Document downloaded from:

http://hdl.handle.net/10251/63159

This paper must be cited as:

Capella Hernández, JV.; Bonastre Pina, AM.; Ors Carot, R.; Peris Tortajada, M. (2015). An interference-tolerant nitrate smart sensor for Wireless Sensor Network applications. Sensors and Actuators B: Chemical. 213:534-540. doi:10.1016/j.snb.2015.02.125.



The final publication is available at http://dx.doi.org/10.1016/j.snb.2015.02.125

Copyright Elsevier

Additional Information

1	An interference-tolerant nitrate smart sensor for Wireless Sensor
2	Network applications
3	
4	Juan V. Capella ^a , Alberto Bonastre ^a , Rafael Ors ^a , Miguel Peris ^{b,*}
5	^a Department of Computer Engineering, Polytechnic University of Valencia, 46071
6	Valencia, SPAIN
7	^b Department of Chemistry, Polytechnic University of Valencia, 46071 Valencia, SPAIN
8	
9	* Corresponding author; <i>e-mail</i> : mperist@qim.upv.es
10	
11	Abstract
12	As a major contaminant in ground water, nitrate determination is a common practice in
13	environmental analysis, especially the continuous and simultaneous monitoring of its
14	concentration at many different points. For this task, sensor networks are a promising
15	tool, although they require the use of sensors with special features, such as those of Ion
16	Selective Electrodes (ISEs). Unfortunately, their measurements are – to a greater or
17	lesser extent – affected by the presence of other coexisting (interfering) ions. A new
18	methodology is then proposed in this work to deal with major interferences (chloride
19	and bicarbonate in the case of nitrate determination), in such a way that the results
20	obtained in the measurements of the content of NO ₃ - with a nitrate selective electrode
21	can be considered as virtually error-free from these interferences. For this purpose, a
22	new sensor node has been developed; it consists of three ISEs (NO ₃ -, Cl-, and HCO ₃ -)
23	coupled to a low-consumption, low-cost microcontroller (a small chip containing all the
24	computer components), which receives and processes all signals coming from the
25	electrodes. This information is suitably treated, as described in detail in this paper, to
26	provide an accurate estimation of the true value of NO ₃ ⁻ concentration.
27	The application of this methodology results in an interference-tolerant nitrate smart
28	sensor capable of being employed within a wireless sensor network in the continuous
29	monitoring of nitrate concentration in aquifers and rivers.
30	
31	
32	Keywords: Smart sensor; Interference tolerance; Nitrate determination; Ion selective
33	electrode; Interfering ion.

1 Introduction

3	5
J	J

36	Nitrate is one of the most common contaminants of ground water, originating mainly
37	from agricultural fertilizer application and release of sewage. As the presence of this
38	species in water presents a well-known risk to health, it seems obvious that the
39	monitoring of nitrate concentration in aquifers and rivers may result fundamental.
40	Several analytical techniques have been used for this purpose [1], ion-selective
41	electrodes (ISEs) being perhaps the most suitable one; in this sense, the literature offers
42	a great deal of references, from early work [2] [3] to recent contributions especially with
43	a view of in-line monitoring [4] [5]. It is straightforward with advantages such as high
44	selectivity, sensitivity, good precision, simplicity, portability, non-destructive analysis,
45	and last but by no means least, low cost and power consumption. The latter makes this
46	technique highly suitable for Wireless Sensor Network (WSN) applications.
47	However, it is a common feature of all analysis methods that when atoms or ions of a
48	different species but with similar properties are also present, i.e. coexist with the atoms
49	or ions of interest, they interfere with measurement. This is also true of analysis using
50	ion selective electrodes, so that if ions similar to the target ions are present, they will –
51	to a greater or lesser extent – affect measurement (when considered in contrast to the
52	target ions, these ions are known as interfering ions). Therefore, when using the ion
53	electrode method, care needs to be taken with regard to mutual interference within each
54	of these groups [6].
55	In this sense, several attempts to deal with this problem have been carried out based on
56	the joint consideration of several ISEs response. Some of these examples include the
57	development of the so-called electronic tongues, arrays of potentiometric sensors (ISEs)
58	coupled to pattern recognition tools. They have been applied to water quality
59	monitoring [7][8], and their performances being compared to those of discrete
60	conventional ion-selective electrodes [9]. On the other hand, recent efforts are focused
61	on the development of artificial neural network (ANN) architectures; they have been
62	applied to raw readings from a chemical sensor multi-probe (e-tongue), comprised of
63	off-the shelf ISEs, to estimate individual ion concentrations in solutions at
64	environmentally relevant concentrations and containing environmentally representative
65	ion mixtures [10]. Nevertheless, all these approaches –though successful-are sometimes

66 too complex (need of high-level computer resources) and time-consuming, and would 67

not be adequate in case of WSN applications. That is why more simple developments,

68 i.e. the utilization of discrete ISEs, are usually preferred.

69 No ion-selective electrodes are completely ion-specific [11]; all are sensitive to other

70 ions having similar physical properties, to an extent which depends on the degree of

71 similarity. Most of these interferences are weak enough to be ignored, but in some cases

the electrode may actually be much more sensitive to the interfering ion than to the

73 desired ion, requiring that the interfering ion be present only in relatively very low

74 concentrations, or entirely absent. In practice, the relative sensitivities of each type of

ion-specific electrode to various interfering ions is generally known and should be

76 checked for each case; however the precise degree of interference depends on many

factors, preventing precise correction of readings. Instead, the calculation of relative

degree of interference from the concentration of interfering ions can only be used as a

79 guide to determine whether the approximate extent of the interference will allow

reliable measurements, or whether the experiment will need to be redesigned so as to

81 reduce the effect of interfering ions.

82 The effects of coexisting ions can be predicted to some extent from the response

membrane material, that is to say, the reactivity of the response membrane material to

the coexisting ions. For example, a solid-state membrane electrode can be seriously

affected by coexisting ions that form insoluble compounds or complex salts with the

material of its response membrane; and a liquid membrane electrode can be affected by

coexisting ions that form ionic associates with the components in its response

88 membrane.

72

75

77

78

80

83

84

85

86

87

90

91

94

95

89 The intensity of the interference produced by an ion species is expressed by the

selectivity coefficient (or by the maximum allowable coexistence factor, this roughly

corresponding to the reciprocal of the selectivity coefficient). The Nicolsky-Eisenman

92 equation (an extension to the Nernst equation) [12] defines the selectivity coefficient k_{ij}

96
$$E_i = E_i^0 + 2.303 \frac{RT}{n_i F} log \left\{ a_i + \sum_j k_{ij} (a_j^{(n_i/n_j)}) \right\}$$

where E is the emf, E^0 the standard electrode potential, n the ionic valency including the 93

sign, a the activity, i the ion of interest (target ion), j the interfering ions and k_{ij} is the

selectivity coefficient. The smaller this value, the better the selectivity with respect to

97	the target ion, <i>i.e.</i> the less the interference by j [13].
98	Hence, in practice, the ratio of target ion to interfering ion concentration is very
99	important. Higher concentrations of target ions result in the interfering ions having a
100	smaller effect, and conversely, lower concentrations result in them having a larger
101	effect. It is then logical that the ideal ISE should be interference-free, although up till
102	now the objective of the major manufacturers of this type of devices consists of keeping
103	these interferences to a minimum. Our aim is then to address cross-ion interferences in
104	such a way that, although ISEs may be only partially selective for their target analyte,
105	we can take advantage of their promising use in in-situ portable sensors.
106	Recent advances in the field of microelectronics and communications allow for the
107	development of modern applications that require new sensors with different
108	requirements from those of the traditional devices; additionally, and owing to their
109	possibilities of data management, they permit to obtain more precise, robust, and
110	powerful systems [14]. Following this line, in this paper we propose a nitrate smart
111	sensor that is able to eliminate major interferences from other species, namely chloride
112	and bicarbonate ions; moreover, it meets the necessary requirements to be utilized
113	within an application based on WSN [15], with all its inherent benefits in the field of
114	chemical analyses.
115	In the next sections, the procedure proposed for the rejection of major interferences in
116	nitrate determination by ISEs will be described in detail, along with the results obtained
117	after its application to discrete samples, with a view to use it in the future within a WSN
118	for environmental analysis purposes.
119	
120	2 Rejection of interferences
121	
122	As mentioned above, one of the main drawbacks while measuring nitrate concentrations
123	by means of ISEs is the interference caused by other similar species, since their
124	presence may cause an incremental deviation on the results obtained. In case of the
125	nitrate electrode [16], the following ions usually interfere (average selectivity
126	coefficients, SC, in brackets): Chloride (6.10 ⁻³), Bicarbonate (5.10 ⁻³), Nitrite (1.10 ⁻³),
127	Acetate (5.10 ⁻⁴), Fluoride (1.10 ⁻⁴), Sulfate (1.10 ⁻⁵). In this sense, the higher the value of
128	SC, the more interference; therefore, in the present work we will consider those two

129	interferences with the highest SC values, i.e. chloride and bicarbonate. Nevertheless, the
130	procedure could easily be applied to other – though less important – potential interfering
131	anions. It should also be remarked that, in our case, the chloride and bicarbonate
132	concentrations have been chosen according to the range found in previous field sample
133	analysis. On the other hand, in the field of Electroanalytical Chemistry it is generally
134	accepted that, at low ionic strengths, (e.g. below 0.01M for monovalent ions and
135	0.001M for divalent ions) the difference between concentration and activity is really
136	small and the use of concentration units instead of activity for the measurements
137	(including calibration) should not cause a significant error in the determinations, even
138	without the use of ionic strength adjustment buffer (ISAB). In our case, all occurring
139	ions being monovalent, the ionic strength of the highest concentrated solution is 0.005,
140	what means that concentrations will be utilized instead of activities with no significant
141	error.
142	The correction system is then based on the evaluation of the accumulative error suffered
143	by the obtained measurement. This error has been found to depend on three factors,
144	namely: the concentrations of nitrate, bicarbonate, and chloride. Our initial hypothesis
145	was that it is possible to estimate, and then compensate, the interference error if [Cl-]
146	and [HCO3-] are known. In this way, additional bicarbonate and chloride ISEs were
147	added to the nitrate ISE in order to measure all three concentrations. It must be
148	considered that the real concentrations are not available, as far as they have to be
149	determined by the measurements – probably affected by the interferences and perhaps
150	other instrumental errors – of these ISEs.
151	As described below, an exhaustive set of experiments (Table 2) has been carried out
152	under laboratory conditions. These experiments consisted of the application of the three
153	ISEs to mixtures prepared with known concentrations of all three anions. As expected,
154	measured nitrate concentrations suffered a relative error ranging from 15 % to 30 %
155	(easily determined as the difference between measured values and true values). For their
156	part, the errors of both bicarbonate and chloride measurements were under 2 % in all
157	instances.
158	In order to obtain reliable results, n replications have been performed for each
159	measurement, n being calculated as follows:
160	The results for each measurement have been considered as random variables (X1,
161	$X_2,, X_n$) with a μ mean value. n simulations have been repeated until an estimation of
162	whas been obtained with a 90 % confidence interval according to the expression:

163

 $\bar{X}(n) \pm t_{n-1,0.95} \sqrt{\frac{S^2(n)}{n}}$ 164

165 where $t_{n-1,0.95}$ represents the upper limit of the Student's t-distribution on n-1 degrees of 166 freedom, and X(n) and $S^{2}(n)$ are the mean and the variance of the results obtained in the 167 different experiments. In general, 5-12 replications were carried out for each 168 169 measurement. 170 We have evaluated different techniques that relation the interferences of chloride and 171 bicarbonate on nitrate measurements, from simple, fixed (predetermined) corrections to 172 more complex techniques. From the corresponding essays, it could be noticed that the 173 former did not prove to be useful in all cases. More complex solutions yielded good 174 results, but their high requirements of computing resources make them unsuitable for 175 WSN. Instead, the proposed methodology brings together simplicity and highly 176 satisfactory results [17]. 177 It assumes that there is some relationship between the concentration of interferents and 178 the error committed in the determination. Bearing this in mind, several experiments 179 were carried out using standard solutions; then, with the results obtained as well as the 180 corresponding true values, different mathematical procedures were evaluated to 181 determine the error committed. Least squares linear regression was chosen since it 182 provided negligible errors in the estimation. According to this procedure, the values 183 provided by the three ISEs used are plotted against (correlated with) the errors in nitrate 184 measurements (true values are known), the corresponding regression line coefficients 185 being shown in Table 1. Linear regression has proved to be satisfactory ($R^2 = 0.9607$). 186 This accuracy is better, for example, than the measurement error. 187 This method also deals with cross-ion interferences, since the regression line is based on 188 measured ISE concentrations, rather than on true concentrations. Thus, the mutual 189 interferences have also been considered. 190 Thereafter, an estimation program was implemented; it applied the previously defined 191 error estimation procedure to the obtained measurements. The estimated error is then

subtracted to the nitrate concentration measured to obtain the corrected value. Once the

latter has been achieved, the error turns out to be under 5 % in all studied cases.

194

192

3 System description

195	3 System description
196	
197	The application of the technique described in the previous section results in the
198	development of a smart sensor node formed by three ISE-type transducers for the
199	determination of nitrate, chloride, and bicarbonate, respectively. The core of the system
200	consists of a low-consumption, low-cost microcontroller; its capacity is nevertheless
201	more than sufficient to carry out all the operations required to apply the developed
202	techniques. This microcontroller is a small integrated circuit that contains all the
203	computer components (CPU, memory, and necessary I/O subsystems), and therefore
204	offers the possibility to implement complete applications using only one chip [18]. The
205	device chosen (ARM Cortex M0) is a 32-bit microcontroller with a high energy
206	efficiency (12.5 μW / MHz) and performance; it has 2 KB RAM memory and 8 KB
207	flash memory, as well as three timers (16 and 32 bits) and an A/D converter (10-bit
208	resolution and 8 channels). Incoming signals from ISEs are adapted/amplified by means
209	of an AD524 Instrumentation Amplifier (Analog Devices).
210	The capture of the information in the sensor node begins in the transducers (nitrate-ISE,
211	chloride-ISE, and bicarbonate-ISE). The analogical signals from these ISEs are
212	conveniently processed and amplified by means of analogical circuits, as shown in
213	Figure 1. The output of this circuit is received by the A/D converters of the
214	microcontroller (Input/Output subsystems) to be discretized. After the suitable
215	conversion, a digital information of each ISE measurement is available and, therefore,
216	able to be processed. From this moment, the error estimation procedure evaluates the
217	interference influence, and the measured nitrate concentration is then corrected by using
218	the proposed method, as described in Section 2.
219	Once the corrected measurement has been calculated, it is transmitted through the
220	communication subsystem, be it periodically, upon request, or in an automatic way
221	when certain conditions are met [19].
222	Since one of the goals was that this interference-tolerant nitrate smart sensor node could
223	be employed in future WSN applications (and thus within the future Internet of Things),
224	both its consumption and maintenance must reach a minimum value [20]. Therefore,
225	low consumption components (the ISEs and the microcontroller) have been utilized;

furthermore, the sensor node has been equipped with an energy harvesting subsystem

that is able to keep it on over long periods of time [2].

228

227

4. Experimental

229

230	
231	All standard and reagent solutions were prepared from analytical reagent grade
232	chemicals (KNO ₃ for nitrate, NaHCO ₃ for bicarbonate, and KCl for chloride) using
233	distilled and deionized water from a Milli-Q water purification system. The deionized
234	water had a specific conductivity less than 0.1 μs cm ⁻¹ .
235	Nitrate, bicarbonate, and chloride concentrations are measured using three different
236	homemade ISEs. When the ISE is immersed in an aqueous solution, a potential is
237	established across the membrane that depends on the relative amounts of analyte in the
238	medium, this potential being read relative to a double-junction reference electrode of the
239	probe. Our ISEs were prepared and conditioned following some of the guidelines
240	provided by the literature [21].
241	In case of nitrate, the ISE employs a silver/silver chloride wire electrode in a custom
242	filling solution. The internal solution is separated from the sample medium by a
243	polyvinylchloride (PVC) membrane, which selectively interacts with NO ₃ ⁻ ions. To
244	prepare this membrane, a mixture of 45 mg tridodecylammonium nitrate, 370 mg
245	dibutylphthalate as plasticizer and 160 mg PVC is thoroughly dissolved in 7 mL
246	tetrahydrofuran. The solution obtained is then poured into small glass rings and the
247	organic solvent is evaporated, thus achieving homogeneous membrane layers with a
248	thickness ranging between 450 and 500 μm . The resulting electrode has the following
249	characteristics: a Nernstian slope of $54 \pm 5 \text{ mV}$ per decade change in activity, a limit of
250	detection of 0.05 mg L ⁻¹ of NO ₃ ⁻ , a response time of less than 10 seconds, a selectivity
251	coefficient for nitrate against chloride ($k_{NO3-, Cl-}$) of 5 × 10 ⁻³ , and a selectivity
252	coefficient for nitrate against bicarbonate ($k_{NO3-, HCO3-}$) of 4 × 10 ⁻³ .
253	The nitrate-ISE was then calibrated with potassium nitrate standard solutions
254	maintained at 25°C and constantly stirred. Figure 2 shows a calibration curve for this
255	anion. All measurements (including calibration) were carried out with 100 mL of test
256	solution in a suitable Pyrex beaker. All glassware was decontaminated, washed several
257	times with double-distilled water and dried in an oven at 150°C overnight prior to use.
258	The validation of the results obtained by the nitrate-ISE was achieved using an official
259	standard method [22] and showed excellent correlation between the two techniques
260	$(R^2 = 0.9948)$ over a large range from 0.1 to 20 mg L^{-1} nitrate-N with no systematic

errors. On the other hand, the drift of the electrode contacting a 9.9 mM nitrate solution

is less than 0.4 mV h⁻¹, measured at constant temperature and with ISE and reference 262 263 electrode continually immersed. 264 As regards bicarbonate, the ISE consists of a polyvinyl chloride tube covered with a thin 265 (10-25 μm) HCO₃-selective membrane made from a mixture containing polyvinyl 266 chloride, di-(2-ethylhexyl) sebacate, trioctyl tin chloride and an H⁺ interference-267 removing trifluroacetophenone (trifluoroacetyldecylbenzene), a liquid solution 268 containing 50 mM phosphate buffer and 0.01 M sodium chloride in the tube, and a lead 269 wire connected to a Ag/AgCl reference electrode positioned in the tube. This ISE has 270 then the following features: a Nernstian slope of 55 ± 5 mV per decade change in activity, a limit of detection of 1.5 mg L⁻¹ of HCO₃⁻, and a response time of less than 20 271 272 seconds. 273 The bicarbonate-ISE was calibrated with sodium bicarbonate standard solutions 274 maintained at 25°C and constantly stirred. Figure 2 shows a calibration curve for this 275 anion. All measurements (including calibration) were carried out with 100 mL of test 276 solution in a suitable Pyrex beaker. All glassware was decontaminated, washed several 277 times with double-distilled water and dried in an oven at 150°C overnight prior to use. 278 The validation of the results obtained by the bicarbonate-ISE was performed using an 279 official standard method [22] and showed excellent correlation between the two techniques (R²= 0.9936) over a large range from 10 to 250 mg L⁻¹ bicarbonate with no 280 systematic errors. It should also be noted that the drift of the electrode contacting a 11.9 281 282 mM bicarbonate solution is less than 0.5 mV h⁻¹, measured at constant temperature and with ISE and reference electrode continually immersed. 283 284 Finally, in the case of the chloride-ISE, the membrane is made from a combination of 285 ([γ-[4,5-dimethyl-3,6-bis(octyloxy)-1,2-phenylene]]bis(trifluoroacetato-O)dimercury), 286 tridodecylmethylammonium chloride, bis(2-ethylhexyl)sebacate, and polyvinylchloride, 287 a reference solution containing 0.1 M potassium chloride, and a wire connected to a Ag/AgCl reference electrode. The ISE so constituted is characterized by a Nernstian 288 slope of 54 ± 5 mV per decade change in activity, a limit of detection of 1 mg L⁻¹ of Cl⁻, 289 290 and a response time of less than 10 seconds. 291 The chloride-ISE was calibrated with potassium chloride standard solutions kept at 292 25°C and constantly stirred. Figure 2 shows a calibration curve for this anion. All 293 measurements (including calibration) were carried out with 100 mL of test solution in a

suitable Pyrex beaker. All glassware was decontaminated, washed several times with

double-distilled water and dried in an oven at 150°C overnight prior to use.

294

296 The validation of the results obtained by the chloride-ISE was performed using an 297 official standard method [22] and showed excellent correlation between the two techniques ($R^2 = 0.9963$) over an approximate range between 1 and 300 mg L⁻¹ chloride 298 299 with no systematic errors. And last but not least, the drift of the electrode contacting a 300 13.4 mM chloride solution is less than 0.4 mV h⁻¹, measured at constant temperature and 301 with ISE and reference electrode continually immersed. 302 Stock solutions of KNO₃ (for nitrate), NaHCO₃ (for bicarbonate), and KCl (for 303 chloride) were prepared by dissolving the appropriate amount of reagent in distilled-304 deionized water. In order to carry out the measurements, the three aforementioned ISEs 305 were then simultaneously immersed in solutions obtained by suitable mixtures of the 306 stock solutions, in such a way that the following concentrations were selected: (a) 0.0, 307 $0.5, 1.0, 1.5, \text{ and } 2.0 \text{ mg L}^{-1} \text{ of nitrate, (b) } 0, 125, 150, 175, \text{ and } 200 \text{ mg L}^{-1} \text{ of }$ bicarbonate, and (c) 0, 50, 100, 150, and 200 mg L⁻¹ of chloride. The contents of the 308 309 three analytes were then measured in combinations of these solutions in threes (Table 310 2). 311 5 Results and discussion 312 313 314 The results achieved are summarized in Figures 3 and 4. Figure 3 shows the values 315 obtained without applying the proposed methodology. The horizontal axis represents the 316 true values of [NO₃-] (solutions were prepared by us following the procedure described 317 in the previous section, and are therefore known values), whereas the vertical axis 318 represents the measured concentrations, i.e. the experimental values provided by the 319 ISEs. Needless to say that an *ideal* nitrate sensor should give rise to a 45-degree line to 320 which the red line of the figure approximates a lot. This line reflects the results obtained 321 with the nitrate-ISE after its calibration and in the absence of interferences (i.e. 322 immersed in a standard nitrate solution). On the other hand, when the interfering anions 323 (chloride and bicarbonate) are added at known concentrations, the measured values start 324 to distance themselves from real concentrations owing to the interferences. 325 Consequently, an uncertainty area appears depending on the concentration of the 326 interfering species. For the concentration ranges of the interferents (see section 4), this 327 uncertainty area is the one between the upper and lower lines.

As it can be inferred from our study, and on the basis of a known nitrate concentration

329	(i.e. any point at the horizontal axis), the response of nitrate-ISE may vary – as a
330	function of the concentration of interferents – in the range resulting from the
331	intersection of the vertical line at this point with the uncertainty area. As an example, for
332	a known value of $[NO_3^-] = 2 \text{ mg } L^{-1}$ the results that could be obtained by measuring
333	with the corresponding ISE would lie between 2.04 mg L ⁻¹ (no interferences) and 2.52
334	mg L^{-1} (corresponding to [Cl ⁻] = 200 mg L^{-1} and [HCO ₃ ⁻] = 200 mg L^{-1}). It can be
335	noticed that – depending on the concentrations of interferents – the error obtained can
336	be significant, up to 26 %.
337	On the other hand, attention should also be paid to the real case of the need to determine
338	(using a NO ₃ ⁻ -ISE) the concentration of nitrate in the presence of an unknown amount
339	of chloride and bicarbonate. In these circumstances, the starting point is the vertical axis
340	(measured values of nitrate-ISE); true nitrate concentrations in the sample may be
341	anyone corresponding to the intersection of a horizontal line passing at this point with
342	the uncertainty area. For instance, for a measured concentration of 2 mg L ⁻¹ , true values
343	of $[NO_3^-]$ in the analyzed sample may be in the range between 1.56 mg L^{-1} (highest
344	concentration of interferents) and 1.96 mg L ⁻¹ (no interferences). The measurement
345	uncertainty is again really high.
346	Figure 4 shows the results obtained after the data treatment of previous measurements
347	with the method described in section 2. The values yielded by a node using this
348	procedure would also generate (as described above for Figure 3) an uncertainty area,
349	although in this case it is significantly smaller. This fact makes the measurements from
350	this node to be virtually free from the effects of the interferences, unlike in case this
351	methodology would not be used.
352	Indeed, if a given value of [NO3-] is now considered and a vertical line is drawn through
353	this point, its intersection with the uncertainty area yields a much lower dispersion of
354	values. In case of the above mentioned example ($[NO_3^-] = 2.00 \text{ mg L}^{-1}$), the values
355	obtained vary between 1.99 mg L ⁻¹ (for 100 mg L ⁻¹ chloride and 125 mg L ⁻¹
356	bicarbonate) and 2.07 mg L ⁻¹ (for 150 mg L ⁻¹ chloride and 200 mg L ⁻¹ bicarbonate), that
357	is to say, with a maximum error of 3.5 %, far from the aforementioned 26 %.
358	Again, we return to the real case of the need to determine (using a NO ₃ -ISE) the
359	concentration of nitrate in the presence of an unknown amount of chloride and
360	bicarbonate. A point is taken at the vertical axis (the value corresponding to the
361	measurement obtained using the proposed methodology), for instance 2.00 mg L^{-1}
362	nitrate; when a horizontal line is drawn through this point, it intersects with the

uncertainty area at values ranging from 1.93 mg L⁻¹ (for 200 mg L⁻¹ chloride and 175 363 mg L⁻¹ bicarbonate) to 1.99 mg L⁻¹ (for 50 mg L⁻¹ chloride and 125 mg L⁻¹ bicarbonate). 364 365 This demonstrates that the resulting nitrate concentration is very close to the true value, 366 the error being thus considerably reduced. 367 6. Conclusions 368 369 370 This work presents a new methodology to develop smart sensors with interference 371 tolerance based on simple Ion Selective Electrodes (ISE). For this purpose, a correction 372 mechanism based on interferent concentration measurement, error estimation, and a 373 correction procedure has been added. This method permits to "reject" the mutual 374 contributions to the analytical signals of ISEs due to the presence of interfering species 375 in the analyzed samples. Basically, it consists of applying statistical techniques –linear 376 regression of experimental data- to adjust the linear coefficients of each interferent on 377 the basis of the experiments observed. 378 This has been incorporated into the design and implementation of a nitrate smart sensor 379 for WSN applications that is able to overcome the problem associated with interfering 380 anions in ground waters, namely, chloride and bicarbonate. The procedure has been 381 applied to the measurements obtained from a nitrate-ISE in the presence of those two 382 anions by means of a microcontroller. The smart sensor node is then composed of the 383 three ISEs used (NO₃⁻, Cl⁻, HCO₃⁻) along with a low-cost, low-consumption 384 microcontroller. All three ISE's are simultaneously immersed in a water sample; the 385 corresponding measurements obtained are then conveniently received and processed by 386 the microcontroller. The final result after applying the proposed method is that the 387 nitrate measurements performed by this smart sensor turn out to be virtually 388 interference-free. 389 Finally, the significance of this proposal lies in the application of automatic procedures 390 implemented in microcontrollers in order to compensate physical deficiencies, such as 391 the impact of interferences, in measurement processes. This proposal may be extended 392 to increase the smartness of sensor nodes in order to enhance their features. 393 New-generation smart sensors applying these principles may be suitable for integration in Wireless Sensor Networks, on which Internet of Things and Ambient Intelligence will 394 395 be based.

396

Acknowledgements

398

397

- 399 The authors gratefully acknowledge the financial support from the Valencian Regional
- 400 Government under Research Project GV/2014/012, Polytechnic University of Valencia
- 401 (Research Project UPV PAID-06-12) and Spanish Government (Research Projects
- 402 CTM2011-29691-C02-01 and TIN2011-28435-C03-0).

403

References

405

- 406 [1] L.M. Nollet (Ed.), Handbook of water analysis, CRC Press, Boca Raton, FL (USA),
- 407 2007.
- 408 [2] G.J. Moody, J.D.R. Thomas, Selective Ion-sensitive Electrodes, Sel. Annu. Rev.
- 409 Anal. 3 (1973) 59-138.
- 410 [3] W. Simon, E. Pretsch, D. Ammann, W.E. Morf, M. Güggi, R. Bissig, M. Kessler,
- Recent developments in the field of ion selective electrodes, Pure Appl. Chem. 42
- 412 (1975) 613-626.
- 413 [4] J.V. Capella, A. Bonastre, R. Ors, M. Peris, In line river monitoring of nitrate
- 414 concentration by means of a Wireless Sensor Network with energy harvesting, Sens.
- 415 Actuators B: Chem. 177 (2013) 419–427.
- 416 [5] J.V. Capella, A. Bonastre, R. Ors, M. Peris, A step forward in the in-line river
- 417 monitoring of nitrate by means of a wireless sensor network, Sens. Actuators B: Chem.
- 418 195 (2014) 396-403.
- 419 [6] G. Dimeski, T. Badrick, A.S. John, Ion Selective Electrodes (ISEs) and interferences
- 420 A review, Clin. Chimica Acta 411 (2009) 309-317.
- 421 [7] A.V. Legin, E.A. Bychkov, B.L. Seleznev, Y.G. Vlasov, Development and analytical
- evaluation of a multisensor system for water quality monitoring, Sens. Actuators B:
- 423 Chem. 27 (1995) 377–379.
- 424 [8] R. Martínez-Máñez, J. Soto, E. García-Breijo, L. Gil, J. Ibáñez, E. Llobet, An
- 425 "electronic tongue" design for the qualitative analysis of natural waters, Sens. Actuators
- 426 B: Chem. 104 (2005) 302–307.

- 427 [9] A.V. Legin, A.M. Rudnitskaya, Y.G. Vlasov, C. di Natale, A. D'Amico, The features
- 428 of the electronic tongue in comparison with the characteristics of the discrete ion-
- 429 selective sensors, Sens. Actuators B: Chem. 58 (1999) 464–468.
- 430 [10] A.V. Mueller, H.F. Hemond, Extended artificial neural networks: Incorporation of a
- 431 priori chemical knowledge enables use of ion selective electrodes for in-situ
- 432 measurement of ions at environmentally relevant levels, Talanta, 117 (2013) 112-118.
- 433 [11] P. Bühlmann, E. Pretsch, E. Bakker, Carrier-Based Ion-Selective Electrodes and
- Bulk Optodes. 2. Ionophores for Potentiometric and Optical Sensors, Chem. Rev. 98
- 435 (1998) 1593-1687.
- 436 [12] C.G. Zoski (Ed.), Handbook of Electrochemistry, Elsevier, Amsterdam, 2007.
- 437 [13] Y. Umezawa, P. Bühlmann, K. Umezawa, K. Tohda, S. Amemiya, Potentiometric
- 438 selectivity coefficients of ion-selective electrodes, Pure and Applied Chemistry, 72
- 439 (2000) 1851-2082.
- 440 [14] S. Oteafy, S.H. Hassanein, Current hindrances in WSNs, John Wiley & Sons, New
- 441 York, NY (USA) 2014.
- 442 [15] J. Suhonen, M. Kohvakka, V. Kaseva, T.D. Hämäläinen, M. Hännikäinen, Low-
- Power Wireless Sensor Networks, Springer, New York, NY (USA), 2012.
- 444 [16] M.J. Moorkroft, J. Davis, R.G. Compton, Detection and determination of nitrate: a
- 445 review, 54 (2001) 785-803.
- 446 [17] Y. Liu, B.C. Seet, A. Al-Anbuky, Ambient Intelligence Context-Based Cross-
- Layer Design in Wireless Sensor Networks, Sensors 14 (2014) 19057-19085.
- 448 [18] O. Casas, M. López, M. Quílez, X. Martinez-Farre, G. Hornero, C. Rovira, M.R.
- Pinilla, P.M. Ramos, B. Borges, H. Marques, P. Girão, Wireless sensor network for
- smart composting monitoring and control, Measurement 47 (2014) 483-495.
- 451 [19] J.V. Capella, A. Bonastre, R. Ors, M. Peris, A wireless sensor network approach for
- distributed in-line chemical analysis of water, Talanta 80 (2010) 1789-1798.
- 453 [20] N. Zaman, K. Ragab, A.B. Abdullah, Wireless Sensor Networks and Energy
- 454 Efficiency: Protocols, Routing and Management, IGI Global, Hershey, PA (USA) 2012.
- 455 [21] E. Pretsch, The new wave of ion-selective electrodes, TrAC Trends Anal. Chem.
- 456 26 (2007) 46–51.
- 457 [22] G.W. Latimer (Ed.), AOAC International, Official Methods of Analysis of AOAC
- 458 International, 19th ed., AOAC, Washington D.C. 2012.

Legends	of figures
Figure 1	Structure of a nitrate smart sensor node. A = signal amplifier & conditioner.
Figure 2	Calibration plots for the determination of nitrate, bicarbonate, and chloride using the corresponding ion-selective electrodes. Activities are expressed in mol L ⁻¹ .
Figure 3	Experimental results obtained in the presence of interferences. Values between parenthesis on the right side of the figure denote the following: ([HCO ₃ -],[Cl-]) All concentrations are expressed in mg L ⁻¹ .
Figure 4	Experimental results obtained with interference tolerance. Values between parenthesis on the right side of the figure denote the following: ([HCO ₃ -],[Cl-]) All concentrations are expressed in mg L-1.
	Figure 1 Figure 2 Figure 3

Table 1.- Coefficients obtained in the regression analysis (concentrations in mg L⁻¹)

473
474
475

475 476	Coefficient	Value
477		
478		
479	Intercept	-0,108336504
480	$[NO_3^-]$	0,123178928
481	[C1 ⁻]	0,000920765
482	[HCO ₃ -]	0,000386705
483		

Table 2.- Initial concentration data set

$[NO_3^-]$, mg L ⁻¹		[Cl ⁻], mg L ⁻¹	_	[HCO ₃ -], mg L ⁻¹	TOTAL
0.5		125		50	
1.0	X	150	X	100	64 experiments
1.5		175		150	04 experiments
2.0		200		50	