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Additional Information

26 depending on the product, species, harvest conditions, processing and storage [1]. The
27 variability in its composition reinforces the importance of specifically evaluating the
28 performance of each potential biomass for a given process.

29 Cassava (*Manihot esculenta*) is one of the main raw materials used in the starch extraction
30 industry, where cassava bagasse appears as a valuable and abundant byproduct. Currently,
31 in Brazil alone, approximately 2 million tons of wet bagasse per year are generated, [2,
32 3]. The differential for cassava bagasse in relation to other residues is associated with its
33 high residual starch content which is a consequence of the low yield of cassava starch
34 extraction [4], which can range from approximately 30 up to 85% (in dry basis) according
35 to the literature [5, 6].

36 Due to its specific chemical composition, cassava bagasse is a potential raw material for
37 bioethanol production. However, well designed pretreatment and hydrolysis needs to be
38 carried out to access the entangled starch and the complex structure of cellulose and
39 hemicellulose. The conversion efficiency of these main macromolecules into small chain
40 carbohydrates must be characterized by high sugar production with low formation of
41 sugar degradation compounds. This can be attained by pretreating the biomass under mild
42 temperatures, using less aggressive acids and under easy-mixing conditions. Phosphoric
43 acid, for example, is able to hydrolyze both lignocellulosic and starchy biomass with
44 reduced sugar degradation [7, 8]. Its use at low temperatures can also prevent the starchy
45 suspension from gelatinizing, which would decrease the mass transfer and the yield of
46 cold hydrolysis of starch granules.

47 In addition to the technological issues, the process efficiency is highly dependent on the
48 correct design of equipment and unit operations. The properties of the raw material is the
49 fundamental information needed to optimize the industrial process. Properties, such as
50 the rheological ones, allow an adequate choice of pumps, accessories, mixers, bioreactors

51 and pipes [9]. However, the study of the properties demands more efforts for the case of
52 heterogeneous aqueous systems which may be influenced by the different amounts of
53 insoluble (biomass particles) and soluble (acid) compounds. While particles can have
54 different sizes and aspect ratios, the presence of an acid can affect how the particles
55 interact with each other during the flow [10, 11].

56 Consequently, the way each of the factors listed above influences the rheological behavior
57 of dilute acid suspensions of cassava bagasse needs to be investigated. Steady-flow curves
58 provide useful data not only for the rheological modelling but also for evaluating the
59 transition between Newtonian and non-Newtonian behavior. Modifications in the
60 suspension compositions may turn Newtonian fluids into viscoplastic fluids with
61 significant initial stress to start the flow, pseudoplastic (or shear-thinning) or dilatant (or
62 shear-thickening) fluids in the presence or absence of yield stress. On the one hand, the
63 shear-thinning fluids show decreasing apparent viscosity as the shear rate is increased.
64 On the other hand, the apparent viscosity of the shear-thickening ones tends to increase
65 with increasing shear rate. Distinct systems may need different sources of energy to attain
66 similar pretreatment conditions, even more so when they include non-conventional
67 techniques such as high-intensity ultrasound. For example, suspensions concentrated in
68 biomass tend to require more energy input to reach the same acoustic effect power
69 ~~obtained~~ in dilute suspensions [12], which is probably related to the increased viscosity
70 [9].

71 Therefore, this study focused on determining and modelling the rheological behavior of
72 different acid suspensions containing previously-characterized cassava bagasse.

73

74 **2. Materials and Methods**

75 **2.1. Cassava bagasse and suspension preparation**

76 Coarsely particulate, dried cassava bagasse was kindly provided by TechnoAmido (São
77 Pedro do Turvo, Brazil). These coarse particles were ground using an M4FH1C knife mill
78 (PHD, Piracicaba, Brazil) at 1750 rpm and then the powdered bagasse was standardized
79 using a sieve (Tyler mesh 100) in order to obtain particles smaller than 0.147 mm.
80 Dilute acid suspensions were prepared varying the phosphoric acid (85% solution;
81 purity>99.95%), (Dinâmica, Diadema, São Paulo), content in distilled water at the
82 following concentrations: 0, 2, 4, 6, 8 and 10% (g of acid per 100 g of dispersant). These
83 solutions were added to cassava bagasse to obtain suspensions with final solids
84 concentrations of 0, 2, 4, 6, 8 and 10% (g of cassava bagasse per 100 g of suspension).
85 Combining the 6 acid solutions with the 6 cassava bagasse concentrations resulted in a
86 total of 36 samples to be analyzed.

87

88 **2.2. Cassava bagasse characterization**

89 **2.2.1. Chemical composition**

90 The cassava bagasse was characterized according to its chemical composition, in
91 triplicate, by determining the protein content by the micro-Kjeldahl method, fat content
92 by Soxhlet extraction, ash content by muffle incineration and moisture content in an oven
93 according to AOAC [13]. Among the carbohydrates, starch content was quantified in a
94 similar way to that described by Paul et al. [14], total fiber content was quantified by the
95 method proposed by Van Soest et al. [15] and total soluble sugars was found by
96 difference. Moreover, the fractions of cellulose, hemicellulose and lignin in the
97 lignocellulose were determined based on the values of insoluble fibers in acid (ADF) and
98 neutral detergent (NDF).

99

100 **2.2.2. Particle size**

101 Particle size of the powdered cassava bagasse was determined in triplicate by laser
102 diffraction (LD) using a MAM 5005 Mastersizer (Malvern Panalytical, Malvern, UK).
103 Distilled water was used as the dispersing medium for the measurements. In this
104 technique, the particles scatter light at an angle of diffraction which is considered
105 inversely proportional to their size. The particle size distribution is initially measured
106 based on volume, expressing the volume percentage of particles in continuous size
107 intervals. Making use of these values, the number-based distribution can be determined
108 by the equivalent sphere assumption.

109 The diameter of the samples was represented as the volume weighted mean diameter
110 ($D_{4,3}$), also known as the De Brouckere mean diameter, defined by Equation (1), and
111 surface weighted mean diameter ($D_{3,2}$) or the so-called Sauter mean diameter, defined by
112 Equation (2). The De Brouckere mean diameter assumes spherical particles of the same
113 volume as the actual particles, while the Sauter mean diameter is defined as the diameter
114 of the sphere with the same surface area as the particle [16]. Polydispersity (width of
115 distribution) was also evaluated according to span value, given in Equation (3). All of
116 them were calculated with respect to the volume-based and number-based distributions.

$$117 \quad D_{4,3} = \frac{\sum n_i \cdot d_i^4}{\sum n_i \cdot d_i^3} \quad (1)$$

$$118 \quad D_{3,2} = \frac{\sum n_i \cdot d_i^3}{\sum n_i \cdot d_i^2} \quad (2)$$

$$119 \quad \text{span} = \frac{D_{0,9} - D_{0,1}}{D_{0,5}} \quad (3)$$

120 where, n_i is the number of particles with d_i diameter (μm), $D_{0,1}$, $D_{0,5}$ and $D_{0,9}$ are additional
121 parameters given by the instrument which represent the maximum particle diameter
122 below which 10%, 50% and 90% of the sample exists, respectively.

123

124 **2.2.3. Particles morphology**

125 The morphology of cassava bagasse particles was evaluated using a BX43 optical
126 microscope (Olympus, Tokyo, Japan) with coupled camera at 4x, 10x and 40x magnitude.
127 Samples were dispersed in distilled water for glass slide preparation and images were
128 acquired from different areas on the microscope slides. In order to identify and to show
129 the presence of residual starch, the bagasse was marked with lugol solution.
130 Besides the visual analyses, particles were also characterized according to the aspect ratio
131 (a_r) given by the relation between length (L) and width (w). Measurements of such
132 characteristic dimensions were done for 100 particles in the different fields of view using
133 the cellSens software version 1.18 (Olympus, Tokyo, Japan).

134

135 **2.3. Rheological measurements**

136 Rheological measurements were carried out in triplicate using an AR-G2 rotational
137 rheometer (TA Instruments, New Castle, USA) with the SPC (Starch Pasting Cell)
138 geometry. This vane-type geometry was used in order to avoid particle sedimentation and
139 slip effects [17]. Approximately 28 mL of each sample were inserted into the equipment
140 and shear rate ramps were set from 1 to 265 s^{-1} in steady flow with 10 points per logarithm
141 interval. The rheological experiments were conducted at five different temperatures:
142 278.13, 288.13, 298.13, 308.13 and 318.13 K, using a controlled temperature bath and a
143 new sample for each assay. Data of shear stress were obtained from Universal Analysis
144 2000 data acquisition system version 4.7 (TA Instruments, New Castle, USA).

145

146 **2.4. Modelling**

147 The means of the triplicate experimental values of the shear stress obtained at each fixed
148 shear rate were calculated for the 36 samples at the temperatures of 278.13, 288.13,

149 298.13, 308.13 and 318.13 K. Average data of shear stress were plotted against shear rate
150 to obtain the corresponding rheograms. The Newton (Equation 4), Bingham (Equation 5),
151 Power Law (Equation 6) and Herschel-Bulkley (Equation 7) models were used to analyze
152 the rheological behavior:

$$153 \quad \tau = \mu \dot{\gamma} \quad (4)$$

$$154 \quad \tau = \tau_0 + \eta_B \dot{\gamma} \quad (5)$$

$$155 \quad \tau = k \dot{\gamma}^n \quad (6)$$

$$156 \quad \tau = \tau_0 + k \dot{\gamma}^n \quad (7)$$

157 In these equations, τ (Pa) is shear stress, $\dot{\gamma}$ (s^{-1}) is shear rate, μ (Pa·s) is the Newtonian
158 viscosity, τ_0 (Pa) is the yield stress, η_B (Pa·s) is the plastic viscosity, k (Pa·s^{*n*}) is the
159 consistency coefficient and n (dimensionless) is the flow behavior index. Fluids with flow
160 behavior index below 1 correspond to shear-thinning fluids while values above 1 indicate
161 shear-thickening behavior. The rheograms were fitted to the Herschel-Bulkley model in
162 order to estimate and to evaluate the rheological parameters, the presence of yield stress
163 and shear-thinning or shear-thickening behavior. The apparent viscosity η_{app} (Pa·s) at a
164 fixed shear rate can be determined from the values of μ after equalizing the shear stress
165 of a non-Newtonian model to the Newton model [18]. The OriginPro 8.0 software
166 (OriginLab Corporation, Northampton, USA) was used to plot the graphs and to carry out
167 the non-linear regressions.

168 The parameters from the Herschel-Bulkley model were submitted to estimated effects
169 analysis and to the analysis of variance using the STATISTICA 10 software (StatSoft
170 Inc., Tulsa, USA) in order to study the influence of the variables on the rheological
171 parameters. The significance of the linear and quadratic terms of cassava bagasse
172 concentration (X_S , %), phosphoric acid content (X_A , %) and absolute temperature (T , K)

173 were analyzed as well as the possible linear interaction among them. The significant
 174 variables ($p < 0.05$) were used to fit the data and provide a final version of the predictive
 175 equation for each parameter. The accuracy of fit was evaluated through the adjusted
 176 determination coefficient (R_{adj}^2), which is a modification of the conventional
 177 determination coefficient R^2 to take into account the number of parameters of the fitted
 178 model, and by the root mean square error ($RMSE$), given respectively by Equation (8) and
 179 Equation (9):

$$180 \quad R_{adj}^2 = 1 - \frac{(m-1)}{(m-p-1)}(1-R^2) \quad (8)$$

$$181 \quad RMSE = \sqrt{\frac{\sum_{m=1}^m (\hat{y}_m - y_m)^2}{m}} \quad (9)$$

182 where m is the number of experimental data, p the number of parameters in the model, R^2
 183 is the determination coefficient, \hat{y}_m the predicted data and y_m the experimental data.

184 The effect of temperature on the apparent viscosity of the suspensions was modelled by
 185 an Arrhenius-type equation (Equation 10):

$$186 \quad \eta_{app,10s^{-1}} = A_0 \exp\left(\frac{E_a}{RT}\right) \quad (10)$$

187 where A_0 is the pre-exponential factor ($\text{Pa}\cdot\text{s}$), E_a is the activation energy ($\text{J}\cdot\text{mol}^{-1}$) and R
 188 is the universal gas constant ($8.314 \text{ J}\cdot\text{mol}^{-1}\cdot\text{K}^{-1}$).

189

190 **3. Results and discussion**

191 **3.1. Chemical characterization**

192 Cassava bagasse samples were chemically characterized (Table 1). The samples have
 193 considerable amounts of residual starch around 50% (in dry basis). This agrees with
 194 results reported by other authors of residual starch in cassava bagasse ranging from 30%

195 up to 85% (in dry basis) [4, 5]. Starch is a polymer with great capacity for releasing
196 fermentable matter. After being hydrolyzed, it can be used as substrate for many
197 applications, including for second generation ethanol production.

198 Similarly as reported by Polachini et al. [19] and Silva et al. [20], the cassava bagasse
199 also showed low fat, ash and protein content. As a consequence of the low concentration
200 of high-value nutrients, cassava bagasse is not often used for animal feed. Consequently,
201 the higher carbohydrate content makes it advantageous for microbial conversion when
202 compared to other residues [4].

203 Fibers represented 46.1% of the bagasse, approximately 92% being cellulose and
204 hemicellulose. In addition to the capacity for releasing sugar from starch, low molecular
205 weight carbohydrates can also be obtained in large amounts from the fibrous matter. Lu
206 et al. [21] showed the high production of glucose and lower amounts of xylose and
207 arabinose from the hydrolysis of starch and lignocellulose from cassava bagasse. On the
208 other hand, it is known that the presence of lignin makes the hydrolyzation of cellulose
209 and hemicellulose difficult. Thus, the low content of lignin observed in cassava bagasse
210 fibers ($\approx 8\%$) offers advantages in contrast with other residues such as sugarcane bagasse,
211 rice straw, corn stover and wheat straw, which all showed 14–23% lignin by dry weight
212 [22].

213

214 **3.2. Particles size determination**

215 Figure 1 shows the particle size distribution, where a monomodal distribution can be seen
216 for both volume-based and number-based distribution.

217 Table 2 presents the average values of the different diameters as well as the span values
218 obtained from the volume-based and number-based distributions. According to the $D_{0,9}$
219 from the number-based distribution, powdered cassava bagasse presented the majority

220 (90%) of particles with particle size smaller than 6.1 μm . However, probably, the presence
221 of a few bigger particles (fibers and agglomerates) resulted in a volume weighted mean
222 diameter ($D_{4,3}$) of approximately 120 μm

223 Concerning the volume-based distribution, the average volume-weighted mean diameter
224 ($D_{4,3}$) was 175 μm . On the other hand, the values for surface-weighted mean diameter
225 $D_{3,2}$ (~60 μm) were much lower than $D_{4,3}$ probably due to the presence of longer, thinner
226 particles instead of more rounded ones [9]. According to Glaser [23], $D_{3,2}$ is an efficient
227 parameter for evaluating hydrolysis performance of non-homogeneous substrates. The
228 same author also stated that biomass with particle size below 200 μm is more accessible
229 to cellulases for further conversion. Consequently, cassava bagasse is an adequate
230 substrate for cellulases since all the mean diameters obtained were lower than this value.
231 A slightly higher span value of 1.935 was a consequence of the varied particle sizes,
232 which can be seen in Figure 1. Moreover, 10% of the volume occupied by the particles
233 ($D_{0,1}$) is attributed to particles smaller than 40 μm and 90% of the volume occupied ($D_{0,9}$)
234 corresponds to particles smaller than 340 μm . Thus, it can be assumed that that 80% of
235 the occupied volume is linked to particles with mean diameter between 40 μm and 340
236 μm . This observation reinforced the difference between the two distributions, which
237 means that more small particles are necessary to occupy the same volume as bigger ones.

238

239 **3.3. Morphological characteristics**

240 Figure 2 shows particles with varied morphologies such as round, rectangular and
241 elongated shapes in the micrographs at different magnifications. They are typical of
242 cellulosic residues, with irregular form and the possible presence of microfibrils. Similar
243 structures have been reported in the literature for peanut shells [9], sugarcane bagasse
244 [24] and even cassava bagasse [25]. In addition to the cellulosic particles, there were also

245 starch granules (black arrows in Figure 2a). Figure 2b shows the cassava bagasse marked
246 with lugol, where the bluish-purple structure indicated starch with iodine. It could be seen
247 that some starch was in the form of granules dispersed or entangled in the fibrous network,
248 which was also observed by Versino et al. [26] and Odoch et al. [27] using scanning
249 electronic microscopy of cassava bagasse. Another fraction seemed to be solubilized as
250 amylose and amylopectin, probably as a result of a partial starch gelatinization and
251 leaching during the cassava processing. The bluish areas also reflect the difficulty in
252 recovering all the cassava starch during its extraction process.

253 The aspect ratio (a_r) of the samples was also determined from the microscopic images.
254 Mean aspect ratio of the sample was found to be 1.98, being the distribution of the aspect
255 ratio shown in the histogram in Figure 3. More than 60% of the particles analyzed had
256 aspect ratios in the range between 1 and 2.5, which is similar to the mean values and the
257 data reported for other agro-industrial residues [28]. The presence of starch granules in
258 relatively spherical form leads to a high concentration of particles with an aspect ratio
259 near to 1.

260

261 **3.4. Flow behavior**

262 The mean shear stress was calculated in all samples for each fixed shear rate at the
263 different temperatures. An average value of the relative standard deviations (standard
264 deviation/mean shear stress) were also calculated in a given flow condition, being these
265 values lower than 3.86% in all of the measurements regarding the 36 samples at the
266 different temperatures. The values obtained were in the range from 0.001 Pa to 8.181 Pa
267 at 1.01 s^{-1} and from 0.184 Pa to 604.91 Pa at 263.34 s^{-1} . In general, as shown in Figure 4,
268 shear stresses increased with the increase of cassava bagasse and acid content and the
269 lowering of temperature. An off-trend displacement was observed between the flow

270 curves of 8% and 10% of solids in Figure 4a, 8% and 10% of acid concentration in Figure
271 4b and of 308.13 K and 318.13 K in Figure 4c. It indicates that some non-linear trend
272 occurred when solids loading, acid concentration and temperature were varied.

273 Figure 4 also shows some rheograms fitted to the Herschel-Bulkley model for different
274 solids concentration, acid content and temperature. In Table 3, some fitting parameters
275 for acid suspensions at intermediate concentration of solids, acid content and
276 temperatures are shown. Full data are provided as Supplementary File (Table S1)

277 Although all data were well-fitted to Herschel-Bulkley model ($R^2 > 0.998$), not all fitting
278 parameters presented large values or physical meaning. For example, acid solutions and
279 suspensions with solids content up to 4% showed relatively low yield stresses (τ_0) and
280 flow behavior index (n) very close to 1. In these conditions, the Herschel-Bulkley model
281 turns into the Newton model and the consistency coefficient can be considered as
282 Newtonian viscosity. Coussot [29] assumed that the interactions between biomass
283 particles are negligible in suspensions with low concentration of solids ($< 4\%$), with
284 Newtonian-like behavior of the dispersant medium predominating.

285 When the solids content and the phosphoric content was 6%, non-Newtonian behavior
286 was more marked. It occurred for suspensions containing powdered peanut shells at
287 higher solids concentration (8%) [9]. Besides the phosphoric acid, the main differences
288 that could be linked to the different types of biomass can be attributed to the chemical
289 composition and to the mean particle size ($D_{3,2} = 178.46 \mu\text{m}$ for peanut shells and $D_{3,2} =$
290 $66.3 \mu\text{m}$ for cassava bagasse), since the aspect ratio was quite similar. Peanut shells are
291 mainly lignocellulosic material while the cassava bagasse is composed by considerable
292 amounts of starch granules and this difference may affect compressibility and frictional
293 forces between the matrices structure [30]. Hou et al. [31] also reported higher apparent
294 viscosities for suspensions containing smaller particles in comparison with suspensions

295 constituted by bigger particles, probably due to an increased specific surface area of
296 thinner particles and the consequent higher particles interaction [32].

297 Regarding the rheological parameters, it was noticed that the yield stress (τ_0) increased
298 and the values of the flow behavior index (n) decreased when phosphoric acid or cassava
299 bagasse concentrations were higher. At the lower temperature with the highest acid and
300 solids concentration the highest τ_0 value (3.21) and the lowest n (0.85) were found. These
301 values are in accordance with previous studies for both lignocellulosic and starchy
302 materials, which fitted the Herschel-Bulkley model to suspensions containing corn stover,
303 peanut shells and amaranth starch [9, 33, 34] with similar trends. As reported in the
304 literature, suspensions containing biomass generally present shear-thinning behavior with
305 significant yield stress when approximately 5% of solids is attained [11]. This trend is
306 likely due to the increased interparticle friction when decreasing the content of interstitial
307 solvent, which influences not only the required energy for starting the flow but also how
308 significant is the particles and aggregates reorganization when shear rate is changed.

309 The maximum value observed for yield stress was approximately 3.22 Pa, indicating that
310 the most concentrated suspensions tested in this work were still able to be mixed by
311 traditional stirred tanks instead of specialized reactors. Roche et al. [35] stated that
312 suspensions with yield stress up to 10 Pa behave as pourable liquids, instead of paste-like
313 characteristics with complex transport properties. These conditions promote the acid-
314 particle interactions since the catalyst is more adequately and uniformly distributed in the
315 dispersant when compared to concentrated materials [33]. In this context, the energy
316 required for moving the materials increases with increasing solids content. The
317 consistency coefficient can increase up to approximately 20 times (from 0.25 to 4.96
318 Pa·s^{*n*}) when the solids concentration varies from 2% to 10% (5 times).

319 In addition to the visible effect of the variables on rheological parameters, an analysis of
 320 the estimated effects was carried out to evaluate its significance ($p < 0.05$ confirmed by
 321 the analysis of variance in Tables S2, S3, S4 and S5 from the Supplementary File). In
 322 general, yield stress (τ_0) and consistency coefficient (k) were linearly affected by all
 323 variables. Additionally, significant quadratic effects were observed for temperature and
 324 biomass concentration, besides the linear interaction $X_S T$ and $X_S X_A$, on both τ_0 and k . On
 325 the other hand, flow behavior index (n) was linearly affected by the biomass and acid
 326 contents and by the linear interaction between them. Solids content also had a quadratic
 327 effect on n , with no significant influence of temperature.

328 Apparent viscosity ($\eta_{app,10s^{-1}}$) was calculated at specific shear rate of 10 s^{-1} , which is
 329 considered adequate for a mixing and stirring process [18, 33]. It ranged from 7.07×10^{-4}
 330 Pa·s for the pure water at 318.13 K up to 3.82 Pa·s for the most solid and acid concentrated
 331 suspension at 278.13 K. These values were linearly influenced by all variables with an
 332 additional quadratic effect of temperature. The linear interactions $X_S T$ and $X_S X_A$ were also
 333 significant, similar to τ_0 and k .

334 In this way, non-linear regression can be done for each rheological parameter as a
 335 simultaneous function of the significant variables. Equation (11), Equation (12), Equation
 336 (13) and Equation (14) that follow were well-fitted to yield stress, consistency coefficient,
 337 flow behavior index and apparent viscosity. They had the fitted determination coefficient
 338 (R_{adj}^2) equal to 0.8596, 0.9744, 0.9392 and 0.9677, and the root mean square error equal
 339 to 0.28 Pa, 0.19 Pa·sⁿ, 0.01 and 0.17 Pa·s, respectively:

$$340 \quad \tau_0 = 25.48 - 0.177T + 0.00031T^2 + 1.2648X_S - 0.00471X_S^2 \quad (11)$$

$$\quad \quad \quad + 0.06515X_A - 0.00371X_S T + 0.00834X_S X_A$$

$$341 \quad k = 25.49 - 0.179T + 0.00031T^2 + 1.8991X_S - 0.01222X_S^2 \quad (12)$$

$$\quad \quad \quad + 0.05198X_A - 0.00591X_S T + 0.00756X_S X_A$$

342 $n = 0.9987 + 0.006588X_S - 0.001617X_S^2 + 0.000187X_A - 0.000553X_SX_A$ (13)

343 $\eta_{app,10s^{-1}} = 24.09 - 0.16795T + 0.00029T^2$ (14)
 344 $+ 1.70511X_S + 0.066X_A - 0.00497X_S T + 0.00296X_SX_A$

344 In any acid solutions and acid suspensions with solids concentration lower than 6%, yield
 345 stress values can be disregarded ($\tau_0 \approx 0$), the flow behavior index can be approximated to
 346 unity ($n \approx 1$) and both k and $\eta_{app,10s^{-1}}$ can be considered as for the Newtonian viscosity.

347

348 3.4.1. Solids concentration

349 Figure 5 shows the yield stress, consistency coefficient and flow behavior index as
 350 functions of cassava bagasse concentration. These graphs are in accordance with the
 351 previous analyses of variance, presenting the linear and quadratic effect of biomass on all
 352 rheological parameters.

353 The values of yield stress increased as the solids concentration increased. Although
 354 authors correlate yield stress with solids concentration by a power relation [11], Laera et
 355 al. [36] states that a linear tendency can be assumed in the case of diluted suspensions.
 356 The yield stress information is useful since it corresponds to the initial energy needed to
 357 move biomass suspensions. The way yield stress behaves as solids concentration changes
 358 can provide important information about the increase in power requirements when
 359 particle settling occurs before the mixer starts moving [11].

360 A similar trend was also observed for the consistency coefficient and for apparent
 361 viscosity. Higher values were obtained with increasing biomass content probably due to
 362 more particle-particle interaction during the flow as a consequence of the number of
 363 particles and the reduction of dispersant as it is absorbed causing the particles to swell
 364 [22]. In addition, mechanical energy is also dissipated by the dispersant when it flows
 365 around the particles when they appear in slightly higher concentrations [37].

366 The flow behavior index had a non-linear decrease as the solids concentration increased.
367 The drop in the n values reflects the decrease in apparent viscosity as the shear rate
368 increased. Polachini et al. [9] related the shear-thinning behavior to the increasing number
369 of collisions among the particles in high loaded suspensions and a consequent deviation
370 from Newtonian properties. Moreover, the breakdown of the aggregate as the shear rate
371 increases can promote the release of interstitial dispersant and particles alignment,
372 providing easy-flow characteristics and the reduction in apparent viscosity values.

373

374 **3.4.2. Phosphoric acid content**

375 The acidification of the medium also had significant influence on the rheological
376 parameters. Figure 6 shows the effect of H_3PO_4 concentration on rheological behavior for
377 different acid suspensions. While the yield stress, consistency coefficient and apparent
378 viscosity increased linearly with increasing acid concentration, the flow behavior index
379 seemed to present a slight less pronounced linear decrease as acid content was increased.
380 The apparent viscosity of the acid solutions increased linearly with increasing acid
381 content. Furthermore, Pääkkö et al. [10] stated that the negative charges of hemicellulose
382 can be neutralized by the H^+ ions. This might reduce electrostatic repulsion among the
383 particles, which leads to an increase in the interfibrillar action and, consequently, in the
384 suspension viscosity. Also, this increase in the particle-particle interaction seemed to
385 slightly enhance the shear-thinning behavior in the higher loaded samples. This could be
386 due to higher particle aggregation and the consequent release of the water trapped within
387 the aggregates at higher shear rates.

388

389 **3.4.3. Temperature dependence**

390 Temperature had significant influence on yield stress, consistency coefficient and
391 apparent viscosity but no effect on the flow behavior index. The values of τ_0 , k and
392 $\eta_{app,10s^{-1}}$ tended to decrease significantly as the temperature increased (Figure 7). The
393 resistance to flow decreased as a consequence of the increase in the degree of molecular
394 thermodynamic movement with increasing temperature. Increasing the temperature keeps
395 the particles away from each other which reduces both viscosity and the specific mass of
396 liquid materials. A similar trend was observed for microalgae slurries and peanut shell
397 suspensions over similar temperature ranges [9, 37].

398 Although the analyses of variance showed quadratic influence of temperature on τ_0 , k and
399 $\eta_{app,10s^{-1}}$, the literature usually correlates the effect of temperature on rheological
400 parameters by an Arrhenius-type equation. Thus, the activation energy of the Arrhenius
401 equation reflects the temperature dependence of each sample. In other words, suspensions
402 with higher E_a have more pronounced variation in viscosity for the same temperature
403 change when compared to the ones with lower E_a .

404 The fitting parameters of an Arrhenius-type equation are shown in Table 4. The goodness
405 of fit was measured by R_{adj}^2 , achieving values greater than 0.8603 for all samples. Ségalen
406 et al. [38] demonstrated that slurries such as sludge follow an Arrhenius-type behavior
407 when they are fully broken, i. e., when non-hydrodynamic forces can be neglected by
408 applying enough shear rate. It means that the suspensions at the studied conditions in our
409 work presented liquid-like characteristics, its transport being easier when compared to
410 solid-like fluids.

411 The flow activation energy of the acid solutions ($X_S = 0\%$) resulted in higher values than
412 the ones for acid suspensions, even presenting a slight decrease with increasing acid
413 concentration. These values are in close agreement with the range established for pure
414 water (13 up to 16 $\text{kJ}\cdot\text{mol}^{-1}$) [39], as it was the major component. These values indicated

415 that the dispersants had their apparent viscosity more strongly affected by temperature in
416 relation to the suspensions. Krokida et al. [40] stated that the addition of non-soluble
417 solids, such as fibers, in the suspensions implies lower E_a when compared to water.

418 The addition of biomass particles to an acidified medium can increase the number of
419 hydrogen bonds related to, e. g. the carboxyl groups in the hemicellulose and hydroxyl
420 groups in the amylose and amylopectin. The higher sensitivity of apparent viscosity with
421 increasing solids content can be linked to the structural changes of the aggregates as a
422 consequence of the buildup or breakdown of the hydrogen bonding and hydrophobic
423 interactions when temperature is altered [41]. Analogously, it could be seen that E_a of the
424 suspensions tend to decrease as the acid concentration increased, probably due to the
425 neutralization of the carboxyl and hydroxyl groups by the H^+ ions.

426 In the aqueous suspensions ($X_A = 0\%$), the activation energy tended to decrease when the
427 solids content increased. The absence of hydrogen ions may raise difficulties in
428 establishing hydrogen bonds among the molecules, leading to charge repulsion by the
429 particles. In addition, the high number of small particles in powdered cassava bagasse
430 could act like a dispersant to facilitate the flow of coarser particles. This phenomenon,
431 observed by Farris [42], could explain the reduced required flow energy as the
432 suspensions become more concentrated.

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434 **4. Conclusions**

435 The high content of residual starch (~49%) and cellulose and hemicellulose (total of
436 ~43%) in the dried bagasse highlighted the need for an adequately designed conversion
437 process not only for converting entangled and dispersed residual starch but also the
438 fermentable matter of lignocellulose. In order to provide important information for the
439 design of equipment and plant operations, rheological behavior of different acid

440 suspensions was studied as a function of solids concentration (0–10%), phosphoric acid
441 in the solution (0–10%) and temperature (278.13–318.13 K). Samples with less than 6%
442 of solids presented Newtonian behavior while the more concentrated ones ($\geq 6\%$) behaved
443 as non-Newtonian fluids with noticeable yield stress ($0 \text{ Pa} < \tau_0 < 3.22 \text{ Pa}$) and shear-
444 thinning characteristics ($0.85 < n < 1$).

445 Resistance to the flow (τ_0 , k and $\eta_{app,10s^{-1}}$) was increased by increasing cassava bagasse
446 loading, acid content and decreasing temperature. Flow behavior index (n) decreased as
447 the biomass and acid concentration increased but no effect of temperature was noticeable.
448 For the range of variables considered, polynomial models could be fitted to provide ready-
449 to-use information. Under the studied conditions, suspensions showed liquid-like
450 properties. These facilitate the mass transport and the access of enzymes when compared
451 to high concentrated pastes. Additionally, in these conditions, hydrolysis and further
452 additional treatments can be enhanced since these conditions allow more efficient
453 propagation of ultrasound in conversion processes assisted by non-conventional
454 technologies.

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464 **6. References**

- 465 [1] K.L. Kenney, W.A. Smith, G.L. Gresham, T.L. Westover, Understanding biomass
466 feedstock variability, *Biofuels*, 4 (2013) 111-127.
- 467 [2] F.A. Fiorda, M.S.S. Soares-Jr, F.A. Silva, L.R.F. Souto, M.V.E. Grossmann, Farinha
468 de bagaço de mandioca: aproveitamento de subproduto e comparação com fécula
469 de mandioca, *Pesquisa Agropecuária Tropical*, 43 (2013) 408-416.
- 470 [3] FAO, Official data of cassava production, Food and Agriculture Organization of the
471 United Nations, 2019.
- 472 [4] A. Pandey, C.R. Soccol, P. Nigam, V.T. Soccol, L.P.S. Vandenberghe, R. Mohan,
473 Biotechnological potential of agro-industrial residues. II: cassava bagasse,
474 *Bioresource Technology*, 74 (2000) 81-87.
- 475 [5] D. Pasquini, E.M. Teixeira, A.A.S. Curvelo, M.N. Belgacem, A. Dufresne, Extraction
476 of cellulose whiskers from cassava bagasse and their applications as reinforcing
477 agent in natural rubber, *Industrial Crops and Products*, 32 (2010) 486-490.
- 478 [6] A.S. Sánchez, Y.L. Silva, R.A. Kalid, E. Cohim, E.A. Torres, Waste bio-refineries for
479 the cassava starch industry: New trends and review of alternatives, *Renewable and*
480 *Sustainable Energy Reviews*, 73 (2017) 1265-1275.
- 481 [7] S. Gámez, J.J. González-Cabriales, J.A. Ramírez, G. Garrote, M. Vázquez, Study of
482 the hydrolysis of sugar cane bagasse using phosphoric acid, *Journal of Food*
483 *Engineering*, 74 (2006) 78-88.
- 484 [8] P. Lenihan, A. Orozco, E. O'Neill, M.N.M. Ahmad, D.W. Rooney, G.M. Walker,
485 Dilute acid hydrolysis of lignocellulosic biomass, *Chemical Engineering Journal*,
486 156 (2010) 395-403.
- 487 [9] T.C. Polachini, A.C.K. Sato, R.L. Cunha, J. Telis-Romero, Density and rheology of
488 acid suspensions of peanut waste in different conditions: An engineering basis for
489 bioethanol production, *Powder Technology*, 294 (2016) 168-176.

- 490 [10] M. Pääkkö, M. Ankerfors, H. Kosonen, A. Nykänen, S. Ahola, M. Österberg, J.
491 Ruokolainen, J. Laine, P.T. Larsson, O. Ikkala, T. Lindström, Enzymatic
492 Hydrolysis Combined with Mechanical Shearing and High-Pressure
493 Homogenization for Nanoscale Cellulose Fibrils and Strong Gels,
494 *Biomacromolecules*, 8 (2007) 1934-1941.
- 495 [11] J.J. Stickel, J.S. Knutsen, M.W. Liberatore, W. Luu, D.W. Bousfield, D.J.
496 Klingenberg, C.T. Scott, T.W. Root, M.R. Ehrhardt, T.O. Monz, Rheology
497 measurements of a biomass slurry: an inter-laboratory study, *Rheol Acta*, 48 (2009)
498 1005-1015.
- 499 [12] T.C. Polachini, G.R.d. Carvalho, J. Telis-Romero, Determination of acoustic fields
500 in acidic suspensions of peanut shell during pretreatment with high-intensity
501 ultrasound, *Brazilian Journal of Chemical Engineering*, 34 (2017) 385-394.
- 502 [13] AOAC, Official Methods of Analysis, Association of Official Analytical Chemists,
503 Washington, D. C., 2005.
- 504 [14] S. Paul, M.K. Das, P. Baishya, A. Ramteke, M. Farooq, B. Baroowa, R. Sunkar, N.
505 Gogoi, Effect of high temperature on yield associated parameters and vascular
506 bundle development in five potato cultivars, *Scientia Horticulturae*, 225 (2017)
507 134-140.
- 508 [15] P.J. Van Soest, J.B. Robertson, B.A. Lewis, Methods for Dietary Fiber, Neutral
509 Detergent Fiber, and Nonstarch Polysaccharides in Relation to Animal Nutrition,
510 *Journal of Dairy Science*, 74 (1991) 3583-3597.
- 511 [16] J.M. Castagnini, N. Betoret, E. Betoret, P. Fito, Vacuum impregnation and air drying
512 temperature effect on individual anthocyanins and antiradical capacity of blueberry
513 juice included into an apple matrix, *LWT - Food Science and Technology*, 64
514 (2015) 1289-1296.

- 515 [17] H.A. Barnes, A review of the slip (wall depletion) of polymer solutions, emulsions
516 and particle suspensions in viscometers: its cause, character, and cure, *Journal of*
517 *Non-Newtonian Fluid Mechanics*, 56 (1995) 221-251.
- 518 [18] J.F. Steffe, *Rheological Methods in Food Process Engineering*, 2 ed., Freeman Press,
519 East Lansing, 1996.
- 520 [19] T.C. Polachini, L.F.L. Betiol, J.F. Lopes-Filho, J. Telis-Romero, Water adsorption
521 isotherms and thermodynamic properties of cassava bagasse, *Thermochimica Acta*,
522 632 (2016) 79-85.
- 523 [20] E.C. Silva, M.P. Cereda, T.A.D. Colman, I.M. Demiate, E. Schnitzler,
524 Characterisation of cassava bagasse in different granulometries from two starch
525 processing plants, *The Journal of Microbiology, Biotechnology and Food Sciences*,
526 5 (2015) 99-102.
- 527 [21] C. Lu, J. Zhao, S.-T. Yang, D. Wei, Fed-batch fermentation for n-butanol production
528 from cassava bagasse hydrolysate in a fibrous bed bioreactor with continuous gas
529 stripping, *Bioresource Technology*, 104 (2012) 380-387.
- 530 [22] B. Volynets, F. Ein-Mozaffari, Y. Dahman, Biomass processing into ethanol:
531 pretreatment, enzymatic hydrolysis, fermentation, rheology, and mixing, *Green*
532 *Processing and Synthesis*, 6 (2017) 1-22.
- 533 [23] R. Glaser, Enzyme-based lignocellulose hydrolyzation—Sauter mean diameter of
534 raw materials as a basis for cellulase performance characterization and yield
535 prediction, *Journal of Biotechnology*, 214 (2015) 9-16.
- 536 [24] A.K. Chandel, F.A. Antunes, V. Anjos, M.J. Bell, L.N. Rodrigues, I. Polikarpov,
537 E.R. de Azevedo, O.D. Bernardinelli, C.A. Rosa, F.C. Pagnocca, S.S. da Silva,
538 Multi-scale structural and chemical analysis of sugarcane bagasse in the process of

539 sequential acid–base pretreatment and ethanol production by *Scheffersomyces*
540 *shehatae* and *Saccharomyces cerevisiae*, *Biotechnol Biofuels*, 7 (2014) 63.

541 [25] A.P. Travalini, E. Prestes, L.A. Pinheiro, I.M. Demiate, Extraction and
542 Characterization of Nanocrystalline Cellulose from Cassava Bagasse, *Journal of*
543 *Polymers and the Environment*, (2017).

544 [26] F. Versino, O.V. López, M.A. García, Sustainable use of cassava (*Manihot*
545 *esculenta*) roots as raw material for biocomposites development, *Industrial Crops*
546 *and Products*, 65 (2015) 79-89.

547 [27] M. Odoch, E.M. Buys, J.R.N. Taylor, Mechanism of cassava tuber cell wall
548 weakening by dilute sodium hydroxide steeping, *Food Chemistry*, 228 (2017) 338-
549 347.

550 [28] S.A. EL-Sayed, M.E. Mostafa, Analysis of Grain Size Statistic and Particle Size
551 Distribution of Biomass Powders, *Waste and Biomass Valorization*, 5 (2014) 1005-
552 1018.

553 [29] P. Coussot, *Rheometry of Pastes, Suspensions and Granular Materials – Application*
554 *in Industry and Environment*, John Wiley & Sons, Hoboken, New Jersey, 2005.

555 [30] D.J. Klingenberg, T.W. Root, S. Burlawar, C.T. Scott, K.J. Bourne, R. Gleisner, C.
556 Houtman, V. Subramaniam, Rheometry of coarse biomass at high temperature and
557 pressure, *Biomass and Bioenergy*, 99 (2017) 69-78.

558 [31] W. Hou, L. Zhang, J. Zhang, J. Bao, Rheology evolution and CFD modeling of
559 lignocellulose biomass during extremely high solids content pretreatment,
560 *Biochemical Engineering Journal*, 105 (2016) 412-419.

561 [32] N.-S. Roh, D.-H. Shin, D.-C. Kim, J.-D. Kim, Rheological behaviour of coal-water
562 mixtures. 1. Effects of coal type, loading and particle size, *Fuel*, 74 (1995) 1220-
563 1225.

- 564 [33] S. Viamajala, J.D. McMillan, D.J. Schell, R.T. Elander, Rheology of corn stover
565 slurries at high solids concentrations – Effects of saccharification and particle size,
566 *Bioresource Technology*, 100 (2009) 925-934.
- 567 [34] A.M. Gozzo, R.L. Cunha, F.C. Menegalli, Extensional and shear viscosity of
568 acidified amaranth starch-sodium caseinate suspensions, *Food Science and*
569 *Technology (Campinas)*, 29 (2009) 587-596.
- 570 [35] C.M. Roche, C.J. Dibble, J.S. Knutsen, J.J. Stickel, M.W. Liberatore, Particle
571 concentration and yield stress of biomass slurries during enzymatic hydrolysis at
572 high-solids loadings, *Biotechnology and Bioengineering*, 104 (2009) 290-300.
- 573 [36] G. Laera, C. Giordano, A. Pollice, D. Saturno, G. Mininni, Membrane bioreactor
574 sludge rheology at different solid retention times, *Water Research*, 41 (2007) 4197-
575 4203.
- 576 [37] H. Chen, Q. Fu, Q. Liao, H. Zhang, Y. Huang, A. Xia, X. Zhu, Rheological properties
577 of microalgae slurry for application in hydrothermal pretreatment systems,
578 *Bioresource Technology*, 249 (2018) 599-604.
- 579 [38] C. Ségalen, E. Dieudé-Fauvel, J. Clément, J.C. Baudez, Relationship between
580 electrical and rheological properties of sewage sludge – Impact of temperature,
581 *Water Research*, 73 (2015) 1-8.
- 582 [39] N. Ouerfelli, Z. Barhoumi, O. Iulian, Viscosity Arrhenius Activation Energy and
583 Derived Partial Molar Properties in 1,4-Dioxane + Water Binary Mixtures from
584 293.15 to 323.15 K, *Journal of Solution Chemistry*, 41 (2012) 458-474.
- 585 [40] M.K. Krokida, Z.B. Maroulis, G.D. Saravacos, Rheological Properties of Fluid Fruit
586 and Vegetable Purree Products: Compilation of Literature Data, *International*
587 *Journal of Food Properties*, 4 (2001) 179-200.

- 588 [41] V.D. Alves, F. Freitas, N. Costa, M. Carvalheira, R. Oliveira, M.P. Gonçalves,
589 M.A.M. Reis, Effect of temperature on the dynamic and steady-shear rheology of a
590 new microbial extracellular polysaccharide produced from glycerol byproduct,
591 Carbohydrate Polymers, 79 (2010) 981-988.
- 592 [42] R.J. Farris, Prediction of the Viscosity of Multimodal Suspensions from Unimodal
593 Viscosity Data, Transactions of The Society of Rheology (1957-1977), 12 (1968)
594 281-301.
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598 **Table 1.** Chemical composition of the powdered cassava bagasse.

General composition	
Moisture (g·100 g ⁻¹ wet matter)	5.2 ± 0.1
Protein (g·100 g ⁻¹ dried matter)	1.37 ± 0.05
Fats (g·100 g ⁻¹ dried matter)	0.30 ± 0.05
Ash (g·100 g ⁻¹ dried matter)	1.12 ± 0.02
Total sugars (g·100 g ⁻¹ dried matter)	1.63 ± 0.03
Starch (g·100 g ⁻¹ dried matter)	49.5 ± 0.8
Fibers (g·100 g ⁻¹ dried matter)	46.1 ± 0.9
Fiber composition*	
Cellulose (g·100 g ⁻¹ dried fiber)	50.9 ± 0.5
Hemicellulose (g·100 g ⁻¹ dried fiber)	41.5 ± 0.4
Lignin (g·100 g ⁻¹ dried fiber)	7.69 ± 0.07

* Based on the values of NDF.

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619 **Table 2.** Parameters of number-based and volume-based particle size distributions for
 620 powdered cassava bagasse.

Parameters	Average values	
	Number based	Volume based
$D_{0,1}$ (μm)	2.1	39.3
$D_{0,5}$ (μm)	2.9	155.5
$D_{0,9}$ (μm)	6.1	340.1
De Brouckere mean diameter – $D_{4,3}$ (μm)	119.7	175.5
Sauter mean diameter – $D_{3,2}$ (μm)	53.4	66.3
Span	1.401	1.935

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640 **Table 3.** Fitting parameters of Herschel-Bulkley model for acid suspensions at
 641 intermediate conditions.

Temperature (K)	Parameters	Cassava bagasse concentration, X_S								
		2%			6%			10%		
		Acid concentration $[H_3PO_4]$, X_A								
		2%	6%	10%	2%	6%	10%	2%	6%	10%
278.13	τ_0 (Pa)	0.2944	0.7862	1.1915	0.9188	1.6739	2.5462	2.2481	2.7290	3.2157
	k (Pa·s ^{<i>n</i>})	0.3730	0.8157	1.2886	1.7530	2.0878	2.2757	4.1297	4.6276	4.9354
	n (-)	1.0033	1.0049	0.9999	0.9687	0.9559	0.9621	0.8889	0.8762	0.8509
	R_{adj}^2	0.9999	0.9999	0.9999	0.9999	0.9999	0.9998	0.9999	0.9999	0.9999
298.13	τ_0 (Pa)	0.2399	0.7788	1.1225	0.6610	0.7291	2.4177	1.1702	1.6657	2.1652
	k (Pa·s ^{<i>n</i>})	0.2965	0.7291	1.2204	1.0540	1.7695	1.6891	2.5929	3.0772	3.4120
	n (-)	0.9990	1.0058	0.9966	0.9809	0.9253	0.9686	0.8878	0.8767	0.8512
	R_{adj}^2	0.9999	0.9999	0.9999	0.9999	0.9994	0.9998	0.9999	0.9999	0.9999
318.13	τ_0 (Pa)	0.2376	0.6803	1.1625	0.4719	1.4434	1.7694	0.7119	1.2045	1.6431
	k (Pa·s ^{<i>n</i>})	0.2469	0.7073	1.1595	0.7370	1.0349	1.6120	1.7670	2.1613	2.6310
	n (-)	1.0019	0.9989	0.9988	0.9809	0.9864	0.9405	0.8938	0.8701	0.8531
	R_{adj}^2	0.9999	0.9999	0.9999	0.9999	0.9998	0.9987	0.9999	0.9999	0.9999

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657 **Table 4.** Fitting parameters of an Arrhenius-type equation for apparent viscosity ($\eta_{app,10s^{-1}}$

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Cassava bagasse concentration, X_S (%)	Parameters	Acid concentration, X_A (%)					
		0	2	4	6	8	10
0	A_0 (Pa·s)	9.82×10^{-7}	9.26×10^{-7}	1.03×10^{-6}	1.02×10^{-6}	2.52×10^{-6}	1.49×10^{-6}
	E_a (kJ·mol ⁻¹)	17.22	17.36	17.24	17.26	15.10	16.40
	R_{adj}^2	0.9925	0.9919	0.9766	0.9793	0.9544	0.9953
4	A_0 (Pa·s)	1.41×10^{-4}	4.34×10^{-4}	0.0240	0.0582	0.1326	0.0472
	E_a (kJ·mol ⁻¹)	19.76	17.92	8.99	7.35	5.72	8.54
	R_{adj}^2	0.9995	0.8604	0.9858	0.9909	0.9651	0.9538
6	A_0 (Pa·s)	6.08×10^{-4}	0.0023	0.0142	0.0256	0.0213	0.0916
	E_a (kJ·mol ⁻¹)	18.16	15.31	11.23	10.14	10.86	7.49
	R_{adj}^2	0.9993	0.9953	0.9954	0.9907	0.9835	0.9758
8	A_0 (Pa·s)	1.01×10^{-3}	0.0026	0.0066	0.0130	0.0223	0.0342
	E_a (kJ·mol ⁻¹)	17.98	16.02	13.97	12.51	11.40	10.52
	R_{adj}^2	0.9995	0.9970	0.9982	0.9840	0.9958	0.9961
10	A_0 (Pa·s)	1.50×10^{-3}	0.0029	0.0061	0.0075	0.0121	0.0236
	E_a (kJ·mol ⁻¹)	17.84	16.38	14.66	14.35	13.27	11.74
	R_{adj}^2	0.9999	0.9964	0.9950	0.9916	0.9978	0.9951

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Table S1. Fitting parameters of Herschel-Bulkley model for acid suspensions.

Temperature (K)	Parameters	Cassava bagasse concentration, X_S						Cassava bagasse concentration, X_S					
		0%						2%					
		Acid concentration [H ₃ PO ₄], X_A						Acid concentration [H ₃ PO ₄], X_A					
		0%	2%	4%	6%	8%	10%	0%	2%	4%	6%	8%	10%
278.13	τ_0 (Pa)	0.0000	0.0000	0.0000	0.0000	-0.0001	0.0000	0.0450	0.2944	0.5319	0.7863	0.9145	1.1915
	k (Pa·s ^{<i>n</i>})	0.0017	0.0017	0.0018	0.0018	0.0018	0.0018	0.1518	0.3730	0.6042	0.8157	1.0818	1.2886
	n (-)	0.9985	0.9995	0.9996	1.0003	0.9996	0.9997	1.0000	1.0033	1.0005	1.0049	0.9959	0.9999
	R_{adj}^2	1.0000	1.0000	1.0000	1.0000	1.0000	1.0000	1.0000	1.0000	1.0000	1.0000	1.0000	1.0000
288.13	τ_0 (Pa)	-0.0002	-0.0002	-0.0002	-0.0001	-0.0001	-0.0001	0.0458	0.2941	0.5737	0.6257	0.9571	1.6040
	k (Pa·s ^{<i>n</i>})	0.0013	0.0013	0.0014	0.0014	0.0014	0.0014	0.0977	0.3250	0.5280	0.8205	1.0194	1.1292
	n (-)	0.9968	0.9986	0.9990	0.9995	0.9988	0.9995	1.0108	1.0016	1.0124	0.9898	0.9997	1.0195
	R_{adj}^2	1.0000	1.0000	1.0000	1.0000	1.0000	1.0000	1.0000	0.9999	0.9999	0.9999	1.0000	0.9999
298.13	τ_0 (Pa)	-0.0001	-0.0001	-0.0002	-0.0001	0.0000	0.0001	0.0140	0.2399	0.4617	0.7788	0.9515	1.1225
	k (Pa·s ^{<i>n</i>})	0.0010	0.0010	0.0010	0.0010	0.0011	0.0011	0.0675	0.2966	0.5229	0.7291	0.9745	1.2204
	n (-)	0.9987	0.9976	0.9975	0.9984	1.0012	1.0015	1.0012	0.9990	1.0000	1.0058	1.0007	0.9966
	R_{adj}^2	1.0000	1.0000	1.0000	1.0000	1.0000	1.0000	1.0000	1.0000	1.0000	1.0000	1.0000	1.0000
308.13	τ_0 (Pa)	0.0000	0.0000	0.0001	0.0000	0.0001	0.0001	0.0044	0.2354	0.4659	0.6713	0.9549	1.2033
	k (Pa·s ^{<i>n</i>})	0.0008	0.0008	0.0009	0.0009	0.0008	0.0009	0.0418	0.2695	0.4954	0.7350	0.9396	1.1801
	n (-)	1.0009	1.0009	1.0011	1.0002	1.0008	1.0004	0.99895	0.99997	1.00023	0.99659	1.00257	0.99914
	R_{adj}^2	1.0000	1.0000	1.0000	1.0000	1.0000	1.0000	0.99999	0.99999	0.99999	0.99999	0.99999	0.99999
318.13	τ_0 (Pa)	0.0000	0.0000	0.0000	0.0000	0.0001	-0.0001	0.00398	0.23763	0.46276	0.68030	0.95586	1.16253
	k (Pa·s ^{<i>n</i>})	0.0007	0.0007	0.0008	0.0008	0.0008	0.0008	0.02181	0.24688	0.47494	0.70733	0.91370	1.15954
	n (-)	0.9990	0.9992	0.9991	0.9999	1.0024	0.9975	0.99999	1.00194	1.00023	0.99887	1.00342	0.99885
	R_{adj}^2	1.0000	1.0000	1.0000	1.0000	1.0000	1.0000	0.99999	0.99999	0.99999	0.99999	0.99999	0.99999

Table S1. Fitting parameters of Herschel-Bulkley model for acid suspensions (cont.).

Temperature (K)	Parameters	Cassava bagasse concentration, X_S						Cassava bagasse concentration, X_S					
		4%						6%					
		Acid concentration [H ₃ PO ₄], X_A						Acid concentration [H ₃ PO ₄], X_A					
		0%	2%	4%	6%	8%	10%	0%	2%	4%	6%	8%	10%
278.13	τ_0 (Pa)	0.2599	0.4495	0.9145	1.0956	1.8596	1.0317	0.6960	0.9188	2.8586	1.6739	0.7760	2.5462
	k (Pa·s ^{<i>n</i>})	0.6957	0.9602	1.0857	1.3575	1.3574	2.0760	1.5026	1.7530	1.4730	2.0878	2.8081	2.2757
	n (-)	1.0000	0.9879	0.9980	0.9805	1.0069	0.9334	0.9989	0.9687	1.0214	0.9559	0.9123	0.9621
	R_{adj}^2	0.9999	0.9999	0.9999	0.9999	0.9995	0.9987	0.9999	0.9999	0.9989	0.9999	0.9995	0.9998
288.13	τ_0 (Pa)	0.2314	0.5131	0.5280	0.8569	1.5400	1.4356	0.3276	0.8076	0.9696	1.1966	1.4638	1.4775
	k (Pa·s ^{<i>n</i>})	0.5158	0.7115	1.0247	1.2155	1.3048	1.6433	1.1906	1.3062	1.5926	1.7940	2.0352	2.3533
	n (-)	1.0015	1.0093	0.9868	0.9796	0.9894	0.9643	0.9886	0.9786	0.9561	0.9586	0.9506	0.9254
	R_{adj}^2	0.9999	0.9999	0.9999	0.9999	0.9999	0.9999	0.9999	0.9999	0.9999	0.9999	0.9999	0.9999
298.13	τ_0 (Pa)	0.1121	0.3466	0.8461	0.8630	0.6271	1.7135	0.4136	0.6611	0.9786	0.7291	2.1004	2.4177
	k (Pa·s ^{<i>n</i>})	0.4014	0.6272	0.7772	1.0703	1.5163	1.3401	0.8984	1.0540	1.2846	1.7695	1.5176	1.6891
	n (-)	0.9990	0.9970	1.0074	0.9813	0.9347	0.9958	0.9910	0.9809	0.9691	0.9252	0.9814	0.9686
	R_{adj}^2	0.9999	0.9999	0.9999	0.9999	0.9995	0.9999	0.9998	0.9999	0.9999	0.9994	0.9998	0.9998
308.13	τ_0 (Pa)	0.0803	0.3243	0.5423	0.7405	1.0065	1.9468	-0.0558	0.4676	0.8977	0.3457	1.6865	1.4404
	k (Pa·s ^{<i>n</i>})	0.3075	0.5300	0.7609	0.9914	1.2097	1.1813	0.8166	0.9371	1.1065	1.6454	1.3523	1.8294
	n (-)	1.0000	0.9999	0.9982	0.9779	0.9799	1.0078	0.9630	0.9654	0.9704	0.9096	0.9775	0.9332
	R_{adj}^2	0.9999	0.9999	0.9999	0.9999	0.9999	0.9993	0.9982	0.9999	0.9999	0.9993	0.9998	0.9999
318.13	τ_0 (Pa)	0.0621	0.0621	0.5196	0.7061	1.3139	1.3139	0.2200	0.4719	0.4472	1.4434	1.4753	1.7694
	k (Pa·s ^{<i>n</i>})	0.2349	0.2349	0.6900	0.9373	0.9975	0.9975	0.5640	0.7370	1.1339	1.0349	1.3775	1.6121
	n (-)	1.0019	1.0019	0.9899	0.9754	0.9958	0.9958	0.9910	0.9809	0.9333	0.9864	0.9446	0.9405

R_{adj}^2	0.9999	0.9999	0.9999	0.9999	0.9999	0.9999	0.9999	0.9998	0.9999	0.9994	0.9998	0.9987	0.9987
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Table S1. Fitting parameters of Herschel-Bulkley model for acid suspensions (cont.).

Temperature (K)	Parameters	Cassava bagasse concentration, X_S						Cassava bagasse concentration, X_S					
		8%						10%					
		Acid concentration $[H_3PO_4]$, X_A						Acid concentration $[H_3PO_4]$, X_A					
		0%	2%	4%	6%	8%	10%	0%	2%	4%	6%	8%	10%
278.13	τ_0 (Pa)	1.2434	1.5029	1.7428	1.9488	2.1958	2.3582	2.0172	2.2481	2.5864	2.7290	2.8890	3.2157
	k (Pa·s ^{<i>n</i>})	2.5922	2.8200	3.0452	3.2976	3.5105	3.7603	3.9539	4.1297	4.2587	4.6276	4.8015	4.9354
	n (-)	0.9467	0.9422	0.9331	0.9251	0.9172	0.9061	0.9019	0.8889	0.8779	0.8762	0.8598	0.8509
	R_{adj}^2	0.9999	0.9999	0.9999	0.9999	0.9999	0.9999	0.9999	0.9999	0.9999	0.9999	0.9999	0.9999
288.13	τ_0 (Pa)	0.8310	1.1086	1.3362	1.5162	1.7556	2.0732	1.3932	1.5755	1.8688	2.0819	2.3055	2.5388
	k (Pa·s ^{<i>n</i>})	1.9812	2.2287	2.4905	2.6319	2.9148	3.1205	3.0392	3.2856	3.3955	3.6324	3.8715	4.0748
	n (-)	0.9431	0.9441	0.9402	0.9141	0.9149	0.9099	0.9022	0.8918	0.8769	0.8667	0.8615	0.8503
	R_{adj}^2	0.9999	0.9999	0.9999	0.9999	0.9999	0.9999	0.9999	0.9999	0.9999	0.9999	0.9999	0.9999
298.13	τ_0 (Pa)	0.6121	0.8473	1.0247	1.2638	1.5065	1.8048	0.9804	1.1702	1.4505	1.6657	1.8925	2.1652
	k (Pa·s ^{<i>n</i>})	1.5454	1.7828	2.0379	2.2555	2.4805	2.6753	2.3847	2.5929	2.7382	3.0772	3.2096	3.4120
	n (-)	0.9481	0.9481	0.9328	0.9231	0.9159	0.9089	0.9022	0.8878	0.8749	0.8767	0.8595	0.8512
	R_{adj}^2	0.9999	0.9999	0.9999	0.9999	0.9999	0.9999	0.9999	0.9999	0.9999	0.9999	0.9999	0.9999
308.13	τ_0 (Pa)	0.4228	0.6548	0.8581	1.0956	1.3355	1.6314	0.6981	0.8985	1.1727	1.3866	1.6181	1.8855
	k (Pa·s ^{<i>n</i>})	1.2375	1.4307	1.7054	1.9325	2.1529	2.3617	1.8916	2.0678	2.3450	2.5340	2.7293	2.9747
	n (-)	0.9487	0.9322	0.9318	0.9241	0.9159	0.9099	0.9022	0.8818	0.8869	0.8697	0.8595	0.8562
	R_{adj}^2	0.9999	0.9999	0.9999	0.9999	0.9999	0.9999	0.9999	0.9999	0.9999	0.9999	0.9999	0.9999
318.13	τ_0 (Pa)	0.3121	0.5444	0.7821	0.9828	1.2206	1.5091	0.5098	0.7120	0.9925	1.2045	1.4493	1.6431

k (Pa·s ⁿ)	0.9846	1.2212	1.4396	1.6845	1.9103	2.1081	1.5287	1.7670	1.9222	2.1614	2.3626	2.6310
n (-)	0.9471	0.9421	0.9343	0.9241	0.9169	0.9090	0.9032	0.8938	0.8782	0.8701	0.8603	0.8532
R_{adj}^2	0.9999	0.9999	0.9999	0.9999	0.9999	0.9999	0.9999	0.9999	0.9999	0.9999	0.9999	0.9999

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Table S2. Analysis of variance (ANOVA) for yield stress:

Factor	Sum of squares	Degree of freedom	Mean square	F	p_{value}
T (linear)	6.92	1	6.92	85.84	<0.001
T (quadratic)	0.4714	1	0.4714	5.85	0.016
X_S (linear)	49.73	1	49.73	616.73	<0.001
X_S (quadratic)	0.3971	1	0.3971	4.92	0.027
X_A (linear)	23.97	1	23.97	297.27	<0.001
T (linear) \times X_S (linear)	5.77	1	5.77	71.62	<0.001
X_S (linear) \times X_A (linear)	1.70	1	1.70	21.12	<0.001
Error	13.87	172	0.08		
Total sum of square	102.83	179			

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Table S3. Analysis of variance (ANOVA) for consistency coefficient:

Factor	Sum of squares	Degree of freedom	Mean square	<i>F</i>	<i>p</i> value
<i>T</i> (linear)	19.12	1	19.12	545.84	<0.001
<i>T</i> (quadratic)	0.4887	1	0.4887	13.95	<0.001
<i>X_S</i> (linear)	183.99	1	183.99	5253.02	<0.001
<i>X_S</i> (quadratic)	2.68	1	2.68	76.44	<0.001
<i>X_A</i> (linear)	16.92	1	16.92	483.10	<0.001
<i>T</i> (linear) × <i>X_S</i> (linear)	14.69	1	14.69	419.36	<0.001
<i>X_S</i> (linear) × <i>X_A</i> (linear)	1.40	1	1.40	39.95	<0.001
Error	6.02	172	0.03		
Total sum of square	245.32	179			

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Table S4. Analysis of variance (ANOVA) for flow index behavior:

Factor	Sum of squares	Degree of freedom	Mean square	<i>F</i>	<i>p</i> _{value}
X_S (linear)	0.3202	1	0.3202	2283.31	<0.001
X_S (quadratic)	0.0469	1	0.0469	334.25	<0.001
X_A (linear)	0.0139	1	0.0139	99.38	<0.001
X_S (linear) \times X_A (linear)	0.0075	1	0.0075	53.36	<0.001
Error	0.0245	175	0.0001		
Total sum of square	0.4130	179			

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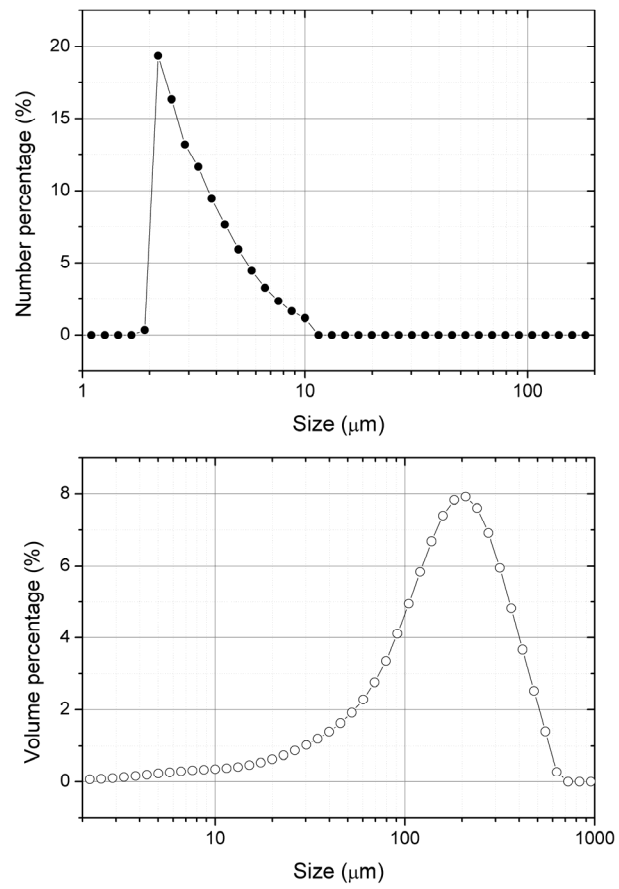
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Table S5. Analysis of variance (ANOVA) for apparent viscosity at shear rate of 10 s^{-1} :

Factor	Sum of squares	Degree of freedom	Mean square	F	p_{value}
T (linear)	15.20	1	15.20	508.16	<0.001
T (quadratic)	0.4205	1	0.4205	14.06	<0.001
X_S (linear)	120.47	1	120.47	4027.48	<0.001
X_A (linear)	13.70	1	13.70	458.27	<0.001
T (linear) \times X_S (linear)	10.36	1	10.36	346.21	<0.001
X_S (linear) \times X_A (linear)	0.2145	1	0.2145	7.17	0.008
Error	5.17	173	0.03		
Total sum of square	165.54	179			

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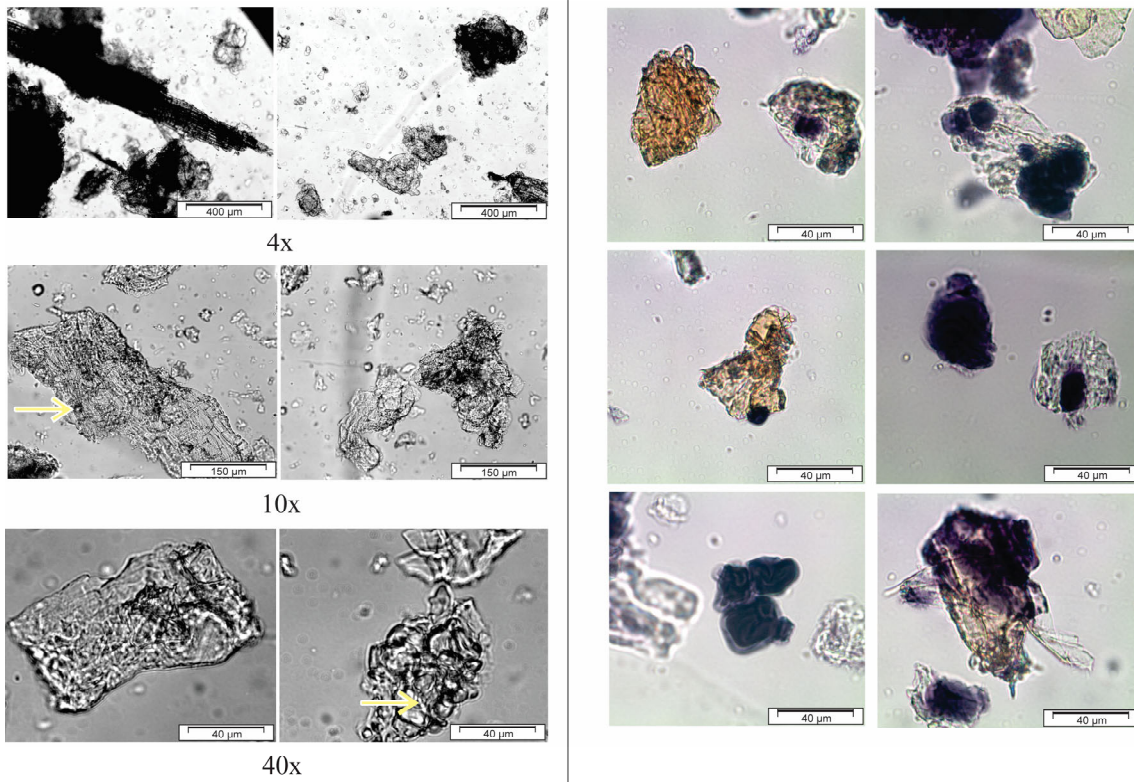


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707 **Figure 1.** Number (A) and volume (B) based particle size distribution for powdered

708 cassava bagasse.

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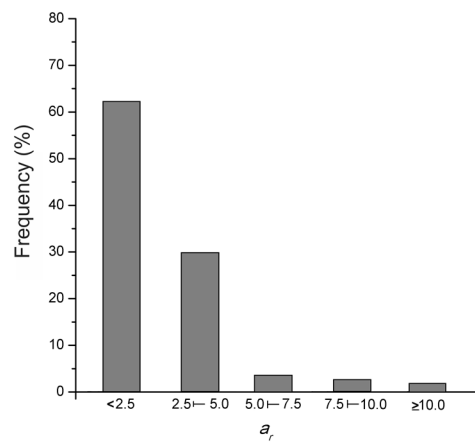


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711 **Figure 2.** Optical microscopic images for cassava bagasse at 4x, 10x and 40x of

712 magnification (A) and for cassava bagasse dyed with lugol at 40x of magnification (B).

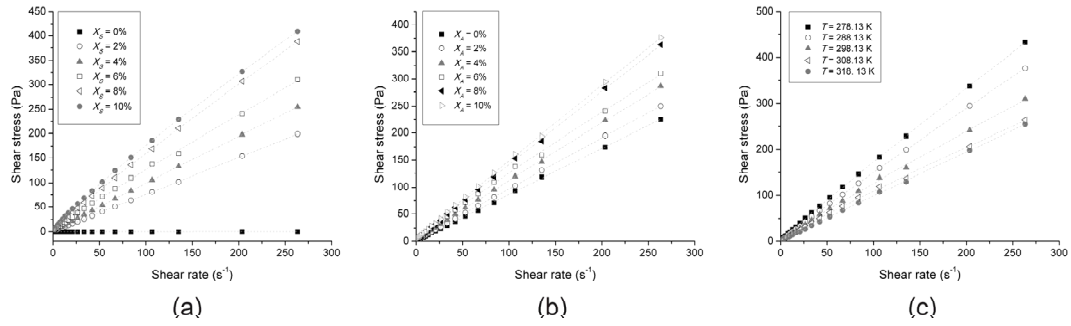
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715 **Figure 3.** Distribution of the aspect ratio (a_r) values for powdered cassava bagasse
 716 particles.

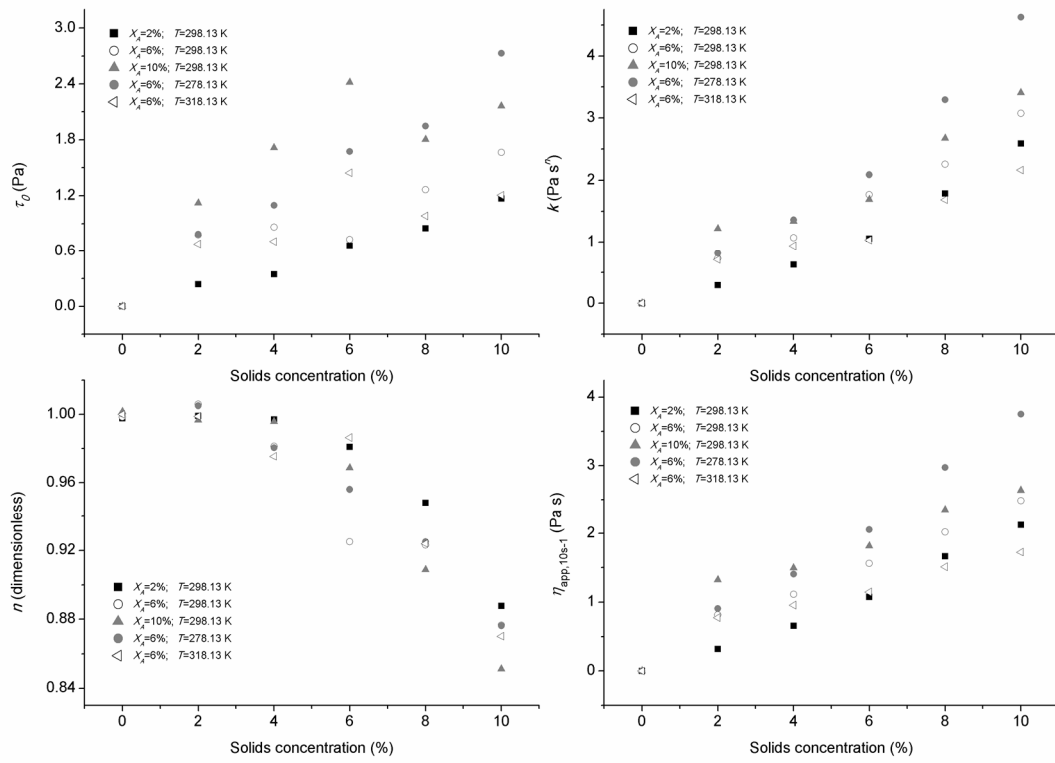
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719 **Figure 4.** Rheograms of the acid suspensions containing cassava bagasse fitted to
 720 Herschel-Bulkley model. (a) $X_A=6\%$ at $T=298.13$ K; (b) $X_S=6\%$ at $T=298.13$ K; and (c)
 721 $X_S=6\%$ and $X_A=6\%$.

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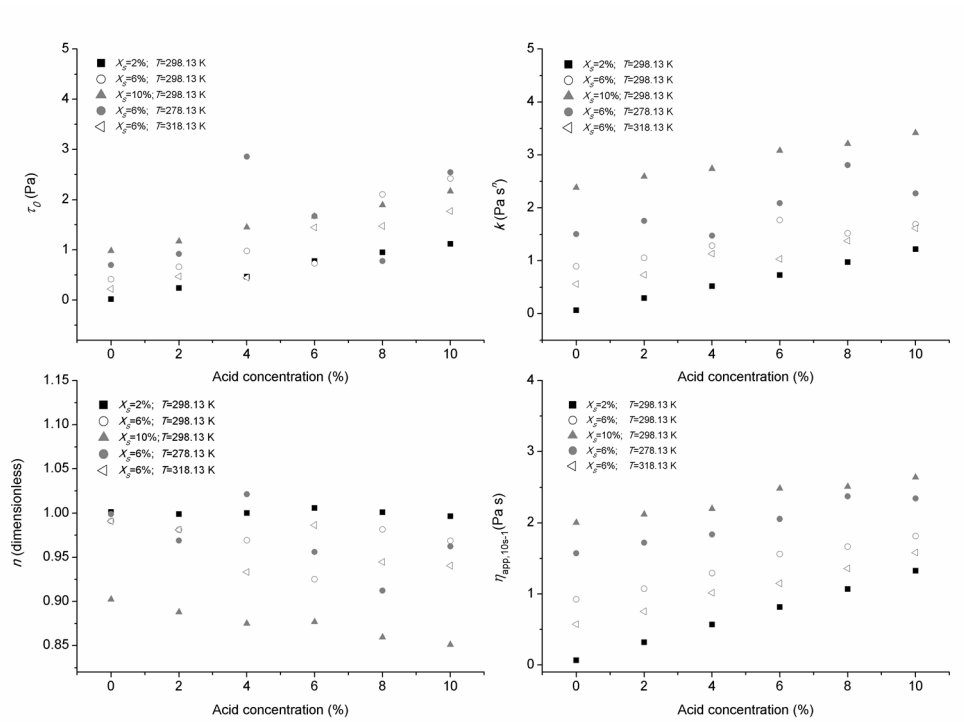


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725 **Figure 5.** Yield stress (τ_0) (A), consistency coefficient (k) (B), flow behavior index (n)726 (C) and apparent viscosity ($\eta_{app,10s^{-1}}$) (D) as functions of cassava bagasse concentration

727 at different acid concentrations and temperatures.

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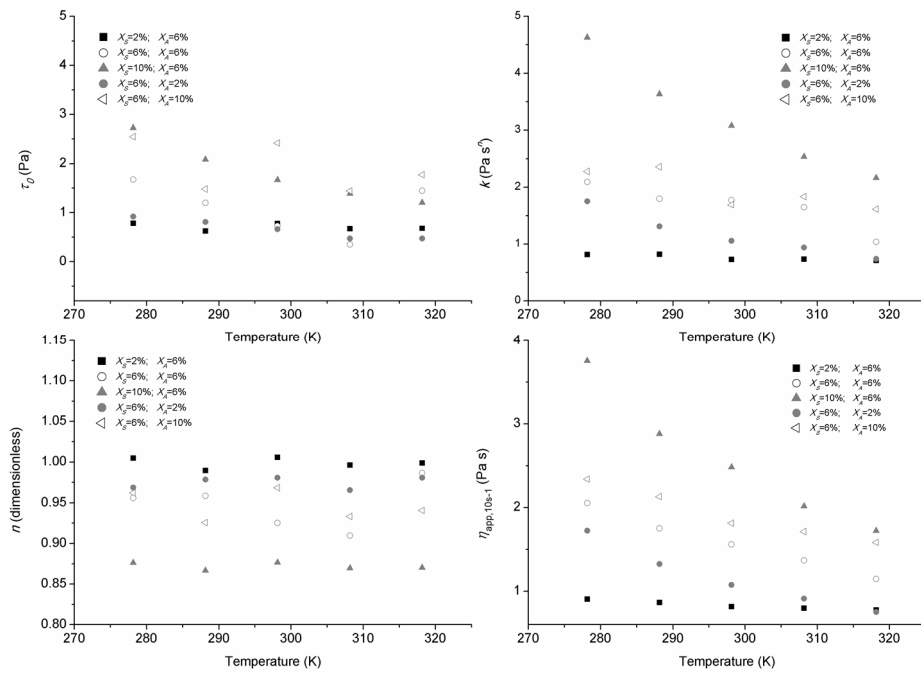
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730 **Figure 6.** Yield stress (τ_0), consistency coefficient (k), flow behavior index (n) and

731 apparent viscosity ($\eta_{app,10s^{-1}}$) as functions of phosphoric acid concentration at different

732 solids concentrations and temperatures.

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735 **Figure 7.** Yield stress (τ_0), consistency coefficient (k), flow behavior index (n) and

736 apparent viscosity ($\eta_{app,10s^{-1}}$) as functions of absolute temperature at different acid and

737 solids concentrations.

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