

INTRODUCTION OF A PRACTICAL LESSON FOR THE EVALUATION OF BIOACTIVE QUALITY IN PLANT MATERIALS ADDRESSED TO STUDENTS IN PLANT BREEDING

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Abstract

According to the new market trends, one of the goals in many modern breeding programmes is the development of varieties with enhanced bioactive properties. Despite the importance of evaluating the content of specific molecules, the analysis of the broad antioxidant capacity is often suitable. For instance, if the goal is to evaluate large quantity of materials, or even to compare different agronomical practices to obtain the highest antioxidant properties. Thus, studies related to plant breeding should include subjects related to the knowledge of analytical methods for the evaluation of the bioactive properties in the materials tested. Theoretical lessons establish the basis for understanding the importance of bioactive properties and the influence of stresses in their content. Nevertheless, practical sessions complement this knowledge and improve the competence of future professionals, providing them a tool for the evaluation of bioactive quality in developing breeding materials. We consider therefore that this competence should be acquired during the Master degree in Plant Breeding offered by our institute. Thus, we propose to introduce a practical lesson in the mandatory subject "Instrumental Techniques" aimed to the comparison of two analytical methods for the analysis of bioactive properties. As plant material, we will use water celery, an herbaceous species with great antioxidant properties. The methodology will include the analysis of the antioxidant activity with the DPPH method and the content in total phenolics, both spectrophotometric methods but measuring different aspects of the bioactive quality. The lesson is divided in three sessions of three hours each, in the course of which students will perform all steps for these analyses, including sample preparation, extraction of bioactive molecules, preparation of the calibration curve, and analysis of results. This practical lesson provides students with the competence to evaluate the antioxidant activity of high number of materials using quick and simple spectrophotometric methodologies, and to deal with real problems such as the need of dilution for samples with very high content of bioactive molecules. Moreover, the practical lesson provides the guidelines to adapt the protocols for different materials and consequently to many breeding programmes, and can be also adapted for food technology students.

Keywords: Antioxidants, Plant Science studies, specific competence, spectrophotometry, total phenolics.

1 INTRODUCTION

Master studies are designed to provide advanced, specialized training of post-graduate students [1]. In Plant Science studies, good knowledge of techniques addressed to analyze nutritional properties of materials is usually needed. In the particular case of Plant Breeding studies, this knowledge can become essential. Traditional breeding has focused mainly on improving the yield of crops [2]. However, nowadays plant breeding programmes also consider other aspects of quality that are of interest for consumers, such as organoleptic and nutritional quality [e.g., 3], together with the introduction of tolerance in crops to pests and illness [2].

Due to the importance of nutritional quality in many breeding programmes, we consider essential that our Plant Breeding master provides students with the competence of analyzing nutritional related traits. It is of particular importance the study of molecules with antioxidant properties. An antioxidant is a molecule able to neutralize other molecules with free radicals, such as reactive oxygen species (ROS). ROS, when are present in excess in the human body, can cause molecular and cellular disorders deriving in several diseases [4]. Within the antioxidants, phenolic compounds deserve special attention. Phenolic compounds include a large number of molecules present in plants as secondary metabolites. From those, flavonoids and phenolic acids, commonly found in fruits and

vegetables, have demonstrated to possess high antioxidant properties [5]. Thus, increasing the content in phenolic compounds and other antioxidant molecules in fruits and vegetables can derive in a better health state of the consumers.

Despite the interest in evaluating the content in specific molecules, analyses of broad phenolic content and antioxidant capacity are often suitable. Measurements of antioxidant capacity and total content in phenolic molecules can be easily performed by using spectrophotometric methodologies [6, 7]. These methodologies allow a quick evaluation of a large quantity of materials in a reduced time. Thus, they can be used in breeding programmes aimed to select genotypes that present the highest antioxidant properties. Furthermore, these methodologies can be also used for establishing proper growing conditions that enhance the antioxidant values (e.g., organic vs. conventional), or for selecting specific genotype x environment interactions [8]. However, the application of these methodologies, although broadly described, involves in many cases problems related to the material processing, extraction and analysis validity. Since these situations are not usually explained in the protocols posted, the study of these situations in practical lessons of the related masters is highly desirable.

Our hypothesis is that the introduction of this practical lesson may contribute to an improved learning process for students of the Master degree in Plant Breeding offered by our Institute. Students should be able to put into practice the theoretical knowledge acquired for enhancing learning [9]. Moreover, we consider that practical lessons are good teaching tools for helping in the complete understanding and fixation of the concepts given in the theoretical lessons. In particular, we think that the development of this practical lesson will help students to understand the concept of antioxidant and phenolic molecules, and the difference and correlation between those concepts. In addition, students will obtain the competence of performing real studies focused on the comparison of different antioxidant properties among materials. Finally, encountering with real problems during the practical lesson will teach students how to solve them, thus improving the learning process [10], and also preparing them for their future careers.

2 THE PRACTICAL SESSION

The "Evaluation of bioactive quality in plant materials" practical lesson requires nine hours to be completed. Thus, the lesson will consist in three sessions of three hours each, taking place in consecutive days. At the beginning of the lesson, the instructor gives an outline with the steps that are being performed during the lesson, including the protocol of the different analytical methods, to each student (Fig. 1).

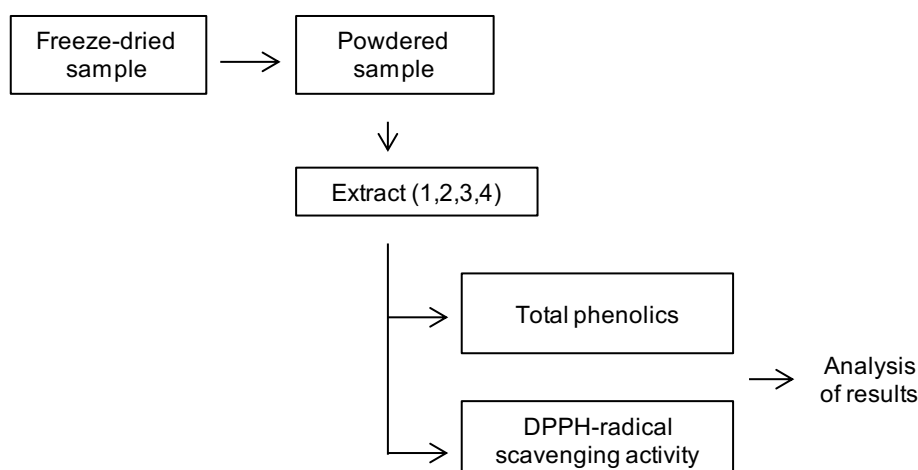


Figure 1. Scheme of the practical lesson.

The first day is aimed to the preparation of solutions and extraction of the antioxidant components. One freeze-dried sample of water celery (*Apium nodiflorum* (L.) Lag.) is provided to each student. This wild vegetable has been chosen due to the great antioxidant properties that the species presents, together with the high content in phenolic compounds in the leaves [11, 12]. Students are first asked to prepare properly the solutions needed for the lesson, considering the volume needed for all the analyses that will be performed in the class, and the different calibration curves that will be used (Table 1). Then, each student follows an extraction protocol according to the instructions of the

teacher (Table 1), considering that there are four possibilities and each extraction protocol must be repeated by, at least, three students. Students are asked to take notes about the exact quantity of material weighted. At the end of this first session, the solutions are stored in adequate conditions according to the type (it is, room temperature, 4°C or -20°C).

Table 1. Example of the solutions and extraction protocol that students are asked to follow.

Solutions needed during the lesson	
Extraction solution 1	Acetone: acetic acid (70:0.5% v/v) in distilled water
Extraction solution 2	Acetone: methanol (35:25% v/v) in distilled water
Extraction solution 3	Methanol 70% (v/v) in distilled water
Extraction solution 4	Distilled water
Diluted Folin-Ciocalteu reagent	Folin-Ciocalteu reagent 10% (v/v) in distilled water, freshly prepared the day of use
DPPH reagent	DPPH reagent 0.025g/L , freshly prepared the day of use
Sodium carbonate solution	Sodium carbonate 60g/L in distilled water
Extraction protocol	
1.	Grind the independent subsample with a pestle.
2.	Weight 0.100 g of the subsample.
3.	Extract with 5 ml of the extraction solution for 1 hour, in continuous stirring.
4.	Centrifuge at 3,500 rpm for 3 min.
5.	Store 1 ml of the supernatant at -20°C.

The second session is used for analyzing the content in total phenolics of the extract by means of the Folin-Ciocalteu methodology [7]. First, one aliquot of the diluted Folin-Ciocalteu reagent is prepared for all the class (Table 1). Students are then asked to take the extracts and follow the protocol as described in the outline, in triplicate, together with the one calibration curve for all the class (Table 2).

Table 2. Example of the protocols that students are asked to follow.

Total phenolics protocol	
1.	Add 500 µl of the diluted Folin-Ciocalteu reagent to 65 µl of the extract.
2.	Incubate for 5 min at room temperature and darkness.
3.	Add 500 µl of the sodium carbonate solution.
4.	Incubate for 60 min at room temperature and darkness.
5.	Transfer 200 µl to a 96-well microplate.
6.	Measure the absorbance in a spectrophotometer at 750 nm.
7.	Repeat the measurement each 5 min until the reaction remains stable.
DPPH-radical scavenging activity protocol	
1.	Add 3.9 ml of the DPPH reagent to 100 µl of the extract.
2.	Incubate for 10 min at room temperature and darkness.
3.	Transfer 200 µl to a 96-well microplate.
4.	Measure the absorbance in a spectrophotometer at 515 nm.
5.	Repeat the measurement each 2 min until the loss of absorbance remains stable.

One 96-well microplate is used for all the class, in which the reactions corresponding to the extracts and the calibration curve are included (Fig. 2). During the measurements with the spectrophotometer, students are asked to take specific notes about the time and the corresponding absorbance.

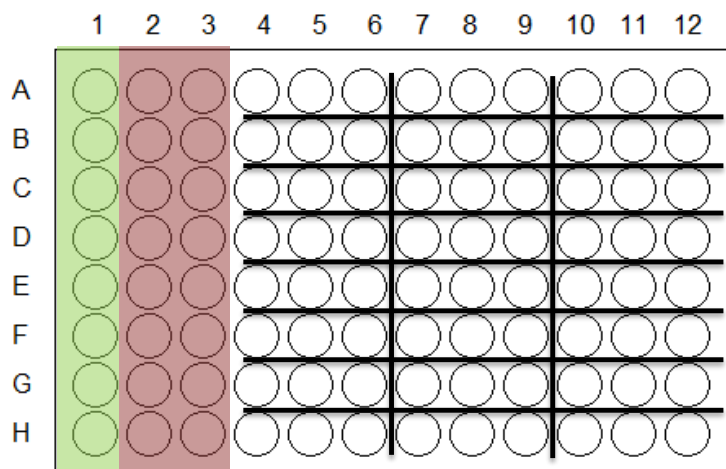


Figure 2. Example of a 96-well microplate filled to read the absorbance. Column 1 is destined to the blank; columns 2 and 3 are destined to the two replications of the calibration curve; and the remaining wells are used for the students, considering that each student fills three consecutive wells with the three replicates performed.

The third session of this practical lesson is used for performing the DPPH-radical scavenging activity protocol [6] and analyzing the results obtained in sessions 2 and 3. First, an aliquot of the DPPH reagent is prepared for all the class (Table 1). Then, students are asked to follow the protocol given in the outline (Table 2), and to fill the 96-well microplate as indicated in Fig. 2. During the measurements, they need to note the loss of absorbance at the corresponding times in which the microplate is read.

The last part of the third session corresponds to the analysis and discussion of results. Each student should be able to give the equation of the calibration curves corresponding to the Folin-Ciocalteu and DPP-radical scavenging activity analyses, and to give a final result for the samples. Then, results are shared with the classmates and discussed. In order to evaluate the knowledge acquired, students are asked to fill a questionnaire (Table 3). The questionnaire includes also some questions addressed to know how students perceive the learning process with this type of practical lessons.

Table 3. Questionnaire filled up by students as evaluation of the practical lesson.

Results obtained		
Content in total phenolics (average \pm SE)	Extract 1	
	Extract 2	
	Extract 3	
	Extract 4	
Considering the results, define the extraction solution that you would use for the measurement of total phenolics by the Folin-Ciocalteu method, and time of incubation before measuring the absorbance		
Content in antioxidant molecules (average \pm SE)	Extract 1	
	Extract 2	
	Extract 3	
	Extract 4	

Considering the results, define the extraction solution that you would use for the measurement of the DPPH-radical scavenging activity, and time of incubation before measuring the loss of absorbance					
Personal questionnaire					
	1 Totally disagree	2 Disagree	3 Not sure	4 Agree	5 Totally agree
The explanations have been clear enough to understand the lesson					
After the lesson, I could perform the analyses without teacher's guidance					
After the activities performed and the explanations provided, I think that I would be able to adjust protocols to new materials of interest					
I think that the lesson has helped to understand the theory concerning the actuation of antioxidants against free radicals and its importance for plants survival as well as for human health					
Comments related to the lesson					

3 CONCLUSIONS

The practical lesson should provide students with the competence to evaluate different antioxidant characteristics of plant materials. We consider that this is one way of contributing the development of breeding programmes focused on nutritional traits [13, 14], since the protocols used allow the quick evaluation of a large number of samples. In addition, the lesson is designed to guide students in the adjustments of the protocols, as well as to understand and solve real problems that can occur. In summary, after this practical lesson, students should feel confident in working with the protocols evaluated adapted to different materials. Finally, this practical session could also be applied to other master degrees related to Plant Science, Plant Industry and Food Technology [15, 16].

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