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Additional Information

# **Ceria nanoparticles with Rhodamine B as a powerful theranostic agent against intracellular oxidative stress**

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† Electronic Supplementary Information (ESI) available: Additional scheme and figures.

## **ABSTRACT**

Ceria nanoparticles with Rhodamine B groups covalently attached on their surface (RhB-CeNPs) were successfully prepared to simultaneously exhibit antioxidant activity and the ability to detect oxidant species. In order to use them for biomedical purposes, the nanoparticles were internalized in two human cell lines (HeLa and Hep3B), confirmed by confocal microscopy. In addition, their biocompatibility was assessed by performing proliferation, viability and apoptosis assays, in which they showed a remarkable lack of toxicity. Thereafter, the antioxidant activity of RhB-CeNPs against reactive oxygen species (ROS) in a model of oxidative stress was demonstrated in HeLa cells using the dichloro-dihydro-fluorescein diacetate (DCFH-DA) assay. RhB-CeNPs exhibited higher cytosolic antioxidant activity than the well-established ceria nanoparticles. Surprisingly, the antioxidant capacity of RhB-CeNPs was evident when the ROS content of the cells increased notably (and was, therefore, harmful for those cells). Furthermore, the ability of RhB-CeNPs as ROS-content sensors was evaluated by measuring oxidative stress in HeLa cells using the intrinsic fluorescence of the Rhodamine B groups present on the nanoparticles. The results with respect to the detection and quantification of ROS were similar to those obtained with DCFH-DA, a typical method of quantifying intracellular ROS. Our results demonstrate the potential of RhB-CeNPs as remarkably biocompatible theranostic agents with application in the field of oxidative stress.

**Keywords:** Theranosis, ceria nanoparticles, oxidative stress, Rhodamine B, reactive oxygen species.

## INTRODUCTION

Oxidative stress is one of the main factors of cellular aging and other cellular disorders.<sup>1-4</sup> It is defined as an imbalance in the production of oxidant species and their effective removal by antioxidants and scavenger enzymes, and is one of the targets of therapies against diseases. In physiological conditions, a homeostatic balance exists between the formation of reactive oxygen species (ROS) and their elimination by endogenous antioxidant scavenging compounds and enzymes.<sup>5</sup> The free radical hypothesis for tissue dysfunction postulates that ROS leads to a modification of lipids, proteins and nucleic acids, which, in turn, contributes to the etiology of different diseases.<sup>6</sup> Importantly, low-to-moderate levels of ROS are physiologically normal and necessary for cell survival, while high levels are detrimental and can even cause cell death. For this reason, there is much current interest in developing new strategies to modulate ROS levels and, thus, control oxidative stress.<sup>7-11</sup> With the aim of exploring in detail the role of ROS, several strategies have been tested *in vivo* and *in vitro* in order to evaluate the benefits of reducing oxidative stress in diseases such as cancer, diabetes, among others, with some beneficial effects being observed. The outcome of these studies has fuelled interest in developing new compounds and materials with antioxidant properties such as nanoparticles to control cellular oxidative stress.

Nanoparticles are used in different fields, including advanced materials, chemistry, biology and medicine, and their synthesis and properties have been topics of continuous study in recent decades. The structure of nanomaterials allows the possibility of incorporating several guests into their cavities when they are porous, or of covalently anchoring several functional components to their surface due to the high percentage of

external atoms. Furthermore, the composition and chemical properties of nanoparticles can be designed to make them inert or reactive versus substrates or the environment. These features make nanoparticles particularly suitable for use in biology and biomedicine, where nanoparticles have been tested as drug delivery systems, and antioxidant or imaging agents, among other potential uses.<sup>12-17</sup> In spite of the range of opportunities offered by nanoparticles, which can exhibit multifunctionality, the most common application in biomedicine only exploits a single property of each one. However, recently there has been increased interest in applying nanomaterials as bifunctional or multifunctional materials. The multifunctionality of nanoparticles derives from their complementary reactivity, which is due to the simultaneous incorporation of different functional groups and molecules in their structure and moreover the intrinsic properties of the modified nanoparticle. In this way, some nanomaterials have been designed to combine optical and magnetic resonance imaging, therapy and optical imaging, radiotherapy and imaging, magnetism and imaging, among other possibilities.<sup>18-22</sup> In this context, one of the emerging applications of nanoparticles is theranosis, which combines therapy and imaging. However, to our knowledge, no one has explored the possibility of using bifunctional nanoparticles as fluorescent biosensors of the concentration of ROS in cells and acting as antioxidants when the ROS concentration overcomes the damage threshold for cells. We believe it is possible to generate “intelligent” nanoparticulate materials in the field of theranostics to detect ROS content and to treat excess ROS, thus modulating antioxidant therapies in a harmless manner. This is the objective of the present work.

Ceria nanoparticles (CeNPs) have been found to be biocompatible with several cellular types, exhibiting antioxidant activity due to their redox properties, which are

attributable to oxygen vacancies on their surface and the presence of  $\text{Ce}^{3+}$  ions in their structure.<sup>23,24</sup> In addition  $\text{CeO}_2$  NPs are stable in aqueous media at neutral pH values. The antioxidant activity of CeNPs can be enhanced by supporting metals in their structure, such as Au.<sup>25</sup> The biocompatibility and redox properties of CeNPs have attracted interest from the field of biomedicine, and these nanoparticles have been proposed for some therapeutic applications. Accordingly, CeNPs have been tested *in vitro* and *in vivo* as quenchers of ROS in some types of cells, and as radiation protectors, cardioprotectors or reducers of endothelial inflammation, among other roles.<sup>26-30</sup>

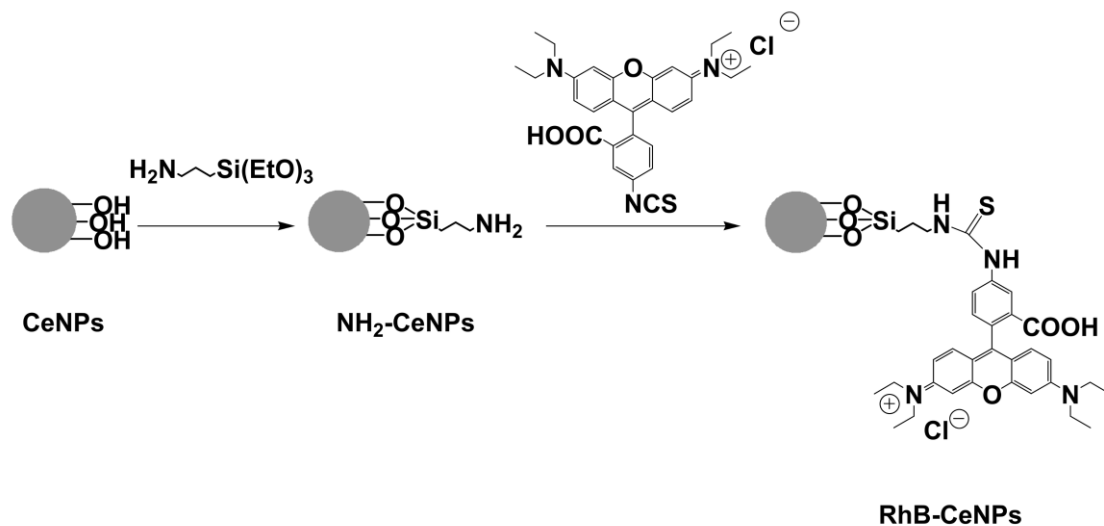
Rhodamine B (RhB) is a fluorophore with typical absorption and fluorescence bands centered approximately at 550 and 580 nm, respectively, and has been used for diverse purposes in chemistry, biology and medicine. The use of RhB derives mainly from its high fluorescence quantum yield and from the quenching of its fluorescence emission by energy transfer or electron transfer processes caused by the interaction of RhB with other compounds.<sup>31-35</sup> In this sense, radical species or hydrogen peroxide ( $\text{H}_2\text{O}_2$ ) can oxidize RhB, causing a decrease in fluorescence intensity that can correlate with the amount of oxidants, and is usually used to quantify the amount of oxidizing species.<sup>36-38</sup> Thus, RhB could be used as a sensor of the oxidant species present in a sample. In addition, some rhodamine dyes are currently used for biomedical purposes such as probes for mitochondria and the endoplasmic reticulum, where it appears to be relatively non-toxic.<sup>39-41</sup>

Due to the capacity of CeNPs as antioxidants and ROS scavengers, and the possibility of covalently attaching RhB to their surface to act as a sensor of oxidant species content,

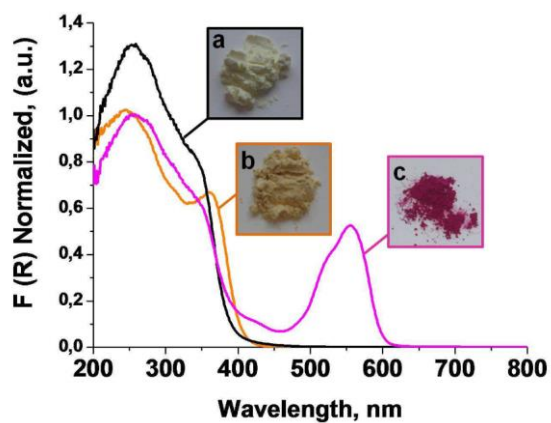
we have developed a novel theranostic nanoparticle to work both as a diagnostic agent of oxidant species content and a form of antioxidant therapy.

## RESULTS AND DISCUSSION

In order to prepare a new class of nanomaterial with properties as both theranostic antioxidant and oxidant content sensor, RhB was covalently attached to CeNPs by modifying their surface. For this purpose, colloidal CeNPs were obtained by hydrolysis of  $\text{Ce}^{4+}$  salts in water at a controlled basic pH and purified by dialysis. The average nanoparticle size was determined from the width of the XRD peaks by applying the Scherrer equation, giving a value of 5.6 nm (Fig. S1, ESI†). Additionally, the XRD pattern of nanoparticles confirmed that the sample corresponded with  $\text{CeO}_2$ , as the XRD coincided with that reported for  $\text{CeO}_2$ . Additionally, the UV-Vis spectrum of CeNPs was recorded. No bands at wavelengths longer than 400 nm were observed for CeNPs (Fig. 1). Once CeNPs had been characterized, covalent functionalization of their surface with amino groups was carried out by silylation of the surface OH groups with 3-aminopropyltriethoxysilane in dry toluene. In this way,  $\omega$ -aminopropyl-functionalized CeNPs ( $\text{NH}_2$ -CeNPs) were obtained. This reaction takes place under neutral conditions and attaches the amino group through strong Ce-O-Si bonds. These amino groups act as tethers that connect RhB to CeNPs. Subsequent reaction of these aminated  $\text{NH}_2$ -CeNPs with RhB isothiocyanate forms a strong thiourea linkage between the organic fluorophore and the inorganic CeNPs, as depicted in Scheme 1. The success of each synthesis step was assessed by chemical combustion analysis, diffuse reflectance UV-Vis spectroscopy (Fig. 1) and FT-IR (Fig. 2).

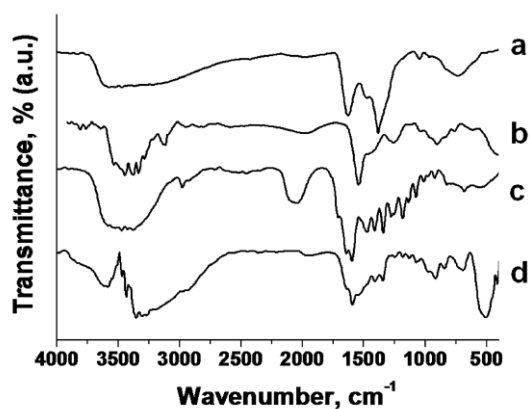


**Scheme 1.** Synthetic route followed to obtain  $\text{NH}_2\text{-CeNPs}$  and  $\text{RhB-CeNPs}$ .



**Fig. 1** Diffuse reflectance UV-Vis absorption spectra (plotted as the Kubelka-Munk function of the reflectivity,  $R$ ) of  $\text{CeNPs}$  (a),  $\text{NH}_2\text{-CeNPs}$  (b) and  $\text{RhB-CeNPs}$  (c).

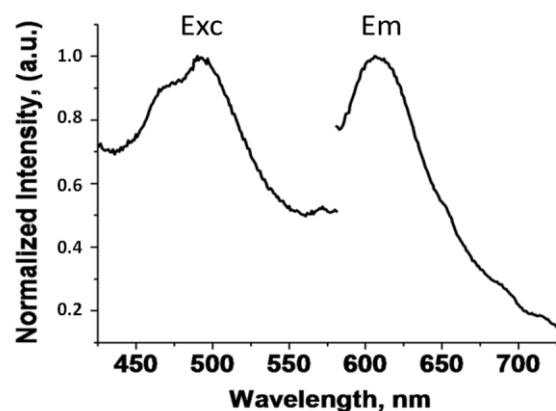




**Fig. 2** FT-IR spectra of a) CeNPs, b) NH<sub>2</sub>-CeNPs, c) Rhodamine B isothiocyanate and d) RhB-CeNPs.

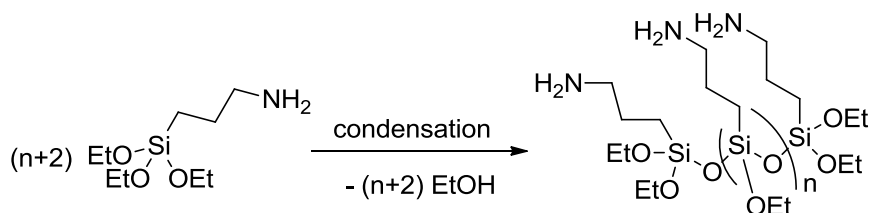
The presence of 3-aminopropyl groups anchored on the ceria surface was evident in the IR spectra of NH<sub>2</sub>-CeNPs; the two vibration bands characteristic of symmetric and asymmetric NH<sub>2</sub> (3270 and 3230 cm<sup>-1</sup>) and CH<sub>2</sub> stretching (2990 and 2950 cm<sup>-1</sup>) which were recorded as weak bands due to the low population of these groups. The loading of aminopropyl silyl groups on NH<sub>2</sub>-CeNPs was 4.1 mmol×g<sup>-1</sup> based on the C and N content in this material, which was determined by combustion chemical analysis. Additionally, the XRD pattern of NH<sub>2</sub>-CeNPs with respect to CeNPs shows that surface functionalization occurred without a significant increase in the average particle of CeNPs that remained at roughly 5.6 nm (Fig. S1, ESI†). Subsequently, covalent attachment of RhB on NH<sub>2</sub>-CeNPs was carried out, producing a change in the colour of the NH<sub>2</sub>-CeNPs sample after functionalization that varied from light yellow to purple and was reflected in the diffuse reflectance spectra of the samples (Fig. 1). This change in the color upon surface functionalization of CeO<sub>2</sub> by ω-aminopropyl groups could indicate the occurrence of some ligand to metal charge transfer complex between amino groups and of the silane and Ce<sup>4+/3+</sup> ions having free coordination positions. As shown

in Figure 1, the RhB-CeNPs sample exhibited an absorption band in the 500-600 nm region due to the anchored RhB dye. This RhB absorption band was somewhat broader than the optical spectrum of the Rhodamine thioisocyanate in solution. This broadness is a general phenomenon observed when organic dyes are supported on solid surfaces and is due to the formation of aggregates and/or the inhomogeneity of the surface in terms of polarity and presence of OH groups. Furthermore, as expected for the presence of RhB in RhB-CeNPs, this dye produced an intense emission. Figure 3 shows the excitation spectrum showing that RhB emission is observed upon excitation in a wide range of wavelengths going from 425 to 575 nm, coinciding with the absorption band (see Figure 1, plot c). No variation of the emission spectrum as a function of the excitation wavelength was observed and the emission maximum appeared at 600 nm. A suitable excitation wavelength was 510 nm.



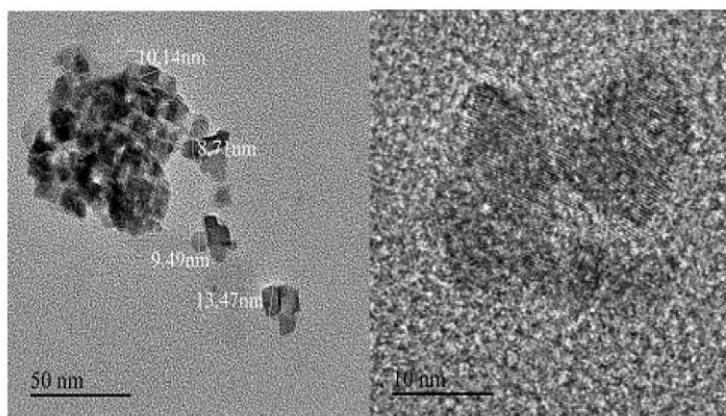
**Fig. 3** Excitation (Exc,  $\lambda_{exc}$  = 510 nm) and fluorescence (Em,  $\lambda_{em}$  600 nm) spectra recorded for RhB-CeNPs.

Additionally, the anchoring of RhB was confirmed by FT-IR spectroscopy, which provided firm evidence of the success of the incorporation. Thus, upon introduction of RhB isothiocyanate (Fig. 2, spectrum d) the isocyanate peak of the RhB isocyanate precursor appearing at  $2055\text{ cm}^{-1}$  disappeared, but the characteristic narrow peaks corresponding with the aromatic region of the xanthene core were clearly resolved in the RhB-CeNPs spectrum (Fig. 2, spectrum d). Combustion chemical analysis of S allowed us to determine the loading of RhB in the resulting RhB-CeNPs, which was  $3.2\text{ mmol}\times\text{g}^{-1}$ , corresponding with about 80 % of the  $\text{NH}_2$  groups present in RhB-CeNPs. Considering that the specific surface area of  $\text{CeO}_2$  is  $180\text{ m}^2\times\text{g}^{-1}$ , these loading results in average about 100 molecules per  $\text{nm}^2$ . This average is clearly much too high and indicates that Scheme 1 is an over simplification and there must be under the preparation conditions a certain degree of self condensation of  $\omega$ -aminopropyl triethoxysilanes as indicated in Scheme 2. These oligosiloxane condensates will be the species covalently attached to the surface of  $\text{CeO}_2$ .



Scheme 2. Condensation of  $\omega$ -aminopropyl triethoxysilane forming oligomers and resulting in a high RhB loading on  $\text{CeO}_2$

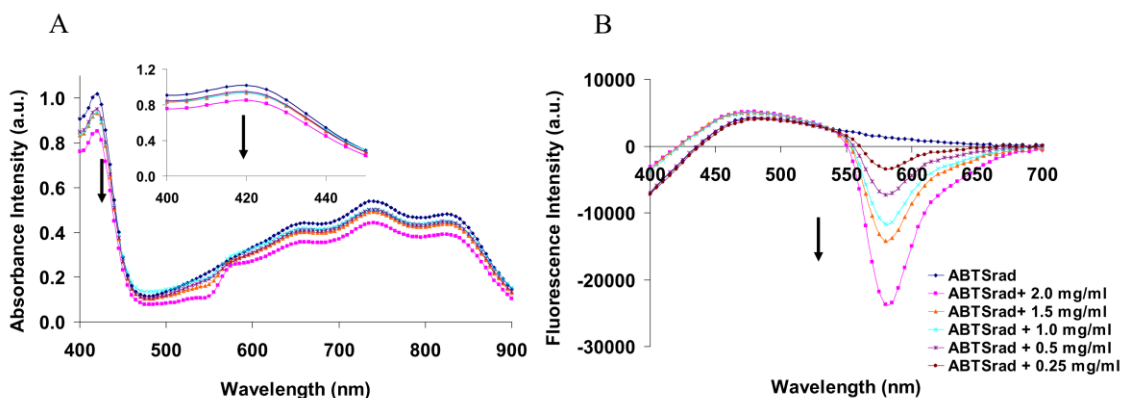
To determine the particle size of RhB-CeNPs, a statistical analysis of RhB-CeNPs was performed by transmission electron microscopy (TEM), the size distribution revealing an average particle size of about 10 nm (Fig. 4).



**Fig. 4** Transmission electron microscopy recorded for RhB-CeNPs.

In addition, RhB-CeNPs were submitted to dynamic light scattering study to determine their size distribution profile in suspension (Fig. S2, ESI†) and their zeta potential value (Fig. S3, ESI†). These data provide information about the surface modification of the CeNPs due to anchor RhB and the size and charge of the nanoparticles in aqueous solution, which is the medium used to biomedical purposes. Hydrodynamic size of RhB-CeNPs in water was about 54 nm, which was higher than the value determined by TEM (Fig. 4), but this may have reflected partial agglomeration of the nanoparticles and/or the presence of a cloud of water molecules solvating the RhB-CeNPs core. It should be noted that even though the surface of pristine nanoparticulate CeO<sub>2</sub> is hydrophilic, upon modification with silanization and RhB dye the hydrophobicity of the particles has to increase. This hydrophobicity is responsible for the occurrence of some agglomeration in water to decrease the surface energy of the colloidal suspension. Regarding the stability of these nanoparticles, the zeta potential value at neutral pH was -25 mV, which is within the limit to be considered a persistent colloid. This zeta potential value seems to be the balance between the intrinsic negative value of CeNPs and the positive charges introduced by RhB.

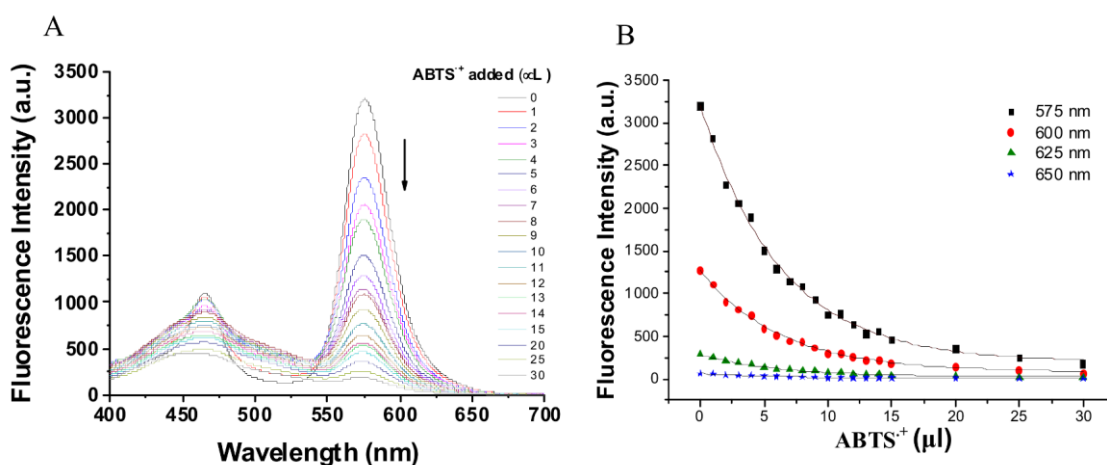
Once the RhB-CeNPs were obtained, their antioxidant capacity was assessed using the ABTS radical scavenger test. This test consists in measuring the decrease of absorbance corresponding to ABTS<sup>•+</sup> (typically 410 nm) when a radical scavenger or antioxidant is added to the solution. The test is carried out by following basically the methodology described by Seeram, *et al.*<sup>42</sup> Thus, initially ABTS<sup>•+</sup> needs to be obtained by passing a colorless 5 mM aqueous solution of ABTS through a filter containing MnO<sub>2</sub>. The resulting green solution of ABTS<sup>•+</sup> is centrifuged at 10000 rpm for 10 min at 20 °C to remove any trace of MnO<sub>2</sub>. Freshly prepared supernatant is used to assess the antioxidant capacity of RhB-CeNPs by monitoring the decrease of the corresponding ABTS<sup>•+</sup> absorbance at 410 nm when different concentrations (0, 0.5, 1, 1.5, and 2 mg/ml) of nanoparticles dispersed in PBS solution are added to the green solution. In order to avoid any possible interference in the UV-Vis ABTS<sup>•+</sup> spectrum due to the addition of RhB-CeNPs, the UV-Vis spectrum of a RhB-CeNPs control solution is measured simultaneously to subtract to the radical absorbance spectrum. As shown in Figure 5A, ABTS<sup>•+</sup> absorbance decreased gradually upon RhB-CeNPs addition. This fact confirms the antioxidant activity of the PBS solution of RhB-CeNPs and, therefore, the suitability of these nanoparticles as antioxidant therapeutic agents.



**Fig. 5** UV-Vis (A) and fluorescent (b,  $\lambda_{ex}$  350 nm) (B) spectra of ABTS radical cation after adding different amounts of RhB-CeNPs. The insert shows the region where the decrease of the band is related with the radical scavenger or antioxidant assessment.

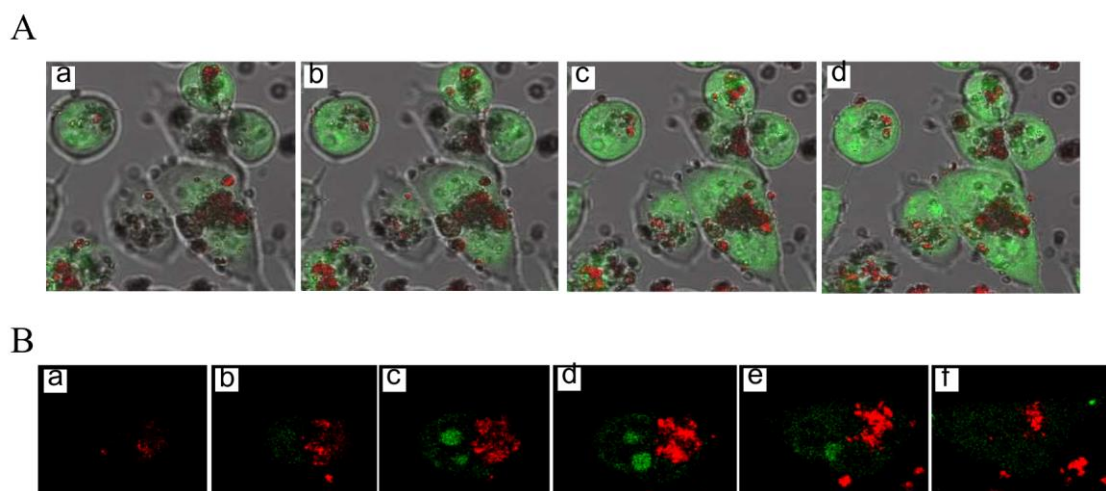
On the other hand, RhB-CeNPs have RhB groups covalent anchored to the CeNPs surface that could be used as sensors to quantify the content of oxidant species. This assumption is based on the well-known capacity of RhB to react with oxidants such as radical species and  $H_2O_2$ , among others. This quenching can be followed by the disappearance of the fluorescence band at 580 nm, after excitation at 350 nm, characteristic of the RhB groups present in RhB-CeNPs. These fluorescence experiments were performed with the same samples used to establish the antioxidant capacity of these nanoparticles (Fig. 5B), after checking for the absence of fluorescence originating from  $ABTS^{\bullet+}$  or ABTS. Additionally, the capacity of RhB-CeNPs as oxidant content sensors was confirmed by a similar experiment in which the RhB-CeNPs content was maintained constant and the concentration of  $ABTS^{\bullet+}$  was increased. For this purpose, the fluorescence spectra were recorded between 400-700 nm after exciting the sample at 350 nm. In this experiment, the characteristic fluorescence intensity of RhB present on RhB-CeNPs decreased progressively following the addition of  $ABTS^{\bullet+}$

(Fig. 6A). The decrease of fluorescence intensity at different wavelengths (for example 575, 600, 625 and 650 nm) followed a first order exponential plot versus the content of ABTS<sup>•+</sup> as an oxidant compound model (Fig. 6B), and confirmed the ability of RhB-CeNPs to quantify the amount of oxidant species. This capacity allows the determination of the concentration of ROS and H<sub>2</sub>O<sub>2</sub>, among others, in solution or in the cells, thus allowing their use as biosensors where the oxidant content may be observed by a decrease of the fluorescent band between 575-650 nm that correspond to RhB-CeNPs.



**Fig. 6** A) Fluorescent spectra of RhB-CeNPs in PBS solution recorded after the excitation at 350 nm and the addition of different aliquots of a 5 mM ABTS<sup>•+</sup> aqueous solution. B) Reduction of the fluorescence intensity of Rh-CeNPs solution at different wavelengths (575, 600, 625, 650 nm) after the addition of different aliquots of the ABTS<sup>•+</sup> aqueous solution. All the fitting are successfully adjusted to a first order exponential plot (575 nm:  $y = 3007.815 \times e^{(-x/6.258)} + 200.502$ ,  $R^2 = 0.9965$ ; 600 nm:  $y = 1185,720 \times e^{(-x/6.212)} + 85.120$ ,  $R^2 = 0,9972$ ; 625 nm:  $y = 260.635 \times e^{(-x/6.351)} + 22.877$ ,  $R^2 = 0.9957$ ; 650 nm:  $y = 59.785 \times e^{(-x/6.281)} + 5.668$ ,  $R^2 = 0.9961$ ).

Before studying the antioxidant behaviour of RhB-CeNPs in cells, their ability to become internalized by cells was confirmed by confocal laser scanning microscopy, by which we monitored the characteristic red fluorescence of the RhB group present in the RhB-CeNPs. Figure 7 shows a selected set of vertical Z confocal fluorescence images showing the intracellular location of RhB-CeNPs nanoparticles (red spots). The cytoplasmic distribution of RhB-CeNPs can be observed clearly in the enlarged image of a single cell shown in Figure 7 (lower panel). These results are compatible with previous reports that similar CeNPs samples can enter cells.<sup>15</sup>



**Fig. 7** Analysis of the cellular up-take of RhB-CeNPs by confocal microscopy. HeLa cells were exposed to the nanoparticles for 3h and live imaging was performed thereafter. Representative images of z-stack (*a* to *d* or *a* to *f*). A) Images show the overlapping of light microscopy and fluorescence microscopy (the cytoplasm was stained with N-Acridine orange (NAO) fluorescing in green). B) Single cell images show the fluorescent green signal coming from the nucleic acid stain SYTO80 whereas RhB-CeNPs autofluoresce in red.



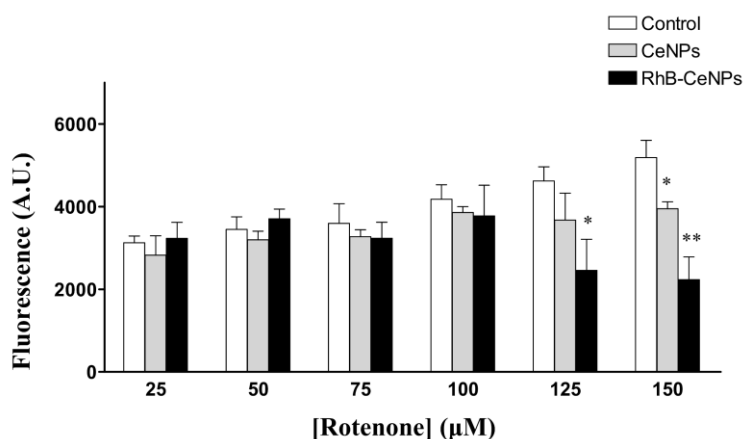
Once RhB-CeNPs had been confirmed as both antioxidant agents and oxidant content sensors in aqueous solution and were shown to be internalized inside cells, their toxicity and capacity as sensors and antioxidants were evaluated in human cell culture models. Cytotoxicity is a major issue in the applicability of nanoparticles in biomedicine. For this reason, several experiments were performed to assess the effect of RhB-CeNPs on the viability, survival and proliferation of cultured cells. Two human cell lines (HeLa and Hep3B) were used in order to avoid cell-type specificity. The results of the cell viability assessment (MTT assay) showed that the addition of CeNPs to the culture medium (incubation for 3 h followed by 21 h recovery) had no effect on the viability of either HeLa or Hep3B cells. RhB-CeNPs significantly diminished the viability of HeLa cells (approximately 50%), whereas only a slight and non-significant decrease was detected in Hep3B cells (Fig. S4, ESI†). Cell survival and exponential proliferation was studied further in both cell lines over a period of 3 days (fluorescence microscopy). RhB-CeNPs did not have a significant effect on cell survival or cell proliferation rate in either of the two cell lines, and results were similar to those observed for the control (untreated cells) at all the time points analyzed (24, 48 and 72 h). This suggests that the effect produced on cell viability in HeLa cells under treatment with RhB-CeNPs did not have major consequences on the survival of these fast proliferating cells over a longer period of time.

In order to confirm and expand the findings of the proliferation/survival experiments showing the lack of a cytotoxic effect, we determined the capacity of RhB-CeNPs to induce apoptosis in HeLa and Hep3B cells (incubation for 3h followed by 21h recovery). The widely used kinase inhibitor staurosporine was employed as a positive control for apoptotic induction. Several typical markers of apoptotic cell death were

analyzed by fluorescence microscopy. An increase in mean Hoechst 44432 fluorescence (nuclear staining), indicative of increased chromatin condensation, which is a hallmark of apoptosis, was detected in both cell types after 24 h-treatment with staurosporine. Of note, neither CeNPs nor RhB-CeNPs provoked any alteration in this parameter (Fig. S5A, ESI†). Similarly, a lack of apoptotic induction was observed when a Bivariate Annexin V/PI analysis was performed. Four distinct subpopulations of cells were evaluated: vital (Annexin V/PI), early apoptotic or classically apoptotic (Annexin V<sup>+</sup>/PI), late apoptotic/necrotic (Annexin V<sup>+</sup>/PI<sup>+</sup>), and damaged or classically necrotic (Annexin V/PI<sup>+</sup>). The results are shown in Figure S5B (See ESI†) (bar charts representing summary of the mean fluorescence data for both fluorochromes in both cell types). Figure S5C (See ESI†) depicts a representative cytogram of Hep3B cells in which the positive control (staurosporine 1  $\mu$ M) triggered typical apoptosis in 45 % of the cells compared to the presence of only 1.2 % apoptotic cells in the negative control. CeNPs did not provoke a significant increase in the apoptotic subcellular population (3.77 % apoptotic cells in the presence of CeNPs). Moreover, there was a lack of a significant increase in the number of necrotic cells. This double staining experiment could not be performed for RhB-CeNPs due to the intense red fluorescence of RhB that interferes with the PI emission of the assay. To sum up, the obtained evidence indicates that short-term exposure to the nanoparticles under study does not trigger cell death in cultured human cells.

The oxidant content of the cells was measured by means of the DCFH-DA assay, which is one of the most usual methods of detecting intracellular ROS for oxidative stress assessment.<sup>7,9,10</sup> The results obtained when we tested the activity of CeNPs and RhB-CeNPs against oxidative stress in HeLa cells are shown in Figure 8. Both CeNPs and

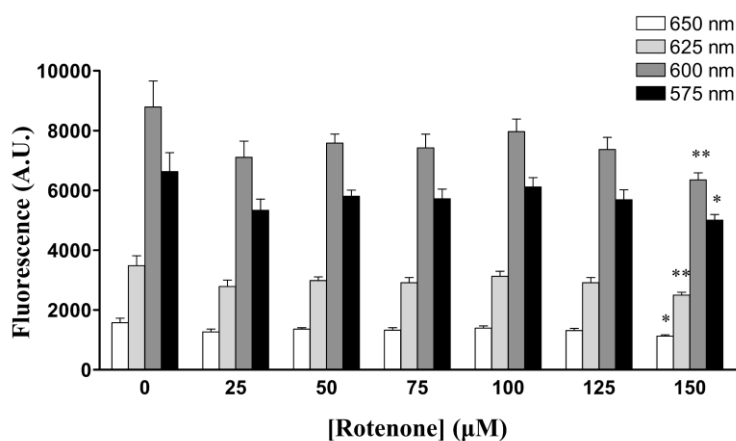
RhB-CeNPs reduced the oxidant content of HeLa cells, but only at high rotenone concentrations (125  $\mu$ M,  $P < 0.05$ ; 150  $\mu$ M,  $P < 0.01$ ), while the latter were more active in reducing cellular oxidative stress. There are numerous reports describing the anti-ROS activity of CeNPs, so it is expected that, when RhB are anchored to the surface of CeNPs, the antioxidant activity of these nanoparticles increases significantly, probably due to the tendency of the RhB group to undergo oxidation, as previously discussed.<sup>15,26,28</sup>



**Fig. 8** Effect of CeNPs (20  $\mu$ g/mL), RhB-CeNPs (20  $\mu$ g/mL) on Rotenone-induced ROS production. Bar charts showing DCFH fluorescence intensity in HeLa cells versus different Rotenone concentration. Data (mean  $\pm$  SEM,  $n=3$ ) were analyzed by 1 way ANOVA test, significance vs control (untreated cells) \*  $p < 0.05$  and \*\*  $p < 0.01$ .

On the other hand, the ability of RhB-CeNPs as ROS content sensors was studied by recording the fluorescence emission of cells which had been previously incubated with the same amount of RhB-CeNPs and after the addition of different rotenone amounts at 575, 600, 625 and 650 nm. As shown in Figure 9, the decrease in the fluorescence

intensity at different wavelengths (575, 600, 625 and 650 nm) owing to RhB-CeNPs was higher when the oxidative stress was increased by higher rotenone concentrations.



**Fig. 9** Effect of applying different concentration of Rotenone on the fluorescence intensity of RhB-CeNPs (20 μg/mL) at different wavelengths (575, 600, 625 and 650 nm) in HeLa cells. Data (mean ± SEM, n=8) were analyzed by 1 way ANOVA test, significance vs control (untreated cells) \* p<0.05 and \*\* p<0.01.

The data correlated with the results of the DCFH-DA probe assay, which is the most usual method of checking the ROS content of cells. These fluorescence intensity data confirm the capacity of RhB-CeNPs as a theranostic nanomaterial that acts simultaneously as a ROS quencher and biosensor. In addition, RhB-CeNPs as sensors have the additional advantage of being able to report at different wavelengths without losing sensor capacity for ROS species.

## **EXPERIMENTAL SECTION**

### **Solvents and reagents**

Unless otherwise indicated, the solvents and reagents employed in both the preparation of materials and in cell culture were purchased from Sigma-Aldrich (Madrid, Spain), Scharlab (Sentmenat, Spain), Life Technologies (Madrid, Spain), or GIBCO (Invitrogen, Eugene, OR) and were used in the form in which they were supplied. The Annexin-V-FLUOS staining kit was from Roche Diagnostics (Sant Cugat del Vallés, Spain). Gases were supplied by Air-Liquide (Valencia, Spain). 96- and 48-well plates were purchased from Falcon BD (Madrid, Spain) and the Nunc 8-well coverslip bottom chamber slides for confocal microscopy were supplied by Thermo Scientific (Madrid, Spain). 2,7-dichlorodihydrofluorescein diacetate (DCFH-DA) was purchased from Life Technologies (Madrid, Spain).

### **Instrumentation**

#### **Synthesis and characterization of nanoparticles**

Combustion chemical analysis (C, H, N, and S) were performed in a FISON CHNOS analyzer. UV-Vis spectra were recorded in transmission mode on a Shimadzu UV-Vis spectrophotometer using quartz cuvettes of 1 cm path length. Fluorescence measurements were performed with a Synergy Mx plate reader fluorimeter (BioTek Instruments, Winooski, VT) or on a JASCO FP-8500 spectrofluorometer (Mary's Court, Easton, MD) provided with a monochromator of variable wavelength in the range of

200-900 nm. FT-IR spectra were recorded on a Nicolet 710 FT-IR spectrophotometer using KBr disks or self-supported wafers (10 mg) compressed to 2 Ton cm<sup>2</sup> for 2 min. Transmission electron microscopy (TEM) images were acquired by a Philips CM300 FEG system with an operating voltage of 100 kV by placing microdrops of CeNPs suspensions directly onto a copper grid coated with carbon film (200 mesh). Powder XRD was recorded with a Philips X'Pert diffractometer using the Cu K radiation at a scan rate of 2° min<sup>-1</sup>. Dynamic Light Scattering used to determine the size and zeta potential of colloidal aqueous solutions of nanoparticles was performed on a Zetasizer Nano ZS (Malvern Instrument, Malvern, UK). 5 mg of nanoparticles were suspended in 100 mL for measurements, which were carried out in an automatic mode. The values of size and zeta potential are the average of 30 runs. The centrifuge used was a Hettich-Zentrifugen one (Tuttlingen, Germany).

## **Cell culture**

The incubator used for cell culture was an IGO 150 (Jouan, Saint-Herblain, France). Confocal images were acquired using a Leica TCS-SP2 confocal laser scanning unit equipped with argon and Helium-Neon laser beams and attached to a Leica DM1RB inverted microscope. In the apoptosis studies, fluorescence was quantified using the IX81 Olympus fluorescence microscope coupled with static cytometry software "ScanR" version 2.03.2 Olympus (Hamburg, Germany) Fluorescence measurements for ROS quantification were performed with a Synergy Mx plate reader fluorimeter (BioTek Instruments, Winooski, VT). Colorimetric analyses (MTT assay) were performed using a multiscan Thermo Labsystems plate-reader spectrophotometer (Thermoscientific, Rockford, IL).

## **Synthesis of Rhodamine B-functionalized CeNPs (RhB-CeNPs)**

### **Synthesis of CeNPs**

CeNPs were prepared by hydrolysis of  $\text{Ce}(\text{NO}_3)_4$  at pH=9 using  $(\text{NH}_4)\text{OH}$  as reported.<sup>15</sup> In brief, an ammonia aqueous solution (1.12 L, 0.8 M) was added under constant stirring and at room temperature over 375 mL of  $\text{Ce}(\text{NO}_3)_4$  (0.8 M), and the dispersion was heated in a poly(ethyleneterephthalate) vessel at 100 °C for 24 h. The resulting yellow solid was filtered and dried under vacuum overnight. The success of the synthesis was determined by XRD, UV-Vis and FT-IR. The surface area of CeNPs measured by isothermal  $\text{N}_2$  adsorption at -198 °C using a Micromeritics ASAP2000 and applying the Brunauer-Emmett-Teller (BET) algorithm was  $180 \text{ m}^2 \times \text{g}^{-1}$ .

### **Preparation of 3-aminopropyl-functionalized CeNPs ( $\text{NH}_2$ -CeNPs)**

Silylation of CeNPs was carried out by stirring a suspension of 1 g of dried CeNPs sample for 3 h under atmospheric pressure) in 15 mL toluene containing 0.5 mL of 3-(aminopropyl)triethoxysilane at reflux temperature for 2 h. Afterwards, the mixture was allowed to cool at room temperature and  $\text{NH}_2$ -CeNPs was decanted and washed with fresh toluene aliquots ( $3 \times 2 \text{ mL}$ ) and allowed to dry, again at room temperature. The percentage of aminopropyl groups was determined by combustion elemental analysis and the success of the functionalization was further assessed by UV-Vis, XRD and FT-IR.

### **Preparation of Rhodamine B-functionalized CeNPs (RhB-CeNPS)**

0.5 g of NH<sub>2</sub>-CeNPs was suspended in 15 mL of CH<sub>2</sub>Cl<sub>2</sub>, to which 0.100 g of Rhodamine B thioisocyanate was added. The suspension was stirred magnetically at reflux temperature for 4 h. After this time, the solid was collected and washed with fresh CH<sub>2</sub>Cl<sub>2</sub> until it became colorless (7×20 mL). The resulting RhB-CeNPs was dried at room temperature under reduced pressure. Following this, nanoparticles were characterized by combustion elemental analysis, TEM, FT-IR, dynamic light scattering and UV-Vis/Fluorescence spectroscopy.

### **Assessment of RhB-CeNPs as antioxidant and radical sensor agents.**

#### **2,2-Azino-bis-3-ethylbenzthiazoline-6-sulfonic acid (ABTS) radical cation solution for radical scavenger-antioxidant assays**

ABTS radical cation (ABTS<sup>•+</sup>) was prepared following the methodology described by Seeram, *et al* with minor changes.<sup>42</sup> In short, a 5 mM water solution of ABTS was passed through a Whatman filter containing MnO<sub>2</sub>. The resulting green solution was centrifuged at 5000 rpm for 10 min at 20 °C and the supernatant solution was used immediately for radical scavenger-antioxidant experiments.

#### **Assessment of the antioxidant behaviour of RhB-CeNPs**

We prepared 50 µl of freshly prepared ABTS<sup>•+</sup> solution and 200 µL of several RhB-CeNPs solutions at different concentrations (0, 0.5, 1, 1.5, and 2 mg/mL) in phosphate



buffered saline solution (PBS). These mixtures were poured into a 96-well plate and stirred for 10 min. After this time, absorption and fluorescence spectra ( $\lambda_{\text{exc}}$  350 nm /  $\lambda_{\text{em}}$  400-700 nm) were recorded. This experiment was performed in triplicate using three replicates in each case.

### **Assessment of RhB-CeNPs as antioxidant sensors**

Aliquots of 1  $\mu\text{L}$  of freshly prepared ABTS<sup>•+</sup> solution were added consecutively to a 0.1 mg/mL solution of RhB-CeNPs in PBS, until a total of 15  $\mu\text{L}$ , and, from that point, aliquots of 5  $\mu\text{L}$  were added up until a total of 30  $\mu\text{L}$ . Each mixture was stirred in the dark at room temperature for 30 min before measuring fluorescence, which was done after sample excitation at 350 nm. Fluorescence was recorded between 400-700 nm.

### **Cell Culture**

Experiments were performed with the human hepatoma Hep3B (ATCC HB-8064) and the human cervical carcinoma HeLa (ATCC CCL-2) cell lines. Reagents employed in cell culture were purchased from GIBCO. Hep3B cells were cultured in minimum essential medium (MEM) supplemented with 1 mM nonessential amino acids, whereas HeLa cells were cultured in Dulbecco's modified Eagle's medium (DMEM) with a high glucose concentration (4.5 mg/mL). Both culture media were supplemented with 10 % heat-inactivated fetal bovine serum (FBS) and the antibiotics penicillin 50 U/mL and streptomycin 50  $\mu\text{g}/\text{mL}$ . Cells were maintained in an incubator at 37 °C in a humidified atmosphere of 5% CO<sub>2</sub>/95% air.

## **Confocal laser scanning microscopy images**

HeLa cells were seeded at 10000 cells/well in Nunc 8-well coverslip bottom chamber slides. Treatment with RhB-CeNPs was performed over 3 h, after which cells were incubated for a further 30 min with 5  $\mu$ M Syto80, a green fluorescent marker which preferentially stains nucleic acids, or 2.5  $\mu$ M 10N nonyl-acridine orange (NAO) in order to stain the cytoplasm. Subsequently, the cell culture was washed twice with Hank's Balanced Salt Solution (HBSS) and immediately submitted to image acquisition using confocal microscopy. The excitation wavelengths for the fluorochromes were 488 nm for Syto 80 and 543 nm for RhB-CeNPs, and the emission apertures for fluorescence detection were 510–540 nm for Syto80 and 575–650 nm for RhB-CeNPs. The Z-section series were obtained by progressive increments of  $1.0 \pm 0.2 \mu\text{m}$ .

## **Proliferation and Viability**

Cells were seeded in 48-well plates one day before the experiment at 15000 and 5000 cells/well for Hep3B and HeLa, respectively. Incubation in the presence of RhB-CeNPs (at 20  $\mu\text{g}/\text{mL}$ ) took place for 3h, after which the medium was refreshed and cells were allowed to proliferate exponentially for 3 days. Cell count was performed at 24, 48, and 72 h by means of fluorescence microscopy coupled with static cytometry. For this, 1  $\mu\text{M}$  of the chromatin-staining dye Hoechst 33342 was added for the last 30 min of treatment and the cells were washed with HBSS. All treatments were performed in duplicate, and 25 images per well were recorded with a IX81 Olympus fluorescence microscope coupled with static cytometry software "ScanR" version 2.03.2.

In addition, the 3-[4,5-dimethylthiazol-2-yl]-2,5-diphenyl tetrazolium bromide (MTT) assay was performed. MTT is a colorimetric assay based on the ability of cells to reduce a soluble yellow tetrazolium salt to blue formazan crystals.[9] This reduction takes place only when mitochondrial reductase enzymes are active and is, thus, a marker of cell viability related to mitochondrial function. For these experiments, cells were seeded in 96-well plates at 20000 and 50000 cells/well for Hep3B and HeLa, respectively. Incubation with RhB-CeNPs (at 20  $\mu\text{g}/\text{mL}$ ) was performed over 3 h, after which the medium was refreshed and cells were further cultured for a total of a 24 h. MTT reagent (Roche Diagnostics, Mannheim, Germany) was added (20  $\mu\text{L}/\text{well}$ ) for the last 4 h of the treatment. Cells were solubilized with dimethyl sulfoxide (DMSO) (100  $\mu\text{L}/\text{well}$ , 5 min, 37  $^{\circ}\text{C}$ ), and absorbance was detected using a Thermo Labsystems Multiscan plate-reader spectrophotometer. Results were obtained by subtracting the intensity of the signal at 690 nm from the 570 nm absorbance, as indicated in the instructions provided with the kit.

### **Apoptosis**

Induction of apoptosis was studied by Bivariate Annexin V/PI analysis for which HeLa and Hep3B cells were seeded in duplicate in 48-well plates at 20000 and 30000 cells/well respectively. Staurosporine (STS), a widely used protein kinase inhibitor, was employed as a positive proapoptotic control using 0.1  $\mu\text{M}$  and 1  $\mu\text{M}$  for HeLa and Hep3B, respectively.[9] Incubation with RhB-CeNPs (at 20  $\mu\text{g}/\text{mL}$ ) was performed over 3 h, after which the medium was refreshed and cells were cultured for a further period of 24 h. The medium was then replaced with HBSS containing 1  $\mu\text{M}$  of Hoechst 33342 to stain nuclei and 0.9  $\mu\text{L}$  Annexin V-fluorescein in order to detect

phosphatidylserine exteriorization, and cells were incubated in the dark for 20 min at 37 °C. Thereafter, 0.3 µL of the chromatin detecting dye propidium iodide (PI) was added (5 min) to label dead or damaged cells. Annexin V and PI solutions were purchased in the form of the Annexin-V-FLUOS staining kit. Analysis was performed of 25 images/well recorded with a IX81 Olympus fluorescence microscope coupled with static cytometry software “ScanR” version 2.03.2.

### **ROS production and assessment of RhB-CeNPs as antioxidant agents and sensors**

To assess ROS content, the day before the experiment HeLa cells were seeded in 96-well plates at 20,000 cells/well in Dulbecco’s Modified Eagle’s Medium (DMEM) containing 10 % heat inactivated fetal bovine serum, 2 mM L-glutamine, 1mM sodium pyruvate, 50 µg/ml penicillin and 50 µg/ml streptomycin.

Culture medium was removed from cells and fresh medium with or without 20 µg/ml RhB-CeNPs and CeNPs were added. Cells were incubated for 3h at 37 °C under gentle agitation. The medium was then replaced with preheated PBS containing 2.5 µM 2,7-dichlorodihydrofluorescein diacetate (DCFH-DA). After incubation for 30 min at 37 °C in the dark under gentle agitation, various concentrations of rotenone, a widely used pharmacological inductor of ROS generation were added and fluorescence was measured at 5 min-intervals over a 30 min period in a Synergy Mx plate reader (BioTek Instruments). DCFH-DA fluorescence intensity was recorded after 30 min of rotenone treatment at 535 nm and after exciting the sample at 485 nm. Fluorescence intensity in each rotenone condition was calculated by subtracting the fluorescence emitted by untreated cells.

To assess the ability of RhB-CeNPs as ROS biosensors we performed the same experiment as described previously, but without the addition of DCFH-DA. Fluorescence intensity of RhB was also measured at 5 min-intervals during a 30 min period after exciting the sample at 543 nm and recording the emission at 575, 600, 625, and 650 nm. The RhB fluorescence values shown are raw data after 30 min of rotenone treatment.

## **CONCLUSION**

In the present study, ceria nanoparticles with rhodamine B groups covalently attached to their surface have been successfully prepared and characterized. These RhB-CeNPs exhibit antioxidant activity and sensor capacity against oxidant species both in solution and human cells submitted to oxidative stress. In addition, we have confirmed that RhB-CeNPs can cross the cellular membrane and are remarkably biocompatible without increasing cell mortality. In these cells, RhB-CeNPs present a higher antioxidant capacity than the well-established CeNPs when there are harmful levels of ROS content, and exhibit a similar sensor capacity to the DCFH-DA probe. Thus, we present RhB-CeNPs as a new class of nanomaterial with theranostic properties for oxidative stress.

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## NOTES AND REFERENCES

- 1 C. Espinet, H. Gonzalo, C. Fleitas, M. J. Menal and J. Egea, *Curr. Drug Targets*, 2015, **16**, 20.
- 2 M. Matsuo, *J. Clin. Bioche. Nutr.*, 2004, **35**, 63.
- 3 R. S. Sohal and R. Weindruch, *Science*, 1996, **273**, 59.
- 4 G. Vitale, S. Salvioli and C. Franceschi, *Nat. Rev. Endocrinol.*, 2013, **9**, 228.
- 5 M. Rocha, N. Apostolova, J. R. Herance, S. Rovira-Llopis, A. Hernandez-Mijares and V. M. Victor, *Med. Res. Rev.*, 2014, **34**, 160.
- 6 J. M. Gutteridge and J. Mitchell, *Br. Med. Bull.*, 1999, **55**, 49.
- 7 C. Gorrini, I. S. Harris and T. W. Mak, *Nat. Rev. Drug Discovery*, 2013, **12**, 931.
- 8 C. Gutierrez-Merino, C. Lopez-Sanchez, R. Lagoa, A. K. Samhan-Arias, C. Bueno and V. Garcia-Martinez, *Cur.r Med. Chem.*, 2011, **18**, 1195.
- 9 R. Martín, C. Menchón, N. Apostolova, V. M. Victor, M. Alvaro, J. R. Herance and H. García, *ACS Nano*, 2010, **4**, 6957.
- 10 L. Raj, T. Ide, A. U. Gurkar, M. Foley, M. Schenone, X. Li, N. J. Tolliday, T. R. Golub, S. A. Carr, A. F. Shamji, A. M. Stern, A. Mandinova, S. L. Schreiber and S. W. Lee, *Nature*, 2011, **475**, 231.
- 11 L. Rochette, M. Zeller, Y. Cottin and C. Vergely, *Biochim. Biophys. Acta*, 2014, **1840**, 2709.
- 12 B. Y. Kim, J. T. Rutka and W. C. Chan, *N. Engl. J. Med.*, 2010, **363**, 2434.
- 13 S. E. Lohse and C. J. Murphy, *J. Am. Chem. Soc.*, 2012, **134**, 15607.
- 14 A. H. Lu, E. L. Salabas and F. Schueth, *Angew. Chem. Int. Ed.*, 2007, **46**, 1222.
- 15 C. Menchón, R. Martín, N. Apostolova, V. M. Victor, M. Alvaro, J. R. Herance and H. García, *Small*, 2012, **8**, 1895.
- 16 T. K. Sau, A. L. Rogach, F. Jaeckel, T. A. Klar and J. Feldmann, *Adv. Mater. (Weinheim, Ger.)*, 2010, **22**, 1805.
- 17 V. Valtchev and L. Tosheva, *Chem. Rev.*, 2013, **113**, 6734.
- 18 J. Della Rocca, D. Liu and W. Lin, *Acc. Chem. Res.*, 2011, **44**, 957.
- 19 D. E. Lee, H. Koo, I. C. Sun, J. H. Ryu, K. Kim and I. C. Kwon, *Chem. Soc. Rev.*, 2012, **41**, 2656.
- 20 J. Liu, X. Zheng, L. Yan, L. Zhou, G. Tian, W. Yin, L. Wang, Y. Liu, Z. Hu, Z. Gu, C. Chen and Y. Zhao, *ACS Nano*, 2015, **9**, 696.
- 21 K. Riehemann, S. W. Schneider, T. A. Luger, B. Godin, M. Ferrari and H. Fuchs, *Angew. Chem. Int. Ed.*, 2009, **48**, 872.
- 22 V. Wagner, A. Dullaart, A. K. Bock and A. Zweck, *Nat. Biotechnol.*, 2006, **24**, 1211.
- 23 F. Esch, S. Fabris, L. Zhou, T. Montini, C. Africh, P. Fornasiero, G. Comelli and R. Rosei, *Science*, 2005, **309**, 752.

- 24 S. Turner, S. Lazar, B. Freitag, R. Egoavil, J. Verbeeck, S. Put, Y. Strauven and G. Van Tendeloo, *Nanoscale*, 2011, **3**, 3385.
- 25 A. Maldotti, A. Molinari, R. Juárez and H. Garcia, *Chem. Sci.*, 2011, **2**, 1831.
- 26 A. Y. Estevez, S. Pritchard, K. Harper, J. W. Aston, A. Lynch, J. J. Lucky, J. S. Ludington, P. Chatani, W. P. Mosenthal, J. C. Leiter, S. Andreescu and J. S. Erlichman, *Free Radic. Biol. Med.*, 2011, **51**, 1155.
- 27 A. Gojova, J. T. Lee, H. S. Jung, B. Guo, A. I. Barakat and I. M. Kennedy, *Inhal. Toxicol.*, 2009, **21**, 123.
- 28 C. Korsvik, S. Patil, S. Seal and W. T. Self, *Chem. Commun. (Camb)*, 2007, **10**, 1056.
- 29 J. Niu, A. Azfer, L. M. Rogers, X. Wang and P. E. Kolattukudy, *Cardiovasc. Res.*, 2007, **73**, 549.
- 30 R. W. Tarnuzzer, J. Colon, S. Patil and S. Seal, *Nano Lett.* 2005, **5**, 2573.
- 31 S. S. Banerjee and D. H. Chen, *Nanotechnology*, 2009, **20**, 185103.
- 32 M. Das, D. Mishra, P. Dhak, S. Gupta, T. K. Maiti, A. Basak and P. Pramanik, *Small*, 2009, **5**, 2883.
- 33 D. Shi, M. Ni, J. Luo, M. Akashi, X. Liu and M. Chen, *Analyst*, 2015, **140**, 1306.
- 34 E. Vlashi, L. Kelderhouse, J. E. Sturgis and P. S. Low, *ACS Nano*, 2013, **7**, 8573.
- 35 L. Yuan, W. Lin, K. Zheng, L. He and W. Huang, *Chem. Soc. Rev.*, 2013, **42**, 622.
- 36 A. Mehrdad and R. Hashemzadeh, *Ultrason. Sonochem.*, 2010, **17**, 168.
- 37 P. Qu, J. Zhao, T. Shen and H. Hidaka, *J. Mol. Catal. A: Chem.*, 1998, **129**, 257.
- 38 X. Zhou, J. Lan, G. Liu, K. Deng, Y. Yang, G. Nie, J. Yu and L. Zhi, *Angew. Chem. Int. Ed.*, 2012, **51**, 178.
- 39 J. H. Kwak, Y. He, B. Yoon, S. Koo, Z. Yang, E. J. Kang, B. H. Lee, S. Y. Han, Y. C. Yoo, K. B. Lee and J. S. Kim, *Chem. Commun. (Camb)*, 2014, **50**, 13045.
- 40 A. Reisch, P. Didier, L. Richert, S. Oncul, Y. Arntz, Y. Mély and A. S. Klymchenko, *Nat. Commun.*, 2014, **5**, 4089.
- 41 P. Reungpatthanaphong, S. Dechsupa, J. Meesungnoen, C. Loetchutinat and S. Mankhetkorn, *J. Biochem. Biophys. Methods*, 2003, **57**, 1.
- 42 N. P. Seeram, S. M. Henning, Y. Niu, R. Lee, H. S. Scheuller and D. Heber, *J. Agric. Food Chem.*, 2006, **54**, 1599.