

Document downloaded from:

<http://hdl.handle.net/10251/166757>

This paper must be cited as:

Souana, K.; Taïbi, K.; Abderrahim, LA.; Amirat, M.; Achir, M.; Boussaid, M.; Mulet, JM. (2020). Salt-tolerance in *Vicia faba* L. is mitigated by the capacity of salicylic acid to improve photosynthesis and antioxidant response. *Scientia Horticulturae*. 273:1-7. <https://doi.org/10.1016/j.scienta.2020.109641>



The final publication is available at

<https://doi.org/10.1016/j.scienta.2020.109641>

Copyright Elsevier

Additional Information

1 **Salt-tolerance in *Vicia faba* is enhanced by the effect of salicylic acid on**
2 **photosynthesis and the antioxidant response**

3
4 **Abstract**

5 Selection and improvement of crops subjected to salinity constitutes an urgent need for
6 increasing agricultural and food production in order to feed the growing human population. The
7 aim of the study is to evaluate the role of salicylic acid application at various concentrations in
8 mitigating the adverse effects of salinity of two faba bean genotypes at the physiological and
9 molecular levels. For this purpose we have selected two genotypes of faba bean (*Vicia faba* L.)
10 and we have applied to them various concentrations of NaCl and salicylic acid in a full factorial
11 design. Growth, water status, gas exchanges, photosynthesis parameters, ions homeostasis and
12 antioxidant enzymes activities were evaluated. One of the genotypes (Aguadulce) performed
13 better under salt stress, but the difference among them was rather quantitative than qualitative. In
14 general, salinity limited growth and several physiological processes. However, faba bean plants
15 have triggered some mechanisms to deal with this constraint throughout the maintenance of
16 water status and the enhancement of antioxidant enzymatic activities. The salt-tolerance of both
17 genotypes at the growth and physiological level was improved by the application of salicylic
18 acid. The beneficial effect of salicylic acid varies according to its concentration, the genotype
19 and the studied parameter. Overall, the genotype Aguadulce performs better with 0.5 mM SA
20 while the genotype Hystal requires 1 mM SA. The genotype Aguadulce displayed better growth,
21 physiological and molecular response under salt stress than the genotype Hystal. Besides,
22 salicylic acid can be considered as potential growth regulator to improve the salt response of
23 both faba bean genotypes. The application of salicylic acid would provide a practical basis for
24 wide cultivation of faba bean in marginal and wastelands under-cultivated and might propose an
25 effective ecological and economical alternative solution to deal with salt-affected soils mainly in
26 arid regions.

1 **Key words**

2 Salt tolerance, salicylic acid, *Vicia faba* L., water status, gas exchanges, photosynthesis, ions
3 homeostasis, antioxidants enzymes.

4

5

6

7 **1. Introduction**

8 Salinity is one of the major threats to food production and agricultural yield in the XXI century
9 and its adverse effects are expected to increase due to anthropogenic global warming and
10 demographic increase. Arid and semiarid regions are more vulnerable to this threat. Recent
11 statistics have revealed that more than 45 million hectares of irrigated land are affected by salt
12 stress and this number is increasing (Isayenkov and Maathuis, 2019).

13

14 Plants subjected to salinity experience a combination of osmotic and ionic stresses that leads
15 regrettably to various damages at the physiological, cellular and molecular levels (Serrano et al.,
16 1999), further causing reduced uptake of nutrients and decline of photosynthetic performance
17 (Taïbi et al., 2016). In addition, salt stress induces the production of reactive oxygen species
18 which damage the biomolecules (lipids, proteins, and nucleic acids) and this causes a decrease of
19 essential enzymatic activities and even causes the degradation of cell membrane systems
20 (Mahajan and Tuteja 2005)

21

22 Salicylic acid is a phenolic phytohormone with roles in plant growth and development,
23 photosynthesis, transpiration, ion uptake and transport. Salicylic acid is involved in endogenous
24 signalling, mediating in plant response against pathogens. an a determinant of plant growth
25 regulation, thereby generating a wide range of metabolic and physiological responses in plants
26 (Khan et al., 2015). Salicylic acid is able to mitigate the effects of stress as induces nutrient

1 uptake, membrane protection, photosynthesis repair, and also can reduce oxidative stress (Khan
2 et al., 2015). There are several reports available highlighting the role of salicylic acid in
3 improving salt stress tolerance in plants (Ahmad et al., 2018; Jayakannan et al., 2013; Khan et
4 al., 2014; Li et al., 2014a; Li et al., 2014b; Nazar et al., 2015).

5
6 Fabaceous plants constitute a suitable solution to feed growing populations, improve human and
7 animal nutrition and enhance soil fertility since they are able to enrich the soil's nitrogen content
8 throughout its symbiotic association with rhizobium (Castro-Guerrero et al., 2016). The faba
9 bean or broad bean, *Vicia faba* L., is an important legume for human nutrition as it is a major
10 source of protein, fiber, vitamins and minerals. This species is very sensitive to salinity and
11 many strategies are being developed to enhance its production and yields in marginal arid lands.

12
13 In the present study, we hypothesised that (i) the extent of the beneficial effects of salicylic acid
14 application varies among genotypes belonging to the same species and that (ii) evaluate the dose-
15 response of the hormone and the physiological and biochemical outcomes under salt stress. To
16 attain these objectives we have evaluated the effect of different concentrations of salicylic acid
17 on two faba bean genotypes at the physiological and molecular levels. Examining the magnitude
18 of intra-specific and genotypic variation for salinity tolerance, and a detailed description of the
19 physiological and molecular response, as well as the effect of salicylic acid could provide vital
20 information to faba bean breeding programs for the selection of the most suited genotypes for
21 introduction in salt affected areas.

22

23 **2. Material and methods**

24 *Plant material and growth conditions*

25 Two genotypes of *Vicia faba* L., Aguadulce and Hystal, widely cultivated in developing countries
26 of the Mediterranean region were surface sterilized using sodium hypochlorite (0.5 %, v/v)

1 during 5 minutes followed by several washes with distilled water. Seeds preparation and
2 conditions of cultivation were performed as described in Taïbi et al. (2016). Four-week-old
3 plants were arranged in factorial block design with eight replicates per treatment, two faba bean
4 genotypes, three salt treatments (i.e. 0, 100 and 200 mM of NaCl) and three salicylic acid
5 ($C_7H_6O_3$, Sigma-Aldrich, C 99.0%) treatments (i.e. 0, 0.5 and 1 mM). Salicylic acid (SA; 2-
6 hydroxybenzoic acid) was dissolved in 0.02 % Tween 20 (Polyoxyethylenesorbitan monolaurate,
7 Sigma Chemicals, UK). Either NaCl or salicylic acid mixed with Tween-20 were both dissolved
8 in the nutrient solution of Hoagland which was applied directly to the pots for irrigation every
9 alternate day. The same volume of Tween-20 was added to the nutrient solution untreated with
10 salicylic acid for homogeneity. Plants were harvested after ten days of treatment. The dry
11 weights of the aerial parts and roots were determined and samples were conserved at $-80\text{ }^{\circ}\text{C}$ for
12 further analyses.

13

14 *Water status measurements*

15 Leaf water potential (Ψ_w , MPa) was measured in eight plants selected randomly per treatment
16 using a Scholander-type pressure pump (PMS 1000, USA). Nevertheless, relative water content
17 (RWC) was assessed on leaves following the formula:

$$18 \text{ RWC (\%)} = [(\text{fresh weight} - \text{dry weight}) / (\text{turgid weight} - \text{dry weight})] \times 100$$

19

20 *Gas exchange and photosynthesis measurements*

21 Instantaneous assessments of stomatal conductance (g_s , $\text{mol m}^{-2} \text{ s}^{-1}$), transpiration (E , $\text{mmol H}_2\text{O}$
22 $\text{m}^{-2} \text{ s}^{-1}$), net CO_2 assimilation (P_n , $\mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$), and instantaneous water use efficiency
23 (WUE_{inst} ; $\mu\text{mol CO}_2 \text{ mmol}^{-1} \text{ H}_2\text{O}$) were also measured on recently expanded leaves of eight
24 plants per treatment by means of a portable photosynthesis (Model LI-6400, LI-COR
25 Biosciences Inc., Lincoln, USA). Besides, maximal photochemical efficiency of PSII (Fv/Fm)
26 was assessed at predawn using a handy PEA portable fluorometer (Hansatech, United Kingdom).

1

2 *Determination of electrolyte leakage*

3 Electrolyte leakage in fresh leaves was determined following Li et al. (2014b) method:

4
$$\text{Electrolyte leakage (\%)} = \frac{(\text{EC}_1 - \text{EC}_0)}{(\text{EC}_2 - \text{EC}_0)} \times 100,$$

5 where EC_0 is the electrical conductivity recorded at 0 time point; EC_1 is the electrical
6 conductivity measured after contents heated at 60 °C; EC_2 is the electrical conductivity after
7 contents heated at 100 °C.

8

9 *Determination of ion content*

10 Concentrations of sodium (Na^+) and potassium (K^+) were determined in oven dried leaf samples,
11 after wet-digesting the samples in $\text{HNO}_3\text{-HClO}_4$ acid mixture (4:1 v/v), by atomic absorption
12 spectrometry (Perkins Elmer, Norwalk, CT, USA) as described by Taïbi et al. (2016).

13

14 *Antioxidant enzymes assay*

15 Antioxidant enzymes were extracted at 4 °C from 500 mg fresh leaf sample in Tris-HCl (100
16 mM, pH 7.5) containing dithiotrol (5 mM), MgCl_2 (10 mM), EDTA (1.0 mM), magnesium
17 acetate (5 mM), PVP (1.5%), PMSF (1.0 mM) and 1 $\mu\text{g/mL}$ aproptinin in a pre-chilled pestle and
18 mortar. The homogenate was centrifuged at 10.000 rpm for 15 minutes at 4°C and the
19 supernatant was used as enzyme source then Superoxide dismutase (SOD), Catalase (CAT),
20 Ascorbate peroxidase (APX) and Glutathione reductase (GR) activity were assayed according to
21 the method described by Cakmak et al. (1993).

22

23 *Statistical analysis*

24 Prior to the analysis, the Levene test was applied to check the analysis of variance requirements.
25 The significance of differences between different treatments was assessed using analysis of
26 variance, at 95% confidence level. The post-hoc Tukey test was used to estimate homogeneous

1 groups. Correlations between the measured parameters were determined based on the coefficient
2 of Pearson.

3

4

5 **3. Results**

6 The effect of salt stress on faba bean seedling growth was evaluated throughout the
7 determination of shoot and root dry weight (Figures 1A and B). In general, dry biomass
8 decreased significantly in comparison to control seedlings for both genotypes in response to the
9 increase of NaCl concentration (p-value<0.01**). Shoot dry biomass decreased by 60 % in
10 Aguadulce and 65 % in Hista while root dry biomass decreased by around 70 % in both
11 genotypes under high salinity. However, the exogenous application of salicylic acid enhances
12 both shoot and root biomass over control and salt stressed seedlings; under high salinity, less
13 decrease in shoot dry biomass was observed in the treatment with 0.5 mM SA (50 % for
14 Aguadulce and 60 % for Hista). However, better effect concerning root dry biomass was noted
15 in the treatment with 1 mM SA for both genotypes.

16

17 **Figure 1.**

18

19 As expected, salinity affects seedlings water status; leaf relative water content (RWC)
20 diminished by more than 32 % for both genotypes and water potential (Ψ_w) decreased
21 significantly by around 153 % in Aguadulce and 135 % in Hista. This finding indicates that
22 seedlings were experiencing drought stress concomitant to salinity. However, salicylic acid
23 application improves significantly seedlings water status through the decrease of water potential
24 to 165 % for Aguadulce and 150 % for Hista and therefore, the recovery of relative water
25 content to 72 % and 74 % respectively in Aguadulce and Hista. Nevertheless, under high

1 salinity, better improving effect of water status was observed through treatment with 0.5 mM SA
2 (Fig. 2A, B).

3

4 **Figure 2.**

5

6 Likewise, stomatal conductance (g_s), transpiration (E), net photosynthesis (Pn) and intercellular
7 CO₂ concentration (C_i) decreased significantly when seedlings were subjected to salt stress
8 (Figures 3 A, B, C, D). The concomitant diminution under high salinity was around 48 % in gas
9 exchange, 52 % in transpiration and intercellular CO₂ concentration, and 34 % in net
10 photosynthesis for both genotypes. Exogenous application of salicylic acid improved
11 significantly these parameters; less decreases were estimated around 34 % and 32 % in gas
12 exchange, 48 % and 53 % for transpiration, 30 % and 31 % in net photosynthesis and 33 % and
13 35 % in intercellular concentration of CO₂ respectively in Aguadulce and Histal subjected to
14 high salinity. Additionally, maximal efficiency of PSII (F_v/F_m) decreased as well under salinity
15 for both genotypes by around 7 % (p-value<0.01**). Nevertheless, exogeneous salicylic acid
16 enhanced the maximal efficiency of PSII under controlled and salt stressed conditions by around
17 2 to 3 %; the highest value obtained under 200 mM NaCl (F_v/F_m=0.77) was observed in the
18 treatment with 0.5 mM SA for the genotype Aguadulce and in the treatment with 1 mM SA for
19 the genotype Histal (Figure 3 E).

20 Subsequent increase in water use efficiency (WUE_{inst}) was induced by salinity and salicylic acid
21 application improves more the obtained values in both the two genotypes (Figure 3 F).

22 Nevertheless, water use efficiency was higher in the genotype Aguadulce under controlled and
23 salt-stressed conditions. This improvement translates a higher capacity to maintain elevated
24 photosynthetic levels in the genotype Aguadulce for comparable transpiration values in both
25 genotypes.

26

1 **Figure 3.**

2

3 Being the last sink, leaves constitute the plant part where salt ions accumulate. Salt stress induced
4 a significant increase of sodium content by around 156 % against a significant decrease of
5 potassium content by around 50 % for both genotypes (p-value<0.001***). However, exogenous
6 salicylic acid supply decreased remarkably sodium content in leaves of both genotypes by around
7 10 % for the concentration 0.5 mM SA and 15 % for the concentration 1 mM SA under high
8 salinity (Fig. 4 A). Concomitantly, potassium content increased for both genotypes by around 30
9 % under 0.5 mM SA and 50 % under 1 mM SA when plants were irrigated with 200 mM NaCl
10 (Fig. 4 B). Consequently, the ratio K^+/Na^+ was improved significantly with salicylic acid treatment
11 (Fig. 4 C).

12

13 In addition, salinity increased the electrolyte leakage by more than four-folds in both genotypes,
14 indicating membrane damage (p-value<0.001***). Treatment of seedlings with salicylic acid
15 diminished widely the electrolyte leakage either in controlled or salt-stressed seedlings. This effect
16 was estimated around 17 % of decrease in Aguadulce and 22 % in Hystal under high salinity in
17 comparison to salt-stressed seedlings untreated with SA (Fig. 4 D).

18

19 **Figure 4.**

20

21 Regarding antioxidant enzymes activity, a significant increase was observed for both genotypes
22 subjected to salt stress in terms of superoxide dismutase (SOD), ascorbate peroxidase (APX),
23 catalase (CAT) and glutathione reductase (GR) activities (p-value<0.001***). The rates of
24 increase were estimated around to 260 % for SOD and APX activity however, CAT activity
25 increased by 210 % in Aguadulce and 224 % in Hystal while GR activity increased by 198 % in
26 Aguadulce and 192 % in Hystal. The application of salicylic acid induced further increase of the

1 enzymatic activity mainly under high salinity. SOD activity increase was estimated around 300 %
2 for Aguadulce and 320 % for Hystal. Nevertheless, APX activity increase was about 280 % for
3 Aguadulce and 270 % for Hystal. By the same, CAT activity increased to 260 % for Aguadulce
4 and 270 % for Hystal while GR activity raised to 240 % for both genotypes. Better effects were
5 observed in the treatment supplied with 0.5 mM SA for APX, CAT and GR. However, SOD
6 activity was higher in the treatment with 1 mM SA (Fig. 5 A, B, C, D).

7

8 **Figure 5.**

9

10 **4. Discussion**

11 Salinity is among the major threats to horticulture by inducing several limitations in plant
12 growth, development and overall productivity. Selection and improvement of salt-tolerant
13 genotypes represents an effective strategy to improve yield in areas suffering from adverse
14 environmental conditions. Therefore, comparison among several genotypes for salt tolerance can
15 be very useful to describe the relevant mechanisms that might help to recognize stress tolerant
16 genotypes and unveiling the limiting factors which can constitute objectives for breeding or
17 molecular breeding (Taïbi et al., 2018; Taïbi et al., 2017; Taïbi et al., 2016). Besides,
18 phytohormones constitute a strong tool to alleviate the adverse effects produced by abiotic stress.
19 Recently, an increasing interest has been focused on the effect of salicylic acid in salinity
20 tolerance.

21

22 In general, salinity induced severe restrictions of physiological and molecular responses of faba
23 bean which affected significantly plant growth. However, salicylic acid restored most of these
24 parameters and improve the plant stress response.

25

1 Severe growth limitations in terms of shoot and root dry biomass were observed under salinity
2 for both genotypes. This finding is one of the most visible effects of salt stress and it is generally
3 attributed to the inhibition of the developmental program and the stimulation of stress response
4 mechanisms (Isayenkov and Maathuis, 2019). Growth arrest under salinity has been already
5 reported in faba bean (Bulut et al., 2011; Dawood and El-Awadi, 2015; Moussa and Hassan,
6 2016) and other leguminous species like common bean (Taïbi et al., 2016; Taïbi et al., 2012;
7 Taïbi et al., 2013a, b). Therefore, biomass accumulation measure may help in the evaluation of
8 the relative degree of salt tolerance among faba bean genotypes. However, the application of
9 salicylic acid enhanced both shoot and root dry biomass in salt-stressed plants. Here we show
10 that the ameliorating effects of salicylic acid could be attributed to its role in membrane
11 protection from salt damage (r-value=0.73**), and enhancement of chlorophyll content and
12 antioxidant enzymes activity (r-value=0.82**) which in turn supports the photosynthesis process,
13 alleviate oxidative stress and regulate ion homeostasis (Khan et al., 2015; Li et al., 2014a; Li et
14 al., 2014b). On the other side, growth limitations could be due to the significant deterioration of
15 relative water content (RWC, r-value=-0.68**) and water potential (Ψ_w , r-value=-0.71**) under
16 salinity. It is well admitted that plants including faba bean respond to salt stress by regulating
17 their osmotic potential and ionic balance throughout the accumulation of organic and inorganic
18 compounds in order to sustain cell turgor and subsequently to maintain growth (Abdul Qados,
19 2011; Nazar et al., 2015). Maintenance of water status through adjustment of water potential is
20 considered as an important adaptation strategy under salinity (Ashraf and Harris, 2013).
21 Application of salicylic acid enhanced leaves diffusive resistance (r-value=0.58**) and control
22 transpiration level (r-value=-0.54**) which might improve water status of the plants (Ahmad et
23 al., 2018).
24 Likewise, gas exchanges and net photosynthesis decreased in plants subjected to salinity (r-
25 value=-0.44*). The stomatal closure, as an earliest response of salt-stressed plants, was
26 significantly marked in the present study leading to the decrease of stomatal conductance and

1 transpiration. In general, higher stomatal conductance enhances CO₂ diffusion into the leaves
2 and improves photosynthetic rate which is an important determinant of growth and yield (Li et
3 al., 2014b). Therefore, the decrease in photosynthesis rates under saline conditions is attributed
4 mostly to the limitation of stomatal conductance (r-value=0.61**). Besides, diminution in
5 photosynthesis might be attributed to perturbations in pigments content and chlorophyll
6 fluorescence, PSII photoinhibition, conformational modifications in membrane-bound ATPase
7 enzyme complex, and the diminution in Rubisco activity (Lawlor and Cornic, 2002).
8 Nevertheless, salicylic acid was found to enhance photosynthetic rates and gas exchanges.
9 Previous studies of Khan et al. (2014) and Ahmad et al. (2018) have reported that salicylic acid
10 increases pigments content, improves the rates of transpiration and photosynthetic electron
11 transport, sustains higher Rubisco and antioxidant enzymatic activities, and boosts the PSII
12 efficiency under salt stress. The maximal quantum yield of PSII phytochemistry (Fv/ Fm)
13 decreased under saline conditions but restored after the addition of salicylic acid. This finding
14 indicates reduced leaves capacity for absorption of excitation energy and diminution in
15 photosynthetic assimilation under salinity (Taïbi et al., 2018; Taïbi et al., 2017). The protective
16 effects of salicylic acid could be associated to the diminution of oxidative damage and the
17 restriction of massive sodium accumulation in leaves (Khan et al., 2015).
18 By the same, regulation of ion homeostasis is one of the main determinants of plants salt
19 tolerance (Isayenkov and Maathuis, 2019). In the present study, salinity increased the content of
20 sodium and decreased the content of potassium. These findings are in agreement with (Ahmad et
21 al., 2018; Bulut et al., 2011; Taïbi et al., 2016). Competition exists for the uptake of sodium and
22 potassium since they share similar physicochemical structure. Under salt conditions, the passive
23 entrance of sodium ions under salinity throughout the plasma membrane could induce membrane
24 depolarization that prevents considerably potassium uptake and increase its leakage through
25 depolarization-activated potassium outward-rectifying channels (Jayakannan et al., 2013). The
26 suppression of potassium uptake induces various cell damages and reduces plant growth since it

1 constitutes an important activator of several cytosolic enzymes and contributes as well for
2 osmotic adjustment (Almeida et al., 2017). Besides, potassium plays a determinant role in
3 regulating stomatal aperture, and this is subjected to a complex regulation so the observed
4 differences may be the underlying cause for the differences observed in water use efficiency and
5 photosynthesis rates. Exogenous salicylic acid supply was found to be very effective in dropping
6 sodium excess and enhancing potassium content in salt-stressed faba bean plants. Salicylic acid
7 reduces the xylemic translocation of sodium to the aerial parts from a part, and the extent of
8 membrane polarization through the stimulation of H⁺-ATPase activity under salt conditions from
9 another part which could, consequently, restore potassium retention and decrease its efflux
10 (Khan et al., 2015). Nevertheless, the higher accumulations of sodium and potassium were
11 characteristics of the genotype Aguadulce. This suggest that it is the ability to uptake and retain
12 potassium that determines faba bean genotypes growth and yield under salt stress.

13 In addition, an increase of electrolyte leakage was associated to salt stress reflecting
14 consequently the occurrence of membrane injuries. Membrane disorganization is due mainly to
15 the decrease of phospholipids and sterols in response to salinity (Wu et al., 1998). Reactive
16 oxygen species attacks linolenic and linoleic polyunsaturated fatty acids and produce complex of
17 lipid hydroxy-peroxides which reduce membrane permeability (Ahmad et al., 2018). It should be
18 noted that membrane protection from salt injuries constitutes a determinant criterion for salinity
19 tolerance mechanisms and salicylic acid demonstrated its benefits to decrease electrolyte
20 leakage. Salicylic acid effects are attributed to the stimulation of the antioxidative system and
21 improvement of calcium uptake (El-Tayeb, 2005).

22 Salt stress affects various physiological and molecular mechanisms associated with plant growth
23 and development. Plants with potent antioxidant system, either constitutive or induced, acquire
24 significant salt-tolerance (Isayenkov and Maathuis, 2019). The activity of the antioxidant enzymes
25 SOD, APX, CAT and GR increased in response to salt stress. The genotype Aguadulce revealed
26 higher SOD and CAT activities while the genotype Histal showed higher APX and GR activities.

1 The role of antioxidant enzymes is to scavenge reactive oxygen species produced by oxidative
2 stress and therefore protect cell membrane integrity (Taïbi et al., 2016). SOD converts superoxide
3 radicals into which will be reduced by CAT to H₂O and oxygen (Demiral and Türkan, 2005).
4 Besides, the increase in GR activity under salt conditions has been also reported (Maribel and
5 Tobita, 1998; Taïbi et al., 2016). The antioxidant enzymatic activities increased in controlled and
6 salt-stressed conditions after salicylic acid treatments. Exogenous application of salicylic acid
7 mitigated salt stress in *S. lycopersicum* throughout the stimulation of GST gene family (Csiszár et
8 al., 2014). The boosted expression of antioxidants genes such as GST1-2, GPX1-2 and ASA-GSH
9 pathway in response to salicylic acid supply restores cell membrane integrity and enhances the
10 content of photosynthetic pigments (Li et al., 2014b). It should be noted that the effect of salicylic
11 acid varied according to the applied concentration; CAT, APX and GR activities were stimulated
12 the maximum at 0.5 mM SA under high salinity while SOD activity was stimulated at 1 mM SA.
13 Taken all together, even if the genotype Aguadulce performed better than the genotype Histal in
14 terms of growth, physiological and molecular response under salt stress, the difference between
15 genotypes remains more quantitative rather than qualitative. Salicylic acid application improved
16 significantly the salt-tolerance of both genotypes throughout the maintenance of cell membrane
17 and photosynthetic process and the diminution of sodium toxicity and oxidative damage.
18 Nevertheless, the action of salicylic acid varies in function of the applied concentration of the
19 hormone, the tested genotype and the studied parameter. Overall, the genotype Aguadulce
20 performs better with the treatment 0.5 mM SA while the genotype Histal manifests greater
21 behaviour with the treatment 1 mM SA.

22

23

24 **5. Conclusion**

1 The intraspecific comparative studies among genotypes for salt-tolerance might help to identify
2 the relevant mechanisms correlating their relative degree of tolerance to salinity and to select the
3 most suited genotypes for very specific environmental conditions.

4 The present study revealed that the difference between genotypes is more quantitative rather than
5 qualitative even if the if the genotype Aguadulce performed better in terms of growth,
6 physiological and molecular response under salt stress. In general, the salt stress promoted several
7 limitations in growth, gas exchanges, photosynthesis, ion homeostasis and other physiological
8 attributes. Nevertheless, faba bean plants have triggered some mechanisms to deal with this
9 constraint throughout the maintenance of water status and the enhancement of antioxidant
10 enzymatic activities. However, the salt-tolerance of both the two genotypes was significantly
11 improved by salicylic acid application which allowed the maintenance of cell membrane and
12 photosynthetic process, restoring of ion homeostasis and the diminution of oxidative damages. The
13 beneficial effect of salicylic acid varies according to its concentration, the tested genotype and the
14 studied parameter. Overall, the genotype Aguadulce performs better under the treatment with 0.5
15 mM SA while the genotype Hystal manifests greater behaviour under the treatment with 1 mM SA.

16 Based on our results, salicylic acid can be considered as potential growth regulator to improve the
17 salt response of faba bean. Plants treatment with salicylic acid might constitute a sustainable
18 approach and effective alternative to restore the adverse effects of salinity. The application of
19 salicylic acid would provide a practical basis for wide cultivation of faba bean and might propose
20 an ecological and economical solution for horticulture in salt-affected soils, mainly in arid regions,
21 for the reclamation of marginal and wastelands under-cultivated.

22

1 6. References

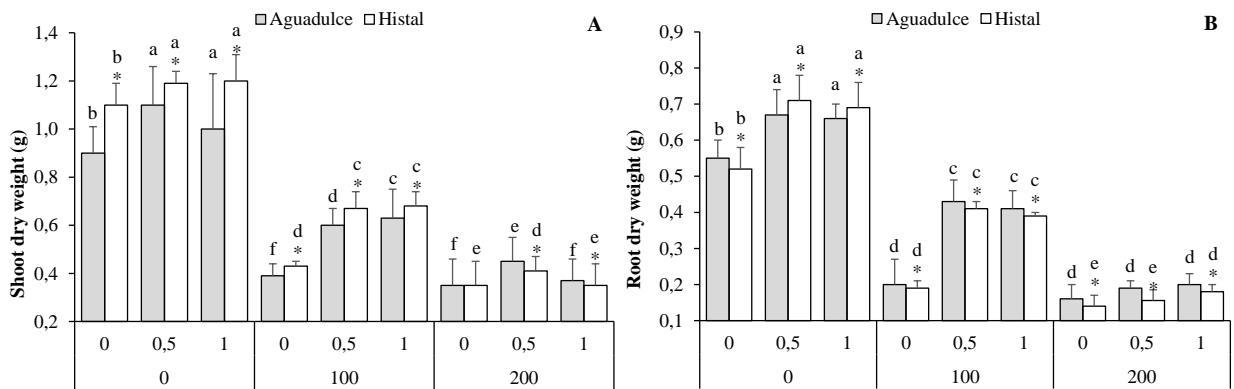
- 2 Abdul Qados, A.M.S., 2011. Effect of salt stress on plant growth and metabolism of bean plant
3 *Vicia faba* (L.). *Journal of the Saudi Society of Agricultural Sciences* 10, 7-15.
- 4 Ahmad, P., Alyemeni, M.N., Ahanger, M.A., Egamberdieva, D., Wijaya, L., Alam, P., 2018.
5 Salicylic Acid (SA) Induced Alterations in Growth, Biochemical Attributes and
6 Antioxidant Enzyme Activity in Faba Bean (*Vicia faba* L.) Seedlings under NaCl Toxicity.
7 *Russian Journal of Plant Physiology* 65, 104-114.
- 8 Almeida, D.M., Oliveira, M.M., Saibo, N.J.M., 2017. Regulation of Na⁺ and K⁺ homeostasis in
9 plants: towards improved salt stress tolerance in crop plants. *Genet Mol Biol* 40, 326-345.
- 10 Ashraf, M., Harris, P.J.C., 2013. Photosynthesis under stressful environments: An overview.
11 *Photosynthetica* 51, 163-190.
- 12 Bulut, F., Akıncı, Ş., Eroğlu, A., 2011. Growth and Uptake of Sodium and Potassium in Broad
13 Bean (*Vicia faba* L.) under Salinity Stress. *Communications in Soil Science and Plant*
14 *Analysis* 42, 945-961.
- 15 Csiszár, J., Horváth, E., Váry, Z., Gallé, Á., Bela, K., Brunner, S., Tari, I., 2014. Glutathione
16 transferase supergene family in tomato: Salt stress-regulated expression of representative
17 genes from distinct GST classes in plants primed with salicylic acid. *Plant Physiology and*
18 *Biochemistry* 78, 15-26.
- 19 Dawood, M.G., El-Awadi, M.E., 2015. Alleviation of salinity stress on *Vicia faba* L. plants via
20 seed priming with Melatonin. *Acta Biológica Colombiana* 20, 223-235.
- 21 Demiral, T., Türkan, İ., 2005. Comparative lipid peroxidation, antioxidant defense systems and
22 proline content in roots of two rice cultivars differing in salt tolerance. *Environmental and*
23 *Experimental Botany* 53, 247-257.
- 24 El-Tayeb, M.A., 2005. Response of barley grains to the interactive effect of salinity and salicylic
25 acid. *Plant Growth Regulation* 45, 215-224.
- 26 Isayenkov, S.V., Maathuis, F.J.M., 2019. Plant Salinity Stress: Many Unanswered Questions
27 Remain. 10.
- 28 Jayakannan, M., Bose, J., Babourina, O., Rengel, Z., Shabala, S., 2013. Salicylic acid improves
29 salinity tolerance in *Arabidopsis* by restoring membrane potential and preventing salt-
30 induced K⁺ loss via a GORK channel. *J Exp Bot* 64, 2255-2268.
- 31 Khan, M.I.R., Asgher, M., Khan, N.A., 2014. Alleviation of salt-induced photosynthesis and
32 growth inhibition by salicylic acid involves glycinebetaine and ethylene in mungbean
33 (*Vigna radiata* L.). *Plant Physiology and Biochemistry* 80, 67-74.

- 1 Khan, M.I.R., Fatma, M., Per, T.S., Anjum, N.A., Khan, N.A., 2015. Salicylic acid-induced
2 abiotic stress tolerance and underlying mechanisms in plants. 6.
- 3 Lawlor, D.W., Cornic, G., 2002. Photosynthetic carbon assimilation and associated metabolism
4 in relation to water deficits in higher plants. *Plant, cell & environment* 25, 275-294.
- 5 Li, L., Zhang, H., Zhang, L., Zhou, Y., Yang, R., Ding, C., Wang, X., 2014a. The physiological
6 response of *Artemisia annua* L. to salt stress and salicylic acid treatment. *Physiol Mol Biol*
7 *Plants* 20, 161-169.
- 8 Li, T., Hu, Y., Du, X., Tang, H., Shen, C., Wu, J., 2014b. Salicylic Acid Alleviates the Adverse
9 Effects of Salt Stress in *Torreya grandis* cv. *Merrillii* Seedlings by Activating Photosynthesis
10 and Enhancing Antioxidant Systems. *PLOS ONE* 9, e109492.
- 11 Maribel, D.-S., Tobita, S., 1998. Antioxidant responses of rice seedlings to salinity stress. *Plant*
12 *Science* 135, 1-9.
- 13 Moussa, H.R., Hassan, M.A.E.-F., 2016. Growth Enhancers to Mitigate Salinity Stress in *Vicia*
14 *fabae*. *International Journal of Vegetable Science* 22, 243-250.
- 15 Nazar, R., Umar, S., Khan, N.A., 2015. Exogenous salicylic acid improves photosynthesis and
16 growth through increase in ascorbate-glutathione metabolism and S assimilation in mustard
17 under salt stress. *Plant Signaling & Behavior* 10, e1003751.
- 18 Taïbi, K., Del Campo, A.D., Vilagrosa, A., Bellés, J.M., López-Gresa, M.P., López-Nicolás,
19 J.M., Mulet, J.M., 2018. Distinctive physiological and molecular responses to cold stress
20 among cold-tolerant and cold-sensitive *Pinus halepensis* seed sources. *BMC Plant Biol* 18,
21 236-236.
- 22 Taïbi, K., del Campo, A.D., Vilagrosa, A., Bellés, J.M., López-Gresa, M.P., Pla, D., Calvete, J.J.,
23 López-Nicolás, J.M., Mulet, J.M., 2017. Drought Tolerance in *Pinus halepensis* Seed
24 Sources As Identified by Distinctive Physiological and Molecular Markers. 8.
- 25 Taïbi, K., Taïbi, F., Ait Abderrahim, L., Ennajah, A., Belkhodja, M., Mulet, J.M., 2016. Effect of
26 salt stress on growth, chlorophyll content, lipid peroxidation and antioxidant defence
27 systems in *Phaseolus vulgaris* L. *South African Journal of Botany* 105, 306-312.
- 28 Taïbi, K., Taïbi, F., Belkhodja, M., 2012. Effects of external calcium supply on the physiological
29 response of salt stressed bean (*Phaseolus vulgaris* L.). *Genetics and Plant Physiology* 2, 177-
30 186.
- 31 Taïbi, K., Taïbi, F., Belkhodja, M., 2013a. Plants growth, water relations and photosynthesis of
32 two bean genotypes *Phaseolus vulgaris* L. treated with NaCl and fluridone. *African Journal*
33 *of Biotechnology* 12, 3811-3821.

- 1 Taïbi, K., Taïbi, F., Belkhodja, M., 2013b. Salinity Effects on the Physiological Response of
- 2 Two Bean Genotypes (*Phaseolus vulgaris* L.). Arab Gulf Journal of Scientific Research 31,
- 3 90-98.
- 4 Wu, J., Seliskar, D.M., Gallagher, J.L., 1998. Stress tolerance in the marsh plant *Spartina patens*:
- 5 Impact of NaCl on growth and root plasma membrane lipid composition. *Physiologia*
- 6 *Plantarum* 102, 307-317.
- 7
- 8

1 **Figures**

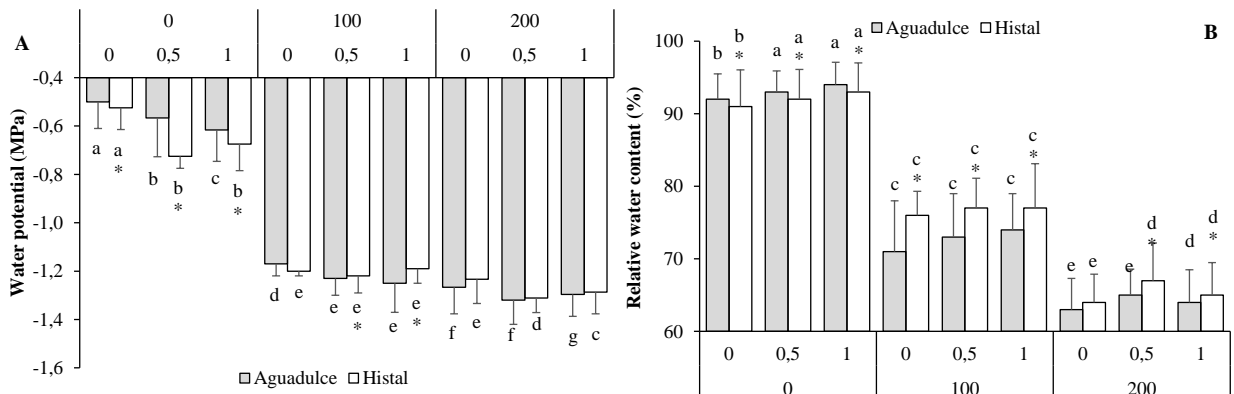
2
3



4
5 **Figure 1.** Effects of salicylic application on shoot (a) and root (b) dry weight of two faba bean
6 genotypes Aguadulce (grey bars) and Hystal (white bars) subjected to salinity.

7 The letters above the bars marks the significant difference among the different treatments with salt and
8 salicylic acid for each genotype following the post hoc Duncan's test. The asterisk shows the significant
9 difference between the two genotypes within the same treatment. Scale bars are mean + SE, being the
10 number of samples $n = 8$.

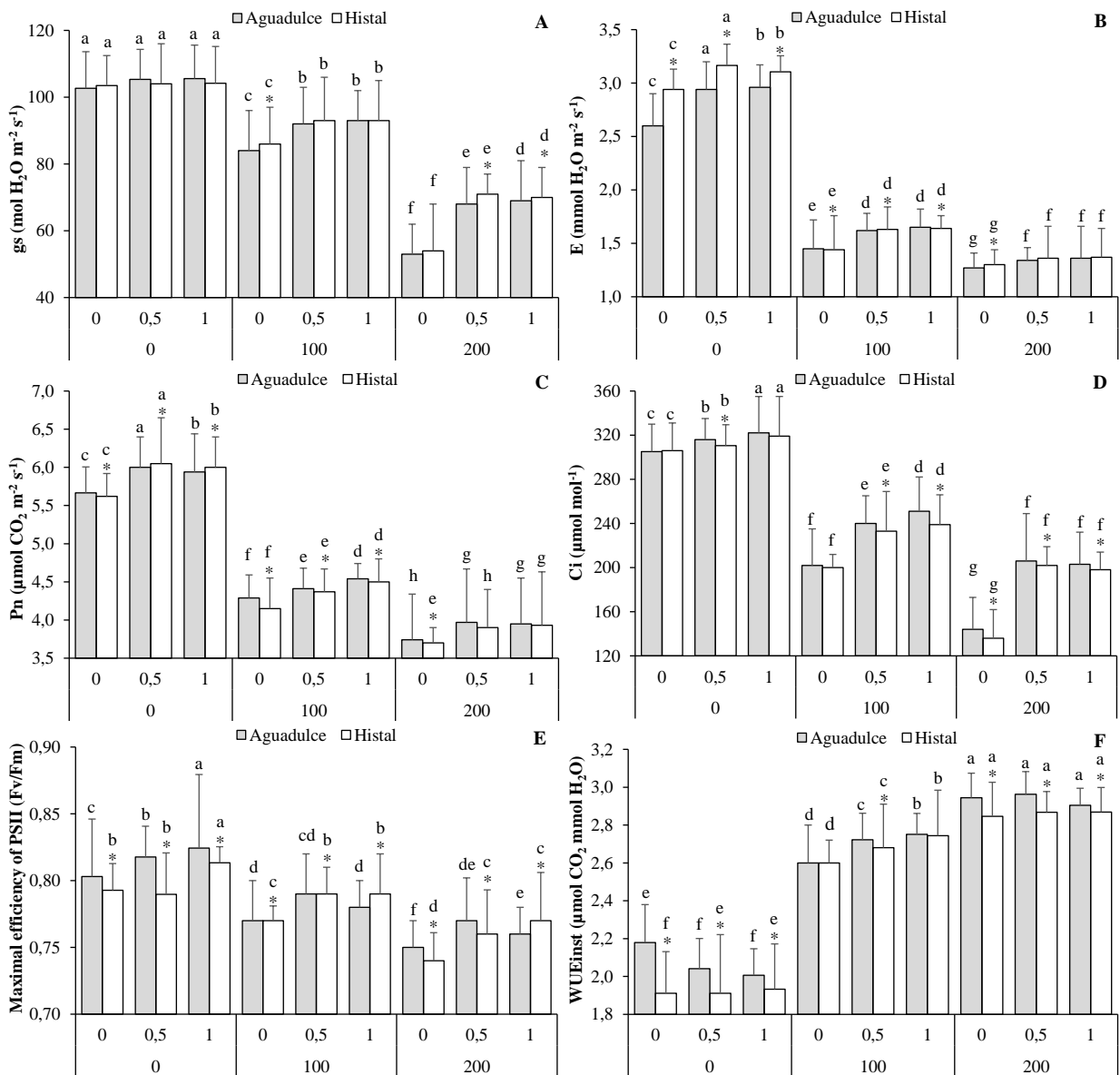
11
12



13
14 **Figure 2.** Effects of salicylic application on leaf water potential (a) and relative water content
15 (b) of two faba bean genotypes Aguadulce (grey bars) and Hystal (white bars) subjected to
16 salinity.

17 The letters above the bars marks the significant difference among the different treatments with salt and
18 salicylic acid for each genotype following the post hoc Duncan's test. The asterisk shows the significant
19 difference between the two genotypes within the same treatment. Scale bars are mean + SE, being the
20 number of samples $n = 8$.

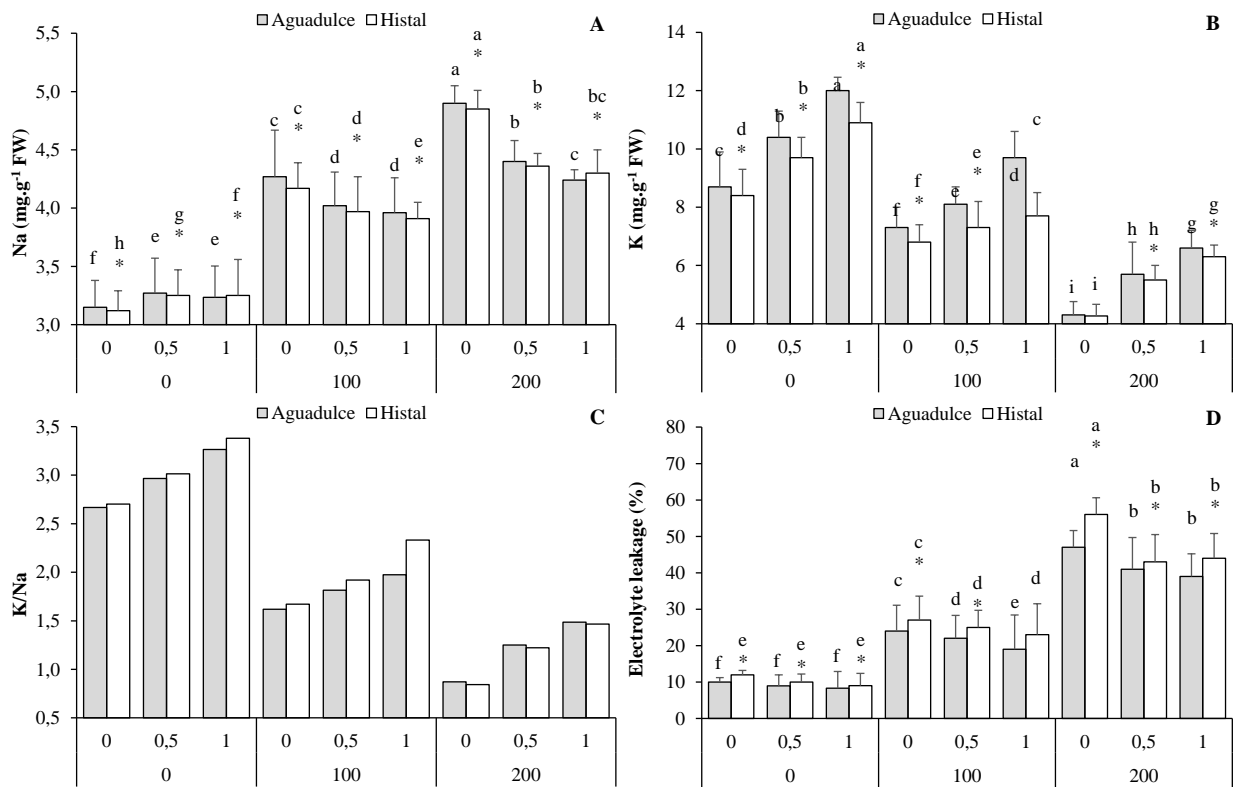
21



1
 2 **Figure 3.** Effects of salicylic application on gas exchanges and photosynthesis of two faba bean
 3 genotypes Aguadulce (grey bars) and Hystal (white bars) subjected to salinity.

4 Stomatal conductance (gs) (a), transpiration (E) (b), Net photosynthesis (Pn) (c), intercellular CO₂
 5 concentration (Ci) (d), Maximal efficiency of PSII (e), and instantaneous water use efficiency (WUEinst)
 6 (f).

7 The letters above the bars marks the significant difference among the different treatments with salt and
 8 salicylic acid for each genotype following the post hoc Duncan's test. The asterisk shows the significant
 9 difference between the two genotypes within the same treatment. Scale bars are mean + SE, being the
 10 number of samples $n = 8$.



1

2 **Figure 4.** Effects of salicylic application on ions content and electrolyte leakage of two faba

3 bean genotypes Aguadulce (grey bars) and Hystal (white bars) subjected to salinity.

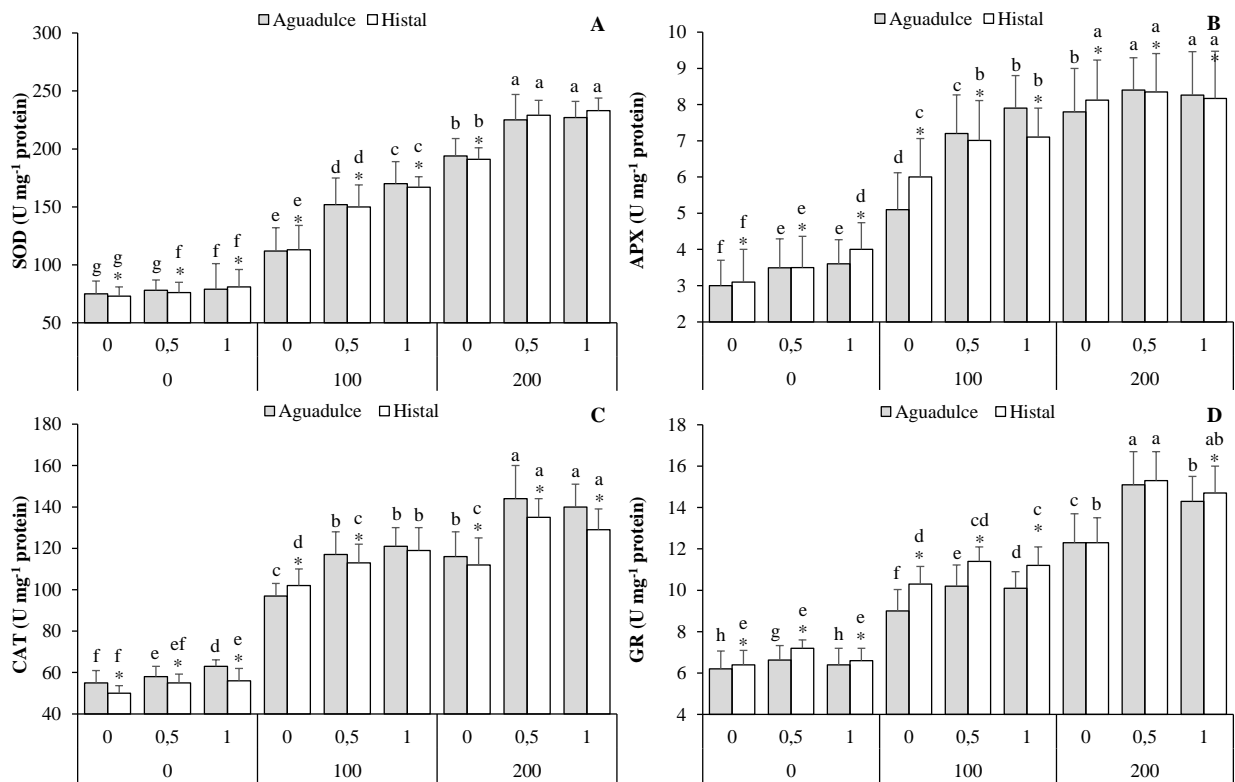
4 Sodium content (Na⁺) (a), potassium content (K⁺) (b), ratio K⁺/Na⁺ (c) and electrolyte leakage (d)

5 The letters above the bars marks the significant difference among the different treatments with salt and

6 salicylic acid for each genotype following the post hoc Duncan's test. The asterisk shows the significant

7 difference between the two genotypes within the same treatment. Scale bars are mean + SE, being the

8 number of samples $n = 8$.



1

2 **Figure 5.** Effects of salicylic application on antioxidant enzymes activities of two faba bean
 3 genotypes Aguadulce (grey bars) and Hystal (white bars) subjected to salinity.

4 Superoxide dismutase (SOD) (a), Ascorbate peroxidase (APX) (b), Catalase (CAT) (c) and Glutathione
 5 reductase (GR) (d).

6 The letters above the bars marks the significant difference among the different treatments with salt and
 7 salicylic acid for each genotype following the post hoc Duncan's test. The asterisk shows the significant
 8 difference between the two genotypes within the same treatment. Scale bars are mean + SE, being the
 9 number of samples $n = 8$.

10