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4

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22

23 **Abstract**

24 Mating disruption (MD) is a species-specific and environmentally friendly pest
25 management tactic based on the release of synthetic sex pheromones aiming to interrupt the
26 mate-finding communication and prevent mating in the target pest. The present work aims
27 provide an overview of the current scientific and technical knowledge on mating disruption
28 of scale pests (Hemiptera: Coccoidea). Biparental scales are suitable targets for mating
29 disruption, as the females have a limited spreading ability, and adult males are short lived
30 and have a narrow window of time for mate searching. In this perspective, delayed mating
31 also plays an important role by reducing female attractiveness and population growth
32 potential. The mechanisms involved in MD of scales are most likely assigned to
33 ‘competitive disruption’ rather than ‘noncompetitive’ mechanisms, although no specific
34 studies addressed this issue. Mating disruption has been commercially developed and
35 increasingly applied against the vine mealybug *Planococcus ficus* (Signoret) (Hemiptera:
36 Pseudococcidae) and the California red scale *Aonidiella aurantii* (Maskell) (Hemiptera:
37 Diaspididae) to a lesser extent. Critical factors affecting MD effectiveness are the pest
38 density and effective disruption late in the season. Mating disruption applied to scale pests
39 is effective in small plots and compatible with biological control and integrated
40 management programs. In conclusion, MD has a high potential for management of scale
41 pests, providing that key factors such as technological advances in pheromone synthesis
42 and pheromone formulations, elucidation of disruption mechanisms, and simplification of
43 the registration process are addressed.

44

45 **Keywords:** Pheromones, Behavioural manipulation methods, IPM, Coccoidea, Mealybugs,
46 Armoured scales

47

48 **1 Introduction**

49 Mating disruption (MD) is a behavioural-modifying tactic of pest management, based on
50 the application of synthetic sex pheromones formulated for release in the air to prevent
51 mating in a target insect pest (Suckling 2000). It is considered an environment-friendly
52 plant protection approach, as sex pheromones are species-specific, non-toxic and active in
53 very small amounts, as compared with other pest management chemicals. In fact, although
54 many pheromone compounds have been registered and applied worldwide, there is no
55 evidence of negative effects on human health, non-target organisms or the environment
56 (Witzgall et al. 2010).

57 While MD was initially proposed as a new approach in late 1960s (Gaston et al. 1967), its
58 commercial application was only possible in the end of 1970s, after industrial-scale
59 synthesis had become available (Doane & Brooks 1981; Cardé & Minks 1995; Witzgall et
60 al. 2010). Since the 1990s, practical implementation of MD has registered an almost
61 exponential growth (Witzgall et al. 2010), though it is still far from reaching its full
62 potential (Miller & Gut 2015).

63 MD has been mainly applied to control lepidopteran pests in various agroecosystems,
64 including vineyards, fruit orchards, cotton and forests, across more than 750,000 ha
65 (Witzgall et al. 2010; Miller & Gut 2015). Only recently, MD has been integrated in pest
66 management of other insect taxa, such as scale insects (Hemiptera, Sternorrhyncha,
67 Coccoomorpha) (Walton et al. 2006; Vacas et al. 2009). Nevertheless, despite the scientific
68 and technical knowledge that has been accumulated, almost no reviews were published on
69 MD of non-lepidopteran pests, including scale insects (e.g., Tabata 2020). The scientific,
70 technological and practical developments in MD of scale insects are reviewed here, aiming
71 at summarizing the current knowledge and stimulating future studies and practical

72 implementation of MD, for a sustainable management of this economically important group
73 of plant insect pests.

74 **2 Economic importance of scale insects**

75 Scale insects or coccoids (hereafter designated as scales) are small, cryptic, piercing-
76 sucking hemipterans, mostly feeding on phloem sap, comprising more than 8,300 species,
77 distributed among 55 families and 1,214 genera (Garcia Morales et al. 2016). Scales
78 include serious pests of various agricultural, forest, and ornamental plants (Kosztarab 1996;
79 Franco et al. 2009; Garcia Morales et al. 2016; Mansour et al. 2017a). The majority of
80 economically important species belong to the two largest families, i.e., armoured scales
81 (Diaspididae, 32% of scale species) and mealybugs (Pseudococcidae, 24% of scale
82 species). The California red scale (CRS) *Aonidiella aurantii* (Maskell) (Diaspididae), the
83 maritime pine bast scale *Matsucoccus feytaudi* Ducasse (Matsucoccidae), and the
84 Bougainvillea mealybug *Phenacoccus peruvianus* Granara de Willink (Pseudococcidae) are
85 examples of important pest scales of *Citrus* spp., *Pinus pinaster* Aiton, *Bougainvillea* spp.
86 and other ornamental plants, respectively (Mendel et al. 2003; Franco et al. 2006; Beltrà et
87 al. 2010).

88 Direct damage is inflicted by feeding activity, as plant sap ingestion and injection of toxic
89 saliva components may result in leaf and fruit discoloration, defoliation, flower and fruit
90 drop, reduction of fruit growth rate, distortion of leaves, new shoots and fruits, and
91 reduction of plant vigour, leading to plant death in extreme cases (Kosztarab 1996; Franco
92 et al. 2009). Indirect damage is the result of honeydew excretion and development of sooty
93 mould (except for Diaspididae), which may reduce photosynthesis and plant growth.

94 Honeydew often attracts ants, which may disrupt the activity of natural enemies and
95 facilitate the spread of scales on the crop (Franco et al. 2009; Quesada et al. 2018). Some
96 scales, mainly mealybugs, are also vectors of plant viruses (Nault 1997; Perilla-Henao &

97 Casteel 2016). For example, several mealybug species, including the vine mealybug
98 (hereafter notated as VMB) *Planococcus ficus* (Signoret) and soft scales (Coccidae) have
99 been recognised as vectors of Grapevine leafroll-associated viruses (GLRaV) (Tsai et al.
100 2008; Mahfoudhi et al. 2009). GLRaV is the most economically important viral disease of
101 grapevines in many producing regions in the world, with an estimated loss of about
102 \$25,000 - \$40,000 per hectare, if no control measures are applied (Atallah et al. 2012).

103 Pest status of a scale is often associated with invasive species. Outside their native range,
104 the populations of invasive scales usually profit from the absence of their natural enemies
105 and, without natural control, often originate outbreaks and economic damage. Some
106 biological traits may favour the invasiveness of scales. Their small size and cryptic
107 behaviour make them difficult to detect in quarantine inspections. Also, parthenogenetic
108 reproduction in some species and high fecundity (e.g., some scales may oviposit up to
109 8,000 eggs) facilitate the establishment of scales in new territories based on just a few
110 females (Pellizzari & Germain 2010). For example, from the 256 scale species considered
111 as pests in the USA, about 75% are non-native, representing 25% of the total number of
112 scale species known in the country (Miller et al. 2005). In Europe, alien species represent
113 near 30% of scale fauna (Pellizzari & Germain 2010) and the EPPO A1 list of pests
114 recommended for regulation as quarantine pests include four scale species, i.e., *Riparsiella*
115 *hibisci* (Kawai & Takagi) (Pseudococcidae), *Margarodes prieskaensis* (Jakubski), *M. vitis*
116 (Philippi) and *M. vredendalensis* De Klerk (Margarodidae) (EPPO 2020). *Riparsiella*
117 *hibisci* and *Margarodes* spp. are considered a serious phytosanitary risk to potted plants
118 and vineyards in the EPPO region, respectively.

119 The main pathway of introduction of alien scales is the horticultural and ornamental trade.
120 It is expected that the growth of global trade will contribute to an increase in the number of
121 new introductions. The observed trend in Europe of the mean number of new alien scales

122 per year corroborates this prediction: 0.7 new alien species per year, in the period 1950-
123 1974; 1.2 in 1975-1999; and 1.3 in 2000-2007 (Pellizzari & Germain 2010). Among these,
124 new alien scales can have a high economic impact. For example, the South African
125 mealybug *Delottococcus aberiae* (De Lotto) was detected in 2009 in Eastern Spain, causing
126 serious damage to citrus crops (Beltrà et al. 2015). Since then, it has been expanding its
127 geographical distribution in the country. Recently, the Farmers Association of Valencia
128 (AVA-ASAJA) estimated that the direct losses from *D. aberiae* in 2020 will be reaching
129 113 million € (PHYTOMA 2020).

130 **3 Identification and analysis of scale sex pheromones**

131 **3.1 Scale sex pheromones identified**

132 Most scales reproduce sexually. The females produce sex pheromone to attract the
133 conspecific males. Specifically, the pheromone chemical structure of 32 scale species
134 belonging to the families Diaspididae, Matsucoccidae, Margarodidae, and Pseudococcidae
135 has been reported to date (see Tab. 1 for references). This number has been increasing
136 enormously since the 2000s, with 68% of these structures reported in the last 20 years (Fig.
137 1). The highest number of reported chemical structures belongs to mealybugs (65.6%),
138 followed by armoured scales (21.9%). The recent increase in the pace of pheromone
139 identification is possibly the result of a combination of different factors, including: the
140 growing number of new alien pest scales; the demand for sustainable pest management
141 solutions in response to the increasing social and political pressures for reducing the use of
142 pesticides; and the technological advances on the identification and synthesis of insect
143 pheromones.

144

145 **3.2 Major patterns in scale sex pheromone structures**

146 Compared with moth sex pheromones, which usually consist of straight-chain 10–18
147 carbon acetates, aldehydes, and alcohols with 0–3 double bonds (Millar 2000), scale
148 pheromones are mainly terpenoid derivatives with unique skeletons (Zou & Millar 2015).
149 Many of the scale pheromones identified to date are different carboxylic esters of
150 monoterpene, hemiterpene or sesquiterpene alcohols but with a common structural feature,
151 the two units of the alcohol moieties are linked with irregular non-head-to-tail connections,
152 whereas monoterpenes are generally composed of two isoprene units coupled by a regular
153 1–4' head-to-tail connection (Breitmaier 2006). The resulting skeletons are mainly related
154 to lavandulol(5-methyl-2-isopropenyl-4-hexenol), maconelliol[(2,2-dimethyl-3-
155 isopropylidencyclobutyl)methanol], chrysanthemol[(2,2-dimethyl-3-
156 isobutenylcyclopropyl)methanol], cyclopentylmethanol and necrodane(1,2,2,3,4-
157 pentamethylcyclopentane) derivatives. Interestingly, some patterns are typical from a
158 specific family (Tab. 1). Acyclic esters of sesquiterpenols are more abundant among the
159 pheromones of the armoured scales, whereas necrodols and esters of lavandulol and
160 chrysanthemol are exclusively found in mealybugs. In contrast, the pheromones of
161 *Matsucoccus* spp. are unsaturated aliphatic ketones and *M. prieskaensis* possesses a
162 tetramethyl primary alcohol. All armoured-scale and mealybug species whose sex
163 pheromone was identified display a wide range of host plants and their pheromone
164 structures indicate that they come from the terpenoid biosynthetic pathway. On the other
165 hand, *Matsucoccus* spp. develop exclusively on a few closely related pine species and the
166 pattern of their pheromone structures suggests the polyketide biosynthetic pathway (Zou &
167 Millar 2015). The site of production and release of sex pheromones in scales is apparently
168 more variable than in lepidopterans. In the case of moths, it is widely documented that the
169 pheromone production occurs in glands located near the tip of the abdomen through
170 modifications of fatty acid biosynthesis pathways (Jurenka 2003). However, it may differ

171 among scales. In armoured scales, pheromones are produced in glands located in the
172 pygidium and released through the rectum and anus (Moreno et al. 1972; Gullan &
173 Kosztarab 1997). In the case of mealybugs, Williams (1985) suggested that the pheromone
174 source could be the translucent pores on the hind legs of adult females, and more recently
175 this hypothesis was tested experimentally and confirmed by Waterworth et al. (2012).
176 Despite the mentioned structural similarities among species in the same family of scales, it
177 is likely that the diversification of scale pheromones is not related to the phylogeny. By
178 building a maximum-likelihood phylogenetic tree, Tabata et al. (2017) found that structural
179 similarities of pheromones among mealybug taxa are discordant with their phylogenetic
180 relationships. These authors described the pheromone of *Dysmicoccus brevipes* (Cockerell)
181 as an aldehyde with a cyclopentane but the pheromone of the most closely related species,
182 *D. neobrevipes* Beardsley, is very different. It is an acyclic acetate, more similar to that of
183 *Planococcus minor* (Maskell). Likewise, cyclobutane structures are found in four different
184 genera of mealybugs (*Planococcus*, *Pseudococcus*, *Phenacoccus*, *Maconellicoccus*) and
185 two of armoured scales (*Acutaspis* and *Aspidiotus*). This discordance between phylogeny
186 and chemical structures probably points out that selection has worked on these families of
187 insects to generate chemical signals that can be clearly discriminated from those of closely
188 related taxa for reproductive isolation (Tabata et al. 2017), which can be especially relevant
189 for coccoids, highly sedentary insects that cannot easily change their host.

190 The only known case of shared structures between scale insect species is (R)-lavandulyl 2-
191 methylbutanoate, one of the pheromone components of the mealybugs *Maconellicoccus*
192 *hirsutus* (Green) and *Phenacoccus madeirensis* Green, consisting of the same enantiomer of
193 lavandulol, but esterified with different 2-methylbutanoic acid enantiomers (S and R,
194 respectively) (Ho et al. 2009). Zhang et al. (2006) found that the compound in the sex
195 pheromone of *P. madeirensis*, (R) (R)-2-methylbutanoate, was inhibitory for the attraction

196 of *M. hirsutus* to (R)-lavandulyl (S)-2-methylbutanoate. Hence, there is no cross-attraction,
197 suggesting a unique chirality recognition system t-lavandulyl that assures the reproductive
198 isolation of the species.

199 Many moth pheromones are blends of several compounds that can be shared between
200 species and the species-specific signals are generally produced by mixing these constituents
201 in different ratios, and there are few examples of moths using unique pheromone
202 components, such as the epoxides produced by some geometrid and lymantriid moth
203 species (Millar 2000). However, all sex pheromones reported to date for scales are species-
204 specific chemicals that create singular communication channels, free of any possible
205 interference with the pheromone channel (Millar et al. 2005a). A single compound
206 generally provides a strong activity. In fact, only seven out of the 32 species reported have
207 pheromone blends of two or three compounds. For example, the first sex pheromone
208 described for a scale species was the binary blend of CRS, composed by two different
209 esters of sesquiterpenols (Roelofs et al. 1977). Other species with binary compound blends
210 as sex pheromones are the pink hibiscus mealybug *M. hirsutus*, with esters of lavandulol
211 and maconelliol (Zhang et al. 2004), *P. madeirensis*, with esters of lavandulol and
212 chrysanthemol (Ho et al. 2009), and *Dysmicoccus grassi* (Leonardi), with two esters of
213 lavandulol (de Alfonso et al. 2012). In other species employing binary blends, such as
214 *Matsucoccus josephi* Bodenheimer & Harpaz (Dunkelblum et al. 1993) and *M. feytaudi*
215 (Einhorn et al. 1990), sex pheromones are mixtures of two geometric isomers. The rarest
216 case is the pheromone blend reported for *Comstockaspis perniciosus* (Comstock), which was
217 finally described as a mixture of three compounds: 3-methylene-7- methyl-7-octen-1-yl
218 propanoate and the E/Z geometric isomers of 3,7-dimethyl-2,7-octadien-1-yl propanoate
219 (Gieselmann et al. 1979b; Anderson et al. 1981). Interestingly, for those species producing
220 blends, each compound has independent attractant activity and their combination does not

221 usually have significant additive or synergistic effects (Roelofs et al. 1977; Anderson et al.
222 1981; Ho et al. 2009). The only exception known is the two-component blend of *M.*
223 *hirsutus*, with both components needed to attract males to the pheromone source (Zhang et
224 al. 2004).

225 As Mori reviewed in 2007, bioactivity of pheromones depends on their chirality and
226 usually a single enantiomer is the responsible for the activity. However, this is not the only
227 case and there is a wide diversity in the recognition of chirality by insects, which was
228 mainly classified by Mori (2007) as follows: 1) the opposite enantiomer does not inhibit the
229 activity of the active stereoisomer; 2) the opposite enantiomer inhibits the response to the
230 active enantiomer; 3) the corresponding diastereomer inhibits the response to the active
231 enantiomer; 4) the opposite enantiomer or diastereomer are also active; 5) the natural
232 pheromone is a mixture of enantiomers or diastereomers and all of them are separately
233 active; 6) different enantiomers or diastereomers are employed by different species; 7) both
234 enantiomers are necessary for activity; 8) one enantiomer is more active than the other but
235 their mixture is synergistic. Most scale sex pheromones can be classified in groups (1) and
236 (4). Unnatural stereoisomers in general have no biological activity or are slightly active
237 (Einhorn et al. 1990; Zhang et al. 2004; Millar et al. 2012; Tabata et al. 2017a; Tabata &
238 Ichiki 2017) and their presence in the mixture does not negatively affect the biological
239 activity of the pheromone. Thus, in many cases it is not necessary to remove the opposite
240 enantiomer generated during pheromone synthesis to be employed in pest management
241 programs and racemates are perfectly active (Hinkens et al. 2001; Zada et al. 2003; El-
242 Sayed et al. 2010; Vacas et al. 2019), which is economically favourable for their
243 implementation. Possible inhibitory effects were reported in a rather few cases: the (S)-
244 isomer of solanone might be responsible for the weak activity of the racemic solanone in
245 the field for *Aulacaspis murrayae* Takahashi (Ho et al. 2014). Similarly, when mixing the

246 unnatural Z-isomer with the *P. minor* pheromone (E-isomer), the effect was inhibitory and
247 the mixture was unattractive, which suggests that stereospecific synthesis of the E-isomer
248 will be required for practical use of this pheromone (Ho et al. 2007).

249

250 **3.3 Techniques for isolation and identification of scale sex pheromones**

251 The classical methods employed since the 1970s for the isolation and identification of sex
252 pheromones consist mainly of volatile collection and chromatographic techniques. For this
253 purpose, an abundant supply of insects is needed because virgin females of scale species
254 release smaller amounts of pheromone, i.e., 0.2-10 ng/day (Levi-Zada et al. 2014; Tabata &
255 Ichiki 2015; 2016; Vacas et al. 2019) and 75 ng/day as the maximum reported (Tabata et al.
256 2012), than female moths, which release 0.3-25 ng/h (Lacey & Sanders 1992; Anfora et al.
257 2005). This implies that tens of thousands (Zada et al. 2003) and even hundreds of millions
258 (Roelofs et al. 1977) of female-day equivalents (quantity of pheromone collected from one
259 female each day) have to be sampled to obtain enough crude quantity that allows isolation
260 of the target compound and the employment of spectroscopic techniques for its
261 identification. To obtain cohorts of virgin females, males have to be eliminated from the
262 population by manually removing their prepupae or pupae, before adult emergence, or
263 treating the rearing substrates with discriminating doses of an insect growth regulator, such
264 as pyriproxifen (Zhang et al. 2004), to prevent males from undergoing full metamorphosis.
265 Once obtained, cohorts of virgin females are aerated, usually together with the rearing
266 substrate (e.g., pumpkin, squash, germinated broad beans, potato sprouts, citrus fruits, or
267 saplings), by passing a purified or clean air stream through a chamber containing the
268 individuals, for the collection of the volatiles they release on adsorbent materials. These are
269 mainly activated charcoal or different resins, such as Porapak Q, Tenax GC or HayeSep Q.
270 Then, the collected substances are extracted with solvent from the adsorbents and the crude

271 extract is first submitted to fractionation by column chromatography (successively eluting
272 with different solvent mixtures), preparative HPLC or preparative GC. The composition of
273 each fraction is studied and candidate compounds are located, guided by biological activity
274 (attraction of males) of each fraction or by comparing the volatile profiles of the extracts
275 from cohorts of virgin females with those of the controls, which can be extracted from
276 cohorts of mated females, immature stages or uninfested rearing substrates.

277 When the target compound is isolated by HPLC or GC from the crude extract or the
278 corresponding fraction, the elucidation of the pheromone structure is then based on
279 spectroscopic data (MS - mass spectrometry or NMR - nuclear magnetic resonance) and
280 chemical microreactions (e.g., hydrogenation, hydrolysis, esterification, ozonolysis) that
281 allow determining, for example, functional groups, number of unsaturations, and position
282 of double bonds. The confirmatory synthesis of the candidate structure supplies the final
283 proof, when matching synthetic and natural chromatographic and spectroscopic data,
284 coupled with the behavioural assays.

285 Other techniques of volatile collections, rather than the classical use of adsorbent resins,
286 offer advantages regarding sensitivity. Solid-phase microextraction (SPME) is a sample
287 preparation technique that integrates sampling and concentration, avoiding the use of
288 solvents (Arthur & Pawliszin 1990) and allowing the direct introduction of the sample into
289 the GC injection port. SPME/GC was first employed for the study of airborne pheromones
290 of coleopterans (Malosse et al. 1995), but it is nowadays a widespread technique. More
291 recently, the automated sequential SPME/GC-MS analysis (SSGA), which consists of a
292 programmable GC-MS autosampler equipped with a SPME syringe, has been employed to
293 study pheromones in Lepidoptera (Levi-Zada et al. 2011) and later in mealybugs (Levi-
294 Zada et al. 2014; 2019).

295

296 **4 Are scales suitable targets for mating disruption?**

297 Only obligate amphimictic insects are potential targets for MD, as facultative or obligate
298 parthenogenesis would deeply reduce the effectiveness of this control tactic. Several
299 species of Margarodidae, Coccidae, Pseudococcidae, and Diaspididae are parthenogenetic
300 (Nur 1971; Miller and Kosztarab 1979; Gullan and Kosztarab 1997), and some species have
301 both sexual and parthenogenetic lineages, including *Aspidiotus nerii* Bouché and *D.*
302 *brevipes* (Andersen et al. 2014; Tabata et al. 2016). Furthermore, hermaphroditism has been
303 reported in *Icerya* spp. (Margarodidae). However, most scales reproduce sexually,
304 including many species of high economic importance, such as the mealybugs *Planococcus*
305 *citri* (Risso), *P. ficus*, *Pseudococcus viburni* (Signoret), *P. calceolariae* (Maskell), and *P.*
306 *longispinus* (Targioni Tozzetti) (James 1937; Huang et al. 2013; Waterworth et al. 2011;
307 Silva et al. 2013).

308 Biparental scales have peculiar biological traits that are expected to make them particularly
309 susceptible to MD (Millar et al. 2005a). First, biparental scales are sexually dimorphic.
310 Adult males are delicate, short-lived (few days at most), neometabolic, winged insects, with
311 no functional mouthparts. In contrast, females are wingless and neotenic, and may live for
312 several months, if unmated or when in dormancy (Gullan & Kosztarab 1997; Franco et al.
313 2009). Scale males seem to display a daily cycle of flight activity. Three different flight
314 patterns have been identified so far: 1) morning flight onsets with sunrise; 2) near sunset; or
315 3) both in early morning and late afternoon (Rice & Moreno 1970; Moreno et al. 1974;
316 Franco et al. 2009). Recent studies using automated sequential SPME GC–MS analysis
317 (SSGA) showed that the emission of sex pheromone by females of *P. citri*, *P. ficus*, and
318 *Nipaecoccus viridis* (Newstead) follows a circadian rhythm (Levi-Zada et al. 2014; 2019),
319 in parallel with the daily flight pattern of the males. Thus, mate location flight is limited to
320 a few hours per day and male scales have a narrow window of opportunity to search for

321 females and to mate during their short life, estimated as less than 12 hours in *P. citri*,
322 although males may mate outside this period, if they succeeded in finding receptive females
323 (Silva et al. 2009; 2013; Mendel et al. 2012). This represents an advantage for MD, as a
324 short-time alteration of mate search would deeply impact the reproductive success of
325 scales. As male scales do not feed and thus are not able to replenish their limited energy
326 budget, the existence of a trade-off between flight activity, mating and longevity is
327 expected, as flight is energy-demanding and thoracic flight muscles expensive to maintain
328 (Denno et al. 1989; Legaspi & Legaspi 1998; Mendel et al. 2012). For example, a trade-off
329 between the number of copulations and longevity was recently demonstrated in
330 *Phenacoccus solenopsis* Tinsley (Tong et al. 2019). Therefore, it is expected that under MD
331 conditions, when exposed to synthetic conspecific pheromone, males may waste their short
332 window of opportunity for mating, by rapidly spending their energy reserves and strongly
333 reducing their chance of mate location and mating performance. Millar et al. (2005)
334 suggested that the pheromone in scale MD will rapidly exhaust the males, removing them
335 from the system, in a similar manner as an insecticide, but without its negative side effects.
336 Furthermore, knowledge on the existence of a circadian rhythm of female pheromone
337 emission and male flight may allow to improve MD, by programming the daily timing of
338 pheromone release (for example in aerosol spray cans, see 7.1.1), and synchronizing
339 pheromone application with female calling and male flight period, thus reducing the
340 amount of pheromone (dose) needed for scale MD.

341 Secondly, as male scales are very sensitive to conspecific sex pheromone, it is expected that
342 the necessary amount of pheromone for obtaining an effective control of scale populations
343 in MD will be relatively small (Millar et al. 2005a). As mentioned before (see 3.3), the
344 amount of pheromone released by females is much smaller in scales than in lepidopterans.

345 Thirdly, the prevalence of migration of mated females is considered the most critical trait to
346 estimate pest susceptibility to pheromone-mediated MD (Cardé & Minks 1995). For
347 example, in the case of lepidopteran pests, the possibility of immigration of fertilised
348 females from habitats surrounding MD plots is considered a major constraint of the method
349 (Ioriatti et al. 2008; Ioriatti & Lucchi 2016; Benelli et al. 2019). In such a case, the
350 effectiveness of MD is dependent on its application in relatively large areas, to reduce the
351 perimeter/area ratio of the treated crop, and consequently the likelihood of female
352 immigration, oviposition and damage. However, in the case of scales this is not an issue, as
353 the adult females are sessile and wingless (Millar et al. 2005a). In fact, MD has been shown
354 to be effective in small plots (> 0.5 ha) for both the VMB (Sharon et al. 2016; Mansour et
355 al. 2017b; Cocco et al. 2018) and the CRS (Vacas et al. 2009; 2010).

356 Other biological traits that may influence the effectiveness of MD in scales will be treated
357 in the section 5. It is known that the success of MD as a pest management tactic is highly
358 dependent on the biological characteristics of the target pest (Gut et al. 2004).

359 **5 Biological traits of scales that may influence the effectiveness of mating disruption**

360 The wingless, and sometimes legless, scale females have limited spreading ability. The
361 dispersal is mostly performed by first-instar nymphs (crawlers), the most mobile stage,
362 displaying morphological and behavioural adaptations for walking and aerial dispersal
363 (Washburn & Washburn 1984). Through wind dispersal, scale crawlers may settle in new
364 host plants up to a few hundred meters from the source (Willard 1974). On the other hand,
365 dispersal by walking is extremely slow and mostly occurs within adjacent plants, as
366 crawlers tend to settle as soon as they find a suitable feeding substrate (Grasswitz & James
367 2008). The short-range dispersal of nymphs and wingless adult females determines that
368 scale pests usually show an aggregated spatial distribution (Meats & Wheeler 2011; Pérez-
369 Rodríguez et al. 2017; Cocco et al. 2018). Differences in the degree of aggregation depend

370 on species-specific dispersal behaviour, such as thigmotaxis and phototaxis, morphological
371 traits of host plants, and natural enemy-pest interactions (Nestel et al. 1995). Aggregated
372 distribution is a potential constraint for effective MD control of scales, as the odds of short-
373 distance fortuitous male-female encounters are expected to increase in dense colonies.

374 Scale males show a positive pheromone dose-response up to a certain pheromone
375 concentration (Branco et al. 2006). Therefore, scale aggregation may promote polygyny
376 (i.e., multiple copulations in males), as it is expected that males will be more attracted by
377 large colonies of virgin females, collectively generating a stronger pheromone signal than
378 single calling females. By locating female colonies, males will increase their chance of
379 multiple mating, minimizing the energy cost of mate searching. This behavioural strategy is
380 particularly beneficial for short-lived insects, such as scale males. A higher number of male
381 copulations and a reduced mating duration and intervals have been observed under
382 laboratory conditions at higher female densities (Silva et al. 2013; Tong et al. 2019).

383 The population age structure of scales may play a role in MD success. For example, in
384 Sardinia (Italy), the overwintering population of VMB is mainly represented by mated
385 females, which start ovipositing in March-April, before the first flight of males (Lentini et
386 al. 2008). Therefore, MD dispensers, which are usually applied in late April-early May
387 before adult male appearance, are ineffective against the first generation and do not prevent
388 the development of the progeny from overwintering mated females. In this perspective, it is
389 of utmost importance the season-long effectiveness of MD dispensers, as a significant
390 reduction of matings in autumn would significantly reduce the proportion of overwintering
391 mated females.

392 Sexual communication in biparental scales is mediated by female sex pheromones. The
393 possibility of intraspecific variation in sex pheromone signals has been reported in the
394 VMB. The sex pheromone of the VMB was identified in Californian populations as a

395 single-component pheromone, i.e., (S)-lavandulyl senecioate (Hinkens et al. 2001). Soon
396 afterward, this and a second component, i.e., (S)-lavandulyl isovalerate, were detected in
397 Israeli populations of the mealybug (Zada et al. 2003). Kol-Maimon et al. (2010) showed
398 that VMB males might respond differently to the two-pheromone components, indicating
399 the existence of different male pherotypes. Based on the three possible behavioural
400 responses (attraction, indifference, repulsion) of mealybug males to each of the two
401 components, nine different pherotypes were defined, including fertile male pherotypes
402 indifferent to both pheromone compounds. Kol-Maimon et al. (2010) compared the
403 pherotypes of VMB populations from eastern (Israel) and western (Portugal) Mediterranean
404 and found that the variability of male response was much higher in eastern (9 pherotypes)
405 than in western (5 pherotypes) populations, where no males were attracted to the isovalerate
406 component. This apparent different pherotype composition in Mediterranean populations,
407 indicating a different genetic makeup, is in accordance with the results of the most recent
408 phylogeographic study on the VMB (Daane et al. 2018). The latter study suggested the
409 existence of two major population groups: 1) a European group, originating in Europe,
410 Tunisia and Turkey, which likely spread in Argentina and South Africa; and 2) a Middle
411 East group, from Israel and Egypt, which was introduced in North America and Mexico.
412 Different degrees of responses of CRS males to wild and laboratory-reared virgin females
413 have been also reported (Tashiro et al. 1969).

414 The existence of male pherotypes in scales may bear practical implications for MD. For
415 example, the effectiveness of the actual MD formulations for the VMB, based on (S)-
416 lavandulyl senecioate, in populations with pherotypes attracted to (S)-lavandulyl isovalerate
417 (e.g., some Israeli populations) may be compromised, as the mating activity of part of
418 males in the mealybug population will not be affected by the pheromone treatment.
419 Furthermore, it is expected that, in such populations, the successive application of MD

420 during several years, will create a selection pressure favouring isovalerate pherotypes,
421 which may alter the genetic structure of local VMB populations and result in resistance to
422 MD.

423 Scales may exhibit different levels of polyandry (i.e., multiple mating in females) and
424 polygyny. Overall, scale males show high mating capacity under laboratory conditions,
425 when exposed to *ad libitum* virgin females, as they are able to mate multiple times, with
426 relatively short mating intervals between consecutive copulations (James 1937; Tashiro &
427 Moffitt 1968; Mendel et al. 1990; Waterworth et al. 2011; Ricciardi et al. 2019; Silva et al.
428 2019; Tong et al. 2019). Polygyny, concurrently with scale clumped distribution, may
429 hinder the effectiveness of MD, as a single male may fertilize several females (Silva et al.
430 2013; 2019; Tong et al. 2019). However, laboratory studies probably overestimated male
431 fecundity. In the field, male performance is expected to be much lower, especially in low
432 population densities, due to the limited energy reserve of males, their short lifespan and the
433 restricted daily flight activity period (Silva et al. 2019). In addition, in mealybug males, the
434 number of copulations decrease over time, likely because of depletion of energy reserves or
435 sperm (Waterworth et al. 2011; Ricciardi et al. 2019).

436 The possibility of female multiple mating (polyandry) in mealybugs has been recently
437 observed in laboratory conditions (Waterworth et al. 2011; Silva et al. 2013) and confirmed
438 by genotype analysis of *P. citri* eggs produced by single females mated with two males
439 (Seabra et al. 2013). Nevertheless, no data are yet available on the frequency of polyandry
440 in field conditions. *Pseudococcus longispinus* females mated up to 8 times in a single day
441 and subsequent copulations occurred up to 23 days after the first event. However,
442 receptivity may be restricted to a shorter period in other species, such as *P. citri*
443 (Waterworth et al. 2011; Silva et al. 2019). After mating, females of VMB and *P. citri*
444 cease pheromone emission within 48 hours, and CRS females become unattractive within

445 24 hours (Tashiro & Moffitt 1968; Levi-Zada et al. 2014). The existence of polyandry in
446 scales may have implications in the effectiveness of MD. Effective MD is expected to have
447 more impact on polyandrous than in monandrous females, as in the absence of multiple
448 mating, none of the benefits of polyandry will occur (Silva et al. 2019). Mated females may
449 compete with virgin females thus reducing the male reproductive success (Waterworth et
450 al. 2011). However, this possibility is unlikely under MD conditions, as virgin females are
451 more frequent and more attractive to males than mated ones (Silva et al. 2019). Data
452 collected by Cocco et al. (2014) in MD plots support this hypothesis.

453 **6 Mechanisms explaining the effectiveness of mating disruption in scales**

454 The effectiveness of MD is dependent on both direct and indirect factors. Direct factors are
455 related to the mechanisms involved in the disruption of pheromone-mediated
456 communication between male and female insects (Miller & Gut 2015). Indirect factors
457 include other mechanisms, “when mating disruption does not disrupt mating”, such as
458 delayed mating (Mori & Evenden 2013).

459 Different MD mechanisms have been postulated (Bartell 1982; Cardé & Minks 1995;
460 Miller et al. 2006a), which can be divided in two main categories: competitive and
461 noncompetitive (Miller et al. 2006a). Competitive disruption includes competitive
462 attraction (false-trail-following, confusion), induced allopatry, and induced arrestment,
463 whereas noncompetitive disruption involves other mechanisms, such as suppressed
464 calling/mating, camouflage, sensory imbalance, induced allochrony, and desensitisation
465 (habituation) (Miller & Gut 2015). More than one disruption mechanism may be involved,
466 depending on the type of MD formulation and insect species (Mori & Evenden 2014;
467 Miller & Gut 2015). However, relatively few studies investigated the mechanisms
468 responsible for MD in particular cases (e.g., Flint & Merkle 1983; Lapointe et al. 2009;
469 Rodriguez-Saona et al. 2010; Mori & Evenden 2014). The meta-analysis carried out by

470 Miller et al. (2006b), based on moth sex-pheromone literature, indicated that competitive
471 disruption is the dominant mechanism.

472 Under MD conditions, other mechanisms unrelated to the disruption of mating may be also
473 involved, such as delayed mating, which may reduce female fitness and thus contribute to
474 MD effectiveness. For example, Mori & Evenden (2013) performed a meta-analysis on the
475 effect of delayed mating in female moth fitness and found a significant decrease in
476 fecundity, fertility, and pre-oviposition period and an increase in female longevity.

477 To the best of our knowledge, no specific studies were carried out to clarify the type of
478 mechanisms responsible for scale MD. Nevertheless, available data suggest that the
479 involved mechanisms are most likely part of competitive disruption. Suckling et al. (2018)
480 found no experimental evidence of habituation in males of *P. calceolariae*, as no significant
481 differences were observed in male response to sex pheromone lures (100 µg) between
482 males pre-exposed to the sex pheromone (1 mg) for 24 h and control males pre-exposed to
483 clean air. On the other hand, in a flight-tunnel simulating MD conditions (16 pheromone
484 lures distributed in 4 x 4 array, with a virgin female in the centre), males of the same
485 mealybug species showed to be attracted to the lures (Ricciardi et al. 2019). This behaviour
486 was associated with a significant decrease in female detection, compared to the control (16
487 rubber septa without pheromone), thus suggesting competitive disruption. Also, maximum
488 CRS captures were obtained in traps baited with pheromone dispensers releasing ca. 300
489 µg/day (Vacas et al. 2017), whereas the calculated minimal release rate for successful MD
490 treatments is 250 µg/day (Vacas et al. 2010).

491 As competitive disruption is a “numbers game”, its outcome in what concerns pest control
492 is dependent on the ratio between the number of pheromone release points (e.g., dispensers)
493 and the number of virgin females, i.e., it is pest-density-dependent, in contrast with
494 noncompetitive disruption mechanisms, which are pest-density-independent (Miller & Gut

495 2015). This prediction is in accordance with field results on MD of scales. In fact, Sharon et
496 al. (2016) observed that the effectiveness of MD, in the management of VMB populations,
497 decreased at high pest densities. Therefore, these results also support the hypothesis of
498 competitive disruption.

499 The fact that no complete shutdown effect (zero captures) has been reported by different
500 authors (e.g., Cocco et al. 2018; Daane et al. 2020; Silva et al. 2020) in male captures
501 registered in pheromone traps installed in MD plots for the VMB is also indirect evidence
502 supporting the hypothesis of competitive disruption mechanisms in scales.

503 Furthermore, in disruption by competition, it is expected that the additional disruption
504 effect resulting from adding more pheromone dispensers in a certain crop area, to be
505 protected by MD, will diminish with the increasing number of dispensers (Miller & Gut
506 2015). Experimental results on scale MD also support this prediction. In a two-years
507 experiment on MD of the VMB, Lucchi et al. (2019) found no dose effect on the
508 pheromone application rate, as no significant differences were found in grape damage
509 among 300, 400, and 500 dispensers/ha (i.e., 54, 72, and 90 g/ha of VMB racemic
510 pheromone, respectively). Vacas et al. (2010) compared the application of 420 and 840
511 dispensers/ha, for MD of CRS, keeping the total release rate in 113 mg/ha/day, and
512 observed no significant differences in fruit damage.

513 Delayed mating has been reported to occur in scale insect populations under MD. In MD
514 conditions for the VMB, Cocco et al. (2018) observed 18.8–66.2% reduction in the
515 percentage of ovipositing females, a mating delay of 5.5–12.5 days, a significant increase
516 of the pre-oviposition period (up to 12.5 days), and in female longevity, a significant
517 decrease in fertility, and no effect on fecundity. Lentini et al. (2018) studied in laboratory
518 conditions the effect of mating delay on the reproductive performance and population
519 growth rates of the VMB. They concluded that only a mating delay longer than 7 days

520 would lead to a reduction in the population growth rates. In the case of CRS, Vacas et al.
521 (2012) found a significant lower number of gravid females in MD plots, in comparison with
522 control, as well as a delay in the development of CRS instars.

523 As in MD conditions unmated females live longer, the increase in female longevity is
524 expected also to affect the relative attractiveness of older females due to a reduction in the
525 emission rate of the sex pheromone. In fact, Levi-Zada et al. (2014) showed that the
526 females of both *P. citri* and VMB have an age-dependent pattern of sex pheromone
527 emission, with a maximum release rate registered for middle-age females. For example, at
528 25 ± 1 °C and 13L:11D photoperiod, pheromone emission in *P. citri* started at 4-6 days-old
529 females, reached the maximum at 9-12 days-old females (2 ng/h), and decreased for older
530 females. In the case of VMB, the emission of pheromone also started at 4-6 days-old
531 females and decreased for females older than 20 days. The maximum amount of sex
532 pheromone was released by 10-20 days-old females (1-2 ng/2h). Females of both mealybug
533 species ceased pheromone release after mating (Levi-Zada et al. 2014). According to these
534 data, and for the same conditions, a reduced attractiveness would be expected for unmated
535 females older than 20 days, in the case of the VMB, although females of 1-28 days old
536 were equally attractive to males in the field (Lentini et al. 2018).

537 Under MD conditions, semiochemical-mediated interactions between scales and their
538 natural enemies may also contribute to its global effectiveness as a pest management tactic.

539 Sex pheromones and volatile cues of scales elicit kairomonal responses in several
540 parasitoids and predators and are exploited for host/prey selection (Branco et al. 2007;
541 Ishaaya & Horowitz 2009; Pekas et al. 2015; Urbina et al. 2018). For example, the encyrtid
542 *Anagyrus vladimiri* Triapitsyn (= *A. sp. near pseudococci*) uses the sex pheromone of the
543 VMB as a chemical cue for host location (Franco et al. 2008; 2011). This kairomonal
544 response of the parasitoid may contribute to enhance biological control of the VMB in MD

545 conditions. In fact, in most of the cases, the level of parasitism of the VMB has been
546 reported to be higher in MD plots compared to control (Walton et al. 2006; Cocco et al.
547 2014; Shapira et al. 2018). However, the higher parasitism level observed in MD plots may
548 be also related with the higher longevity of VMB females, which are thus expected to be
549 exposed to parasitisation for a longer period. This is apparently the explanation for the
550 increased parasitism of CRS by *Aphytis melinus* DeBach, observed under MD conditions.
551 The presence of the pheromone in MD treated plots was reported to delay the development
552 of CRS allowing a higher parasitism rate compared with untreated plots and a significant
553 higher total predation and parasitism in MD treated plots (Vacas et al. 2012; Vanaclocha et
554 al. 2012). Although the hypothesis that the CRS sex pheromone could attract *A. melinus*
555 was raised by Sternlicht (1973), later Morgan & Hare (1998) found no evidence of a
556 kairomonal response of the parasitoid to the host sex pheromone, in olfactometer
557 experiments. More recently, in field experiments, Pekas et al. (2015) also revealed no
558 attractant effect of CRS pheromone for both *Aphytis lepidosaphes* Compere and *A. melinus*,
559 although a positive response was observed for *A. chrysomphali* Mercet.

560 **7 Development and application of mating disruption formulations in pest** 561 **management of scales**

562 Here we summarize the accumulated knowledge and recent developments on the
563 implementation of MD for the control of two major scale pests, VMB and CRS, for which
564 this pheromone-based management tactic is already practiced with commercial
565 formulations.

566 **7.1. The vine mealybug**

567 The VMB is the most economically important mealybug species infesting grapevine (*Vitis*
568 *vinifera* L.) worldwide (Walton & Pringle 2004; Franco et al. 2009; Daane et al. 2012;
569 Reineke & Thiéry 2016; Mansour et al. 2018). The isolation, identification and synthesis of

570 the sex pheromone of the VMB (Hinkens et al. 2001; Millar et al. 2002) allowed its
571 application for pest management purposes. Since then, different MD formulations have
572 been developed, tested and applied for the control of its populations in many grape-growing
573 areas in Europe, North Africa, Middle East, and North and South America (Walton et al.
574 2006; Miano et al. 2011; Cocco et al. 2014; 2018; Sharon et al. 2016; Mansour et al. 2017b;
575 Lucchi et al. 2019).

576

577 **7.1.1. Formulations**

578 Over the last two decades, four formulations of MD have been tested and whenever
579 appropriate, implemented as a control tactic against VMB, in different grape-producing
580 areas in Europe, North Africa, Middle East, or America. These formulations exploiting
581 different materials and technologies include (Tab. 2): 1) sprayable microencapsulated
582 formulation (Checkmate®VMB-F); 2) membrane dispensers (Checkmate®VMB-XL); 3)
583 aerosol spray cans (Puffer®); and 4) rope (reservoir) dispensers (Isonet®PF). A double
584 rope dispenser (Isonet®LPF) is also under evaluation for the combined MD of VMB and
585 the European grapevine moth *Lobesia botrana* (Den. & Schiff.) (Baba et al. 2019; Ricciardi
586 et al. submitted).

587 **7.1.2. Dose effectiveness**

588 The first MD test against VMB was carried out in California table grapes using a sprayable
589 microencapsulated formulation, containing 16.3 and 10.8% of active ingredient (a.i.) by
590 weight of racemic lavandulyl senecioate, in 2003 and 2004, respectively (Walton et al.
591 2006). Three to four applications of 10.7 g a.i./ha were carried out, between April and
592 August, corresponding to a total of 32.1 g a.i./ha and 53.5 g a.i./ha per season, respectively
593 (Tab. 2). In addition, a delayed dormant (February) application of chlorpyrifos or an in-
594 season (June) application of buprofezin were carried out in the experiments of 2003 and

595 2004, respectively. A reduction in trap catches of adult males, mealybug density (only in
596 2003) and crop damage was obtained, but the formulation showed a relatively short (3 to 5
597 weeks) effective lifetime. More recently, Suterra developed a new microencapsulated
598 formulation (Checkmate®VMB-F), which was tested by applying 4-5 monthly pheromone
599 treatments, between late May and early September, at 12.4 g a.i./ha per treatment,
600 corresponding to a total of 49.4-61.8 g a.i./ha per season (Haviland 2017a; 2017b). The
601 possibility of being applied by conventional sprayers, along with insecticides or fungicides
602 (except those containing oil-based products, emulsifiable concentrates or including
603 organosilicone surfactants) and the inexistence of pre-harvest intervals, residues, or
604 international maximum residue limits for exported fruits, constitutes the major advantages
605 of microencapsulated formulations. In addition, the cost of each treatment
606 (Checkmate®VMB-F) corresponds to about 20% of the cost of membrane dispensers
607 (Checkmate®VMB-XL). That is, in the maximal number of five applications per season it
608 has a similar cost to that of membrane dispenser system. As a rule, microencapsulated
609 formulations are not permitted in organic farming. However, some exceptions for its use in
610 organic table grape fields in California have been granted by some organic-certifying
611 organizations in 2019 and 2020 (KCDA 2020).

612 Membrane dispensers (Checkmate® VMB-XL) loaded with 150 mg of the racemic sex
613 pheromone have been tested against VMB populations in different grapevine regions and
614 using different number of dispensers (Tab. 2). In California (USA), Langone et al. (2014)
615 applied ca. 400 dispensers per ha (62-93 g a.i./ha), in combination with a delayed dormant
616 application (before pheromone dispenser set up) of chlorpyrifos and a post-harvest
617 treatment of spirotetramat. A total of 620-625 dispensers per ha was tested in Sardinia
618 (Italy) (62.5-93.8 g a.i./ha; Cocco et al. 2014; 2018), Israel (93.8 g a.i./ha; Sharon et al.
619 2016), and Central-South Tunisia (93 g a.i./ha; Mansour et al. 2017b). In the case of

620 Sardinia, MD was combined with a treatment of chlorpyrifos or lambda-cyhalothrin, and in
621 Tunisia, with imidacloprid. In all experiments, MD significantly reduced male captures in
622 pheromone traps and mealybug densities on grapevines. Field lifetime of pheromone
623 membrane dispensers was estimated to be 130-150 days in Sardinia, with a mean
624 pheromone release rate of 484 mg/ha/day (Cocco et al. 2014; 2018), and 120 days in
625 warmer Central-South Tunisia (Mansour et al. 2017b).

626 The aerosol spray cans (Puffer®) controlled by programmable chips were also tested
627 against VMB populations, in California vineyards at a rate of ca. 4.9 spray cans per ha, in
628 combination with a delayed dormant application of chlorpyrifos and a post-harvest
629 application of spirotetramat. Spray devices were timed to release 1.3 g a.i./day throughout
630 the entire growing season. In addition, membrane dispensers were installed in the treated
631 plot perimeter, emitting about one-eighth as much pheromone as the aerosol spray cans.

632 Pheromone treated plots showed a reduction in male trap captures and grapevine damage,
633 in comparison with control plots (Langone 2013; Langone et al. 2014).

634 More recently, pheromone rope dispensers (Isonet® PF) have been tested against VMB in
635 Italian vineyards. Field trials conducted at a dose of 90 g a.i./ha (500 dispensers x 180 mg
636 a.i./ha) resulted in a noteworthy delayed mating and decrease in the number of matings, as
637 the number of ovipositing females was significantly reduced (Cocco et al. 2018).

638 Furthermore, this formulation, applied over consecutive years, significantly reduced VMB
639 density and showed a field lifetime of about 200 days and a release rate of 385 mg/ha/day
640 (Cocco et al. 2018). Similar results were observed in a three-year study in Portugal, with
641 the same dosage (Silva et al. 2020). Lucchi et al. (2019) compared the efficacy of different
642 pheromone dosages, i.e., 300, 400 or 500 rope dispensers per ha (180 mg of racemic
643 lavandulyl senecioate per dispenser, i.e., 54, 72, and 90 g a.i./ha, respectively), in northern
644 and southern Italian vineyards. They observed that all pheromone treatments significantly

645 decreased VMB density on grape bunches, and found no significant differences among
646 dosages. These results suggest that the minimal effective dose for this MD formulation
647 (Isonet®PF) is 300 dispensers per ha.

648 Recently, based on experiments carried out in California, between 2004 and 2007, Daane et
649 al. (2020) provided further information on the effectiveness of different MD formulations,
650 including sprayable formulation, membrane dispensers and rope dispensers. They
651 concluded that sprayable formulation was slightly more effective than dispensers, for the
652 same pheromone dose.

653 The new double rope dispenser (Isonet®LPF) was evaluated in 2017 and 2018 by testing
654 400, 500 and 600 units/ha at three study sites located in Southern (Sicily), Central
655 (Tuscany) and Northern (Veneto) Italy (Ricciardi et al. submitted). Trials were performed
656 by monitoring *L. botrana* and VMB populations in wine and table grape vineyards
657 managed with MD and no-treated control vineyards. MD results showed a significant
658 reduction of the number of infested inflorescences, as well as of the number of *L. botrana*
659 nests and VMB individuals per inflorescence compared with untreated controls. No
660 significant differences were found between the three dosages of Isonet® LPF dispensers.
661 Performing MD against both insect species using a single dispenser reduced the labour
662 costs, the amount of plastic tools used in the field, as well as the insecticide treatments.

663 **7.1.3. Timing of mating disruption application**

664 No specific studies have been carried out to determine the optimal application timing of
665 MD. However, most studies on MD of VMB testing membrane and rope dispensers report
666 a field deployment before the first seasonal flight of males (Cocco et al. 2014; 2018; Sharon
667 et al. 2016; Mansour et al. 2017b), which depends on climatic conditions. For example, it
668 usually occurs in March, April, and mid-May in Israel, Central-South Tunisia, and Sardinia
669 (Italy), respectively. Setting up the pheromone treatment before first male flight is in

670 accordance with best practices applied for lepidopteran pests (Ioriatti et al. 2008). In this
671 perspective, it is of utmost importance to consider the season-long effective lifespan of
672 MD dispensers, which as mentioned before was estimated as 120-150 days, in membrane
673 dispensers, and about 200 days, in rope dispensers (Cocco et al. 2014; 2018; Mansour et al.
674 2017b). In the case of microencapsulated formulations, the effective lifetime is only about
675 3-5 weeks, which impose up to 5 treatments per year, to cover the whole season (Haviland
676 2017a; 2017b). Above all, an effective disruption of male-female communication and a
677 consequent reduction of mating activity in autumn are expected to markedly reduce the
678 proportion of overwintering mated females, and thus the size of VMB population escaping
679 MD in the following spring. In fact, membrane dispensers reduced the percentage of
680 ovipositing females in October to 40-50%, whereas rope dispensers were more effective
681 late in the season by reducing the percentage of matings to 0-10% (Cocco et al. 2018). The
682 reduction of overwintering mated females by MD applied for consecutive years led to an
683 increased effectiveness of this control method resulting in a reduction of VMB density
684 (Sharon et al. 2016; Cocco et al. 2018). The cumulative effectiveness of MD in reducing
685 the population density of VMB is in accordance with findings on lepidopteran pests
686 (Stockel et al. 1994; Varner et al. 2001).

687 In hot-climate areas, the build-up of VMB populations starts as early as February and up to
688 nine generations per year have been estimated (Sharon et al. 2017). Under such conditions,
689 a single release of pheromone dispensers is not sufficient to disrupt mealybug male activity
690 for the whole season. The application of membrane dispensers twice a year, i.e. February
691 and August, induced a year-round effective MD and a significant reduction of VMB
692 population in the following year (Sharon et al. 2017).

693 The results of a four-year study carried out in California (Daane et al. 2020) suggested that
694 season-long or late-season coverage of MD is probably more important than the applied
695 pheromone dose, for the control of the VMB.

696 **7.1.4. Compatibility of mating disruption with biological and chemical control**

697 MD is considered a very selective pest management tactic, with no expected negative
698 impact on non-target organisms, as it is based on the use of sex pheromones, which are
699 species-specific semiochemicals. However, in case of parasitoids or predators showing
700 kairomonal responses to the sex pheromone of their host/prey, we may expect some
701 negative side effects. For example, it was expected that the host location process of the
702 parasitoid *A. vladimiri*, which displays a high kairomonal response to VMB sex
703 pheromone, might be disrupted by VMB MD (Franco et al. 2008; 2011). Nevertheless, the
704 experimental data available on MD of the VMB do not support this hypothesis. In fact,
705 experiments showed no significant differences, in the level of parasitisation of VMB by *A.*
706 *vladimiri*, between MD and control vineyards (Walton et al. 2006), or even an increase of
707 parasitism level in pheromone-treated plots (Daane et al. 2006; Cocco et al. 2014),
708 suggesting that MD does not disrupt parasitism or may even enhance it. More recently,
709 Shapira et al. (2018) tested whether the use of MD to control the VMB and *L. botrana*
710 affects hymenopteran parasitoids and spiders in wine-producing vineyards in Israel. The
711 results showed that MD did not influence the abundance, diversity and community
712 composition of most parasitoids and spiders. The sampled hymenopterans were mainly
713 parasitoids of leafhoppers, whiteflies, leafmining dipterans, and thrips. In the case of the
714 VMB parasitoid *A. vladimiri*, the number of female wasps captured in traps baited with the
715 pheromone of the VMB was significantly lower in MD than in control vineyards. However,
716 the parasitism of the VMB was only detected in MD vineyards. This apparent contradictory
717 result is most probably explained by a competitive effect between pheromone traps used to

718 monitor *A. vladimiri* and other pheromone sources associated with MD dispensers, in a
719 similar way to that occurring with mealybug males, due to the kairomonal attraction of the
720 parasitoid to the VMB pheromone. Overall, available data clearly indicate that MD is
721 compatible and may even enhance biological control of the VMB and other grapevine
722 pests.

723 As in moth pest species (Cardé & Minks 1995), the effectiveness of MD in the control of
724 VMB populations is density-dependent, as its efficacy decreases at high pest densities
725 (Sharon et al. 2016). Therefore, at moderate-high VMB densities MD should be combined
726 with chemical or biological control tactics, following an IPM approach (Mansour et al.
727 2017b; 2018; Lucchi & Benelli 2018). In fact, MD of the VMB has been often associated
728 with insecticide applications in IPM programs. All the available formulations increased the
729 effectiveness of insecticides commonly used in VMB control, namely chlorpyrifos,
730 buprofezin, imidacloprid, and spirotetramat (Walton et al. 2006; Cocco et al. 2014; 2018;
731 Langone et al. 2014; Haviland 2017a; Mansour et al. 2017b, 2018). Nonetheless, MD was
732 effective in reducing the mealybug density also as a stand-alone control tactic (Sharon et al.
733 2016; Lucchi et al. 2019).

734 **7.2. California red scale**

735 Once the pheromone of CRS was identified, it was used for the monitoring of the scale. The
736 first trials of MD were conducted in the early 1980s, using rubber septa pheromone
737 dispensers (Barzakay et al. 1986; Hefetz et al. 1988). These dispensers were loaded with
738 low amounts of pheromone (below 6 mg) and needed replacement every 2 months.

739 Although the treatment reached male capture reduction in monitoring traps, the efficacy in
740 terms of pest damage reduction was not proved. Later, in the early 2000s, a new
741 formulation with 0.4 mg/dispensers was registered by the United States EPA and
742 commercialised under the name Red Scale DownTM. These dispensers installed at a density

743 of 250 units per ha, and replaced every 3 months, showed low to moderate efficacy in
744 orchards with low infestation levels (Sousa et al. 2008). Twenty years after the first tests,
745 MD studies were started over in Spain employing mesoporous dispensers and using
746 pheromone loads over 50 mg per dispenser (Vacas et al. 2009).

747 **7.2.1. Formulations**

748 There are currently two main formulations commercially available for MD of CRS (Tab.
749 2): mesoporous and membrane dispensers. Both types of formulations are passive
750 dispensers (usually applied at a rate of 300-600 units per ha), in which the pheromone is
751 continuously released, regardless of the time of day or the pest flight activity.
752 Mesoporous dispensers were developed by Vacas et al. (2009), consisting of cylindrical
753 tablets of clay material on which the pheromone is retained, not only by physical methods,
754 but also by the chemical interaction of the emitter matrix with the pheromone (Domínguez-
755 Ruiz et al. 2008). These dispensers have been available in the market with several
756 trademarks, such as Scalebur® (EPA SL), Dardo® (Syngenta Agro SA) and, more recently,
757 Masslure®AoAu (Massó) (Tab. 2). They exhibit good performance during long periods and
758 are less temperature-dependent compared with rubber septa or polymeric dispensers
759 (Domínguez-Ruiz et al. 2008). It should be taken into account that a temperature-dependent
760 dispenser wastes a high amount of pheromone in the warmer hours of the day when the
761 flight activity of CRS males is very low (Gieselmann 1990). The main disadvantage of
762 these mesoporous dispensers is the affinity that some corvid birds, especially magpies, have
763 for the pheromone tablets. In areas where these birds are abundant, the dispensers are
764 pecked and thrown to the ground, with the consequent loss of pheromone sources and
765 efficacy of the treatment.

766 Membrane dispensers are based in a plastic recipient containing the pheromone, with a
767 semipermeable membrane that regulates its emission. They are commercially available as

768 CheckMate®CRS Dispenser (Suterra LCC) (Tab. 2). This kind of dispenser is more
769 sensitive to high range temperature variations, although their average pheromone release
770 rate is substantially constant under the typical temperatures of Mediterranean climates
771 where citrus crops are cultivated.

772 **7.2.2. Dose effectiveness**

773 Several studies have been carried out to calculate the quantity of pheromone required for an
774 effective MD of CRS, but the information about dispensers' release rate was not always
775 complete. First field trials conducted by Hefetz et al. (1988) demonstrated that a reduction
776 of male captures was achieved by placing 400 rubber septa per ha, with a total load of 30
777 mg of pheromone per tree, for the whole season. However, no data on the quantity of
778 pheromone released were given. In studies carried out with rubber septa emitters, the
779 amount of residual pheromone after 2-3 months varied between 36% (McQuate et al. 2019)
780 and 72% (Smit et al. 1997). Even in a longer period of 6 months, rubber septa loaded with
781 pheromones could contain up to 71% of initial load (Zhang et al. 2013). Taking the most
782 unfavourable scenario, we could ensure that the pheromone emitters described by Hefetz et
783 al (1988) would contain at least one third of their initial charge at the end of their useful
784 life. Calculating a total pheromone dose of 12 g/ha/season and considering one third of
785 residual pheromone not emitted, we can assume that 8 g/ha/season (6 months) were
786 effectively released, with a mean rate of 44 mg/ha/day to reach the MD showed in this
787 work. However, no fruit damage assessments were performed and the effect of MD was
788 only evaluated regarding male capture reduction in pheromone traps placed in the treated
789 vs. control area.

790 A second work demonstrated that a pheromone release rate over 105 mg/ha/day was the
791 most suitable to reach effective MD (Vacas et al. 2009). Results showed that release rates
792 of 16.8 mg/ha/day did not originate male catch reduction in monitoring traps, whereas

793 release rates of 42 mg/ha/day achieved moderate male catch reduction, but the level of MD
794 in the last case was not enough to reduce fruit damage in the pheromone-treated areas.
795 Another study conducted in commercial orchards demonstrated that release rates of 113
796 mg/ha/day were more effective than oil treatments, achieving 70% fruit damage reduction
797 in the MD treated plots, during the first year of treatment (Vacas et al. 2010).
798 In the trials described above, the number of dispensers per tree was one, with a plant
799 density of 400-450 trees per ha. Hefetz et al. (1988) suggested that using four dispensers
800 per tree, with a quarter of the pheromone load, and maintaining the total dose of
801 pheromone, could lead to a higher reduction in male catches. Vacas et al. (2010), in MD
802 trials with a pheromone release rate of 113 mg/ha/day, registered no significant differences
803 in fruit damage between 420 and 840 dispensers per ha. Therefore, a single dispenser per
804 tree is sufficient for effective MD, with dispenser densities over 400/ha and ensuring a
805 pheromone release rate over 113 mg/ha/day.

806 **7.2.3. Timing of mating disruption application**

807 The date of dispenser deployment is always a key point for the success of MD. A general
808 rule in case of moth pests is setting up the dispensers before the first generation male flight,
809 to prevent early potential mating and the build-up of the population. However, this rule has
810 not been demonstrated in other insects, including scales. In CRS, this may not be the best
811 strategy. CRS can complete three to five generations per year (Grout et al. 1989). In the
812 Mediterranean region, three generations usually occur and a possible fourth generation
813 could take place in some areas and warmer autumns. First male flight takes place in spring
814 and CRS populations in the first generation usually show low densities, whereas the third
815 and occasional fourth generations are abundant and directly responsible for fruit infestation,
816 in most cases. Vacas et al. (2015) conducted a two-year study to adjust the timing of
817 dispenser application and assess the importance of controlling the first generation of CRS.

818 Results demonstrated that the control of the first CRS generation is not essential for
819 achieving a good efficacy, and applying the pheromone just before the second male flight
820 can generate at least the same efficacy. Furthermore, considering that there is a period of
821 30-40 days without male activity, between the first and the second CRS male flights,
822 dispenser deployment before the first flight will originate a wasting of about 2 months of
823 pheromone emission, when compared with the application before the second male flight.
824 Therefore, according to the results obtained by Vacas et al. (2015), between the two timing
825 options for MD of CRS, i.e., disrupting the first generation or completely disrupting the
826 third or even the fourth generation, the last one seems to be the optimal management tactic.
827 Another point to consider is the cumulative effect of MD. One of the main advantages of
828 MD is the reduction of pest populations year after year (Cardé & Minsk 1995). This
829 cumulative effect can only be achieved if MD is acting during the main part of the pest
830 population growth. For this reason, when the cost of the pheromone or the lifespan of the
831 dispensers do not allow keeping them active in the field during the whole year, the
832 deployment of pheromone dispensers should be timed to cover the generations that are
833 most responsible for population growth, that is the third and fourth generations, in the case
834 of CRS.

835 **7.2.4. Plot shape and size requirements for mating disruption**

836 The minimum size of the treated plots and the distance to untreated areas are also important
837 factors for MD success. Although the migration of fertile females from outside the
838 pheromone-treated plots is not an issue in CRS, as the females are sessile, the airborne
839 pheromone concentration is lower in the edge of the treated plots, and thus males may be
840 able to find receptive females and mate. In field trials conducted by Vacas et al. (2009), a
841 buffer area of 15 m around the treated plot was delimited to obtain the best performance of
842 CRS MD. However, in the case of plots treated with MD at the lowest pheromone doses, a

843 higher fruit damage was observed in the buffer area. For this edge effect, MD treatment is
844 not recommended in plots of less than 0.5 ha and it is necessary to avoid narrow and
845 elongated shapes that do not allow fulfilling the requirements of the edges.

846 **7.2.5. Compatibility of mating disruption with biological and chemical control**

847 It is important to know whether and how MD treatment can influence mortality caused by
848 parasitoids (mainly *A. melinus*) and predators (e.g., *Rhyzobius lophanthae* (Blaisdell)) of
849 CRS. As mentioned earlier, the hypothesis raised by Sternlicht (1973) that CRS sex
850 pheromone could attract *Aphytis* species was confirmed for *A. chrysomphali*, an endemic
851 parasitoid of the Mediterranean area (Pekas et al. 2015). In contrast, no attractant effect was
852 detected for the main *Aphytis* species, *A. melinus* and *A. lepidosaphes*. In the same way,
853 Morgan & Hare (1998) and Vacas et al. (2012) demonstrated that the presence of CRS
854 pheromone in the environment does not affect *A. melinus* mating behaviour or its capacity
855 to parasitize or feed on CRS. Moreover, the presence of the pheromone in MD plots was
856 reported to delay the development of CRS, allowing a higher parasitism rate compared with
857 untreated plots and a significant higher total predation and parasitism in MD plots (Vacas et
858 al. 2012, Vanaclocha et al. 2012)

859 Another important point to highlight is the high compatibility of MD with chemical control.
860 In fact, several studies demonstrated that MD efficacy is always higher with low to medium
861 pest pressure due to the accidental encounters between males and females that occur when
862 the populations are very high in competitive mating disruption, as it has been described in
863 section 6 (Sharon et al. 2016). In such high populations, Vacas et al. (2010) showed that a
864 chemical treatment (e.g., mineral oil) in the first CRS generation, combined with MD in the
865 second and third generations engendered better results than chemical or MD treatments
866 alone.

867 **8 Actual worldwide use of mating disruption against scales**

868 Mating disruption is currently applied worldwide in about 184,500 ha against VMB
869 (129,500 ha) and CRS (55,000 ha). Suterra formulations for the VMB (CheckMate®VMB-
870 XL, CheckMate®VMB-F) are used in ca. 120,000 ha, mostly (more than 100,000 ha) in
871 California, but also in Argentina, Uruguay, South Africa, Spain, and Italy, among other
872 countries. In California, the most used product is the microencapsulated formulation
873 CheckMate®VMB-F. CheckMate®VMB-XL has been available since 2010 and was the
874 first registered product worldwide in California (Suterra LLC, pers. comm. 2020). In
875 Europe, MD of VMB using Shin-Etsu formulation Isonet®PF is employed in 9,500 ha,
876 namely in Italy (6,000 ha), Spain (3,000 ha), and Greece (500 ha) (Shin-Etsu Chemical Co.
877 Ltd, pers. comm. 2020).

878 In relation to CRS, Suterra formulations are applied in ca. 50,000 ha worldwide. Most of
879 that area is covered by CheckMate®CRS, especially in the USA. In California, where it has
880 been registered since April 2016, this product has been used in more than 40,000 ha. It is
881 available or under registration process in the main citrus-producing countries of Latin
882 America and South Africa and is expected to be also available soon in some Southern
883 European countries. Suterra produces another passive dispenser that is commercialised only
884 in Spain, as well as an aerosol formulation that is seldom used in California (Suterra LLC,
885 pers. comm. 2020). In Spain, MD is applied against CRS in ca. 5,000 ha, using different
886 trademark formulations, such as Dardo (Syngenta), Scalebur (EPA SL) and Masslure
887 (Massó). MD formulations for CRS have been registered in Europe since December 2016
888 (EPA SL, pers. comm. 2020).

889 **9 Future prospects**

890 The expansion of MD as a sustainable approach for the management of scale pests depends
891 on future developments on technological, scientific and legal aspects, such as economic
892 synthesis of pheromones, availability of efficient formulations, clarification of MD

893 mechanisms, and suitable registration procedures. That is, all aspects that may improve MD
894 effectiveness facilitate its practical application and reduce the costs.

895 The cost of synthetic pheromones is often regarded as the key limiting factor for MD. In the
896 case of scales, several species have pheromones that are structurally complex and difficult
897 to synthesize at the industrial level, which may compromise practical application of MD
898 (Rodriguez-Saona et al. 2009; Tabata 2020). Therefore, technological advances simplifying
899 the methods used for the synthesis of scale pheromones, such as the invention of a new
900 synthetic method for the lavandulol-related pheromones, e.g., VMB and *Planococcus*
901 *krauniae* (Kuwana) (Tabata 2020), can be of critical importance. The level of purity in
902 synthetic pheromones may also influence production cost, as increasing purity requires
903 more rigorous distillation processes (Hinkens et al. 2001). Daane et al. (2020) recently
904 obtained similar efficacy in field trials using 99% and 95% chemically pure racemic
905 lavandulyl senecioate in MD against the VMB.

906 The development of more efficient formulations, less temperature-dependent and with a
907 longer effective lifespan, should be further explored. For example, the release rate of
908 pheromone from dispensers can be adjusted through membrane permeability (Daane et al.
909 2020). Multispecies formulations, such as Isonet® LPF (Ricciardi et al. submitted), are
910 another approach, which may contribute to reducing application costs of MD. Suckling et
911 al. (2016) reported an average return on investment of six to one in the use of a
912 multispecies formulation (ISOMATE® 4-Play™) for MD of the codling moth *Cydia*
913 *pomonella* (L.) and several leafrollers in New Zealand. The use of aerosol spray cans for
914 MD in scales deserves further investigation, as this type of formulation has advantages over
915 passive formulations, including lower application cost, and the possibility of synchronizing
916 the pheromone release with the period of circadian activity of the target pest (Benelli et al.

917 2019; Daane et al. 2020). As mentioned earlier, male flight activity and female emission of
918 pheromone in scales are limited to a few hours per day.

919 Further studies are also needed for optimising the pheromone dose (e.g., dependent on the
920 number of dispensers per ha and pheromone load of each dispenser; or the number of
921 applications per season and the application rate, in the case of flowable formulations), and
922 better defining the seasonal coverage of MD in scales (i.e., identification of critical periods,
923 optimisation of coverage strategy; Daane et al. 2020).

924 The elucidation of MD mechanisms in scales, besides the scientific interest per se, has
925 important practical implications. For example, in competitive disruption, which is pest-
926 density-dependent, the suppression of pest reproduction is not expected in high population
927 densities, whereas in non-competitive disruption mating will be strongly suppressed even in
928 high pest pressure. Further related examples are discussed by Miller & Gut (2015).

929 Finally, further developments in legal aspects related with MD formulations are still needed
930 to facilitate their registration and use, as part of the global trend aiming to reduce our
931 dependence on pesticides (Lechenet et al. 2017; Brühl & Zaller 2019; Möhring et al. 2020).

932 Regulatory requirements for pheromones should take into consideration the expected low
933 risk for human health and environment of these semiochemicals, due to their specific
934 properties, namely the specificity, the use in low doses close to natural concentrations, and
935 the rapid dissipation and/or degradation (European Commission 2016). In the USA,
936 pheromones are considered biopesticides, which generally require much less data to register
937 than a conventional pesticide. In fact, new biopesticides are often registered in less than a
938 year, compared with an average of more than three years for conventional pesticides (EPA
939 2020). However, in Europe, pheromones are classified as Plant Protection Products and fall
940 under Regulation 1107/2009, which has been mainly designed for classical synthetic
941 pesticides and does not completely consider the specific nature of pheromones, although

942 the registration process of Straight Chain Lepidopteran Pheromones has been simplified
943 (European Commission 2014). Nevertheless, this is not the case for other groups, including
944 scale pheromones. For example, the registration process for the approval of the VMB
945 pheromone for MD in Europe started four years ago and is still in progress (EFSA 2020).

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954

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Table 1. List of identified scale sex pheromones.

Family/species	Pheromone compound	Reference	Type	Molecular formula
Diaspididae				
<i>Acutaspis albopicta</i> (Cockerell)	[(1S,3S)-2,2-dimethyl-3-(prop-1-en-2-yl)cyclobutyl]methyl (R)-2-methylbutanoate	Millar et al. (2012)	Ester-sesquiterpenic cyclobutane	C ₁₅ H ₂₆ O ₂
<i>Aonidiella aurantii</i> (Maskell)	3-methyl-6-isopropenyl-9-decen-1-yl acetate (Z)-3-methyl-6-isopropenyl-3, 9-decadien-1-yl acetate	Roelofs et al. (1977)	Sesquiterpenol ester	C ₁₆ H ₂₈ O ₂ C ₁₆ H ₂₆ O ₂
<i>Aonidiella citrina</i> (Coquillett)	(E)-3,9-dimethyl-6-isopropyl-5,8-decadien-1-yl acetate	Gieselmann et al. (1979a)	Sesquiterpenol ester	C ₁₇ H ₃₀ O ₂
<i>Aspidiotus nerii</i> Bouché	(1R,2S)-cis-2-isopropenyl-1-(4'-methyl-4'-penten-1'-yl)cyclobutaneethanol acetate	Einhorn et al. (1998)	Cyclobutane sesquiterpenol ester	C ₁₇ H ₂₈ O ₂
<i>Aulacaspis murrayae</i> Takahashi	(5R,6E)-5-isopropyl-8-methyl-6,8-nonadien-2-one	Ho et al. (2014)	Nor-sesquiterpene ketone	C ₁₃ H ₂₂ O
<i>Pseudaulacaspis pentagona</i> (Targioni Tozzetti)	(R,Z)-3,9-dimethyl-6-isopropenyl-3,9-decadien-1-ol propionate	Heath et al. (1979)	Sesquiterpenol ester	C ₁₈ H ₃₀ O ₂
<i>Comstockaspis perniciosus</i> (Comstock)	(Z)-3,7-dimethyl-2,7-octadien-1-yl propanoate 3-methylene-7-methyl-7-octen-1-yl propanoate (E)-3,7-dimethyl-2,7-octadien-1-yl propanoate	Gieselmann et al. (1979b) Anderson et al. (1981)	Esters of geraniol and nerol	C ₁₃ H ₂₂ O ₂ C ₁₃ H ₂₂ O ₂ C ₁₃ H ₂₂ O ₂
Margarodidae				
<i>Margarodes prieskaensis</i> (Jakubski)	(2R,4R,6R,8R)-2,4,6,8-tetramethylundecan-1-ol	Burger et al. (2017)	Tetramethyl primary alcohol	C ₁₅ H ₃₂ O
Matsucoccidae				
<i>Matsucoccus feytaudi</i> Ducas	(E,E)-8,10-(3S,7R)-3,7,9-trimethyldodecadien-6-one (8Z, 10E)-3,7,9-trimethyl-8,10-dodecadien-6-one	Einhorn et al. (1990)	Unsaturated aliphatic ketones	C ₁₅ H ₂₆ O C ₁₅ H ₂₆ O
<i>Matsucoccus josephi</i> Bodenheimer & Harpaz	(2E,4E,8E)-4,6-dimethyl-2,4,8-decatrien-7-one (2E,4Z,8E)-4,6-dimethyl-2,4,8-decatrien-7-one	Dunkelblum et al. (1993)		C ₁₂ H ₁₈ O C ₁₂ H ₁₈ O
<i>Matsucoccus matsumurae</i> (Kuwana) (= <i>Matsucoccus resinosa</i> (Kuwana), <i>Matsucoccus thunbergiana</i> Miller & Park)	(2E, 4E)-4,6,10,12-tetramethyl-2,4-tridecadien-7-one	Lanier et al. (1989)		C ₁₇ H ₃₀ O
Pseudococcidae				

<i>Crisicoccus matsumotoi</i> (Siraiwa)	3-methyl-3-butenyl-5-methylhexanoate	Tabata et al. (2012)	Hemiterpenol ester	C12H22O2
<i>Delottococcus aberiae</i> (De Lotto)	(4,5,5-trimethyl-3-methylenecyclopent-1-en-1-yl)methyl acetate	Vacas et al. (2019)	β -necrodol ester	C12H18O2
<i>Dysmicoccus brevipes</i> (Cockerell)	(-)-(anti-1,2-dimethyl-3-methylenecyclopentyl)acetaldehyde	Tabata et al. (2017)	Cyclopentane/aldehyde	C10H16O
<i>Dysmicoccus grassii</i> (Leonardi)	(R)-5-Methyl-2-(prop-1-en-2-yl)-hex-4-enyl acetate	de Alfonso et al. (2012)	Lavandulol ester	C13H22O2
	(R)-5-Methyl-2-(prop-1-en-2-yl)-hex-4-enyl propionate			C12H20O2
<i>Dysmicoccus neobrevipes</i> Beardsley	(+)-(E)-2-isopropyl-5-methylhexa-3,5-dienyl acetate	Tabata & Ichiki (2015)	Acyclic ester	C12H20O2
<i>Ferrisia virgata</i> (Cockerell)	(Z)-((1S,3R)-2,2-dimethyl-3-(2-methylprop-1-enyl)cyclopropyl) 2-methylbut-2-enoate	Tabata & Ichiki (2017)	Chrysanthemol ester	C14H22O2
<i>Maconellicoccus hirsutus</i> (Green)	(R)-2,2-dimethyl-3-(1-methylethylidene)cyclobutylmethyl (S)-2-methylbutanoate	Zhang et al. (2004)	Cyclobutane/maconelliol ester	C15H26O2
	(R)-2-isopropenyl-5-methyl-4-hexenyl (S)-2-methylbutanoate		Lavandulol ester	C15H26O2
<i>Nipaecoccus viridis</i> (Newstead)	2,2,3,4-tetramethyl-3-cyclopentenyl-methyl isobutyrate	Levi-Zada et al. (2019)	γ -necrodol ester	C14H24O2
<i>Phenacoccus madeirensis</i> Green	trans-(1R,3R)-chrysanthemyl (R)-2-methylbutanoate	Ho et al. (2009)	Chrysanthemol ester	C15H26O2
	(R)-2-isopropenyl-5-methyl-4-hexenyl (R)-2-methylbutanoate		Lavandulol ester	C15H26O2
<i>Phenacoccus solenopsis</i> Tinsley	(2,2-dimethyl-3-isopropylidene)cyclobutylmethyl 3-methyl-2-butenolate	Tabata et al. (2016)	Maconelliol ester	C15H24O2
<i>Planococcus citri</i> (Risso)	(1-R-cis)-(+)-2,2-dimethyl-3-(1-methylethenyl)cyclobutanemethanol acetate	Bierl-Leonhardt et al. (1981)	Cyclobutane/ester	C12H20O2
<i>Planococcus ficus</i> (Signoret)	(S)-5-methyl-2-(prop-1-en-2-yl)-hex-4-enyl 3-methyl-2-butenolate	Hinkens et al. (2001)	Lavandulol ester	C15H24O2
<i>Planococcus kraunhia</i> (Kuwana)	2-isopropylidene-5-methyl-4-hexen-1-yl butyrate	Sugie et al. (2008)	Lavandulol ester	C14H24O2
<i>Planococcus minor</i> (Maskell)	(E)2-isopropyl-5-methyl-2,4-hexadienyl acetate	Ho et al. (2007)	Lavandulol ester	C12H20O2
<i>Pseudococcus baliteus</i> Lit	2-((S)-1,2,2-trimethyl-3-cyclopentenyl)-2-oxoethyl (S)-2-methylbutyrate	Tabata et al. (2020)	Ester of α -hydroxyketone	C15H24O3
<i>Pseudococcus calceolariae</i> (Maskell)	(1R,2R)-[2,2-dimethyl-3-(2-methylprop-1-enyl)cyclopropyl]methyl (R)-2-acetoxy-3-methylbutanoate	El-Sayed et al. (2010)	Chrysanthemol ester	C17H28O4
<i>Pseudococcus comstocki</i> (Kuwana)	2,6-dimethyl-3-acetoxy-1,5-heptadiene	Negishi et al. (1980)	Lavandulol ester/norterpenol	C11H18O2
<i>Pseudococcus cryptus</i> Hempel	(1R,3R)-3-isopropenyl-2,2-dimethylcyclobutylmethyl 3-methyl-3-butenolate	Arai et al. (2003)	Cyclobutane/ester	C15H24O2
<i>Pseudococcus longispinus</i> (Targioni Tozzetti)	2-(1,5,5-trimethylcyclopent-2-en-1-yl)ethyl acetate	Millar et al. (2009)	Ester of 1,2,2-trimethylcyclopentane	C12H20O2
<i>Pseudococcus maritimus</i> (Ehrhorn)	(R,R)-trans-(3,4,5,5-tetramethylcyclopent-2-en-1-yl)methyl 2-methylpropanoate	Figadere et al. (2007)	α -necrodol ester	C14H24O2
<i>Pseudococcus viburni</i> (Signoret)	(1R,2R,3S)-(2,3,4,4-tetramethylcyclopentyl)methyl acetate	Millar et al. (2005b)	Ester of 2,3,4,4-tetramethylcyclopentane	C12H22O2

1 **Table 2.** Worldwide application of mating disruption in the control of the vine mealybug
 2 (VMB) and California red scale (CRS): formulations and grape-growing areas.

3

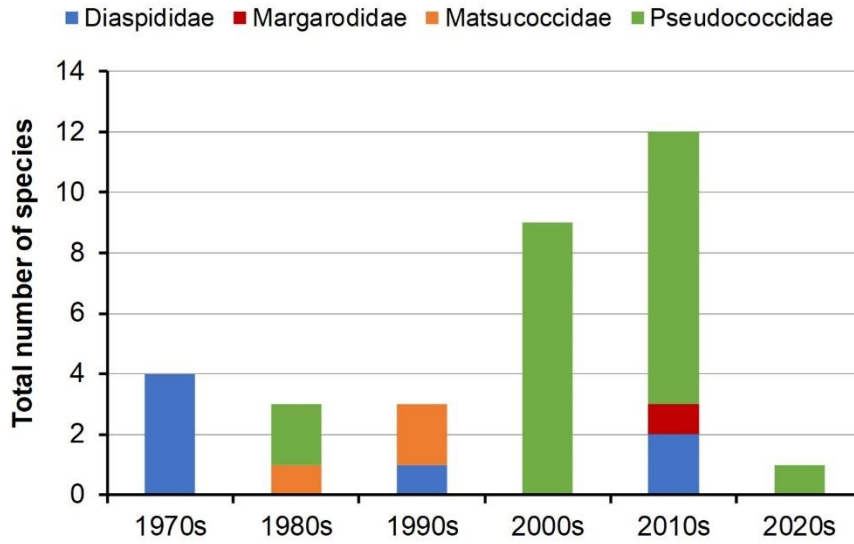
Scale species	Type of formulation	Trade name	Country or region	References
VMB	Sprayable microencapsulated formulation	Checkmate®VMB-F (Suterra LLC)	California (USA)	Daane et al. (2006; 2020); Walton et al. (2006); Haviland (2017a; 2017b)
	Membrane (reservoir) dispensers	Checkmate®VMB-XL (Suterra LLC)	California (USA), Israel, Italy, Tunisia	Cocco et al. (2014; 2018); Langone et al. (2014); Sharon et al. (2016); Mansour et al. (2017a); Daane et al. (2020)
	Aerosol spray cans	Puffer® (Suterra LLC)	California (USA)	Langone et al. (2014)
	Rope dispensers	Isonet®PF (Shin-Etsu Chemical Co. Ltd)	Italy	Cocco et al. (2018); Lucchi et al. (2019); Daane et al. (2020)
CRS	Mesoporous dispensers	Scalebur® (EPA SL); Dardo® (Syngenta Agro SA); Masslure®AoAu (Massó)	Spain, Portugal	Vacas et al. (2009; 2010)
	Membrane dispensers	CheckMate®CRS Dispenser (Suterra LCC)	California (USA), Uruguay	Casado et al. (2018)

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6 **Fig. 1.** Evolution of the total number of sex pheromones described for scale species
7 reported by families, from seventies to date.

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