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Additional Information

**Indirect calculation of monoclonal antibodies in nanoparticles using the radiolabeling process with Technetium 99 metastable as primary factor:
alternative methodology for the entrapment efficiency**

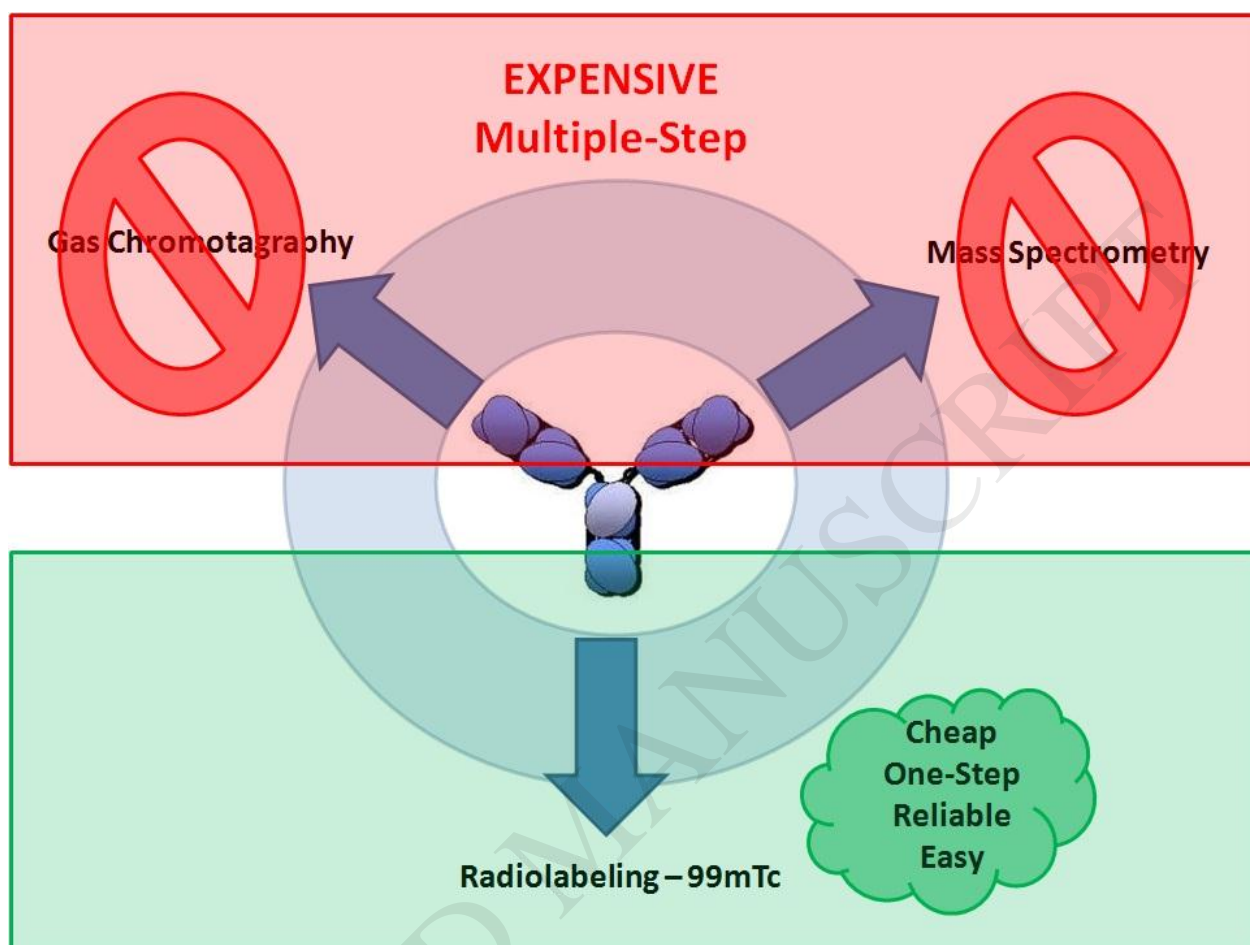
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Graphical Abstract



HIGHLIGHTS

- Novel methodology for the development of magnetic core mesoporous silica
- Radiolabeling process of monoclonal antibodies with ^{99m}Tc
- Efficient methodology to calculate the Incorporation of monoclonal antibodies into nanoparticles by radiolabeling process

Abstract

The use of monoclonal antibodies (Mab) in the current medicine is increasing. Antibody-drug conjugates (ADCs) represents an increasingly and important modality for treating several types of cancer. In this area, the use of Mab associated with nanoparticles is a valuable strategy. However, the methodology used to calculate the Mab entrapment, efficiency and content is extremely expensive. In this study we developed and tested a novel very simple one-step methodology to calculate monoclonal antibody entrapment in mesoporous silica (with magnetic core) nanoparticles using the radiolabeling process as primary methodology. The magnetic core mesoporous silica were successfully developed and characterized. The PXRD analysis at high angles confirmed the presence of magnetic cores in the structures and transmission electron microscopy allowed to determine structures size (58.9 ± 8.1 nm). From the isotherm curve, a specific surface area of $872 \text{ m}^2/\text{g}$ was estimated along with a pore volume of $0.85 \text{ cm}^3/\text{g}$ and an average pore diameter of 3.15 nm . The radiolabeling process to proceed the indirect determination were well-done. Trastuzumab were successfully labelled ($>97\%$) with Tc-99m generating a clear suspension. Besides, almost all the Tc-99m used (labelling the trastuzumab) remained trapped in the surface of the mesoporous silica for a period as long as 8 hours. The indirect methodology demonstrated a high entrapment in magnetic core mesoporous silica surface of Tc-99m-trastuzumab. The results confirmed the potential use from the indirect entrapment efficiency methodology using the radiolabeling process, as a one-step, easy and cheap methodology.

Keywords: nanoparticles, Magnetic Core Mesoporous Silica Nanoparticles doped with Trastuzumab and labeled with Tc-99m, cancer, oncology, smart device.

1. Introduction

The use of monoclonal antibodies (Mabs) in the current medicine is increasing each day. Monoclonal antibody-based treatment is used in a great variety of disease, with especial attention in oncology [1-3]

Antibody-drug conjugates (ADCs) are monoclonal antibodies attached to biologically active drugs by chemical linkers with labile bonds. It represents an increasingly and important modality for treating several types of cancer. The impact of ADCs in this field is due the exquisite specificity of antibodies that deliver the conjugated cytotoxic agent to targeted tumor cells, thus reducing the systemic toxicity associated with traditional chemotherapeutic treatments. ADCs are distinguished on the basis of the drug, linker, and also the amino acid residue of attachment on the antibody [4-6]. In the field of ADC, the use of Mab associated with nanoparticles is very common [7-9]. In this specific ADC case the calculation of Mab entrapment efficiency is of vital importance and in many cases requires a specific and expensive methodology.

The primary method to evaluate the mAb purification can be made by Protein A chromatography, followed by two or three subsequent chromatographic polishing steps [10]. In this case the chromatographic column continues to be favored, and has been the focus of continuous improvements

through development of higher capacity resins and use of mixed-mode sorbents. Another methodology involves mass spectroscopy or mass spectroscopy-associated liquid chromatography [11,12]. In both cases, the methodology used to calculate the Mab entrapment, efficiency and content is extremely expensive. In this study we developed and tested a novel and very simple one-step methodology to calculate monoclonal antibody entrapment efficiency in mesoporous silica nanoparticles using the radiolabeling process as primary methodology.

2. Materials and methods

2.1. Preparation of magnetic core mesoporous silica nanoparticles (MSN)

2.1.1. Reagents and materials

Iron(III)chloride hexahydrate, iron(II)tetrachloride hexahydrate, oleic acid, hexadecyltrimethylammonium bromide (CTAB) and tetraethyl orthosilicate (TEOS) were purchased from Sigma. Ammonia solution (32%), ethanol and ethyl acetate were purchased from Scharlau. Chloroform was obtained from Acros Organics. Distilled water was used in all reactions.

2.1.2. Synthesis of oleate-coated iron oxide nanoparticles

Iron oxide nanoparticles (Fe_3O_4 magnetite nanocrystals) were obtained by a modified coprecipitation method [1]. Briefly, 12 g of iron(III)chloride hexahydrate were mixed with 4.9 g of iron(II)chloride tetrahydrate in 50 ml of water at 80 °C

under a flow of argon and mechanical stirring. Ammonia solution 32% (19.53 ml) was carefully added and the mixture turned completely dark. Oleic acid (2.13 ml) was added after 30 min and the reaction was left stirring at 80 °C for another 90 min. The reaction was cooled down and centrifuged at 9500 rpm for 10 min. The resulting black precipitate was washed three times with distilled water and three times with ethanol and then dried under vacuum overnight. In order to prevent their oxidation, the oleate-coated iron oxide nanoparticles were kept in chloroform giving a dark brown ferrofluid.

2.1.3. Synthesis of magnetic core MSNs

In a typical procedure, 100 mg of CTAB were dissolved in 10 ml of water, followed by addition of 0.74 ml of the ferrofluid (8.88 mg/ml). The mixture was placed in a probe sonicator (Branson 450 Sonifier) for 2 min, giving an oil-in-water emulsion. Then, the mixture was heated to 65 °C to evaporate the chloroform and achieve an effective phase transfer from chloroform to water. The resulting transparent aqueous suspension was added to a solution of 30 ml of water and 0.548 ml of ammonia (32%), which was then, heated up to 75 °C. Then, 0.5 ml of tetraethyl orthosilicate (TEOS) was added dropwise followed by addition of 3 ml of ethyl acetate. The reaction was stirred at 350 rpm and 75 °C during 3 h. Then, the reaction mixture was placed on an ice bath and the nanoparticles were collected by centrifugation (9500 rpm, 10 min). Afterward, the sample was washed with ethanol twice and dried under vacuum overnight. The final magnetic core MSNs were calcined in air at 550 °C for 5 h.

2.2. Characterization of magnetic core MSNs

2.2.1. Powder X-ray diffraction

The synthesised materials were characterised by powder X-ray diffraction (PXRD), transmission electron microscopy (TEM) and N₂ adsorption-desorption analysis. PXRD measurements were obtained using a Bruker AXS D8 Advance diffractometer equipped with CuK α radiation and working at 40 kV/40 mA. PXRD measurements were performed at high angle ($2\theta = 15^\circ - 68^\circ$) and low angle range ($2\theta = 1.3^\circ - 8.3^\circ$).

2.2.2. Transmission electron microscopy

TEM images were taken on a 100 kV JEOL JEM-1010 microscope operated with AMT image capture engine software. TEM samples were prepared by adding 10 μ l of nanoparticles suspended in distilled water onto carbon-coated copper grids. The statistical analysis of the data obtained from TEM images was performed using Origin Pro software.

2.2.3. N₂ adsorption-desorption

N₂ adsorption-desorption measurements were conducted in a TriStar II Plus surface area and porosity analyzer from Micromeritics. The specific surface area of the material was determined from the adsorption-desorption isotherm by applying the BET (Brunauer-Emmett-Teller) model. The pore volume and average pore size was estimated by using the BJH (Barrett-Joyner-Halenda) model.

2.3. Doping magnetic core mesoporous silica nanoparticles with trastuzumab

Magnetic core mesoporous silica nanoparticles were loaded with trastuzumab. In order to get the loaded magnetic core mesoporous silica nanoparticles, 200 µg (200 µg/1 mL) of Herceptin® (trastuzumab) was stirred (24 hrs) at room temperature with 100 µg of magnetic core mesoporous silica nanoparticles. After this period the magnetic core mesoporous silica nanoparticles soaked with trastuzumab solution was dried under low pressure at 30°C temperature for 12 hours, until completely dry. The dried powder containing solely magnetic core mesoporous silica nanoparticles loaded with trastuzumab were individualized and set aside for later labeling with Tc-99m. The calculation to determine the amount of trastuzumab necessary to completely dope the magnetic core mesoporous silica nanoparticles was done considering the size of the magnetic core mesoporous silica nanoparticles (100 nm) applied to the circumference volume (Cv) equation (1):

$$\text{Equation 1: } Cv = \pi r^3$$

Where: Cv: circumference volume

π : constant

R: Radius of the circumference

In this case, we have that the weight of one single magnetic core mesoporous silica nanoparticle is about: 1×10^{-15} g. Thus, in 100 µg of magnetic core mesoporous silica nanoparticles we have approximately 10^{12-13} nanoparticles with a surface area approximately 872 m²/g. This means that using an excess of Mab (200 µg) that has a molar mass of 145.531,5 g mol⁻¹ we have a ratio of 3:1 trastuzumab/magnetic core mesoporous silica nanoparticles.

2.4. Trastuzumab labeling process with Tc-99m

The labeling process was done by the direct radiolabeling process as described previously by Pascual et al. [13], Cerqueira-Coutinho et al. [14] and Rosa et al. [15]. In this methodology we used 150 µg of trastuzumab. Briefly, 100 µCi (approximately 300 µL) of Tc-99m was incubated with a stannous chloride (SnCl₂) solutions (80 µg/mL) (Sigma-Aldrich) for 20 minutes at room temperature. Then this solution was incubated with 150 µg trastuzumab for another 10 minutes in order to label their structures.

2.5. Quality control of the labeling process with Tc-99m

In order to confirm the efficiency of the trastuzumab labeling process, paper chromatography was done using Whatman paper n^o 1 using 2 µl of the labeled-nanoparticle and acetone (Sigma-Aldrich) as mobile phase. The radioactivity of the strips was verified in a γ-counter (Perkin Elmer Wizard® 2470, Shelton, CT City, State).

2.6. Indirect entrapment efficiency calculation – Tc-99m EE%

To perform the entrapment efficiency of the trastuzumab into the magnetic core mesoporous silica nanoparticles we performed the Tc-99m entrapment efficiency. In this direction, we previously labeled the trastuzumab with Tc-99m and then, doped the magnetic core mesoporous silica nanoparticles as described in the section: doping magnetic core mesoporous silica nanoparticles with radiolabeled trastuzumab (Tc-99m-trastuzumab).

2.7. Planar imaging from chromatographic system

Planar images from the chromatographic system using Tc-99m-trastuzumab (3.7 MBq in 0.2 mL) were done in triplicate. The images were integrated for 5 min centered at 140 KeV, with a Millennium Gamma Camera (GE Healthcare, Cleveland, USA), using a 15% window. The images were processed using OsiriX software.

3. Statistical analysis

Statistical analyses were performed using Origin Pro 8 (OriginLab, USA) software. Results are shown as means \pm standard deviation (S.D.). P-values less than 0.05 were considered significant.

4. Results and discussion

4.1. Characterization of magnetic core MSNs

The structure periodicity of the mesoporous material was confirmed by PXRD, which showed a sharp peak at the low-angle region for both as-made (**S0-1**) and calcined (**S0-2**) magnetic core MSNs (Figure 1). The slight shift of the peak to higher angles indicates shrinkage of the silica matrix due to the condensation of silanol groups during the calcination process. PXRD analysis at high angles confirmed the presence of magnetic cores within the structure (see inset in Figure 1).

The mesoporous structure of **S0-2** magnetic core MSNs was also analysed by transmission electron microscopy and the size of the primary nanoparticles was determined by image analysis (58.9 ± 8.1 nm, $n = 100$). The data was

represented in a histogram, which shows the particle size distribution of the **S0-2** nanoparticles (Figure 2 A and B).

The N₂ adsorption-desorption isotherms of the magnetic core MSNs presented a typical type IV behaviour (Figure 3), characteristic of mesoporous materials. From the isotherm curve, a specific surface area of 872 m²/g was estimated along with a pore volume of 0.85 cm³/g and an average pore diameter of 3.15 nm.

4.2. Trastuzumab labeling process with Tc-99m

The trastuzumab were successfully labelled (>97%) with Tc-99m generating a clear suspension.

4.3. Quality control of the labeling process with Tc-99m

The effectiveness of the labeling process was confirmed by paper chromatography which indicated almost no significant dissociation of technetium-99m from the trastuzumab for a period as long as 8 hours, as shown in Table 1:

4.4. Planar imaging from chromatographic system

In order to corroborate the paper chromatography a planar imaging from the paper chromatographic system has been performed, as shown in Figure 4.

The results showed an ascending movement with a great amount in the base corroborating the formation of the complex between the Tc-99m and the trastuzumab.

4.5. Entrapment efficiency – Tc-99m EE%

In order to calculate the amount of Tc-99m that has been absorbed/doped into the magnetic core mesoporous silica nanoparticles (together with the trastuzumab) we calculated the entrapment efficiency (EE%) of Tc-99m-trastuzumab into the magnetic core mesoporous silica nanoparticles, using the equation(2)::

Equation 2:

$${}^{99m}\text{TcEE}\% = \frac{\text{total amount of Tc - } {}^{99m}\text{Tc used}}{\text{total amount of Tc - } {}^{99m}\text{Tc in the supernatant}} \times 100$$

The result found for the Tc-99m EE% was 97,5% ±0,8. This result confirmed that almost all the Tc-99m used (labelling the trastuzumab) was trapped in the surface of the mesoporous silica.

The quality control of the solely trastuzumab labeling process with Tc-99m showed that over 97% of the Tc-99m remains connected with trastuzumab for a

period as long as 8 hours (as shown in Table 2), corroborating our decision to use this indirect way to calculate the amount of trastuzumab.

5. Conclusions

The results confirmed the potential use from the indirect entrapment efficiency methodology using the radiolabeling process. This methodology although involves the use of radioactive material can be an easy, rapid and efficient methodology that can be used to calculate Mab adsorption/entrapment efficiency into nanoparticles systems.

Conflict of Interest

The authors state that do not have any conflicts of interest

Acknowledgements

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Figures

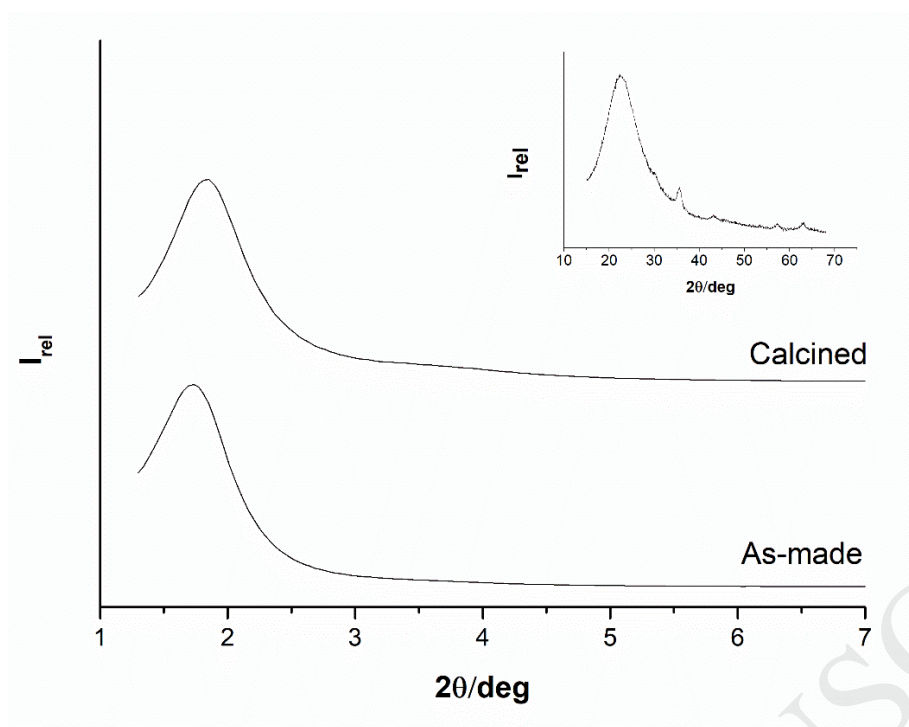


Figure 1. Powder X-ray diffraction patterns of (bottom) as-made magnetic core MSNs (**S0-1**) and calcined magnetic core MSNs (**S0-2**). Inset shows the peaks corresponding to magnetite nanocrystals and the characteristic broad peak of amorphous silica.

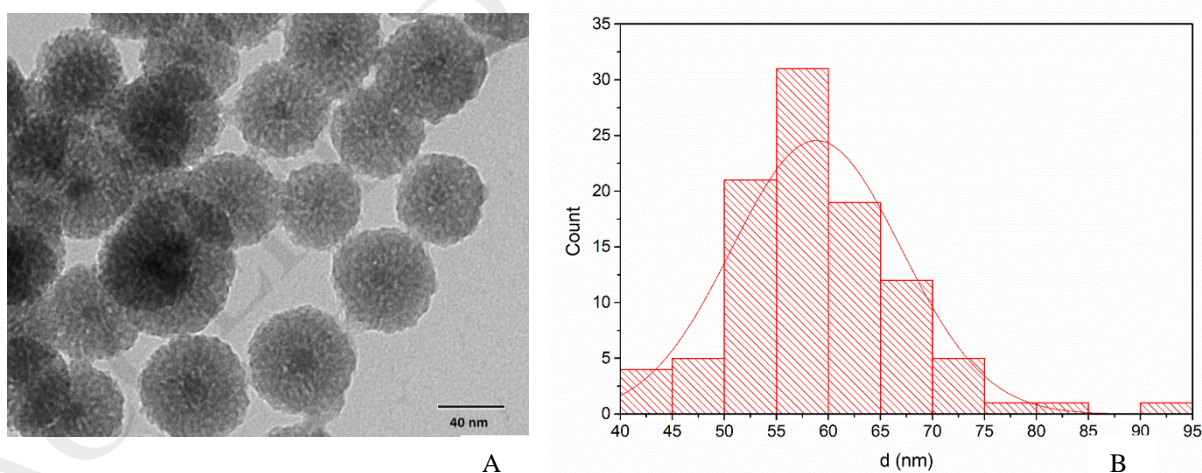


Figure 2. A: Transmission electron microscopy images of calcined **S0-2** nanoparticles, showing the spherical shape and the magnetic core. **B:** Histogram and normal size distribution of calcined **S0-2** nanoparticles determined by transmission electron microscopy image analysis ($n=100$).

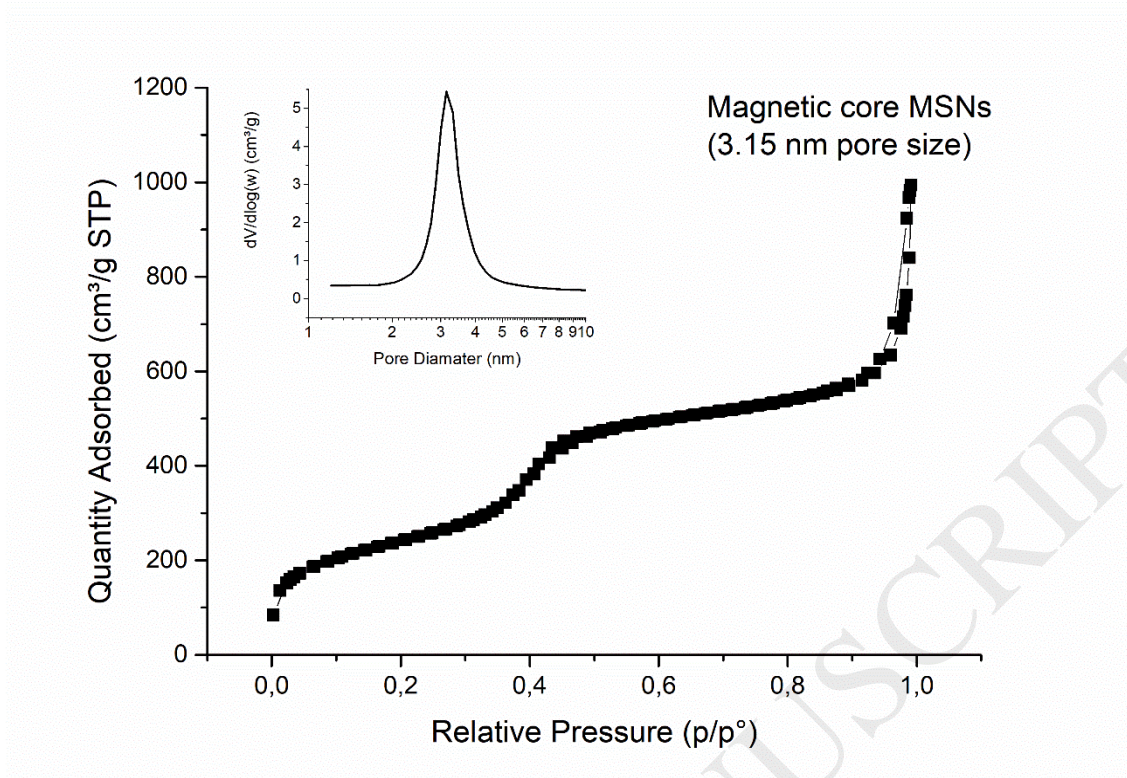


Figure 3. N₂ adsorption-desorption isotherm of calcined **S0-2** nanoparticles. Inset shows the pore size distribution of the material.

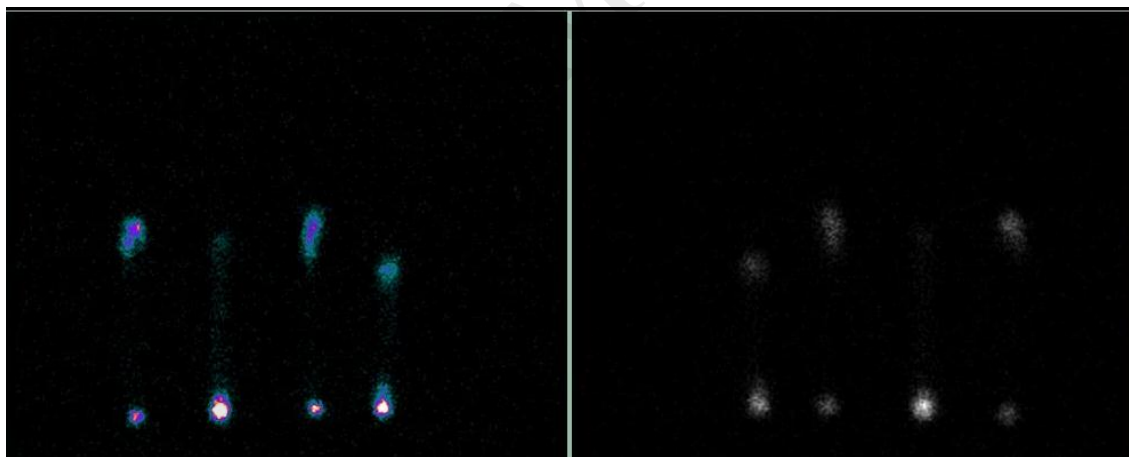


Figure 4. Planar imaging from the chromatographic system corroborating the formation of the complex Tc-99m-trastuzumab and the quality control from the labeling process.

Table 1. Percentage of labeled trastuzumab observed over time, after ascending chromatograms of Tc-99m compared with free pertechnetate ($\text{NaTc-}^{99\text{m}}\text{O}_4^-$).

Time (h)	Labeling (%)Trastuzumab
0	98.6± 0.7%
1	97.8± 0.5%
2	97.3± 1.0%
4	97.0± 0.8%
8	97.1± 0.5%

Table 2: Percentage of labeled trastuzumab observed over time, after ascending chromatograms of Tc-99m compared with free pertechnetate ($\text{NaTc-}^{99\text{m}}\text{O}_4^-$).

Time (h)	Labeling (%) Trastuzumab
0	99.1 ± 0.4%
1	98.7 ± 0.9%
2	97.9 ± 0.8%
4	98.4 ± 1.1%
8	97.5 ± 0.5%