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Long-term implications of feed energy source in different genetic types of reproductive rabbit females. II. Immunologic status.

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Short title: Diet x genetic in does: Immunologic status

Papers in this same series:

Long-term implications of feed energy source in different genetic types of reproductive rabbit females. I. Resources acquisition and allocation

Long-term implications of feed energy source in different genetic types of reproductive rabbit females. III. Fitness and productivity

Abstract

Genetic selection and nutrition management have played a central role in the development of commercial rabbitry industry over the last few decades, being able to affect productive and immunological traits of the animals. However, the implication of different energy sources in animals from diverse genetic lines achieving such evolutionary success remains still unknown. Therefore, in this work, 203 female rabbits housed and bred in the same conditions were used from their first artificial insemination until their fifth weaning. The animals belonged to three different genetic types diverging greatly on breeding goal s (H line, hyper-prolific (n = 66); LP line, robust (n = 67) and R line, selected for growth rate (n = 67), and were assigned to two experimental diets, promoting major differences in energy source (cereal starch or animal fat). The aims of this work were to: (1) characterise and describe blood leukocyte populations of three lines of rabbit does in different physiological stages during their reproductive period: first artificial insemination, first weaning, second parturition and fifth weaning; and (2) study the possible influence of two different experimental diets on the leukocyte populations in peripheral blood. Flow cytometry analyses were performed on blood samples taken from females at each different sampling stade. Lymphocyte populations at both weanings were characterised by significantly lower counts of total, $CD5^+$ and $CD8^+$ lymphocytes (–19.8, –21.7 and –44.6%; $P<0.05$), and higher counts of monocytes and granulocytes (+49.2 and +26.2%; $P<0.05$) than in the other stages. Females had higher blood counts of lymphocytes B, $CD8^+$ and $CD25^+$ and lower counts of $CD4^+$ at first than at fifth weaning (+55.6, +85.8, +57.5, –14.5%; $P<0.05$). G/L ratio was higher at both weanings ($P<0.05$), and $CD4^+/CD8^+$ ratio increased

progressively from the 1AI to the 5W (P<0.001). Regarding the effect of genetic type in blood leukocyte counts, LP animals presented the highest counts for total, B, $CD5^+$ and $CD8^+$ lymphocytes (+16.7, +31.8, +24.5 and +38.7; P<0.05), but R rabbits showed the highest counts for monocytes and granulocytes (+25.3 and +27.6; P<0.05). The type of diet given during the reproductive life did not affect the leukocyte population counts. These results indicate that there are detectable variations in the leukocyte profile depending on the reproductive stage of the animal (parturition, weaning or none of them). Moreover, foundation for reproductive longevity criteria allows animals to be more capable of adapting to the challenges of the reproductive cycle from an immunological viewpoint.

Keywords: immunological challenge, genetic type, flow cytometry, animal fat, cereal starch.

Implications

The description of the normal immunological variations in rabbit does from three very common commercial genetic lines during their reproductive life entails an important and basic step in order to perform further comprehensive studies on how these animals may develop different strategies to successfully overcome productive and reproductive challenges. Moreover, the assignment of an appropriate nutrition is a critical issue in the rabbit industry and major efforts and resources are currently focused on this field. Therefore, finding out if different energy sources influence the ability of these animals to organise effective immunological responses is of great interest for farmers and researchers.

Introduction

Relevant advances in genetic selection, reproductive management and feeding systems (Pascual, 2010) have allowed the rabbitry industry to evolve greatly in the last few decades. Genetic selection by productive longevity has resulted in an effective increase in the number of long-living animals, able to maintain high reproductive performance throughout their productive life. However, a long life for these animals is burdened with challenges and their ability to survive is grounded on the maintenance of a reliable and stable health and accurate management of body resources in constant, unpredictable variation. Indeed, the evolutionary success achieved by genetic types founded by productive longevity is mainly attributable to their ability to successfully overcome productive, environmental and immunological challenges (Pascual *et al.*, 2013). So, animals from these genetic lines have been considered more robust than the rest (García-Quirós *et al.*, 2014), understanding the concept of robustness in farm animals as defined by Knap (2005): 'The ability to combine a high production potential with resilience to stressors, allowing for unproblematic expression of a high production potential in a wide variety of environmental conditions'. In fact, these animals are not only able to adapt to short-term challenges, but can also integrate their adaptations over time to adapt to long-term patterns (e.g. temperature stress, intense reproductive rhythm or recurring pathogens). However, it is uncertain what the mechanisms are that evolution has reached in these animals to address their disparate needs. Previous studies point to the metabolism (Savietto *et al.*, 2015) and immunity (Guerrero *et al.*, 2011; Ferrian *et al.*, 2012) as the main factors responsible for organising

effective responses that allow them to maintain high reproductive performance during successive lactations.

Notwithstanding the evidenced impact of genetic selection on the robustness of the animal, it has also been suggested that the use of a fat-enriched lactation diet could contribute to improving the maturity of the immune system of young rabbits at weaning (García-Quirós *et al.*, 2014) and, therefore, their general health status towards the growing period. In this conceptual framework, this is the second of three consecutive papers (see companion papers Arnau-Bonachera *et al.*, 2017a and 2017b) that were designed to provide a context in which animals from three different genetic types and fed with two distinct diets - but housed and bred in the same conditions- could be systematically studied and compared throughout their reproductive life (from the first artificial insemination to the sixth parturition). In that context, this paper is mainly focused on the study of the immunological status of the animals. Therefore, the specific aims of this work were to (1) characterise and describe blood leukocyte populations and their evolution during the abovementioned reproductive period of three lines of rabbit does differing greatly in animal type; and (2) study the possible influence of two different experimental diets, promoting major differences in the energy source (fat or starch), on the leukocyte populations in peripheral blood.

Material and Methods

Animals

The Committee of Ethics and Animal Welfare of the UPV approved this study. All animals were handled according to the principles of animal care published by Spanish Royal Decree 1201/2005 (BOE, 2005; BOE = Official Spanish State Gazette). The experiment involved a total of 203 female rabbits *(Oryctolagus cuniculus)* which were used from their first artificial insemination (AI) until their fifth weaning (from December 2012 to April 2013). Rabbit does belonged to three genetic types developed at the Institute for Animal Science and Technology of the Universitat Politècnica de València (UPV), differing greatly in breeding goals. Line H (n = 66), founded and selected by hyper-prolific criteria (Cifre *et al.*, 1998); line LP (n=67), characterised by a high robustness (Sánchez *et al.*, 2008; Pascual *et al.*, 2013); and line R (n =70), selected for growth rate during the fattening period (Estany *et al.*, 1992).

Diets

Two experimental diets were formulated and pelleted, according to the recommendations of De Blas and Mateos (2010) for reproductive rabbit does, promoting major differences in energy source. CS diet was prepared using cereal starch [237 g of starch and 21 g of ether extract (EE) per kg dry matter (DM)], whereas in the AF diet, part of the starch was replaced by animal fat (105 g of starch and 86 g of EE per kg DM). Nevertheless, both diets were isoenergetic and isoproteic [approx. 11.3 MJ of digestible energy (DE) and 126 g of digestible protein per kg of DM]. Further details of the diets and the

139 methodology used to characterise them can be found in Arnau-Bonachera *et*
140 *al.,* (2017)

141 *Experimental procedure*

142 Animals were housed under conventional environmental conditions (average
143 daily temperatures varying from 13.3 to 26.1 ºC), with an alternating cycle of 16
144 h of light and 8 h of darkness. At 19 weeks of age, all the rabbit females were
145 inseminated (with pooled semen from their respective line) and housed in
146 individual cages (700 x 500 x 320 mm) provided with a nest for litters from 28th
147 day of gestation. After the first parturition, all animals from the three genetic
148 types were randomly assigned to one of the reproductive diets. Until this point,
149 all the animals had received the same commercial diet for reproductive rabbit
150 does. Both experimental diets were provided *ad libitum* and the animals were
151 alternately allocated from within genetic type and reproduction diet throughout
152 the experimental farm. Litters were standardised to 8-9 kits at first parturition
153 and 9-11 onwards. Females were inseminated at 11 days postpartum (dpp) and
154 weaned at 30 dpp. Non-pregnant females were re-inseminated 21 days
155 afterwards, up to a maximum of three times. Blood samples were taken from
156 females at different physiological stages: first AI (1AI, at the start of the
157 reproductive life), first weaning (1W, potential immunological risk moment),
158 second parturition (2P, a moment described as immunologically critical, Ferrian
159 *et al.*, 2012) and fifth weaning (5W, same stage as first weaning but an ulterior
160 reproductive cycle). Diurnal variations in haematological parameters were
161 minimised by collecting blood at approximately the same time (9:00 h-10:00 h).

162 *Flow cytometry analysis*

Flow cytometry analysis was performed 1 h after sampling using 1 mL of peripheral blood drawn from the median artery of the ear, using vacuum tubes with EDTA. Prior to any other procedure, the white blood cell (WBC) count was determined using a haematology analyser (MEK-6410, Nihon Kohden, Japan). Then, blood was transferred to a 50 mL tube, in which 40 mL of ammonium chloride lysing solution at 4°C was added to isolate WBC. After 6 min of incubation in the dark, samples were centrifuged at 400 g for 5 min at room temperature. The supernatant was eliminated and the pellet was carefully resuspended in 1 mL of phosphate-buffered saline 1x (PBS). The density of the suspension was adjusted to 10^6 cells per mL by counting with Neubauer chamber. Primary monoclonal antibodies were added (Table 1), and incubated for 20 min at room temperature in the dark. Then, the pellet was washed with 1 mL of PBS, and centrifuged again in the same conditions mentioned above. Thereafter, secondary antibodies (Rat anti-mouse IgG 2a+b Phycoerythrin [VMRD, Inc. Exalpha] and Goat anti-mouse IgM: R-Phycoerythrin-human adsorbed [AbDSerotec]) were added, and incubated for 20 min at room temperature in the dark. One mL of PBS was added before running the flow cytometer. The outcome WBC suspensions were analysed in a Cytomics FC500 flow cytometer (Beckman Coulter, Brea, CA). The common leukocyte antigen CD14 and CD45 expression was used for the “lymphogate” setup as previously described (Jeklova *et al.*, 2007; Guerrero *et al.*, 2011). Calculation of total lymphocyte and respective subsets counts were performed as the product of WBC count and specific populations percentages, as described by Hulstaert *et al.* (1994) and Guerrero *et al.* (2011).

Statistical analysis

188 The asymmetrical distribution of the original data led to the logarithmic
189 transformation of data from all variables, except from the ratios G/L and
190 $CD4^+/CD8^+$, which were directly obtained from the counts (without logarithmic
191 transformation). Data from transformed variables were then analysed using a
192 mixed model (SAS Institute, 2002) including genetic type (H, LP, R), diet (AF,
193 CS), physiological stages (1AI, 1W, 2P, 5W) and their interactions as fixed
194 effects, and the permanent effect of each rabbit female (p) and the error term
195 (e) as random effects. Random effects were assumed to have an average of
196 zero and a variance of σ_p^2 for permanent, and σ_e^2 for the error term. This way, it
197 is possible to model variance among animals by using a compound symmetric
198 structure for the variance-covariance matrix of the residuals (R), when a
199 repeated measure experiment is performed. As diets were offered from the first
200 parturition on, when the effect of the diet was studied, first insemination data
201 (previous control to diet offering) was removed from the analysis.
202

Results

Table 2 shows the blood leukocyte population counts of all rabbit does, at the different physiological stages controlled from the first insemination to the fifth weaning. Lymphocyte populations at both weanings were characterised by lower counts of total, $CD5^+$ and $CD8^+$ (−19.8, −21.7 and −44.6%; $P<0.05$) and higher counts of monocytes and granulocytes (−49.2 and −26.2%; $P<0.05$) than in the other controls. Females had higher blood counts of lymphocytes B, $CD8^+$ and $CD25^+$ and lower of $CD4^+$ at first than at fifth weaning (+55.6, +85.8, +57.5, −14.5%; $P<0.05$). Although no great differences were found for leukocyte counts between first AI and second parturition, $CD25^+$ was higher for the latter (+64.8%; $P<0.05$). With reference to ratio G/L, it was higher at both weanings (on average 1.71 *vs.* 1.15 for the other controls; $P<0.05$), and the ratio $CD4^+/CD8^+$ was progressively increasing from the 1IA to the 5W ($P<0.001$).

Regarding effect of genetic type in blood leukocyte counts (Table 3), LP rabbit does presented the average highest counts for total, B, $CD5^+$ and $CD8^+$, (+16.7, +31.8, +24.5 and +38.7, respectively; $P<0.05$). This scenario relates mainly to the higher count of these lymphocyte populations at the second parturition of LP females (Fig. 1a, 1b, 1c and 1e). However, R rabbit does showed the highest counts for granulocytes (+27.6%; $P<0.05$). Granulocyte counts were always the highest for R females (Fig. 1h), and although H females showed a higher monocyte count at 1AI, values for R females were greater from first to fifth weaning (Fig. 1g). Moreover, R animals showed the highest G/L ratio, due to their greater G/L value at the 5W (2.54 *vs.* 1.38 on average for the other genotypes; $P<0.05$) (Fig. 2a). In addition, H females presented the highest $CD4^+/CD8^+$ ratio at 5^{th} weaning. Although no differences were observed at 1AI

228 (Fig. 2b), the $CD4^+/CD8^+$ ratio of H females increased progressively throughout
229 the period of study, reaching the highest differences at 5W (9.17 *vs.* 6.16 for the
230 other genotypes; $P<0.05$). Table 4 shows that the type of diet given during the
231 reproductive life did not affect the leukocyte population counts. However, two
232 interactions between the genetic type and the diet for total lymphocytes and
233 granulocytes were observed. Genetic type did not affect total lymphocyte counts
234 when fed with AF diet, but H rabbit does showed significantly lower counts
235 when fed with CS diet (Fig. 3a). Regarding the granulocyte counts, the lowest
236 values were obtained for LP females when animals were fed with AF and for H
237 females when fed with CS (Fig. 3b).

Discussion

The study of haematological parameters and lymphocyte subsets through flow cytometry analyses has been widely used to determine the physiological and pathological changes in the peripheral blood leukocyte subpopulations in different species. Specifically, in rabbits, there are several studies reporting these parameters as adequate indicators for the immunological state of animals of diverse ages and conditions: conventional or SPF animals, neonatal to pubescent rabbits, primiparous rabbit does and adult rabbits (Jeklova *et al.*, 2007; Jeklova *et al.*, 2009; Guerrero *et al.*, 2011).

It is well established that leukocyte subpopulations vary with aging. At early stages, newborns start their life with a competent, but still naïve immune system, in which protection provided by the immune mechanisms and by transferred maternal antibodies plays an important role (Kampen *et al.*, 2006). In rabbitry, the moment of first mating has frequently been identified as a crucial point in development of the young females. This is the last item of 'pure' data on the animal, a sign of the animal soma that is probably related to their productive potential. From this moment on, all their productive records will be conditioned by their reproductive history (Pascual *et al.*, 2013), and specific immune responses will be developed over time against different infectious, environmental or productive challenges. Therefore, all results obtained in this study at 1W, 2P and 5W are compared to a reference sampling control set at the age of first mating (1AI). This scenario allows us to compare the evolution of animals throughout their reproductive life (from 1AI to 5W), housed, fed and bred in the same conditions, aiming to obtain specific, measurable information about the immunological and productive traits of the same group of animals in

263 certain crucial stages. Studies on the evolution of the immune system indicate
264 that stress responses, immunity and inflammation are deeply interconnected
265 and constitute an integrated defence network capable of coping with most
266 stressors (Franceschi *et al.*, 2000; Larbi *et al.*, 2008). Even further, previous
267 studies suggest that immune aging profiles described in laboratory and
268 domestic mammals may generalise to more complex consequences and could
269 develop fitness costs under natural conditions (Nussey, *et al.*, 2012).
270 As previously reported (Wells *et al.*, 1999; Guerrero *et al.*, 2011; Ferrian *et al.*,
271 2012), the present study evidences that leukocyte populations varied
272 throughout the rabbit does' productive cycle, reaching different levels at the four
273 distinct control moments sampled. Therefore, it is worth discussing them one by
274 one. Firstly, it is interesting to analyse the productive and reproductive
275 conditions characterising each sampling moment. In that sense, animals at first
276 weaning are influenced by great challenging needs for the production of milk
277 and to be able to cope with their gestation, as they overlap both stages (milking
278 and gestation). As a consequence, they increase their feed intake, and show a
279 moderate level of mobilisation, similar to the observed in 1AI but lower than in
280 2P (see results shown in the first paper of this same series, by Arnau-
281 Bonachera *et al.*, 2017). In rabbit does, the risk of culling peaks during the two
282 first lactations, especially at the end of pregnancy (Rosell and de la Fuente,
283 2009). This period includes two of our moments of sampling: first weaning and
284 second parturition. Other species, such as dairy cows, are also more vulnerable
285 to infectious diseases around calving due to immune suppression during this
286 period (Meglia *et al.*, 2005).

287 In this sense, second parturition has been specifically described as a
288 physiological state that is especially challenging for rabbit does during their
289 reproductive life (Ferrian *et al.*, 2012), as it is not only a reproductive challenge
290 but also a crucial period of risk of infections and cellular and tissue damage.
291 However, at this point LP females show higher counts for most lymphocyte
292 populations (total, B, $CD5^+$, $CD4^+$ and $CD8^+$), the significant increase in $CD25^+$
293 (+64.8 %) being especially notable. All these changes may be related with the
294 immune system being more capable of adapting to the challenges of the
295 productive cycle in LP animals than in the other genetic types.
296 Particularly, $CD4^+CD25^+$ is a population of regulatory T cells (Tregs) which are
297 considered as T-activated cells, although there is still no clear consensus on the
298 definition of Tregs. It is known that these cells are essential for maintaining
299 peripheral tolerance, preventing autoimmune diseases and limiting chronic
300 inflammatory diseases (Chen *et al.*, 2016). These traits favour successfully
301 confronting different challenging physiological stages. However, regulatory
302 activity has also been described in T cells with low expression of CD25, which
303 means that high expression of CD25 itself is not enough to characterise all
304 Tregs (Dejaco *et al.*, 2006). Therefore, other specific markers are being used to
305 identify Tregs. In the last few years, FOXP3 has been established to be the
306 most specific marker of Tregs (Sakaguchi, 2005). Unfortunately, in this study
307 Tregs counts were determined by marking positive for CD25 T cells population,
308 but FOXP3 should have been used for a more specific determination of Tregs
309 prevalence. This fact is due to the limiting availability of commercial antibodies
310 against FOXP3 suitable to be used in rabbits' flow cytometry.

311 Following with the comparison between genetic lines at second parturition, it is
312 worth to mention that, due to the increase in the number of lymphocytes but not
313 of granulocytes, the G/L ratio was lower for LP animals. As numbers of
314 neutrophils and lymphocytes oscillate in opposite directions under stressful
315 conditions, researchers have often considered the ratio of one to the other as a
316 composite measure of the stress response (Davis *et al.*, 2008). G/L ratio is a
317 stress indicator that is known to increase in the presence of various stressors,
318 diseases or infections (Davis *et al.*, 2008). In other species, and even in birds, it
319 has also been shown that parents that make intense reproductive effort have
320 high G/L values (Horak *et al.* 1998). Moreover, high G/L ratios have been
321 associated in birds with susceptibility to infection (Al-Murrani *et al.*, 2002) or low
322 survival to the next breeding season (Kilgas *et al.*, 2006). These associations
323 can make G/L ratios valuable for predicting future problems in both populations
324 and individuals. Although leukocyte profiles do not indicate the number of
325 granulocytes or lymphocytes that are available in reserve in other body
326 compartments, or how many would be released or redistributed in response to a
327 stress or infectious agent (Davis *et al.*, 2008), the fact that seems clear is that
328 rabbit does from LP line reach the 2P in an immunologically less stressful
329 situation than the other lines.
330 Arnau-Bonachera *et al.* (2017) reported in the first paper of this series a higher
331 mobilisation throughout the first reproductive cycle, reaching the 2P in better
332 conditions and suffering less stress during the following cycle. In the same
333 sense, the immunological data described in this work back up the hypothesis
334 that animals from LP lines are more robust than the other genetic types, as they
335 are able to adapt to reproductive challenges by using their body reserves more

336 accurately. Moreover, they seem to be able to manage their body reserves as if
337 they were predicting future needs. In that sense, the metabolic profile of LP line
338 in 2P is characterised by a higher level of glucose and a lower level of NEFAs,
339 showing great differences in the ratio glucose/NEFAs when compared to H and
340 R females at 2P. In fact, T lymphocytes specifically require glucose uptake for
341 cell survival, size, activation and cytokine production, and they consume it at
342 high rates in a function-dependent manner (Maclaver *et al.*, 2008). The close
343 association between glucose metabolism and lymphocyte function has been
344 suggested to introduce the possibility of several pathologies resulting from the
345 inability of these cells to meet their nutrient demands under a given condition
346 (Wasinki *et al.*, 2014). In this group of animals, the direct correlation between
347 the level of glucose (Arnau-Bonachera *et al.*, 2017a) and the number of
348 lymphocytes is observed regardless of the genotype and the temperature
349 (+0.23±0.11; $P<0.05$). Therefore, higher counts of total, $CD5^+$ and $CD8^+$
350 lymphocytes in LP females at 2P seems to be associated to the peak of glucose
351 shown at that moment and not to any of the other factors included in Arnau-
352 Bonachera *et al.* (2017a). In other words, LP rabbit does managed to have
353 higher levels of glucose available at the most challenging time of their
354 reproductive life, which implies a guaranteed supply of nutrients for the
355 activation and function of lymphocytes. This mechanism of adaptation may be
356 suggested as one of the factors contributing to increase the robustness of these
357 animals, which may consequently be likely to live longer, although longevity is
358 not their criteria of selection. Our data reinforce the hypothesis that the animals
359 from a line founded by screening for reproductive longevity (LP line), under
360 normal favourable breeding conditions, develop a greater immunological ability

361 to confront reproductive challenges and to confer animals a more robust nature
362 (Ferrian *et al.*, 2012; García-Quirós *et al.*, 2014).
363 Regarding weaning, differences between lymphocyte populations from the first
364 and the fifth weaning were detected. Taking into account that both sampling
365 moments represent the same type of reproductive challenge (weaning), we
366 hypothesised that the effect of the aging may be one of the main factors that
367 caused these variations. Some changes regarding aging in the leukocyte
368 populations have been described in other species. One of the most reported
369 data items is the $CD4^+/CD8^+$ ratio, which in our study is decreased in animals at
370 fifth weaning compared to first weaning, as it has been described as a normal
371 effect of aging in other species (i.e.: mice, Callahan *et al.*, 1993; cattle, Ayoub
372 and Yang, 1996; humans, Castelo-Branco and Soveral, 2014). In this work, the
373 H line showed the highest increase in the $CD4^+/CD8^+$ ratio at 2P, which can be
374 considered as one of the signs related with an earlier aging of their immune
375 system (see also the third paper of this same series, by Arnau-Bonachera *et al.*,
376 2017).
377 In reference to the interaction between genetic type-diet and leukocyte
378 populations, few remarkable data were found. The only statistically significant
379 data observed were the decrease of total lymphocyte in H animals with CS diet,
380 and the increase of granulocytes in LP females fed with CS diet. Both facts are
381 probably related to the way of managing their body resources. H rabbit does are
382 very dependent of their body condition, as they need to be able to feed very
383 large litters. However, excessive fat deposits can also be counterproductive, as
384 they diminish fertility and increase mortality (Arnau-Bonachera *et al.*, 2017a and
385 2017b). On the contrary, LP animals do not depend on their body condition as

much as H animals, mainly because they have developed several different mechanisms to modulate their responses and keep the energy homeostasis balance without reducing their fertility, while being able to maintain most of their litter alive until weaning.

Despite the significant statistical nature of our data from the study, we are aware that sometimes the variation in the values of health and immunological traits combined with productive and reproductive parameters are difficult to interpret, as the meaningfulness of the changes in particular values is largely unknown. Moreover, it must be taken into account that aging is a complex and multi-factorial process, and defective immune responses in aged and multiparous animals are likely to be caused by the interaction of accumulated weaknesses throughout the immune system rather than to one individual aspect of a single immune cell type function (Plowden *et al.*, 2004a and 2004b). However, the observed relationships, though suggestive, are not able to firmly indicate a causal link between some aspects of the immunological condition during the reproductive life of animals from three different genetic types. Therefore, further research would be important in order to establish a correlation between this type of data and future survival probability. In fact, similar hypotheses have been previously considered, suggesting that age-dependent differences in immunity may become targets for natural selection in other species of mammals (Nussey *et al.*, 2012).

Conclusions

The present study has evidenced that leukocyte populations vary throughout the rabbit doe's productive cycle. According to our results, oscillations were different depending on the genetic line and the stage of the reproductive cycle.

However, the interaction between genetic type and diet did not cause important changes in leukocyte populations. Animals founded for high robustness (LP line) showed greater ability to adapt immunologically to the reproductive challenges than those selected by hyper-prolificacy (H line) or by growth rate (R line). Differences among lines were especially remarkable at a critical physiological moment such as the second parturition. Although genetic, management and nutritional strategies developed over the last few decades have brought valuable advances in the rabbit industry, it seems that they have also caused undesired consequences affecting, among other factors, the ability of the animals to maintain a stable and competent immunological status throughout their productive life.

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Table 1 *Monoclonal antibodies used for the flow cytometry analysis of this study*

Monoclonal antibody	Iso.	Spec.	Cell labelling	Clone	Ref.	Comp.
Mouse anti-rabbit T lymphocytes: FITC[1]	IgG1	CD5	T cell	KEN-5	Kotani *et al*. (1993a)	Abd Serotec
Mouse anti-rabbit α-pan B	IgM	IgM	B cell	MRB143A	Davis and Hamilton (2008)	VMRD Inc.
Mouse anti-rabbit CD4	IgG2a	CD4	T cell subset	KEN-4	Kotani *et al*. (1993a)	Abd Serotec
Mouse anti-rabbit α-CD8	IgG2a	CD8	T cell subset	ISC27A	Davis and Hamilton (2008)	VMRD Inc.
Mouse anti-rabbit CD25	IgG2b	CD25	Activated T cells	KEI-ALPHA1	Kotani *et al*. (1993b)	Abd Serotec
Mouse anti-human CD14: FITC	IgG2a	CD14	Monocytes & granulocytes	TÜK4	Jacobsen *et al*. (1993)	Abd Serotec
Mouse anti-rabbit α-CD45	IgM	CD45	All leukocytes	ISC76A	Davis and Hamilton (2008)	VMRD Inc.

Iso. = Isotype; Spec. = Specificity; Ref. = References; Comp. = Company

[1] Clon KEN-5 recognises rabbit T lymphocytes and immunoprecipitates. This antibody recognises rabbit CD5, but does not bind to rabbit CD5 transfectants. Known rabbit CD5 antibodies also show binding to most B lymphocytes, which are not labelled by this clone (information obtained from datasheet).

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Table 2 *Evolution of the leukocyte counts in the blood of rabbit females (least square mean; log_{10} 10^6/L)*

	Stade (S)[1]					
	1AI	1W	2P	5W	SEM[2]	P-value
n	203	130	96	65		
Total lymphocytes (L)	3.47^{b}	3.38^{a}	3.47^{b}	3.38^{a}	0.018	<0.001
Lymphocytes B	1.41^{bc}	1.43^{c}	1.31^{ab}	1.24^{a}	0.045	0.002
Lymphocytes T CD5^{+}	3.30^{b}	3.17^{a}	3.27^{b}	3.18^{a}	0.019	<0.001
CD4^{+}	3.04^{b}	2.97^{a}	3.07^{b}	3.04^{b}	0.019	<0.001
CD8^{+}	2.69^{c}	2.49^{b}	2.56^{c}	2.22^{a}	0.025	<0.001
CD25^{+}	1.10^{a}	1.26^{b}	1.31^{b}	1.07^{a}	0.040	<0.001
Monocytes	2.37^{a}	2.58^{b}	2.44^{a}	2.58^{b}	0.026	<0.001
Granulocytes (G)	3.41^{a}	3.55^{b}	3.46^{a}	3.53^{b}	0.021	<0.001
G/L[3]	1.02^{a}	1.65^{b}	1.27^{a}	1.76^{b}	0.113	<0.001
CD4^{+}/CD8^{+}[3]	2.46^{a}	3.22^{b}	3.43^{b}	7.21^{c}	0.174	<0.001

n: Number of records per trait.

[1] Stade (S): 1AI: at the first artificial insemination; 1W: at the weaning of the first lactation; 2P: at the second parturition; 5W: at the weaning of the fifth lactation.

[a,b,c] Means in a row not sharing superscripts significantly differ at $P<0.05$.

[2] Pooled standard error of means.

[3] G/L and CD4^{+}/CD8^{+} ratios were directly obtained from the counts (no logarithmic transformation).

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Table 3 *Effect of genetic type on the leukocyte counts in the blood of rabbit females (least square mean; log_{10} 10^6/L)*

	Genetic type (G)[1]				P-value	
	H	LP	R	SEM[2]	G	G×S[3]
n	155	181	156			
Total lymphocytes (L)	3.39[a]	3.46[b]	3.43[ab]	0.016	0.010	0.005
Lymphocytes B	1.30[a]	1.42[b]	1.31[a]	0.036	0.027	0.118
Lymphocytes T CD5$^+$	3.22[a]	3.28[b]	3.19[a]	0.017	<0.001	0.006
CD4$^+$	3.04[b]	3.07[b]	2.98[a]	0.017	0.002	0.016
CD8$^+$	2.44[a]	2.58[b]	2.46[a]	0.024	<0.001	0.001
CD25$^+$	1.20[ab]	1.12[a]	1.23[b]	0.033	0.052	0.111
Monocytes	2.48[ab]	2.45[a]	2.55[b]	0.025	0.016	0.002
Granulocytes (G)	3.45[a]	3.46[a]	3.57[b]	0.019	<0.001	0.530
G/L[4]	1.35[a]	1.25[a]	1.69[b]	0.102	0.006	<0.001
CD4$^+$/CD8^{+4}	4.98[b]	3.47[a]	3.86[a]	0.178	<0.001	<0.001

n: Number of records per trait.

[1] Genetic type (G): line H, founded by litter size at birth and selected by litter size at weaning during 17 generations; line LP, founded by reproductive longevity criteria by selecting females from commercial farms that had a minimum of 25 parturitions with more than 7.5 kits born alive per parity and then selected by litter size at weaning for 7 generations; line R, founded and selected during 25 generations by average daily gain from the 4th to the 9th week of life.

[a,b,c] Means in a row not sharing superscripts significantly differ at $P<0.05$.

[2] Pooled standard error of means.

[3] S: Stade (see Table 2).

[4] G/L and CD4$^+$/CD8$^+$ ratios were directly obtained from the counts (no logarithmic transformation).

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Table 4 *Effect of diet on the leukocyte counts in the blood of rabbit females (least square mean; log_{10} 10^6/L)*

	Diet (D)[1]			P-value		
	AF	CS	SEM[2]	D	D×S[3]	G×D[4]
n	222	211				
Total lymphocytes (L)	3.41	3.43	0.025	0.615	0.332	0.005
Lymphocytes B	1.28	1.33	0.028	0.287	0.316	0.595
Lymphocytes T $CD5^+$	3.22	3.22	0.015	0.976	0.276	0.066
$CD4^+$	3.04	3.04	0.015	0.964	0.681	0.090
$CD8^+$	2.42	2.44	0.020	0.576	0.403	0.259
$CD25^+$	1.22	1.24	0.038	0.811	0.288	0.564
Monocytes	2.58	2.53	0.022	0.193	0.639	0.110
Granulocytes (G)	3.53	3.53	0.016	0.775	0.553	0.004
G/L[5]	1.60	1.62	0.105	0.929	0.386	0.737
$CD4^+/CD8^+$[5]	4.85	4.85	0.166	0.254	0.253	0.577

n: Number of records per trait.

[1] Diet (D): CS, mainly based on cereal starch (247 g of starch and 21 g of ether extract (EE) per kg dry matter (DM)); AF, mainly based on animal fat (104 g of starch and 85 g of EE per kg DM).

[2] Pooled standard error of means.

[3] S: Stade (see Table 2).

[4] G: Genetic type (see Table 3).

[5] G/L and $CD4^+/CD8^+$ ratios were directly obtained from the counts (no logarithmic transformation).

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Figure 1 Interaction Genetic type × Control for the (a) total lymphocytes (b) lymphocytes B (c) lymphocytes T $CD5^+$, (d) $CD4^+$, (e) $CD8^+$, (f) $CD25^+$, (g) monocytes, and (h) granulocytes counts in blood of reproductive rabbit females. Genetic type: (☐ line H, characterised by hyper-prolificacy; ▢ line LP, characterised by functional hyper-longevity, and ■ line R, characterised by daily gain). ^{a,b,c,d,e} Means for a genetic type within a stade not sharing superscripts significantly differ at $P<0.05$.

Figure 2 Interaction Genetic type × Control for the (a) ratio Granulocytes/Lymphocytes and (b) ratio CD4+/CD8+ in the blood of reproductive rabbit females. Genetic type: (☐ line H, characterised by hyper-prolificacy; ▢ line LP, characterised by functional hyper-longevity, and ■ line R, characterised by daily gain). ^{a,b,c,d} Means for a genetic type within a stade not sharing superscripts significantly differ at $P<0.05$.

Figure 3 Interaction Genetic type × Diet for the (a) total lymphocytes and (b) granulocytes counts in blood of reproductive rabbit females. Genetic type: (☐ line H, characterised by hyper-prolificacy; ▢ line LP, characterised by functional hyper-longevity, and ■ line R, characterised by daily gain). Diet: CS, mainly based on cereal starch; AF, mainly based on animal fat. ^{a,b,c} Means for a genetic type within a diet type not sharing superscripts significantly differ at $P<0.05$.

Figure 1

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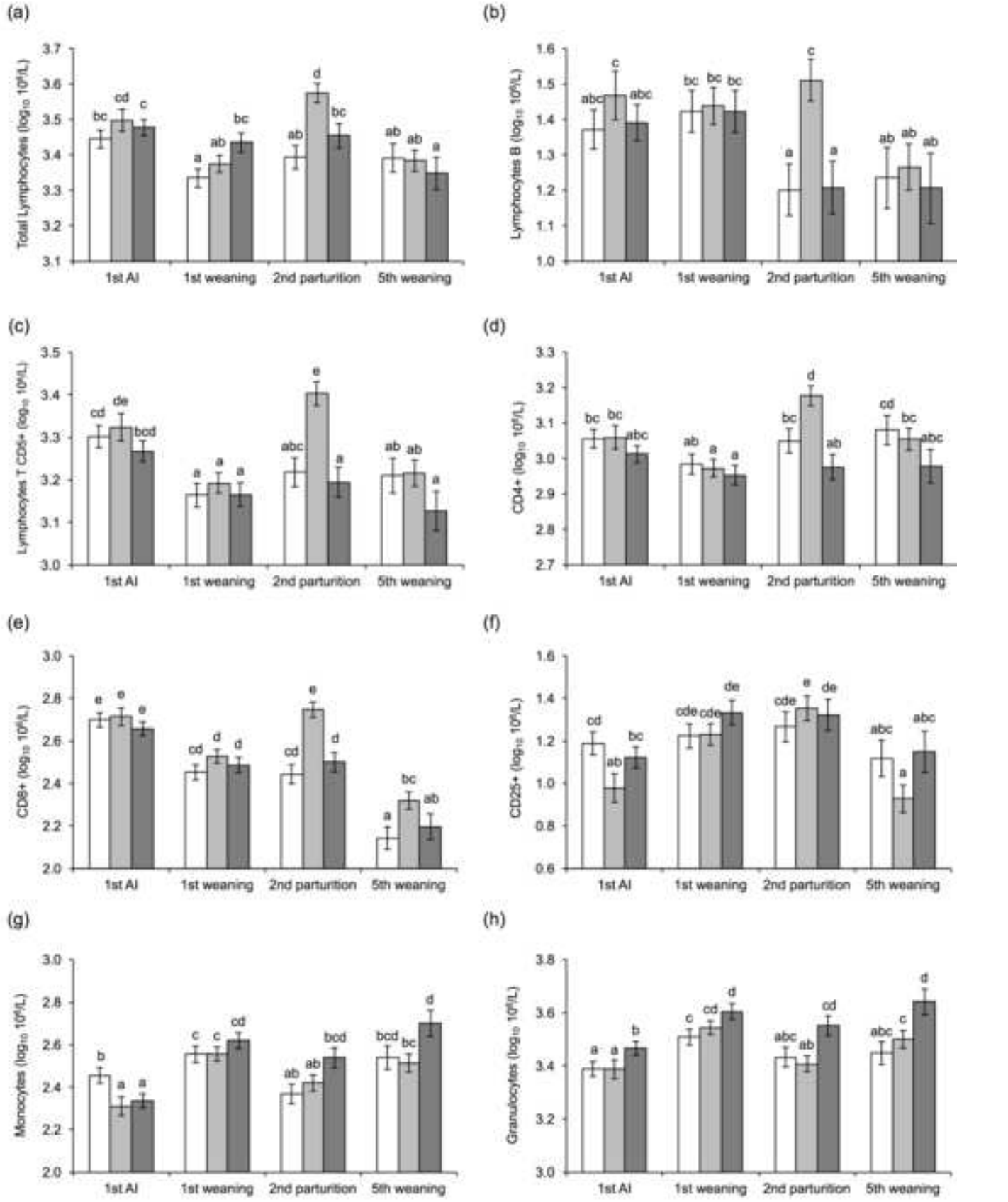

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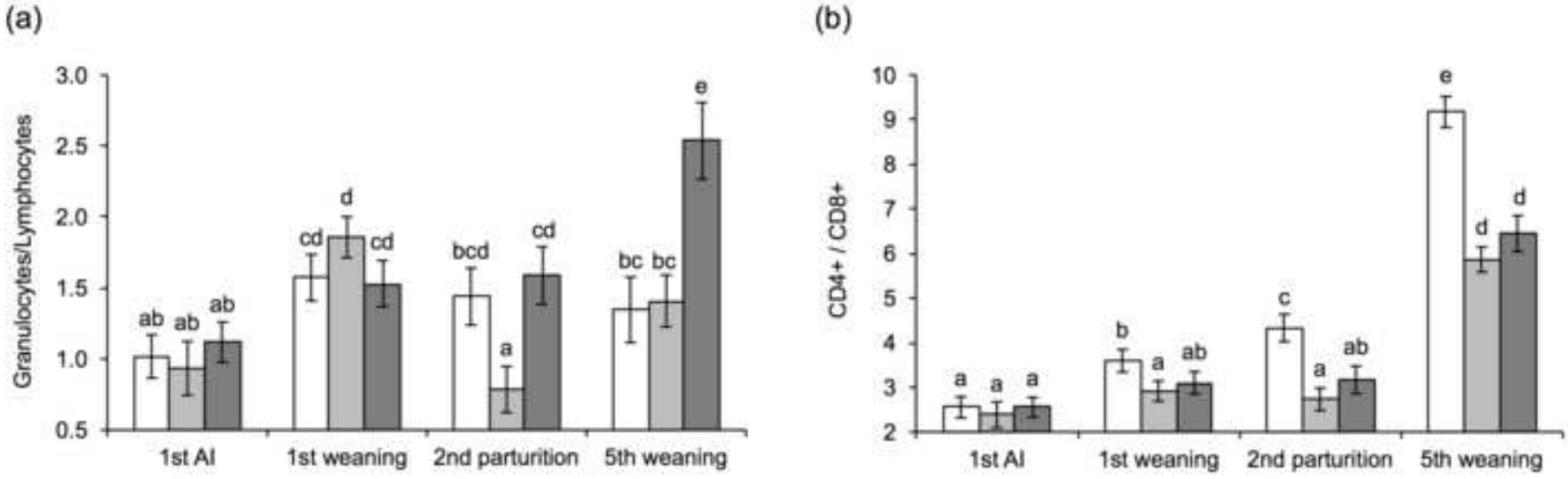

Figure 3

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