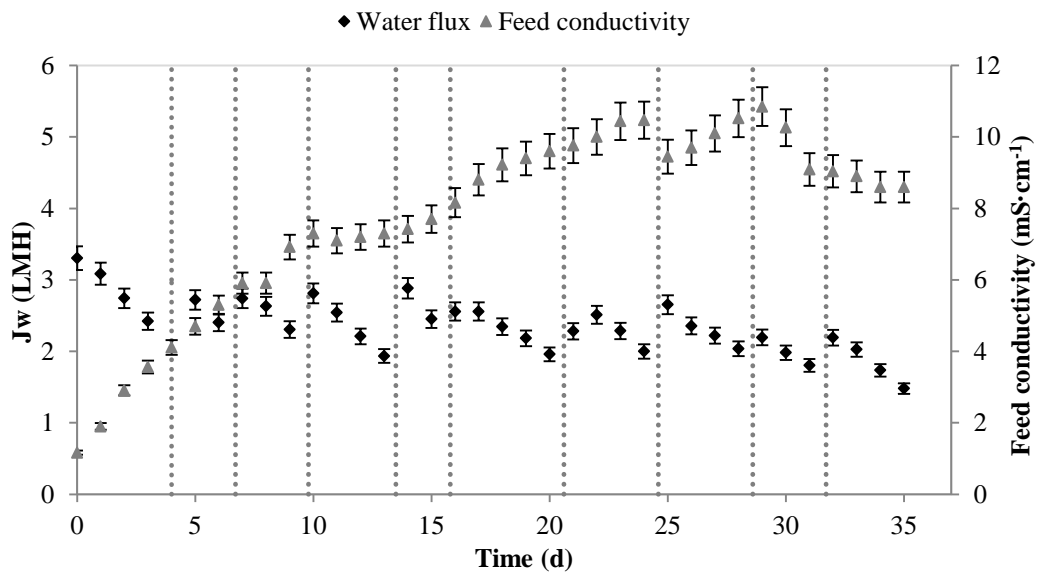
314



315

316

317 b)



318

319

Figure 3: Water flux and feed solution conductivity evolution for a) test 1 and b) test 2.

320

321

322

323

324 3.3. Performance of the OMBR experiments

325 Organic matter and nutrients content in the mixed liquor supernatant were evaluated in
326 order to assess the OMBR performance. Fig. 4 shows COD removal efficiencies for
327 both tests during the experimental period. It is important to highlight that the COD
328 removal was measured on the basis of the soluble COD in the bioreactor instead of
329 calculating the COD content in the DS, since it is considered that FO membrane also
330 rejects organic matter. For test 1, COD removal efficiencies were higher than 90% and
331 slightly decreased at the end of the experiment. By contrast, test 2 had COD removal
332 efficiencies lower than test 1 (between 97.65% and 77.92%). The accumulation of non-
333 biodegradable organic matter (in this case cellular debris since the wastewater influent
334 was a solution of peptone and meat extract) in the biological reactor due to its rejection
335 by the FO membrane (practically 100%) led to a slight and gradual decrease of the COD
336 removal efficiency during the experiment [31,32]. In addition, this decrease was higher
337 in the test 2 than in the test 1, since the conductivity increase in the reactor was higher,
338 leading to biomass inhibition. These results were in concordance with previous studies,
339 such as Huang et al. [33], Qiu et al. [34] and Pathak et al. [35] , who reported COD
340 removal efficiencies operating forward OMBR around 90%.

341 Nutrients removal was not studied in test 1 since no anoxic phase in the OMBR was
342 included. However, $\text{NH}_4^+\text{-N}$ and TP content in the supernatant were measured to ensure
343 that there were enough nutrients to carry out the biological process and also to detect

344 their eventual accumulation in the reactor. The $\text{NH}_4^+\text{-N}$ and TP concentration ranges in
345 the supernatant were $3\text{-}8\text{ mg}\cdot\text{L}^{-1}$ and $1\text{-}4\text{ mg}\cdot\text{L}^{-1}$, respectively.

346 By contrast, the removal of nutrients was analyzed in test 2 as it is shown in Fig. 5.
347 Thus, this test was divided into two stages: a first aerobic stage (until 18th-day of
348 operation) and a second anoxic/aerobic stage (from 18th-day of operation on). The study
349 of the nutrients removal in tests 2 was of special interest due to the reverse ammonium-
350 nitrogen flux increasing its concentration in the biological reactor.

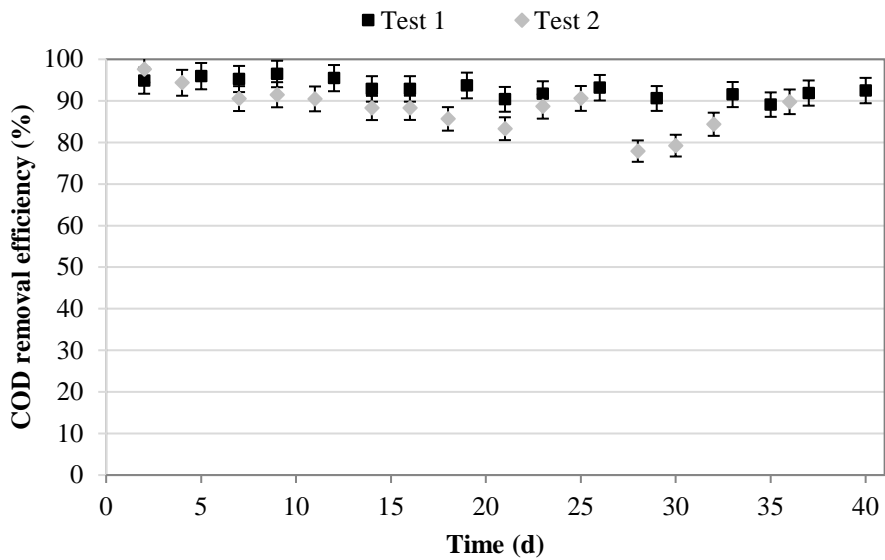
351 On the one hand, during the first stage (only aerobic reaction) it is observed an increase
352 of the TN concentration. This was due to the TN rejection by the membrane in spite of
353 the nutrient biomass assimilation. Nitrification process occurred in the reactor since
354 $\text{NO}_3\text{-N}$ concentration increased. Finally, these nitrogen compounds were rejected by FO
355 membrane and, as a result, TN concentration increased in the bioreactor [36] as it is
356 observed in Fig. 5. This trend was also observed by Luo et al. [19], who operated an
357 OMBR and nitrification process was developed in the biological treatment.

358 On the other hand, from 18th-day of operation, when anoxic phase was included, both
359 $\text{NO}_3\text{-N}$ and TP concentration decreased indicating that there was denitrification and
360 that phosphorous assimilation was enhanced. Specifically, the removal of TP was
361 80.5%. Similar results were published by Pathak et al. [35] who obtained a TP removal
362 in the bioreactor of 81.22%.

363 Focusing on SO_4^{2-} and $\text{NH}_4^+\text{-N}$, i.e. the main components of the DS, it has to be
364 highlighted that their high reverse flux due to Fick's law was responsible for the
365 conductivity increase in the biological reactor. The periodical sludge withdrawals seem
366 to be critical in order to avoid the sulfates precipitation (mainly as calcium sulfate) on
367 the membrane surface due to the high sulfates concentration (Fig. 3.b).

368 Concerning the pH, a slight increase (0.94 and 0.78 units for test 1 and 2, respectively)
369 was observed during the OMBR operation mainly due to reverse draw solute flux. The
370 forward diffusion of protons from the bioreactor into the DS, in combination with the
371 reverse transport of cations (like sodium in the case of test 1) to maintain the
372 electroneutrality of the mixed liquor, led to a pH rise in the biological reactor [19].
373 However, this did not affect the biological process (pH values remained in the
374 appropriate range for biological activity).

375



376

377

Figure 4: COD removal efficiency during the experimental period for both experiments.

378

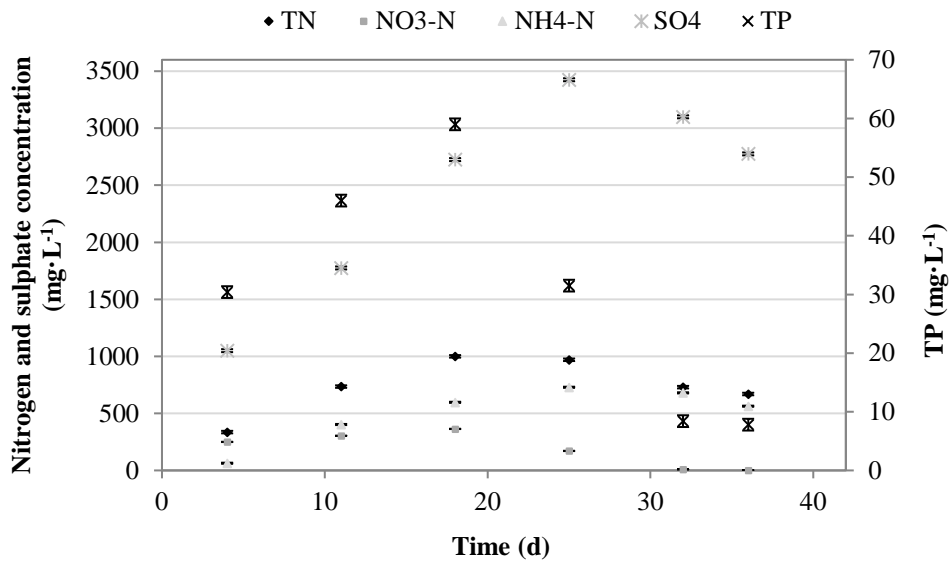


Figure 5: Nutrients and SO₄ evolution in the feed solution during the experimental period.

3.4. Biomass characteristics

Salinity build-up in the bioreactor could alter the biomass characteristics during OMB operation. Previous studies [31,37] have reported that higher salt concentrations could increase the endogenous respiration of microorganisms. The biomass could increase the secretion of organic cellular substances, which would lead to an enhancement of SMP and eEPS production with the aim of cell protection. Fig. 6 represents the SMP and eEPS concentrations in terms of proteins and carbohydrates during the experimental period. DNA has also been included in the SMP in order to evaluate eventual cell lysis. It can be clearly observed that SMP (proteins, carbohydrates and DNA) concentrations were higher in test 2 than in test 1. This result confirms the higher stress of the biomass in test 2 caused by the higher salinity than in test 1 (Fig. 3) [38]. By contrast, this fact had no influence on the eEPS concentration, which was stable and very similar in both tests probably due to the low working organic loads. Similar results were published by Luo et al. [19] related with EPS content in a OMBR. These authors reported that there

397 was no significant variation in the EPS content probably due to a balance between its
398 release and hydrolysis.

399

400 However, for test 1 and from sample 3 (at that time the F/M ratio was increased to 0.24
401 g COD·g SS⁻¹·d⁻¹), SMP concentrations were constant and eEPS decreased with the
402 operation time. This indicates that biomass was adapted to salinity build-up. Thus, the
403 increase of the organic load resulted in a lower eEPS release. These results agree with
404 those reported by Wang et al. [39].

405

406 From Fig. 6.a and 6.b, it has to be highlighted that the concentration difference between
407 proteins and carbohydrates was high. In this way, Sabia et al. [40] published that the
408 ratio between proteins and carbohydrates in SMP dramatically increased with the sludge
409 retention time (SRT). These results were in agreement with those obtained in this study,
410 since in both tests the SRT was increasing since the only sludge withdrawals were
411 carried out to take the samples.

412

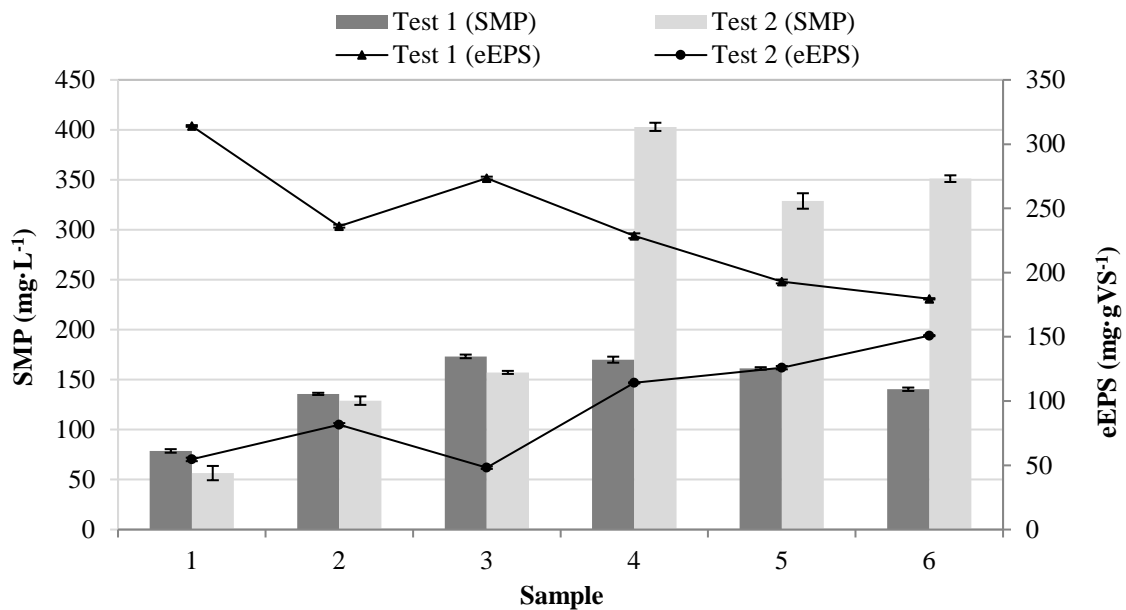
413 Finally, Fig. 6.c relates the measured DNA concentrations in the supernatant from both
414 tests. DNA content indicates that cell lysis occurs [41]. Cell lysis was much higher in
415 test 2, which is in concordance with the above explained results. The high stress caused
416 by the quick conductivity increase and the high increase of the ammonium nitrogen
417 concentration were responsible for the cell lysis. Furthermore, it was detected a direct
418 relation between DNA and protein concentrations, what corroborates that measured
419 proteins could come from the released cellular material from the bacteria, as previously
420 published Zuriaga et al. [42].

421

422 Regarding to physic-chemical properties of sludge, CST is an interesting index for
 423 sludge filterability [43]. Normalized CST with respect to TSS (nCST) was calculated to
 424 investigate the effect of salts concentration on the filterability of the sludge as shown in
 425 Table 1. In general terms, the low CST values obtained for both tests indicated good
 426 sludge quality from the point of view of its filterability and dewaterability. Thus, it was
 427 observed that the presence of salts concentration in the mixed liquors did not affect the
 428 filterability of the sludge since the values were very similar during the experimental
 429 period for both tests. According to Zhang et al. [44], EPS concentration is considered to
 430 be an important factor affecting sludge filterability. These authors published that CST
 431 decreased as increases EPS concentration. This fact was due to the improvement of
 432 sludge dewaterability as a consequence of the increase of flocs size. The same trend was
 433 observed in results detailed in Table 1 when CST values of samples 1 and 2 are
 434 compared.

435

436 a)

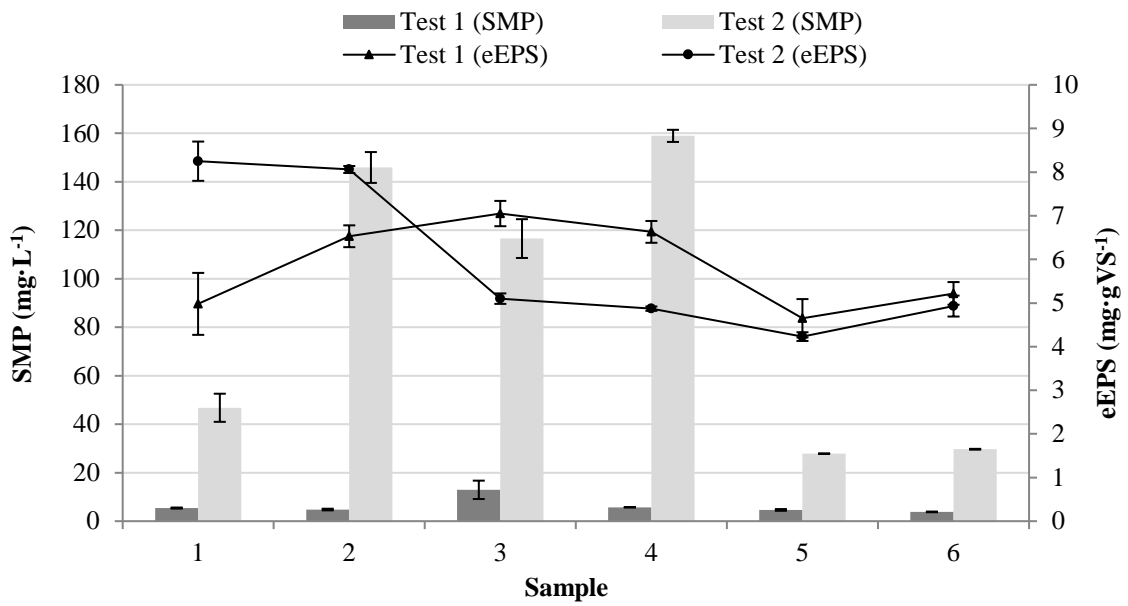


437

438

439

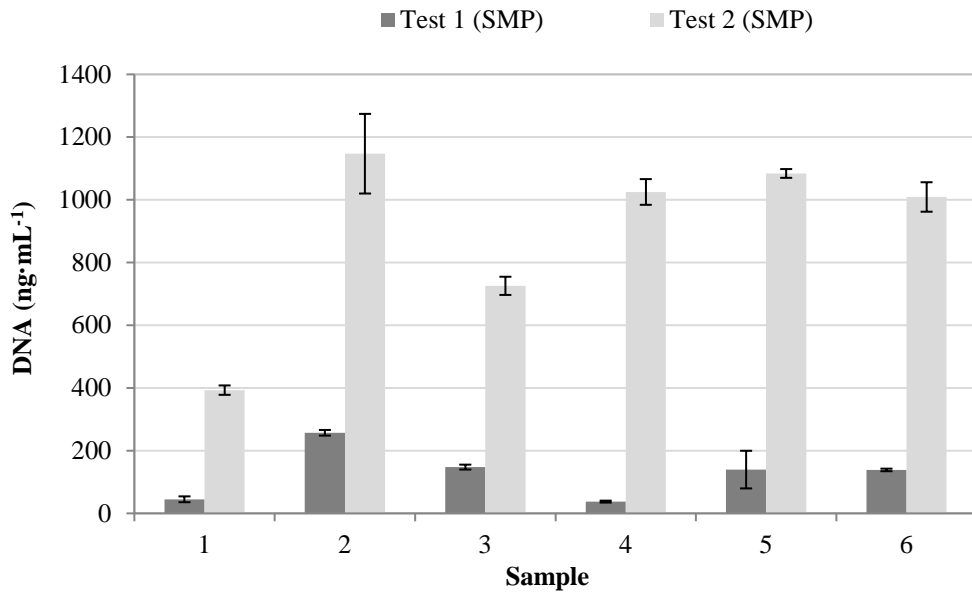
440 b)



441

442

443 c)



444

445 **Figure 6: SMP and eEPS concentrations in terms of a) proteins, b) carbohydrates and c) DNA.**

446

447

Table 1: nCST for both test during the experimental time.

nCST (s·L·gTSS ⁻¹)	
Test 1	Test 2

Sample 1	3.08 ± 0.15	2.51 ± 0.11
Sample 2	4.92 ± 0.20	2.19 ± 0.10
Sample 3	4.79 ± 0.21	1.98 ± 0.08

448

449

450 3.5. Membrane microscopy analysis

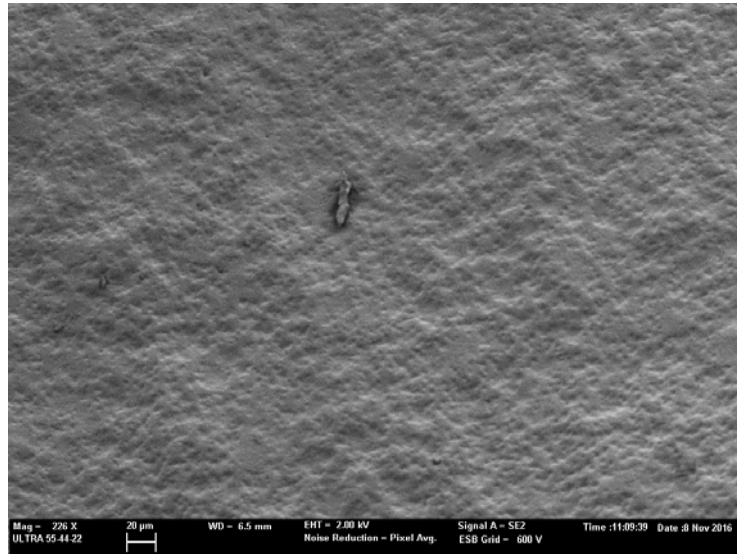
451 According to Gu et al. [45], CTA FO membranes were covered by a fouling layer
452 during the process but foulants could be easily removed applying a backflushing step.
453 However, in this study and for both tests, FO membranes were fouled and the foulants
454 after 40 operation days could not be totally removed by backflushing (the water flux
455 was not totally restored up to the initial value). To check the membrane fouling, surface
456 morphology of the fouled FO membrane used for test 1 is illustrated in Fig. 7 since no
457 significant differences were found between the two used membranes. Besides, an
458 elemental composition of fouling layer is shown in Fig. 8, Fig. 8.a for membrane used
459 in test 1 and Fig. 8.b for membrane used in test 2.

460 From FE-SEM images it was observed that a fouling layer was detected on the fouled
461 FO membrane after the experimental period and foulant clusters were distributed over
462 the membrane surface. EDX results are illustrated in Fig. 8. In addition to the high
463 peaks of C and O coming from the membrane material, low peaks for nitrogen,
464 phosphorous, calcium, magnesium, chloride, sulphur and sodium were also detected. It
465 has to be highlighted that the S and Ca peaks are slightly higher in the membrane of the
466 tests 2 due to a higher sulphate reverse flux that entailed the accumulation of sulphates
467 in the reactor with the subsequent calcium sulphate precipitation.

468 According to Zhao et al. [46], inorganic compounds could also form precipitates in the
469 organic fouling layer, even forming bridges between divalent cations and organic
470 molecules (e.g. proteins, carbohydrates) on the membrane surface.

471

472



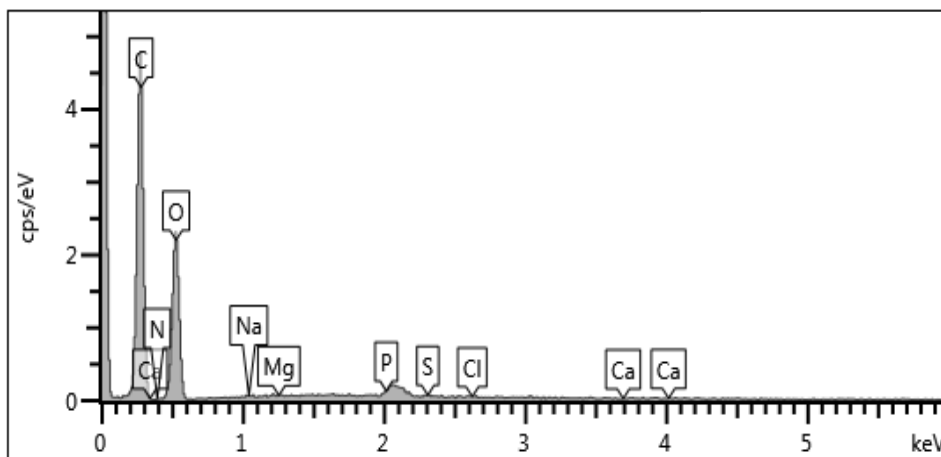
473

474

Figure 7: FE-SEM image of active layer of fouled membrane used in test 1.

475

476 a)



477

478

479 b)

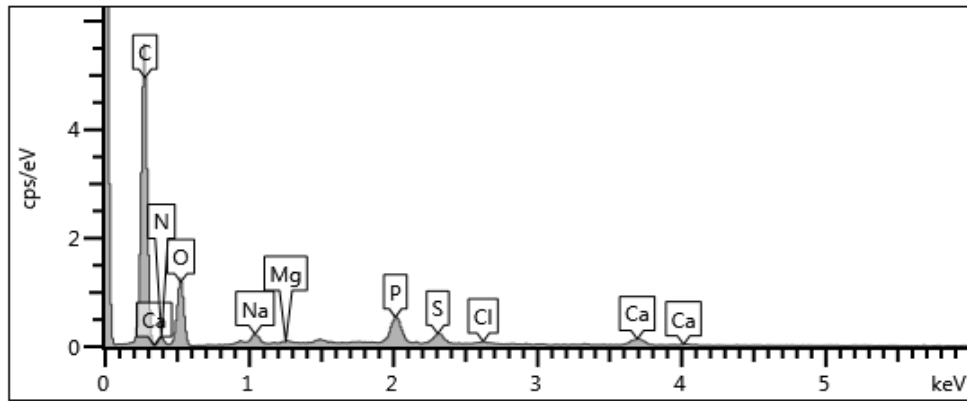


Figure 8: EDX analysis of fouled membranes a) test 1 and b) test 2.

480

481

482

483

484 3.6. Statistical analysis

485 A simple linear regression analysis was calculated with the software STATGRAPHICS
 486 to confirm that there is relationship between COD removal efficiency (dependent
 487 variable) and the feed solution conductivity (independent variable). The coefficient R^2
 488 shows the variability in the response that is represented by the fitted model, indicating
 489 how well the model fits the response variable. Eq. 4 and 5 shows a linear regression
 490 between the COD removal efficiency and the feed solution conductivity for test 1 and 2,
 491 respectively. The R^2 for Eq. 4 and 5 were 56.66% and 65.13%, respectively. In addition,
 492 the sign of the effect indicates if it produces an increase (+) or a decrease (-) of the
 493 variable response.

$$494 \text{ COD removal (\%)} = 99.46 - 1.24 \times \text{Feed Conductivity (mS/cm)} \quad (4)$$

495

$$496 \text{ COD removal (\%)} = 105.55 - 2.27 \times \text{Feed Conductivity (mS/cm)} \quad (5)$$

497

498 Finally, if p-values were lower than 0.05, it was considered that independent variables
499 (in this case, feed solution conductivities) were statistically significant at the 95.0%
500 confidence level. In this case, for both tests, feed solution conductivity is statistically
501 significant since p-values were lower than 0.05 (0.0005 and 0.0002 for test 1 and 2,
502 respectively).

503 In addition, one-way ANOVA was analyzed to study if COD removal efficiencies
504 (response variable) and the draw solution tested (named factor) were independents or
505 not. In this way, from STATGRAPHICS results it can be concluded that there were
506 statistically significant differences between the mean COD removal efficiency value
507 from test 1 and from test 2 since p-value was 0.0011. In addition, the F-ratio differs
508 significantly of 1 (F-ratio was 12.9959) indicating that the null hypothesis of equality of
509 means of the COD removal efficiencies was rejected.

510

511

512 **Conclusions**

513

514 Results reported here demonstrate that both DS tested were useful to carry out an
515 OMBR process for treating simulated wastewater, since results in terms of COD
516 removal efficiencies were always higher than 80% in spite of the salt concentration
517 increase in the reactor. Comparing both DS, salt reverse flux was higher when the
518 industrial wastewater was used as DS. With industrial waste water as DS, the COD
519 removal efficiency was slightly lower since SMPs were produced at a higher extent due
520 to the cellular stress caused but the sharper salt concentration increment. The SEM and

521 EDX observations on the FO fouled membrane indicated the presence of a fouling layer
522 combining organic and inorganic scaling.
523 Nevertheless, the less membrane fouling and the higher water flux in comparison with
524 the NaCl solution make the use of the ammonium absorption effluent as DS attractive
525 for the OMBR operation. In addition, the ammonium-nitrogen from the DS could be
526 eliminated at a certain extent, since after its permeation through the membrane nitrogen
527 could be nitrified and finally denitrified in the reactor.

528

529

530

531 **Acknowledgments**

532

533 This study was supported by the Spanish Ministry of Economy and Competitiveness
534 through the project RTC-2015-3582-5-AR.

535

536 **References**

537 [1] M.A. Shannon, P. W. Bohn, M. Elimelech, J.G. Georgiadis, B.J. Mariñas, A.M.
538 Mayes, Science and technology for water purification in the coming decades, Nature
539 452 (2008) 301-310.

540

541 [2] N.T. Hancock, P. Xu, M.J. Roby, J.D. Gomez, T.Y. Cath, Towards direct
542 potable reuse with forward osmosis: Technical assessment of long-term process
543 performance at the pilot scale, J. Membr. Science. 445 (2013) 34–46.

544

545 [3] X. Wang, V.W.C. Chang, C.Y. Tang, Osmotic membrane bioreactor (OMBR)
546 technology for wastewater treatment and reclamation: Advances, challenges, and
547 prospects for the future, *J. Memb. Sci.* 504 (2016) 113–132.

548 [4] A. Achilli, T.Y. Cath, E.A. Marchand, A.E. Childress, The forward osmosis
549 membrane bioreactor: A low fouling alternative to MBR processes, *Desalination*. 239
550 (2009) 10–21.

551

552 [5] T.Y. Cath, A.E. Childress, M. Elimelech, Forward osmosis: Principles,
553 applications, and recent developments, *J. Memb. Sci.* 281 (2006) 70–87.

554

555 [6] T.S. Chung, S. Zhang, K.Y. Wang, J. Su, M.M. Ling, Forward osmosis
556 processes: Yesterday, today and tomorrow, *Desalination* 287 (2012) 78–81.

557

558 [7] R.W. Holloway, A.S. Wait, A. F. da Silva J. Herron, M.D. Schutter, K. Lampi,
559 T.Y. Cath, Long-term pilot scale investigation of novel hybrid ultra fi ltration-osmotic
560 membrane bioreactors, *Desalination* 363 (2015) 64–74.

561

562 [8] R.W. Holloway, A. Achilli, T.Y. Cath, The osmotic membrane bioreactor: a
563 critical review, *Environ. Sci. Water Res. Technol.* 1 (2015) 581–605.

564

565 [9] E.A. Bell, R.W. Holloway, T.Y. Cath, Evaluation of forward osmosis membrane
566 performance and fouling during long-term osmotic membrane bioreactor study, *J.*
567 *Memb. Sci.* 517 (2016) 1–13.

568

- 569 [10] M. Xie, W.E. Price, L.D. Nghiem, Rejection of pharmaceutically active
570 compounds by forward osmosis: Role of solution pH and membrane orientation, *Sep.*
571 *Purif. Technol.* 93 (2012) 107–114.
- 572 [11] L.A. Hoover, W.A. Phillip, A. Tiraferri, N.Y. Yip, M. Elimenech, Forward with
573 Osmosis: Emerging Applications for Greater Sustainability, *Environ. Sci. Technol.* 45
574 (2011) 9824–9830.
- 575
- 576 [12] K.L. Hickenbottom, N.T. Hancock, N.R. Hutchings, E.W. Appleton, E.G.
577 Beaudry, P. Xu, T.Y. Cath, Forward osmosis treatment of drilling mud and fracturing
578 wastewater from oil and gas operations, *Desalination*. 312 (2013) 60–66.
- 579
- 580 [13] F. Meng, S.R. Chae, A. Drews, M. Kraume, H.-S. Shin, F. Yang, Recent
581 advances in membrane bioreactors (MBRs): membrane fouling and membrane material,
582 *Water Res.* 43 (2009) 1489–1512.
- 583
- 584 [14] W.C.L. Lay, Q. Zhang, J. Zhang, D. McDougald, C. Tang, R. Wang, Y. Liu,
585 A.G. Fane, Study of integration of forward osmosis and biological process: Membrane
586 performance under elevated salt environment, *Desalination*. 283 (2011) 123–130.
- 587
- 588 [15] H. Yamamura, K. Kimura, Y. Watanabe, Mechanism involved in the evolution
589 of physically irreversible fouling in microfiltration and ultrafiltration membranes used
590 for drinking water treatment, *Environ. Sci. Technol.* 41 (2007) 6789–6794.
- 591
- 592 [16] A. Drews, Membrane fouling in membrane bioreactors. Characterisation,
593 contradictions, cause and cures, *J. Memb. Sci.* 363 (2010) 1–28.

594

595 [17] S. Judd, *The MBR book: Principles and Applications of Membrane Bioreactors*
596 *for Water and Wastewater Treatment*, Ed Elsevier, Amsterdam, The Netherlands, 2010.

597 [18] X. Wang, Y. Chen, B. Yuan, X. Li, Y. Ren, Impacts of sludge retention time on
598 sludge characteristics and membrane fouling in a submerged osmotic membrane
599 bioreactor, *Bioresour. Technol.* 161 (2014) 340–347.

600

601 [19] W. Luo, F.I. Hai, W.E. Price, M. Elimelech, L.D. Nghiem, Evaluating ionic
602 organic draw solutes in osmotic membrane bioreactors for water reuse, *J. Memb. Sci.*
603 514 (2016) 636–645.

604

605 [20] R. Malheiro, A. Sousa, S. Casal, A. Bento, J.A. Pereira, Cultivar effect on the
606 phenolic composition and antioxidant potential of stoned table olives, *Food Chem.*
607 *Toxicol.* 49 (2011) 450–457.

608

609 [21] E. Ferrer-Polonio, J.A. Mendoza-Roca, A. Iborra-clar, J.L. Alonso-Molina, L.
610 Pastor-Alcañiz, Biological treatment performance of hypersaline wastewaters with high
611 phenols concentration from table olive packaging industry using sequencing batch
612 reactors, *J. Ind. Eng. Chem.* 43 (2016) 44–52.

613

614 [22] APHA, AWWA, WEF, *Standard Methods for the Examination of Water and*
615 *Wastewater*, 21st ed., American Public Health Association, Washington DC, 2005.

616

617 [23] E. Zuriaga-Agustí, A. Bes-Piá, J.A. Mendoza-Roca, J.L. Alonso-Molina,
618 Influence of extraction methods on proteins and carbohydrates analysis from MBR

619 activated sludge flocs in view of improving EPS determination, *Sep. Purif. Technol.*
620 112 (2013) 1–10.

621 [24] B. Frolund, R. Palmgren, K. Keiding, P. Halkjaer Nielsen, Extraction of
622 extracellular polymers from activated sludge using a cation exchange resin, *Water Res.*
623 (1996) 1749–1758.

624

625 [25] E. Yang, K.J. Chae, A.B. Alayande, K.-Y. Kim, I.S. Kim, Concurrent
626 performance improvement and biofouling mitigation in osmotic microbial fuel cells
627 using a silver nanoparticle-polydopamine coated forward osmosis membrane, *J. Memb.*
628 *Sci.* 513 (2016) 217–225.

629

630 [26] T. Takahashi, M. Yasukawa, H. Matsuyama, Highly condensed polyvinyl
631 chloride latex production by forward osmosis: Performance and characteristics, *J.*
632 *Memb. Sci.* 514 (2016) 547–555.

633

634 [27] S. Zou, Y.N. Wang, F. Wicaksana, T. Aung, P.C.Y. Wong, A.G. Fane, C.Y.
635 Tang, Direct microscopic observation of forward osmosis membrane fouling by
636 microalgae: Critical flux and the role of operational conditions, *J. Memb. Sci.* 436
637 (2013) 174–185.

638

639 [28] X. Wang, C. Wang, C.Y. Tang, T. Hu, X. Li, Y. Ren, Development of a novel
640 anaerobic membrane bioreactor simultaneously integrating micro filtration and forward
641 osmosis membranes for low- strength wastewater treatment, *J. Memb. Sci.* 527 (2017)
642 1–7.

643

- 644 [29] G. Qiu, S. Zhang, D. S. S. Raghavan, S. Das, Y.P. Ting, Towards high through-
645 put biological treatment of municipal wastewater and enhanced phosphorus recovery
646 using a hybrid microfiltration- forward osmosis membrane bioreactor with hydraulic
647 retention time in sub-hour level, *Bioresour. Technol.* 219 (2016) 298–310.
- 648
- 649 [30] W. Luo, H. V Phan, M. Xie, F.I. Hai, W.E. Price, M. Elimelech, L.D. Nghiem,
650 Osmotic versus conventional membrane bioreactors integrated with reverse osmosis for
651 water reuse: Biological stability, membrane fouling, and contaminant removal, *Water*
652 *Res.* 109 (2017) 122–134.
- 653
- 654 [31] L. Chen, Y. Gu, C. Cao, J. Zhang, J-W. Ng, C. Tang, Performance of a
655 submerged anaerobic membrane bioreactor with forward osmosis membrane for low-
656 strength wastewater treatment, *Water Research* 50 (2014) 114-123.
- 657
- 658 [32] G. Qiu, Y.M. Law, Y.M.S. Das, Y.P Ting, Direct and complete phosphorus
659 recovery from municipal wastewater using a hybrid microfiltration-forward osmosis
660 membrane bioreactor process with seawater brine as draw solution, *Environ. Sci.*
661 *Technol.* 49 (2015) 6156–6163.
- 662
- 663 [33] L. Huang, D.J. Lee, J.Y. Lai, Forward osmosis membrane bioreactor for
664 wastewater treatment with phosphorus recovery, *Bioresour. Technol.* 198 (2015) 418–
665 423.
- 666
- 667 [34] G. Qiu, S. Zhang, D.S. Srinivasa Raghavan, S. Das, Y-P. Ting, The potential of
668 hybrid forward osmosis membrane bioreactor (FOMBR) processes in achieving high

669 throughput treatment of municipal wastewater with enhanced phosphorus recovery,
670 Water Res. 105 (2016) 370–382.

671 [35] N. Pathak, L. Chekli, J. Wang, Y. Kim, S. Phuntsho, S. Li, N. Ghaffour, T.
672 Leiknes, H. Shon, Performance of a novel baffled osmotic membrane bioreactor-
673 microfiltration hybrid system under continuous operation for simultaneous nutrient
674 removal and mitigation of brine discharge, *Article in press*, Bioresour. Technol. (2017).

675

676 [36] R.W. Holloway, A.E. Childress, K.E. Dennett, T.Y. Cath, Forward osmosis for
677 concentration of anaerobic digester centrate., Water Res. 41 (2007) 4005–4014.

678

679 [37] W.C.L. Lay, Y. Liu, A.G. Fane, Impacts of salinity on the performance of high
680 retention membrane bioreactors for water reclamation: A review, Water Res. 44 (2010)
681 21–40.

682

683 [38] W. Luo, F.I. Hai, J. Kang, W.E. Price, W. Guo, H.H. Ngo, K. Yamamoto, L.D.
684 Nghiem, Effects of salinity build-up on biomass characteristics and trace organic
685 chemical removal: implications on the development of high retention membrane
686 bioreactors, Bioresour. Technol. 177 (2015) 274–281.

687

688 [39] X. Wang, Y. Zhao, B. Yuan, Z. Wang, X. Li, Y. Ren, Comparison of biofouling
689 mechanisms between cellulose triacetate (CTA) and thin-film composite (TFC)
690 polyamide forward osmosis membranes in osmotic membrane bioreactors, Bioresour.
691 Technol. 202 (2016) 50–58.

692

693 [40] G. Sabia, M. Ferraris, A. Spagni, Effect of solid retention time on sludge
694 filterability and biomass activity: Long-term experiment on a pilot-scale membrane
695 bioreactor treating municipal wastewater, *Chem. Eng. J.* 221 (2013) 176–184.

696 [41] E. Zuriaga-Agustí, G. Garrido-Mauri, J.A. Mendoza-Roca, A. Bes-Piá, J.L.
697 Alonso-Molina, Reduction of the sludge production in a sequencing batch reactor by
698 addition of chlorine dioxide: Influence on the process performance, *Chem. Eng. J.* 209
699 (2012) 318–324.

700

701 [42] E. Zuriaga-Agustí, J.A. Mendoza-Roca, A. Bes-Piá, J.L. Alonso-Molina, I.
702 Amorós-Muñoz, Sludge reduction by uncoupling metabolism: SBR tests with para-
703 nitrophenol and a commercial uncoupler., *J. Environ. Manage.* 182 (2016) 406–411.

704

705 [43] G.W. Chen, W.W. Lin, D.J. Lee, Capillary suction time (CST) as a measure of
706 sludge dewaterability, *Water Sci. Technol.* 34 (1996) 443–448.

707

708 [44] Z. Zhang, Y. Zhou, J. Zhang, S. Xia, S.W. Hermanowicz, Effects of short-time
709 aerobic digestion on extracellular polymeric substances and sludge features of waste
710 activated sludge, *Chem. Eng. J.* 299 (2016) 177–183.

711

712 [45] Y. Gu, L. Chen, J.-W. Ng, C. Lee, V.W.-C. Chang, C.Y. Tang, Development of
713 anaerobic osmotic membrane bioreactor for low-strength wastewater treatment at
714 mesophilic condition, *J. Memb. Sci.* 490 (2015) 197–208.

715

716 [46] Y. Zhao, L. Song, S.L. Ong, Fouling behavior and foulant characteristics of
717 reverse osmosis membranes for treated secondary effluent reclamation, *J. Memb. Sci.*
718 349 (2010) 65–74.
719