

Document downloaded from:

<http://hdl.handle.net/10251/161694>

This paper must be cited as:

Giménez, JB.; Martí, N.; Bouzas, A.; Ferrer, J.; Seco, A. (2020). A mathematical approach to predict the solids concentration in anaerobic membrane bioreactors (AnMBR): Evaluation of the volatile solids solubilization. *Journal of Environmental Management*. 271:1-8.  
<https://doi.org/10.1016/j.jenvman.2020.110983>



The final publication is available at

<https://doi.org/10.1016/j.jenvman.2020.110983>

Copyright Elsevier

Additional Information

1 **A MATHEMATICAL APPROACH TO PREDICT THE SOLIDS**  
2 **CONCENTRATION IN ANAEROBIC MEMBRANE**  
3 **BIOREACTOS (AnMBR): EVALUATION OF THE VOLATILE**  
4 **SOLIDS SOLUBILIZATION.**

5

6 Juan B. Giménez<sup>a\*</sup>, Nuria Martí<sup>b</sup>, Alberto Bouzas<sup>b</sup>, José Ferrer<sup>a</sup>, Aurora Seco<sup>b</sup>.

7

8 <sup>a</sup>Institut Universitari d'Investigació d'Enginyeria de l'Aigua i Medi Ambient,  
9 IIAMA, Universitat Politècnica de València, Camí de Vera s/n, 46022 Valencia, Spain  
10 (e-mail: [juagiga5@iiama.upv.es](mailto:juagiga5@iiama.upv.es); [jferrer@hma.upv.es](mailto:jferrer@hma.upv.es))

11 <sup>b</sup>Departament d'Enginyeria Química, Escola Tècnica Superior d'Enginyeria,  
12 Universitat de València, Avinguda de la Universitat s/n, 46100 Burjassot, Valencia,  
13 Spain (e-mail: [nuria.marti@uv.es](mailto:nuria.marti@uv.es); [alberto.bouzas@uv.es](mailto:alberto.bouzas@uv.es); [aurora.seco@uv.es](mailto:aurora.seco@uv.es))

14 **Abstract**

15 Anaerobic Membrane Bioreactors (AnMBR) are gaining attention as a suitable  
16 approach for sustainable low-strength wastewater treatment, as they bring together the  
17 advantages of both anaerobic treatments and membrane bioreactors. However,  
18 increasing the sludge retention time (SRT) necessary to favor hydrolysis increases the  
19 suspended solids concentration potentially leading to decreased permeate flux.  
20 Therefore, the availability of a mathematical approach to predict the solids  
21 concentration within an AnMBR can be very useful. In this work, a mathematical model

---

\* Corresponding Author: tel. +34 96 354 30 85, e-mail: [juagiga5@iiama.upv.es](mailto:juagiga5@iiama.upv.es)

22 describing the volatile solids concentration within the reactor as a function of the  
23 operating parameters and the influent characteristics is developed. The solubilization of  
24 organic particulates was clearly influenced by temperature and the SRT, whereas the  
25 hydraulic retention time influence was negligible. Furthermore, the activation energy  
26 value of about  $20 \text{ kJ}\cdot\text{mol}^{-1}$  confirms the idea that diffusion of hydrolytic enzymes from  
27 the bulk solution to the particle surface is the rate-limiting step of hydrolysis.

28 **Keywords:** Particulates hydrolysis, mathematical modelling, solubilization constant,  
29 solids prediction, AnMBR.

## 30 **1. INTRODUCTION.**

31 The current trend in wastewater management is moving towards the development of  
32 sustainable technologies aimed to reclaim both water and resources (i.e. nutrients and  
33 energy) from wastewater (Daigger, 2008; Kleerebezem and van Loosdrecht, 2007;  
34 Larsen et al., 2009; Li and Yu, 2011). Anaerobic treatment is regarded as one of the  
35 most attractive options for the development of sustainable wastewater treatment  
36 systems, since they have the potential to both being net energy producers and enabling  
37 nutrients to be recovered whilst reducing the excess-sludge production (Chen, 2020).

38 Nevertheless, anaerobic treatment of Municipal wastewater (MWW) streams requires  
39 an efficient biomass retention method. Otherwise, the combination of typical high flow-  
40 rates and low Chemical Oxygen Demand (COD) concentrations on MWW, and the low  
41 growth rates of organisms carrying out anaerobic degradation of organic matter at  
42 moderate temperatures (medium latitude temperate climates) would result in prohibitive  
43 reaction volumes to maintain the required sludge retention time (SRT).

44 Anaerobic membrane bioreactors (AnMBR) exhibit an outstanding capacity for  
45 particulate matter retention, enabling AnMBR to efficiently decouple hydraulic

46 retention time (HRT) from SRT. However, one important issue in the design of MBR-  
47 based systems is particles, since it affects the performance of the membrane.

48 Hydrolysis of particulate organic material has been considered the rate-limiting step  
49 in anaerobic digestion (Nabi et al., 2020, 2019; Pavlostathis and Giraldo-Gomez, 1991),  
50 although some authors have emphasized that the hydrolytic process still remains as the  
51 least well-defined step (Gavala et al., 2003; Miron et al., 2000). The general kinetic  
52 term of hydrolysis encompasses disintegration, solubilization and enzymatic hydrolysis  
53 processes in most of the practical cases presented in literature (Batstone et al., 2002a).  
54 Furthermore, hydrolysis rates decline with temperature (Lettinga et al., 2001), requiring  
55 longer SRTs for hydrolysis to occur at sub-mesophilic temperatures. However,  
56 increasing the SRT, while keeping the HRT constant, increases the suspended solids  
57 concentration, potentially leading to decreased permeate flux (Smith et al., 2012).  
58 Therefore, an accurate prediction of the solids concentration might be very useful to  
59 achieve a proper AnMBR operation.

60 In spite of the complexity of anaerobic processes, mechanistic-modelling based on  
61 the understanding of underlying biological processes is nowadays a well-established  
62 modelling approach (Batstone et al., 2015; Robles et al., 2018). Furthermore, most of  
63 the empirical/ semi-empirical modelling approaches tackling the physical process (i.e.  
64 filtration) can accurately reproduce the sludge effect over the membrane performance.  
65 However, these models use parameters requiring specific laboratory procedures and  
66 equipment (e.g. SMP), that might result in continuity issues when interfaced to a given  
67 biological model (Robles et al., 2013). Conversely, some authors (Ludwig et al., 2012;  
68 Robles et al., 2013; Sarioglu et al., 2012) are developing new models easily combinable  
69 with biological models, using the most critical variables to membrane fouling as model  
70 parameters (i.e., shear intensity and solids concentration).

71 This work aims at describing a mathematical approach to predict the solids  
72 concentration within AnMBR, depending on the operating conditions. A 700-days  
73 period, during which the system performance was evaluated under different operating  
74 conditions, was used in order to fit the experimental data to the model prediction. A  
75 non-linear regression was used to estimate the volatile-solids solubilization constant to  
76 assess its variability with the operating conditions.

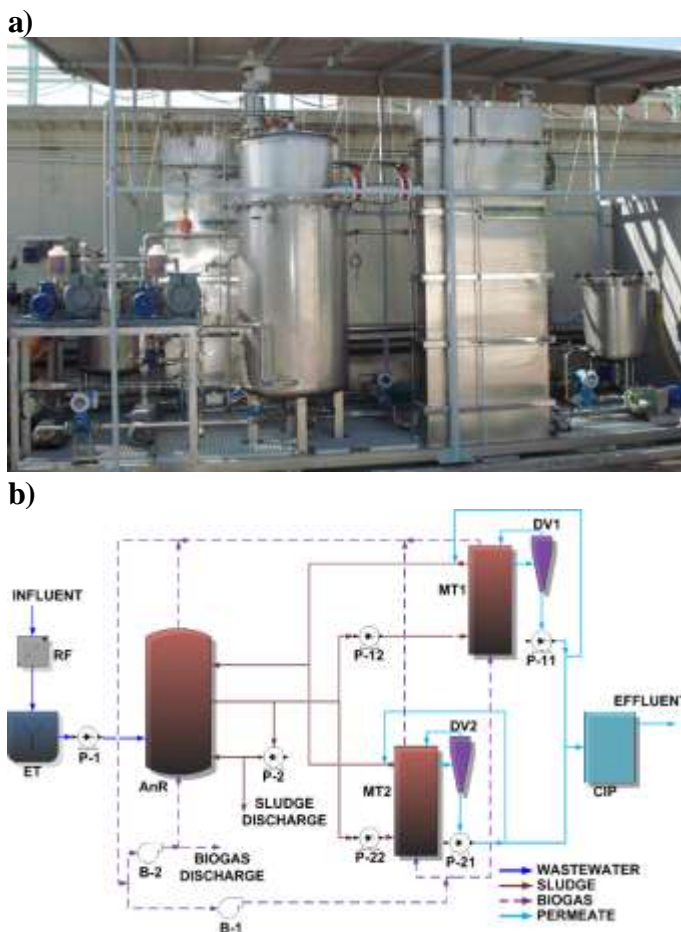
## 77 **2. MATERIALS AND METHODS.**

### 78 **2.1. Demonstration-scale plant description.**

79 The present work was carried out in a demonstration-scale plant placed in the  
80 “Barranco del Carraixet” wastewater treatment plant (WWTP), in Alboraya (Valencia),  
81 Spain. The demonstration-scale plant was fed with the effluent of the pre-treatment  
82 (grease and sand removal) of the full-scale WWTP. The AnMBR mainly consisted of a  
83 1.3 m<sup>3</sup> jacketed anaerobic reactor connected to two 0.8 m<sup>3</sup> membrane tanks. Each  
84 membrane tank included one commercial ultrafiltration hollow-fiber membrane system  
85 (PURON®, Koch Membrane Systems, 0.05 µm pore size, 30 m<sup>2</sup> total filtering area). A  
86 rotofilter with a screen size of 0.5 mm was installed as pre-treatment. One equalization  
87 tank (0.3 m<sup>3</sup>) and one Clean-In-Place tank (0.2 m<sup>3</sup>) were also included as main elements  
88 of the pilot plant. A fraction of the produced biogas (0.5 Nm<sup>3</sup> per cubic meter of reactor  
89 volume) was continuously recycled to the anaerobic reactor in order to improve the  
90 stirring conditions and to favor the stripping of the produced gases from the liquid  
91 phase, whereas another fraction of the produced biogas (0.2-0.3 Nm<sup>3</sup>/h per m<sup>2</sup> of  
92 membrane) was also continuously recycled to the bottom of the hollow-fiber  
93 membranes in order to minimize the cake layer formation. Also, a degassing vessel was  
94 installed between each membrane tank and the corresponding vacuum pump to recover  
95 the bubbles of biogas in the permeate. A temperature control system was also

196 implemented, consisting of a warm-water tank equipped with a 6 kW resistance and a  
 197 thermostat switch that maintained the water temperature at 65 °C. The temperature in  
 198 the anaerobic reactor was effectively controlled by a water pump that enabled the warm  
 199 water to flow through the reactor jacket depending on the difference between the reactor  
 200 temperature and the set point.

201 Figure 1 shows a general view (a), as well as a process flow diagram (b) of the  
 202 demonstration-scale plant. Further details on the demonstration-scale plant components  
 203 and operation can be found in Giménez et al. (2011).



204 Figure 1. (a) General view and (b) Process Flow Diagram of the AnMBR demonstration plant. RF:  
 205 Rotofilter; ET; Equalization Tank; AnR: Anaerobic Reactor; B: Blower; P: Pump; MT: Membrane Tank;  
 206 DV: Degassing Vessel; CIP: Clean-In-Place.

## 207 2.2. Volatile Suspended Solids mass balance.

208 The general form of the volatile solids mass balance can be written according to

209 Equation 1:

$$(Q_{WS} \cdot [VS]_R + Q_E \cdot [VS]_E) - Q_I \cdot [VS]_I + V_R \cdot \frac{d[VS]_R}{dt} = V_R \cdot \rho_H \quad \text{Equation 1}$$

110 Where  $Q_{WS}$ ,  $Q_E$  and  $Q_I$  are the waste sludge, effluent and influent volumetric flow  
 111 rates, respectively.  $[VS]_R$ ,  $[VS]_E$  and  $[VS]_I$  stand for the volatile-solids concentration in  
 112 the reactor, effluent and influent, respectively.  $V_R$  is the volume of the reactor and  $\rho_H$   
 113 stands for the hydrolysis rate. The hydrolysis process of particulate organic matter  
 114 encompasses several stages (i.e., enzyme production, diffusion, adsorption, reaction and  
 115 enzymatic deactivation). However, first order kinetics are the most widespread in order  
 116 to describe the hydrolysis process, since more complex kinetic equations only provide  
 117 slightly better results (Batstone et al., 2002a). Therefore, the hydrolysis kinetics can be  
 118 described by Equation 2:

$$\rho_H = k_H \cdot [SS_{BD}] \quad \text{Equation 2}$$

119 Where  $[SS_{BD}]$  stands for the biodegradable suspended solids concentration (mg  
 120  $SS \cdot L^{-1}$ ), and  $k_H$  is the first order hydrolysis constant ( $day^{-1}$ ) that encompasses the  
 121 cumulative effect of every single process involved in the overall enzymatic hydrolysis  
 122 process. The rate of hydrolysis depends on pH, temperature, concentration of  
 123 hydrolyzing biomass and characteristics of particulates (Pavlostathis and Giraldo-  
 124 Gomez, 1991). In order to set the volatile solids mass balance out, the following  
 125 assumptions have been made:

- 126 1. Biodegradable suspended solids are comprised within the volatile suspended  
 127 solids (VSS) fraction.
- 128 2. The dissolved fraction of the volatile solids is negligible as compared to the  
 129 suspended fraction. Therefore,  $[VSS] \cong [VS]$ .
- 130 3. The biodegradable fraction of the volatile solids is constant during each period.

131 Therefore, according to assumption 1, a change in the biodegradable suspended  
 132 solids will induce the same change in the VSS, so it is possible to rewrite the hydrolysis  
 133 rate expression as in Equation 3:

$$\rho_H = k_H \cdot [SS_{BD}] \equiv k_H \cdot [VSS_{BD}] \equiv k_H \cdot f_{BD} \cdot [VSS] \quad \text{Equation 3}$$

134  $f_{BD}$  standing for the biodegradable fraction of the volatile solids within the reactor.

135 Therefore, considering assumption 2 and 3, the hydrolysis rate can be expressed as in

136 Equation 4:

$$\rho_H \cong k_H \cdot f_{BD} \cdot [VS] \equiv k_{Sol,VS} \cdot [VS] \quad \text{Equation 4}$$

137  $k_{Sol,VS}$  being (Equation 5):

$$k_{Sol,VS} = k_H \cdot f_{BD} \quad \text{Equation 5}$$

138 Equation 4 states that the hydrolysis rate can be described as a function of the VS  
 139 concentration within the reactor. The proportionality factor, the so-called VS  
 140 solubilization constant ( $k_{Sol,VS}$ ), encompasses both the first order hydrolysis constant  
 141 and a factor that stands for the biodegradable fraction of the volatile solids. The  
 142 hydrolysis constant reflects the cumulative effect of every step involved in the complete  
 143 enzymatic hydrolysis process (enzyme production, diffusion, adsorption, reaction and  
 144 deactivation; (Batstone et al., 2002a)) which has been reported to be a function of pH,  
 145 temperature, hydrolytic biomass concentration and physio-chemical features of the  
 146 substrate, among other parameters (Pavlostathis and Giraldo-Gomez, 1991), whereas  
 147 the biodegradable fraction of the volatile solids depends on the biodegradable  
 148 particulates accumulated within the reactor. Indeed, both parameters will depend on the  
 149 influent MWW features, as well as on the selected operating conditions.

150 The resulting general expression for the volatile solids mass balance in continuous  
 151 stirred tank reactors (CSTR) is given by Equation 6:



$$(Q_{WS} \cdot [VS]_R + Q_E \cdot [VS]_E) - Q_I \cdot [VS]_I + V_R \cdot \frac{d[VS]_R}{dt} = V_R \cdot \rho_H$$

Equation 6

$$\equiv V_R \cdot k_{Sol,VS} \cdot [VS]_R$$

152 A further development of the VS and non-volatile solids (NVS) mass balances in  
 153 AnMBR, where  $[VS]_E$  is negligible, resulted in Equation 7 and Equation 8 for the  
 154 estimation of the VS and NVS concentration within the reactor, respectively:

$$[VS]_R = - \frac{[VS]_I \cdot SRT}{(k_{Sol,VS} \cdot SRT - 1) \cdot HRT}$$

Equation 7

$$+ \left( [VS]_R^0 + \frac{[VS]_I \cdot SRT}{(k_{Sol,VS} \cdot SRT - 1) \cdot HRT} \right) \cdot \exp \left[ \left( k_{Sol,VS} - \frac{1}{SRT} \right) \cdot t \right]$$

$$[NVS]_R = \frac{[NVS]_I \cdot SRT}{HRT}$$

Equation 8

$$+ \left( [NVS]_R^0 - \frac{[NVS]_I \cdot SRT}{HRT} \right) \cdot \exp \left[ - \frac{t}{SRT} \right]$$

155 Where  $[NVS]$ , HRT and SRT stand for the non-volatile solids concentration, the  
 156 hydraulic and the solids retention times, respectively. Subscripts have been used in the  
 157 same way than in Equation 1, whereas the superscript “0” has been used to denote the  
 158 concentration at the beginning of the considered period. Experimental data for  $[VS]_I$ ,  
 159  $[NVS]_I$ ,  $[VS]_R^0$  and  $[NVS]_R^0$  in the different periods can be found in table S1 (support  
 160 material).

161 From the above expressions for the estimation of the VS and NVS concentration, it is  
 162 possible to estimate the VS percentage according to Equation 9:

$$\% VS_R = \frac{[VS]_R}{[VS]_R + [NVS]_R} \cdot 100 = \frac{[VS]_R}{[TS]_R} \cdot 100$$

Equation 9

### 163 2.3. Experimental design.

164 The present work comprises 700 days of AnMBR demonstration plant operation that  
 165 was sub-divided into 10 different operating periods. HRT and SRT were easily  
 166 controlled by modifying the influent and waste-sludge flow-rates, respectively.

167 Temperature was controlled from period P1 to period P5, and in periods P9 and P10.  
 168 Conversely, the temperature control was disconnected during periods P6 to P8, enabling  
 169 the temperature of the system to evolve according to the ambient temperature. Table 1  
 170 shows the operating conditions that were set in each period.

<b>Period</b>	<b>T (°C)</b>	<b>HRT (h)</b>	<b>SRT (d)</b>
<b>P1</b>	33	17	71
<b>P2</b>	33	12	71
<b>P3</b>	33	9	74
<b>P4</b>	33	6	71
<b>P5</b>	20	24	69
<b>P6</b>	29	28	42
<b>P7</b>	23	14	39
<b>P8</b>	17	12	29
<b>P9</b>	20	12	29
<b>P10</b>	20	12	40

171 Table 1. Operating conditions in the 10 periods evaluated.

172 **2.4. Analytical methods.**

173 Total solids (TS), Volatile solids (VS), Total suspended solids (TSS) and volatile  
 174 suspended solids (VSS) for the influent and for the mixed liquor were determined  
 175 according to Standard Methods (APHA et al., 2012).

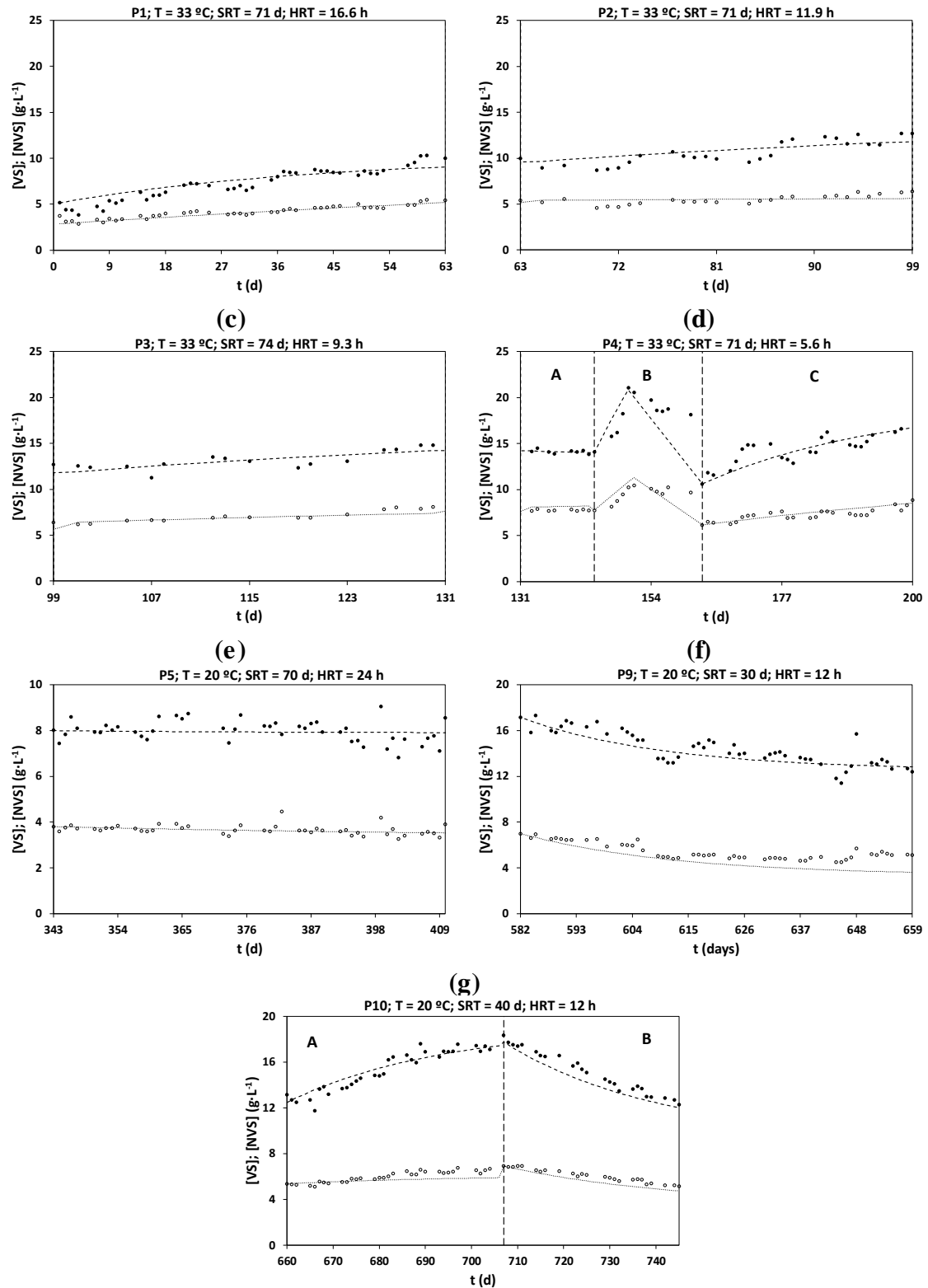
176 **3. RESULTS AND DISCUSSION.**

177 **3.1. Influence of the operating conditions over the VS solubilization process.**

178 As previously stated, both the hydrolysis constant and the biodegradable fraction of  
 179 the volatile solids depend on the influent MWW features, as well as on the selected  
 180 operating conditions. Figure 2 (a) to (g) show the evolution of both the experimental  
 181 (dots) and the estimated (lines) values for the VS and the NVS concentration during  
 182 periods P1 to P5, P9 and P10.

(a)

(b)



● [VS]<sub>Experimental</sub> --- [VS]<sub>Estimated</sub> ○ [NVS]<sub>Experimental</sub> ..... [NVS]<sub>Estimated</sub>

183 Figure 2 Time-evolution of the experimental and fitted Volatile solids [VS] and the Non-Volatile solids  
 184 [NVS] concentration.

185 As it can be seen in Figure 2, the estimated NVS concentration ( $[NVS]_{Estimated}$ )

186 accurately followed the experimental NVS concentration ( $[NVS]_{Experimental}$ ) trend,

187 indicating that the average influent NVS concentration-values ( $[NVS]_i$ ; Table S1) used  
 188 in the NVS mass balance (Equation 8) were representative of the actual influent NVS  
 189 concentration during the different periods. Likewise, it can be assumed that the average  
 190 influent VS concentration values ( $[VS]_i$ ; Table S1) were also representative of the actual  
 191 influent VS concentration during each period. Therefore, the VS mass balance  
 192 (Equation 7) was used to adjust the volatile-solids solubilization constant ( $k_{Sol, VS}$ ) by  
 193 means of the least-squares, non-linear fitting of the estimated VS concentration data  
 194 ( $[VS]_{Estimated}$ ), to the experimental data ( $[VS]_{Experimental}$ ). An evaluation of the influence  
 195 of the operating conditions over  $k_{Sol, VS}$  is presented below.

### 196 **3.1.1. Hydraulic Retention Time.**

197 In order to assess the effect of the HRT on the VS solubilization process, the VS  
 198 solubilization constant was estimated from the non-linear fitting of the VS balance  
 199 concentration data to the analytical VS concentration data during periods P1, P2, P3 and  
 200 P4. As shown in Table 1, SRT and temperature were set to 70 days and 33°C,  
 201 respectively, whereas HRT was decreased from 17 to 6 hours. Table 2 shows the  
 202 estimated values for the VS solubilization constant during periods P1 to P4.

<b>Period</b>	<b>HRT (h)</b>	<b><math>k_{Sol, VS}</math> (d<sup>-1</sup>)</b>
<b>P1</b>	16.6	0.0186
<b>P2</b>	11.9	0.0068
<b>P3</b>	9.3	0.0067
<b>P4</b>	5.6	0.0181

203 Table 2. Variation of the Volatile Solids solubilization constant ( $k_{Sol, VS}$ ) with HRT.

204 The VS solubilization constant was similar during periods P1 and P4, despite the  
 205 three-fold reduction in the HRT. These results evidenced the negligible influence of the  
 206 HRT over the hydrolysis process. On the other hand, the VS solubilization constants  
 207 during periods P2 and P3 were lower than for periods P1 and P4, the lowest value being  
 208 that of period P3.

209 The reduced VS solubilization constant during periods P2 and P3 was attributed to  
210 the domestic-featured influent wastewater, given that P2 and P3 corresponded to the  
211 summer period, in which the industrial fraction of wastewater is significantly reduced.  
212 Indeed, a clear difference in the organic load and composition of the influent wastewater  
213 was observed between summer and the rest of the year (Giménez, 2014), indicating that  
214 the biodegradability of the domestic fraction of the MWW was lower than the industrial  
215 fraction. These results suggest that it was the different influent wastewater features  
216 during summer rather than the HRT what influenced the volatile-solids solubilization  
217 constant  $k_{Sol, VS}$ , by means of the likely different volatile solids biodegradable fraction in  
218 the reactor ( $f_{BD}$ ).

### 219 **3.1.2. Temperature.**

220 Previous works (Giménez et al., 2014, 2012) have reported a clear influence of  
221 temperature on the volatile solids percentage within AnMBR, indicating an uneven  
222 solubilization of the volatile solids as a result of the hydrolysis-rate variation with  
223 temperature. Additionally, the variation in the hydrolysis rate affected the availability of  
224 substrates in the subsequent stages of the anaerobic degradation. Indeed, transitory VFA  
225 accumulation was observed at the beginning of period P5 (data not shown), which was  
226 attributed to the acetogenic and methanogenic unbalanced activities derived from the  
227 temperature change, rather than from the change in the hydrolysis rate. During these  
228 transitory impaired methanogenic-activity periods the high retention capacity of the  
229 membranes helped to wash the VFA out of the system, preventing the AnMBR  
230 acidification (reactor pH was virtually constant). The eventual depletion of VFA  
231 indicated that the balance between the different microbial communities was restored,  
232 and that hydrolysis was again the limiting step in the anaerobic degradation of organic  
233 matter. Thus, both the biogas and waste sludge productions varied accordingly. In this

234 work, the influence of temperature variations over the VS solubilization process was  
235 evaluated by comparing the VS solubilization constants estimated in periods P4 and P5.  
236 The SRT was set at 70 days, whereas temperature was decreased from 33°C, in period  
237 P4, to 20 °C, in P5. HRT was higher in period P5 than in period P4, however, the  
238 solubilization process was not affected by this operating parameter, as previously  
239 reported.

240 Figure 2 (d) and (e) shows the evolution of both the experimental (dots) and  
241 estimated (dashed lines) values for the VS and NVS concentration during periods P4  
242 and P5.

243 The estimated VS solubilization constant dropped by 26 % with the temperature  
244 decrease, reducing its value from 0.0200 day<sup>-1</sup> at 33 °C (P4) to 0.0148 day<sup>-1</sup> at 20 °C  
245 (P5). The data were fitted to a Van't Hoff-Arrhenius equation, given by the Equation  
246 10:

$$k_{Sol,VS} = A \cdot e^{-\frac{E_a}{R \cdot T}} \quad \text{Equation 10}$$

247 Where A stands for the pre-exponential factor (day<sup>-1</sup>), E<sub>a</sub> for the activation energy  
248 (kJ·mol<sup>-1</sup>) and R being the universal gases constant (0.008314 kJ·mol<sup>-1</sup>·K<sup>-1</sup>). The  
249 experimental data fitting resulted in the values for the Van't Hoff-Arrhenius parameters  
250 shown in Equation 11:

$$k_{Sol,VS} = 17,78 \cdot e^{-\frac{17,28}{R \cdot T}} \quad \text{Equation 11}$$

251 Veeken and Hamelers (1999) determined the activation energy for the hydrolysis of  
252 4 selected bio-wastes (i.e. leaves, straw, orange peelings and grass). Hydrolysis  
253 constants were determined using the non-linear, least-squares method to fit cumulative  
254 methane production data to a first-order kinetics expression. Batch biomethane potential  
255 tests were carried out at 20, 30 and 40 °C, and it was assumed that hydrolysis was the  
256 limiting step of the anaerobic digestion and that intermediary products were being

257 consumed as soon as they were produced. These authors attributed the accurate fitting  
258 of the first-order kinetic model to a total occupation of the enzyme adsorption sites by  
259 hydrolytic enzymes that were whether already present in the bio-waste component  
260 surface or rapidly produced by fast-growing hydrolytic microorganisms. The hydrolysis  
261 rates followed an Arrhenius-type behavior, resulting in an average activation energy in  
262 the order of  $64 \text{ kJ}\cdot\text{kmol}^{-1}$ , which is a typical value for enzyme kinetics, confirming that  
263 the enzyme concentration must have been exceeding the concentration of degradable  
264 surface-sites. Conversely, the results obtained in the present work for the Van't Hoff-  
265 Arrhenius equation parameters derived from the VS solubilization constants at  $33 \text{ }^\circ\text{C}$   
266 and at  $20 \text{ }^\circ\text{C}$ , suggest that diffusion of hydrolytic enzymes from the bulk solution to the  
267 particle surface was rather the rate-limiting step of hydrolysis of MWW particulates,  
268 since the activation energy for diffusion is in the order of  $20 \text{ kJ}\cdot\text{mol}^{-1}$ . The observed  
269 difference was attributed to the combination of two factors. On the one hand, according  
270 to previous studies (Moñino et al., 2016), the average size of organic particulates in the  
271 MWW used in this study was more than a couple orders of magnitude lower ( $d_{50} =$   
272  $0,0398 \text{ mm}$ ) than that of the bio-wastes used by Veeken and Hamelers (1999) in their  
273 study ( $5\text{-}10 \text{ mm}$ ), leading to a much higher concentration of enzyme adsorption sites.  
274 On the other hand, in continuous operation, hydrolytic enzymes must reach their  
275 adsorption site, regardless of the mechanism used (Batstone et al., 2002b), before the  
276 enzymatic reaction can take place. The combination of these two factors resulted in a  
277 shift of the hydrolysis limiting step towards enzyme diffusion rather than enzymatic  
278 reaction.

279 According to Tchobanoglous et al. (2003), if a rate coefficient is known at one  
280 temperature, it may be calculated at another (within the range over which the

281 temperature dependent coefficient rises with increases in temperature) by rearranging  
282 the Arrhenius equation as shown in Equation 12:

$$k_{T_2} = k_{T_1} \cdot e^{\frac{E_a \cdot (T_2 - T_1)}{R \cdot T_1 \cdot T_2}} \quad \text{Equation 12}$$

283 Because the mesophilic temperature range is small when T is expressed in K, the  
284 term ( $R \cdot T_1 \cdot T_2$ ) in Equation 12 does not vary significantly and may be considered to be  
285 constant. Therefore, Equation 12 can be rewritten as Equation 13:

$$k_{T_2} = k_{T_1} \cdot \theta^{(T_2 - T_1)} \quad \text{Equation 13}$$

286 Where  $\theta$  is known as the temperature coefficient, and can be calculated by means of  
287 Equation 14:

$$\theta = e^{\frac{E_a}{R \cdot T_1 \cdot T_2}} \quad \text{Equation 14}$$

288 In the present study,  $\theta$  was found to be 1.0234, resulting in the temperature-  
289 correction expression given by Equation 15:

$$k_{T_2} = k_{T_1} \cdot 1.0234^{(T_2 - T_1)} \quad \text{Equation 15}$$

291

### 292 **3.1.3. Sludge Retention Time.**

293 The influence of the SRT over the VS solubilization process was assessed during  
294 periods P5, P9 and P10, during which the SRT was set to 70, 30 and 40 days,  
295 respectively, and the temperature remained constant in 20 °C. The HRT in P5 was  
296 higher than in periods P9 and P10, however the influence of this operating parameter  
297 was negligible, as previously reported.

298 Figure 2 (e) to (g) show the evolution of both the experimental (dots) and the  
299 estimated (dashed lines) values for the VS and NVS concentrations during periods P5,  
300 P9 and P10. Table 3 shows the estimated values for the VS solubilization constant



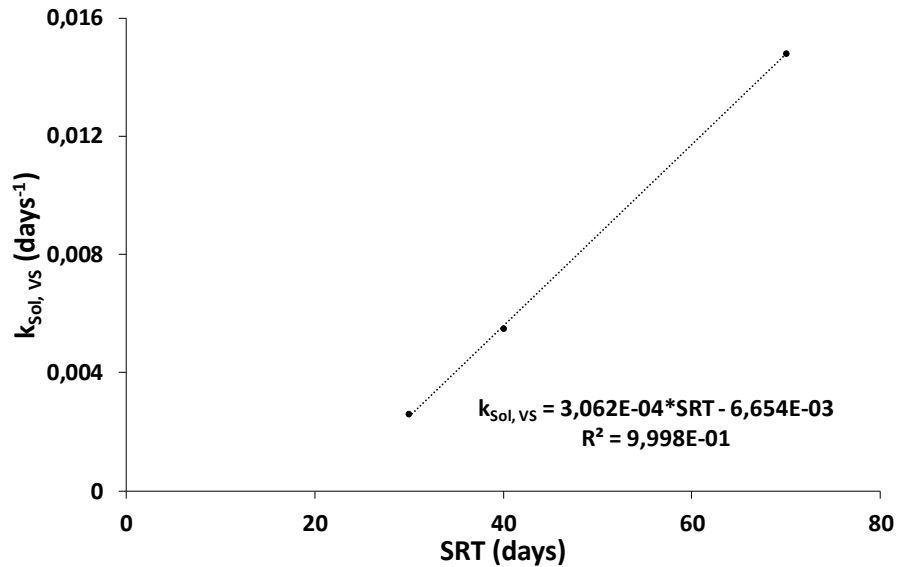
301 obtained from the non-linear fitting of the model to the analytical data during periods  
302 P5, P9 and P10.

<b>Period</b>	<b>SRT (d)</b>	<b><math>k_{Sol,VS}</math> (d<sup>-1</sup>)</b>
<b>P5</b>	70	0.0148
<b>P10</b>	40	0.0055
<b>P9</b>	30	0.0026

303 Table 3. Variation of the VS solubilization constant with SRT.

304 The values obtained for  $k_{Sol,VS}$  in periods P5, P9 and P10, during which the SRT was  
305 set to 70, 30 and 40 days, respectively, indicate that the hydrolysis rate declined with  
306 the reduction of the SRT. These results stated that the influence of the SRT over the  
307 hydrolysis process was not only related to the extent of the hydrolysis achieved as a  
308 function of the contact time between the organic particulate matter and the enzymes  
309 carrying out the hydrolysis process within the reactor. In addition, the hydrolysis rate  
310 was also affected by the SRT, which was attributed to a shift in the hydrolytic activity.  
311 Indeed, previous studies demonstrated that an increase in the SRT entailed an increase  
312 in the relative abundance of some hydrolytic microorganisms (Durán et al., 2018).  
313 These results are in agreement with those reported by Veeken and Hamelers (1999),  
314 which indicated that the hydrolysis rate depends, among others, on the concentration of  
315 hydrolytic biomass.

316 Figure 3 shows the correlation between the SRT and the VS solubilization constant.  
317 A linear regression analysis was performed (see Table S2; support material), and the  
318 analysis of variance stated that there exists a statistically-significant linear correlation  
319 between both parameters within the range of SRT studied (P-value = 0.0083).  
320 Furthermore, the Pearson's correlation coefficient ( $r = 0.9999$ ) indicated that the  
321 correlation was strong, with 99,98 % of the variability in  $K_{Sol,VS}$  being explained by the  
322 model, as evidenced by the coefficient of determination ( $R^2$ ).



323

324 Figure 3. Correlation between  $k_{Sol, VS}$  and SRT at 20°C.

325 As stated in section 3.1.2, VFA accumulation was observed at the beginning of  
 326 periods P5 (SRT = 70 days). It has been hypothesized that the VFA accumulation was  
 327 attributed to a temperature decrease rather than to a hydrolysis-rate variation. Likewise,  
 328 VFA accumulated at the beginning of period P9 (SRT = 30 days) in spite of the lower  
 329 hydrolysis rate, confirming this hypothesis. Conversely, no VFA accumulation was  
 330 observed during period P10 (SRT = 40 days), indicating that the balance between the  
 331 different microbial groups was not affected by the enhanced hydrolysis derived from the  
 332 SRT increase from period P9 to period P10. Alternatively, previous works (Giménez et  
 333 al., 2014, 2012) reported that COD mass balances in the different periods indicated that  
 334 the biogas and waste sludge productions were completely dependent on the hydrolysis  
 335 extent, as long as hydrolysis was the limiting step.

336

### 337 3.2. Model Validation

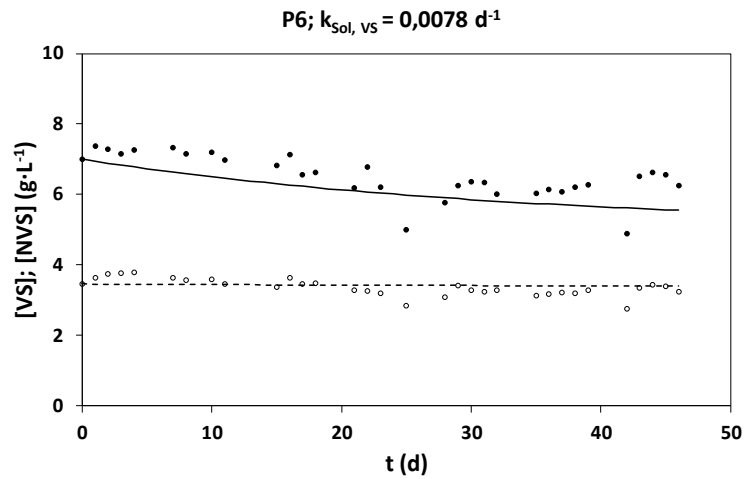
338 Periods P6, P7 and P8 were used to validate the model prediction. To this aim, the  
 339 volatile solids solubilization constants were calculated from the mathematical  
 340 expression found for the influence of the SRT (see Figure 3). Experimental data used to

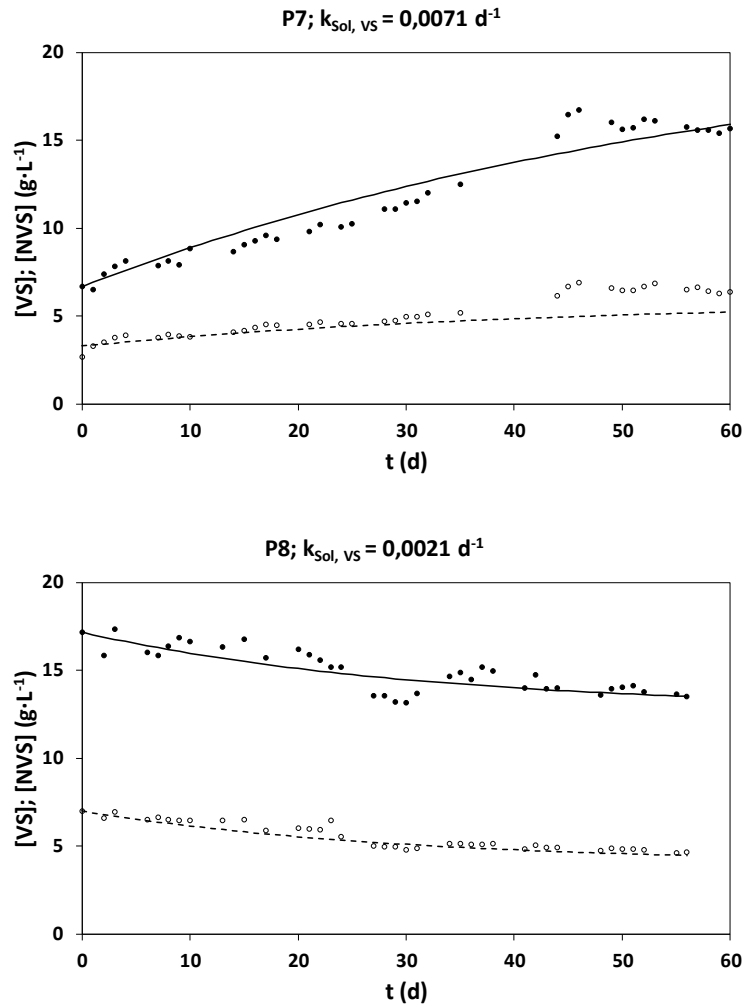
341 calibrate the influence of SRT were obtained during operation at 20 °C. Therefore, the  
 342 calculated  $k_{Sol, VS}$  were further corrected for temperature with equation 14. Table 4  
 343 shows the operating parameters set in each period and the estimations for the volatile  
 344 solids solubilization constant.

Period	T (°C)	HRT (d)	SRT (d)	$k_{Sol, VS}$ (d <sup>-1</sup> )
<b>P6</b>	29	1.44	42	0.0078
<b>P7</b>	23	0.64	43	0.0071
<b>P8</b>	17	0.63	29	0.0021

345 Table 4. Operating conditions and  $k_{Sol, VS}$  estimated for periods P6, P7 and P8.

346 The time-course evolution for both the volatile and non-volatile solids concentration  
 347 in the reactor was compared to that predicted by Equation 7 and Equation 8,  
 348 respectively, using the  $k_{Sol, VS}$  previously estimated.  
 349





350 Figure 4. Model validation

351 Figure 4 shows the time-course evolution for both the experimental and the predicted  
 352 volatile and non-volatile solids concentration values. This figure shows that the model  
 353 calibration and further validation on the basis of experimental results allowed to  
 354 accurately predict the volatile and non-volatile solids concentrations in an AnMBR  
 355 treating real MWW, depending on the HRT, the SRT and the temperature of operation.

356 The applicability of the model is limited to specific MWW in the range of SRT and  
 357 temperatures tested in this work. Nevertheless, it is worth of mention that the inclusion  
 358 of additional parameters (e.g. biodegradable fraction of the volatile solids,  $f_{BD}$ ) would  
 359 enable to extend the applicability of the model to a wider range of influent  
 360 characteristics. Furthermore, in spite of the wide knowledge regarding the effect of

361 temperature on the hydrolysis rate, additional operation data at different temperatures  
362 could assist in the calculation of a more accurate Van't-Hoff- Arrhenius-equation  
363 activation energy, which would be useful to determine the limiting stage of the  
364 hydrolysis process in AnMBR treating MWW.

#### 365 **4. CONCLUSIONS.**

366 This work presents a mathematical approach to predict the solids concentration  
367 within an AnMBR, depending on the influent wastewater characteristics and the  
368 operating conditions.

369 The influence of the HRT on the VS solubilization was negligible, as evidenced by  
370 the close values for the VS solubilization constant, despite the three-fold reduction of  
371 the HRT. Therefore, lower  $k_{Sol,VS}$  during summer periods were attributed to the different  
372 influent wastewater features rather than to HRT.

373 The hydrolysis extent was higher with longer SRT, as a result of the extended contact  
374 time between the organic particulate matter and the enzymes carrying out the hydrolysis  
375 process. Furthermore, the shift in the relative abundance of hydrolytic microorganisms  
376 with SRT also influenced the hydrolysis rate.

377  $k_{Sol,VS}$  decreased by 26 % with temperature decrease from 33 °C to 20 °C. Data fitting  
378 to an Arrhenius equation resulted in an activation energy of 17,28 kJ·mol<sup>-1</sup> suggesting  
379 that diffusion of hydrolytic enzymes from the bulk solution to the adsorption site is the  
380 rate-limiting step of hydrolysis.

381 The model calibration and further validation on the basis of experimental results  
382 allowed to accurately predict the volatile and non-volatile solids concentrations in an  
383 AnMBR treating real MWW, depending on the operating HRT, SRT and temperature.

## 384 **ACKNOWLEDGEMENTS.**

385 This research work was supported by the Spanish Research Foundation (CICYT)  
386 under Grants CTM2008-06809-C02-01 and CTM2008-06809-C02-02); Comunitat  
387 Valenciana Regional Government under Grant GVACOMP2009-285.

## 388 **REFERENCES.**

- 389 APHA, AWWA, WEF, 2012. Standard Methods for the Examination of Water &  
390 Wastewater.
- 391 Batstone, D.J., Keller, J., Angelidaki, I., Kalyuzhnyi, S.V., Pavlostathis, S.G., Rozzi, A.,  
392 Sanders, W.T.M., Siegrist, H., Vavilin, V.A., 2002a. The IWA Anaerobic  
393 Digestion Model No 1 (ADM1), Water Science and Technology. IWA  
394 Publishing.
- 395 Batstone, D.J., Keller, J., Angelidaki, I., Kalyuzhnyi, S.V., Pavlostathis, S.G., Rozzi, A.,  
396 Sanders, W.T.M., Siegrist, H., Vavilin, V.A., 2002b. Anaerobic Digestion Model  
397 No. 1.
- 398 Batstone, D.J., Puyol, D., Flores-Alsina, X., Rodríguez, J., 2015. Mathematical  
399 modelling of anaerobic digestion processes: applications and future needs. Rev.  
400 Environ. Sci. Bio/Technology 14, 595–613. [https://doi.org/10.1007/s11157-015-](https://doi.org/10.1007/s11157-015-9376-4)  
401 9376-4
- 402 Chen, C., 2020. Anaerobic membrane bioreactors for sustainable and energy-efficient  
403 municipal wastewater treatment, in: Current Developments in Biotechnology and  
404 Bioengineering. pp. 335–366. [https://doi.org/10.1016/B978-0-12-819852-0.00014-](https://doi.org/10.1016/B978-0-12-819852-0.00014-2)  
405 2
- 406 Daigger, G.T., 2008. New approaches and technologies for wastewater management, in:  
407 The Bridge. Vol. 38-2: Technologies for Clean Water. pp. 38–45.

408 Durán, F., Zamorano-López, N., Barat, R., Ferrer, J., Aguado, D., 2018. Understanding  
409 the performance of an AnMBR treating urban wastewater and food waste via  
410 model simulation and characterization of the microbial population dynamics.  
411 *Process Biochem.* 67, 139–146. <https://doi.org/10.1016/j.procbio.2018.02.010>

412 Gavala, H.N., Yenal, U., Skiadas, I. V., Westermann, P., Ahring, B.K., 2003.  
413 Mesophilic and thermophilic anaerobic digestion of primary and secondary sludge.  
414 Effect of pre-treatment at elevated temperature. *Water Res.* 37, 4561–4572.  
415 [https://doi.org/10.1016/S0043-1354\(03\)00401-9](https://doi.org/10.1016/S0043-1354(03)00401-9)

416 Giménez, J.B., 2014. Estudio del tratamiento anaerobio de aguas residuales urbanas en  
417 biorreactores de membranas. University of Valencia.

418 Giménez, J.B., Martí, N., Ferrer, J., Seco, A., 2012. Methane recovery efficiency in a  
419 submerged anaerobic membrane bioreactor (SAnMBR) treating sulphate-rich  
420 urban wastewater: Evaluation of methane losses with the effluent. *Bioresour.*  
421 *Technol.* 118, 67–72. <https://doi.org/10.1016/j.biortech.2012.05.019>

422 Giménez, J.B., Martí, N., Robles, A., Ferrer, J., Seco, A., 2014. Anaerobic treatment of  
423 urban wastewater in membrane bioreactors: Evaluation of seasonal temperature  
424 variations. *Water Sci. Technol.* 69, 1581–1588.  
425 <https://doi.org/10.2166/wst.2014.069>

426 Giménez, J.B., Robles, A., Carretero, L., Durán, F., Ruano, M. V, Gatti, M.N., Ribes, J.,  
427 Ferrer, J., Seco, A., 2011. Experimental study of the anaerobic urban wastewater  
428 treatment in a submerged hollow-fibre membrane bioreactor at pilot scale.  
429 *Bioresour. Technol.* 102, 8799–806. <https://doi.org/10.1016/j.biortech.2011.07.014>

430 Kleerebezem, R., van Loosdrecht, M.C.M., 2007. Mixed culture biotechnology for  
431 bioenergy production. *Curr. Opin. Biotechnol.* 18, 207–12.  
432 <https://doi.org/10.1016/j.copbio.2007.05.001>

433 Larsen, T.A., Alder, A.C., Eggen, R.I.L., Maurer, M., Lienert, J., 2009. Source  
434 Separation: Will We See a Paradigm Shift in Wastewater Handling? *Environ. Sci.*  
435 *Technol.* 43, 6121–6125. <https://doi.org/10.1021/es803001r>

436 Lettinga, G., Rebac, S., Zeeman, G., 2001. Challenge of psychrophilic anaerobic  
437 wastewater treatment. *Trends Biotechnol.* 19, 363–370.  
438 [https://doi.org/10.1016/S0167-7799\(01\)01701-2](https://doi.org/10.1016/S0167-7799(01)01701-2)

439 Li, W.-W., Yu, H.-Q., 2011. From wastewater to bioenergy and biochemicals via two-  
440 stage bioconversion processes: a future paradigm. *Biotechnol. Adv.* 29, 972–82.  
441 <https://doi.org/10.1016/j.biotechadv.2011.08.012>

442 Ludwig, T., Gaida, D., Keysers, C., Pinnekamp, J., Bongards, M., Kern, P., Wolf, C.,  
443 Sousa Brito, A.L., 2012. An advanced simulation model for membrane bioreactors:  
444 Development, calibration and validation. *Water Sci. Technol.* 66, 1384–1391.  
445 <https://doi.org/10.2166/wst.2012.249>

446 Miron, Y., van Lier, J.B., Lettinga, G., 2000. Miron 2000. *Water Res.* 34, 1705–1713.

447 Moñino, P., Jiménez, E., Barat, R., Aguado, D., Seco, A., Ferrer, J., 2016. Potential use  
448 of the organic fraction of municipal solid waste in anaerobic co-digestion with  
449 wastewater in submerged anaerobic membrane technology. *Waste Manag.* 56,  
450 158–165. <https://doi.org/10.1016/j.wasman.2016.07.021>

451 Nabi, M., Zhang, G., Li, F., Zhang, P., Wu, Y., Tao, X., Bao, S., Wang, S., Chen, N.,  
452 Ye, J., Dai, J., 2020. Enhancement of high pressure homogenization pretreatment  
453 on biogas production from sewage sludge: a review. *Desalin. Water Treat.* 175,  
454 341–351. <https://doi.org/10.5004/dwt.2020.24670>

455 Nabi, M., Zhang, G., Zhang, P., Tao, X., Wang, S., Ye, J., Zhang, Q., Zubair, M., Bao,  
456 S., Wu, Y., 2019. Contribution of solid and liquid fractions of sewage sludge  
457 pretreated by high pressure homogenization to biogas production. *Bioresour.*



458 Technol. 286, 121378. <https://doi.org/10.1016/j.biortech.2019.121378>

459 Pavlostathis, S.G., Giraldo-Gomez, E., 1991. Kinetics of anaerobic treatment: A critical  
460 review. *Crit. Rev. Environ. Control* 21, 411–490.  
461 <https://doi.org/10.1080/10643389109388424>

462 Robles, Á., Ruano, M.V., Charfi, A., Lesage, G., Heran, M., Harmand, J., Seco, A.,  
463 Steyer, J., Batstone, D.J., Kim, J., Ferrer, J., 2018. A review on anaerobic  
464 membrane bioreactors ( AnMBRs ) focused on modelling and control aspects.  
465 *Bioresour. Technol.* 270, 612–626. <https://doi.org/10.1016/j.biortech.2018.09.049>

466 Robles, A., Ruano, M. V., Ribes, J., Seco, A., Ferrer, J., 2013. A filtration model  
467 applied to submerged anaerobic MBRs (SAnMBRs). *J. Memb. Sci.* 444, 139–147.  
468 <https://doi.org/10.1016/j.memsci.2013.05.021>

469 Sarioglu, M., Insel, G., Orhon, D., 2012. Dynamic in-series resistance modeling and  
470 analysis of a submerged membrane bioreactor using a novel filtration mode.  
471 *Desalination* 285, 285–294. <https://doi.org/10.1016/j.desal.2011.10.015>

472 Smith, A.L., Stadler, L.B., Love, N.G., Skerlos, S.J., Raskin, L., 2012. Perspectives on  
473 anaerobic membrane bioreactor treatment of domestic wastewater: a critical  
474 review. *Bioresour. Technol.* 122, 149–59.  
475 <https://doi.org/10.1016/j.biortech.2012.04.055>

476 Tchobanoglous, G., Burton, F.L., Stensel, H.D., Metcalf & Eddy, I., Burton, F., 2003.  
477 *Wastewater Engineering: Treatment and Reuse*, McGraw-Hill higher education.  
478 McGraw-Hill Education.

479 Veeken, A., Hamelers, B., 1999. Effect of temperature on hydrolysis rates of selected  
480 biowaste components. *Bioresour. Technol.* 69, 249–254.  
481