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Additional Information

# 1 **A redox-sensitive nanofluidic diode based on nicotinamide-** 2 **modified asymmetric nanopores**

3 Mubarak Ali,<sup>a,b,\*</sup> Ishtiaq Ahmed,<sup>c</sup> Patricio Ramirez,<sup>d</sup> Saima Nasir,<sup>a</sup> Salvador Mafe,<sup>e</sup>  
4 Christof M. Niemeyer,<sup>c</sup> and Wolfgang Ensinger<sup>a</sup>

5 *<sup>a</sup>Technische Universität Darmstadt, Fachbereich Material- u. Geowissenschaften, Fachgebiet*  
6 *Materialanalytik, Alarich-Weiss-Str. 2, D-64287 Darmstadt, Germany*

7 *<sup>b</sup>GSI Helmholtzzentrum für Schwerionenforschung, Planckstr. 1, D-64291 Darmstadt, Germany*

8 *<sup>c</sup>Karlsruhe Institute of Technology (KIT), Institute for Biological Interfaces (IBG-1), Hermann-von-*  
9 *Helmholtz-Platz, D-76344 Eggenstein-Leopoldshafen, Germany*

10 *<sup>d</sup>Departament de Física Aplicada, Universitat Politècnica de València, E-46022 València, Spain*

11 *<sup>e</sup>Departament de Física de la Terra i Termodinàmica, Universitat de València, E-46100 Burjassot,*  
12 *Spain.*

13

14 \* Corresponding author: E-mail address: M.Ali@gsi.de (M. Ali)

15

## 16 **Abstract**

17 We demonstrate a redox-sensitive nanofluidic diode whose ion rectification is  
18 modulated by the oxidation and reduction of chemical moieties incorporated on its  
19 surface. To achieve this goal, we have first synthesized the chemical compounds 1-(4-  
20 aminobutyl)-3-carbamoylpyridin-1-ium (Nic-BuNH<sub>2</sub>) and 3-carbamoyl-1-(2,4-  
21 dinitrophenyl)pyridinium (Nic-DNP). Then, the surface of track-etched single  
22 asymmetric nanopores is decorated with the redox-sensitive Nic-BuNH<sub>2</sub> and Nic-DNP  
23 molecules using carbodiimide coupling chemistry and Zincke reaction, respectively.  
24 The success of the modification reactions is monitored through the changes in the  
25 current–voltage ( $I$ – $V$ ) curves prior to and after pore functionalization. Upon exposing

1 the modified pore to solutions of hydrogen peroxide (oxidizing agent) and sodium  
2 dithionite (reducing agent) the surface charge is reversibly modulated from positive to  
3 neutral, leading to measurable changes in the electronic readout of ion current passing  
4 through the nanopore. On oxidation, the quaternary nicotinamide units impart positive  
5 charge to the pore surface, resulting in the ion current rectification (anion-selective  
6 pore). On the contrary, the complementary reduced dihydronicotinamide moieties  
7 resulted in the loss of surface charge and ohmic behaviour (non-selective pore). The  
8 experimental results are further theoretically described by using Poisson-Nernst-  
9 Planck equations.

10

11 *Keywords:* synthetic nanopores; redox reaction; nicotinamide; current rectification;  
12 surface functionalization; track-etching.

13

## 14 **1. Introduction**

15 Ion channels and nanopores have received much attention because of their  
16 unique transport properties. In living organisms, ion channels regulate the flow of ions  
17 (e.g.,  $\text{Na}^+$ ,  $\text{K}^+$ , and  $\text{Cl}^-$ ) and small organic molecules into and out of the cell  
18 membrane to facilitate the chemical and electrical communication with the  
19 extracellular environment. Biological pores are highly selective and can discriminate  
20 between ions with similar size and same charge [1]. Therefore, they are frequently  
21 employed as model systems to investigate transport phenomena at the nanoscale [2,  
22 3]. Among the various protein pores, the  $\alpha$ -hemolysin self-assembled in a lipid bilayer  
23 has been extensively used for the detection and analysis of biomolecules [4, 5].  
24 However, the fragility and sensitivity of the embedding lipid bilayer towards harsh

1 conditions (temperature, pH and salt concentrations) restrain its use for more practical  
2 purposes. On the contrary, robust synthetic nanopores have recently attracted a great  
3 deal of interest in the area of nano/biotechnology. The structural (size and shape) and  
4 chemical (fixed charges) characteristics of synthetic nanopores have some similarities  
5 with their biological counterparts [6-8]. Asymmetric nanopores display ionic transport  
6 properties such as ion current rectification, current gating, and ion selectivity [9-12].

7       The electrical charge on the pore surface is one of the key parameters that  
8 determine the ion current rectification in conical nanopores [9-11]. To date, a variety  
9 of responsive molecules and functional groups that respond to external stimuli such as  
10 ions [13, 14], biomolecules [15-18], light [19-22], pH [3, 23-25], temperature [26, 27]  
11 and combinations of pH and temperature [28, 29] have been immobilized onto the  
12 pore surface. Upon the application of an external stimulus, the modified pore  
13 undergoes changes in the effective diameter and surface charge polarity, resulting in  
14 the variation of ionic flux across the membrane. Recently, ionic permeation across  
15 membranes has also been modulated through redox reactions occurring in  
16 nanoconfined geometries.[30-37] For this purpose, a variety of redox sensitive  
17 moieties have been functionalized on the inner pore walls. For example, Vansco and  
18 co-workers have demonstrated the fabrication of redox-responsive porous  
19 polyelectrolyte membranes by the complexation of poly(ferrocenylsilane) based  
20 poly(ionic liquid)s and poly(acrylic acid) [36]. Miller and Martin have demonstrated  
21 the tuning of electroosmotic flow through carbon nanotube membranes via depositing  
22 a thin film of redox polymer poly(vinylferrocene) on the inner walls [34]. Recently,  
23 Mitta et al, miniaturized a redox sensitive nanofluidic diode based on an asymmetric  
24 gold-coated nanopore through the electrochemical polymerization of aniline on the  
25 pore surface [35]. Moreover, we have also demonstrated the occurrence of redox

1 reaction in the confined environment upon the addition of hydrogen peroxide in the  
2 electrolyte solution in contact with horseradish peroxidase enzyme-modified  
3 nanopores [37]. To broaden the scope and application spectrum of these nanoporous  
4 systems, the synthesis and anchoring of more complex functional molecules, e.g.,  
5 oxidation-reduction (redox) sensitive moieties, on the pore surface constitutes a  
6 challenge for current techniques.

7 In living organisms, the majority of biological processes involves reversible  
8 oxidation and reduction (redox) reactions to store and release biological energy. The  
9 biologically relevant redox couples nicotinamide adenine dinucleotide  
10 ( $\text{NAD}^+/\text{NADH}$ ) and nicotinamide adenine dinucleotide phosphate ( $\text{NADP}^+/\text{NADPH}$ ),  
11 play a key role in cellular functions [38, 39]. Both contain the nicotinamide ring as a  
12 redox center on which oxidation and reduction processes occur. Oxidation of the  
13 substrate takes place by the removal of two hydrogen atoms (dehydrogenation) from  
14 the nicotinamide moiety. The oxidized nicotinamide ( $\text{NAD}^+$  or  $\text{NADP}^+$ ) accepts a  
15 hydride ion and subsequently transforms into the reduced nicotinamide ( $\text{NADH}$  or  
16  $\text{NADPH}$ ). The electrical charge on nicotinamide ring is dependent on the redox  
17 reaction, i.e., it is positively charged in oxidized and uncharged (neutral) in reduced  
18 form.

19 Previously, Imanishi and his co-workers have demonstrated the water  
20 permeation through porous membranes grafted with redox-responsive polymer  
21 brushes containing nicotinamide moieties [40]. Furthermore, Ishihara *et al.*  
22 investigated the insulin permeation rate via oxidation of nicotinamide moiety in a  
23 redox polymer grafted on a porous membrane [41]. Taking into account the redox-  
24 mediated switching of surface charge, we are interested in the possible modulation of

1 the transmembrane rectified ion flux by decorating nicotinamide units onto the inner  
2 surface and walls of the asymmetric nanopores.

3 We demonstrate the construction of a redox-sensitive nanofluidic diode based on  
4 single asymmetric nanopores. To achieve this goal, the chemical compounds 1-(4-  
5 aminobutyl)-3-carbamoylpyridinium (Nic-BuNH<sub>2</sub>) and 3-carbamoyl-1-(2,4-  
6 dinitrophenyl)pyridiniumchloride (Nic-DNP) are synthesized and anchored onto the  
7 pore surface via carbodiimide coupling chemistry and Zincke reaction, respectively.  
8 The pore surface properties are tuned by exposing the modified pore to solutions of  
9 hydrogen peroxide (oxidant) and sodium dithionite (reductant), which leads to  
10 measurable changes in the electronic readout of the current–voltage ( $I$ – $V$ )  
11 characteristics. The experimental results are also described theoretically by using a  
12 model based on the Poisson-Nernst-Planck equations.

13

## 14 **2. Materials and methods**

### 15 **2.1 Materials**

16 Polymer membranes of polyethylene terephthalate (PET) (Hostaphan RN 12,  
17 Hoechst) of thickness 12  $\mu$ m were irradiated at the linear accelerator UNILAC (GSI,  
18 Darmstadt) with single swift heavy ions (Au) of kinetic energy 11.4 MeV/nucleon.  
19 Then, the ion tracked membranes were subjected to soft UV light irradiation on each  
20 side for 60 minutes. The UV irradiation process sensitized the latent tracks of heavy  
21 ions for the subsequent etching process.

22 All the chemicals and reagents were of analytical grade and used as received  
23 without further purification. *N*-(3-dimethylaminopropyl)-*N*-ethylcarbodiimide  
24 hydrochloride (EDC), pentafluorophenol (PFP), nicotinamide (Nic), 2,4-

1 dinitrochlorobenzene (DNCB), *tert*-butyl *N*-(4-aminobutyl)carbamate, hydrogen  
2 peroxide (H<sub>2</sub>O<sub>2</sub>), sodium dithionite, sodium bicarbonate, sodium hydroxide,  
3 potassium chloride and trifluoroacetic acid were purchased from Sigma-Aldrich,  
4 Schnelldorf, Germany.

## 5 **2.2 Fabrication of single asymmetric nanopores**

6 The latent ion damage tracks in polymer membranes were converted into conical  
7 nanopores through the asymmetric track-etching technique reported by Apel and co-  
8 workers [42]. A custom-made conductivity cell with three chambers was employed  
9 for the simultaneous fabrication of single-pore and multipore membranes. A single-  
10 shot membrane and a membrane irradiated with a flux of 10<sup>7</sup> ions/cm<sup>2</sup> were placed on  
11 both sides of the middle chamber of the conductivity cell and clamped tightly. The  
12 middle compartment contained apertures on both sides and was filled with an etching  
13 solution (9 M NaOH). The other two compartments were filled with an acidic etch  
14 stop solution (1 M KCl + 1 M HCOOH). Gold electrodes were inserted on both sides  
15 of the single-ion irradiated membrane and a potential of –1 V was applied across the  
16 membrane to monitor the etching process at room temperature. During this process,  
17 the current remained zero until the etchant permeated through the whole length of the  
18 membrane. After the breakthrough point, an increase in ionic current flowing through  
19 the nascent pore was observed. The etching process was stopped as the current  
20 reached to 0.5 ± 0.1 nA. Subsequently, the membranes were thoroughly washed with  
21 stopping solution in order to neutralize the etchant, followed by deionized water. The  
22 etched membranes were then dipped in deionized water overnight in order to remove  
23 the residual salts.

24

## 1 **2.3 Synthesis of 1-(4-aminobutyl)-3-carbamoylpyridin-1-ium (Nic-BuNH<sub>2</sub>) (6)**

2 The chemical compounds 3-carbamoyl-1-(2,4-dinitrophenyl)pyridiniumchloride (Nic-  
3 DNP) referred as Zincke reagent (**3**) and 1-(4-aminobutyl)-3-carbamoylpyridin-1-ium  
4 (Nic-BuNH<sub>2</sub>) (**6**) were synthesized according to the reported method with slight  
5 modifications (see supporting information for detail) [43, 44].

## 6 **2.4 Functionalization of nanopore surface**

### 7 2.4.1 Carbodiimide coupling chemistry

8 The carboxylic groups on the pore surface were first activated by exposing the  
9 single pore membrane to an ethanolic solution of *N*-(3-dimethyl-aminopropyl)-*N*-  
10 ethylcarbodiimide (EDC; 100 mM)/pentafluorophenol (PFP; 200 mM) for 1 h at room  
11 temperature. After activation, the samples were exposed to a solution of Nic-BuNH<sub>2</sub>  
12 (50 mM) prepared in ethanol and the reaction was allowed to occur overnight. Finally,  
13 the functionalized membrane was washed several times with ethanol followed by  
14 deionized water.

### 15 2.4.2 Zincke reaction

16 In this case, the carboxylic acid groups were first derivatized with ethylenediamine  
17 via EDP/PFP coupling chemistry. Subsequently, the aminated single pore membrane  
18 was exposed to a Nic-DNP (Zincke reagent; 40 mM) solution prepared in methanol  
19 for 24 h. Then, the membrane was washed carefully with methanol followed by  
20 deionized water to remove the physically adsorbed Nic-DNP molecules from the pore  
21 surface.

## 22 **2.5 Current-voltage measurements**

23 The carboxylated and Nic-modified single-pore membrane was fixed between  
24 the two halves of the conductivity cell. Both halves of the cell were filled with a 0.1



1 M KCl solution at pH 6. An Ag/AgCl electrode was inserted into each half-cell  
2 solution and a picoammeter/voltage source (Keithley 6487, Keithley Instruments,  
3 Cleveland, Ohio, USA) was used to apply the desired transmembrane potential and  
4 measure the ionic current across the membrane. In the case of the asymmetric  
5 nanopore, the ground electrode was placed on the base side of the pore. In order to  
6 record the  $I$ - $V$  curves, a scanning triangle voltage signal from  $-2$  to  $+2$  V was used.

## 7 **2.6 Reduction of nicotinamide**

8 A solution of reductant  $\text{Na}_2\text{S}_2\text{O}_4$  (20 mM) was prepared in an aqueous  $\text{NaHCO}_3$   
9 (50 mM). For the reduction of nicotinamide into dihydronicotinamide groups on the  
10 pore surface, the modified pore was exposed to  $\text{Na}_2\text{S}_2\text{O}_4$  solution for 2 h. Then the  
11 pore was washed several times with water.

## 12 **2.7 Oxidation of dihydronicotinamide**

13 A 20 mM solution of oxidizing agent ( $\text{H}_2\text{O}_2$ ) was prepared in water. Then, the  
14 pore was exposed to  $\text{H}_2\text{O}_2$  solution for 2 h. During this time, the reduced  
15 dihydronicotinamide was oxidized back to nicotinamide moieties on the pore surface.  
16 The pore was washed with deionized water before  $I$ - $V$  measurement.

## 17 **2.8 Theoretical modeling**

18 The experimental  $I$ - $V$  curves can be described in terms of a theoretical model  
19 previously developed based on the Poisson and Nernst-Planck (PNP) equations

$$20 \quad \nabla^2 \phi = \frac{F}{\varepsilon} (c_{\text{Cl}^-} - c_{\text{K}^+}) \quad (1)$$

$$21 \quad \nabla \cdot \vec{J}_i = -\nabla \cdot [D_i (\nabla c_i + z_i c_i \nabla \phi)] = 0, i = \text{K}^+, \text{Cl}^- \quad (2)$$

22 where  $D_i$ ,  $c_i$ ,  $\vec{J}_i$ , and  $z_i$  are, respectively, the diffusion coefficient, the local  
23 concentration, the flux and the charge number of ion  $i$  ( $i = \text{K}^+$  and  $\text{Cl}^-$ ), and  $\varepsilon$  and  $\phi$  are  
24 the electrical permittivity and the local electric potential of the solution. Integration of

1 Equations (1)-(2) permits the calculation of the ionic fluxes at a given voltage. Once  
2 the ionic fluxes have been calculated, the electric current passing through any  
3 arbitrary section of radius  $a$  in the pore

$$4 \quad I = \sum_i z_i F \pi a^2 J_i . \quad (3)$$

5 can be obtained. Additional details can be found in Reference [45].

6 Assuming a conical geometry for the pore and infinite dilution values for the  
7 diffusion coefficients, the unknown model parameters are the diameters of the pore  
8 base and tip,  $D$  and  $d$ , respectively, and the concentration of fixed charges on the pore  
9 surface,  $\sigma$  (in elementary charges per square nanometer). In our experiments, the  
10 model parameters are assessed according to the following scheme. First, the pore base  
11 diameter is determined using SEM techniques with a membrane sample containing a  
12 large number of pores (ca.  $10^7$  pores  $\text{cm}^{-2}$ ) which was etched at the same time as the  
13 single pore sample. The tip diameter is calculated from the  $I-V$  curve of the as-  
14 prepared pore sample separating two 1 M KCl solutions. Under these conditions, the  
15 electrolyte ions screen the surface charges ( $\sigma = 0$ ) leading to a quasi-linear I-V curve.  
16 Parameters  $D$  and  $d$  are assumed to remain constant for a given sample after  
17 successive modifications of the pore surface charges. With this assumption, the only  
18 unknown parameter ( $\sigma$ ) is calculated by fitting the theoretical results to the  
19 experiments.

20 It must be mentioned that the 1-dimensional PNP model used here contains a number  
21 of approximations: the electrolyte is treated as an ideal solution, ignoring the finite  
22 size of ions, diffusion boundary layers at the membrane-solution interfaces that might  
23 lead to additional potential drops are not included in the model, and the ionic diffusion  
24 coefficients within the nanopore are similar to those in the bathing solutions. In  
25 addition, the shape of the nanopore might not be exactly conical and the surface

1 charge density could not be homogeneously distributed along the pore, as assumed in  
2 the model. Therefore, the conclusions concerning the nanopore structure obtained  
3 from the fitting procedure should be valid only qualitatively. While more realistic 2-D  
4 models and simulations handling the electrostatic correlations beyond the mean field  
5 approximation have been reported [46, 47], the pore geometry and the distribution of  
6 fixed charges along the pore cannot be unambiguously determined and these models  
7 include more unknown parameters than the simple 1-D approach used here.

8  
9

### 10 **3. Results and discussion**

11 In PET membranes, the fabrication of asymmetric nanopores was realized via  
12 the track-etching method. Swift heavy ion irradiation and chemical etching of ion  
13 tracks leads to the generation of carboxylic acid ( $-\text{COOH}$ ) groups on the pore surface.  
14 These groups served as starting units to further tune the chemical characteristics of the  
15 surface. The chemical compounds, 1-(4-aminobutyl)-3-carbamoylpyridin-1-ium (Nic-  
16  $\text{BuNH}_2$ ) and 3-carbamoyl-1-(2,4-dinitrophenyl)pyridinium (Nic-DNP) having  
17 nicotinamide moiety were synthesized (Scheme 1A). The Zincke reagent (**3**) was  
18 synthesized by reacting nicotinamide (**1**) and 2,4-dinitrochlorobenzene (**2**). The  
19 Zincke reagent **3** was then coupled with tert-butyl-4-aminobutylcarbamate (**4**) to  
20 prepare the Nic-BuNHBoc (**5**). The boc group was removed by treating compound **5**  
21 with trifluoroacetic acid to obtain Nic-BuNH<sub>2</sub> (**6**).

22 The as-prepared pores are used to covalently couple the carboxyl groups and  
23 amino groups of Nic-BuNH<sub>2</sub> on the channel surface through EDC/PFP coupling  
24 chemistry (Scheme 1B(a)). The Nic-DNP molecules were decorated on the EDA-  
25 modified pore surface using Zincke reaction (Scheme 1B(b)). The presence of

1 nicotinamide moiety onto the pore surface was confirmed by measuring the  $I-V$   
2 curves before and after chemical modification. For recording the  $I-V$  curves, the  
3 single-pore membrane (as-prepared/modified) was assembled between the two  
4 compartments of a conductivity cell. The electrolyte solution (0.1 M KCl) prepared in  
5 tris buffer (10 mM, pH 6.0) was filled in both compartments of the conductivity cell  
6 on either side of the membrane. The electrodes are arranged in such a way that high  
7 and low ionic currents were recorded for the positive and negative voltages,  
8 respectively.

9 It is well-known that conical nanopores exhibit cation selectivity and current  
10 rectification phenomena. This means that under an applied potential cations  
11 preferentially flow from the tip opening towards the base opening of the pore due to  
12 the presence of negatively charged carboxylate ( $-\text{COO}^-$ ) groups on the nanopore  
13 surface [9-11].

14 Figure 1A shows the  $I-V$  curves of a single asymmetric nanopore pore before  
15 and after the immobilization of nicotinamide moieties using EDP/PFP coupling  
16 chemistry. The modification process resulted in the switching of pore surface charge  
17 from negative to positive due to the presence of positively charged quaternary  
18 pyridinium units. This fact caused a reverse in the current rectification, i.e., the  
19 NicBu-modified pore preferentially transported anions from the tip to the base, as  
20 shown by the reverse  $I-V$  curve obtained compared with the case of the as-prepared  
21 pore of Figure 1A. This confirmed the successful attachment of nicotinamide moieties  
22 on the pore surface and walls. The theoretical results of Figure 1B were calculated  
23 following the procedure depicted in section 2.8 to determine the pore base ( $D = 400$   
24 nm) and tip ( $d = 24$  nm) diameters. The concentrations of surface charge where  $\sigma =$   
25  $-0.2 e/\text{nm}^2$  for the as-prepared pore and  $\sigma = 0.12 e/\text{nm}^2$  for the NicBu-modified pore,

1 where  $e$  is the elementary charge. These results indicate a partial substitution (~60%)  
2 of the original carboxylate groups of the as-prepared pore by the nicotinamide  
3 moieties of the NicBu-modified pore.

4 Once the nanopore surface was successfully functionalized with nicotinamide  
5 moieties, we proceeded to study the redox reactions inside the confined environment.  
6 The immobilized nicotinamide moieties were in oxidized state (quaternary pyridinium  
7 form) and the net charge on pore surface was positive, as evidenced from the current  
8 rectification reversal. The reduction of nicotinamide moieties was performed by  
9 replacing the electrolyte solution (100 mM KCl) used in the  $I-V$  measurements with  
10 sodium dithionite ( $\text{Na}_2\text{S}_2\text{O}_4$ ) prepared in sodium bicarbonate solution. The pore  
11 remained in contact with the reductant solution in dark for a time period. During the  
12 course of the reduction process, the oxidized (positively charged) pyridinium ring of  
13 nicotinamide was converted to 1,4-dihydropyridine (uncharged/ neutral) form,  
14 resulting in the loss of pore surface charges, as shown in Figure 2A. To avoid any  
15 interference from the reducing agent, the reductant solution was replaced by pure 0.1  
16 M KCl in both half cells for each  $I-V$  measurement. Moreover, the reproducibility of the  
17 redox system was validated by performing the same experiment on two single pore  
18 membrane samples (data not shown here).

19 Figure 2B depicts the  $I-V$  curves of the nanopore with nicotinamide moiety in  
20 the oxidized and reduced states. Upon reduction, the pore becomes nonselective due  
21 to the existence of uncharged dihydronicotinamide groups on the surface, as  
22 evidenced from the linear  $I-V$  characteristics (the net surface charge on the pore walls  
23 was then zero. From the  $I-V$  curves (1<sup>st</sup> cycle), the reduction process switched the  
24 nanopore from an “ON” state characterized by a high rectified ion flux to an “OFF”  
25 state with a low non-rectified current.

1           The dihydronicotinamide can be reoxidized back to nicotinamide in successive  
2 cycles by exposing the modified pore to a solution of an oxidizing agent. We  
3 employed hydrogen peroxide ( $\text{H}_2\text{O}_2$ ) for the oxidation of dihydronicotinamide. The  
4 oxidation process resulted in the generation of positive charges on the pore walls.  
5 Eventually, the pore became anion-selective and rectified the ion current as evidenced  
6 in the  $I-V$  curves (2<sup>nd</sup> cycle). This procedure was repeated again, giving the results of  
7 the  $I-V$  curves (3<sup>rd</sup> cycle). The redox-sensitive interconversion of nicotinamide  $\leftrightarrow$   
8 dihydronicotinamide is responsible for the polarity of the pore surface, which in turn  
9 affects the membrane permselectivity. This fact suggested that the redox controlled  
10 reversible nicotinamide/dihydronicotinamide interconversion provided a chemical  
11 tool to modulate the electronic readout of ionic current flowing through the nanopore,  
12 as shown in the experiments of Figure 2B.

13           Figure 2C shows the results obtained with the theoretical model in the  
14 successive reversible nicotinamide/dihydronicotinamide interconversion cycles. The  
15 pore diameters used in the calculations ( $D = 400$  nm and  $d = 24$  nm) were the same as  
16 in Figure 1B. The calculated surface charge densities (see the insets in the curves)  
17 reveal a partial recovery of the positive charges after the successive redox cycles.  
18 Progressive pore reutilization along several cycles decreases the effective number of  
19 charged groups leading to decreasing values of  $\sigma$ , probably because of partial  
20 regeneration.

21           The second approach employed to immobilize nicotinamide moieties on the pore  
22 surface was based on Zincke reaction (Scheme 1B(b)) [48, 49]. To this end, the  
23 carboxylic acid groups on the as-prepared pore surface of a new sample were first  
24 derivatized with ethylenediamine (EDA) using carbodiimide coupling chemistry. The  
25 presence of amine groups on the pore surface was confirmed by recording the  $I-V$

1 curve. The EDA-modified pore exhibited anion-selectivity, as reflected from its  
2 reversed  $I-V$  curve (Figure 3A). Subsequently, the EDA-modified pore was exposed  
3 to a methanolic solution of Nic-DNP compound (Zincke reagent) [49]. During the  
4 course of reaction, the amine groups on the pore surface acted as nucleophile and  
5 incorporated at the 6-position of nicotinamide group of Nic-DNP compound, resulting  
6 in the generation of 2,4-dinitroaniline. The pore functionalized through Zincke  
7 reaction, in the following denoted as “Nic-modified pore”, still exhibited reversed  
8 current rectification due to the existence of the positively charged nicotinamide  
9 moiety. When Nic-modified pore was exposed to reductant solution, the nicotinamide  
10 moiety transformed into its complementary reduced neutral dihydronicotinamide, as  
11 suggested by the linear  $I-V$  characteristic of the pore (Figure 3A). The theoretical  
12 results of Figure 3B were calculated again following the procedure described in  
13 section 2.8 and resulted in the pore diameters  $D = 240$  nm and  $d = 24$  nm and the  
14 surface charge densities indicated in the figure inset. In this case, the model results  
15 suggest the substitution of the amine groups by the nicotinamide groups of the pore  
16 surface.

17 We also performed a control experiment to demonstrate the success of Zincke  
18 reaction, the switching of pore surface polarity, and the concomitant changes in the  
19 current rectification observed in Figure 3 due to the reduction of nicotinamide. For  
20 this purpose, another pore modified with EDA was exposed to a solution of reducing  
21 agent under the same experimental conditions of Figure 3.  $I-V$  curves in Figure 4 did  
22 not show any significant change in the current rectification on exposure to reductant  
23 solution. These experimental results further confirmed the nicotinamide  
24 immobilization on the pore surface via Zincke reaction and the subsequent reduction  
25 inside the confined geometry.

1

## 2 **4. Conclusions**

3 In summary, we have demonstrated the fabrication of a redox-sensitive  
4 nanofluidic diode whose transport properties can be reversibly tuned via oxidation and  
5 reduction processes occurring inside the pore. The chemical compounds Nic–BuNH<sub>2</sub>  
6 and Nic–DNP were synthesized and functionalized on the pore surface via  
7 carbodiimide coupling chemistry and Zincke reaction, respectively. After the  
8 nicotinamide functionalization, the surface polarity was switched from negative to  
9 positive, resulting in the reverse of the current rectification characteristic of the  
10 nanopore. The reduction of nicotinamide into dihydronicotinamide (neutral) moieties  
11 led to the loss of current rectification and the pore behaved like an ohmic resistor. The  
12 rectification behaviour was subsequently recovered by re-oxidation of  
13 dihydronicotinamide into nicotinamide moieties. In summary, the redox reactions  
14 allowed us to reversibly switch the nanopore inner environment from hydrophilic and  
15 conducting (“*ON*” state) to a hydrophobic and nonconducting (“*OFF*” state). Model  
16 calculations allow for the determination of the key transport parameters of the  
17 functionalized pores.

18

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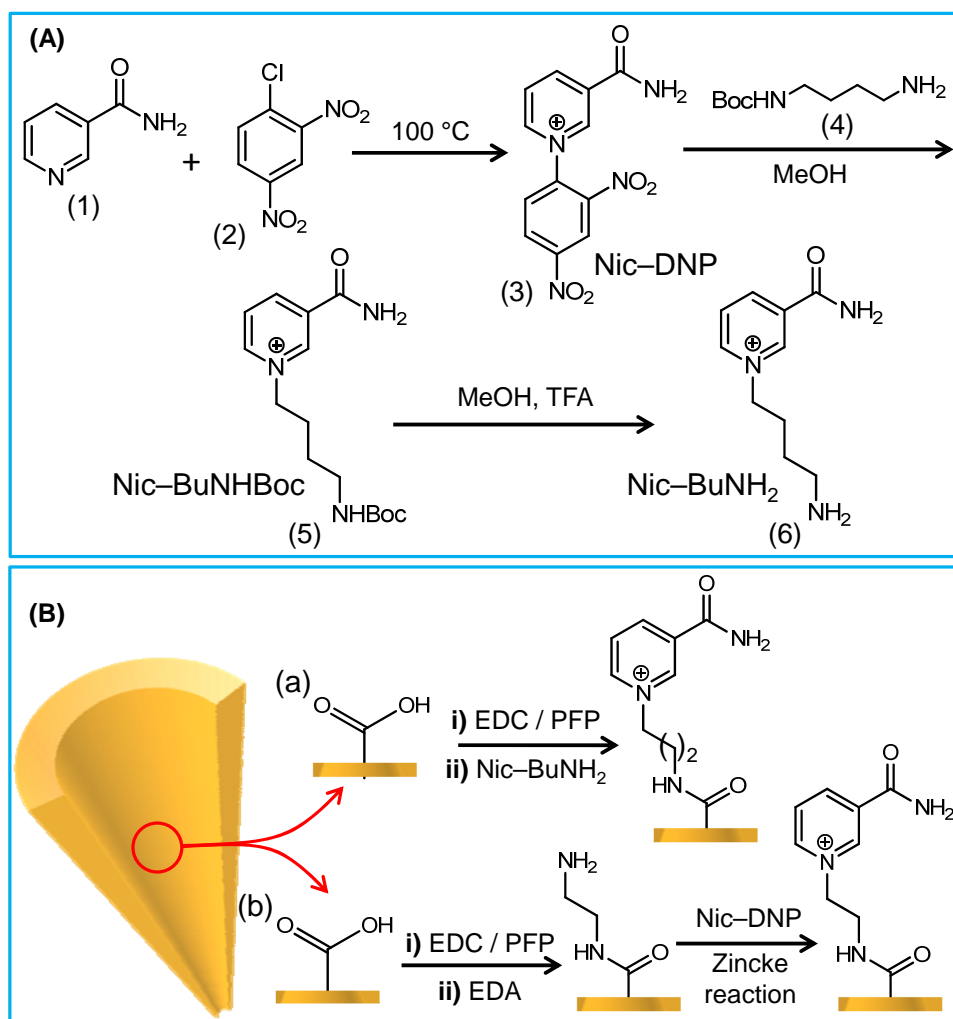
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## 1 Figures and Legends

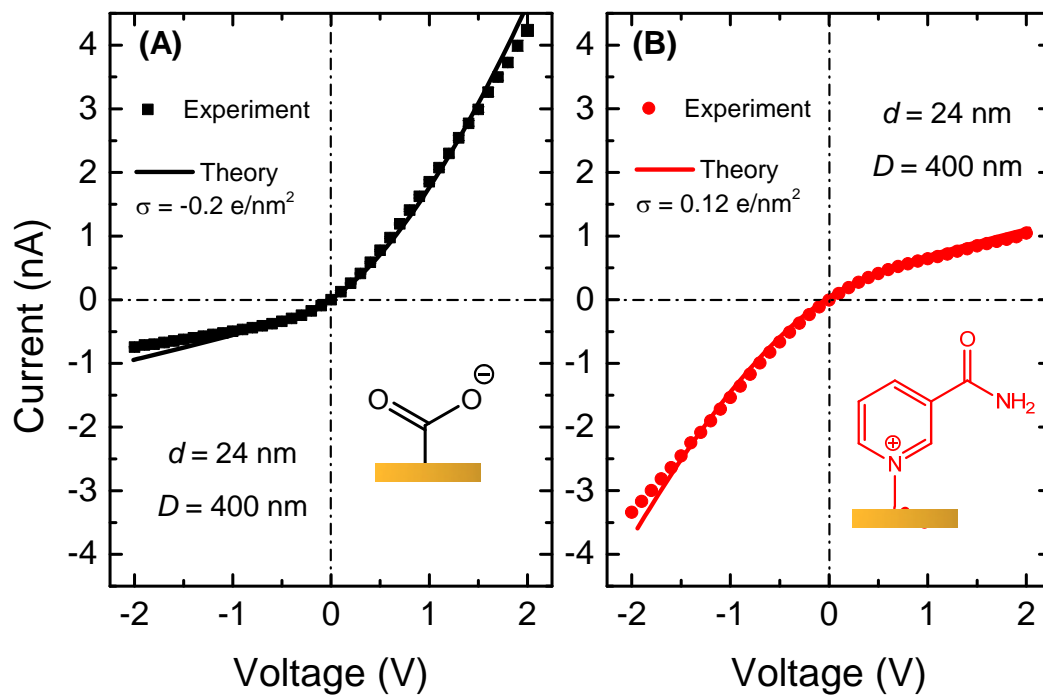


2

3 **Scheme 1.** (A) Reaction scheme for the synthesis of Nic-DNP (3) and Nic-BuNH<sub>2</sub> (6)  
4 compounds. (B) The functionalization of the pore surface carboxylic acid groups with  
5 Nic-BuNH<sub>2</sub> molecules via carbodiimide coupling chemistry (a) and immobilization of  
6 Nic-DNP moieties through Zincke reaction (b).

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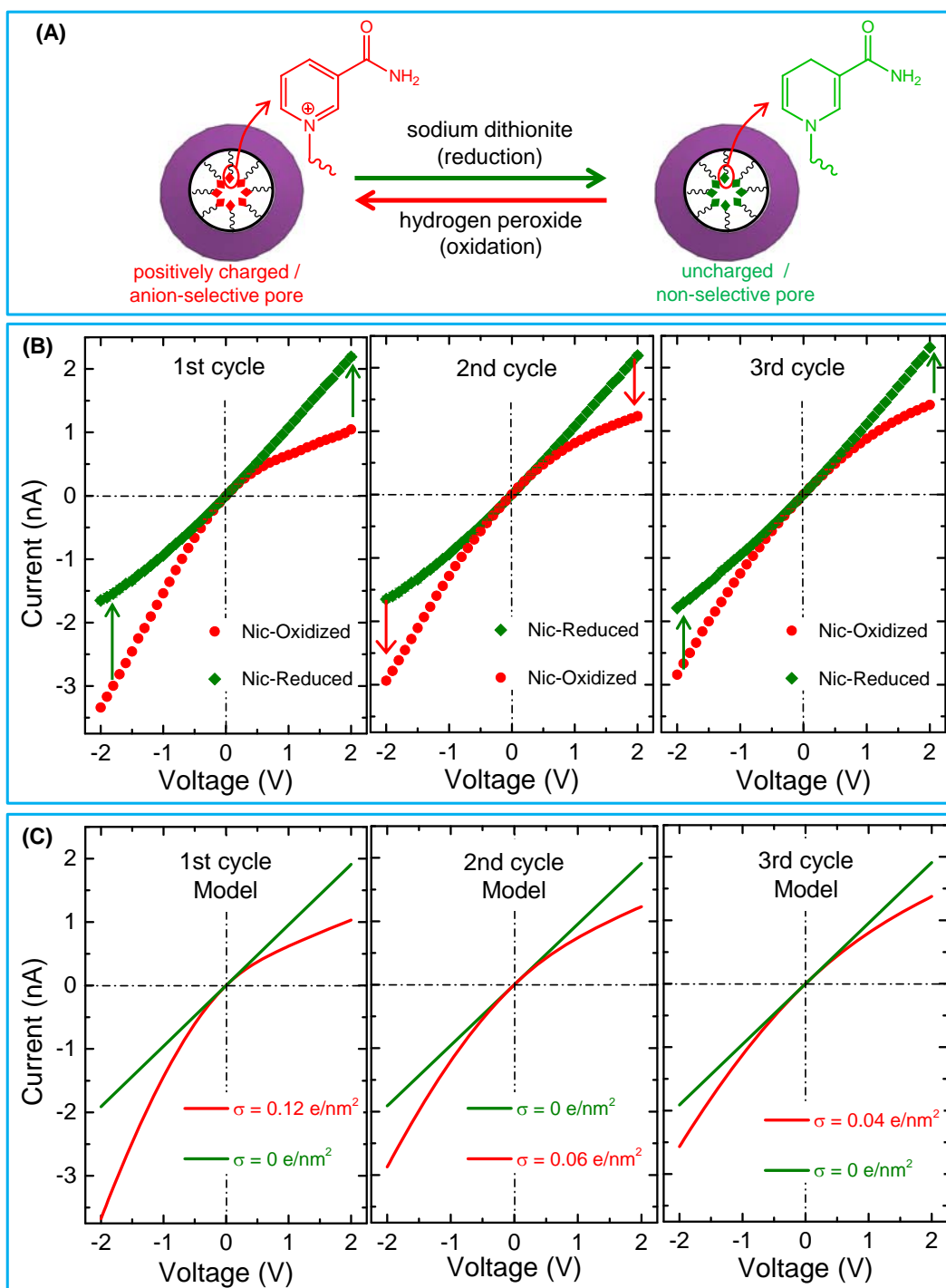
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1

2 **Figure 1.** Experimental and theoretical  $I$ - $V$  curves of single conical nanopore  
 3 measured in aqueous 0.1M KCl (pH 6.0) with carboxylate groups (A) and  
 4 nicotinamide moieties (B) on the pore surface.

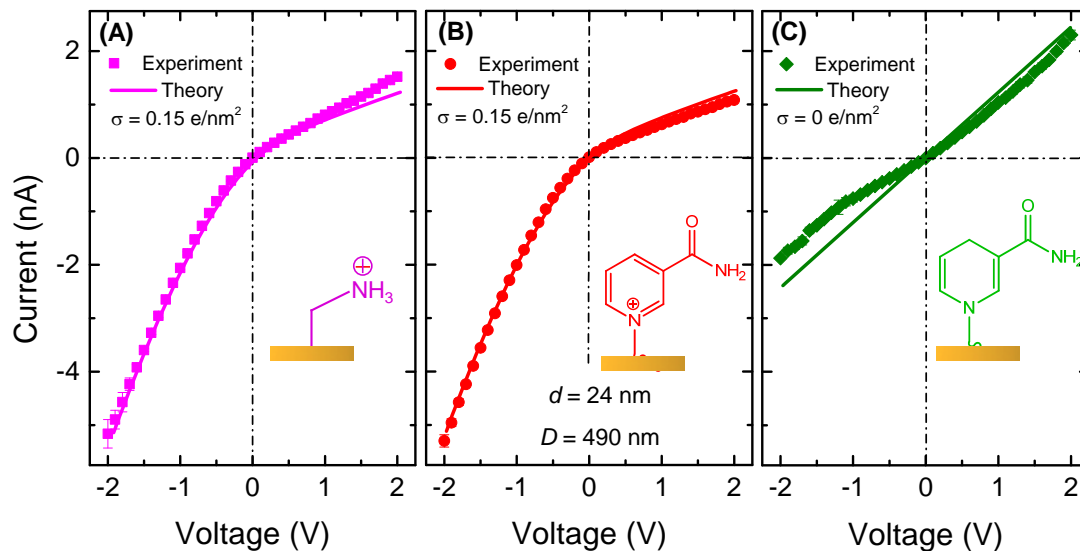
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2 **Figure 2.** (A) Scheme for the oxidation and reduction of nicotinamide moieties on the  
 3 inner pore walls. (B) Experimental  $I-V$  curves of the NicBu-modified pore exhibiting  
 4 the reversible switching between the oxidized (positively charged) and reduced  
 5 (uncharged) states of nicotinamide moieties measured in aqueous 0.1M KCl (pH 6.0),  
 6 separately. (C) Theoretical  $I-V$  curves corresponding to the experimental data. Pore  
 7 diameters  $D$  and  $d$  were the same as those in Figure 1B.

8



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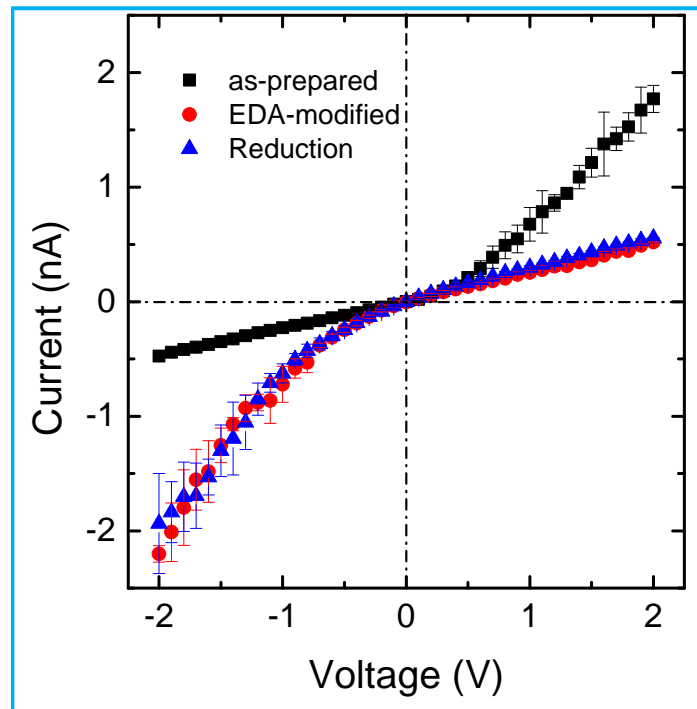
2 **Figure 3.** Experimental and theoretical  $I$ - $V$  curves of single conical nanopore  
 3 containing the amine groups (A) and the nicotinamide moieties in the oxidized (B)  
 4 and reduced (C) states measured in an aqueous 0.1M KCl (pH 6.0) solution,  
 5 separately.

6

7



1



2

3 **Figure 4.**  $I$ - $V$  curves of as-prepared pore and EDA-modified conical nanopore after  
4 exposure to dithionite solution measured in aqueous 0.1 M KCl (pH 6.0) solution.

5