Document downloaded from:

http://hdl.handle.net/10251/176231

This paper must be cited as:

Martinez-Perez-Cejuela, H.; Mompo-Rosello, O.; Crespi-Sanchez, N.; Palomino Cabello, C.; Catalá-Icardo, M.; Simo-Alfonso, EF.; Herrero-Martinez, JM. (2020). Determination of benzomercaptans in environmental complex samples by combining zeolitic imidazolate framework-8-based solid-phase extraction and high-performance liquid chromatography with UV detection. Journal of Chromatography A. 1631:1-9. https://doi.org/10.1016/j.chroma.2020.461580



The final publication is available at https://doi.org/10.1016/j.chroma.2020.461580

Copyright Elsevier

Additional Information

| 1 | Determination of ber | zomercaptans in environmental complex samples by | | | | | |
|----|--|---|--|--|--|--|--|
| 2 | combining zeolitic imid | lazolate framework-8-based solid-phase extraction and | | | | | |
| 3 | high-performa | nce liquid chromatography with UV detection | | | | | |
| 4 | | | | | | | |
| 5 | H. Martínez-Pérez-Cejuel | a ^{a+} , O. Mompó-Roselló ^{a+} , N. Crespí-Sánchez ^b , C. Palomino | | | | | |
| 6 | Cabello ^b , M. Catalá-Icardo ^c , E.F. Simó-Alfonso ^a , J.M. Herrero-Martínez ^a * | | | | | | |
| 7 | | | | | | | |
| 8 | ^a Department of Analytical | Chemistry, University of Valencia, C/Dr. Moliner, 50, 46100- | | | | | |
| 9 | Burjassot, Valencia, Spain. | | | | | | |
| 10 | ^b Department of Chemistry, | University of the Balearic Islands, Cra. Valldemossa km 7.5, | | | | | |
| 11 | 07122 Palma de Mallorca, Spain. | | | | | | |
| 12 | ^c Instituto de Investigación para la Gestión Integrada de Zonas Costeras, Campus de | | | | | | |
| 13 | Gandía, Universitat Politèc | cnica de València, C/ Paranimf no 1, 46730 Grao de Gandia, | | | | | |
| 14 | Valencia, Spain. | | | | | | |
| 15 | | | | | | | |
| 16 | *Corresponding author: | Dr. José Manuel Herrero-Martínez | | | | | |
| 17 | | Tel.: +34963544062 | | | | | |
| 18 | | Fax: +34963544436 | | | | | |
| 19 | | e-mail: jmherrer@uv.es | | | | | |
| 20 | | | | | | | |
| 21 | ⁺ Both authors contributed e | equally to this manuscript. | | | | | |
| 22 | | | | | | | |

23 Abstract

24 In this work, the synthesis of zeolitic imidazolate framework-8 (ZIF-8) crystals and their 25 subsequent application as effective sorbents for extraction and preconcentration of 26 several benzomercaptans from environmental complex samples is described. These 27 materials were prepared by solvothermal approach varying the concentration of n-28 butylamine modulator to modify the surface of the metal-organic framework. The 29 resulting materials were characterized by scanning and transmission electron microscopy, 30 powder X-ray diffraction and Fourier transform infrared spectroscopy. The ZIF-8 31 material that gave the best features was selected as extractive phase and the influence of 32 various parameters (sample pH and elution solvent composition, among others) on the 33 extraction efficiency of target compounds were investigated. Under the optimal 34 conditions of the method, the tested analytes (2-mercaptobenzothiazole, 2-35 mercaptobenzoxazole and 2-mercapto-6-nitrobenzothiazole) were retained and eluted 36 quantitatively with alkaline 50:50 (v:v) methanol-water mixture. Using the proposed method, low limits of detection, in the range of 16-21 ng L^{-1} for aqueous samples and 0.4-37 0.5 µg kg⁻¹ for soil samples, were achieved whereas the precision (expressed as relative 38 39 standard deviation) was lower than 7%. The resulting solid-phase extraction protocol, 40 using the zeolitic material as sorbent, was combined with liquid chromatography and 41 ultraviolet-vis detector and successfully applied to determine traces of these organic 42 pollutants in environmental samples.

Keywords: Metal-organic frameworks; environmental pollutants; soil and aqueous samples; solid-phase extraction; sorbent; sample preparation

46 1. Introduction

47 Metal-organic frameworks (MOFs) have aroused a great interest over the last decades. 48 These coordination polymers are a class of highly porous materials composed of metal 49 ions or clusters linked by organic ligands. Their crystalline structures, including their pore 50 sizes and surfaces, can be controlled by a careful selection of their precursors. MOFs are 51 characterized by a large specific surface area and intrinsic porosity, as well as profuse 52 active sites [1]. Many applications have been proposed for this type of materials, and 53 surely, their number will grow in the coming years. Some of those applications include 54 gas/electrical storage and catalysis [2], removal of contaminants in air or water by 55 adsorption and separation [3-4], sensors [5] and sorbents for sample preparation [6]. In 56 particular, zeolitic imidazolate frameworks (ZIFs) are a subfamily of MOFs consisting of 57 Zn or Co metal ions tetrahedrally coordinated by anionic imidazolate ligands [7, 8]. ZIF-58 8 [Zn(2-methylimidazolate)₂] has been widely used due to its easy synthesis, high thermal 59 and chemical stabilities [9, 10], great pore volume and surface area [9]. ZIF-8 shows a 60 very large adsorption capacity, which is attributed to its high porosity and the different 61 ways in which it can interact with adsorbates, such as π -stacking, hydrogen bonding and 62 ionic interactions [11, 12]. Such good features have become this material in one of the 63 most attractive MOFs for the extraction and preconcentration of analytes. Related to this, 64 solid-phase microextraction (SPME) [6] has been commonly reported as a suitable 65 sample preparation technique in the extraction of amines [13], polycyclic aromatic 66 hydrocarbons [14], BTEX [15] and fluoroquinolones [16] from environmental water 67 samples. Despite its advantages, SPME has certain disadvantages such as the limited fiber 68 lifetime, its fragility and its relatively high cost. In this sense, other alternatives based on 69 conventional solid-phase extraction (SPE) format (syringe/cartridges) [17], magnetic SPE 70 [18] or the use of membrane as support medium [19-21] have been developed. However, 71 the preparation of homogenous ZIF-8 membranes is a difficult task presenting some 72 drawbacks such as possible sedimentation phenomena of MOF crystals during the 73 preparation stage, the formation of bubbles during stirring (poor reproducibility) and 74 limited reusability (blockage of pores of membrane). Besides, in most of these works, the 75 selectivity of sorbent has been attributed to the hydrophobic and π -interactions of solutes 76 with the 2-methylimidazole ligands instead of taking advantages of their tunable pore size 77 and feasible chemical functionalization [22].

78 The pollution of the environment by organic compounds is always of great concern due 79 to the toxicity risks for aquatic organisms and human posed by their occurrence and

80 increasing concentration in water bodies. Among organic pollutants, benzomercaptans, 81 such as 2-mercaptobenzothiazole (MBT) and 2-mercaptobenzoxazole (MBO), deserve 82 special attention due to their broad use in many industrial processes. These compounds 83 are commonly used as vulcanization accelerators in the rubber industry, as biocorrosion 84 inhibitors, as coating agents of metallic surfaces, and as biocides in medical applications 85 [23-24]. As consequence, such compounds have been found in different environmental 86 compartments such as surface waters, wastewater effluents, sewage treatment plants, 87 soils, roadsides, etc. These poorly biodegradable pollutants may cause allergic reactions, 88 induce tumours, and be toxic to aquatic organisms [25-26]. According to the NSF 89 International Standard/American National Standard (NSF/ANSI 60-2016), the drinking 90 water criteria (Annex C) for MBT is maximum contamination levels or maximum 91 allowable concentration, respectively of total allowable concentration is $20 \,\mu g \, L^{-1}$, single product allowable concentration is 2 μ g L⁻¹ [27]. In order to reach these low 92 93 concentrations, determination techniques mainly based on LC combined with mass 94 spectrometry detectors have been reported [28-31]. In any case, sample preparation 95 techniques are required in order to reduce matrix interferences and to improve sensitivity 96 to benzomercaptans in environmental complex samples, being SPE the most common 97 technique used for extraction and preconcentration of these kind of analytes [23, 24, 28, 98 30, 31]. However, SPE shows some disadvantages, such as limited selectivity and poor 99 reusability.

100 In order to overcome the above-mentioned drawbacks, there is a recent interest concerned 101 to functionalized MOFs, which are obtained by incorporation of ligands including 102 additional functional groups (-OH, -NH₂, ...). Particularly, amine-functionalized MOFs 103 have mainly attracted attention since they improved MOF water stability and provided 104 the possibility of forming hydrogen bonds and even electron transfer capability towards 105 analytes [32]. Recently, these materials have applied for capturing of several polar guests 106 from different matrices. Thus, NH₂-MIL-101(Fe) and NH₂-MIL-53(Al) have been used 107 for the removal of imidacloprid in water [33] and to selectively recognize tetracyclines 108 from milk [34], respectively. Also, amino-functionalized UiO-66 combined with 109 magnetic Fe₃O₄ microspheres has been used for the selective extraction of bisphenols in 110 river water samples [35]. In these works, the introduction of amino moieties is 111 accomplished by one-pot solvothermal synthesis MOFs, which has the advantage of 112 avoiding the complicated steps required in post-modification approach. Alternatively, 113 amino moieties can be incorporated through the addition of organic amines as modulators

in the synthesis mixture of MOF [22]. However, this approach has not been explored inliterature with sample treatment purposes.

116 In this work, amine-modulated ZIF-8 (nano)crystals (NH₂-ZIF-8) was used as sorbent for 117 the extraction of these target compounds (MBO, MBT and its nitro derivative) in 118 environmental samples. The incorporation of n-butylamine molecules to the framework 119 structure resulted appropriate to tailor the surface properties improving the selectivity of 120 the extraction of these polar compounds. Although this type of material has been 121 previously used in hydrophobic compounds in dispersive mode with sophisticated 122 technique (GC-MS) [36], its application to hydrophilic compounds in cartridge format 123 with accessible instrumentation (LC-UV) has not been explored. Taking into account the 124 key role of butylamine (BA) as modulator, in this work, its content on crystal structure 125 and posterior interaction with analytes was studied. The resulting MOF that provided the 126 best features was selected as SPE sorbent and parameters affecting the extraction process 127 such sample pH and eluent solvent composition (among others) were investigated. 128 Finally, the applicability of the method for the analysis of these compounds by HPLC-129 UV was evaluated in environmental samples obtained from different sources.

130

131 **2. Experimental section**

132 2.1. Reagents and materials

133 Zinc nitrate hexahydrate, 2-methylimidazole (Hmim), n-butylamine (BA), ethylamine 134 and hexylamine were purchased from Sigma-Aldrich (Milwaukee, WI, USA). Sodium 135 hydroxide was obtained from Panreac Química SA (Barcelona, Spain). Acetonitrile 136 (ACN), methanol (MeOH), acetic acid (HAcO) and hydrochloric acid were from VWR 137 International Eurolab (Barcelona, Spain). The benzomercaptans selected in this work 138 were: 2-mercaptobenzoxazole (MBO), 2-mercaptobenzothiazole (MBT) and 6-139 nitrobenzo[d]thiazole-2-thiol (NMBT), all purchased from Sigma-Aldrich. The structures 140 of these compounds are given in Table S1. Deionized water was prepared in Crystal B30 141 EDI Adrona deionizer (Riga, Latvia). All other reagents were of analytical grade unless 142 otherwise stated. 1 mL empty propylene disposable SPE cartridge and frits $(1/16', 20 \,\mu\text{m})$ 143 were from Análisis Vínicos (Tomelloso, Spain). Individual standard solutions of benzomercaptans (1000 mg L⁻¹) were prepared in MeOH and stored at 4 °C until its use. 144 145 Working standard solutions were done daily by appropriate dilution from these stock 146 solutions.

148 2.2. Instrumentation

149 Scanning electron microscopy (SEM) images were obtained with a Hitachi S-4800 150 electron microscope (Ibaraki, Japan) equipped with a retrodispersive electron detector 151 and an energy dispersive spectrometer (EDAX Genesis 4000). A transmission electron 152 microscope (TEM) coupled to a digital camera AMT RX80 model JEM-1010 JEOL 153 (Akishima, Japan) was used to obtain the transmission electron micrographs. Adsorption-154 desorption isotherms of nitrogen were registered in a S5 Micromeritics ASAP2020 155 instrument (Norcross, USA) at 77 K. The Brauner-Emmet-Teller (BET) model was used 156 to calculate the specific surface area using the low-pressure range. Powder X-ray 157 Diffraction (p-XRD) patterns were obtained in a D8 Advance A25 diffractometer 158 (Bruker). Attenuated total reflection Fourier-transform infrared (FT-IR) spectra of 159 powdered materials were registered with a DuraSamplIR II accessory from Smiths 160 Detection Inc. (Warrington, UK) equipped with a nine reflection diamond/ZnSe 161 DuraDisk plate, installed on a Bruker FT-IR spectrometer (Bremen, Germany) model 162 Tensor 27. The zeta potential measurements were performed with Zetasizer Nano ZS 163 equipment (Malvern Instruments, Malvern, UK). NH₂-ZIF-8-BA 1 was dispersed in distilled water at a concentration of 1 g L⁻¹ and sonicated during 1 min. Measurements 164 were carried out per triplicate at 25 °C. The final zeta potential value was estimated from 165 166 the particle mobility using the Smoluchowski model.

167 Chromatographic separation of standards and samples was performed on a 1260 Infinity 168 II HPLC from Agilent Technologies (Waldbronn, Germany) with a UV-Vis diode-array 169 detector. The separation conditions of benzomercaptans were adapted from Parham et al. 170 [30]. The chromatographic column was a Kromasil C18 (250×4.0 mm, 5 µm particle 171 size, Análisis Vínicos). Separation was done under gradient elution mode (at flow rate of 172 1.0 mL min⁻¹) using as solvents: (A) water and (B) ACN containing both 0.1% (v/v) of 173 HAcO. The gradient elution and the detection wavelength programs are given in 174 supplementary information (see Table S2).

175

176 2.3. Synthesis of ZIF-8 crystals

177 ZIF-8 materials were synthesized following protocols reported in the literature by 178 Cravillon *et al.* [22, 37]. To obtain ZIF-8 (nano)crystals (without modulator), two 179 methanolic solutions were needed. Solution (A) composed of $Zn(NO_3)_2 \cdot 6H_2O$ (9.87 180 mmol) in MeOH (200 mL) and solution (B) of 2-methylimidazole (79.04 mmol) in MeOH 181 (200 mL). The later solution was added to the former under stirring. After keeping at

182 room temperature for 24 h, a white dispersion of ZIF-8 was formed, and separated by 183 centrifugation at 6000 rpm for 10 min. Then, the resulting solid was washed with MeOH 184 three times and dried at 40°C overnight. On the other hand, ZIF-8 nanocrystals using BA 185 (NH₂-ZIF-8-BA) as modulator agent were prepared as given elsewhere [22]. A ligand 186 solution (B) (9.874 mmol of Hmim in 50 mL MeOH) including the modulator agent 187 (9.874 mmol of n-BA) was poured slowly into a solution (A) containing the metallic 188 cation (2.469 mmol of Zn(NO₃)₂·6H₂O in 50 mL of MeOH) under stirring and left at 189 room temperature for 24 h. The resulting white solid (namely, NH₂-ZIF-8-BA 1) was 190 recovered and washed with the same protocol that is described above for ZIF-8 191 (nano)crystals. Also, different molar contents of BA as modulator (5 and 20 mmol, NH₂-192 ZIF-8-BA 2 and 3, respectively) were tested by keeping constant the molar ratio of other 193 starting reagents. Additionally, other alkyl amines (ethyl- and hexylamine) were 194 investigated as modulator agent (at 10 mmol) following the abovementioned protocol for 195 NH₂-ZIF-8-BA materials.

196

197 2.4. Sample collection and treatment

The sampling of water was performed from different locations in Valencia.
Approximately, 500 mL were collected in dark containers and refrigerated at 4°C until
their use.

201 Several soil samples from different sources were collected, air-dried at room temperature, 202 crushed and then sieved through a 1 mm sieve. These samples were stored on 203 polypropylene bags in darkness at 25°C. The pH, humidity and electrical conductivity of 204 soil samples were measured following ISO rules [38]. pH and electrical conductivity were 205 measured in a 1:5 (soil:deionized water) extract shaken for 5 min and measured after 2 h. 206 For the determination of humidity, 1 g of each sample was heated at 70°C overnight until 207 constant weight. These soil parameters are listed in Table S3. The extraction of 208 benzomercaptans from soil samples was adapted from Zhang et al [39], as follows. Briefly, 0.1 g approx. of each sample was weighed and spiked at 500 μ g kg⁻¹ with the 209 210 investigated pollutants. After vortex-assisted and ultrasonic homogenization, the samples 211 were allowed to stand with the analytes for 2 h. Then, 5 mL of MeOH were added, shaken 212 during 20 min and centrifuged at 6000 rpm for 3 min. The supernatant was collected and 213 the extraction procedure was repeated twice. All methanolic fractions were combined and 214 evaporated at low pressure up to small volume (ca. 100 μ L) and reconstituted up to 5 mL 215 with water before application of SPE protocol described below.

216

217 2.5. Extraction procedure

218 A SPE protocol using the developed materials as sorbents was applied to the extraction 219 of aromatic benzomercaptans in environmental samples. For the preparation of SPE 220 devices, 20 mg of the metal-organic framework was packed between two frits into 1 mL 221 empty polypropylene cartridges. Prior to loading, the sewage waters were sonicated and 222 centrifuged, and the extraction was carried out following the protocol described above. 223 The material was firstly conditioned with MeOH (500 μ L) and water (500 μ L), and after 224 that, a known volume of standard solution or sample (5 mL and 25 mL for soil and water 225 samples, respectively) was passed through the cartridge. The sorbent was finally washed 226 with water (500 μ L). The retained analytes were later eluted with 250 μ L of 50% (v/v) MeOH containing 0.5 mol L⁻¹ NaOH. The extraction units were regenerated with water 227 228 (10 mL) in order to eliminate the NaOH excess. Prior to HPLC injection, the elution 229 fraction was properly neutralized with hydrochloric acid.

230

231 **3. Results and discussion**

232 3.1. Preliminary considerations

As it was mentioned in the Introduction, the selection of the adequate MOF as SPE sorbent is a key aspect since it strongly influences on the selective retention and the way of interaction with the target analytes. In this sense, ZIF-8 material, due to its pore size and the kind of interactions it can establish with the analytes, has advantageous features making it a good SPE phase candidate. As shown in Fig. S1, the molecular sizes of MBO and MBT are smaller than the pore size of ZIF-8 (3.4 Å) [40, 41]; whereas for NMBT, a partial embedding process may happen into ZIF-8 structure.

240 Regarding the interaction forces involved, there are different possible interaction modes 241 between this MOF and benzomercaptans: i) hydrophobic effects and π -interaction 242 between the imidazole rings of ZIF-8 and the aromatic moieties of these analytes (see 243 structures in Fig. S1); ii) hydrogen bonding between NH groups in ZIF-8, and certain 244 hydrogen-bond accepting groups (e.g. ether, organo sulfurs, nitro, among others) present 245 in the analytes (Fig. S1) thus enhancing the hydrophilic binding and improving the 246 selectivity; and iii) electrostatic interaction between ZIF-8 and benzomercaptan 247 molecules. In this sense, the surface charge of ZIF-8 nanocrystals and the ionization 248 degree of benzomercaptans (see Table S1) should be considered (as we discussed below). 249 Additionally, other potential contributions due to the amine moiety such as cation- π

interaction between aromatic ring and amino protonated ZIF-8 structure can occur [42],
which have been previously reported with other amino functionalized MOFs and
compounds with structural similarities [43]. In this regard, the synergetic contribution of
BA modulator of the NH₂-ZIF-8-BA in these last interactions can be explained by its
presence onto the surface of ZIF-8 crystals as previously demonstrated [36].

255 Indeed, the use of modulators during MOF synthesis has proven to be a promising strategy 256 through which the internal MOF structure (crystal size and morphology), and outer 257 surface functionalities, can be controlled by using monodentate ligands [44]. The 258 modulators then compete with conventional multidentate ligands for coordination to the 259 metal cations [45, 46]. In order to study the influence of BA upon the crystal growth of 260 ZIF-8 and other features, several materials were synthesized varying its molar ratio. 261 Taking into account all these considerations, the ability of these synthesized materials as 262 SPE phases is here evaluated (see Table 1).

263

264 *3.2. Characterization of material*

265 The morphology and crystal size of the prepared materials (ZIF-8 and NH₂-ZIF-8 with 266 different modulators) were investigated by SEM and TEM analysis, respectively. Fig. 1A 267 and B shows the SEM micrograph of the ZIF-8 (without modulator) and NH₂-ZIF-8-BA 268 1 (corresponding to *ca*. 10 mmol of BA) materials, respectively. In both images, a 269 polyhedral shape of the nanocrystals was evidenced, which is in agreement with the 270 typical form of ZIF-8 nanocrystals [40, 41]. Also, SEM images of MOFs prepared at other 271 BA contents were taken (Fig. S2). As it can be seen, the morphology did not change 272 substantially with modulator content. The TEM images of the resulting materials for this 273 set of experiments are summarized in Fig. S2, and the mean sizes of the crystals are given 274 in Table 1. As it can be seen, an increase of the concentration of BA modulator led to the 275 increased mean size of the resulting crystals. The tendency observed for BA-modulated 276 MOFs is consistent with that reported in previous studies focused on the addition of 277 monocarboxylic acid modulators [44, 47]. These results can be explained taking into 278 account that modulating ligands can tailor crystal nucleation and growth via both 279 coordination and deprotonation equilibria [22]. Thus, the addition of a basic ligand (L) as 280 BA (pK_a value in aqueous media is 10.7) can deprotonate the [Zn(Hmim)_mL_n] species, 281 and thus producing an acceleration of rate of ligand exchange reactions, resulting in a 282 higher nucleation rate and therefore in a smaller final crystal size. When a low amount of 283 BA is added a, a large number of nuclei were formed and rapidly grow at the same time

affording smaller crystals. However, higher concentration of BA provide a slow nucleation (fewer nuclei) of the MOF giving as result larger crystals. This is due to this additive starts to have a relevant role as a competitive ligand in the complex formation. On the other hand, in absence of modulating ligand (BA), there is not a deprotonation reaction, which led to small concentration of $[Zn(Hmim)_mL_n]$ species decreasing the nucleation rate, and consequently in a large final crystal size.

Additionally, energy dispersive X-ray spectroscopy (EDX) of the synthesized materials were performed. As shown in Table 1, nitrogen content increased with increasing BA content, which confirmed not only the presence of BA onto the surface of ZIF-8 crystals but also the possibility of tailoring the loading of surface amine moieties.

To determine the surface area, nitrogen adsorption/desorption isotherms of the synthesized materials were performed. They exhibited typical Type I isotherms (a representative example is depicted in Fig. S3), which are indicative of their microporous nature. The specific surface area, calculated using the BET equation, are shown in the Table 1. As it can be observed, high surface areas (> 1000 m²/g) were obtained from BA contents of 10 mmol or higher.

Furthermore, X-ray powder diffraction patterns, depicted in Fig. S4, were in good agreement with the simulated one, thus indicating that the current materials were successfully prepared. Also, FT-IR spectra of ZIF-8 materials were taken (Fig. S5). All the spectra display the typical bands of the ZIF-8 MOF [36, 40], corroborating their successful preparation. In addition, a small band close to 2900 cm⁻¹ appeared in the NH₂-ZIF-8-BA, which can be assigned to the aliphatic stretching C–H of BA.

306 Next, the synthesized MOFs were evaluated as SPE sorbents in order to study the 307 influence of modulation and choose the phase that provided the highest extraction 308 efficiency. As shown in Table 1, the NH₂-ZIF-8-BA 1 showed high recoveries jointly 309 with exposed surfaces having enough amino moieties for the selective capture of analytes. 310 Additionally, this material was superior to that synthesized without modulator, which 311 clearly demonstrated the important contribution of the amount of attached BA onto ZIF-312 8 crystals in the extraction efficiency of these pollutants. It is evident from the previous 313 discussion and experimental results that amine group of BA modulator plays a key role 314 in the crystal growth of MOF as well as in his subsequent retention properties. For this 315 purpose, other organic amines (ethyl and hexyl) with similar basicity (pKa values ranged 316 between 10.6-10.9) were studied as modulators using the same conditions of NH₂-ZIF-8-317 BA 1. The characterization of the resulting materials gave mean sizes of the crystals similar to those obtained with BA modulated material and surface area values ranged between 1150 and 1324 m2 g⁻¹. Both materials were tested as SPE sorbents, giving recoveries up to 90%. These results suggested that the alkyl chain has not a relevant effect, whereas the introduction of amine in the final framework is an important issue to improve the retention performance. At sight of these results, NH₂-ZIF-8-BA 1 was selected for further studies.

324

325 *3.3. Optimization of SPE protocol*

In order to obtain the maximal extraction performance, various parameters such as sample pH value, elution solvent composition and volume were investigated. During optimization of SPE conditions, an aqueous solution containing 250 μ g L⁻¹ of each benzomercaptan was employed as a test mixture.

330 Sample pH is an important variable that influences the extraction performance since it 331 affects the ZIF-8 charge surface as well as the analyte charge state. According to previous 332 reports, ZIF-8 material is charged positively in the pH range of 2-10 due to its imidazole 333 moieties [48, 49]. In any case, zeta potential measurements of the selected material were 334 performed at loading/elution conditions (Fig. S₆). Thus, the positive value of this 335 parameter at loading step (pH close to 7.0) implied a positive surface charge of MOF 336 nanoparticles, whereas that it reversed from positive to negative when eluting solution 337 was tested (pH > 12). These results were consistent with the above-mentioned studies. 338 On the other hand, the heterocyclic benzomercaptans may exist as thione and/or thiol 339 forms, and the thiol form can be further ionized in aqueous solutions (see pKa values in 340 Table S1). In this study, the effect of the sample pH value was evaluated from 5 to 9 (data 341 not shown), by considering aspects such as the structural integrity of this framework (pH 342 > 4) [50] as well as the usual pH range found in surface water systems (6.5 to 8.5). 343 Recoveries in the tested pH range were almost constant for each benzomercaptan. These 344 results can be explained as follows. On the one side, the existence of electrostatic 345 interactions can be present taking into account the surface charge of MOF and the 346 ionization state of benzomercaptans. Particularly, when the solution pH was below 6.0, 347 these compounds are predominantly as neutral (molecular) species, while the surface of 348 ZIF-8 remained positive, which made the electrostatic attraction between them become 349 weak. When solution pH increased from 6.0 to 8.5, the anionic form of benzomercaptans 350 begin to have greater significance due to progressive deprotonation of these molecules 351 (see pK_a values), and the electrostatic attraction between these solutes and ZIF-8 became

a main factor in the interaction mechanism. Nevertheless, the recovery values of target compounds on ZIF-8 along the tested pH range remained nearly constant. This fact suggested that the factor controlling the retention of probe compounds was not just electrostatic interaction, and there should be other modes of action that influence on the adsorption process (such as π -stacking and hydrogen bonding interactions). Therefore, no pH adjustment was required in sample solutions.

358 The selection of an appropriate eluting solvent is of major concern for the optimization 359 of the SPE process. To obtain the highest eluting efficiencies, several eluting solvents 360 were investigated (see Fig. 2, left part). Desorption solvents as pure MeOH gave very low 361 recovery values of analytes (< 8%). As can be observed, when MeOH-water mixtures 362 (50/50, v/v) were used, higher recoveries were obtained (up to 29%), which subsequently 363 increased up to around 79% when using mixtures of MeOH-water basified with 0.1 M 364 NaOH. This increment can be explained taking into account the surface charge of MOF 365 and the ionization state of benzomercaptans. As it was mentioned before, the point of zero 366 charge for ZIF-8 is approximately at pH 9.8 [48, 49]. Consequently, at pH above this 367 value, an electrostatic repulsion between the negatively charged ZIF-8 surfaces and the 368 benzomercaptan anions is produced, which favors the desorption of these analytes from 369 the sorbent. Consequently, the content of NaOH (from 0.1 to 0.5 M) in MeOH-water 370 mixtures was optimized. It was found that a content of 0.5 M NaOH gave recoveries 371 ranging between 97 and 108%. So, as in agreement with previous studies [50], the MOF 372 structure was stable at this pH, a 0.5 M NaOH concentration was selected for further 373 studies.

The volume of eluent can affect not only the elution efficiency, but also the concentration of the analytes in the eluent. Therefore, the effect of the solvent volume (100-1000 μ L) was also investigated (Fig. S7). As can be seen, recoveries of analytes higher than 87.3% (< 6.1% relative standard deviation, RSDs) were obtained for 250 μ L, and this value was selected for further experiments.

Another essential parameter in a SPE protocol, especially to monitor trace pollutants in environmental analysis, is the breakthrough volume. Thus, the effect of sample volume was evaluated using the NH₂-ZIF-8 as sorbent. Several volumes (in the range 1-100 mL) were loaded by keeping constant the total amount of each pollutant (500 ng) (see Fig. 2, right part). As can be seen, the extraction recoveries of all the benzomercaptans were higher than 80% up to 50 mL, however, higher volumes caused a diminution in the extraction efficiency. Consequently, considering the minimum elution volume found (250 μ L) and the maximum sample loading volume (25 mL) with average recoveries values

387 *ca.* 100%, a preconcentration factor of 100 was estimated.

- 388 Finally, the reusability of sorbent was also evaluated, and the obtained results indicated
- that the MOF could be reused at least 10 times without significant decrease of efficiency
- 390 extraction (between 87-104% with RSD below to 8%).
- 391
- 392 *3.4. Figures of merit and application to real samples*
- The developed SPE procedure using NH₂-ZIF-8-BA 1 as sorbent, combined with HPLC-393 394 UV detection, was validated in terms of linearity, sensitivity and precision under the previously optimized experimental conditions. The obtained results are summarized in 395 396 Table 2. As can be seen from this table, a good linearity range (r > 0.998) was observed in the chromatographic dynamic range of 7-2500 μ g L⁻¹ for all analytes. The limits of 397 398 detection (LOD) and quantification (LOQ) (after applying the extraction protocol) were 399 experimentally obtained as the concentration of the analyte that provided a signal-to-noise 400 ratio (S/N) of 3 and 10, respectively. Thus, the LODs for aqueous samples varied from 16 ng L^{-1} to 21 ng L^{-1} , whereas the LOQs were in the range 52 ng L^{-1} and 69 ng L^{-1} . In 401 the case of soil samples, the LOD and LOQ values ranged 0.4-0.5 μ g kg⁻¹ and 1.3-1.7 μ g 402 kg⁻¹, respectively. 403
- 404 The precision of the method (intra and inter-units), expressed as relative standard 405 deviation (RSD, %) was also determined from standard solutions at a concentration level 406 of 250 μ g L⁻¹ of each analyte and subjected to the SPE protocol. RSD values comprised 407 between 2.0 and 7.0% were found.
- 408 To study the applicability of the developed SPE-HPLC-UV method using NH₂-ZIF-8-BA 409 1 as sorbent, the developed method was applied to monitor the target analytes in 410 environmental complex matrices. The samples were analyzed in order to find any 411 potential presence of the analytes. The results (see Table 3) showed that none of the target 412 benzomercaptan was detected in the nonspiked real samples. Next, validation samples 413 were prepared using these samples fortified with the three target compounds at concentration level of 5-500 µg L⁻¹. As shown in Table 3, the recoveries of analytes were 414 415 satisfactory, ranging between 74 and 117%. In order to discard matrix effect in sewage 416 water, particularly for that one from Gandía (influent), standard addition calibration curves were done. Several spiked samples at different levels (25-100 µg L⁻¹) were 417 418 percolated through cartridges and the slopes obtained in the resulting calibration curves 419 were not statistical different from those found with the external calibration method

420 (confidence level of 95%) As representative examples, Fig. 3 (left) shows the 421 chromatograms of a water sample (from influent sewage water) spiked with the analytes 422 without and with SPE pretreatment. On the other hand, the right part of Fig. 3 illustrates 423 the eluted fraction corresponding to a blank soil sample and that spiked with the analytes. 424 As observed, an effective enrichment of analytes of interest in both matrices was 425 evidenced, which demonstrated the performance of the synthesized sorbent to be used as 426 preconcentration purposes in complex samples.

427

428 3.5. Comparison with other commercial sorbents and extraction methodologies

Next, a comparison in terms of extraction performance with one commercial sorbent
(silica gel) commonly used for the extraction of benzomercaptans [51, 52] was made.
This generic sorbent gave retention values below 50% (see Fig. S8), which underlines the
convenience of using our synthesized sorbent.

433 Finally, the developed material was compared with other extraction sorbents of target 434 analytes in environmental samples (see Table 4). Regarding aqueous samples analysis, 435 the recovery values obtained were quite similar to those found in most reported studies; 436 however, in certain works [23, 28] these values were lower than 65%. Regarding the 437 LODs, our values were better than those described by Parham et al. [24], and similar to 438 those reported in refs. [28, 30, 31] using a sophisticated and high-cost MS detector. Other 439 advantage found is the less amount (20 mg) of sorbent required compared with single-use 440 commercial SPE sorbents (commonly up to 150-500 mg) [28, 30, 31]. This amount was 441 also similar to a nanomaterial-based method using CuNPs [23, 24] probably due to 442 surface area-to-volume ratio; however, its reusability was quite lower than that found in 443 the present study.

Likewise, in soil monitoring, the present method gave higher recoveries than those described in literature [53, 54]. Concerning LODs, our protocol provided comparable LOD values [53], or higher using MS detection [54]. In any case, this drawback does not reduce the good features of our protocol such as the easy and cheap preparation of the sorbent combined with the accessible equipment required, make this protocol a feasible method to monitor benzomercaptans and their derivatives in environmental samples.

450

451 **4. Conclusions**

452 In this research, n-butylamine modulated ZIF-8 nanocrystals were successfully 453 synthesized, characterized and applied as SPE sorbent to the extraction of

454 benzomercaptans in environmental matrices. Prior to the SPE optimization, the addition 455 of BA (at several molar ratios) as modulator to the reaction mixture was done to evaluate 456 its impact on the crystal growth and adsorption capacity of the resulting MOFs. As a result 457 of this study, NH₂-ZIF-8 containing 10 mmol of BA as modulator was the most 458 appropriate sorbent. Then, several experimental parameters of the SPE protocol (such as 459 sample pH, desorption solvent composition, among others) were investigated in detail 460 using this MOF as SPE phase. The highly selective and efficient retention of 461 benzomercaptans on this MOF take advantages of multiple interactions between these 462 targets and the framework.

463 Moreover, the fruitful combination of NH₂-ZIF-8-BA 1 followed by HPLC-UV analysis 464 led to high extraction recovery values, low LODs, excellent enrichment factors and 465 satisfactory reusability. All these figures of merits prove that the present protocol 466 constitutes a simple, cost-effective, and appropriate methodology for extraction and 467 preconcentration of these pollutants in complex environmental samples. As far as we 468 know, this study reports the first study of influence of modulator content on the 469 morphology and extraction performance of organic pollutants and the subsequent 470 application of the best modulated ZIF-8 for the extraction of benzomercaptans and 471 demonstrate that it could be a promising SPE sorbent for further applications.

472

473 Acknowledgements

474 The authors would like to gratefully acknowledge the financial support received from the 475 project RTI2018-095536-B-I00 (Ministry of Science, Innovation and Universities, 476 Spain). Spanish Agencia Estatal de Investigación (AEI-Spain) and the European Funds 477 for Regional Development (FEDER-European Union) are also gratefully acknowledged 478 for financial support through Project CTQ2016-77155-R (AEI/FEDER, UE).). H. M. P-479 C. and N.C. thank the support from the Spanish Ministry of Science, Innovation and 480 Universities for FPU pre-doctoral fellowships. The authors extend their appreciation to 481 MCIU for granting the Spanish Network of Excellence in Sample preparation (RED2018-482 102522-T). This article is based upon work from the Sample Preparation Task Force and 483 Network, supported by the Division of Analytical Chemistry of the European Chemical 484 Society.

485

486 **Declaration of competing interest:** None.

487 References

- 488 [1] M. Safaei, M. M. Foroughi, N. Ebrahimpoor, S. Jahani, A. Omidi, M. Khatami. A
- 489 review on metal-organic frameworks: synthesis and applications. Trends Anal. Chem. 490 118 (2019) 401-425.
- [2] C. C. Hou, Q. Xu. Metal-organic frameworks for energy. Adv. Energy Mater. 9 491 492 (2019) 1801307.
- 493 [3] L. Joseph, B. M. Jun, M. Jang, C. M. Park, J. C. M.Senmache, A. J. H. Maldonado,
- 494 A. Heyden, M. Yu, Y. Yoon. Removal of contaminants of emerging concern by metal-
- 495 organic framework nanoadsorbents: A review. Chem. Eng. J. 369 (2019) 928-946.
- 496 [4] P. Kumar, E. Vejerano, A. Khan, G. Lisak, J. H. Ahn, K. H. Kim. Metal organic
- 497 frameworks (MOFs): Current trends and challenges in control and management of air 498
- quality. Korean J. Chem. Eng., 36 (2019) 1839-1853.
- 499 [5] A. Shahat, H. M. A. Hassan, H. M. E. Azzazy. Optical metal-organic framework
- 500 sensor for selective discrimination of some toxic metal ions in water. Anal. Chim. Acta. 501 793 (2013) 90-98.
- 502 [6] P. R. Bautista, I. P. Fernández, J. Pasán, V. Pino. Are metal-organic frameworks able
- 503 to provide a new generation of solid-phase microextraction coatings? - A review. Anal. 504 Chim. Acta 939 (2016) 26-41.
- 505 [7] X. Gong, Y. Wang, T. Kuang. ZIF-8-based membranes for carbon dioxide capture 506 and separation. ACS Sustainable Chem. Eng. 5 (2017) 11204-11214.
- 507 [8] X. L. Xu, H. Wang, J.B. Liu, H. Yan. The applications of zeolitic imidazolate 508 framework-8 in electrical energy storage devices: a review. J. Mater. Sci. Mater Electron 509 28 (2017) 7532-7543.
- 510 [9] J. Troyano, A. C. Sánchez, C. Avci, I. Imaz, D. Maspoch. Colloidal metal-organic
- 511 framework particles: the pioneering case of ZIF-8. Chem. Soc. Rev. 48 (2019) 5534-512 5546.
- [10] H. Zhang, D. Liu, Y. Yao, B. Zhang, Y. Lin. Stability of ZIF-8 membranes and 513 514 crystalline powders in water at room temperature. J. Membr. Sci. 485 (2015) 103-111.
- 515 [11] H. Tanaka, S. Ohsaki, S. Hiraide, D. Yamamoto, S. Watanabe, M. T. Miyahara,
- 516 Adsorption-Induced structural transition of ZIF-8: a combined experimental and
- 517 simulation study. J. Phys. Chem. C. 118 (2014) 8445-8454.
- 518 [12] Y. Pan, D. Heryadi, F. Zhou, L. Zhao, G. Lestari, H. Su, Z. Lai, Tuning the crystal
- 519 morphology and size of zeolitic imidazolate framework-8 in aqueous solution by
- 520 surfactants, Cryst. Eng. Comm. 13 (2011) 6937-6940.

- 521 [16] H. Lan, T. Rönkkö, J. Parshintsev, K. Hartonen, N. Gan, M. Sakeye, J. Sarfraz, M.
- 522 L. Riekkola. Modified zeolitic imidazolate framework-8 as solid-phase microextraction.
- 523 Arrow coating for sampling of amines in wastewater and food samples followed by gas

524 chromatography-mass spectrometry. J. Chromatog. A. 1486 (2017) 76-85.

- [14] J. Kong, F. Zhu, W. Huang, H. He, J. Hu, C. Sun, Q. Xian, S. Yang. Sol–gel based
 metal-organic framework zeolite imidazolate framework-8 fibers for solid-phase
 microextraction of nitro polycyclic aromatic hydrocarbons and polycyclic aromatic
 hydrocarbons in water samples. J. Chromatogr. A. 1603 (2019) 92-101.
- 529 [15] F. Maya, M. Ghani. Ordered macro/micro-porous metal-organic framework of type
- 530 ZIF-8 in a steel fiber as a sorbent for solid-phase microextraction of BTEX. Microchim.
 531 Acta. 186 (2019) 425.
- 532 [16] J. Pang, Y. Liao, X. Huang, Z. Ye, D. Yuan. Metal-organic framework-monolith 533 composite-based in-tube solid phase microextraction on-line coupled to high-534 performance liquid chromatography-fluorescence detection for the highly sensitive 535 monitoring of fluoroquinolones in water and food samples. Talanta. 199 (2019) 499-506. 536 [17] M. Rio, C. P. Cabello, V. Gonzalez, F. Maya, J. B. Parra, V. Cerdà, G. T. Palomino. 537 Metal oxide assisted preparation of core-shell beads with dense metal-organic 538 framework coatings for the enhanced extraction of organic pollutants. Chem. Eur. J. 22 539 (2016) 11770-11777.
- [18] X. Liu, Z. Sun, G. Chen, W. Zhang, Y. Cai, R. Kong, X. Wang, Y. Suo, J. You.
 Determination of phthalate esters in environmental water by magnetic zeolitic
 imidazolate framework-8 solid-phase extraction coupled with high-performance liquid
 chromatography. J. Chromatogr. A. 1409 (2015) 46-52.
- 544 [19] D. Ge, H. K. Lee. Zeolite imidazolate frameworks 8 as sorbent and its application to 545 sonication-assisted emulsification microextraction combined with vortex-assisted porous 546 membrane-protected micro-solid-phase extraction for fast analysis of acidic drugs in
- environmental water samples. J. Chromatogr. A. 1257 (2012) 19-24.
- 548 [20] D. Ge, H. K. Lee. Sonication-assisted emulsification microextraction combined with
- vortex-assisted porous membrane-protected micro-solid-phase extraction using mixed
 zeolitic imidazolate frameworks 8 as sorbent. J. Chromatogr. A. 1263 (2012) 1-6.
- 551 [21] Y. Wang, S. Jin, Q. Wang, G. Lu, J. Jiang, D. Zhu. Zeolitic imidazolate framework-
- 552 8 as sorbent of micro-solid-phase extraction to determine estrogens in environmental
- 553 water samples. J. Chromatogr. A. 1291 (2013) 27-32.

- [22] J. Cravillon, R. Nayuk, S. Springer, A. Feldhoff, K. Huber, M. Wiebcke. Controlling
 zeolitic imidazolate framework nano- and microcrystal formation: insight into crystal
- 556 growth by time-resolved in situ static light scattering. Chem. Mater. 23 (2011) 2130-2141.
- [23] H. Parham, F. Khoshnam. Highly efficient and simultaneous removal of 2mercaptobenzothiazole and 2-mercaptobenzoxazole from water samples by copper oxide
 nanoparticles. J. Chem. Technol. Biotechnol. 88 (2013) 1736-1743.
- 560 [24] H. Parham, F. Khoshnam. Solid phase extraction-preconcentration and high
 561 performance liquid chromatographic determination of 2-mercapto-(benzothiazole,
 562 benzoxazole and benzimidazole) using copper oxide nanoparticles. Talanta. 114 (2013)
 563 90–94.
- 564 [25] T. Sorahan. Cancer risks in chemical production workers exposed to 2-565 mercaptobenzothiazole. Occup. Environ. Med. 66 (2009) 269-273.
- 566 [26] M. H. Whittaker, A. M. Gebhart, T. C. Miller, F. Hammer. Human health risk
 567 assessment of 2-mercaptobenzothiazole in drinking water. Toxicol. Ind. Health. 20 (2004)
 568 149-163.
- 569 [27] NSF International Standard/American National Standard for drinking water
 570 additives. Drinking water treatment chemicals health effect. NSF International, USA.
 571 March 9, 2016 (NSI/ANSI 60-2016).
- 572 [28] A. Kloepfer, M. Jekel, T. Reemtsma. Determination of benzothiazoles from complex
 573 aqueous samples by liquid chromatography–mass spectrometry following solid-phase
 574 extraction. J. Chromatogr. A. 1058 (2004) 81-88.
- 575 [29] E. Fries. Determination of benzothiazole in untreated wastewater using polar-phase
 576 stir bar sorptive extraction and gas chromatography–mass spectrometry. Anal. Chim.
 577 Acta. 689 (2011) 65-68.
- 578 [30] I. Carpinteiro, B. Abuin, M. Ramil, I. Rodríguez, R. Cela. Simultaneous 579 determination of benzotriazole and benzothiazole derivatives in aqueous matrices by 580 mixed-mode solid-phase extraction followed by liquid chromatography-tandem mass 581 spectrometry. Anal Bioanal Chem. 402 (2012) 2471-2478.
- [31] C. H. Loi, F. Busetti, K. L. Linge, C. A. Joll. Development of a solid-phase extraction
 liquid chromatography tandem mass spectrometry method for benzotriazoles and
- benzothiazoles in wastewater and recycled water. J. Chromatogr. A. 1299 (2013) 48-57.
- 585 [32] Z. Xiang, C. Fang, S. Leng, D. Cao. An amino group functionalized metal–organic
- 586 framework as a luminescent probe for highly selective sensing of Fe 3+ ions. J. Mater.
- 587 Chem. A. 2 (2014) 7662-7665.

- 588 [33] C. Gecgel, U. B. Simsek, B. Gozmen, M. Turabik. Comparison of MIL-101(Fe) and
- amine-functionalized MIL-101(Fe) as photocatalysts for the removal of imidacloprid in
 aqueous solution. J. Iran. Chem. Soc. 16 (2019) 1735–1748.
- 591 [34] C. Li, L. Zhu, W. Yang, X. He, S. Zhao, X. Zhang, W. Tang, J. Wang, T. Yue, Z. Li.
- 592 Amino-functionalized Al–MOF for fluorescent detection of tetracyclines in milk. J. Agr.
- 593 Food Chem. 67 (2019) 1277-1283.
- 594 [35] Y. Yuan, X. Zheng, H. Lin, Y. Li, M. Yang, X. Liu, C. Deng, Z. Fan. Development
- 595 of a hydrophilic magnetic amino-functionalized metal-organic framework for the highly
- efficient enrichment of trace bisphenols in river water samples. Talanta, 211 (2020)120713.
- 598 [36] F. Maya, C. P. Cabello, S. Clavijo, J. M. Estela, V. Cerdà, G. T. Palomino. Zeolitic
- 599 imidazolate framework dispersions for the fast and highly efficient extraction of organic
- 600 micropollutants. RSC Adv. 5 (2015) 28203-28210.
- 601 [37] J. Cravillon, S. Münzer, S. J. Lohmeier, A. Feldhoff, K. Huber, M. Wiebcke. Rapid
- room-temperature synthesis and characterization of nanocrystals of a prototypical zeolitic
- 603 imidazolate framework. Chem. Mater. 21 (2009) 1410-1412.
- 604 [38] ISO (2005). Soil Quality–Determination of pH (ISO 10390: 2005). ISO (1994). Soil
- 605 Quality—Determination of the Specific Electrical Conductivity. ISO (2003) Soil quality
- 606 -- Determination of soil water content as a volume fraction on the basis of known dry
- 607 bulk density -- Gravimetric method (ISO 16586:2003).
- 608 [39] Z. Zhang, N. Ren, Y. F. Li, T. Kunisue, D. Gao, K. Kannan. Determination of
- 609 benzotriazole and benzophenone UV filters in sediment and sewage sludge. Environ. Sci.
- 610 Technol. 45 (2011) 3909-3916.
- 611 [40] C. Y. Sun, C. Qin, X. L. Wang, G. S. Yang, K. Z. Shao, Y. Q. Lan, Z. M. Su, P.
- 612 Huang, C. G. Wang, E. B. Wang. Zeolitic imidazolate framework-8 as efficient pH-
- 613 sensitive drug delivery vehicle. Dalton Trans. 23 (2012) 6906-6909.
- 614 [41] F. Yan, Z. Y. Liu, J. L. Chen, X. Y. Sun, X. J. Li, M. X. Su, B. Li, B. Di. Nanoscale
- 615 zeolitic imidazolate framework-8 as a selective adsorbent for theophylline over caffeine
- 616 and diprophylline. RSC Adv. 4 (2014) 33047-33054.
- 617 [42] J.C. Ma, D.A. Dougherty, The cation-π interaction, Chem. Rev. 97 (1997) 1303–
 618 1324.
- 619 [43] I. Ahmed, N.A. Khan, J.W. Yoon, J.S. Chang, S.H. Jhung, Protonated MIL-125-
- 620 NH2: Remarkable Adsorbent for the Removal of Quinoline and Indole from Liquid Fuel,
- 621 ACS Appl. Mater. Interfaces. 9 (2017) 20938–20946. [44] A. Schaate, P. Roy, A. Godt,

- 622 J. Lippke, F. Waltz, M. Wiebcke, P. Behrens. Modulated synthesis of Zr-based metal-
- 623 organic frameworks: from nano to single crystals. Chem. Eur. J. 17 (2011) 6643-6651
- 624 [45] Q. Liu, L. N. Jin, W. Y. Sun. Coordination modulation induced and ultrasonic-
- 625 assisted synthesis of size-controlled microporous metal-imidazolate framework crystals
- 626 with enhanced adsorption performance. Cryst. Eng. Comm. 15 (2013) 8250-8254.
- 627 [46] G. Wißmann, A. Schaate, S. Lilienthal, I. Bremer, A. M. Schneider, P. Behrens.
- Modulated synthesis of Zr-fumarate MOF. Micropor. Mesopor. Mat. 152 (2012) 64-70.
- 629 [47] S. Diring, S. Furukawa, Y. Takashima, T. Tsuruoka, S. Kitagawa. Controlled
- multiscale synthesis of porous coordination polymer in nano/micro regimes. Chem.
 Mater. 22 (2010) 4531-4538.
- 632 [48] Y. N. Wu, M. Zhou, B. Zhang, B. Wu, J. Li, J. Qiao, X. Guan, F. Li. Amino acid
- assisted templating synthesis of hierarchical zeolitic imidazolate framework-8 for
 efficient arsenate removal. Nanoscale. 6 (2014) 1105-1112.
- 635 [49] J. Li, Y. N. Wu, Z. Li, B. Zhang, M. Zhu, X. Hu, Y. Zhang, F. Li. Zeolitic imidazolate
- framework-8 with high efficiency in trace arsenate adsorption and removal from water.J. Phys. Chem. C. 118 (2014) 27382-27387.
- [50] C. S. Wu, Z. H. Xiong, C. Li, J. M. Zhang. Zeolitic imidazolate metal organic
 framework ZIF-8 with ultra-high adsorption capacity bound tetracycline in aqueous
 solution. RSC Adv. 5 (2015) 82127-82137.
- 641 [51] Y. Liu, Q. H. Zou, M. X. Xie, J. Han. A novel approach for simultaneous
- determination of 2-mercaptobenzimidazole and derivatives of 2-thiouracil in animal
 tissue by gas chromatography/mass spectrometry. Rapid Commun. Mass Spectrom. 21
 (2007) 1504–1510.
- [52] M. Lõhmus, K. Kallaste, B. Le Bizec. Determination of thyreostats in urine and
 thyroid gland by ultra high performance liquid chromatography tandem mass
 spectrometry. J. Chromatogr. A 1216 (2009) 8080-8089.
- 648 [53] A. G. Asimakopoulos, A. Ajibola, K. Kannan, N. S. Thomaidis. Occurrence and
- 649 removal efficiencies of benzotriazoles and benzothiazoles in a wastewater treatment plant
- 650 in Greece. Sci. Total Environ. 452-453 (2013) 163-171.
- 651 [54] J. Zhang, X. Zhang, L. Wu, T. Wang, J. Zhao, Y. Zhang, Z. Men, H. Mao.
- 652 Occurrence of benzothiazole and its derivates in tire wear, road dust, and roadside soil.
- 653 Chemosphere 201 (2018) 310-317.
- 654

Table 1. Influence on different BA content modulator of textural parameters ofsynthesized MOFs and their extraction performances.

| MOF | Modulator | Particle size (nm) | Surface area (m ² g ⁻¹) | N amount (atomic, %) | Recoveries ^a (%) | |
|-----------------------------|-------------|--------------------|---|-------------------------|-----------------------------|--|
| ZIF-8 | None | 89 ± 6 | 1456 | 45.3 | 60-63 | |
| NH ₂ -ZIF-8-BA 1 | BA, 10 mmol | 61 ± 2 | 1365 | 52.6 | 94-102 | |
| NH ₂ -ZIF-8-BA 2 | BA, 5 mmol | 47 ± 5 | 856 | 47.7 | 64-77 | |
| NH ₂ -ZIF-8-BA 3 | BA, 20 mmol | 72 ±7 | 1213 | 56.3 | 99-102 | |

^aExtraction conditions: sample concentration, 500 µg L⁻¹; volume, 5 mL; eluting solvent, 0.5 mL of 50:50
MeOH:H₂O (v/v) with 0.5 M NaOH.

Table 2. Figures of merit of the developed material used as SPE sorbent in the extraction

and analysis of benzomercaptans.

| Analyte | Calibration range (µg L ⁻¹) | Within-device recovery (%) ± RSD (n=3) | Between-device recovery (%) ± RSD (n=3) | LOD ^a (µg L ⁻¹) | LOQ ^a (µg L ⁻¹) | |
|---------|--|--|---|---|---|--|
| MBO | 5 - 2500 | 109 ± 2 | 107 ± 2 | 1.6 | 5 | |
| MBT | 7 - 5000 | 95 ± 7 | 101 ± 1 | 2 | 7 | |
| NMBT | 7 - 5000 | 100 ± 7 | 95 ± 1 | 2 | 7 | |

^aInstrumental values (without applying the extraction protocol)

- **Table 3.** Recovery study of benzomercaptans in spiked environmental samples analyzed
- following the recommended SPE protocol. Recovery (%) \pm SD (n=3)^a.

| | Environmental samples | | | | | | | | | | | |
|---------|------------------------------------|----------|------------------------------------|-----------|-----------|-------------------------------------|----------------|-------------|--------------------|-------------|-------------------|------------|
| Analyte | Gandia sewage water (influent) | | Paterna sewage water (effluent) | | Tap water | | Soil 1 (Sueca) | | Soil 2 (Burjassot) | | Soil 3 (Valencia) | |
| - | Spiked level (µg L ⁻¹) | | | | | Spiked level (µg kg ⁻¹) | | | | | | |
| | - | 5 | - | 5 | - | 5 | - | 500 | - | 500 | - | 500 |
| MBO | < LOD | 76 ± 2 | < LOD | 94 ± 2 | < LOD | 117 ± 9 | < LOD | 109 ± 2 | < LOD | 100 ± 7 | < LOD | 102 ± 3 |
| MBT | < LOD | 76 ± 5 | < LOD | 82 ± 2 | < LOD | 81 ± 4 | < LOD | 103 ± 1 | < LOD | 107 ± 1 | < LOD | 98 ± 6 |
| NMBT | < LOD | 74 ± 5 | < LOD | 75 ± 6 | < LOD | 74 ± 8 | < LOD | 93 ± 1 | < LOD | 84 ± 10 | < LOD | 82 ± 3 |

^aThe recovery values were calculated by dividing the concentration found by theconcentration added (calibration curves).

676 **Table 4.** Comparison with other reported methods for benzomercaptans extraction and677 analysis.

| Analytes | Material (sorbent amount) | Method | Sample matrix | Recoveries (%) | LOD (ng L ⁻¹ / ng kg ⁻¹) | Reusability | EF ^a | Reference |
|-------------------------------|--|----------------------|------------------------------|--------------------|--|-------------|----------------------------------|-----------|
| MBT, MBO | Copper oxide nanoparticles (14 mg) | SPE / HPLC-DAD | Industrial water samples | 56-93 - 3 | | | [23] | |
| MBT, MBO, MBI | Copper oxide nanoparticles (14 mg) | SPE / HPLC-DAD | Environmental water sample | 96-104 | 1900-2700 | 3 | 200 | [24] |
| MBT | Oasis HLB cartridge (200 mg) | SPE / HPLC-MS | Municipal wastewater samples | samples 60-70 16.7 | | - | 7-14 | [28] |
| MBT | Oasis MAX (150 mg) | SPE / HPLC-MS | Environmental water samples | 80-118 | 1-4 | - | 20-100 | [30] |
| MBT | Oasis HLB (500 mg) | SPE / HPLC-MS | Water samples | 70-85 | 7 | - | 21-83 | [31] |
| BT, HBT, MTBT, ABT | Strata TM -X cartridges (200 mg) | SPE / HPLC- MS/MS | Dewatered sewage sludge | 50-116 | 40 - 13000 | - | - | [51] |
| MBT, HBT, BT, ABT, MTBT | Poly-Sery HLB cartridge (60 mg) | SPE / UPLC- MS/MS | Road dust / Roadside soil | 56-105 | 20-120 | - | - | [52] |
| MBO, MBT, NMBT | NH2-ZIF-8-BA 1 (20 mg) | SPE / HPLC-UV | Environmental samples | 74-117 | 16-21 / 400-500 | 10 | 4 ^b ;100 ^c | This Work |

678

^aThe EF from the overall SPE protocol was calculated using the ratio between the final (eluent)

680 extract and the maximum sample volume load (for aqueous samples) or the amount of solid

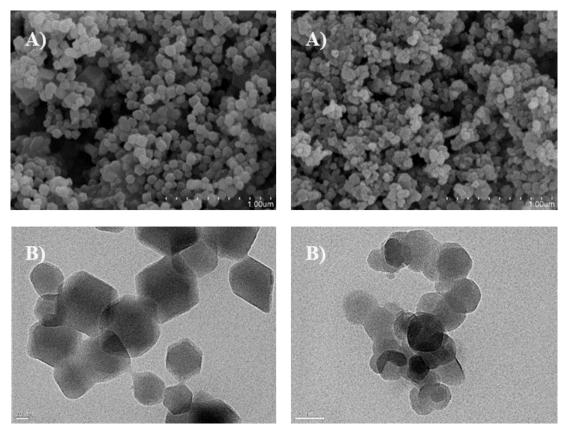
681 weighed (for soil samples); ^bsoil samples; ^caqueous samples

682

683 EF: enrichment factor. MBT: 2-mercaptobenzothiazole, MBO: 2-mercaptobenzoxazole, MBI: 2-

684 mercaptobenzimidazole, NMBT: 2-mercapto-6-nitrobenzothiazole, HBT: 2-

685 hidroxybenzothiazole, MTBT: 2-methyl-thiobenzothiazole, ABT: 2-aminobenzothiazole.



688 Figure 1. SEM (A) and TEM (B) micrographs of ZIF-8 (left part) and NH₂-ZIF-8-BA 1

- 689 (right part) materials.
- 690

687

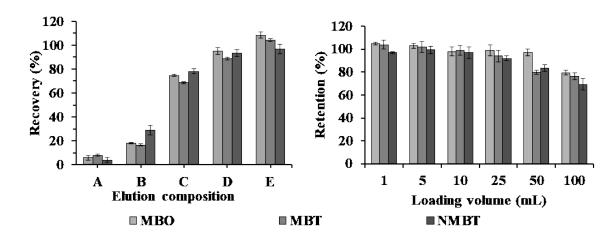
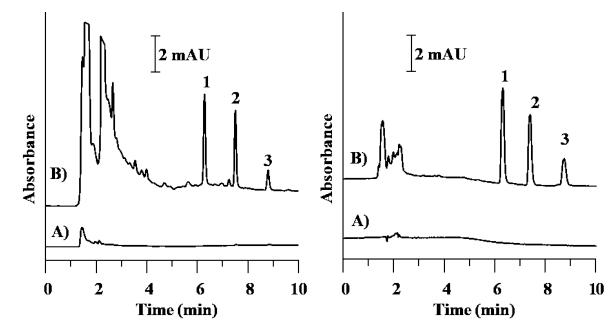




Figure 2. Effect of elution solvent composition on the recovery values of analytes using NH₂-ZIF-8 as SPE sorbent (left part),eluent compositions: A) MeOH, B) 50:50 MeOH:H₂O (v/v), C) 50:50 MeOH:H₂O (v/v) with 0.1 M NaOH, D) 50:50 MeOH:H₂O (v/v) with 0.2 M NaOH and E) 50:50 MeOH:H₂O (v/v) with 0.5 M NaOH; effect of loading sample volume on the extraction efficiency of benzomercaptans (right part). Error bar = SD (n = 3).





699

Figure 3. HPLC-UV chromatograms of sewage water (Gandía's plant) (left part) and extracts of soil samples (right part) spiked with 5 μ g L⁻¹ of each benzomercaptan, respectively, without applying (A) and applying the proposed SPE protocol (B). The right part shows an extract of soil sample without (A) and with spiking of analytes at 500 μ g kg⁻¹ (B) subjected to the SPE treatment. Chromatographic conditions are given in Experimental Section and Supplementary Information. Peak identification: 1, MBO; 2, MBT; 3, NMBT.