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Additional Information

1 **Fate and removal of microplastics in unplanted lab-scale**
2 **vertical flow constructed wetlands**

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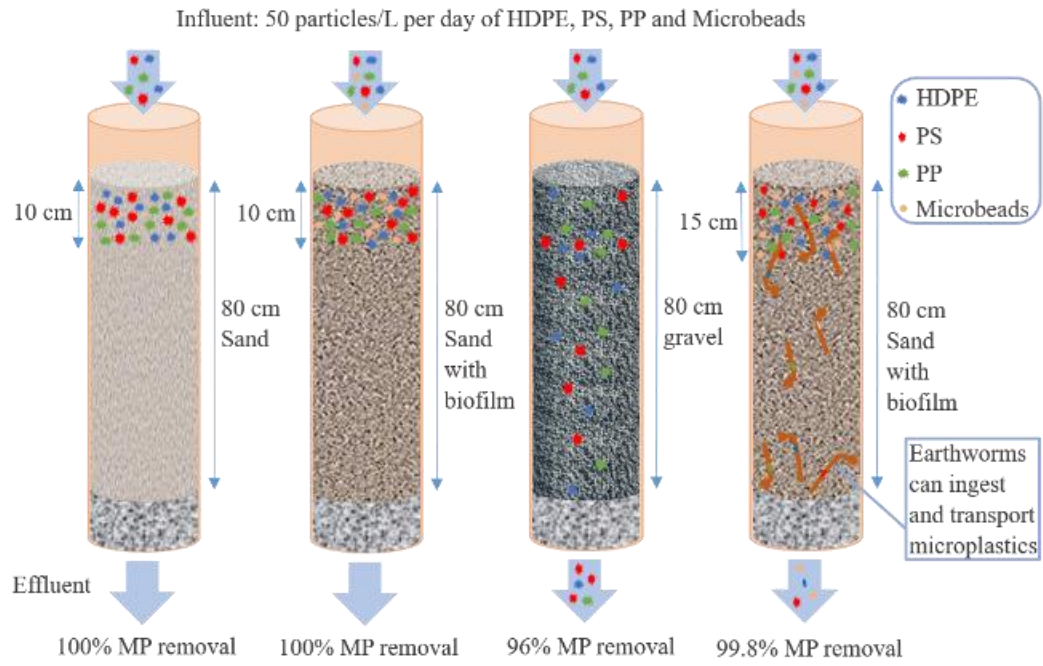
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1. Porous media of vertical-flow constructed wetlands (VFCWs) can remove microplastics.
2. Small size and spherical microplastics showed higher mobility in lab-scale VFCWs.
3. Microplastics were distributed throughout the full height of gravel-filled VFCWs.
4. Sand-filled VFCWs retained more than 98.8% of microplastics in the top 10 cm layer.
5. Earthworms can transport microplastics to the bottom of VFCWs and egest them.

15 **Abstract**

16 Many studies have reported conventional wastewater treatment plants as one of the
17 main sources of microplastics (MPs). However, constructed wetlands (CWs) as a
18 nature-based wastewater treatment system have received little attention. This study
19 investigated the influence of biofilm, media type and earthworms on the fate and
20 removal of MPs in a short-term (45d) experiment with unplanted lab-scale vertical
21 flow CWs (VFCWs). In sand-filled VFCWs, MPs were retained in the first 10 cm,
22 and the removal efficiency was 100%, regardless of the presence of a biofilm. When
23 gravel was used as filling material, the removal efficiency of MPs was stable at 96%,
24 but the MPs were distributed throughout the 80 cm high VFCWs. In the presence of
25 earthworms, the maximum depth that MPs reached within sand-filled VFCWs
26 increased from 10 to 15 cm. Furthermore, the MPs concentration at a depth of 3-6 cm
27 and 6-10 cm increased 2 and 10 fold respectively compared to the same VFCWs
28 without earthworms. Although no MPs were detected in the sand from deep layers
29 (15-80 cm), transport of MPs from top to the bottom by earthworms was found, and a
30 few MPs were detected in the effluent, leading to a removal efficiency of 99.8%. This
31 study indicated that both a higher media grain size and the presence of earthworms
32 have a small effect on the removal efficiency of MPs in VFCWs, but the effect on the

33 distribution of MPs was considerably. Longer-term studies in full-scale CWs are
34 advised to perform under the influence of more practical factors.

35 **Keywords:** Microplastics; Constructed wetlands; Biofilm; Media; Earthworms

36 **1. Introduction**

37 Microplastics (MPs) pollution was first proposed by Thompson et al. (2004) and has
38 gradually become a focus of global research. MPs are commonly defined as plastic
39 particles smaller than 5 mm (Arthur et al., 2009). It was reported that MPs could be
40 accumulated by fish, birds and mammals which are feeding on aquatic organisms or
41 living in aquatic environments. The ingestion of MPs could be caused by the
42 organisms' inability to distinguish MPs from food, or through predation of lower
43 trophic creatures containing these particles (Fendall and Sewell, 2009; Ivar do Sul and
44 Costa, 2014). Several impacts of MPs on organisms have been reported, such as
45 reduction of the predatory performance and efficiency (de Sá et al., 2015), oxidative
46 stress (Della Torre et al., 2014), neurotoxicity (Ribeiro et al., 2017), genotoxicity
47 (Della Torre et al., 2014), growth delay (Redondo-Hasselerharm et al., 2018) and
48 causing death (Li et al., 2016). Human beings are also exposed to the ubiquitous MPs
49 (Li et al., 2020), and its potential negative effects (Smith et al., 2018).

50 There are two main sources of MPs: primary MPs which are plastics that appear
51 directly in the environment at micron scale, including microbeads from personal care
52 products and microplastic powder used in paint and sand blasting (Cole et al., 2011),
53 and secondary MPs which are formed from the large size plastic waste after aging in
54 the natural environment (Rillig, 2012).

55 Human activities introduce large quantities of plastic waste into the environment,
56 which is mainly a result of mismanagement of the waste system such as industrial
57 pollution, urban surface runoffs and domestic wastewater. Previous studies have
58 reported that domestic washing and personal care products were the major sources of
59 MPs (De Falco et al., 2018; Fendall and Sewell, 2009). These MPs from households
60 could be directly discharged into the sewer system with wastewater and end up at
61 wastewater treatment plants (WWTPs) where the technologies used were not designed
62 for MPs removal. Several studies have reported the MPs removal in each stage of
63 WWTPs (Ben-David et al., 2021; Sun et al., 2019). The pre-treatment and primary
64 treatment removed the most of MPs, with removal efficiencies of 35% - 59% in pre-
65 treatment and 50% - 98% in primary treatment. After secondary treatment and tertiary
66 treatment, MPs in wastewater were further decreased to 0.2% - 14% and 0.2% - 2%
67 relative to the influent respectively. The total removal efficiencies of MPs in WWTPs

68 were reported to above 88% without tertiary treatment and to be up to over 97% with
69 tertiary treatment (Murphy et al., 2016; Sun et al., 2019). Despite this large reduction,
70 WWTPs are still releasing millions of MPs into receiving water every day (Mason et
71 al., 2016; Murphy et al., 2016). It was reported that MPs concentration significantly
72 increased in the downstream of WWTPs, which means that effluent from WWTPs is a
73 major source of the MPs in the environment (Estahbanati and Fahrenfeld, 2016;
74 McCormick et al., 2014).

75 Constructed Wetlands (CWs), as a robust nature-based solution for wastewater
76 treatment (Rousseau et al., 2008), have been widely applied for small communities in
77 Europe. As these CWs often discharge in small water courses with low dilution rates,
78 the environmental impact can be quite high. CWs have a relatively good removal
79 efficiency on total suspended solids (TSS) (over 80%) (Dotro et al., 2017) when
80 compared with other stages in WWTPs (i.e. 50% - 70% in primary treatment and 80%
81 - 90% in bio-treatment) (Silva et al., 2014). MPs as solid and micron particles can be
82 considered to have the same properties as TSS in wastewater. Therefore, it can be
83 expected that CWs may also have a good performance on MPs removal. Up to now,
84 few studies have reported MPs removal in CWs. Wang et al. (2020) have studied MPs
85 removal in a horizontal subsurface flow CW used as tertiary treatment in a small

86 community. The MP concentrations in wastewater were significantly reduced from
87 6.45 to 0.77 particles/L after tertiary treatment and the removal efficiency of the
88 global WWTP was further increased to 98%. Wei et al. (2020) also have investigated
89 four horizontal subsurface flow CWs which also served as tertiary treatments in
90 Hangzhou, China. The results suggested that CWs could strengthen MPs removal in
91 WWTPs.

92 Nevertheless, it can be expected that the impact of different factors in CWs on MPs
93 removal may vary largely, such as the biofilm present in vertical flow constructed
94 wetlands (VFCWs), the choice of media type (sand or gravel) and the presence of
95 macroinvertebrates. The biofilm in VFCWs has been reported to reduce the effective
96 porosity (Zhao et al., 2009), which is expected to influence the removal and
97 distribution of MPs. Also, the different possible media (e.g. sand and gravel) have
98 different size, which would result in varied filtration capacities (Crittenden et al.,
99 2012). As for macroinvertebrates and particularly earthworms, it has been reported
100 that they were successfully introduced into CWs to remediate clogging problems (Li
101 et al., 2012). However, the life and health of earthworms can be influenced, even with
102 low concentrations of MPs (Huerta Lwanga et al., 2016). It can thus be hypothesized
103 that MPs in CWs could have an effect on the clogging remediation of CWs by

104 affecting the earthworms. On the other hand, the earthworms may have an influence
105 on the removal and distribution of MPs in CWs due to bioturbation. It has been
106 reported that earthworms could be able to transport particles typically of millimeter
107 size (0.5-1 mm), such as plant seeds and sand particles (Shumway and Koide, 1994).
108 A recent study also showed that earthworms could transport MPs in soil from the
109 surface to a depth of 10 cm (Rillig et al., 2017b).

110 The hypotheses of this study are (i) small size media and presence of biofilms can
111 enhance the removal and retention capacity of VFCWs for MPs, (ii) earthworms in
112 VFCWs can play an important role in MPs transport. The result of this study is
113 expected to fill part of the knowledge gap and to give directions for further research
114 on full-scale CW where additional factors are anticipated to play a role.

115 **2. Materials and methods**

116 2.1 Materials

117 The MPs used in this study were polystyrene (PS) particles, high-density polyethylene
118 (HDPE) particles, polypropylene (PP) particles and PE microbeads (Table 1 and Fig.
119 1). Apart from the microbeads (45-53 μm) which were bought from Cospheric
120 (California, US), the other three kinds of MPs were made from daily utensils which

121 are the main sources of plastic waste: plastic cups (PS), caps (HDPE) of water bottle
122 and food containers (PP). To facilitate later identification, a different bright color was
123 selected for every MP type. The selected utensils were first shredded into small pieces
124 by a scissor. Then the pieces were further cut in a kitchen blender into microparticles.
125 After that, the particles were collected and screened by a stack of sieves (5600 μm ,
126 425 μm , 125 μm , 75 μm) (Wang et al., 2020). The particles remaining on the 125 μm
127 and 75 μm sieves were collected. Before use, these MPs were separated by a
128 sonication equipment in case they stuck together. The sand and gravel used in this
129 study were bought from a DIY shop (Kortrijk, Belgium), and characterized in the lab.
130 The average diameter of sand was 356 μm , and the size fractions were 10% < 231
131 μm , 50% < 400 μm , 90% < 668 μm . The diameter of the gravel (pebble) used was
132 2000 μm < 30% < 5600 μm and 5600 μm < 70% < 8000 μm . Earthworms were
133 purchased from the company Eco-cult Worms (Roeselare, Belgium), including a mix
134 of two species: *Eisenia fetida* and *Lumbricus castaneus*. Before the earthworms were
135 added to CWs, they were washed with distilled water and kept on moistened filter
136 paper at 22 ± 1 °C for 24 hours to void their gut contents (Wang et al., 2019).

137 2.2 Experimental design

138 To investigate the effect of media type, biofilm and earthworms on MPs fate and
139 removal, 10 unplanted lab-scale VFCWs (Fig. S1) with a height of 100 cm and a
140 diameter of 12.5 cm were constructed in a thermostatic room (22 ± 1 °C). Each
141 VFCW was packed with 80 cm of the respective media for the main biologically
142 active layer, and 5 cm gravel for a lower drainage layer. In order to separate particles
143 from liquids of the effluent, funnels containing 40 μ m sieves were attached to the
144 bottom of each column. The solids on the funnels were collected for MPs analysis
145 every 5 days. To avoid difficulties with the analyses, the top of the columns was
146 covered with perforated aluminum foil to prevent MPs contamination from the air
147 (e.g. operation accident with MPs used, fibers from clothes) and to prevent the
148 earthworms from escaping. The specific conditions of each VFCW are shown in
149 Table 2. VFCW1 and VFCW2 were run as blank control group (fed with tap water
150 without MPs). VFCW3 and VFCW4 were filled with sand and run with tap water.
151 VFCW5 and VFCW6 were filled with sand and run with synthetic wastewater to
152 grow biofilms. VFCW7 and VFCW8 were filled with gravel and run with tap water.
153 VFCW9 and VFCW10 were filled with sand and run with synthetic wastewater and
154 earthworms were added after 20 days operation, allowing thus sufficient time for
155 organic matter to accumulate inside the wetlands for earthworms to survive on. It

156 should be noted that vegetation was not selected as a variable in this initial study, to
157 have maintain a clear focus on the processes of interest, i.e. physical filtration
158 influenced by pore size, effect of biofilm development, and effect of earthworm
159 bioturbation. In addition, there are two processes were vegetation is expected to have
160 an impact, i.e. (1) physical disturbance of the top layer by wind-induced movement,
161 and (2) formation of new channels by root growth decay. The former could not be
162 mimicked at lab-scale, and the latter was estimated to be insignificant in a short-term
163 study.

164 Synthetic wastewater was prepared with a simple formula, because its main purpose
165 was to stimulate biofilm growth, and not to represent real wastewater (Hoagland and
166 Arnon, 1950). Briefly, synthetic wastewater was prepared adding sugar (160 mg/L),
167 milk (0.16 mL/L) and a nutrient solution (0.4 mL/L) to tap water, the latter based on
168 Hoagland and Arnon (1950). The proportions of sugar and milk were estimated to
169 have a concentration of biological oxygen demand (BOD) of approximately 150 mg/l,
170 typical for the effluent of primary treatment. The fertilizer solution contained 21.85
171 mg/mL NO₃-N, 17.25 mg/mL NH₄-N, 29.9 mg/mL carbamide, 57.5 mg/mL P₂O₅, 69
172 mg/mL K₂O, 0.046 mg/mL Cu-EDTA, 0.4255 mg/mL Fe-EDTA, 0.0345 mg/mL Mn-
173 EDTA, 0.01725 mg/mL Mo-EDTA and 0.1725 mg/mL Zn-EDTA. The tap water was

174 dechlorinated by settling down for at least 1 hour before being introduced into the
175 VFCWs. The density of synthetic wastewater was $1.001 \pm 0.001 \text{ g/cm}^3$ (20 °C). As the
176 density is very similar to tap water, the transport of MPs in the VFCW would not be
177 affected. The hydraulic loading rate was set at a typical 0.08 m/d for VFCWs (Dotro
178 et al., 2017), with the VFCWs being fed twice per day manually (0.5 L per feeding),
179 and the feeding events were practical paused during weekends. The MPs were added
180 with the synthetic wastewater or the tap water, 50 particles/L for each kind of MP.
181 Each VFCW was run for 45 days.

182 2.3 MPs extraction methods

183 2.3.1 Extracting MPs from the effluent

184 The effluents of VFCWs were collected every 5 days for MPs analysis. The
185 separation method was adopted from a previous study (Wang et al., 2020). For the
186 effluent, the particles on the meshes in the funnels were flushed by distilled water into
187 beakers which were put in the oven at 70 °C to dry up. After that, 20 ml high-density
188 solution (CaCl_2 1.34 g/cm^3) was added to the beakers and they were mixed with the
189 help of a glass bar by hand (Stolte et al., 2015). Then the liquids were transferred
190 from beakers to vials. To make sure all particles were recovered, the beakers were
191 washed 3 times with the CaCl_2 solution, and the washing liquids were also added to

192 the vials. Next, the vials were centrifuged for 5 min at 3500 rpm, and the supernatants
193 were vacuum filtrated with glass microfibers filters (Particle retention 1.5 μm , 47 mm
194 diameter, VWR) to collect the MPs from the samples. The vials were then filled again
195 with high-density solution and centrifuged one more time to ensure complete MP
196 recovery. Finally, the filters were placed in aluminum containers and put in the oven
197 at 70 °C for 5 min to dry before MP counting.

198 2.3.2 Extracting MPs from the solids

199 At the end of the experiment, the MPs inside the CWs were also analyzed with a
200 separation method adopted from O'Connor et al. (2019). The solids were analyzed by
201 layer. According to the previous study, the division of layers in the upper region was
202 relatively dense. The division of layers in deeper regions was based on the analytical
203 result of MPs in the above layer. So the CWs filled with sand were divided into 7
204 layers, which were 0-3, 3-6, 6-10, 10-15, 15-30, 30-50 and 50-80 cm, and CWs with
205 gravel were divided into 9 layers which were 0-3, 3-6, 6-10, 10-15, 15-20, 20-30, 30-
206 40, 40-50 and 50-80 cm. Each layer was first mixed to homogenize it and weighed
207 before drying (W_0), then two subsamples (30 g) were taken to assess humidity and
208 loss on ignition (LOI), and two more subsamples (30 g) were taken to extract MPs.
209 The solids samples were put in a flask and the same high-density CaCl_2 solution was

210 added. Then the flasks containing the sand samples were put on a magnetic stirrer and
211 thoroughly stirred for 20 minutes in order to separate the MPs from the sand and have
212 them floating on the surface. The flasks containing the gravel samples were sealed
213 and shaken by hand and then inverted in order to let the MPs afloat. This shaking and
214 inverting step was repeated 3 times. After that, the solution was left to settle for 2
215 hours and the supernatant was collected to be further analyzed as before. The
216 concentration of MPs was calculated by Eq. (1):

$$217 \quad c = \frac{N_0 \times W_0}{W_1 \times V} \quad (1)$$

218 where c is the MPs concentration (particles/cm³), N_0 is the number of particles
219 detected in each subsample, W_0 is the total weight of each layer, W_1 is the fixed
220 subsample weight 30 g, V is the total volume of each layer (cm³) and calculated by
221 height and diameter.

222 2.3.3 Extracting MPs from the earthworms

223 At the end of the experiment, earthworms were collected from the VFCWs by layer
224 and analyzed for MPs. The separation method was adopted from Wang et al. (2019).
225 Earthworms were first picked out with the help of tweezers and then washed with
226 distilled water thoroughly to remove the particles stuck to their surface. After that, the

227 earthworms were put on wet filter paper in a clean beaker. The beaker was then put in
228 the thermostatic room (22 ± 1 °C) for 48 h to let the earthworms egest. Then the
229 beakers and earthworms were washed again with distilled water and the resulting
230 liquid fractions were analyzed by the same method as used on the effluent. The
231 earthworms were transferred to another clean beaker and put in an oven at 70 °C to
232 dry off to obtain the dry weight. Then 10% KOH solution was added to the beakers
233 for digestion in a 70 °C oven for 24 h (Kühn et al., 2017). After digestion, the samples
234 were screened by a 40 µm sieve to remove the KOH solution. The residue on the
235 sieve was then transferred into glass beakers with distilled water and the following
236 MPs extraction procedure from water sample was as described in section 2.3.1. The
237 MPs concentration inside earthworms and in excreta of earthworms were expressed in
238 MPs per gram of earthworms (dry weight).

239 2.4 MPs counting

240 The filters which contained MPs were counted under stereomicroscopes with
241 augmentations between x14 and x70 (SZM and SDZ-PL, Kyowa, Japan). The filters
242 were read from left to right, then down one row from right to left and so on (Hidalgo-
243 Ruz et al., 2012). During the counting, MPs were classified into PS, HDPE, PP and
244 microbeads by color.

245 2.5 Quality control and contamination prevention

246 During all steps, including the setting up of CWs, sampling and analysis, materials
247 made of plastic were avoided as much as possible. The filters and beakers containing
248 the sample were always covered with aluminum foil when not being tested, to avoid
249 contamination. Blanks, CW1 (filled with sand) and CW2 (filled with gravel), were
250 run for the same period as the experimental group and fed with tap water. The
251 analyses of MPs in effluent and its distribution were also performed in blanks, and no
252 typical MPs used in this study were found. At the end of the experiment, the recovery
253 rate of MPs in VFCWs was calculated for quality control. Each kind of MPs inside
254 VFCWs, effluent and earthworms were compared with the total MPs in the influent to
255 obtain the recovery rate.

256 2.6 Analytical methods

257 The size of sand was measured by Mastersizer S (Malvern, Netherland) and the size
258 of gravel was measured by mechanical sieving. Chemical oxygen demand (COD) in
259 wastewater and effluent was determined every 5 days and measured by Hach kits
260 (Closed tube digestion, Hach) according to standard methods (APHA, 2005). The

261 humidity and loss on ignition (LOI) were measured by oven and muffle furnace

262 (Touch et al., 2017). Each test was run in duplicate.

263 2.7 Statistical analysis

264 Statistical analysis was performed using SPSS 24 software (SPSS Inc., USA). The

265 significant differences of MPs distribution by layers between the VFCWs were tested

266 by paired t-test and one-way analysis of variance followed by post-hoc (Tukey's) test.

267 The significance level was 0.05 ($p < 0.05$).

268 **3. Results and discussion**

269 3.1 General performance of different VFCWs

270 COD removal by VFCWs presumably indicates the existence of a biofilm inside

271 VFCWs. In this study, the biofilm was designed to develop in sand-filled VFCW5/6

272 and 9/10 by feeding synthetic wastewater. The COD concentration in the influent of

273 VFCW9/10 was a bit higher than VFCW5/6 in order to provide enough organic

274 matter for earthworms. The results are shown in Fig. 2a and 2b; the average removal

275 efficiency of COD in VFCW5/6 and 9/10 was 90.8% and 93.7%, respectively. This

276 indicated the growth and activity of biofilm. The biofilm could also enhance the water

277 retention, lead to higher humidity, and increase volatile content of solids. As shown in

278 Fig. 3, both VFCW5/6 and 9/10 had higher humidity and LOI than VFCW3/4 and 7/8,
279 this also suggested the presence of biofilm.

280 The MPs removal was different between the VFCWs. In sand-filled VFCW3/4 and
281 5/6, no MPs were detected in the effluent and the removal efficiency thus reached
282 100%. In gravel-filled VFCW7/8, MPs were detected in the effluent, suggesting that
283 MPs could penetrate the CWs filled with gravel. MPs were already detected in the
284 first sample of effluent of VFCW7/8 and the average total concentration was 0.2
285 particles/L (Fig. 2c). With the continued operation of the VFCWs, the PS particle
286 concentration in the effluent noticeably increased and reached 4.9 particles/L on day
287 41. Due to the lower relative density, HDPE particle (0.95 g/cm^3) concentrations were
288 much smaller than PS (1.05 g/cm^3) in the effluent. The highest concentration of
289 HDPE in the effluent was 0.6 particles/L. Because of their lowest density of only 0.90
290 g/cm^3 , the PP particles were hardly found in the effluent, and the average
291 concentration of PP in effluent was only 0.07 particles/L. These three kinds of MPs
292 had similar shape and size, thus the relative density could be the major effector on the
293 removal of MPs. Low density MPs have more hindered penetration under infiltration
294 because of the high buoyancy effect.

295 In sand-filled VFCW9/10, earthworms were added, and they influenced the removal
296 of MPs. In the effluent of both VFCW9 and VFCW10, MPs were detected from day
297 25 onwards which was 5 days after adding the earthworms. During the operation of
298 VFCW9 and 10, there was a 50 days suspension of feeding due to the COVID-19
299 pandemic from day 25. But the earthworms inside were still alive until day 75. At the
300 end of the experiment, the MP concentrations in the effluent remained at a very low
301 level. The average concentration of MPs from day 25 to the end was 0.4 particles/L,
302 and the removal efficiency still reached 99.8% (Fig. 2d). Furthermore, only PS
303 particles, HDPE particles and microbeads were detected in the effluent. Similar as
304 gravel-filled VFCWs, PP particles were not found in the effluent due to the low
305 relative density (0.90 g/cm^3). The proportion of microbeads reached 65%, while the
306 proportions of PS and HDPE particles were 30% and 5%.

307 3.2 Influence of biofilm on the fate of MPs

308 VFCW3, 4, 5 and 6 were filled with sand and used to investigate the influence of
309 biofilm on the fate of MPs. VFCW3 and VFCW4 were fed with tap water and the
310 others were fed with synthetic wastewater to develop the biofilm. The movement of
311 MPs in the VFCWs could be influenced by external conditions such as flow and
312 porosity, and MP characteristics such as density, shape and size (Yao et al., 1971). PP,

313 HDPE and PS particles were granular morphotypes, and the microbeads were
314 spherical morphotypes (Table 1). Granular and spherical morphotypes are expected to
315 have better mobility than fragments or filaments (O'Connor et al., 2019). However, in
316 this group, none of the four kinds of MPs fully flowed through the sand filled CWs as
317 in none of the effluents were any MPs detected during the 45 days. According to the
318 distribution analysis (Fig. 4a and 4b), there was no obviously difference in the
319 distribution of PP, HDPE and PS particles between VFCWs with and without biofilm
320 ($p > 0.05$). Most of the MPs were detected in the top 6 cm layer. This agrees quite
321 well with a previous study by O'Connor et al. (2019) who investigated MPs' vertical
322 migration in a sandy soil with wet-dry cycles, and found that the maximum depth that
323 MPs could reach was 7.5 cm after twelve 83 mm infiltration events. In layer 1 (depth
324 between 0 and 3 cm) and 2 (depth between 3 and 6 cm), the average total MPs
325 concentrations in VFCW3/4 without biofilm were 17.2 and 1.9 particles/cm³,
326 respectively. And the average concentrations of the same MPs in VFCW5/6 with
327 biofilm were 17.8 and 1.7 particles/cm³, respectively. The slight difference between
328 VFCW3/4 and VFCW5/6 could be caused by the biofilm (Vymazal, 2018), which
329 slightly enhanced the filtering capability in the first layer. However, in the deeper
330 layer 3 (depth between 6 and 10 cm) in VFCW5/6 with biofilm, the microbeads

331 concentration was 0.096 particles/cm³ and higher than PP, HDPE and PS ($p < 0.05$).

332 The major differences between microbeads and others were the particle size and

333 morphotype. The size of microbeads was 45-53 μm which was much smaller than the

334 other MPs used (75-425 μm). Recent studies also showed that smaller sized MPs had

335 higher permeability (Rillig et al., 2017a).

336 The result showed that CWs filled with sand had a great removal capacity for MPs

337 and that the removal efficiency for these four kinds of MPs reached 100%. The

338 biofilm slightly enhanced the filtering capacity of CWs, leading to a tiny difference of

339 MPs distribution. MPs were mostly retained in the first 6 cm. Nevertheless, due to the

340 smaller size and spherical morphotype, microbeads had the highest concentration in

341 the layer of 6-10 cm. O'Connor et al. (2019) also reported that the PE particles with an

342 average size of 21 μm they used for the vertical migration experiments were the most

343 mobile of the MPs tested, travelling to a maximum depth of 7.5 cm.

344 3.3 Influence of media type on the fate of MPs

345 Gravel-filled VFCW7 and 8 were used to investigate the influence of media type on

346 the fate of MPs. The VFCWs were fed with tap water. The distribution of MPs in

347 gravel-filled VFCWs was highly different from sand-filled VFCWs ($p < 0.05$), with

348 PS, HDPE and PP particles being distributed throughout the VFCWs. Layer 3 (depth
349 between 6 and 10 cm) retained the most MPs, with an average total concentration of
350 3.98 particles/cm³ (Fig. 4c). In the top 30 cm layer, the concentrations of PP, HDPE
351 and PS particles were similar. On the contrary, in the lower layer (50-80 cm), the
352 concentration of PS particles was higher than PP and HDPE ($p < 0.05$). In the deepest
353 layer, the concentration of PS was 0.081 particles/cm³, while the concentrations of
354 HDPE and PP were 0.032 and 0.024 particles/cm³, respectively. These results are
355 consistent with the results of concentrations in the effluent. Therefore, relative density
356 is still the major factor influencing the distribution of MPs, and the particles with low
357 relative density (lower than water) could be retained to a higher degree in VFCWs.
358 Compared with the VFCWs filled with sand, the gravel-filled VFCWs have a low
359 capacity to retain the MPs, and high-density MPs more easily passed the VFCWs. In
360 real-life applications, gravel-filled VFCWs usually have a dense sludge layer on top,
361 known as French-style CWs which was reported to have a high capacity for TSS
362 removal (Milot et al., 2016). Therefore, it is reasonable to assume that the sludge
363 layer can enhance the MPs retention capacity of gravel-filled VFCWs.
364 The MPs were distributed throughout the VFCWs by water percolation, but the lowest
365 removal efficiency of MPs for gravel-filled VFCWs still surmounted 96%. The result

366 indicated that the diameter of wetland media also played an important role in
367 distribution and removal of MPs. A previous study also reported the effect of porous
368 media diameter on filtration (Tarbuck et al., 2005). The filtering effect happens when
369 the pore spaces of medium are too narrow to allow the particles to pass.
370 Consequently, gravel had a much higher average diameter than sand, which increased
371 the depth of penetration of MPs (Crittenden et al., 2012), leading to a reduction of
372 retention capacity of VFCWs.

373 3.4 Influence of earthworm activity on the fate of MPs

374 VFCW9 and 10 were filled with sand and populated with earthworms. These VFCWs
375 were used to investigate the influence of earthworm activity on the fate of MPs. The
376 distribution of MPs inside the VFCWs was also changed due to the activities of the
377 earthworms. The maximum depth at which MPs were detected in the sand was
378 increased to layer 4 (depth between 10 and 15 cm), and the average total
379 concentration of MPs was 0.27 particles/cm³ (Fig. 4d). MPs were not detected in the
380 sand of deeper layers (15-80 cm) but shown in the effluent, which could be due to the
381 extremely small concentrations of MPs transported by earthworms. However, the MPs
382 concentrations in layer 2 (depth between 3 and 6 cm) and 3 (depth between 6 and 10
383 cm) also increased a lot compared to VFCW5/6 which were also fed with synthetic

384 wastewater but without earthworms. The average total MPs concentration of
385 VFCW9/10 in layer 2 was 5.57 particles/cm³, which was about 2 times higher than the
386 average of VFCW5/6. Likewise, the average total concentration of MPs in layer 3 was
387 1.31 particles/cm³, which was 10 times higher than the average of VFCW5/6.

388 Apparently, the earthworms increased the mobility of MPs considerably. A recent
389 study also reported the transport of MPs by earthworms in soil (Rillig et al., 2017b).
390 They used PE particles for a transportation experiment, and the results showed that
391 earthworms could transport them from the surface down to a depth of 10 cm. In this
392 study, the depth of transport was higher and reached 15 cm. On the one hand, when
393 earthworms move in the CWs, they could create numerous tunnels (Huerta Lwanga et
394 al., 2017; Yang et al., 2019). With the tunnels, the MPs could move down more easily
395 with the infiltrating water. On the other hand, the macroinvertebrates could ingest the
396 MPs and deposit them elsewhere when defecating. Also, the MPs could attach to the
397 body of macroinvertebrates and thus be transported (Rillig et al., 2017b).

398 The distribution of earthworms was different between VFCW9 and 10. In VFCW9,
399 18, 1, 0, 0, 2, 0, 17 earthworms were found at a depth of 0-3, 3-6, 6-10, 10-15, 15-30,
400 30-50, 50-80 cm, respectively (Fig. 5). However, no earthworms were found in the
401 bottom layer of CW10. This partial migration of earthworms in CW9 may be caused

402 by the effect of humidity and oxygen (Chuang and Chen, 2008). During the 50 days
403 suspension of feeding caused by COVID-19 pandemic, the bottom layer could
404 provide earthworms with the same moisture and oxygen to survive as the top layer. In
405 VFCW9, the earthworms were mainly distributed in the first layer and the last layer. It
406 suggested that they could move throughout the VFCWs from top to bottom. The
407 results further showed that MPs were present in the excreta of earthworms (Fig. S2),
408 and the concentrations were 40.00, 68.26, 30.49 and 10.55 particles/g dw at the depths
409 of 0-3, 3-6, 15-30 and 50-80 cm, respectively. The presence of MPs in the excreta
410 directly confirmed the ingestion of MPs by earthworms. The concentrations of MPs in
411 excreta of earthworms in the upper layer were higher than in the lower layer owing to
412 the high abundance of MPs in the upper layer. According to the distribution of MPs
413 inside the VFCWs, there was no MP detected in the layer between 50 and 80 cm, but
414 the MPs in the excreta found in this layer reached 10.55 particles/g dw. This
415 suggested that earthworms could carry the MPs from the top to the bottom. The
416 distribution of earthworms was different in VFCW10. There were 15, 8, 10 and 4
417 earthworms at the depths of 0-3, 3-6, 6-10 and 10-15 cm, respectively. No earthworm
418 was found in the deeper layers. The MP concentrations in the excreta of earthworms
419 were 25.30, 18.62, 20.85 and 21.44 particles/g dw. The highest concentration was also

420 found in the top layer which was due to the high abundance of MPs. Both in VFCW9
421 and 10, no MPs were detected in the digestion residue of earthworms after egestion
422 for 48 h. This suggested that MPs might be egested completely from the gut of
423 earthworms. Previous studies also confirmed the ingestion and excretion of MPs by
424 earthworms (Huerta Lwanga et al., 2017; Wang et al., 2019), and observed the
425 transportation of MPs. With the presence of earthworms, the distribution of MPs in
426 VFCWs changed considerably when compared to VFCW5/6 without earthworms.
427 Also, the removal efficiency slightly decreased from 100% to 99.8%, which could be
428 caused by the migration of earthworms. This indicates that the bioturbation of
429 earthworms could reduce the MPs removal by VFCWs.

430 In addition, the total alive earthworms collected at the end of the experiment were 38
431 and 37 in VFCW9 and 10, respectively. The earthworm population had declined a bit
432 as a few might have escaped from the bottom of the VFCWs. No reproduction
433 occurred. Previous studies also reported that MPs had no significant effects on
434 survival of earthworms (*Lumbricus terrestris* and *Eisenia andrei Bouché*) (Huerta
435 Lwanga et al., 2016; Rodriguez-Sejjo et al., 2017), but their growth rate was
436 significantly reduced after 60 days of exposure to MPs (28, 45 and 60% dw/ dw), and
437 tissue injury was induced (Huerta Lwanga et al., 2016). This is consistent with the

438 result of this experiment; the survival of earthworms had not been affected. But the
439 effect on tissue injury and reproduction of earthworms needs to be further
440 investigated.

441 The results convincingly showed that activities of earthworms could considerably
442 influence the distribution of MPs inside VFCWs ($p < 0.05$), but the MPs removal
443 efficiency of VFCWs was stable at 99.8%. MPs could reach deeper layer (10-15 cm)
444 when compared to the same type of VFCWs (5/6) without earthworms. However, the
445 MPs were still most abundant in the first 10 cm layer, and the concentration of MPs in
446 the first 10 cm layer was 98.8% of the total concentration inside VFCWs. In this case,
447 only the first 10 cm layer of VFCWs would be highly polluted with MPs. When
448 needing to manage the wetland media because of clogging, a special management
449 would thus be required to avoid the dispersion of MPs from the first 10 cm layer
450 solids. Also, it has been reported that microorganisms such as bacteria and fungi can
451 degrade MPs (Yuan et al., 2020), the abundant MPs and organic matter in the top
452 layer (0-10 cm) could favor degradation.

453 3.5 MPs balance

454 Based on the distribution of MPs inside the VFCWs and their respective effluent
455 concentrations, the total amount of MPs of each VFCW was estimated and compared
456 with the total MPs in the influent to ensure the reliability of the data. As shown in
457 Table 3, all the MPs, except the microbeads in sand-filled VFCW9/10 with
458 earthworms, reached a recovery rate more than 89%. The difference may be due to the
459 uneven distribution of MPs in the solids. However, in the sand-filled VFCW9/10 with
460 earthworms, the recovery rate of microbeads only reached 78%, while this rate was
461 90% in sand-filled VFCW5/6 with biofilm. This decrease could be related to the
462 activities of earthworms. This can be explained as follows. First, the size of
463 microbeads ingested by earthworms was very small (only 45-53 μm), so they could
464 have served as the nuclei for the aggregates in earthworm casts (Shipitalo and Protz,
465 1989). In this case, the microbeads could be missed in the analysis due to their
466 invisibility. The second possibility is the efficiency of the applied KOH digestion
467 method. The residue after digestion still wrapped this small size microbeads which
468 made it difficult to detect. Overall, the recovery rate of MPs reached a considerable
469 degree, which supports the reliability of the analyzed data.

470 **4. Conclusion**

471 In this study, the effects of biofilm, media type and earthworms on the fate and
472 removal of MPs in unplanted lab-scale VFCWs were investigated over a time span of
473 45 days. The sand-filled VFCWs had a high removal capacity for MPs. MPs were
474 retained in the first 10 cm, and the removal efficiency was 100%. The growth of
475 biofilm slightly enhanced the filtering capacity of VFCWs, leading to a tiny
476 difference in MPs distribution. When gravel was used to fill the VFCWs, the removal
477 efficiency of MPs was 96%, and the MPs were distributed throughout the VFCWs. In
478 the presence of earthworms, the maximum depth that MPs reached in VFCWs
479 increased from 6-10 cm to 10-15cm, while the MP concentrations at the depth of 3-6
480 cm and 6-10 cm respectively increased 2 and 10 times compared to the same type
481 VFCWs without earthworms. Although MPs were not found in the sand of deep
482 layers (15-80 cm), MPs were found in the excreta of earthworms present in the
483 bottom layer and also in the effluent, but the removal efficiency of MPs was still
484 99.8%. Also, the concentration of MPs in the first 10 cm layer was still 98.8% of the
485 total MPs concentration in VFCWs with earthworms. Accumulation of large
486 quantities of MPs in the first 10 cm layer of sand-filled VFCWs could be expected
487 after a long run, which should be properly managed to avoid the dispersion of MPs.

488 It should be noted that other important factors in full-scale CWs were not considered
489 in this study, such as the vegetation, soil composition, hydraulic loading rate and flow
490 patterns. These are critical parameters in CWs and expected to affect the fate and
491 removal of MPs. Likewise, natural weather (e.g. rain and wind), plant root system and
492 bioturbation from abundant organisms were missing in lab-scale VFCWs. We
493 recommend that further studies could consider more factors from the real-world, there
494 is still a large knowledge gap in real-scale CWs. Also, various shapes and more types
495 of MPs warrant further research in this regard.

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506 **References**

- 507 APHA. Standard methods for the examination of water and wastewater. Washington
508 DC: American Public Health Association/American Water Works
509 Association/Water Environment Federation, 2005.
- 510 Arthur C, Baker J, Bamford HA. Proceedings of the International Research Workshop
511 on the Occurrence, Effects, and Fate of Microplastic Marine Debris,
512 September 9-11, 2008, University of Washington Tacoma, Tacoma, WA,
513 USA, 2009.
- 514 Ben-David EA, Habibi M, Haddad E, Hasanin M, Angel DL, Booth AM, et al.
515 Microplastic distributions in a domestic wastewater treatment plant: Removal
516 efficiency, seasonal variation and influence of sampling technique. *Science of
517 The Total Environment* 2021; 752: 141880.
518 <https://doi.org/10.1016/j.scitotenv.2020.141880>.
- 519 Chuang S-C, Chen JH. Role of diurnal rhythm of oxygen consumption in emergence
520 from soil at night after heavy rain by earthworms. *Invertebrate Biology* 2008;
521 127: 80-86. <https://doi.org/10.1111/j.1744-7410.2007.00117.x>.
- 522 Cole M, Lindeque P, Halsband C, Galloway TS. Microplastics as contaminants in the
523 marine environment: A review. *Marine Pollution Bulletin* 2011; 62: 2588-
524 2597. <https://doi.org/10.1016/j.marpolbul.2011.09.025>.
- 525 Crittenden JC, Trussell RR, Hand DW, Howe KJ, Tchobanoglous G. Granular
526 Filtration. *MWH's Water Treatment: Principles and Design, Third Edition*,
527 2012, pp. 727-818. <https://doi.org/10.1002/9781118131473.ch11>.
- 528 De Falco F, Gullo MP, Gentile G, Di Pace E, Cocca M, Gelabert L, et al. Evaluation
529 of microplastic release caused by textile washing processes of synthetic
530 fabrics. *Environmental Pollution* 2018; 236: 916-925.
531 <https://doi.org/10.1016/j.envpol.2017.10.057>.
- 532 de Sá LC, Luís LG, Guilhermino L. Effects of microplastics on juveniles of the
533 common goby (*Pomatoschistus microps*): Confusion with prey, reduction of
534 the predatory performance and efficiency, and possible influence of
535 developmental conditions. *Environmental Pollution* 2015; 196: 359-362.
536 <https://doi.org/10.1016/j.envpol.2014.10.026>.
- 537 Della Torre C, Bergami E, Salvati A, Faleri C, Cirino P, Dawson KA, et al.
538 Accumulation and Embryotoxicity of Polystyrene Nanoparticles at Early Stage
539 of Development of Sea Urchin Embryos *Paracentrotus lividus*. *Environmental
540 Science & Technology* 2014; 48: 12302-12311.
541 <https://doi.org/10.1021/es502569w>.

542 Dotro G, Langergraber G, Molle P, Nivala J, Puigagut J, Stein O, et al. Treatment
543 Wetlands: IWA Publishing, 2017. <https://doi.org/10.2166/9781780408774>.

544 Estahbanati S, Fahrenfeld NL. Influence of wastewater treatment plant discharges on
545 microplastic concentrations in surface water. *Chemosphere* 2016; 162: 277-
546 284. <https://doi.org/10.1016/j.chemosphere.2016.07.083>.

547 Fendall LS, Sewell MA. Contributing to marine pollution by washing your face:
548 Microplastics in facial cleansers. *Marine Pollution Bulletin* 2009; 58: 1225-
549 1228. <https://doi.org/10.1016/j.marpolbul.2009.04.025>.

550 Hidalgo-Ruz V, Gutow L, Thompson RC, Thiel M. Microplastics in the Marine
551 Environment: A Review of the Methods Used for Identification and
552 Quantification. *Environmental Science & Technology* 2012; 46: 3060-3075.
553 <https://doi.org/10.1021/es2031505>.

554 Hoagland DR, Arnon DI. The water-culture method for growing plants without soil.
555 *Circular*. California Agricultural Experiment Station 1950; 347: 32 pp.

556 Huerta Lwanga E, Gertsen H, Gooren H, Peters P, Salánki T, van der Ploeg M, et al.
557 Microplastics in the Terrestrial Ecosystem: Implications for *Lumbricus*
558 *terrestris* (Oligochaeta, Lumbricidae). *Environmental Science & Technology*
559 2016; 50: 2685-2691. <https://doi.org/10.1021/acs.est.5b05478>.

560 Huerta Lwanga E, Gertsen H, Gooren H, Peters P, Salánki T, van der Ploeg M, et al.
561 Incorporation of microplastics from litter into burrows of *Lumbricus terrestris*.
562 *Environmental Pollution* 2017; 220: 523-531.
563 <https://doi.org/10.1016/j.envpol.2016.09.096>.

564 Ivar do Sul JA, Costa MF. The present and future of microplastic pollution in the
565 marine environment. *Environmental Pollution* 2014; 185: 352-364.
566 <https://doi.org/10.1016/j.envpol.2013.10.036>.

567 Kühn S, van Werven B, van Oyen A, Meijboom A, Bravo Rebolledo EL, van
568 Franeker JA. The use of potassium hydroxide (KOH) solution as a suitable
569 approach to isolate plastics ingested by marine organisms. *Marine Pollution*
570 *Bulletin* 2017; 115: 86-90. <https://doi.org/10.1016/j.marpolbul.2016.11.034>.

571 Li D, Shi Y, Yang L, Xiao L, Kehoe DK, Gun'ko YK, et al. Microplastic release from
572 the degradation of polypropylene feeding bottles during infant formula
573 preparation. *Nature Food* 2020; 1: 746-754. <https://doi.org/10.1038/s43016-020-00171-y>.

574

575 Li H-X, Getzinger GJ, Ferguson PL, Orihuela B, Zhu M, Rittschof D. Effects of
576 Toxic Leachate from Commercial Plastics on Larval Survival and Settlement
577 of the Barnacle *Amphibalanus amphitrite*. *Environmental Science &*
578 *Technology* 2016; 50: 924-931. <https://doi.org/10.1021/acs.est.5b02781>.

579 Li YK, Liu YZ, Li GB, Xu TW, Liu HS, Ren SM, et al. Surface topographic
580 characteristics of suspended particulates in reclaimed wastewater and effects
581 on clogging in labyrinth drip irrigation emitters. *Irrigation Science* 2012; 30:
582 43-56. <https://doi.org/10.1007/s00271-010-0257-x>.

583 Mason SA, Garneau D, Sutton R, Chu Y, Ehmann K, Barnes J, et al. Microplastic
584 pollution is widely detected in US municipal wastewater treatment plant
585 effluent. *Environmental Pollution* 2016; 218: 1045-1054.
586 <https://doi.org/10.1016/j.envpol.2016.08.056>.

587 McCormick A, Hoellein TJ, Mason SA, Schlupe J, Kelly JJ. Microplastic is an
588 Abundant and Distinct Microbial Habitat in an Urban River. *Environmental
589 Science & Technology* 2014; 48: 11863-11871.
590 <https://doi.org/10.1021/es503610r>.

591 Millot Y, Troesch S, Esser D, Molle P, Morvannou A, Gourdon R, et al. Effects of
592 design and operational parameters on ammonium removal by single-stage
593 French vertical flow filters treating raw domestic wastewater. *Ecological
594 Engineering* 2016; 97: 516-523. <https://doi.org/10.1016/j.ecoleng.2016.10.002>.

595 Murphy F, Ewins C, Carbonnier F, Quinn B. Wastewater Treatment Works (WwTW)
596 as a Source of Microplastics in the Aquatic Environment. *Environmental
597 Science & Technology* 2016; 50: 5800-5808.
598 <https://doi.org/10.1021/acs.est.5b05416>.

599 O'Connor D, Pan S, Shen Z, Song Y, Jin Y, Wu W-M, et al. Microplastics undergo
600 accelerated vertical migration in sand soil due to small size and wet-dry
601 cycles. *Environmental Pollution* 2019; 249: 527-534.
602 <https://doi.org/10.1016/j.envpol.2019.03.092>.

603 Redondo-Hasselerharm PE, Falahudin D, Peeters ETHM, Koelmans AA. Microplastic
604 Effect Thresholds for Freshwater Benthic Macroinvertebrates. *Environmental
605 Science & Technology* 2018; 52: 2278-2286.
606 <https://doi.org/10.1021/acs.est.5b05416>.

607 Ribeiro F, Garcia AR, Pereira BP, Fonseca M, Mestre NC, Fonseca TG, et al.
608 Microplastics effects in *Scrobicularia plana*. *Marine Pollution Bulletin* 2017;
609 122: 379-391. <https://doi.org/10.1016/j.marpolbul.2017.06.078>.

610 Rillig MC. Microplastic in Terrestrial Ecosystems and the Soil? *Environmental
611 Science & Technology* 2012; 46: 6453-6454.
612 <https://doi.org/10.1021/es302011r>.

613 Rillig MC, Ingraffia R, de Souza Machado AA. Microplastic incorporation into soil in
614 agroecosystems. *Frontiers in plant science* 2017a; 8: 1805.
615 <https://doi.org/10.3389/fpls.2017.01805>.

616 Rillig MC, Ziersch L, Hempel S. Microplastic transport in soil by earthworms.
617 Scientific Reports 2017b; 7: 1362. [https://doi.org/10.1038/s41598-017-01594-](https://doi.org/10.1038/s41598-017-01594-7)
618 [7](https://doi.org/10.1038/s41598-017-01594-7).

619 Rodriguez-Seijo A, Lourenço J, Rocha-Santos TAP, da Costa J, Duarte AC, Vala H,
620 et al. Histopathological and molecular effects of microplastics in Eisenia
621 andrei Bouché. Environmental Pollution 2017; 220: 495-503.
622 <https://doi.org/10.1016/j.envpol.2016.09.092>.

623 Rousseau DPL, Lesage E, Story A, Vanrolleghem PA, De Pauw N. Constructed
624 wetlands for water reclamation. Desalination 2008; 218: 181-189.
625 <https://doi.org/10.1016/j.desal.2006.09.034>.

626 Shipitalo MJ, Protz R. Chemistry and micromorphology of aggregation in earthworm
627 casts. Geoderma 1989; 45: 357-374. [https://doi.org/10.1016/0016-](https://doi.org/10.1016/0016-7061(89)90016-5)
628 [7061\(89\)90016-5](https://doi.org/10.1016/0016-7061(89)90016-5).

629 Shumway DL, Koide RT. Seed preferences of Lumbricus terrestris L. Applied Soil
630 Ecology 1994; 1: 11-15. [https://doi.org/10.1016/0929-1393\(94\)90019-1](https://doi.org/10.1016/0929-1393(94)90019-1).

631 Silva C, Quadros S, Ramalho P, Alegre H, Rosa MJ. Translating removal efficiencies
632 into operational performance indices of wastewater treatment plants. Water
633 Research 2014; 57: 202-214. <https://doi.org/10.1016/j.watres.2014.03.025>.

634 Smith M, Love DC, Rochman CM, Neff RA. Microplastics in Seafood and the
635 Implications for Human Health. Current Environmental Health Reports 2018;
636 5: 375-386. <https://doi.org/10.1007/s40572-018-0206-z>.

637 Stolte A, Forster S, Gerdt G, Schubert H. Microplastic concentrations in beach
638 sediments along the German Baltic coast. Marine Pollution Bulletin 2015; 99:
639 216-229. <https://doi.org/10.1016/j.marpolbul.2015.07.022>.

640 Sun J, Dai X, Wang Q, van Loosdrecht MCM, Ni B-J. Microplastics in wastewater
641 treatment plants: Detection, occurrence and removal. Water Research 2019;
642 152: 21-37. <https://doi.org/10.1016/j.watres.2018.12.050>.

643 Tarbuck EJ, Lutgens FK, Tasa D, Linneman S. Earth: an introduction to physical
644 geology: Pearson/Prentice Hall Upper Saddle River, 2005.

645 Thompson RC, Olsen Y, Mitchell RP, Davis A, Rowland SJ, John AW, et al. Lost at
646 sea: where is all the plastic? Science(Washington) 2004; 304: 838.
647 <https://doi.org/10.1126/science.1094559>.

648 Touch N, Hibino T, Takata H, Yamaji S. Loss on Ignition-Based Indices for
649 Evaluating Organic Matter Characteristics of Littoral Sediments. Pedosphere
650 2017; 27: 978-984. [https://doi.org/10.1016/S1002-0160\(17\)60487-9](https://doi.org/10.1016/S1002-0160(17)60487-9).

651 Vymazal J. Does clogging affect long-term removal of organics and suspended solids
652 in gravel-based horizontal subsurface flow constructed wetlands? Chemical

653 Engineering Journal 2018; 331: 663-674.
654 <https://doi.org/10.1016/j.cej.2017.09.048>.

655 Wang J, Coffin S, Sun C, Schlenk D, Gan J. Negligible effects of microplastics on
656 animal fitness and HOC bioaccumulation in earthworm *Eisenia fetida* in soil.
657 Environmental Pollution 2019; 249: 776-784.
658 <https://doi.org/10.1016/j.envpol.2019.03.102>.

659 Wang Q, Hernández-Crespo C, Santoni M, Van Hulle S, Rousseau DPL. Horizontal
660 subsurface flow constructed wetlands as tertiary treatment: Can they be an
661 efficient barrier for microplastics pollution? Science of The Total
662 Environment 2020; 721: 137785.
663 <https://doi.org/10.1016/j.scitotenv.2020.137785>.

664 Wei S, Luo H, Zou J, Chen J, Pan X, Rousseau DPL, et al. Characteristics and
665 removal of microplastics in rural domestic wastewater treatment facilities of
666 China. Science of The Total Environment 2020; 739: 139935.
667 <https://doi.org/10.1016/j.scitotenv.2020.139935>.

668 Yang X, Lwanga EH, Bemani A, Gertsen H, Salanki T, Guo X, et al. Biogenic
669 transport of glyphosate in the presence of LDPE microplastics: A mesocosm
670 experiment. Environmental Pollution 2019; 245: 829-835.
671 <https://doi.org/10.1016/j.envpol.2018.11.044>.

672 Yao K-M, Habibian MT, O'Melia CR. Water and waste water filtration. Concepts and
673 applications. Environmental Science & Technology 1971; 5: 1105-1112.
674 <https://pubs.acs.org/doi/abs/10.1021/es60058a005>.

675 Yuan J, Ma J, Sun Y, Zhou T, Zhao Y, Yu F. Microbial degradation and other
676 environmental aspects of microplastics/plastics. Science of The Total
677 Environment 2020; 715: 136968.
678 <https://doi.org/10.1016/j.scitotenv.2020.136968>.

679 Zhao L, Zhu W, Tong W. Clogging processes caused by biofilm growth and organic
680 particle accumulation in lab-scale vertical flow constructed wetlands. Journal
681 of Environmental Sciences 2009; 21: 750-757. [https://doi.org/10.1016/S1001-](https://doi.org/10.1016/S1001-0742(08)62336-0)
682 [0742\(08\)62336-0](https://doi.org/10.1016/S1001-0742(08)62336-0).

683

1 TABLES AND FIGURES

2 Table 1 Characteristics of used MPs. PS, HDPE and PP granules were home-made,

3 the PE spheres were purchased from Cospheric (California, US).

Polymer	PS	HDPE	PP	PE
Size (μm)	75-425	75-425	75-425	45-53
Color	Red	Blue	Green	Orange
Morphotype	Granular	Granular	Granular	Sphere
Density (g/cm^3)	1.05	0.95	0.90	1.00

4 Table 2 Experimental design.

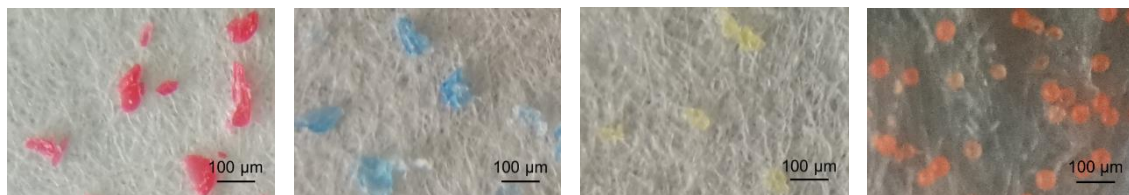
Number	Media type	Influent	MPs	Earthworms
VFCW1	Sand	Tap water	-	-
VFCW2	Gravel	Tap water	-	-
VFCW3	Sand	Tap water	PS, PP, HDPE	-
VFCW4	Sand	Tap water	PS, PP, HDPE	-
VFCW5	Sand	Synthetic wastewater	PS, PP, HDPE, Microbeads	-
VFCW6	Sand	Synthetic wastewater	PS, PP, HDPE, Microbeads	-
VFCW7	Gravel	Tap water	PS, PP, HDPE	-
VFCW8	Gravel	Tap water	PS, PP, HDPE	-
VFCW9	Sand	Synthetic wastewater	PS, PP, HDPE, Microbeads	40 individuals added
VFCW10	Sand	Synthetic wastewater	PS, PP, HDPE, Microbeads	40 individuals added

5

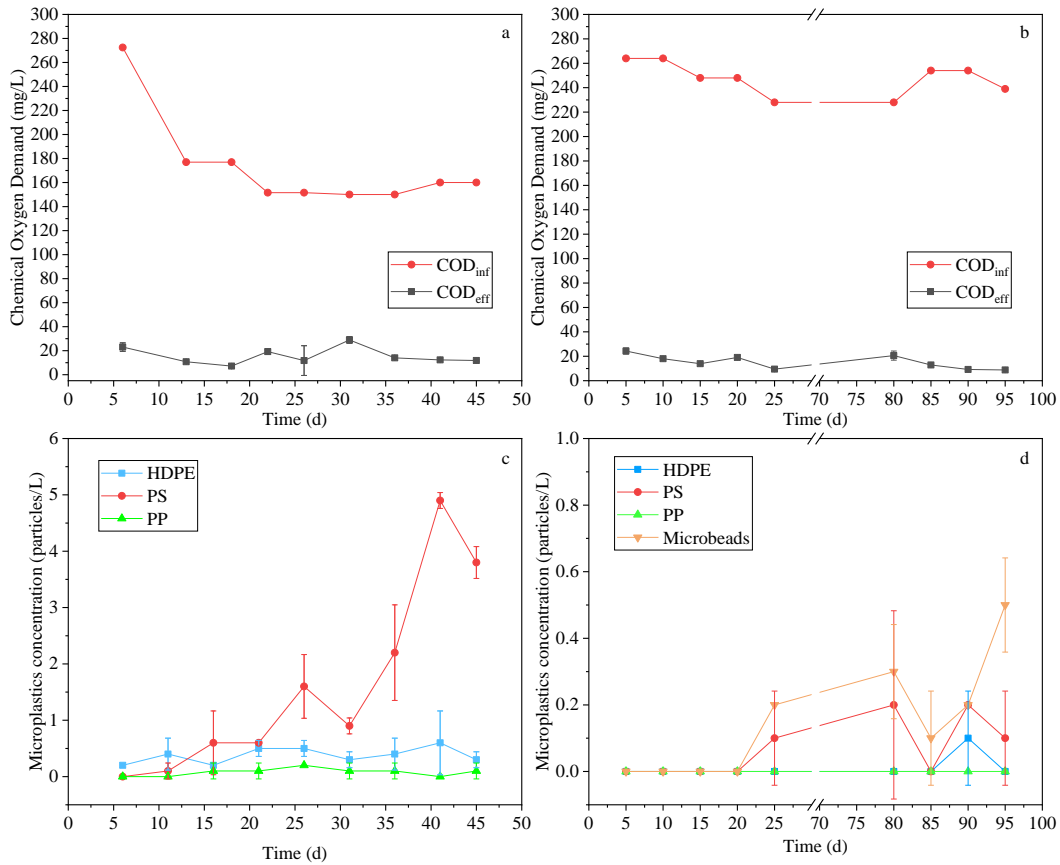
6 Table 3 The average recovery rates (average \pm stand error) of different MPs in
 7 VFCWs, the recovery rates were estimated by the MPs in the influent, effluent, solids
 8 and earthworms.

CWs	PS (%)	HDPE (%)	PP (%)	Microbeads (%)
Sand-filled VFCWs without biofilm	99.5 \pm 0.5	95.0 \pm 0.4	90.3 \pm 1.0	-
Sand-filled VFCWs with biofilm	101.5 \pm 1.2	98.7 \pm 1.2	91.4 \pm 4.2	90.0 \pm 2.3
Gravel-filled VFCWs	98.4 \pm 6.1	90.3 \pm 9.0	89.1 \pm 8.1	-
Sand-filled VFCWs with earthworms	102.3 \pm 2.2	99.4 \pm 4.1	94.0 \pm 8.3	78.1 \pm 4.9

9
 10



11 a b c d
 12 Fig. 1. Stereomicroscopic photographic images of the four MPs used, (a) PS, (b)
 13 HDPE, (c) PP, (d) Microbeads.



14

15 Fig. 2. (a) COD concentration in the influent and effluent of sand-filled VFCW5/6, (b)

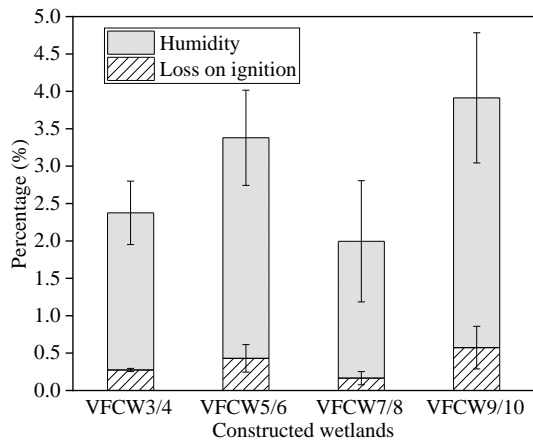
16 COD concentration in the influent and effluent of sand-filled VFCW9/10, (c) MPs

17 concentration in the effluent of gravel-filled VFCW7/8, (d) MPs concentration in the

18 effluent of sand-filled VFCW9/10. The concentration of each kind of MPs in influent

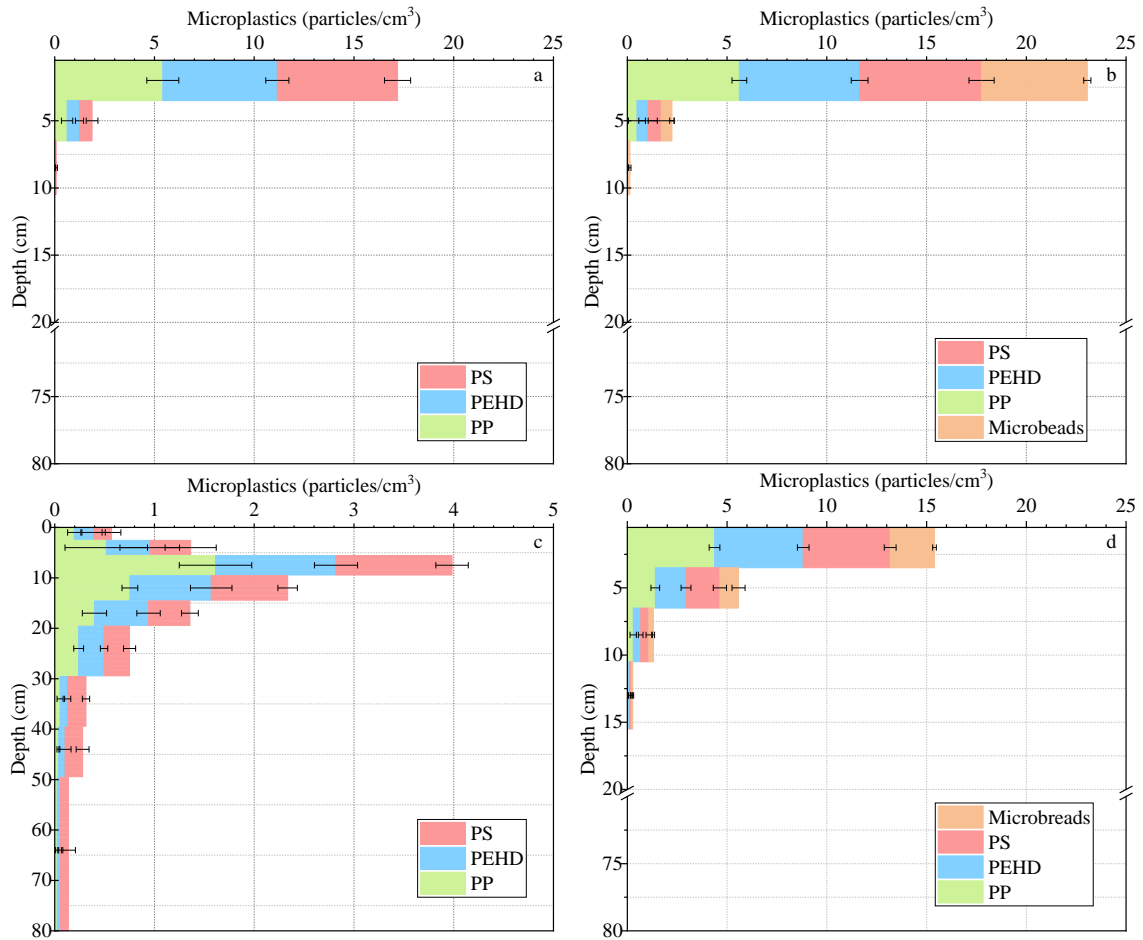
19 was 50 particles/L per day.

20



21

22 Fig. 3. Humidity and Loss on ignition of VFCWs.

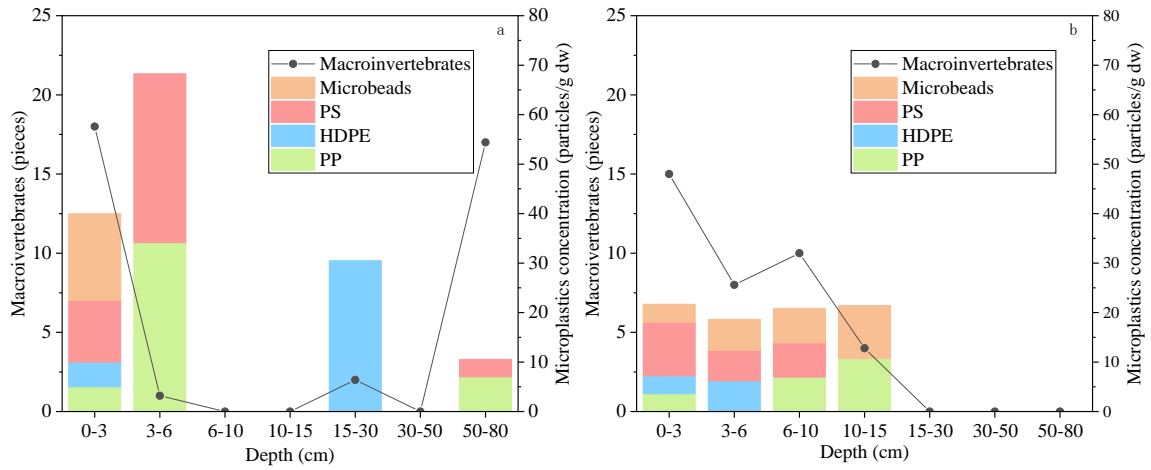


23

24 Fig. 4. Distribution of MPs inside VFCWs, (a) sand-filled VFCW3/4 without biofilm,

25 (b) sand-filled VFCW5/6 with biofilm, (c) gravel-filled VFCW7/8, (d) sand-filled

26 VFCW9/10 with earthworms.

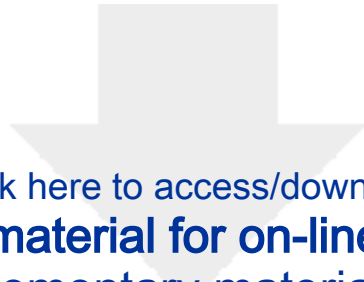


27

28 Fig. 5. Distribution of earthworms and MPs content in the excreta of earthworms in

29 sand-filled VFCWs with earthworms, (a) VFCW9, (b) VFCW10.

30



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Declaration of interests

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

The authors declare the following financial interests/personal relationships which may be considered as potential competing interests: