



High-Grade Glioma Treatment Response Monitoring Biomarkers: A Position Statement on the Evidence Supporting the Use of Advanced MRI Techniques in the Clinic, and the Latest Bench-to-Bedside Developments. Part 1: Perfusion and Diffusion Techniques

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Objective: Summarize evidence for use of advanced MRI techniques as monitoring biomarkers in the clinic, and highlight the latest bench-to-bedside developments.

Methods: Experts in advanced MRI techniques applied to high-grade glioma treatment response assessment convened through a European framework. Current evidence regarding the potential for monitoring biomarkers in adult high-grade glioma is reviewed, and individual modalities of perfusion, permeability, and microstructure imaging are discussed (in Part 1 of two). In Part 2, we discuss modalities related to metabolism and/or chemical composition, appraise the clinic readiness of the individual modalities, and consider post-processing methodologies involving the combination of MRI approaches (multiparametric imaging) or machine learning (radiomics).

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Results: High-grade glioma vasculature exhibits increased perfusion, blood volume, and permeability compared with normal brain tissue. Measures of cerebral blood volume derived from dynamic susceptibility contrast-enhanced MRI have consistently provided information about brain tumor growth and response to treatment; it is the most clinically validated advanced technique. Clinical studies have proven the potential of dynamic contrast-enhanced MRI for distinguishing post-treatment related effects from recurrence, but the optimal acquisition protocol, mode of analysis, parameter of highest diagnostic value, and optimal cut-off points remain to be established. Arterial spin labeling techniques do not require the injection of a contrast agent, and repeated measurements of cerebral blood flow can be performed. The absence of potential gadolinium deposition effects allows widespread use in pediatric patients and those with impaired renal function. More data are necessary to establish clinical validity as monitoring biomarkers. Diffusionweighted imaging, apparent diffusion coefficient analysis, diffusion tensor or kurtosis imaging, intravoxel incoherent motion, and other microstructural modeling approaches also allow treatment response assessment; more robust data are required to validate these alone or when applied to post-processing methodologies.

Conclusion: Considerable progress has been made in the development of these monitoring biomarkers. Many techniques are in their infancy, whereas others have generated a larger body of evidence for clinical application.

Keywords: magnetic resonance imaging, glioma, perfusion, diffusion, pseudoprogression, monitoring biomarkers, glioblastoma, high-grade glioma

1 INTRODUCTION

High-grade gliomas (adult-type diffuse WHO grade 3 and 4 gliomas) account for up to 85% of all new cases of malignant primary brain tumors diagnosed every year, with an incidence of approximately 5/100,000 person years in Europe and North America (1). Of these, approximately 70% are either "glioblastomas, isocitrate dehydrogenase (IDH)-wildtype" or "astrocytoma, IDH-mutant, grade 4" (2). Approximately 15% are "astrocytoma, IDH-mutant, grade 3" and approximately 10% are "oligodendroglioma, IDH-mutant and (chromosome) 1p/ 19q deleted, grade 3." Malignant primary brain tumors cause the greatest number of years of life lost than any other cancer (3). Grade 4 glioma is particularly devastating: The median survival without any treatment is less than six months and with standardof-care treatment is only 14.6 months (4). In adults aged up to 70 years with good performance status, maximal safe tumor resection followed by radiotherapy with concomitant and adjuvant temozolomide has been recommended as the standard-of-care treatment for grade 4 glioma since 2005 (4-6).

During and after treatment, "monitoring biomarkers" are measured serially and are required to detect any change in the extent of glioma infiltration or provide evidence of treatment response (7). Magnetic resonance imaging (MRI) is particularly useful in determining treatment response as it is noninvasive and captures the entire tumor volume as well as adjacent tissues. Furthermore, in nearly all high-grade gliomas, the integrity of the blood-brain barrier (BBB) is disrupted. Following intravenous administration of gadolinium-based contrast agents (GBCA), the hydrophilic contrast molecules diffuse out of the vessel lumen and accumulate within the extravascular extracellular space, manifesting as contrast-enhancing hyperintense regions on T_1 weighted sequences (8). Subsequently, MRI has been incorporated into recommendations for determining treatment response in clinical trials (9). In these recommendations, treatment response assessment is based on simple linear metrics of contrast-enhancing tumor, specifically, the product of the maximal perpendicular crosssectional dimensions (in "measurable" lesions, which are defined as > 10 mm in all perpendicular dimensions). The recommendations are based on expert opinion informed by observational studies and derived from the biologically plausible assumption that changes in tumor size identify progression of disease, potentially before it becomes clinically apparent, resulting in a lead time improvement for the rapeutic intervention (10). Indeed, there may be benefits in changing management before the development of irreversible disability or before the extent of tumor precludes intervention. Some justification for enhancement as a disease proxy has been inferred from data showing that enhancing tumor size and extent of resection are "prognostic biomarkers" (7) at both first presentation and recurrence (11, 12).

The trial assessment recommendations, incorporated in a less stringent form during routine clinical assessment (13), allow for an early change in treatment strategy, for example, termination of ineffective treatments or switching to second-line therapies (14). However, there are four important challenges using conventional structural MRI protocols. First, there is a paucity of evidence that earlier diagnosis of disease recurrence using conventional structural MRI influences prognosis (10). It is noted that individual enhancing tumor growth trajectories vary between individuals with the same histological tumor type.

Second, seemingly simple measurements can still be challenging because tumors have a variety of shapes, tumors may be confined to a cavity rim, and the edge of tumors may be difficult to define (15). For example, large, cyst-like gliomas are common and are often "non-measurable" unless a solid peripheral nodular component fulfils the above "measurable" criteria.

Third, there is a lack of biological specificity for contrast enhancement, which can lead to false positive, false negative, and indeterminate results, particularly relating to post-treatment related pseudophenomena in glioma (10). In high-grade glioma, pseudoprogression is an early post-treatment-related effect typically occurring within six months of finishing concomitant temozolomide and radiotherapy (Figure 1), whereas pseudoresponse typically occurs after anti-angiogenic agents such as bevacizumab have been administered. False positive progression and false negative treatment response are manifest as an increase or decrease in MRI contrast-enhancing volume, respectively. Delayed treatment effects, such as increased enhancement due to radiation necrosis, can similarly cause false positive progression. Other examples of non-specificity include post-operative peritumoral parenchymal enhancement following operative "tissue handling" or following operative infarction.

Fourth, due to the non-specificity of changes in contrastenhancing volume, treatment response assessment typically is made in a retrospective manner as confirmatory imaging is required to demonstrate a sustained increase (**Figure 1**) or a sustained decrease in enhancing volume. This leads to a delay in diagnosis.

Clearly, contemporaneous, accurate, and reliable monitoring biomarkers are required for high-grade glioma treatment response assessment. Due to the non-specific nature of contrast enhancement, histopathology is sometimes employed as a combined or alternative monitoring biomarker; often it is considered the reference standard-of-treatment response (Figure 2). However, there are several challenges when using histopathology as a monitoring biomarker. First, there is a paucity of evidence that earlier diagnosis of viable tumor using biopsy influences prognosis (2). Second, repeat tissue sampling has a high risk of procedure-related morbidity in patients with highgrade glioma compared to other systemic cancers (20). Third, biopsy has a potential drawback of sampling bias (21). Fourth, there is a need to improve and better systematize the application of histological and molecular analysis to diffuse glioma in the recurrent setting; currently, it is not standardized, causing a variety of interobserver diagnostic interpretations given the background of extensive post-therapy-related changes (2).

An emerging alternative approach is to harness the potential value of circulating biomarkers (including circulating tumor cells, exosomes, and microRNAs) to monitor disease progression in glioma patients (22). For example, a recent study found that serum micro-RNA levels did not increase in

cases of pseudo-progression, and increased levels were associated with tumor progression (23). However, as with any potential monitoring blood or cerebral spinal fluid biomarker, potential use requires further evaluation and validation in larger scale prospective studies before implementation into routine clinical practice can be envisaged.

Another promising approach, which concords with an impetus to derive an evidence-base for follow-up imaging of high-grade gliomas driven both by researchers as well as patients and caregivers (24-26), is to use advanced imaging techniques. Considerable technical developments have occurred in the last three decades; however, clinical translation is far from ubiquitous. A European survey in 2016 suggested dynamic susceptibility contrast-enhanced (DSC) MRI and 1H-magnetic resonance spectroscopy was used in the clinic routinely in 82% and 80% of hospitals respectively (27). However, with only 3% survey response rate, it is unlikely to be representative of European practice. A 2020 survey of all UK neurooncology centers with 100% response rate from all three lead specialists (neuroradiology, neuro-oncology, and neurosurgery) within all centers showed that only 10% of centers routinely used any advanced MRI technique and a third of centers used such techniques in selected cases (13). It is noteworthy that many respondents suggested that advanced imaging techniques would improve their practice.

The purpose of this position statement is to summarize the evidence for the use of advanced MRI techniques as monitoring biomarkers in adult high-grade glioma in the clinic, and to highlight the latest bench-to-bedside developments.

2 MATERIALS AND METHODS

Clinicians, engineers, and physicists with expertise in advanced MRI techniques applied to high-grade glioma, convened virtually through the European Cooperation in Science and Technology (COST) Glioma MR Imaging 2.0 (GliMR) initiative (28) from July 2020 through July 2021, which also included experts from outside of Europe. A working group analyzed the available evidence for potential high-grade glioma monitoring biomarkers derived from advanced imaging techniques. The consensus decision was to focus on monitoring biomarkers that can reliably differentiate post-treatment-related effect from true tumor progression during (or before) the point when contrast enhancement on T_1 -weighted MRI images first increases. False-negative progression (pseudoresponse) during second line treatment is a concern in the United States but rarely in Europe, as the European Medicines Agency concluded that progression-free survival bevacizumab trial outcome measures are inherently confounded and the use of bevacizumab is not supported (29). Therefore, pseudoresponse was not the focus of the position statement.

Potential biomarkers were derived predominantly from the individual image acquisition modalities of perfusion and/or permeability (using DSC MRI techniques, dynamic contrast enhanced [DCE] and arterial spin labeling [ASL]), microstructure [diffusion MRI techniques], metabolism and/or chemical



FIGURE 1 | Panels of axial T₁-weighted images after contrast administration in two patients with glioblastoma, IDH-wildtype. (A–D) demonstrate tumor progression.
(A) Preoperative appearance of an occipital glioblastoma. (B) MRI performed five days after resection with no enhancement suggestive of residual tumor. (C) The patient continued with standard-of-care radiotherapy and temozolomide. Within three months of radiotherapy, a new enhancing lesion could be seen at the margin of the postoperative cavity. (D) The enhancing lesion had increased in size three months later and was confirmed to represent tumor recurrence after repeat surgery.
(E–H) demonstrate pseudoprogression. (E) Preoperative appearance of an insular glioblastoma. (F) Postoperative appearance 24 hours after surgery showing blood degradation products, with no enhancement suggestive of residual tumor. (G) The patient continued with standard-of-care radiotherapy and temozolomide. MRI performed within six months of radiotherapy demonstrated a new, contrast-enhancing lesion. (H) Follow-up MRI at monthly intervals showed a gradual decrease in the size of the contrast-enhancing lesion without a change to standard-of-care temozolomide or corticosteroid use. The image shown here is the MRI performed four months later.



FIGURE 2 | Hematoxylin and eosin-stained biopsy samples showing three distinct histopathological patterns from three patients who were treated for glioblastoma, IDH-wildtype. All biopsies were obtained from a contrast-enhancing region that had increased in size on serial T_1 -weighted images during follow-up imaging. (A) A fully viable tumor recurrence with dense cellularity and pseudopalisading necrosis. This corresponds to tumor progression. (B) Depopulated tumor with necrotizing treatment effect. This corresponds to the radiological appearances of pseudoprogression. (C) Nearly absent tumor cell compartment with extensive necrotizing treatment effect and hyalinizing vasculopathy. This corresponds to the delayed treatment effect, radiation necrosis. Pseudoprogression and radiation necrosis are two well-documented forms of "post-treatment related effect." Pseudoprogression generally occurs within the six months following completion of chemoradiotherapy, and resolves or stabilizes without additional treatment, whereas radiation necrosis generally occurs beyond six months, up to several years after radiotherapy, and is often more severe and progressive (16-19). Images courtesy of Dr. J Huse, Department of Pathology, MD Anderson Cancer Center, United States.

composition (magnetic resonance spectroscopy and chemical exchange saturation transfer [CEST] technique}, and MRI combined with positron emission tomography [PET]).

We also considered post-processing methodologies involving the combination of MRI approaches (multiparametric imaging) and image feature analysis techniques using machine learning (radiomics).

We agreed the work would be a *position statement* (30) promoting discussion on emerging topics and indicate the evidence gaps, strengths and limitations. Together this would provide the foundation for producing a multi-stakeholder downstream *guideline* (30) beyond the current position statement.

Specifically, it was agreed that advanced imaging technique analyses would

- summarize methodology,
- highlight strengths and weaknesses,
- · determine clinical diagnostic accuracy, and
- expound the state-of-the-art and future developments.

Finally, we assessed the incorporation into national and/or international technical and/or clinical guidelines and indicated the current level of development and clinical readiness (presented in Part 2).

Advanced imaging technique analyses were compiled by subject matter experts and incorporated into a manuscript and circulated to the working group members. Edits and feedback were incorporated until all authors were in agreement with the content, and a position statement was produced summarizing the evidence for the use of advanced MRI techniques as monitoring biomarkers in the clinic, and highlighting the latest bench-to-bedside developments.

To determine clinical diagnostic accuracy, we performed MEDLINE (including PubMed), Embase and Cochrane Register searches for recent systematic reviews and meta-analyses, favoring those which followed Preferred Reporting Items for Systematic Reviews and Meta-Analysis: Diagnostic Test Accuracy (PRISMA-DTA) methodology (31). We also performed searches to analyze individual clinical studies related to each advanced imaging technique since the time of the included systematic review; if a systematic review was published before 2015, we confined our searches to 2015–2021. Search terms are listed in Supplementary Table S1. Given that the position statement describes a broad range of studies involving several imaging approaches (a range of MRI advanced techniques, PET, and post-processing methodologies) and several target conditions (pseudoprogression, radiation necrosis, or a combination of both) a PRISMA-DTA analysis addressing a specific question on diagnostic accuracy was beyond our remit. Nonetheless, components of the PRISMA-DTA methodology have been incorporated where practicable.

3 RESULTS

3.1 An Overview of Perfusion and Permeability

A hallmark of cancers is angiogenesis, the formation of new tumor vessels to sustain metabolic demands of the growing tumor (32). Its main trigger is hypoxia *via* HIF-1 (hypoxia inducible factor 1) release and cascades of other pro-angiogenic factors, e.g., vascular endothelial growth factor. Compared to normal vessels, tumor

vessels are larger, more irregular, and, importantly, highly permeable. The increased permeability leads to a disrupted BBB, allowing the extravasation of macromolecules (such as contrast agents) across the capillary wall (33). The glioma vasculature will thus exhibit increased perfusion, blood volume, and permeability compared with normal brain tissue.

Although the exact definition may depend on the method applied (34), perfusion is generally defined as the passage of blood through the tissue microcirculation. In the brain, cerebral blood flow (CBF) is traditionally used synonymously with perfusion, and both are measured in units of volume of blood per volume or weight of tissue per time (ml/100 ml/min or ml/100 mg/ min). On the other hand, the term perfusion imaging is often used in a wider sense and also includes other aspects of the vascular function of an organ or tissue such as blood volume or vessel permeability. In the brain, the most commonly measured perfusion parameters are CBF and cerebral blood volume (CBV). In order to distinguish perfusion parameter maps (showing regional distribution within the brain) from global brain measurements, in imaging they are sometimes referred to as regional CBF and CBV (rCBF and rCBV, respectively). In a possibly confusing fashion, measures of CBF and CBV that are obtained voxelwise relative to the rest of the brain, but not in absolute units, are also often referred to as rCBF and rCBV, respectively. The latter definition of rCBF/ rCBV is commonly used in the context of the perfusion imaging techniques described here. For this reason, we chose to use CBF and CBV to refer to absolute measurements of perfusion and blood volume, while keeping rCBF and rCBV for relative versions of these parameters. Moreover, perfusion imaging measurements of brain tumors are commonly normalized to a reference, for example healthy appearing brain of the contralateral hemisphere. In this case, normalized values are referred to as nRCBF/nRCBV, respectively. Another perfusion related parameter is vessel permeability, which may be quantified in terms of a transfer constant (K^{trans}) or a permeability surface area product (PS), depending on the measurement technique and associated modeling approach.

DSC and DCE techniques are based on dynamic MR imaging during the bolus passage of a GBCA to provide images of perfusion- and permeability-related parameters. DSC is a T_2/T_2^* method most often used to assess vascular proliferation in terms of CBV. Conversely, DCE is a T_1 -weighted method, which in its most widely used form provides information on a combination of tissue perfusion and vessel permeability as measured by K^{trans} in addition to plasma volume (Vp). Alternatively, arterial spin labeling (ASL) is a completely noninvasive and potentially quantitative technique without GBCA, which is based on labeling arterial water in order to provide an endogenous, freely diffusible perfusion tracer (35). ASL provides perfusion-weighted images, and it can be used to quantify tissue perfusion (CBF).

3.2 DSC-MRI

3.2.1 Methodology

With DSC-MRI, T_2 - or T_2^* -weighted images are acquired with high temporal resolution during the bolus administration of a GBCA (**Figure 3**). The bolus passage induces a gradient of susceptibility between the intravascular and extravascular tissue compartments causing a transient decrease in the signal intensity (36). Most often T_2^* -weighted gradient-echo (GRE) echo planar imaging (EPI) methods are used, which are sensitive to vessels of all diameters (37). T_2 -weighted images, obtained with spin-echo (SE) EPI methods, provide CBV maps primarily sensitive to microvessels. Subsequent descriptions will be given for GREbased methods unless otherwise noted. The same basic principles for data acquisition and processing apply for SE-based methods.

As further illustrated in **Figure 3**, the DSC signal time course, S(t) is converted to a change in the T_2^* relaxation rate ($\Delta R2^*(t)$),

$$\Delta R2^{*}(t) = \frac{-1}{TE} \ln\left(\frac{S(t)}{S_{b}}\right),$$
 Equation 1

where S_b is the mean of the baseline (pre-bolus) signal intensity whose integration provides an estimate of the relative cerebral blood volume (rCBV).

$$rCBV = \int \Delta R2^*(t)dt$$
 Equation 2

This measure is termed "relative" because it provides a voxelwise value *relative* to the rest of the brain.

Alternatively, an absolute measure of CBV can be determined from the ratio of the tissue ($C_t(t)$) (which is assumed equivalent to $\Delta R2^*(t)$) and arterial concentration-time curve ($C_a(t)$) first-pass areas,

$$CBV = \kappa \frac{\int C_t(\tau) d\tau}{\int C_a(\tau) d\tau},$$
 Equation 3

where κ is the scale factor accounting for the density of brain tissue and the differences in hematocrit between capillaries and large vessels (38). As indicated, an absolute measure of CBV requires the determination of Ca(t), also referred to as the arterial input function (AIF). More often, for purposes of comparison, the rCBV will be normalized by dividing the rCBV in each image voxel by the mean rCBV determined from a reference tissue region of interest (ROI), such as normal appearing white matter giving normalized rCBV (nRCBV) values. Like absolute CBV, the nRCBV maps provide greater consistency for the comparison of rCBV across time and patients. Note that in some instances the term nRCBV may be used interchangeably with normalized CBV (nCBV), with the implicit assumption that the CBV is relative. In all cases, an explicit description of the steps used to create CBV should be provided, with clear definitions of nomenclature provided. Finally, another newer approach termed "standardization" has been devised to calibrate the rCBV maps directly (39) without requiring a normalizing ROI. Standardized rCBV (sRCBV) has demonstrated greater consistency across time (39) and improved repeatability compared with nRCBV (40). Both nRCBV and sRCBV have been used with success in several brain tumor clinical trials (41–43).

3.2.2 Correction for Leakage Effects

A key assumption of DSC-MRI is that GBCA remains compartmentalized within the intravascular space, causing a



susceptibility difference between the intravascular and extravascular compartments. However, with brain tumors, which often have a disrupted BBB, this assumption is violated. GBCA extravasation causes changes to the extravascular T_1 and T_2 (GBCA causes a change in the relaxation rate of the local tissue, which is inversely proportional to T_1 and T_2 [$R_1 = 1/T_1$; $R_2 = 1/T_2$) in ways that can profoundly affect the DSC-MRI time course (44) and, thus, the accuracy of the rCBV estimate (45, 46). Therefore, different MRI acquisition and post-processing approaches have been used to diminish and/or correct for confounding GBCA leakage effects. One common approach is to administer a single-dose preload of GBCA before the collection of the DSC-MRI data, during which a second dose of GBCA is administered. The preload serves to diminish the effects of contrast extravasation that would occur during the DSC-MRI acquisition (45). One study demonstrated that use of a single-dose preload (0.1 mmol Gd/kg) with a 5-minute incubation time before the DSC-MRI dose, provided the best distinction in rCBV between tumor and treatment effect (47). Yet, even with a contrast agent preload, a correlation with tumor grade was only evident when a post-processing correction for leakage effects was also applied (46). The Boxerman-Schmainda-Weisskoff method (48) has become one of the most widely used leakage correction methods, but several modifications have since been proposed that deserve further consideration (49–51).

More recently, a population-based digital reference object of brain tumor tissue was developed to evaluate the entire parameter space of tissue condition and imaging parameter settings (52, 53). The digital reference object results confirmed that the double-dose approach using a single-dose (0.1 mmol/kg) preload and single-dose DSC with Boxerman-Schmainda-Weisskoff correction was the most accurate and most robust method, being fairly insensitive to slight variations in parameter settings. Yet, another potential lower GBCA dose method emerged. By simply using a lower flip angle (30°) with TE = 30 ms (at 3 Tesla [T]) or TE = 54 ms (at 1.5 T), it was predicted that rCBV results comparable to the double-dose reference standard could be obtained. This was experimentally confirmed at 3 T in a multisite study of brain tumor patients (54). These results helped to inform a recent consensus recommendation specifically for DSC-MRI data acquisition in high-grade glioma, which includes both the double-dose reference standard and single-dose methods (55). Clinical MRI results together with rCBV maps, obtained using the double-dose reference standard from a patient with a grade 2 oligodendroglioma, are shown in **Figure 4**.

3.2.3 Evidence from Clinical Studies

A systematic review and meta-analysis of studies between 2005 and 2015 included 17 DSC-MRI studies and concluded that individual studies, taken in an isolated form, showed encouraging results to differentiate tumor progression from treatment-related changes, with sensitivities and specificities in the 80%-90% range (56). Yet, widespread use has been hampered by the wide range of proposed rCBV thresholds reported (0.9-2.15). This variation is largely attributed to the statistically significant heterogeneity in how the DSC-MRI data are collected and analyzed (56). However, the studies reviewed preceded the recently published consensus recommendation that describes how best to acquire DSC-MRI data, and that a correction for leakage should be included. With widespread implementation of this consensus acquisition, it is hoped that greater consistency in mean rCBV thresholds will follow. A recent multisite study supports this contention (57). Several sites used their own postprocessing platform to process the same DSC-MRI data whose acquisition was consistent with the subsequently published consensus recommendation regarding acquisition. A common threshold applicable to all sites could be determined to distinguish high- from low-grade tumor. This suggests that, with greater consistency in the acquisition of DSC-MRI data, agreement on thresholds to distinguish treatment-related effects from residual or recurrent tumor is also possible.

The practice of using the mean rCBV determined from the enhancing lesion ROI is another often-overlooked factor that may contribute to the wide range of reported rCBV thresholds. Rarely is a lesion composed of pure tumor or pure treatment effect. Instead, there is an admixture of tumor and treatment effect that will be unique for each patient and each cohort of patients. Yet, all studies described in the meta-analysis (56) used the mean rCBV from an ROI. Consequently, it is not surprising that each study reports a different threshold value because each



study comprises a unique set of patients. Even if the acquisition and post-processing methods were equivalent, it is still likely that each study would report a different threshold to distinguish tumor from treatment effect.

One proposed research solution to the determination of a widely accepted single threshold, is to use image-localized biopsy and spatially histopathologic correlation as described in two retrospective studies of high-grade brain tumors (58, 59). In these studies, only tissue confirmed to be either pure tumor or pure treatment effect were used to determine an rCBV threshold for distinction. In both studies, the consensus acquisition protocol was used, and the rCBV threshold was determined to be the same to distinguish high-grade tumor from treatment effect. A significant difference in tumor and treatment effect was also found for standardized rCBV (59). Biopsy-determined rCBV thresholds can be used to create voxelwise maps that distinguish tumor from non-tumor for the determination of the overall fraction of tumor burden within the enhancing lesion. These maps, referred to as fractional tumor burden (FTB) maps (60), provide a unique ability to visualize the admixture of tumor and post-treatment effects (Figure 5). The potential of FTB to predict response to treatment (61) and to distinguish between tumor or treatment effect, as confirmed by histopathologic examination of resected tissue samples (62), has been demonstrated. More recently, a prospective study (63) using image-localized biopsy tissue validated the ability of rCBV to predict tumor content (0-100%), demonstrating similar performance of nRCBV and sRCBV for the creation of FTB. Given that sRCBV does not require choosing a reference ROI, this represents an important step toward workflow optimization for the creation of FTB maps.

3.2.4 Future Developments

Other DSC-MRI perfusion metrics have also demonstrated promise for providing clinically relevant information to evaluate treatment response. In a preclinical study, the distribution of mean transit times, determined from DSC-MRI, was shown to be dosedependent and predictive of response to the anti-angiogenic agent Sugen11657 (64). In a related study, capillary transit time heterogeneities together with indices of estimated tumor oxygenation, both determined from the DSC-MRI data, demonstrated promise for predicting unfavorable therapeutic effects in patients with recurrent glioblastoma (65). Combined GRE and SE sequences, which can provide measures of mean vessel diameter, have been shown to correlate with brain tumor grade (45, 46) and response to anti-angiogenic treatment (64). Multi-echo perfusion sequences, such as SPICE (44) or SAGE (66), measure GRE and/or SE at multiple echo times (TEs), enabling implicit correction of T_1 leakage effects and the data for determining both DSC-MRI and DCE-MRI perfusion parameters maps (67). Though the technology is still under development (68), these newer perfusion methods are likely to provide a wealth of information relevant to treatment monitoring in brain tumors. Furthermore, with the recent development of the brain tumor DSC-MRI digital reference object (52, 53), which can serve as a benchmark for all DSC-MRI methods (69), translation of these technologies for clinical use should take place more quickly than previously possible. The additional role to predict response to treatments is also emerging, as mentioned above, in the context of FTB.

3.2.5 Summary, Strengths, and Weaknesses

Measures of rCBV derived from DSC-MRI have consistently demonstrated the ability to provide information about brain tumor growth and response to treatment that is not available with current structural MRI methods. Of all advanced techniques, DSC is the most extensively studied for clinically utility. With a power-injector available and a cannula placed for contrast agent administration, DSC-MRI data are easy to collect using commonly available imaging methods and requires only an extra 2-3 minutes of scanning. Despite the ease of use and valuable information provided, the widespread clinical use of DSC-MRI has been hampered by the need to correct for confounding leakage effects, which in turn has led to a variety of ways to collect and process this data. However, great strides have been made to overcome these limitations, with a recent publication clearly outlining how best to collect this data (Boxerman consensus) and a strong recommendation that leakage correction must be used in the post-processing. In this





regard, the Boxerman-Schmainda-Weisskoff leakage correction (48) method has been the most commonly used and currently is the most widely recommended approach. Therefore, consensus regarding the collection and processing of DSC-MRI is being reached. Remaining challenges include the fact that the DSC-MRI acquisition is most often based on EPI. Although EPI provides the necessary temporal resolution, the spatial resolution is less good and significant signal dropout and geometric distortions can occur at air-tissue interfaces, such

as near sinuses, with post-surgical blood products, or at resection sites. Therefore, efforts are underway to develop higher resolution fast imaging techniques with less sensitivity to these unwanted susceptibility effects. These efforts include the development of spiral-based sequences (44) or parallel imaging methods such as SENSE (70). With a complete solution composed of consistent acquisition and post-processing methods, all obtained with high temporal and spatial resolution, DSC-MRI should be widely accepted as a robust addition to standard brain tumor exam to assess treatment response.

3.3 DCE-MRI 3.3.1 Methodology

DCE-MRI is based on dynamic T_1 -weighted imaging after intravenous administration of a GBCA (**Figure 6**). The contrast agent will increase the relaxation rate $R_1 = 1/T_1$, and the change in relation rate is linearly related to the concentration of the contrast agent *C* by

$$R_1 = R_1^0 + r_1 C,$$
 Equation 4

where R_1^0 is the relaxation rate before arrival of the contrast agent and r_1 is the field strength specific relaxivity of the contrast agent. The exact relationship between signal (S) and change in relaxation rate depends on the sequence used. For the commonly used 3D T_1 -weighted spoiled gradient recalled echo sequence, with short TE << T_2^* so that T_2^* effects are negligible, the measured signal is related to the contrast agent by:

$$S(t) = M_o \sin(\alpha) \frac{1 - e^{-\frac{TR}{T_1(t)}}}{1 - \cos(\alpha)e^{-\frac{TR}{T_1(t)}}}$$
$$= M_o \sin(\alpha) \frac{1 - e^{-TR(R_1^0 + r_1 C(t))}}{1 - \cos(\alpha)e^{-TR(R_1^0 + r_1 C(t))}}, \qquad \text{Equation 5}$$

where α is the flip angle and M_o is the equilibrium magnetization. Only T_1 , and thus R_1^0 , are unknown and are

usually estimated by preceding T_1 mapping (typically obtained by performing T_1 imaging at variable flip angles).

Various tracer kinetics models can be applied to the measured arterial and tissue concentration curves [see Ref. (71) for a comprehensive review]. The choice (and complexity) of the model, and thus the requirements for the imaging protocol, are dependent on the tissue and pathology of interest. For brain tumors, the most widely applied three-parameter twocompartment model (i.e., the extended Tofts' model, Figure 7) provides voxelwise estimates of plasma volume fraction (V_p), interstitial volume (V_e), and K^{trans} (72), and thus information on permeability and blood volume, both indicators of neoangiogenesis. For reliable estimation of K^{trans} from this model, a temporal resolution of < 10 sec (ideally ≤ 5 sec) and a total scan duration of 5 minutes following a single dose (0.1 mmol/kg) of GBCA are recommended in the proposed revised Quantitative Imaging Biomarkers Alliance (QIBA) profile (73). In the extended Tofts' model, the parameter K^{trans} reflects both tissue perfusion (F) and PS. Except in extreme cases where permeability is very low (F >> PS and $K^{trans} \approx PS$) or very high (PS >> F and $K^{trans} \approx F$), the parameter K^{trans} is a mixture of F and PS. The errors/bias of the parameter estimates derived from the extended Tofts' model may be substantial and depend on both the tissue (e.g., differences in low- compared to high-grade tumors) and the experimental conditions, e.g., temporal resolution (74). Still, this model may provide useful estimates of $V_{\rm p},\,K^{\rm trans}$, and $V_{\rm e}$ also at low temporal resolution on 1.5 T systems.







Good repeatability of DCE using the extended Tofts' model has been reported in normal tissue of both healthy subjects and patients with brain tumors (75), and moderate-to-good repeatability of K^{trans} similar to that of DSC nRCBV was found in contrast enhancing lesions of patients with glioblastoma during treatment (76). Nonetheless, a cross-vendor and multisite reproducibility study of quantitative T_1 imaging in a phantom found variable reproducibility dependent on both sequence and protocol used, field strength and range of T_1 values (77). Furthermore, whilst studies of DCE reading and analysis reported good to excellent inter- and intraobserver agreement (78, 79), there was considerable influence from the software used (78).

3.3.2 Future Developments

The current standard implementation of DCE using the extended Tofts' model described in the recent QIBA guidelines (73) may be considered state-of-the-art for DCE brain tumor imaging, but it has changed little in recent years. Methods taking advantage of better signal-to-noise ratio (SNR) at 3 T allow higher temporal resolution, permitting quantification of F, e.g., by model freedeconvolution (80), and thus separation of F and K^{trans} by the four-parameter two-compartment exchange models (2CXM) (81, 82). Estimation of F is (among other factors) highly sensitive to errors related to undersampling of the input function; therefore, high temporal resolution (ideally 1-2 sec) is required and may be achieved by limiting coverage or spatial resolution. Simpler (model independent) approaches are based on initial area under the signal curve (iAUC), and reflect the initial accumulation of contrast by the tissue (irrespective of the mechanism). This approach alleviates the need for T_1 mapping, measurement of arterial input function, and kinetic modeling, and it is possibly more robust across scanners and implementations (83, 84).

3.3.3 Strengths and Weaknesses

DCE-MRI is the standard perfusion method outside the brain, e.g., prostate and breast, but it is less widely used than DSC-MRI in the brain. Nonetheless, DCE-MRI has several advantages compared with DSC-MRI for post-treatment imaging of brain tumors. First, DCE-MRI is a T_1 -weighted imaging technique and hence is less sensitive to the susceptibility artifacts in tissue adjacent to the skull base as well as within treated tumors that limit the coverage of T_2^* -weighted imaging in DSC-MRI (85). In fact, DCE-MRI might provide useful image quality in > 80% of uninterpretable DSC-MRI cases (86). Second, unlike T2*weighted imaging, T_1 -weighted contrast leakage effects are not a cause of error when using DCE-MRI. Instead, they allow explicit modeling of permeability and can provide absolute parameter maps of tissue microvascular properties. Third, T_1 weighted imaging in DCE allows easier and more accurate estimation of an AIF for kinetic modeling than EPI used for DSC-MRI. This and the conversion of blood and tissue signal to concentration of GBCA permit absolute quantification of physiological parameters, although quantitative parameter estimates probably are highly model specific. The main technical disadvantages of DCE-MRI compared with DSC-MRI are lower temporal resolution, need for T_1 mapping, and increased scan duration. Clinically, there are no generally established cut-off points or diagnostic criteria using the standard implementation (the extended Tofts' model). In the case of 2CXM analysis, a single small study investigated 2CXM DCE for pre-radiotherapy prediction of post-radiotherapy tumor progression (87), but no published clinical data regarding diagnostic accuracy are currently available, means of implementation are not standardized, and scanner and software solutions for advanced kinetic modeling are limited.

3.3.4 Evidence From Clinical Studies

Compared with DSC-MRI, the available evidence from clinical studies is more limited. **Supplementary Table S2** shows studies of diagnostic accuracy for differentiating disease progression related and treatment-related effects. Two partially overlapping meta-analyses (56, 88) calculated pooled sensitivity and specificity of 88–89% and 85–86%, respectively, based in combination on a total of 10 studies published from 2011 to 2015 including 371 patients. It is noteworthy that the diagnostic accuracy was higher in studies with model independent analysis (88) than in studies with two- or three-parameter models, while most recent studies (and clinical practice) typically employ three-parameter models. Clinical multicenter brain tumor studies

using DCE have been performed for tumor grading (79) and for evaluation of antiangiogenic treatment of recurrent glioma (76).

Several studies have made comparisons of the various DCE-MRI parameters with each other and with DSC-MRI. Optimal cut-off values determined by receiver operating curve (ROC) analysis and associated diagnostic accuracies from the individual studies are shown in Supplementary Table S2. Two large retrospective studies from the same group, each including more than 150 patients, found similar overall accuracy (area under receiver operating curve [AUROC]) of DSC-MRI and model free DCE-MRI for separation of progressive disease from pseudoprogression (89) and radiation necrosis (90), with DSC-MRI tending to provide higher specificity and lower sensitivity compared with DCE-MRI. Smaller retrospective studies comparing model-based analysis of DCE-MRI with DSC-MRI have reported variable findings, although single DCE-MRI parameters tend to provide lower diagnostic accuracy compared with DSC-MRI (Supplementary Table S2). The only prospective study comparing three-parameter analysis of DCE-MRI with DSC-MRI found AUROC of Vp and the model free analysis (iAUC) to be similar, and both slightly lower than DSC-CBV, whereas K^{trans} yielded the lowest AUROC (91). A prospective study including both high-grade glioma and metastases also found AUROC of \boldsymbol{V}_{p} superior to \boldsymbol{K}^{trans} , and suggest improved accuracy when combing the Vp and K^{trans} (92). Other, mainly smaller, retrospective studies of three-parameter analysis have suggested K^{trans} or V_e outperform V_p (Supplementary Table S2), while a single study found no diagnostic value of any DCE-MRI parameter (93). The evidence does not converge toward recommending a specific parameter or cut-off point, but a visual analysis of parameter maps comparing tumor to normal tissues may be a clinically acceptable approach (94).

In summary, clinical studies have proven the potential of DCE-MRI for distinguishing treatment-related effects from recurrence, but the optimal DCE acquisition protocol, mode of analysis, and parameter of highest diagnostic value and optimal cut-off points remain to be established.

3.4 ASL

3.4.1 Methodology

ASL is a perfusion imaging technique based on magnetically labeling water spins as they flow through the large brainsupplying arteries at the cervical spine level using radiofrequency (RF) pulses. An image of the brain is acquired after allowing for sufficient time for the labeled spins to reach the brain capillaries. The signal difference in magnetization between labeled images and non-labeled control images yields perfusion-weighted images using kinetic model estimations to calculate the quantitative CBF map (95). Typically, multiple label and control repetitions are acquired in alternating order, and their average subtraction value is used. A variety of ASL pulse sequences and associated analysis methods have been developed, which employ different strategies to control for unwanted labeling/subtraction effects and to overcome the intrinsically low SNR of the ASL signal. For a recent review focused on the ASL basic concepts and the current state-of-theart acquisition and analysis approaches, see Ref. (35). New developments in ASL are also detailed by van Osch et al. (96).

3.4.1.1 Labeling Schemes

The two main labeling schemes are continuous ASL (CASL) (97) and pulsed ASL (PASL) (98, 99), illustrated in Figure 8. While CASL relies on the flow-driven adiabatic inversion of arterial water spins as they flow through the labeling plane under the continuous application of a RF field, PASL is based on the instantaneous inversion of water spins within a wide slab upon the application of an adiabatic inversion RF pulse. A derivation of CASL was more recently developed, pseudo-continuous ASL (pCASL), which uses a train of RF pulses to induce flow-driven adiabatic inversion of arterial water spins (100). In this way, it retains the SNR advantage of CASL compared with PASL while allowing implementation in standard clinical MRI systems not possible with CASL. The advantages of pCASL relative to both CASL and PASL have made this the labeling scheme of choice for clinical applications (95). More advanced and purposespecialized labeling schemes employ non-spatially selective labeling, which can be achieved by inverting spins based on their velocity, or acceleration, rather than their spatial position: velocity-selective ASL and acceleration-selective ASL (101).

3.4.1.2 Image Acquisition

Fast image readout techniques should be employed, most commonly these are multi-slice 2D EPI but also segmented 3D methods such as 3D GRASE or 3D stack-of-spirals. While the latter are recommended for their higher SNR, the former is less sensitive to motion. Other acquisition parameters need to be selected, namely regarding background suppression and vascular crushing gradients, which may be used to minimize signal contributions from static and macrovascular spins, respectively. A critical choice is that of the time delay between labeling and image readout-post-labeling delay (PLD) in pCASL/CASL and inversion time (TI) in PASL (see Figure 8A). Most often, a single delay acquisition is performed: in this case, the delay should be long enough to ensure that the labeled spins have reached the tissues by the time signal is read out, so that it is not sensitive to variations in the arterial transit time or bolus arrival time, illustrated in Figure 8. Additionally, in PASL, saturation pulses should be applied in order to control the label duration, using QUIPSS II (102) or Q2TIPS (103) approaches (see Figure 8A). Because the label, and hence the perfusion signal, decay with longitudinal relaxation (T_1 of arterial blood up to the capillaries and T_1 of tissue after exchange), the trade-off between insensitivity to arterial transit time and sufficient SNR must be considered when choosing the value of PLD/TI. Alternatively, multiple delays may be sampled and an appropriate kinetic model fitted to the data to estimate CBF as well as arterial transit time. For multi-delay acquisitions, optimal sampling schemes (104, 105) or more efficient sampling schemes such as time-encoded pCASL (106), should be considered.

3.4.1.3 Perfusion Quantification

For perfusion quantification, tracer kinetic models have been developed with different levels of complexity. A standard kinetic model of ASL signals was originally developed by Buxton assuming a single well-mixed compartment and instantaneous exchange of water between blood and tissue (107) (see **Figure 8B**). Various



extensions of this model have since been proposed, namely to account for restricted water exchange/vessel permeability using two compartment models (108, 109), macrovascular (as well as microvascular) contributions (110), partial volume effects (111), or flow dispersion (112). Simplifications of the standard model have also been proposed (95) to obtain CBF measurements based on single-delay ASL acquisitions:

In pCASL:
$$CBF = \frac{\lambda \Delta M e^{\frac{PLD}{T_{1b}}}}{2\alpha T_{1b} M_{0b} (1 - e^{-\frac{r}{T_{1b}}})}$$
 Equation 6

In PASL:
$$CBF = \frac{\lambda \Delta M e^{\frac{TI}{T_{1b}}}}{2\alpha TI_1 M_{0b}}$$
 Equation 7

where ΔM is the control-label magnetization difference, M_{0b} is the equilibrium magnetization of arterial blood, T_{1b} [s] is the longitudinal relaxation time constant of arterial blood, λ is the blood/tissue partition coefficient for water, τ [s] is the label bolus duration, and α is the labeling efficiency. In order to quantify CBF in absolute units, calibration of ASL measurements must be performed. This involves normalizing the control-label difference images, ΔM , by the equilibrium magnetization of arterial blood, M_{0b} (113). Since M_{0b} cannot be measured directly *in vivo*, different methods have been proposed to estimate it based on the measurement of the tissue M_0 , typically using a proton-density image acquired with the same readout as the ASL images but with a long repetition time (TR).

3.4.1.4 Consensus Recommendations

In an attempt to harmonize ASL perfusion measurements across studies, a consensus paper was published in 2015 with recommendations regarding the most appropriate implementation for clinical applications (95). In particular, the combination of pCASL with a segmented 3D readout was recommended for its superior SNR; however, PASL and 2D readouts still are often used. The use of background suppression without vascular crusher gradients is also recommended. Although ASL may be performed at 1.5 T, the recommended field strength is 3 T. In this paper, a calibration procedure and a simplified kinetic model were also proposed for CBF

quantification based on single delay measurements. Although the consensus paper resulted in part from the efforts of a European network focused on dementia, the recommendations were made for clinical applications more generally. In fact, this consensus paper resulted in the implementation of a recommended sequence by MRI vendors, and this has indeed been adopted by an increasing number of studies. However, the consensus paper did not take tumor studies into key account, nor does it discuss peculiarities of tumor ASL imaging, which may influence the optimal application of ASL in tumor imaging under the recommended conditions. Several studies have assessed the test-retest repeatability of ASL CBF measurements, including the recommended single-delay pCASL implementation (114, 115) but also more complex protocols (116, 117). A reproducibility study has been performed for ASL CBF measurements obtained with commercial implementations of ASL across from major MRI vendors (118). This, however, preceded the consensus paper and results may be improved now that vendors have generally adopted these recommendations. Regarding multisite reproducibility, only one study has partially investigated this by assessing CBF in specific brain regions (115). All of these reliability studies have been performed on healthy subjects, and no studies in patients with brain tumors have been reported.

3.4.2 Strengths and Weaknesses

In contrast to DSC-MRI and DCE-MRI, ASL techniques do not require the injection of a contrast agent. As a consequence, fast repeated measurements on individual patients can be performed. Moreover, the absence of potential gadolinium deposition effects also allows ASL to be more widely used in pediatric patients and in patients with impaired renal function. Relative to DSC-MRI, ASL has the additional advantage that perfusion measurements are not tampered by BBB permeability effects. Even after leakage effects have been corrected for, DSC perfusion imaging is based on GBCA as an intravascular tracer, which limits its ability to yield true perfusion measurements. In contrast, ASL is based on water as a diffusible tracer, allowing it to truly measure perfusion at the capillary level (**Figure 9**). Additionally, susceptibility effects leading to signal dropout and geometric distortions are less prominent since shorter echo times are used.



FIGURE 9 | Three-months post-radiotherapy follow-up in a 58-year-old patient diagnosed with a glioblastoma, IDH-wildtype, studied on a 3T GE MRI system. (A) The post-GBCA T_1 -weighted series shows thick rim enhancement, which could theoretically also be pseudoprogression after recent completion of standard-ofcare radiotherapy and concomittant temozolomide. (B) The DSC rCBV map, acquired with 2D EPI (TE/TR = 45/1500 ms), indicates general hyperperfusion of the enhancing outer tumor rim, which makes early true progression more likely and is thus decisive for further therapy planning. (C) The ASL CBF map, which was acquired with 3D pCASL using white paper settings, shows hyperperfusion concordant with the DSC rCBV map but without the need of gadolinium administration.

Despite the potential advantages relative to DSC-MRI and DCE-MRI, the main weakness of ASL remains its intrinsically low SNR. This has led to the proliferation of methodological options and limited accessibility in clinical sites, which have hindered its wider application so far. Besides the general lack of robustness that results from the low SNR, ASL is also more sensitive to motion artifacts than DSC-MRI or DCE-MRI, due to the inherent image subtraction as well as the frequent use of 3D readout. Additionally, single-delay ASL acquisitions may have macrovascular artifacts or underestimation of tissue perfusion in areas with prolonged arterial transit times, e.g., in patients with macrovascular stenosis.

3.4.3 Evidence From Clinical Studies

An overview of clinical studies published is provided in **Supplementary Table S3**. All studies employed commercial pCASL sequences with a single delay. Three of the listed studies were part of a recent meta-analysis of a total of 160 patients, which reported a pooled sensitivity and specificity of 79% and 78%, respectively to differentiate true progression from post-treatment related effects by ASL (119). All other studies also reported reported fair to good diagnostic accuracies for differentiating true progression from post-treatment related effects. The largest retrospective study included 69 patients (both low and high-grade gliomas) and reported sensitivity and specificity of 74% and 82%, respectively, for separation of progression from post-treatment related effects (120). In the two largest prospective studies, one study found both high sensitivity (94%) and specificity (92%) for separation of post-

treatment related effects from true progression among 42 patients with treated high-grade gliomas (121), while the other reported low sensitivity (65%) but very high specificity (100%) among 58 patients (both low- and high-grade gliomas) in the context of pseudoprogression (122).

3.4.4 Future Developments

Studies exploring the utility of ASL for the identification of posttreatment related effects in high-grade glioma are limited to basic implementations of the technique and have technical and clinical variabilities as well as small sample sizes. Despite this, ASL continues to undergo substantial advances yet to be fully exploited in this context (96). In particular, advanced ASL techniques based on multiple-delay ASL measurements combined with appropriate kinetic models may be employed to obtain measurements of perfusion-related parameters besides rCBF (35). Of special interest in glioma is the putative possibility to obtain vessel permeability measurements by using multicompartment kinetic models incorporating water exchange between blood and tissue (108, 109). For now, the biggest challenge is less the technical development and more the harmonization of current protocols, which would lead to clearer comparison between studies with the potential downstream effect of generating a higher level of evidence.

3.5 An Overview of Diffusion Techniques

Various diffusion MRI techniques can be applied to glioma treatment response assessment including standard diffusion-weighted imaging (DWI), apparent diffusion coefficient (ADC)

analysis, diffusion tensor imaging (DTI), diffusion kurtosis imaging (DKI), intravoxel incoherent motion (IVIM), as well as a multitude of microstructural modeling approaches, such as neurite orientation dispersion and density imaging (NODDI) (123) or restricted spectrum imaging (RSI) (124); free water imaging (FWI) (125); and vascular, extracellular and restricted diffusion for cytometry in tumors (VERDICT) (126).

The basic DWI sequence achieves diffusion contrast based on different gradient pulses in between the excitation and refocusing pulses of a basic SE experiment (127). This basic sequence produces images that highlight diffusional water molecule motion by signal loss. It has been extended to capture information of diffusion in different directions (128) and over different time periods. The gradient strength and duration are expressed in different b-values with a unit of s/mm². Typical bvalues for brain imaging are up to a single (also called single-shell) b-value of 1000 s/mm² and the signal decay up to this b-value can be approximated by a Gaussian distribution. Notably, lower (50-250 s/mm²) b-values are associated with signal from microvascular perfusion rather than Brownian diffusion motion, which is important in, for example, IVIM (129), a method that aims to recover the pseudo-diffusion coefficient of blood using several bvalues (also called multi-shell) in its modeling approach. Note that multi-shell acquisitions may also be used in other modeling approaches. Higher b-values of > 1000 s/mm² are associated with more restricted and hindered environments employed in non-Gaussian (such as DKI) and microstructural models.

The ADC is the best-known diffusion parameter. By scaling the diffusion image with a non-weighted image, quantitative maps can be produced. This quantitative technique is available on any commercial MRI machine, which makes the integration in clinical trials attractive. Qualitative visual interpretation is also possible and

gives an impression of cellular density and water content and can help in clinical differential diagnostics (Figure 10).

Extending the diffusion methodology to sensitize the signal over different directions, the DTI approach has seen widespread application due to its methodological and mathematical simplicity and elegance (130). DKI models (131) focus on the deviation from the Gaussian signal. This model is more sensitive to heterogeneity and complex tissue architecture, rendering parameters that are correlated with biological tissue properties and reflecting the organization of tissue microstructure (132). This approach may deliver biomarkers more relevant to disease interpretation, such as in heterogeneous environments, and dense architecture, such as solid tumors (133). A different approach to overcome the limitation of the Gaussian model is by explicitly modeling it. The FWI model can be used both to visualize the Gaussian signal (free water) and to remove it (free water corrected) (125) using a multi-shell acquisition. This may be useful to distinguish a large area of edema or necrosis from residual tumor and can be used to make DTI fiber tracking in the presence of such processes more accurate (134). Other multi-shell models aim to advance the understanding of neuronal diffusion parameters, such as NODDI (123) or RSI (124). Although NODDI has been designed entirely for use in normal brain tissue, the RSI model has been adapted for use in tumors (135).

3.6 DWI

3.6.1 Methodology

The b-value dependent diffusion signal in a classical diffusion acquisition is calculated based on the Stejskal-Tanner equation (127), where S_b is the signal depending on the apparent diffusion coefficient ADC and the diffusion gradient length δ , separation Δ , and strength G. The b-value is defined as $b = \gamma^2 \cdot G^2 \cdot \delta^2 (\Delta - \frac{\delta}{3})$,





where γ is the gyromagnetic ratio. Assuming a pure Gaussian diffusion of water and a log-linear signal decay with increasing b-value:

$$S_b = S_0 e^{-b \cdot ADC}$$
 Equation 8

By acquiring a diffusion weighted image S_b and an unweighted image S_0 , the apparent diffusion coefficient ADC can be calculated as:

$$ADC = \log \frac{S_b}{S_0} \cdot -\frac{1}{b}$$
 Equation 9

If more than one DWI S_b exists, ADC may also be derived by calculating a least squares fit to the Stejskal-Tanner equation.

To sensitize the diffusion image S_{b} , the diffusion gradient pulse may be played out on each of the three gradient axes individually, or on all three gradients at the same time (so-called tetrahedral scheme) (136). If played out on each axis individually, the images need to be averaged before the ADC map is calculated. Therefore, to calculate an ADC image, four separate volumes have to be acquired: the unweighted image S_0 and three S_b images for each of the individual gradient axes. To reduce the time it takes to acquire the sets of images, EPI (137) is commonly performed. EPI uses blipped line encoding to fill out k-space more quickly. Although this technique is incredibly fast, as with perfusion EPI acquisition, it is sensitive to susceptibility changes in the local magnetic field, often leading to artifacts near large tissue-air boundaries e.g., close to the frontal sinuses of the skull.

3.6.2 Strengths and Weaknesses

Diffusion-weighted sequences usually have a fast acquisition time, which makes them suitable for anxious or poorly compliant patients, both of which are common in the brain tumor patient population. DWI sequences are globally ubiquitous and well-standardized across centers with good dissemination of clinical knowledge of image interpretation compared to more advanced methods. They allow the discrimination of differential diagnoses such as stroke mimicking tumor progression or infection in the surgical cavity. On the other hand, DWI is affected by geometric distortions close to bone and air, which can affect diagnostic quality in adjacent brain structures. Similar artifacts can occur in the resected surgical cavity due to blood products.

3.6.3 Evidence From Clinical Studies

Progressive disease in high-grade gliomas is characterized by tumor hypercellularity manifesting as low ADC values (138– 141), which confer poor survival (142–144). In contrast, an increase in ADC may indicate response to chemoradiotherapy (138, 145, 146) (**Figure 11**). Similar findings have been seen in other brain tumors, in tumors outside the central nervous system, and after high-grade glioma immunotherapy (138, 147–149). It is noteworthy that high ADC values have also been associated with the peritumoral T_2 hyperintensity of glioblastoma, albeit inconsistently. A variety of advanced analyses using histogram-based techniques, either at a fixed time or longitudinally, capture tumor heterogeneity and can be





applied to ADC maps to characterize therapy (138, 150, 151). One study showed that the fifth percentile on a cumulative histogram of ADC, obtained at a b-value of 3,000 sec/mm², was lower in progressive disease than in pseudoprogression (P < 0.001) (151). In two single-site studies, a clear cut-off of a mean ADC proved to be successful in predicting pseudoprogression (152, 153) with ADC $\leq 1300 \times 10^{-6}$ mm²/s (sensitivity 100%, specificity 100%) and $\leq 1313 \times 10^{-6}$ mm²/s (sensitivity 98.3%, specificity 100.0%), respectively for true progression.

A recent meta-analysis, which contained six eligible studies with a total of 214 patients, concluded that quantitative ADC is an effective approach for differentiation of glioma recurrence from pseudoprogression, and can be used as an auxiliary tool to diagnose glioma progression: sensitivity was 0.95 (95% confidence interval [CI] = 0.89 - 0.98), specificity was 0.83 (95% CI = 0.72 - 0.91) (154). Another meta-analysis showed that quantitative ADC has moderate diagnostic performance in differentiating glioma recurrence from radiation necrosis (sensitivity: 0.82 (95% CI: 0.75 - 0.87); specificity: 0.84 (95% CI: 0.76 - 0.91). The authors recommended not using diffusion MRI alone in differentiating between glioma recurrence and radiation necrosis (155). The latter analysis was supported by a meta-analysis comparing advanced imaging techniques that gave sensitivity and specificity results of 0.71 (95% CI: 0.60 - 0.80) and 0.84 (95% CI: 0.77 - 0.93), respectively, providing the lowest diagnostic accuracy among advanced MRI techniques analyzed (also MRS, DSC-MRI and DCE-MRI) (156). Ordinary ADC maps were found to be inferior in spatially discriminating treatment response compared with DSC-derived perfusion measures (59). Another study could not find any difference between perfusion and diffusion metrics in the ability to discriminate between recurrent tumor and pseudoprogression (157). In a different study, neither DSC-MRI nor DWI were found to be predictive of tumor recurrence (158).

Incorporating diffusion metrics into radiomics models improved diagnostic performance for identifying pseudoprogression over a model with only structural imaging volumes (89). A retrospective study employing a dictionary-learning approach was able to train its model to identify pseudoprogression from DTI data (159). The performance accuracy of radiomics and machine learning approaches in MRI are described in detail in Part 2.

3.6.4 Future Developments

A few studies have used longitudinal ADC maps, namely parametric response maps (PRMs) (160) or functional diffusion maps (fDMs, **Figure 11**) (145), to discriminate progression from pseudoprogression (161–163). Whilst these promising methods have been at the research stage for more than a decade, more evidence is required to translate these as routine clinical tools.

Since the initial development of fDMs (145, 164, 165), many other studies have assessed the potential of fDMs in treatment response evaluation in glioma (164, 166, 167). A study of 50 patients with glioma proved that the rate of change in fDM "hypercellular" volumes within hyperintense fluid-attenuated inversion recovery (FLAIR) regions of interest, predicted tumor progression, time-toprogression, and overall survival for both cytotoxic and antiangiogenic treatments earlier than standard anatomical imaging (167). Another study including 20 brain tumor patients found that fDMs predicted patient treatment response at three weeks from the start of chemoradiotherapy, revealing that early changes in tumor diffusion values could be used as a prognostic indicator of subsequent treatment response (146). The preliminary conclusion regarding fDMs is that this imaging biomarker has the potential to track treatment response in brain tumor patients in both enhancing and non-enhancing tumors. A recent study employing ADC histogram analysis, found that low-ADC tumor volume shrinkage has the same predictive power as fDMs (168).

A small study assessing the use of PRMs with "relative" ADC, where the relative ADC represents a ratio between tumor and contralateral normal-appearing tissue, showed the potential in differentiating true progression from pseudoprogression (169). Voxel-based PRMs of ADC were used to predict early tumor progression (170). Compared with traditional response criteria (Macdonald or RANO criteria (171)), a PRM has the potential to allow individualization of treatment seven to eight weeks earlier than the traditional response criteria (172), since PRM stratification can be determined during the radiotherapy and concomitant temozolomide treatment phase.

Given that DWI is acquired in most routine MRI examinations, post-processing methodologies leveraging multiparametric combinations, also described in Part 2, have often incorporated diffusion techniques. This may be with or without the application of machine learning approaches. For example, a machine learning model that used FLAIR and DWI images was able to predict tumor recurrence with higher accuracy than using either modality by itself (173). Likewise, analysis with a multiparametric clustering approach was shown to be superior to a single-modality approach (174, 175).

In summary, study designs will likely improve as large prospective studies are performed using published study protocols and this will provide a higher level of evidence for their translation as routine monitoring biomarkers for high-grade gliomas (176). Analyses are also likely to become more complex using histogram analyses or machine learning approaches, which can integrate longitudinal and multiparametric data. The beginnings of this development can already be seen (177).

3.7 IVIM and DKI

3.7.1 Methodology

IVIM and DKI are based on standard diffusion acquisition techniques. Standard DWI with ADC maps as a reference assume that water diffusion in tissues is Gaussian and that the diffusion signal will fall log-linearly with increasing b-value. However, the reality is different due to cell membrane interactions in anisotropic nerve fibers (and glial cells and their products such as myelin) and intravascular water in capillaries. Both IVIM and DKI try to assess the non-Gaussian and perfusion components in the diffusion signal, IVIM at the low end of the b-value axis of the signal-decay curve, assessing the perfusion-dependent component, and DKI on the high end of the b-value axis, above 1000 mm²/s.

The physiological premise behind IVIM is that due to the complex networks of capillaries, the flowing water-bound

protons appear to move randomly, thereby simulating a Gaussian diffusion process. This pseudo-diffusion component can be expressed through various extensions of the basic Stejskal-Tanner equation, of which the biexponential model is possibly the most frequently applied in IVIM (178):

$$\frac{S(b)}{S(0)} = f \cdot e^{-b \cdot (D^* + D_{blood})} + (1 - f) \cdot e^{-b \cdot D_{tissue}}$$
Equation 10

with f being the capillary blood perfusion fraction in the diffusion signal, D^* the pseudo-diffusion coefficient in the capillaries, and D true diffusion in either blood or tissue.

However, model fitting can become more stable, and thus reliable for application, if the model is simplified by separating diffusion into the fast pseudo-diffusion component of flowing water molecules, D^* , and the slow component of water molecules moving by thermal diffusion, D, without further subdivision of tissue and blood components:

$$\frac{S(b)}{S(0)} = f \cdot e^{-b \cdot D^*} + (1 - f) \cdot e^{-b \cdot D}$$
 Equation 11

At high b-values, cell membranes become an influential factor on the diffusion signal. Therefore, the kurtosis effect, or the width and amplitude deformation of the Gaussian diffusion probability curve, needs to be incorporated as a dimensionless kurtosis factor K (131):

$$\frac{S(b)}{S(0)} = f \cdot e^{-bD^*} + (1 - D + f) \cdot e^{-bD + \frac{1}{6}b^2D^2K}$$
 Equation 12

The choice of b-values influences the shape of the fitting curve and therefore the parameter results. For IVIM, this means that more b-values should be chosen in the spectrum below 250 mm^2 / s. Currently, there is no consensus on how many or exactly which b-values should be chosen for IVIM and DKI measurements.

3.7.2 Strengths and Weaknesses

IVIM and DKI are both relevant techniques in neuro-oncological imaging because they can potentially assess important tumor tissue properties such as microperfusion and cellularity. They are also non-invasive. Multiple b-value acquisitions, however, can extend scan time. Choosing fewer b-values will shorten scan time but potentially will reduce the accuracy of the signal curve fit. Furthermore, IVIM and DKI, like all diffusion techniques, are heavily influenced by scanner-dependent field homogeneity as well as magnetic distortion caused by the tissue examined, which can limit their application e.g. in tissues close to air-filled spaces. There are many other limitations in the models used for postprocessing, which can either underestimate or overestimate IVIM and DKI parameters. A detailed elaboration on the issues of fitting and parameter calculation for both IVIM and DKI was published in 2017 (131). The influence of model parameter selection to calculate the perfusion fraction in IVIM, for example, has been shown in gliomas (179).

3.7.3 Evidence From Clinical Studies

The ADC and perfusion fraction (f) extracted using a previously validated simplified IVIM model (150, 180) were used to assess the

prediction of early progression in IDH-wildtype glioblastoma (181). Higher ADC_{T2-FLAIR} at baseline, lower $f_{T2-FLAIR}$ at the 10th (of 30) radiation fraction, and lack of increase in ADC_{T2-FLAIR} at the 20th radiation fraction compared with baseline, were associated with early progression. IVIM metrics were not associated with overall survival, while higher $f_{T2-FLAIR}$ at the 10th radiation fraction was associated with longer progression-free survival (181). In a study of 51 patients (182), a histogram analysis of IVIM metrics was able to differentiate true progression from pseudoprogression, with ROC analyses identifying the 90th percentile for perfusion to be discriminant with a sensitivity of 87.1%, and a specificity of 95.0%. A comparison study of different ADC modeling approaches found that all computed metrics, including those using the IVIM model, can differentiate tumor recurrence from pseudoprogression (100).

In a recent study of 32 glioblastomas, DKI showed high diagnostic accuracy, up to 88%, for differentiating between tumor progression and post-treatment related effects (183). In a comparison with DTI, DKI was found to be superior in predicting recurrence (184). DKI recently was applied to investigate treatment response and pseudoprogression in several studies (184–186) and these found that DKI was superior in assessing treatment response when compared to ADC and DTI. However, one of the studies found only limited benefit when compared to standard ADC measurements (185).

3.7.4 Future Developments

It is potentially possible to render a tumor angiogram and elastographic information based on IVIM and DKI measurements (178). Because vascular and elastographic tumor imaging are upcoming techniques in glioma imaging (187, 188), IVIM and DKI may be applied to such advanced applications. A more important development in the near future will be determing the optimal choice of the number of b values needed for a reliable fit for IVIM and DKI (189).

3.8 DTI

3.8.1 Methodology

By sampling the diffusion signal with at least six different directions, the information can be grouped into a so-called tensor, a multi-dimensional mathematical construct that adheres to mathematical operators (139) and can be used to determine diffusion magnitude (mean diffusivity) and degrees of diffusion anisotropy (fractional anisotropy [FA]). The estimated directionality can be visualized through post processing techniques using ellipsoids or fibers. DTI obtained by acquiring repeated DWI in different non-collinear directions can lead to some quantitative parameters, such as FA, mean diffusivity, axial diffusivity, and radial diffusivity. FA is an index between 0 and 1 to assess the degree of diffusion asymmetry in a voxel in terms of its eigenvalues. Because anatomical tracts are anisotropic, FA maps show where tracts are passing, or changing in disease, e.g., through destruction of these due to tumor cell invasion or sometimes treatment. A limitation of the diffusion tensor model is the assumption that the diffusion of water will always be Gaussian (190). This is true for diffusion within a singular environment (i.e., without different compartments), but

due to imaging voxels being relatively large, it rarely points to a complete picture in human tissue.

3.8.2 Strengths and Weaknesses

All scanner manufacturers have protocols to acquire DTI data and calculate mean diffusivity and FA maps as part of a postprocessing step on the console. However, there is no common framework and a lack of consensus regarding how many directions or which b-values shells should be chosen. As in DWI, EPI-related artifacts such as geometric distortions and signal loss present challenges in superimposing DTI-derived maps on structural imaging. Although tractography is currently less relevant as a monitoring biomarker and largely beyond the scope of this position statement, it is noted that some scanner manufacturers offer the ability to perform tractography. These algorithms are often a "black-box," making comparison and reproducibility difficult. Commercial and non-commercial software packages exist, employing both deterministic and probabilistic algorithms. It is noteworthy that these require considerable training and present challenges when integrating the process with clinical workflows.

3.8.3 Evidence From Clinical Studies

After standard-of-care radiotherapy and concomittant temozolomide, when compared with pseudoprogression, the FA in contrast-enhancing tumor was higher in true tumor progression (162, 191, 192) and even appears to increase when compared to previous measurements (163). Whilst higher FA in progression has been postulated to occur due to the orientation of overproduced extracellular matrix in glioblastoma, other authors reported no difference in FA when pseudoprogression and progression were compared (141, 193). DTI-based maps of tissue isotropy and anisotropy have also been successfully used to predict recurrence of glioblastoma (194).

3.8.4 Future Developments

Clinical studies aiming to optimize the multiple b-value shells and number of directions selected, are required before consensus standardization will support translation into the clinic routinely. This is being enabled by the dissemination of sequences and greater accessibility of post-processing pipelines without the need for advanced programming skills.

3.9 Other Advanced Diffusion Techniques 3.9.1 Methodology

Multiple other advanced MRI techniques are based on DWI. Many of them focus on modeling diffusion in different tissue environments to disentangle the signal from "hindered, restricted, and isotropic compartments" (195). The hypothesis is that these compartments reflect actual tissue compartments, such as the extra-axonal or extracellular space (hindered diffusion), intracellular, or intra-axonal space (restricted), and fluid, such as edema or cerebrospinal fluid (isotropic). Many of these models, such as the NODDI technique, are designed based on assumptions in healthy white matter (123). Therefore, their use may be limited in glioblastomas. The RSI model (124) offers parametric indices to be calculated in a similar way to NODDI, such as with neurite density and dispersion indices, but RSI does allow more flexibility that may make it more appropriate for tumor imaging. While the tissue in RSI is modeled as cylinders, similar to other DWImicrostructure techniques targeted at white matter, the relatively large spectrum of size and orientation modeled makes it sensitive to a large range of geometries beyond the cylindrical shape (124). In addition, removal of the isotropic signal contribution can aid in disentangling enhancing tumor volume from edema. Disentangling the isotropic compartment is also of interest in the FWI technique. Here, a modeling approach that is initialized with a constant diffusivity of free water can be used to create maps of free water as well as produce a diffusion tensor that has the isotropic signal removed. Integration with the tensor approach lends its application to improving tractography results in the presence of a conspicuous amount of edema.

VERDICT is a model that infers tumor cell microstructure from DWI measurements (126). This model derives multiple compartments (intracellular, intravascular and extracellular– extravascular spaces) and has recently been used to investigate treatment response in animal models of glioma (196, 197). It has subsequently been adapted in human glioblastoma studies in treatment-naive patients (198). The VERDICT model assumes tumor cells are spherical and can model both cell radii and densities, making it an attractive option for the use in treatment response monitoring.

3.9.2 Strengths and Weaknesses

Even though the advanced radiological imaging and multiparametric imaging approach is promising in the context of post-treatment related effects or specifically for pseudoprogression and radiation necrosis, efforts to establish uniform practices and protocols are again necessary. As in DWI and DTI, geometric distortions are also a challenge and there is no consensus on how many directions and which b-value shells are optimal with which particular microstructural technique. Other factors, such as the choice of TE and TR that is limited by either hardware or sequence, also add difficulty to the reproduction of results.

Many microstructural imaging approaches require lengthy protocols or uncommon scanner hardware options, such as parallel transmit or head coils with a large number of channels. In addition, the high b-values required often necessitate imaging close to the noise baseline, which requires lengthy averaging or noise modeling in post-processing, which is not a trivial challenge in clinical imaging.

3.9.3 Evidence From Clinical Studies

The evidence in clinical studies of brain tumor treatment response is very limited. However, FWI has most recently been used to successfully predict glioblastoma recurrence (199). Furthermore, RSI has been used to show treatment effects (135, 200, 201) as well as investigating overall and progressionfree survival in gliomas (202). NODDI has been used together with serial DTI imaging to observe tumor changes up to two months before recurrence (203). For completeness in describing these emerging techniques, it is also noted that RSI was used to differentiate true progression from immune-mediated pseudoprogression (**Figure 12**) in a proof-of-concept case





study. A 64-year-old man had follow-up imaging over several months. He developed pathologically proven pseudoprogression as well as a secondary primary lesion in a separate location. The authors found that RSI was able to differentiate true progression from pseudoprogression while ADC could not (204). However, whilst RSI is supported by marginally more clinical evidence than these other emerging techniques, it has yet to be adopted in large-scale glioblastoma research studies, thereby allowing the strengths of the technique to be demonstrated.

4 DISCUSSION

Contemporaneous, accurate, and reliable monitoring biomarkers are required for high-grade glioma treatment response assessment as the use of conventional structural MRI protocols is limited by important challenges. The current evidence regarding the potential for monitoring biomarkers based on advanced MRI techniques is reviewed, and the individual modalities of perfusion, permeability, and microstructural imaging are described (these will be complemented by modalities related to metabolism and/or chemical composition discussed in Part 2). Considerable developments have been made in advanced MRI methodology. Although some techniques have evolved and matured over three decades, several new state-of-the-art methods are poised to contribute to the imaging armamentarium. However, limitations remain for all techniques. Good-quality evidence of clinical diagnostic accuracy is typically lacking. Clinical implementation of standardized tools generally remains challenging, and some recent techniques are in their infancy. The readiness of individual modalities in terms of technical validation, clinical evidence, acceptance and implementation are summarized in Part 2.

High-grade glioma vasculature exhibits increased perfusion, blood volume, and permeability compared with normal brain tissue. Measures of CBV derived from DSC-MRI have consistently provided information about brain tumor growth and response to treatment; it is the most clinically validated advanced technique. Clinical studies have proven the potential of DCE-MRI for distinguishing post-treatment related effects from recurrence, but the optimal acquisition protocol, mode of analysis, and parameter of highest diagnostic value and optimal cut-off points remain to be established. These same challenges are present to some extent with other perfusion techniques. ASL techniques do not require the injection of a contrast agent, and fast repeated measurements can be performed. Moreover, the absence of potential gadolinium deposition effects allows widespread use in pediatric patients and those with impaired renal function. Although the current review is focused on adult high-grade glioma, several studies have shown the utility of ASL perfusion imaging in pediatric brain tumors (205). DWI, ADC, DTI, DKI, IVIM, and other microstructural modeling approaches also allow treatment response assessment; more robust data is required to validate these techniques alone or when applied to post-processing methodologies. (Post-processing methodologies involving the combination of MRI approaches [multiparametric imaging] or machine learning are summarized in Part 2). Finally, for all parametric imaging techniques, the clinical application of cutoffs based on the literature is complicated by differences in how a ROI is defined and what particular descriptive statistic value (maximum, mean, etc.) within the ROI is used. Using a fixed image analysis approach, a multi-center study did show that excellent agreement of cut-offs between sites and platforms may be achieved (57). Nevertheless, until methods are standardized, generally accepted cut-off values may be difficult to apply in the clinical setting. Alternatively, clinical departments may rely on replicating approaches for generation and analysis of parametric images described in individual studies reporting the cut-off used.

In conclusion, considerable progress has been made in the development of advanced MRI monitoring biomarkers. As we approach the mid-twenty-first century, the techniques will have matured and serve as robust clinic-ready assays for treatment response assessment in personalized medicine regimens, which themselves will evolve in parallel. For now, more research and collaboration are needed to provide standardized and evidencedbased tools for this uncommon but disproportionately devastating disease.

AUTHOR CONTRIBUTIONS

All authors have contributed to the conception of the two parts of the article, revised them critically and approved the submitted versions. Authors TB and OH served as overall editors. The individual sections were drafted by: Introduction (TB), DSC-MRI (KS and MÁ-T), DCE-MRI (OH), ASL (PF and VK), diffusion techniques (RN, FR, and VK), spectroscopy (ECW and GH), CEST (EAHW), multiparametric imaging (OH), clinical readiness (OH),

REFERENCES

- Ostrom QT, Gittleman H, Liao P, Rouse C, Chen Y, Dowling J, et al. CBTRUS Statistical Report: Primary Brain and Central Nervous System Tumors Diagnosed in the United States in 2007-2011. *Neuro Oncol* (2014) 16(Suppl 4):iv1–63. doi: 10.1093/neuonc/nou223
- Haider AS, van den Bent M, Wen PY, Vogelbaum MA, Chang S, Canoll PD, et al. Toward a Standard Pathological and Molecular Characterization of Recurrent Glioma in Adults: A Response Assessment in Neuro-Oncology Effort. *Neuro Oncol* (2020) 22:450–6. doi: 10.1093/neuonc/noz233
- Burnet NG, Jefferies SJ, Benson RJ, Hunt DP, Treasure FP. Years of Life Lost (YLL) From Cancer Is an Important Measure of Population Burden — and Should be Considered When Allocating Research Funds. *Br J Cancer* (2005) 92:241–5. doi: 10.1038/sj.bjc.6602321

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SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fonc.2022.810263/ full#supplementary-material

- Stupp R, Mason WP, van den Bent MJ, Weller M, Fisher B, Taphoorn MJB, et al. Radiotherapy Plus Concomitant and Adjuvant Temozolomide for Glioblastoma. N Engl J Med (2005) 352:987–96. doi: 10.1056/ NEJMoa043330
- Weller M, van den Bent M, Tonn JC, Stupp R, Preusser M, Cohen-Jonathan-Moyal E, et al. European Association for Neuro-Oncology (EANO) Guideline on the Diagnosis and Treatment of Adult Astrocytic and Oligodendroglial Gliomas. *Lancet Oncol* (2017) 18:e315–29. doi: 10.1016/ S1470-2045(17)30194-8
- Stupp R, Hegi ME, Mason WP, van den Bent MJ, Taphoorn MJB, Janzer RC, et al. Effects of Radiotherapy With Concomitant and Adjuvant Temozolomide Versus Radiotherapy Alone on Survival in Glioblastoma in a Randomised Phase III Study: 5-Year Analysis of the EORTC-NCIC Trial. *Lancet Oncol* (2009) 10:70025–7. doi: 10.1016/S1470-2045(09)70025-7

- Cagney DN, Sul J, Huang RY, Ligon KL, Wen PY, Alexander BM. The FDA NIH Biomarkers, EndpointS, and Other Tools (BEST) Resource in Neuro-Oncology. *Neuro Oncol* (2018) 20:1162–72. doi: 10.1093/neuonc/nox242
- Cao Y, Sundgren PC, Tsien CI, Chenevert TT, Junck L. Physiologic and Metabolic Magnetic Resonance Imaging in Gliomas. J Clin Oncol (2006) 24:1228–35. doi: 10.1200/JCO.2005.04.7233
- Wen PY, Macdonald DR, Reardon DA, Cloughesy TF, Sorensen AG, Galanis E, et al. Updated Response Assessment Criteria for High-Grade Gliomas: Response Assessment in Neuro-Oncology Working Group. J Clin Oncol (2010) 28:1963–72. doi: 10.1200/JCO.2009.26.3541
- Booth TC, Thompson G, Bulbeck H, Boele F, Buckley C, Cardoso J, et al. A Position Statement on the Utility of Interval Imaging in Standard of Care Brain Tumour Management: Defining the Evidence Gap and Opportunities for Future Research. *Front Oncol* (2021) 11:620070. doi: 10.3389/ fonc.2021.620070
- Ellingson BM, Harris RJ, Woodworth DC, Leu K, Zaw O, Mason WP, et al. Baseline Pretreatment Contrast Enhancing Tumor Volume Including Central Necrosis is a Prognostic Factor in Recurrent Glioblastoma: Evidence From Single and Multicenter Trials. *Neuro Oncol* (2017) 19:89– 98. doi: 10.1093/neuonc/now187
- Lacroix M, Abi-Said D, Fourney DR, Gokaslan ZL, Shi W, DeMonte F, et al. A Multivariate Analysis of 416 Patients With Glioblastoma Multiforme: Prognosis, Extent of Resection, and Survival. J Neurosurg (2001) 95:190–8. doi: 10.3171/jns.2001.95.2.0190
- Booth TC, Luis A, Brazil L, Thompson G, Daniel RA, Shuaib H, et al. Glioblastoma Post-Operative Imaging in Neuro-Oncology: Current UK Practice (GIN CUP Study). *Eur Radiol* (2021) 31:2933–43. doi: 10.1007/ s00330-020-07387-3
- Dhermain FG, Hau P, Lanfermann H, Jacobs AH, van den Bent MJ. Advanced MRI and PET Imaging for Assessment of Treatment Response in Patients With Gliomas. *Lancet Neurol* (2010) 9. doi: 10.1016/S1474-4422 (10)70181-2
- Booth TC, Williams M, Luis A, Cardoso J, Ashkan K, Shuaib H. Machine Learning and Glioma Imaging Biomarkers. *Clin Radiol* (2020) 75. doi: 10.1016/j.crad.2019.07.001
- Brandsma D, Stalpers L, Taal W, Sminia P, van den Bent MJ. Clinical Features, Mechanisms, and Management of Pseudoprogression in Malignant Gliomas. *Lancet Oncol* (2008) 9:453–61. doi: 10.1016/S1470-2045(08)70125-6
- Booth TC, Larkin TJ, Yuan Y, Kettunen MI, Dawson SN, Scoffings D, et al. Analysis of Heterogeneity in T2-Weighted MR Images can Differentiate Pseudoprogression From Progression in Glioblastoma. *PloS One* (2017) 12: e0176528. doi: 10.1371/journal.pone.0176528
- Radbruch A, Fladt J, Kickingereder P, Wiestler B, Nowosielski M, Bäumer P, et al. Pseudoprogression in Patients With Glioblastoma: Clinical Relevance Despite Low Incidence. *Neuro Oncol* (2015) 17:151–9. doi: 10.1093/neuonc/ nou129
- Booth TC, Waldman AD, Jefferies S, Jäger R. Comment on "The Role of Imaging in the Management of Progressive Glioblastoma. A Systematic Review and Evidence-Based Clinical Practice Guideline". J Neurooncol (2014) 118:435–60. doi: 10.1007/s11060-013-1330-0
- Dhawan S, Venteicher AS, Butler WE, Carter BS, Chen CC. Clinical Outcomes as a Function of the Number of Samples Taken During Stereotactic Needle Biopsies: A Meta-Analysis. J Neurooncol (2021) 154:1– 11. doi: 10.1007/s11060-021-03785-9
- Müller DMJ, Robe PAJT, Eijgelaar RS, Witte MG, Visser M, de Munck JC, et al. Comparing Glioblastoma Surgery Decisions Between Teams Using Brain Maps of Tumor Locations, Biopsies, and Resections. JCO Clin Cancer Inf (2019) 3:1–12. doi: 10.1200/CCI.18.00089
- Müller Bark J, Kulasinghe A, Chua B, Day BW, Punyadeera C. Circulating Biomarkers in Patients With Glioblastoma. *Br J Cancer* (2020) 122:295–305. doi: 10.1038/s41416-019-0603-6
- Morokoff A, Jones J, Nguyen H, Ma C, Lasocki A, Gaillard F, et al. Serum microRNA is a Biomarker for Post-Operative Monitoring in Glioma. *J Neurooncol* (2020) 149:391–400. doi: 10.1007/s11060-020-03566-w
- Lawrie TA, Kernohan A, Jenkinson MD, Thompson G. Interval Brain Imaging for Adults With Cerebral Glioma. *Cochrane Database Syst Rev* (2018) 12:CD013137. doi: 10.1002/14651858.CD013137.pub2

- The National Institute for Health and Care Excellence. Guideline Ng99. Available at: https://www.nice.org.uk/guidance/ng99 (Accessed 1 March 2019).
- The James Lind Alliance Priority Setting Partnerships . Available at: http://: www.jla.nihr.ac.uk/priority-setting-partnerships/neuro-oncology/ (Accessed 1 August 2018).
- Thust SC, Heiland S, Falini A, Jäger HR, Waldman AD, Sundgren PC, et al. Glioma Imaging in Europe: A Survey of 220 Centres and Recommendations for Best Clinical Practice. *Eur Radiol* (2018) 28:3306–31. doi: 10.1007/ s00330-018-5314-5
- Clement P, Booth T, Borovečki F, Emblem KE, Figueiredo P, Hirschler L, et al. GliMR: Cross-Border Collaborations to Promote Advanced MRI Biomarkers for Glioma. J Med Biol Eng (2021) 41:1–11. doi: 10.1007/ s40846-020-00582-z
- European Medicine Agency. *Refusal Assessment Report for Avastin* (2010). Available at: https://www.ema.europa.eu/en/documents/variation- report/ avastin-h-c-582-ii-0028-epar-refusal-assessment-report-variation_en.pdf (Accessed 15 April 2019).
- 30. Europarat C of E. Developing a Methodology for Drawing Up Guidelines on Best Medical Practices - Recommendation Rec13 and Explanatory Memorandum (2002). Strasbourg: Council of Europe Publications (2002).
- McInnes MDF, Moher D, Thombs BD, McGrath TA, Bossuyt PMthe PRISMA-DTA Group, et al. Preferred Reporting Items for a Systematic Review and Meta-Analysis of Diagnostic Test Accuracy Studies: The PRISMA-DTA Statement. *JAMA* (2018) 319:388–96. doi: 10.1001/ jama.2017.19163
- Hanahan D, Weinberg RA. Hallmarks of Cancer: The Next Generation. Cell (2011) 144:646–74. doi: 10.1016/j.cell.2011.02.013
- Ahir BK, Engelhard HH, Lakka SS. Tumor Development and Angiogenesis in Adult Brain Tumor: Glioblastoma. *Mol Neurobiol* (2020) 57:2461–78. doi: 10.1007/s12035-020-01892-8
- Michael Peters A. The Precise Physiological Definition of Tissue Perfusion and Clearance Measured From Imaging. *Eur J Nucl Med Mol Imaging* (2018) 45:1139–41. doi: 10.1007/s00259-018-3982-7
- 35. Jezzard P, Chappell MA, Okell TW. Arterial Spin Labeling for the Measurement of Cerebral Perfusion and Angiography. J Cereb Blood Flow Metab (2018) 38:603–26. doi: 10.1177/0271678X17743240
- Rosen BR, Belliveau JW, Vevea JM, Brady TJ. Perfusion Imaging With NMR Contrast Agents. *Magn Reson Med* (1990) 14:249–65. doi: 10.1002/ mrm.1910140211
- Boxerman JL, Bandettini PA, Kwong KK, Baker JR, Davis TL, Rosen BR, et al. The Intravascular Contribution to fMRI Signal Change: Monte Carlo Modeling and Diffusion-Weighted Studies *In Vivo. Magn Reson Med* (1995) 34:4–10. doi: 10.1002/mrm.1910340103
- Perkio J, Aronen HJ, Kangasmaki A, Liu Y, Karonen J, Savolainen S, et al. Evaluation of Four Postprocessing Methods for Determination of Cerebral Blood Volume and Mean Transit Time by Dynamic Susceptibility Contrast Imaging. *Magn Reson Med* (2002) 47:973–81. doi: 10.1002/mrm.10126
- Bedekar D, Jensen T, Schmainda KM. Standardization of Relative Cerebral Blood Volume (rCBV) Image Maps for Ease of Both Inter- and Intrapatient Comparisons. Magn Reson Med (2010) 64:907–13. doi: 10.1002/mrm.22445
- Prah MA, Stufflebeam SM, Paulson ES, Kalpathy-Cramer J, Gerstner ER, Batchelor TT, et al. Repeatability of Standardized and Normalized Relative CBV in Patients With Newly Diagnosed Glioblastoma. *Am J Neuroradiol* (2015) 36:1654–61. doi: 10.3174/ajnr.A4374
- 41. Schmainda KM, Zhang Z, Prah M, Snyder BS, Gilbert MR, Sorensen AG, et al. Dynamic Susceptibility Contrast MRI Measures of Relative Cerebral Blood Volume as a Prognostic Marker for Overall Survival in Recurrent Glioblastoma: Results From the ACRIN 6677/RTOG 0625 Multicenter Trial. *Neuro Oncol* (2015) 17:1148–56. doi: 10.1093/neuonc/nou364
- Gerstner ER, Zhang Z, Fink JR, Muzi M, Hanna L, Greco E, et al. ACRIN 6684: Assessment of Tumor Hypoxia in Newly Diagnosed Glioblastoma Using 18f-FMISO PET and MRI. *Clin Cancer Res* (2016) 22:5079–86. doi: 10.1158/1078-0432.CCR-15-2529
- 43. Schmainda KM, Prah MA, Marques H, Kim E, Barboriak DP, Boxerman JL. Value of Dynamic Contrast Perfusion MRI to Predict Early Response to Bevacizumab in Newly Diagnosed Glioblastoma: Results From ACRIN 6686

Multi-Center Trial. Neuro Oncol (2020) 23:314–23. doi: 10.1093/neuonc/ noaa167

- 44. Paulson ES, Prah DE, Schmainda KM. Spiral Perfusion Imaging With Consecutive Echoes (SPICE) for the Simultaneous Mapping of DSC- and DCE-MRI Parameters in Brain Tumor Patients: Theory and Initial Feasibility. *Tomography* (2016) 2:295–307. doi: 10.18383/j.tom.2016.00217
- 45. Donahue KM, Krouwer HGJ, Rand SD, Pathak AP, Marszalkowski CS, Censky SC, et al. Utility of Simultaneously Acquired Gradient-Echo and Spin-Echo Cerebral Blood Volume and Morphology Maps in Brain Tumor Patients. *Magn Reson Med* (2000) 43:845–53. doi: 10.1002/1522-2594 (200006)43:6<845::AID-MRM10>3.0.CO;2-J
- 46. Schmainda KM, Rand SD, Joseph AM, Lund R, Ward BD, Pathak AP, et al. Characterization of a First-Pass Gradient-Echo Spin-Echo Method to Predict Brain Tumor Grade and Angiogenesis. *Am J Neuroradiol* (2004) 25:1524–32.
- 47. Hu LS, Baxter LC, Pinnaduwage DS, Paine TL, Karis JP, Feuerstein BG, et al. Optimized Preload Leakage-Correction Methods to Improve the Diagnostic Accuracy of Dynamic Susceptibility-Weighted Contrast-Enhanced Perfusion MR Imaging in Posttreatment Gliomas. Am J Neuroradiol (2010) 31:40–8. doi: 10.3174/ajnr.A1787
- Boxerman JL, Schmainda KM, Weisskoff RM. Relative Cerebral Blood Volume Maps Corrected for Contrast Agent Extravasation Significantly Correlate With Glioma Tumor Grade, Whereas Uncorrected Maps do Not. *Am J Neuroradiol* (2006) 27:859–67.
- Bjornerud A, Sorensen AG, Mouridsen K, Emblem KE. T1- and T2*-Dominant Extravasation Correction in DSC-MRI: Part I-theoretical Considerations and Implications for Assessment of Tumor Hemodynamic Properties. J Cereb Blood Flow Metab (2011) 31:2041–53. doi: 10.1038/ jcbfm.2011.52
- Quarles CC, Ward BD, Schmainda KM. Improving the Reliability of Obtaining Tumor Hemodynamic Parameters in the Presence of Contrast Agent Extravasation. *Magn Reson Med* (2005) 53:1307–16. doi: 10.1002/ mrm.20497
- 51. Leu K, Boxerman JL, Cloughesy TF, Lai A, Nghiemphu PL, Liau LM, et al. Improved Leakage Correction for Single-Echo Dynamic Susceptibility Contrast Perfusion MRI Estimates of Relative Cerebral Blood Volume in High-Grade Gliomas by Accounting for Bidirectional Contrast Agent Exchange. AJNR Am J Neuroradiol (2016) 37:1440–6. doi: 10.3174/ ajnr.A4759
- 52. Semmineh NB, Bell LC, Stokes AM, Hu LS, Boxerman JL, Quarles CC. Optimization of Acquisition and Analysis Methods for Clinical Dynamic Susceptibility Contrast MRI Using a Population-Based Digital Reference Object. Am J Neuroradiol (2018) 39:1981–8. doi: 10.3174/ajnr.A5827
- Semmineh NB, Stokes AM, Bell LC, Boxerman JL, Quarles CC. A Population-Based Digital Reference Object (DRO) for Optimizing Dynamic Susceptibility Contrast (DSC)-MRI Methods for Clinical Trials. *Tomography* (2017) 3:41–9. doi: 10.18383/j.tom.2016.00286
- Schmainda KM, Prah MA, Hu LS, Quarles CC, Semmineh N, Rand SD, et al. Moving Toward a Consensus DSC-MRI Protocol: Validation of a Low-Flip Angle Single-Dose Option as a Reference Standard for Brain Tumors. *Am J Neuroradiol* (2019) 40:626–33. doi: 10.3174/ajnr.A6015
- 55. Boxerman JL, Quarles CC, Hu LS, Erickson BJ, Gerstner ER, Smits M, et al. Consensus Recommendations for a Dynamic Susceptibility Contrast MRI Protocol for Use in High-Grade Gliomas. *Neuro Oncol* (2020) 22:1262–75. doi: 10.1093/neuonc/noaa141
- Patel P, Baradaran H, Delgado D, Askin G, Christos P, John Tsiouris A, et al. MR Perfusion-Weighted Imaging in the Evaluation of High-Grade Gliomas After Treatment: A Systematic Review and Meta-Analysis. *Neuro Oncol* (2017) 19:118–27. doi: 10.1093/neuonc/now148
- 57. Schmainda KM, Prah MA, Rand SD, Liu Y, Logan B, Muzi M, et al. Multisite Concordance of DSC-MRI Analysis for Brain Tumors: Results of a National Cancer Institute Quantitative Imaging Network Collaborative Project. *AJNR Am J Neuroradiol* (2018) 39:1008–16. doi: 10.3174/ajnr.A5675
- Hu LS, Eschbacher JM, Dueck AC, Heiserman JE, Liu S, Karis JP, et al. Correlations Between Perfusion MR Imaging Cerebral Blood Volume, Microvessel Quantification, and Clinical Outcome Using Stereotactic Analysis in Recurrent High-Grade Glioma. *Am J Neuroradiol* (2012) 33:69–76. doi: 10.3174/ajnr.A2743

- Prah MA, Al-Gizawiy MM, Mueller WM, Cochran EJ, Hoffmann RG, Connelly JM, et al. Spatial Discrimination of Glioblastoma and Treatment Effect With Histologically-Validated Perfusion and Diffusion Magnetic Resonance Imaging Metrics. J Neurooncol (2018) 136:13–21. doi: 10.1007/ s11060-017-2617-3
- 60. Hu LS, Eschbacher JM, Heiserman JE, Dueck AC, Shapiro WR, Liu S, et al. Reevaluating the Imaging Definition of Tumor Progression: Perfusion MRI Quantifies Recurrent Glioblastoma Tumor Fraction, Pseudoprogression, and Radiation Necrosis to Predict Survival. *Neuro Oncol* (2012) 14:919– 30. doi: 10.1093/neuonc/nos112
- Prah MA, Connelly JM, Schmainda KM. MRI-Perfusion Derived Fractional Tumor Burden (FTB) is Predictive of Overall and Progression Free Survival in Newly Diagnosed Glioblastoma Following Concomitant Chemoradiotherapy. Proc Int Soc Magnetic Resonance Med (Honolulu Hawaii) 707.
- 62. Iv M, Liu X, Lavezo J, Gentles AJ, Ghanem R, Lummus S, et al. Perfusion MRI-Based Fractional Tumor Burden Differentiates Between Tumor and Treatment Effect in Recurrent Glioblastomas and Informs Clinical Decision-Making. *AJNR Am J Neuroradiol* (2019) 40:1649–57. doi: 10.3174/ ajnr.A6211
- 63. Hoxworth JM, Eschbacher JM, Gonzales AC, Singleton KW, Leon GD, Smith KA, et al. Performance of Standardized Relative CBV for Quantifying Regional Histologic Tumor Burden in Recurrent High-Grade Glioma: Comparison Against Normalized Relative CBV Using Image-Localized Stereotactic Biopsies. *AJNR Am J Neuroradiol* (2020) 41:408–15. doi: 10.3174/ajnr.A6486
- 64. Quarles CC, Schmainda KM. Assessment of the Morphological and Functional Effects of the Anti-Angiogenic Agent SU11657 on 9L Gliosarcoma Vasculature Using Dynamic Susceptibility Contrast MRI. Magn Reson Med (2007) 57:680–7. doi: 10.1002/mrm.21193
- Bonekamp D, Mouridsen K, Radbruch A, Kurz FT, Eidel O, Wick A, et al. Assessment of Tumor Oxygenation and its Impact on Treatment Response in Bevacizumab-Treated Recurrent Glioblastoma. J Cereb Blood Flow Metab (2017) 37:485–94. doi: 10.1177/0271678X16630322
- Stokes AM, Quarles CC. A Simplified Spin and Gradient Echo Approach for Brain Tumor Perfusion Imaging. *Magn Reson Med* (2016) 75:356–62. doi: 10.1002/mrm.25591
- Quarles CC, Gore JC, Xu L, Yankeelov TE. Comparison of Dual-Echo DSC-MRI- and DCE-MRI-Derived Contrast Agent Kinetic Parameters. *Magn Reson Imaging* (2012) 30:944–53. doi: 10.1016/j.mri.2012.03.008
- Han M, Yang B, Fernandez B, Lafontaine M, Alcaide-Leon P, Jakary A, et al. Simultaneous Multi-Slice Spln- and Gradient-Echo Dynamic Susceptibility Perfusion-Weighted MRI of Gliomas. *Nmr BioMed* (2021) 34:e4399. doi: 10.1002/nbm.4399
- 69. Bell LC, Semmineh N, An H, Eldeniz C, Wahl R, Schmainda KM, et al. Evaluating the Use of rCBV as a Tumor Grade and Treatment Response Classifier Across NCI Quantitative Imaging Network Sites: Part II of the DSC-MRI Digital Reference Object (DRO) Challenge. *Tomogr (Ann Arbor Mich)* (2020) 6:203–8. doi: 10.18383/j.tom.2020.00012
- Skinner JT, Robison RK, Elder CP, Newton AT, Damon BM, Quarles CC. Evaluation of a Multiple Spin- and Gradient-Echo (SAGE) EPI Acquisition With SENSE Acceleration: Applications for Perfusion Imaging in and Outside the Brain. *Magn Reson Imaging* (2014) 32:1171–80. doi: 10.1016/ j.mri.2014.08.032
- Sourbron SP, Buckley DL. Classic Models for Dynamic Contrast-Enhanced MRI. NMR BioMed (2013) 26:1004–27. doi: 10.1002/nbm.2940
- 72. Tofts PS, Brix G, Buckley DL, Evelhoch JL, Henderson E, Knopp MV, et al. Estimating Kinetic Parameters From Dynamic Contrast-Enhanced T1-Weighted MRI of a Diffusable Tracer: Standardized Quantities and Symbols. J Magn Reson Imaging (1999) 10:223–32. doi: 10.1002/(SICI) 1522-2586(199909)10:3<223::AID-JMRI2>3.0.CO;2-S
- QIBA Profile: 4 DCE-MRI Quantification (DCEMRI-Q). Available at: http:// qibawiki.rsna.org/images/1/1f/QIBA_DCE-MRI_Profile-Stage_1-Public_ Comment.pdf ref. accessed Feb 1, 2021
- Luypaert R, Sourbron S, de Mey J. Validity of Perfusion Parameters Obtained Using the Modified Tofts Model: A Simulation Study. *Magn Reson Med* (2011) 65:1491–7. doi: 10.1002/mrm.22728
- 75. Artzi M, Liberman G, Blumenthal DT, Bokstein F, Aizenstein O, Ben Bashat D. Repeatability of Dynamic Contrast Enhanced Vp Parameter in Healthy

Subjects and Patients With Brain Tumors. *J Neurooncol* (2018) 140:727–37. doi: 10.1007/s11060-018-03006-w

- Schmainda KM, Prah MA, Marques H, Kim E, Barboriak DP, Boxerman JL. Value of Dynamic Contrast Perfusion MRI to Predict Early Response to Bevacizumab in Newly Diagnosed Glioblastoma: Results From ACRIN 6686 Multicenter Trial. *Neuro Oncol* (2021) 23:314–32. doi: 10.1093/neuonc/ noaa167
- 77. Bane O, Hectors SJ, Wagner M, Arlinghaus LL, Aryal MP, Cao Y, et al. Accuracy, Repeatability, and Interplatform Reproducibility of T₁ Quantification Methods Used for DCE-MRI: Results From a Multicenter Phantom Study. *Magn Reson Med* (2018) 79:2564–75. doi: 10.1002/ mrm.26903
- Conte GM, Castellano A, Altabella L, Iadanza A, Cadioli M, Falini A, et al. Reproducibility of Dynamic Contrast-Enhanced MRI and Dynamic Susceptibility Contrast MRI in the Study of Brain Gliomas: A Comparison of Data Obtained Using Different Commercial Software. *Radiol Med* (2017) 122. doi: 10.1007/s11547-016-0720-8
- Anzalone N, Castellano A, Cadioli M, Conte GM, Cuccarini V, Bizzi A, et al. Brain Gliomas: Multicenter Standardized Assessment of Dynamic Contrast-Enhanced and Dynamic Susceptibility Contrast MR Images. *Radiology* (2018) 287:933–94. doi: 10.1148/radiol.2017170362
- Larsson HBW, Hansen AE, Berg HK, Rostrup E, Haraldseth O. Dynamic Contrast-Enhanced Quantitative Perfusion Measurement of the Brain Usingt1-Weighted MRI at 3T. J Magn Reson Imaging (2008) 27:754–62. doi: 10.1002/jmri.21328
- Sourbron S, Ingrisch M, Siefert A, Reiser M, Herrmann K. Quantification of Cerebral Blood Flow, Cerebral Blood Volume, and Blood-Brain-Barrier Leakage With DCE-MRI. *Magn Reson Med* (2009) 62:205–17. doi: 10.1002/mrm.22005
- Larsson HBW, Courivaud F, Rostrup E, Hansen AE. Measurement of Brain Perfusion, Blood Volume, and Blood-Brain Barrier Permeability, Using Dynamic Contrast-Enhanced T₁-Weighted MRI at 3 Tesla. *Magn Reson Med* (2009) 62:1270–81. doi: 10.1002/mrm.22136
- Narang J, Jain R, Arbab AS, Mikkelsen T, Scarpace L, Rosenblum ML, et al. Differentiating Treatment-Induced Necrosis From Recurrent/Progressive Brain Tumor Using Nonmodel-Based Semiquantitative Indices Derived From Dynamic Contrast-Enhanced T1-Weighted MR Perfusion. *Neuro Oncol* (2011) 13:1037–46. doi: 10.1093/neuonc/nor075
- 84. Suh CH, Kim HS, Choi YJ, Kim N, Kim SJ. Prediction of Pseudoprogression in Patients With Glioblastomas Using the Initial and Final Area Under the Curves Ratio Derived From Dynamic Contrast-Enhanced T1-Weighted Perfusion MR Imaging. *Am J Neuroradiol* (2013) 34:2278–86. doi: 10.3174/ajnr.A3634
- Henriksen OM, Larsen VA, Muhic A, Hansen AE, Larsson HBW, Poulsen HS, et al. Simultaneous Evaluation of Brain Tumour Metabolism, Structure and Blood Volume Using [18F]-Fluoroethyltyrosine (FET) PET/MRI: Feasibility, Agreement and Initial Experience. Eur J Nucl Med Mol Imaging (2016) 43:103–12. doi: 10.1007/s00259-015-3183-6
- Heo YJ, Kim HS, Park JE, Choi C-G, Kim SJ. Uninterpretable Dynamic Susceptibility Contrast-Enhanced Perfusion MR Images in Patients With Post-Treatment Glioblastomas: Cross-Validation of Alternative Imaging Options. *PloS One* (2015) 10:e0136380. doi: 10.1371/journal.pone.0136380
- Lundemann M, Munck af Rosenschöld P, Muhic A, Larsen VA, Poulsen HS, Engelholm S-A, et al. Feasibility of Multi-Parametric PET and MRI for Prediction of Tumour Recurrence in Patients With Glioblastoma. *Eur J Nucl Med Mol Imaging* (2019) 46:603–13. doi: 10.1007/s00259-018-4180-3
- Okuchi S, Rojas-Garcia A, Ulyte A, Lopez I, Ušinskienė J, Lewis M, et al. Diagnostic Accuracy of Dynamic Contrast-Enhanced Perfusion MRI in Stratifying Gliomas: A Systematic Review and Meta-Analysis. *Cancer Med* (2019) 8:5564–73. doi: 10.1002/cam4.2369
- Park JE, Kim HS, Goh MJ, Kim SJ, Kim JH. Pseudoprogression in Patients With Glioblastoma: Assessment by Using Volume-Weighted Voxel-Based Multiparametric Clustering of MR Imaging Data in an Independent Test Set. *Radiology* (2015) 275:792–802. doi: 10.1148/radiol.14141414
- Kim HS, Goh MJ, Kim N, Choi CG, Kim SJ, Kim JH. Which Combination of MR Imaging Modalities Is Best for Predicting Recurrent Glioblastoma? Study of Diagnostic Accuracy and Reproducibility. *Radiology* (2014) 273:831–43. doi: 10.1148/radiol.14132868

- 91. Zakhari N, Taccone MS, Torres CH, Chakraborty S, Sinclair J, Woulfe J, et al. Prospective Comparative Diagnostic Accuracy Evaluation of Dynamic Contrast-Enhanced (DCE) vs. Dynamic Susceptibility Contrast (DSC) MR Perfusion in Differentiating Tumor Recurrence From Radiation Necrosis in Treated High-Grade Gliomas. J Magn Reson Imaging (2019) 50:573–82. doi: 10.1002/jmri.26621
- 92. Hatzoglou V, Yang TJ, Omuro A, Gavrilovic I, Ulaner G, Rubel J, et al. A Prospective Trial of Dynamic Contrast-Enhanced MRI Perfusion and Fluorine-18 FDG PET-CT in Differentiating Brain Tumor Progression From Radiation Injury After Cranial Irradiation. *Neuro Oncol* (2016) 18:873–80. doi: 10.1093/neuonc/nov301
- 93. Park YW, Ahn SS, Kim EH, Kang S-G, Chang JH, Kim SH, et al. Differentiation of Recurrent Diffuse Glioma From Treatment-Induced Change Using Amide Proton Transfer Imaging: Incremental Value to Diffusion and Perfusion Parameters. *Neuroradiology* (2021) 63:363–72. doi: 10.1007/s00234-020-02542-5
- Fussell D, Young RJ. Role of MRI Perfusion in Improving the Treatment of Brain Tumors. *Imaging Med* (2013) 5:407–26. doi: 10.2217/iim.13.50
- 95. Alsop DC, Detre JA, Golay X, Günther M, Hendrikse J, Hernandez-Garcia L, et al. Recommended Implementation of Arterial Spin-Labeled Perfusion Mri for Clinical Applications: A Consensus of the ISMRM Perfusion Study Group and the European Consortium for ASL in Dementia. *Magn Reson Med* (2015) 73:102–16. doi: 10.1002/mrm.25197
- 96. van Osch MJP, Teeuwisse WM, Chen Z, Suzuki Y, Helle M, Schmid S. Advances in Arterial Spin Labelling MRI Methods for Measuring Perfusion and Collateral Flow. J Cereb Blood Flow Metab (2018) 38:1461–80. doi: 10.1177/0271678X17713434
- Williams DS, Detre JA, Leigh JS, Koretsky AP. Magnetic Resonance Imaging of Perfusion Using Spin Inversion of Arterial Water. *Proc Natl Acad Sci USA* (1992) 89:212–6. doi: 10.1073/pnas.89.1.212
- Kwong KK, Chesler DA, Weisskoff RM, Donahue KM, Davis TL, Ostergaard L, et al. Mr Perfusion Studies With T1-Weighted Echo Planar Imaging. *Magn Reson Med* (1995) 34:878–87. doi: 10.1002/mrm.1910340613
- Kim S-G. Quantification of Relative Cerebral Blood Flow Change by Flow-Sensitive Alternating Inversion Recovery (FAIR) Technique: Application to Functional Mapping. *Magn Reson Med* (1995) 34:293–301. doi: 10.1002/ MRM.1910340303
- 100. Liao D, Liu Y, Tang B, Wang D, Li X, Dai B, et al. Comparison of DWI Based on Monoexponential, Biexponential and Stretched-Exponential Models in Differentiating Tumor Recurrence and Pseudoprogression of Glioblastoma. *Chin J Med Imaging Technol* (2019) 35:1450–5. doi: 10.13929/j.1003-3289.201903059
- 101. Wong EC, Cronin M, Wu W-C, Inglis B, Frank LR, Liu TT. Velocity-Selective Arterial Spin Labeling. *Magn Reson Med* (2006) 55:1334–41. doi: 10.1002/MRM.20906
- 102. Wong EC, Buxton RB, Frank LR. Implementation of Quantitative Perfusion Imaging Techniques for Functional Brain Mapping Using Pulsed Arterial Spin Labeling. NMR BioMed (1997) 10:237–49. doi: 10.1002/(SICI)1099-1492(199706/08)10:4/5<237::AID-NBM475>3.0.CO;2-X
- 103. Luh W-M, Wong EC, Bandettini PA, Hyde JS. QUIPSS II With Thin-Slice TI 1 Periodic Saturation: A Method for Improving Accuracy of Quantitative Perfusion Imaging Using Pulsed Arterial Spin Labeling (1999). Available at: http://citeseerx.ist.psu.edu/viewdoc/download?doi=10.1.1.139.499&rep= rep1&type=pdf (Accessed December 7, 2018).
- 104. Woods JG, Chappell MA, Okell TW. A General Framework for Optimizing Arterial Spin Labeling MRI Experiments. *Magn Reson Med* (2018) 81:2474– 248. doi: 10.1002/mrm.27580
- 105. Santos N, Sanches JM, Sousa I, Figueiredo P. Optimal Sampling and Estimation in PASL Perfusion Imaging. *IEEE Trans BioMed Eng* (2011) 58:3165–74. doi: 10.1109/TBME.2011.2164916
- 106. Teeuwisse WM, Schmid S, Ghariq E, Veer IM, Van Osch MJP. Time-Encoded Pseudocontinuous Arterial Spin Labeling: Basic Properties and Timing Strategies for Human Applications. *Magn Reson Med* (2014) 72:1712–22. doi: 10.1002/mrm.25083
- 107. Buxton RB, Frank LR, Wong EC, Siewert B, Warach S, Edelman RR. A General Kinetic Model for Quantitative Perfusion Imaging With Arterial Spin Labeling. *Magn Reson Med* (1998) 40:383–96. doi: 10.1002/ mrm.1910400308

- 108. Lawrence KS St, Frank JA, McLaughlin AC. Effect of Restricted Water Exchange on Cerebral Blood Flow Values Calculated With Arterial Spin Tagging: A Theoretical Investigation. *Magn Reson Med* (2000) 44:440–9. doi: 10.1002/1522-2594(200009)44:3<440::AID-MRM15>3.0.CO;2-6
- 109. Parkes LM, Tofts PS. Improved Accuracy of Human Cerebral Blood Perfusion Measurements Using Arterial Spin Labeling: Accounting for Capillary Water Permeability. *Magn Reson Med* (2002) 48:27-41. doi: 10.1002/mrm.10180
- 110. Chappell MA, MacIntosh BJ, Donahue MJ, Günther M, Jezzard P, Woolrich MW. Separation of Macrovascular Signal in Multi-Inversion Time Arterial Spin Labelling MRI. *Magn Reson Med* (2010) 63:1357–65. doi: 10.1002/ mrm.22320
- 111. Chappell MA, Groves AR, MacIntosh BJ, Donahue MJ, Jezzard P. Woolrich MW. Partial Volume Correction of Multiple Inversion Time Arterial Spin Labeling MRI Data. *Magn Reson Med* (2011) 65:1173–83. doi: 10.1002/ mrm.22641
- 112. Chappell MA, Woolrich MW, Kazan S, Jezzard P, Payne SJ, MacIntosh BJ. Modeling Dispersion in Arterial Spin Labeling: Validation Using Dynamic Angiographic Measurements. *Magn Reson Med* (2013) 69:563–70. doi: 10.1002/mrm.24260
- 113. Pinto J, Chappell MA, Okell TW, Mezue M, Segerdahl AR, Tracey I, et al. Calibration of Arterial Spin Labeling Data—Potential Pitfalls in Post-Processing. *Magn Reson Med* (2020) 83:1222–34. doi: 10.1002/mrm.28000
- 114. Baas KPA, Petr J, Kuijer JPA, Nederveen AJ, Mutsaerts HJMM, van de Ven KCC. Effects of Acquisition Parameter Modifications and Field Strength on the Reproducibility of Brain Perfusion Measurements Using Arterial Spin-Labeling. Am J Neuroradiol (2021) 42:109–15. doi: 10.3174/ajnr.A6856
- 115. Almeida JRC, Greenberg T, Lu H, Chase HW, Fournier J, Cooper CM, et al. Test-Retest Reliability of Cerebral Blood Flow in Healthy Individuals Using Arterial Spin Labeling: Findings From the EMBARC Study. *Magn Reson Imaging* (2018) 45:26–33. doi: 10.1016/j.mri.2017.09.004
- 116. Mezue M, Segerdahl AR, Okell TW, Chappell MA, Kelly ME, Tracey I. Optimization and Reliability of Multiple Postlabeling Delay Pseudo-Continuous Arterial Spin Labeling During Rest and Stimulus-Induced Functional Task Activation. J Cereb Blood Flow Metab (2014) 34:1919–27. doi: 10.1038/jcbfm.2014.163
- 117. Sousa I, Vilela P, Figueiredo P. Reproducibility of the Quantification of Arterial and Tissue Contributions in Multiple Postlabeling Delay Arterial Spin Labeling. J Magn Reson Imaging (2014) 40:1453–62. doi: 10.1002/ jmri.24493
- 118. Mutsaerts HJMM, van Osch MJP, Zelaya FO, Wang DJJ, Nordhøy W, Wang Y, et al. Multi-Vendor Reliability of Arterial Spin Labeling Perfusion MRI Using a Near-Identical Sequence: Implications for Multi-Center Studies. *Neuroimage* (2015) 113:143–52. doi: 10.1016/j.neuroimage.2015.03.043
- 119. Wang L, Wei L, Wang J, Li N, Gao Y, Ma H, et al. Evaluation of Perfusion MRI Value for Tumor Progression Assessment After Glioma Radiotherapy. *Med (Baltimore)* (2020) 99:e23766. doi: 10.1097/MD.00000000023766
- 120. Wang YL, Chen S, Xiao HF, Li Y, Wang Y, Liu G, et al. Differentiation Between Radiation-Induced Brain Injury and Glioma Recurrence Using 3D pCASL and Dynamic Susceptibility Contrast-Enhanced Perfusion-Weighted Imaging. *Radiother Oncol* (2018) 129:68–74. doi: 10.1016/j.radonc.2018.01.009
- 121. Razek KAAA, El-Serougy L, Abdelsalam M, Gaballa G, Talaat M. Differentiation of Residual/Recurrent Gliomas From Postradiation Necrosis With Arterial Spin Labeling and Diffusion Tensor Magnetic Resonance Imaging-Derived Metrics. *Neuroradiology* (2018) 60:169–77. doi: 10.1007/s00234-017-1955-3
- 122. Pellerin A, Khalifé M, Sanson M, Rozenblum-Beddok L, Bertaux M, Soret M, et al. Simultaneously Acquired PET and ASL Imaging Biomarkers may be Helpful in Differentiating Progression From Pseudo-Progression in Treated Gliomas. *Eur Radiol* (2021) 2021:1–11. doi: 10.1007/S00330-021-07732-0
- 123. Zhang H, Schneider T, Wheeler-Kingshott CA, Alexander DC. NODDI: Practical *In Vivo* Neurite Orientation Dispersion and Density Imaging of the Human Brain. *Neuroimage* (2012) 61:1000–16. doi: 10.1016/ j.neuroimage.2012.03.072
- 124. White NS, Leergaard TB, D'Arceuil H, Bjaalie JG, Dale AM. Probing Tissue Microstructure With Restriction Spectrum Imaging: Histological and Theoretical Validation. *Hum Brain Mapp* (2013) 34:327–46. doi: 10.1002/ hbm.21454

- Pasternak O, Sochen N, Gur Y, Intrator N, Assaf Y. Free Water Elimination and Mapping From Diffusion MRI. *Magn Reson Med* (2009) 62:717–30. doi: 10.1002/mrm.22055
- 126. Panagiotaki E, Walker-Samuel S, Siow B, Johnson SP, Rajkumar V, Pedley RB, et al. Noninvasive Quantification of Solid Tumor Microstructure Using VERDICT MRI. *Cancer Res* (2014) 74:1902–12. doi: 10.1158/0008-5472.CAN-13-2511
- 127. Stejskal EO, Tanner JE. Spin Diffusion Measurements: Spin Echoes in the Presence of a Time-Dependent Field Gradient. J Chem Phys (1965) 42:288– 92. doi: 10.1063/1.1695690
- 128. Moseley ME, Cohen Y, Kucharczyk J, Mintorovitch J, Asgari HS, Wendland MF, et al. Diffusion-Weighted MR Imaging of Anisotropic Water Diffusion in Cat Central Nervous System. *Radiology* (1990) 176:439–45. doi: 10.1148/radiology.176.2.2367658
- 129. Le Bihan D, Breton E, Lallemand D, Grenier P, Cabanis E, Laval-Jeantet M. MR Imaging of Intravoxel Incoherent Motions: Application to Diffusion and Perfusion in Neurologic Disorders. *Radiology* (1986) 161:401–7. doi: 10.1148/radiology.161.2.3763909
- Alexander AL, Lee JE, Lazar M, Field AS. Diffusion Tensor Imaging of the Brain. *Neurotherapeutics* (2007) 4:316–29. doi: 10.1016/j.nurt.2007.05.011
- 131. Jensen JH, Helpern JA, Ramani A, Lu H, Kaczynski K. Diffusional Kurtosis Imaging: The Quantification of Non-Gaussian Water Diffusion by Means of Magnetic Resonance Imaging. *Magn Reson Med* (2005) 53:1432–40. doi: 10.1002/mrm.20508
- 132. Maiter A, Riemer F, Allinson K, Zaccagna F, Crispin-Ortuzar M, Gehrung M, et al. Investigating the Relationship Between Diffusion Kurtosis Tensor Imaging (DKTI) and Histology Within the Normal Human Brain. *Sci Rep* (2021) 11:8857. doi: 10.1038/s41598-021-87857-w
- 133. Raab P, Hattingen E, Franz K, Zanella FE, Lanfermann H. Cerebral Gliomas: Diffusional Kurtosis Imaging Analysis of Microstructural Differences. *Radiology* (2010) 254:876–81. doi: 10.1148/radiol.09090819
- 134. Gong S, Zhang F, Norton I, Essayed WI, Unadkat P, Rigolo L, et al. Free Water Modeling of Peritumoral Edema Using Multi-Fiber Tractography: Application to Tracking the Arcuate Fasciculus for Neurosurgical Planning. *PloS One* (2018) 13:e0197056. doi: 10.1371/journal.pone.0197056
- 135. Khan UA, Rennert RC, White NS, Bartsch H, Farid N, Dale AM, et al. Diagnostic Utility of Restriction Spectrum Imaging (RSI) in Glioblastoma Patients After Concurrent Radiation-Temozolomide Treatment: A Pilot Study. J Clin Neurosci (2018) 58:136–41. doi: 10.1016/j.jocn.2018.09.008
- 136. Conturo TE, McKinstry RC, Akbudak E, Robinson BH. Encoding of Anisotropic Diffusion With Tetrahedral Gradients: A General Mathematical Diffusion Formalism and Experimental Results. *Magn Reson Med* (1996) 35:399–412. doi: 10.1002/mrm.1910350319
- 137. Mansfield P. Multi-Planar Image Formation Using NMR Spin Echoes. J Phys C Solid State Phys (1977) 10:L55–8. doi: 10.1088/0022-3719/10/3/004
- Aquino D, Gioppo A, Finocchiaro G, Bruzzone MG, Cuccarini V. MRI in Glioma Immunotherapy: Evidence, Pitfalls, and Perspectives. J Immunol Res (2017) 2017::5813951. doi: 10.1155/2017/5813951
- Basser PJ, Mattiello J, LeBihan D. MR Diffusion Tensor Spectroscopy and Imaging. *Biophys J* (1994) 66:259–67. doi: 10.1016/S0006-3495(94)80775-1
- 140. Hrabe J, Kaur G, Guilfoyle D. Principles and Limitations of NMR Diffusion Measurements. J Med Phys (2007) 32:34–42. doi: 10.4103/0971-6203.31148
- 141. Sundgren PC, Fan X, Weybright P, Welsh RC, Carlos RC, Petrou M, et al. Differentiation of Recurrent Brain Tumor Versus Radiation Injury Using Diffusion Tensor Imaging in Patients With New Contrast-Enhancing Lesions. *Magn Reson Imaging* (2006) 24:1131–4. doi: 10.1016/ j.mri.2006.07.008
- 142. Shiroishi MS, Boxerman JL, Pope WB. Physiologic MRI for Assessment of Response to Therapy and Prognosis in Glioblastoma. *Neuro Oncol* (2016) 18:467–78. doi: 10.1093/neuonc/nov179
- 143. Nakamura H, Murakami R, Hirai T, Kitajima M, Yamashita Y. Can MRI-Derived Factors Predict the Survival in Glioblastoma Patients Treated With Postoperative Chemoradiation Therapy? *Acta Radiol* (2013) 54:214–20. doi: 10.1258/ar.2012.120525
- 144. Saraswathy S, Crawford FW, Lamborn KR, Pirzkall A, Chang S, Cha S, et al. Evaluation of MR Markers That Predict Survival in Patients With Newly Diagnosed GBM Prior to Adjuvant Therapy. J Neurooncol (2009) 91:69–81. doi: 10.1007/s11060-008-9685-3

- 145. Moffat BA, Chenevert TL, Lawrence TS, Meyer CR, Johnson TD, Dong Q, et al. Functional Diffusion Map: A Noninvasive MRI Biomarker for Early Stratification of Clinical Brain Tumor Response. *Proc Natl Acad Sci* (2005) 102:5524–9. doi: 10.1073/pnas.0501532102
- 146. Østergaard L, Sorensen AG, Kwong KK, Weisskoff RM, Gyldensted C, Rosen BR. High Resolution Measurement of Cerebral Blood Flow Using Intravascular Tracer Bolus Passages. Part II: Experimental Comparison and Preliminary Results. *Magn Reson Med* (1996) 36. doi: 10.1002/ mrm.1910360511
- 147. Vrabec M, Van Cauter S, Himmelreich U, Van Gool SW, Sunaert S, De Vleeschouwer S, et al. MR Perfusion and Diffusion Imaging in the Follow-Up of Recurrent Glioblastoma Treated With Dendritic Cell Immunotherapy: A Pilot Study. *Neuroradiology* (2011) 53:721–3. doi: 10.1007/s00234-010-0802-6
- 148. Song J, Kadaba P, Kravitz A, Hormigo A, Friedman J, Belani P, et al. Multiparametric MRI for Early Identification of Therapeutic Response in Recurrent Glioblastoma Treated With Immune Checkpoint Inhibitors. *Neuro Oncol* (2020) 22:1658–66. doi: 10.1093/neuonc/noaa066
- 149. Lupo JM. Diffusion MRI as an Early Marker of Response to Immune Checkpoint Inhibitors. *Neuro Oncol* (2020) 22:1557–8. doi: 10.1093/ neuonc/noaa224
- 150. Schmainda KM. Diffusion-Weighted MRI as a Biomarker for Treatment Response in Glioma. CNS Oncol (2012) 1:169–80. doi: 10.2217/cns.12.25
- 151. Chu HH, Choi SH, Ryoo I, Kim SC, Yeom JA, Shin H, et al. Differentiation of True Progression From Pseudoprogression in Glioblastoma Treated With Radiation Therapy and Concomitant Temozolomide: Comparison Study of Standard and High- *B* -Value Diffusion-Weighted Imaging. *Radiology* (2013) 269:831–40. doi: 10.1148/radiol.13122024
- 152. Bulik M, Kazda T, Slampa P, Jancalek R. The Diagnostic Ability of Follow-Up Imaging Biomarkers After Treatment of Glioblastoma in the Temozolomide Era: Implications From Proton MR Spectroscopy and Apparent Diffusion Coefficient Mapping. *BioMed Res Int* (2015) 2015:641023. doi: 10.1155/2015/641023
- 153. Kazda T, Bulik M, Pospisil P, Lakomy R, Smrcka M, Slampa P, et al. Advanced MRI Increases the Diagnostic Accuracy of Recurrent Glioblastoma: Single Institution Thresholds and Validation of MR Spectroscopy and Diffusion Weighted MR Imaging. *NeuroImage Clin* (2016) 11:316–21. doi: 10.1016/j.nicl.2016.02.016
- 154. Yu Y, Ma Y, Sun M, Jiang W, Yuan T, Tong D. Meta-Analysis of the Diagnostic Performance of Diffusion Magnetic Resonance Imaging With Apparent Diffusion Coefficient Measurements for Differentiating Glioma Recurrence From Pseudoprogression. *Med (Baltimore)* (2020) 99:e20270. doi: 10.1097/MD.00000000020270
- 155. Zhang H, Ma L, Shu C, Wang Y, Dong L. Diagnostic Accuracy of Diffusion MRI With Quantitative ADC Measurements in Differentiating Glioma Recurrence From Radiation Necrosis. J Neurol Sci (2015) 351:65–71. doi: 10.1016/j.jns.2015.02.038
- 156. van Dijken BRJ, van Laar PJ, Holtman GA, van der Hoorn A. Diagnostic Accuracy of Magnetic Resonance Imaging Techniques for Treatment Response Evaluation in Patients With High-Grade Glioma, a Systematic Review and Meta-Analysis. *Eur Radiol* (2017) 27:4129–44. doi: 10.1007/ s00330-017-4789-9
- 157. Prager AJ, Martinez N, Beal K, Omuro A, Zhang Z, Young RJ. Diffusion and Perfusion MRI to Differentiate Treatment-Related Changes Including Pseudoprogression From Recurrent Tumors in High-Grade Gliomas With Histopathologic Evidence. Am J Neuroradiol (2015) 36:877–85. doi: 10.3174/ ajnr.A4218
- 158. Khalifa J, Tensaouti F, Lotterie JA, Catalaa I, Chaltiel L, Benouaich-Amiel A, et al. Do Perfusion and Diffusion MRI Predict Glioblastoma Relapse Sites Following Chemoradiation? J Neurooncol (2016) 130:181–92. doi: 10.1007/s11060-016-2232-8
- 159. Zhang J, Yu H, Qian X, Liu K, Tan H, Yang T, et al. Pseudo Progression Identification of Glioblastoma With Dictionary Learning. *Comput Biol Med* (2016) 73:94–101. doi: 10.1016/j.compbiomed.2016.03.027
- 160. Galbán CJ, Chenevert TL, Meyer CR, Tsien C, Lawrence TS, Hamstra DA, et al. The Parametric Response Map is an Imaging Biomarker for Early Cancer Treatment Outcome. Nat Med (2009) 15:572–6. doi: 10.1038/nm.1919
- 161. Kashimura H, Inoue T, Beppu T, Ogasawara K, Ogawa A. Diffusion Tensor Imaging for Differentiation of Recurrent Brain Tumor and Radiation

Necrosis After Radiotherapy—Three Case Reports. Clin Neurol Neurosurg (2007) 109:106-10. doi: 10.1016/j.clineuro.2006.04.005

- 162. Wang S, Martinez-Lage M, Sakai Y, Chawla S, Kim SG, Alonso-Basanta M, et al. Differentiating Tumor Progression From Pseudoprogression in Patients With Glioblastomas Using Diffusion Tensor Imaging and Dynamic Susceptibility Contrast MRI. Am J Neuroradiol (2016) 37:28–36. doi: 10.3174/ajnr.A4474
- 163. Qian X, Tan H, Zhang J, Zhao W, Chan MD, Zhou X. Stratification of Pseudoprogression and True Progression of Glioblastoma Multiform Based on Longitudinal Diffusion Tensor Imaging Without Segmentation. *Med Phys* (2016) 43:5889. doi: 10.1118/1.4963812
- 164. Hamstra DA, Chenevert TL, Moffat BA, Johnson TD, Meyer CR, Mukherji SK, et al. Evaluation of the Functional Diffusion Map as an Early Biomarker of Time-to-Progression and Overall Survival in High-Grade Glioma. *Proc Natl Acad Sci* (2005) 102:16759–64. doi: 10.1073/pnas.0508347102
- 165. Moffat BA, Chenevert TL, Meyer CR, Mckeever PE, Hall DE, Hoff BA, et al. The Functional Diffusion Map: An Imaging Biomarker for the Early Prediction of Cancer Treatment Outcome. *Neoplasia* (2006) 8:259–67. doi: 10.1593/neo.05844
- 166. Ellingson BM, Malkin MG, Rand SD, LaViolette PS, Connelly JM, Mueller WM, et al. Volumetric Analysis of Functional Diffusion Maps is a Predictive Imaging Biomarker for Cytotoxic and Anti-Angiogenic Treatments in Malignant Gliomas. J Neurooncol (2011) 102:95–103. doi: 10.1007/s11060-010-0293-7
- 167. Kelly PJ, Daumas-Duport C, Kispert DB, Kall BA, Scheithauer BW, Illig JJ. Imaging-Based Stereotaxic Serial Biopsies in Untreated Intracranial Glial Neoplasms. J Neurosurg (1987) 66:865–74. doi: 10.3171/jns.1987.66.6.0865
- 168. Chenevert TL, Malyarenko DI, Galbán CJ, Gomez-Hassan DM, Sundgren PC, Tsien CI, et al. Comparison of Voxel-Wise and Histogram Analyses of Glioma ADC Maps for Prediction of Early Therapeutic Change. *Tomogr (Ann Arbor Mich)* (2019) 5:7–14. doi: 10.18383/j.tom.2018.00049
- 169. Reimer C, Deike K, Graf M, Reimer P, Wiestler B, Omar Floca R, et al. Differentiation of Pseudoprogression and Real Progression in Glioblastoma Using ADC Parametric Response Maps. *PloS One* (2017) 12:e0174620. doi: 10.1371/journal.pone.0174620
- 170. Yoon RG, Kim HS, Kim DY, Hong GS, Kim SJ. Apparent Diffusion Coefficient Parametric Response Mapping MRI for Follow-Up of Glioblastoma. *Eur Radiol* (2016) 26:1037–47. doi: 10.1007/s00330-015-3896-8
- Macdonald DR, Cascino TL, Schold SC, Cairncross JG. Response Criteria for Phase II Studies of Supratentorial Malignant Glioma. J Clin Oncol (1990) 8:1277–80. doi: 10.1200/JCO.1990.8.7.1277
- 172. Chenevert TL, Ross BD. Diffusion Imaging for Therapy Response Assessment of Brain Tumor. *Neuroimaging Clin N Am* (2009) 19:559–71. doi: 10.1016/j.nic.2009.08.009
- 173. Bacchi S, Zerner T, Dongas J, Asahina AT, Abou-Hamden A, Otto S, et al. Deep Learning in the Detection of High-Grade Glioma Recurrence Using Multiple MRI Sequences: A Pilot Study. J Clin Neurosci (2019) 70:11–3. doi: 10.1016/j.jocn.2019.10.003
- 174. Chang PD, Chow DS, Yang PH, Filippi CG, Lignelli A. Predicting Glioblastoma Recurrence by Early Changes in the Apparent Diffusion Coefficient Value and Signal Intensity on FLAIR Images. Am J Roentgenol (2017) 208:57–65. doi: 10.2214/AJR.16.16234
- 175. Yoon RG, Kim HS, Koh MJ, Shim WH, Jung SC, Kim SJ, et al. Differentiation of Recurrent Glioblastoma From Delayed Radiation Necrosis by Using Voxel-Based Multiparametric Analysis of MR Imaging Data. *Radiology* (2017) 285:206–13. doi: 10.1148/radiol.2017161588
- 176. Liu Z-C, Yan L-F, Hu Y-C, Sun Y-Z, Tian Q, Nan H-Y, et al. Combination of IVIM-DWI and 3D-ASL for Differentiating True Progression From Pseudoprogression of Glioblastoma Multiforme After Concurrent Chemoradiotherapy: Study Protocol of a Prospective Diagnostic Trial. BMC Med Imaging (2017) 17:10. doi: 10.1186/s12880-017-0183-y
- 177. Shin KE, Ahn KJ, Choi HS, Jung SL, Kim BS, Jeon SS, et al. DCE and DSC MR Perfusion Imaging in the Differentiation of Recurrent Tumour From Treatment-Related Changes in Patients With Glioma. *Clin Radiol* (2014) 69: e264–72. doi: 10.1016/j.crad.2014.01.016
- Le Bihan D. What can We See With IVIM MRI? *Neuroimage* (2019) 187:56– 67. doi: 10.1016/j.neuroimage.2017.12.062

- 179. Keil VC, Mädler B, Gielen GH, Pintea B, Hiththetiya K, Gaspranova AR, et al. Intravoxel Incoherent Motion MRI in the Brain: Impact of the Fitting Model on Perfusion Fraction and Lesion Differentiability. J Magn Reson Imaging (2017) 46:1187–99. doi: 10.1002/jmri.25615
- 180. Conklin J, Heyn C, Roux M, Cerny M, Wintermark M. Federau C. A Simplified Model for Intravoxel Incoherent Motion Perfusion Imaging of the Brain. Am J Neuroradiol (2016) 37:2251–7. doi: 10.3174/ajnr.A4929
- 181. Jabehdar Maralani P, Myrehaug S, Mehrabian H, Chan AKM, Wintermark M, Heyn C, et al. Intravoxel Incoherent Motion (IVIM) Modeling of Diffusion MRI During Chemoradiation Predicts Therapeutic Response in IDH Wildtype Glioblastoma. *Radiother Oncol* (2021) 156:258–65. doi: 10.1016/j.radonc.2020.12.037
- 182. Kim HS, Suh CH, Kim N, Choi CG, Kim SJ. Histogram Analysis of Intravoxel Incoherent Motion for Differentiating Recurrent Tumor From Treatment Effect in Patients With Glioblastoma: Initial Clinical Experience. *Am J Neuroradiol* (2014) 35:490–7. doi: 10.3174/ajnr.A3719
- 183. D'Amore F, Grinberg F, Mauler J, Galldiks N, Blazhenets G, Farrher E, et al. Combined 18f-FET PET and Diffusion Kurtosis MRI in Posttreatment Glioblastoma: Differentiation of True Progression From Treatment-Related Changes. *Neuro Oncol Adv* (2021) 3:vdab044. doi: 10.1093/noajnl/vdab044
- 184. Wu XF, Liang X, Wang XC, Qin JB, Zhang L, Tan Y, et al. Differentiating High-Grade Glioma Recurrence From Pseudoprogression: Comparing Diffusion Kurtosis Imaging and Diffusion Tensor Imaging. *Eur J Radiol* (2021) 135:109445. doi: 10.1016/j.ejrad.2020.109445
- 185. Chakhoyan A, Woodworth DC, Harris RJ, Lai A, Nghiemphu PL, Liau LM, et al. Mono-Exponential, Diffusion Kurtosis and Stretched Exponential Diffusion MR Imaging Response to Chemoradiation in Newly Diagnosed Glioblastoma. J Neurooncol (2018) 139:651–9. doi: 10.1007/s11060-018-2910-9
- 186. Baek HJ, Kim HS, Kim N, Choi YJ, Kim YJ. Percent Change of Perfusion Skewness and Kurtosis: A Potential Imaging Biomarker for Early Treatment Response in Patients With Newly Diagnosed Glioblastomas. *Radiology* (2012) 264:834–43. doi: 10.1148/radiol.12112120
- 187. Wang X, Li F, Wang D, Zeng Q. Diffusion Kurtosis Imaging Combined With Molecular Markers as a Comprehensive Approach to Predict Overall Survival in Patients With Gliomas. *Eur J Radiol* (2020) 128:108985. doi: 10.1016/j.ejrad.2020.108985
- Juan-Albarracín J, Fuster-Garcia E, García-Ferrando GA, García-Gómez JM. ONCOhabitats: A System for Glioblastoma Heterogeneity Assessment Through MRI. Int J Med Inform (2019) 128:53–61. doi: 10.1016/ j.ijmedinf.2019.05.002
- 189. Hu Y-C, Yan L-F, Han Y, Duan S-J, Sun Q, Li G-F, et al. Can the Low and High B-Value Distribution Influence the Pseudodiffusion Parameter Derived From IVIM DWI in Normal Brain? *BMC Med Imaging* (2020) 20:14. doi: 10.1186/s12880-020-0419-0
- 190. Niendorf T, Dijkhuizen RM, Norris DG, Van Lookeren Campagne M, Nicolay K. Biexponential Diffusion Attenuation in Various States of Brain Tissue: Implications for Diffusion-Weighted Imaging. *Magn Reson Med* (1996) 36:847–57. doi: 10.1002/mrm.1910360607
- 191. Xu J-L, Li Y-L, Lian J-M, Dou S, Yan F-S, Wu H, et al. Distinction Between Postoperative Recurrent Glioma and Radiation Injury Using MR Diffusion Tensor Imaging. *Neuroradiology* (2010) 52:1193–9. doi: 10.1007/s00234-010-0731-4
- 192. Alexiou GA, Zikou A, Tsiouris S, Goussia A, Kosta P, Papadopoulos A, et al. Comparison of Diffusion Tensor, Dynamic Susceptibility Contrast MRI and 99mtc-Tetrofosmin Brain SPECT for the Detection of Recurrent High-Grade Glioma. *Magn Reson Imaging* (2014) 32:854–9. doi: 10.1016/ j.mri.2014.04.013
- 193. Agarwal A, Kumar S, Narang J, Schultz L, Mikkelsen T, Wang S, et al. Morphologic MRI Features, Diffusion Tensor Imaging and Radiation Dosimetric Analysis to Differentiate Pseudo-Progression From Early Tumor Progression. J Neurooncol (2013) 112:413–20. doi: 10.1007/s11060-013-1070-1
- 194. Price SJ, Jena R, Burnet NG, Carpenter TA, Pickard JD, Gillard JH. Predicting Patterns of Glioma Recurrence Using Diffusion Tensor Imaging. *Eur Radiol* (2007) 17:1675–84. doi: 10.1007/s00330-006-0561-2

- 195. Panagiotaki E, Schneider T, Siow B, Hall MG, Lythgoe MF, Alexander DC. Compartment Models of the Diffusion MR Signal in Brain White Matter: A Taxonomy and Comparison. *Neuroimage* (2012) 59:2241–54. doi: 10.1016/ j.neuroimage.2011.09.081
- 196. Roberts T, Hyare H, Hipwell B, Ianus A, Breen-norris J, Panagiotaki E, et al. Quantification of Tumour Microstructure in Low and High-Grade Brain Tumours Using VERDICT MRI: An Initial Feasibility Study. *Neuro Oncol* (2018) 20(Suppl. 1):i16. doi: 10.1093/neuonc/nox238.072
- 197. Roberts TA, Hyare H, Agliardi G, Hipwell B, d'Esposito A, Ianus A, et al. Noninvasive Diffusion Magnetic Resonance Imaging of Brain Tumour Cell Size for the Early Detection of Therapeutic Response. Sci Rep (2020) 10:9223. doi: 10.1038/s41598-020-65956-4
- 198. Zaccagna F, Riemer F, Priest AN, McLean MA, Allinson K, Grist JT, et al. Non-Invasive Assessment of Glioma Microstructure Using VERDICT MRI: Correlation With Histology. *Eur Radiol* (2019) 29:5559–66. doi: 10.1007/ s00330-019-6011-8
- 199. Metz MC, Molina-Romero M, Lipkova J, Gempt J, Liesche-Starnecker F, Eichinger P, et al. Predicting Glioblastoma Recurrence From Preoperative MR Scans Using Fractional-Anisotropy Maps With Free-Water Suppression. *Cancers (Basel)* (2020) 12:728. doi: 10.3390/cancers12030728
- 200. McDonald CR, Delfanti RL, Krishnan AP, Leyden KM, Hattangadi-Gluth JA, Seibert TM, et al. Restriction Spectrum Imaging Predicts Response to Bevacizumab in Patients With High-Grade Glioma. *Neuro Oncol* (2016) 18:1579–90. doi: 10.1093/neuonc/now063
- 201. Farid N, Almeida-Freitas DB, White NS, McDonald CR, Muller KA, VandenBerg SR, et al. Restriction-Spectrum Imaging of Bevacizumab-Related Necrosis in a Patient With GBM. *Front Oncol* (2013) 3 SEP:258. doi: 10.3389/fonc.2013.00258
- 202. Latysheva A, Geier OM, Hope TR, Brunetti M, Micci F, Vik-Mo EO, et al. Diagnostic Utility of Restriction Spectrum Imaging in the Characterization of the Peritumoral Brain Zone in Glioblastoma: Analysis of Overall and Progression-Free Survival. *Eur J Radiol* (2020) 132:109289. doi: 10.1016/ j.ejrad.2020.109289
- 203. Jin Y, Randall JW, Elhalawani H, Al Feghali KA, Elliott AM, Anderson BM, et al. Detection of Glioblastoma Subclinical Recurrence Using Serial Diffusion Tensor Imaging. *Cancers (Basel)* (2020) 12:568. doi: 10.3390/ cancers12030568
- 204. Daghighi S, Bahrami N, Tom WJ, Coley N, Seibert TM, Hattangadi-Gluth JA, et al. Restriction Spectrum Imaging Differentiates True Tumor Progression From Immune-Mediated Pseudoprogression: Case Report of a Patient With Glioblastoma. *Front Oncol* (2020) 10:24. doi: 10.3389/fonc.2020.00024
- 205. Delgado AF, De Luca F, Hanagandi P, van Westen D, Delgado AF. Arterial Spin-Labeling in Children With Brain Tumor: A Meta-Analysis. Am J Neuroradiol (2018) 39:1536–42. doi: 10.3174/ajnr.A5727

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