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1 Assessment of grape cluster yield components based on 3D
2 descriptors using stereo vision

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25 **Abstract**

26 Wine quality depends mostly on the features of the grapes it is made from. Cluster and berry
27 morphology are key factors in determining grape and wine quality. However, current practices
28 for grapevine quality estimation require time-consuming destructive analysis or largely
29 subjective judgment by experts.

30 The purpose of this paper is to propose a three-dimensional computer vision approach to
31 assessing grape yield components based on new 3D descriptors. To achieve this, firstly a partial
32 three-dimensional model of the grapevine cluster is extracted using stereo vision. After that a
33 number of grapevine quality components are predicted using SVM models based on new 3D
34 descriptors. Experiments confirm that this approach is capable of predicting the main cluster
35 yield components, which are related to quality, such as cluster compactness and berry size
36 ($R^2 > 0.80$, $p < 0.05$). In addition, other yield components: cluster volume, total berry weight
37 and number of berries, were also estimated using SVM models, obtaining prediction R^2 of 0.82,
38 0.83 and 0.71, respectively.

39

40 **Keywords:** grape quality; cluster yield components; *Vitis vinifera* L; non-invasive
41 technologies; stereo-vision; 3D descriptors;

42

43 1. Introduction

44 Due to the economic importance of the wine industry worldwide, innovative methods and
45 technologies are being developed and applied to increase the quality of wine. The quality of
46 wine is partially subjective, as it depends on the consumer's taste and preferences. However, it
47 also depends on objective parameters. Of all the factors that influence the quality of the wine,
48 the most important is the quality and features of the grapes it is made from.

49 Cluster compactness and berry size are two key factors of grapevine fruit quality. Current
50 practices to assess these quality parameters require time-consuming destructive analysis or
51 largely subjective judgments by experts. Unfortunately, in addition to the lack of objectivity of
52 these parameters, the short and limited time available for analysis during harvest time and the
53 lack of measurement tools as well as their high cost, among other factors, make it difficult to
54 assess grape quality. For this reason, developing non-destructive grape supervision analysis,
55 which increases the objectivity (Tello & Ibáñez, 2014) or automates the estimation (Roscher et
56 al., 2014), represents a huge technological advance compared to the current practices.

57 Nowadays, 2D vision systems are widely used in the agri-food industry/agribusiness with
58 proven results (Benlloch, Agustí, Sanchez, & Rodas, 1995; Brosnan & Sun, 2004; Diago, Sanz-
59 Garcia, Millan, Blasco, & Tardaguila, 2014...). Moreover, huge advances in 3D sensor
60 technology are leading to new opportunities and challenges. Specifically, one of the sectors
61 which can clearly benefit from 3D computer vision technology is viticulture; for example, to
62 forecast the quality of wine grapes (Whalley & Shanmuganathan, 2013).

63

64 1.1 Three-dimensional computer vision methods

65 There are many methods for extracting 3D information from 2D images. These methods can be
66 classified into active and passive methods. Active methods emit light patterns into the scene to
67 analyze their behavior, while passive methods only analyze the behavior of ambient light.

68

69 Some active methods are based on structured light (Udomkun, Nagle, Mahayothee, & Müller,
70 2014; Verdú et al., 2013). The simplest form of this is the projection of light stripes onto the
71 surface, enabling the 3D position of the surface to be calculated from its intersection with the
72 stripes.

73

74 Passive methods, such as stereo photogrammetry (Cyganek & Siebert, 2011), consist of two
75 camera views of the surface from slightly different locations. Corresponding features are
76 matched between the two images and the 3D surface is then constructed by triangulation. An
77 advantage of photogrammetry over structured light is that the natural appearance of the
78 surface is captured as a normal part of the process. A disadvantage is that it relies on there
79 being enough features in the surface texture (visual appearance) for matching to take place.

80

81 Although stereoscopy is very simple and has a low cost when compared to other 3D
82 techniques, not many studies using this technique for food inspection can be found in the
83 available literature. This is mainly because automatic matching of points from stereo pairs is a
84 difficult task for computer-based image analysis. However, there are some interesting works
85 using stereoscopy for detecting weeds (Sachez & Marchant, 2000), measuring the thickness of
86 wheat grains (Sun, Berman, Coward, & Osborne, 2007), estimating firmness in salmon

87 (Quevedo & Aguilera, 2010) and measuring the volume of apple slices in a drying process
88 (Sampson, Chang, Rupasinghe, & Zaman, 2014). In all these works, edge features are used to
89 solve the correspondence problem.

90
91

92 1.2 Grapevine quality components

93 Cluster architecture and berry morphology and distribution are key factors in determining
94 grape quality. On the one hand, compact clusters show favorable conditions for the
95 development of different grape pests and diseases (especially *Botrytis cinerea*). On the other
96 hand, the number of interior berries increases with cluster compactness. These interior berries
97 may not receive the sunlight needed to achieve an adequate phenolic maturity, leading to a
98 heterogeneous ripeness of the cluster. At present, cluster compactness is visually estimated by
99 experts using OIV descriptor No. 204 (OIV, 2007). This descriptor categorizes a cluster into one
100 of five groups, quantified by 1, 3, 5, 7 and 9, where number 1 indicates “berries in grouped
101 formation with many visible pedicels” and number 9 indicates “misshapen berries”. This
102 estimation is subjective, but objective compactness estimations are required to allow
103 comparisons between different works. Recently (Tello & Ibáñez, 2014) have evaluated some
104 destructive methods for an objective estimation of grape cluster compactness.

105

106 Berry size is related to the skin-to-pulp ratio of the berry and the concentration of skin-located
107 compounds that play a key role in wine quality. Two pieces of research have been published
108 recently regarding the estimation of grape berry size. (Cubero et al., 2014) presents a method
109 to estimate the size and weight of isolated wine-grapes taking into account the peduncle.
110 (Roscher et al., 2014) detects circular structures on in-field images which are potentially
111 berries and classifies them into the ‘berry’ or ‘non-berry’ class by utilizing a conditional random
112 field. These two works estimate berry sizes from one image using a calibration object with a
113 known size, which is located at a certain distance from the berries in the scene. Instead of this,
114 it will be interesting to use a stereoscopic system to estimate berry size (without the constraint
115 of always having a known calibration object in the scene).

116

117 This work contributes to accomplishing an objective and non-destructive estimation of the
118 main grape yield components. To achieve this, a new approach has been developed to
119 generate a partial three-dimensional model of the grape cluster using 3D computer vision
120 technology. The approach is intended to be used alongside an inspection system consisting of
121 two cameras arranged in a stereoscopic fashion. The system outputs two images of the grapes,
122 one from each camera. These images, along with the camera calibrations, are then used to
123 provide a partial 3D reconstruction of a grape cluster. Once this 3D model is obtained, 3D
124 descriptors are automatically extracted to estimate the main cluster quality components and
125 other yield components.

126

127 The experiments, obtained plant material and images are presented in Sections 2.1 and 2.2. In
128 Section 2.3 the proposed 3D reconstruction approach is presented. Section 2.4 explains the 3D
129 descriptors and the models proposed to estimate quality and yield components. The
130 experiments and the results obtained are shown and discussed in Section 3. Finally, the
131 conclusions of the paper are presented in Section 4.

132

133 2. Material and methods

134 2.1. Description of grapevine cluster samples

135 The study was carried out with 100 grapevine clusters from ten different red grapevine (*Vitis*
136 *vinifera* L.) varieties: Grenache, Pinot Noir, Graciano, Monastrell, Mencia, Bobal, Cabernet
137 Sauvignon, Tempranillo, Merlot and Carignan. All clusters were collected at Vitis Navarra
138 Nursery vineyards (Navarra, Spain), just prior harvest in 2011.

139 At a first stage, cluster compactness was rated according to OIV descriptor No. 204 (OIV, 2007)
140 by a panel formed by 10 experienced judges. Secondly, yield components, including the
141 number of berries per cluster, cluster volume, and cluster and berry weight, were destructively
142 measured at the University of La Rioja (Spain). The weight of each cluster was determined
143 using a scale (Blauscal, AC-5000), and the morphological volumes were measured through the
144 volume of water displaced by immersion in a bucket filled with water. Once clusters were
145 destemmed by hand, the number of berries was counted. In addition, 15 berries per cluster
146 were randomly chosen to measure their length and width using digital calipers (Mitutoyo, CD-
147 15DCX). For the latter two features, the average of the 15 measures was used.

148 2.2. Image acquisition system

149 Images were taken using a Bumblebee2 stereo camera (Point Grey Research Inc, Richmond,
150 BC, Canada) model BB2-08S2C-25 configured in automatic mode (white balance, gain and
151 shutter). Images were synchronized and had a resolution of 1024 x 768 RGB color pixels from 0
152 to 255 values per channel.

153 Image acquisition was performed in lab conditions but the image acquisition set-up was
154 designed to be near field conditions. The sample grape cluster was fixed from its peduncle, as
155 if hanging from the vine (Fig. 1). The illuminants employed were four pairs of fluorescent tubes
156 (Osram L 18W/965 BIOLUX) with a color temperature of 6500 K. They were distributed and
157 oriented at different angles, which produced highlights on the sample (Fig. 2). The distance
158 between illuminant and samples was around 0.35 m. It was decided that the light intensity
159 should not be changed because it would have a lower effect than the color changes between
160 samples. The camera can be placed at a distance of 0.10 m to 0.25 m from the sample. In this
161 set-up, the furthest distance was chosen to work with the lowest resolution of the sample
162 projection (around 170x250 pixels). As a result, this set-up resulted in a difficult image
163 acquisition scenario.

164 A controlled background was needed to calculate one of the proposed 3D descriptors for
165 assessing compactness.

166

167

168 Image acquisition was performed using the program PGR FlyCapture v.1.22 (Point Grey
169 Research Inc, Richmond, BC, Canada).

170 2.3. Stereoscopic 3D reconstruction of grape clusters

171 The proposed approach is intended to be used alongside an inspection system consisting of
172 two cameras arranged in a parallel stereoscopic fashion. The system outputs two images of the
173 grape cluster, one from each camera. These images, along with the known camera calibrations,
174 are then used to automatically obtain a partial 3D reconstruction of a grape cluster, which can
175 be refined manually by a user.

176

177 Unfortunately, this approach has some inherent difficulties. Firstly, some of the grapes in the
178 internal layers of the cluster are totally or partially hidden behind other berries. This fact
179 impedes the reconstruction of the berries that are completely hidden. However the 3D
180 descriptors can still be extracted from the reconstructed grape berries. This also means that
181 the grape berries that are partially hidden will be harder to generate due to missing
182 information. In addition, for it to be feasible to solve the problem, grape berries were modeled
183 mathematically as perfect spheres. In fact, though, they may actually be slightly deformed,
184 more akin to an ovoid, which may be somewhat troublesome in certain extreme cases.

185

186 In order to overcome these difficulties, several computer vision techniques were used. These
187 included: i) the 3D implementation of the Hough transform (Woodford, Pham, Maki, Perbet, &
188 Stenger, 2014), ii) feature detection and iii) feature matching. Also, the approach presented
189 here involved extensive use of direct and inverse perspective projection at various stages of
190 the automatic process.

191 A new 3D reconstruction tool has been developed in C++ using Microsoft Visual Studio 2010
192 Professional and the Qt 4.8.0 UI framework. Image processing was performed using the
193 openCV library v.2.3.1. This 3D reconstruction tool provides a clearly defined workflow,
194 including an automatic approach and a manual refinement part. In the automatic phase, the
195 tool generates as many correct grapes as possible without user supervision. The effectiveness
196 of this method varies greatly on a case by case basis, as some grape varieties may be easier to
197 reconstruct than others, depending primarily on how prominent the difficulties discussed
198 above are.

199 2.3.1. Automated approach

200 This approach automatically generates a partial three-dimensional model of the grape berries
201 without the user's help. In order to do so, the following problems were solved.

202

203 Firstly, the original images were rectified to accomplish a parallel camera configuration, where
204 the baseline ($b = 0.1197$ m) was aligned with the Xs-axis and therefore the epipolar constraints
205 could be easily applied to solve the correspondence problem. In this case, only vertical edge
206 features were used, since the horizontal edges belong to the same epipolar line and this
207 increases the risk of obtaining wrong correspondences.

208 Edges are features which fit perfectly in scenarios with a high degree of variability in color or
209 illumination. In this case, vertical edges were extracted from the rectified images and for each

210 vertical edge in the rectified images a vector of descriptors was estimated. This vector consists
211 of the gradient module (m_i), the sines (s_i) and the cosines (c_i) of the gradient orientation. The
212 range of distances allowed between the camera and the sample, described at image
213 acquisition set-up as shown in Fig. 1, achieved accuracy with reconstruction errors smaller
214 than 1 mm at occlusion edges. Therefore, the occlusion edges of the berries can be used to
215 solve the reconstruction problem obtaining low errors.

216 The measure of similarity between features in the left and right images was calculated by the
217 Euclidean distance between their normalized vector of descriptors [m_i , s_i , c_i]. Feature matching
218 consists of an optimization problem where features in a row in the left image must be
219 matched with features in the same row in the right image. This correspondence problem was
220 solved using dynamic programming, where a tree of possible solutions is explored. A node of
221 this tree contains two column indexes (one for each image), a list of matching pairs and the
222 correspondence cost. The two indexes pointed to the features that the algorithm was trying to
223 correspond in this node (Fig. 3).

224 Each node generated three new branches. The first branch took into account the case when
225 the feature in the left image was occluded and therefore did not correspond to any feature in
226 the right image. In this case, the son node incremented its left index and added a constant to
227 its correspondence cost. The second branch considered the case when the feature in the left
228 image matched with the feature in the right image. In this case the node incremented its two
229 indexes, included the new matching pair in the list and added the similarity distance to its
230 correspondence cost. And finally, the third branch assumed that the feature in the right image
231 was occluded and therefore did not correspond to any feature in the left image. In this last
232 case, the node incremented its right index and added a constant to its correspondence cost.

233 The algorithm expanded the tree and found the optimal solution, which was the list of
234 matching pairs on the leaf node with the minimum cost.

235 For each matching pair of vertical edges, a 3D point was obtained by triangulation. In
236 addition, others features (not only vertical edges) could be reconstructed. For example,
237 points between berries, which have a dark appearance in the two images. The 3D points,
238 reconstructed using vertical edges, were the input to a Hough transform which had been
239 adapted to detect spheres in the four dimensional parameter space (x_c , y_c , z_c , r). After the
240 accumulation phase, a minimum number of votes were set for detecting spheres. A
241 refinement step was performed in order to detect and remove some wrong spheres, such
242 as overlapping spheres or spheres which include reconstructed points between berries. It
243 is important to mention that Hough transform is a robust technique against possible noise
244 like highlights.

245 2.3.2. Manual refinement

246 In the manual phase, the user could reconstruct additional berries using the 3D reconstruction
247 tool. This could be achieved by two methods. The first required the user to input the
248 coordinates (position and radius) of the grape. While this method certainly worked, it may be
249 somewhat unintuitive and slow. For this reason, an alternative, more visual method was

250 designed and developed. In this case, in order to reconstruct a grape the user just needed to
251 specify 5 points of its border in one of the original images and its center in the other one. With
252 these data, the tool generated a 3D grape berry automatically, without the need for any
253 further action. This enabled the user to reconstruct a grape simply by entering a total of 6
254 points in a totally visual, intuitive and quick way. Furthermore, the tool assisted this process by
255 providing the user with a visual aid in the form of previsualizations of the final result in both of
256 the original images and axis restrictions whenever possible. With this method the user would
257 not need to be an expert to use the tool, as the process is simple, easy and straightforward.

258 One of the objectives of this work was to make the 3D reconstruction tool as intuitive as
259 possible. Therefore, the tool has a graphical interface (GUI) that enables the user to visualize
260 the model and interact with it. The GUI that can be seen in Fig. 4 and it is divided into two
261 main parts: the 2D viewports and the 3D viewport.

262 On the one hand, there is a 2D viewport, where all the information related to the 2D space,
263 such as the original images of the grape, is shown. As mentioned above, the reconstruction
264 process takes two images of the grape as input; therefore, the tool has two 2D viewports, one
265 for each camera. These viewports can also be used by the user to enter input to the tool, such
266 as the 6 points required in the manual reconstruction phase.

267 A 2D viewport also has some features that are not directly related to the reconstruction
268 process but that can greatly help the user. The most important of these features is the
269 projection of the model onto the original images. The result is a new image which consists of
270 the original image with the projection of the current model drawn over it. With this new image
271 the user can easily check if the model correctly fits the grapes in the original images.

272 On the other hand, the 3D viewport provides a fully three-dimensional visualization of the
273 current model. The user can select individual grapes and edit their parameters, as well as
274 navigate through the 3D space. The 3D viewport is immediately updated with new information
275 to reflect the current state of the model. This means that, whenever the user adds a new grape
276 using the system explained above, the 3D viewport will show the changes on the fly. The same
277 applies to the automatic phase: the moment it finishes its execution, the 3D viewport will be
278 updated.

279 Finally, the reconstructed model can be saved in an open, easily-readable format, meaning
280 that the models can be easily used by other applications that want to display or perform
281 certain operations on them.

282

283 2.4. 3D descriptors and models

284 There are many morphological descriptors for the characterization of the grapevine cluster
285 which can be measured manually. However, with the cluster 3D model, non-destructive
286 descriptors can be calculated automatically (see Table 1). In this work, six 3D descriptors were
287 extracted automatically from the 3D model: cluster volume, berry size, number of berries per
288 cluster, concavity measure, intersection between berries, and number of berries per area. The

289 last 3D descriptors are new descriptors proposed in this work to assess grapevine cluster
290 compactness that cannot be measured by hand.

291 In order to calculate the concavity measurement (holes between berries) an image
292 segmentation should first be performed. This segmentation was performed in two steps:
293 firstly, the definition of a region of interest (ROI) and secondly, classification of pixels into
294 three classes: background, holes and berries.

295 The first step, ROI definition, calculates the difference between the convex hull of the cluster
296 and the berries from the 3D model. This difference estimates the potential concavities. These
297 estimated potential concavities were extracted from the 2D images based on the projection of
298 the 3D model. The convex hull of the cluster was calculated for the image based on a 2D
299 Delaunay triangulation. This triangulation was calculated from the projected sphere centers of
300 the 3D model. Then, using the triangles obtained, a convex hull was created. However, some
301 triangles from this convex hull were removed to ensure a better fit to the projected shape of
302 the cluster, specifically focusing on removing large concavities. This refinement consisted of
303 removing the triangles that had at least one vertex belonging to the convex hull and in which
304 the longest edges are three times longer than their heights. Pixels inside this refined convex
305 hull were used as the first region. The second part of this step was to make a second region as
306 the union of the ellipses from the projection of the 3D model. Finally, the ROI was defined as
307 the difference between the first region and the second region.

308 Fig. 5 shows an example of this step on the Bobal sample 1. The projected berries can be seen
309 in green, the Delaunay triangulation in blue, the convex hull in red and the removed triangles
310 in purple. In Fig. 6, pixels inside the ROI were colored in red.

311 The second step was the color segmentation of the pixels in the ROI into 3 classes. This pixel
312 classification was based on the k -nearest-neighbors pattern recognition approach (Sánchez,
313 Albarracín, Grau, Ricolfe, & Barat, 2008). Five different grape colors were self-trained for each
314 pair of images using a k -means clustering of the pixel colors inside the projected ellipses of the
315 3D model. Five colors were also trained for the background determination. Then, for each pixel
316 inside the ROI a difference to these trained colors was calculated. If the lowest difference in
317 absolute values was with a background color, then it was classed as background. If the lowest
318 difference was with a self-trained grape color, then if the difference was positive (pixel color
319 was brighter than self-trained grape color) it was classified as grape color, otherwise it was
320 classified as a hole with the difference as an approximate measure of the depth.

321 The depth value for all the hole pixels was accumulated plus the accumulated value of all the
322 background pixels inside the ROI, taking their depth as 255. The mean of both images was
323 calculated for each sample. This value was called the concavity measure and it is the only 3D
324 descriptor that requires a controlled background.

325 The intersection between spheres was approximated using the Eq. 1 where c is the center of
326 the sphere, r its radius and n the total number of grapes in the 3D model.

$$d = \begin{cases} \left| \|c_1 - c_2\| - r_1 - r_2 \right|, & \text{if } \|c_1 - c_2\| - r_1 - r_2 < 0 \\ 0, & \text{if } \|c_1 - c_2\| - r_1 - r_2 \geq 0 \end{cases}$$

$$I = d * d * \min(r_1, r_2) \quad (1)$$

$$Im = \frac{1}{n} * \sum_{i=1}^n I_i$$

327

328 The number of berries per area was approximated as the area of the ROI divided by the
329 number of berries. This value was averaged using the stereo pair of images.

330 The cluster volume was calculated using the information from the 3D model following this
331 equation (Eq. 2).

$$V = \sum_{i=1}^n \frac{4 * \pi * r_i^3}{3} \quad (2)$$

332

333 Another value extracted from the 3D model was the berry size as the mean volume of the
334 grape berries in the 3D model (Eq. 3).

$$V_m = \frac{1}{n} * V \quad (3)$$

335 The number of berries was the number of reconstructed berries in the 3D model for each
336 sample.

337 The 3D descriptors calculations were performed using our own code developed using the
338 image processing toolbox of Matlab R2008a (The Mathworks, Natick, Massachusetts, USA) and
339 loading the previously reconstructed 3D models.

340 2.5. Statistical analysis

341 *Kendall's Tau-b* (Kendall, 1970) correlation coefficients were calculated between the 3D
342 descriptors and the compactness measured by the visual evaluation panel.

343 Support Vector Machine regression models (using nu-support vector regression) (Schölkopf,
344 Smola, Williamson, & Bartlett, 2000) were used to predict compactness and some
345 morphological components (berry size, cluster volume, cluster weight and number of berries)
346 using the calculated 3D descriptors as input data. The model is based on a number of support
347 vectors (samples selected from the calibration set) restricted by the nu parameter and non-
348 linear model coefficients which define the non-linear mapping of variables (calculated 3D
349 descriptors).

350 The samples were pseudo randomly divided 2/3 for building the model and 1/3 for testing it. It
351 was adjusted so that all the different grape species were represented for building and testing.

352 To estimate the fit of the calibration data with the models developed, the Root-Mean-Square
353 Error of Calibration (RMSEC) was used. It is defined as:

$$RMSEC = \sqrt{\frac{\sum_{i=1}^n (\hat{y}_i - y_i)^2}{n}} \quad (4)$$

354 Where \hat{y}_i are the values of the predicted variable when all samples are included in the model
355 formation, y_i are the known values and n is the total number of samples.

356 A random cross-validation method was employed to evaluate the models developed for the
357 samples used in the calibration model. In this method, subsets of 8 ($n/8$) random samples are
358 used to test the model developed without them. This method was iterated three times with
359 different samples and its results averaged to achieve more reliable validations.

360 The Root-Mean-Square Error of Cross-Validation (RMSECV) was used to evaluate and compare
361 the accuracy of the different SVM models developed using the random cross-validation
362 method described previously. RMSECV is based on Eq. 4, but in this case the parameter \hat{y}_i was
363 the value of the variable estimated using a model that was built without the removed group of
364 samples ($i \in RemovedSamples$).

365 The prediction accuracy of the model was estimated using the root-mean-square error of
366 prediction (RMSEP). RMSEP is calculated exactly as in Eq.4, except that the estimates \hat{y}_i refer
367 to 33 new samples which were not involved in model building.

368 All statistical procedures were performed on PLS Toolbox 6.5 (Eigenvector Research Inc.,
369 Wenatchee, Washington, USA), a toolbox extension within the Matlab 7.6 computational
370 environment (The Mathworks, Natick, Massachusetts, USA).

371

372 3. Results and discussion

373 The efficiency of the 3D reconstruction tool was tested by generating the 3D model of 100
374 clusters of grapes of different varieties. All models were successfully generated using a
375 combination of the automatic and manual phase (Fig. 7). The process of obtaining an almost
376 exact reconstruction of the visible part of the cluster never took more than 10 minutes, often
377 less depending on the complexity of the cluster.

378 Applying only the automatic reconstruction phase, the mean success rate was 20% correctly
379 reconstructed berries per view (approximately 10 berries) (Fig. 8). The lowest rate was 10%
380 when the automatic phase was applied to “Tempranillo” clusters. The rest of the varieties
381 provided better reconstruction ratios, the best case being “Bobal”, with 27.78%.

382 Using the aforementioned features of the 2D viewport the model developed was checked
383 against the original images of the cluster, obtaining successful results with projected errors
384 lower than 5 pixels (Fig. 9). Taking into account the acquisition set-up defined in Fig. 1, it was

385 proven that if the error of projected edges is lower than 5 pixels, the reconstruction error will
386 be lower than 1 mm. The only visible grapes that could not be generated were the ones that:
387 a) were visible only in one image and hidden in the other, thus making it impossible to
388 generate a 3D version of it; or b) could not be properly identified because of visual noise. All
389 the other visible grapes could be reconstructed with minimal error.

390 At this point, it is important to highlight the advantages of having a digitalized, 3D model of the
391 grapes. The most important one is the fact that the desired 3D descriptors can be extracted
392 automatically, of course. However, it is important to note that it also allowed the model to be
393 saved easily, virtually forever.

394 The benefits of this implication are significant because it is possible to create a record of
395 models of different wine varieties over the years. Moreover, it is also possible to recover a
396 previously analyzed model and extract new descriptors for it that were not needed before.

397 To correctly assess these benefits one can compare it to the traditional system where only
398 measures or images can be stored. This means that in order to analyze the grape clusters with
399 new criteria it is necessary to perform the measurement again. Apart from the time needed to
400 do this, those grape clusters may not be available any more.

401

402 3.1. Cluster quality components

403 A Kendall tau study was performed to correlate with compactness for the new 3D descriptors
404 concavity measure, intersection between berries and number of berries per area. It was also
405 calculated for the index CI-12, which (Tello & Ibáñez, 2014) claims can be used for
406 compactness measures. As it can be seen in Table 2, the CI-12 result confirmed a good
407 correlation, as reported by the authors. The concavity measure descriptor presented the
408 highest Pearson correlation coefficient, whereas the number of berries per area was poorly
409 correlated.

410 Fig. 10 and Fig. 11 depict the image segmentation results of two samples of Cabernet Franc
411 with two different classes of compactness 3, a loose cluster, and 7, corresponding to a tight
412 cluster. Hole depths inside the cluster were coded as blue being deeper the higher the value.
413 As it can be seen, there were significantly more background and hole pixels in Fig. 10 than in
414 Fig. 11. This information was collected by the previously defined concavity measurement.
415 Therefore, it seems logical that there was a high correlation value with the compactness
416 obtained with the Kendall tau study.

417 The results of the calibrated SVM model for predicting compactness using these three 3D
418 descriptors: concavity measure, intersection between berries and number of berries per area,
419 can be seen in Table 3. The inclusion of the 3D descriptors intersection between berries and
420 number of berries per area in the SVM model for grape cluster compactness enhanced the
421 results by around 10% in comparison with using the concavity measure alone. Although the
422 Kendal tau results in a poor direct correlation for the 3D descriptor of number of berries per

423 area (Table 2), it helped to model some extreme cases, which explains the better results when
424 it is used.

425 It should be noted that only one model was generated for ten different grapevine cultivars,
426 which increases the complexity of the problem. Prediction results were moderate (R^2 Pred =
427 0.80), but it can be observed in Fig. 12 that the maximum error for test samples was around
428 ± 2 so it was misclassified in the worst case as one class above or below.

429 Another SVM model was built regarding the berry size quality component. The results (Table 3)
430 were good, with an error in calibration and prediction of less than 0.23 cm^3 and an R^2 above
431 0.82. The model built was very robust because the samples used for the prediction fit even
432 better than those used in the calibration (RMSEP < RMSEC). It should be mentioned, however,
433 that for the compactness descriptors, this 3D descriptor was calculated based only on the
434 automatic reconstruction of the 3D model and it did not need a controlled background.

435

436 3.2. Other cluster yield components

437 Three different SVM models were also built for each of the morphological descriptors
438 estimated (cluster volume, total berry weight and number of berries). These models were built
439 using the same distribution of samples for calibration and testing as those used for the quality
440 components. Just one 3D descriptor was used for each yield component (the 3D descriptor
441 with the same name as the yield component, except for weight, which was calculated with the
442 cluster volume). As can be seen in Table 3, the results for cluster volume and total berry
443 weight showed a determination coefficient (R^2) in prediction higher than 0.82. For the manual
444 measurement of cluster volume, the complete cluster (including pedicels and branches) was
445 measured, while for the image method employed, only the information about the
446 reconstructed berries was used, so it seems coherent that a better result was obtained using
447 berry weight alone. The outcomes were lower for number of berries (approx. 24 berries of
448 error in prediction) probably because many berries were occluded inside the cluster (especially
449 for the samples with greater compactness).

450

451 4. Conclusions

452 The research focused on estimating grape yield components using stereo vision. Due to the
453 difficulty of making the correct correspondence between the pair of images, a 3D
454 reconstruction tool was developed to obtain an accurate 3D model of the samples with a
455 reconstruction error of less than 1 mm. It has been proven that the tool actually achieves the
456 goal of reconstructing the model successfully while complying with the standards of usability,
457 speed and user-friendliness. In fact, using the 3D models obtained only from the automatic
458 approach, berry size was estimated by an SVM model with an R^2 in prediction higher than 0.82.
459 Once three-dimensional models were available, new 3D descriptors were extracted from them
460 to assess the compactness quality component: concavity measure, intersection between
461 berries and number of berries per area. These descriptors were evaluated and compared with
462 the state-of-the-art index CI-12 on 100 different samples from 10 different varieties. The

463 concavity measure descriptor gave a Kendal tau correlation of -0.71 ($p < 0.01$) compared with
464 the 0.52 ($p < 0.01$) obtained by CI-12. An SVM was developed using these new 3D descriptors
465 with an R^2 in prediction higher than 0.80, with a maximum classification error of one class. In
466 addition, other yield components: cluster volume, total berry weight and number of berries,
467 were estimated using SVM models, obtaining the following R^2 in prediction, respectively: 0.82,
468 0.83 and 0.71.

469
470 The results achieved show the capability of this technique for solving the problem of having an
471 accurate and objective tool for measuring cluster compactness. In addition, the fact that the
472 berry size quality component was estimated automatically without a controlled background
473 made this technique very feasible for use under field conditions. It would be very useful, for
474 example, to assess grapevine quality components in inter varietal studies (such as genetic
475 association studies).

476
477 Grape clusters were measured in a difficult image acquisition scenario with the aim of
478 simulating field conditions. Future work will focus on extrapolating these results into the field
479 scenario.

480

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486

487 References

- 488 Benlloch, J., Agustí, M., Sanchez, A., & Rodas, A. (1995). Colour segmentation
489 techniques for detecting weed patches in cereal crops. *Proc. of the 4th*
490 *Workshop on Robotics in Agriculture and the Food-Industry*, 71–81.
- 491 Brosnan, T., & Sun, D.-W. (2004). Improving quality inspection of food
492 products by computer vision—a review. *Journal of Food Engineering*,
493 *61*(1), 3 – 16. doi:[http://dx.doi.org/10.1016/S0260-8774\(03\)00183-3](http://dx.doi.org/10.1016/S0260-8774(03)00183-3)
- 494 Cubero, S., Diago, M. P., Blasco, J., Tardáguila, J., Millán, B., & Aleixos, N.
495 (2014). A new method for pedicel/peduncle detection and size
496 assessment of grapevine berries and other fruits by image analysis.
497 *Biosystems Engineering*, *117*(0), 62–72.
498 doi:[10.1016/j.biosystemseng.2013.06.007](https://doi.org/10.1016/j.biosystemseng.2013.06.007)
- 499 Cyganek, B., & Siebert, J. P. (2011). *An introduction to 3D computer vision*
500 *techniques and algorithms*. John Wiley & Sons.
- 501 Diago, M. P., Sanz-Garcia, A., Millan, B., Blasco, J., & Tardaguila, J. (2014).
502 Assessment of flower number per inflorescence in grapevine by image
503 analysis under field conditions. *Journal of the Science of Food and*
504 *Agriculture*, *94*(10), 1981–1987. doi:[10.1002/jsfa.6512](https://doi.org/10.1002/jsfa.6512)
- 505 Kendall, M. (1970). Rank Correlation Methods (4th edn.) Griffin. London, UK.
- 506 Quevedo, R., & Aguilera, J. M. (2010). Computer Vision and Stereoscopy for
507 Estimating Firmness in the Salmon (Salmon salar) Fillets. *Food and*

508 *Bioprocess Technology*, 3(4), 561–567. doi:10.1007/s11947-008-0097-
509 3

510 Roscher, R., Herzog, K., Kunkel, A., Kicherer, A., Töpfer, R., & Förstner, W.
511 (2014). Automated image analysis framework for high-throughput
512 determination of grapevine berry sizes using conditional random
513 fields. *Computers and Electronics in Agriculture*, 100, 148–158.

514 Sacher, A. J., & Marchant, J. A. (2000). Fusing 3D information for crop/weeds
515 classification. In *Pattern Recognition, 2000. Proceedings. 15th*
516 *International Conference on* (Vol. 4, pp. 295–298 vol.4).
517 doi:10.1109/ICPR.2000.902917

518 Sampson, D. J., Chang, Y. K., Rupasinghe, H. P. V., & Zaman, Q. U. (2014). A
519 dual-view computer-vision system for volume and image texture
520 analysis in multiple apple slices drying. *Journal of Food Engineering*,
521 127(0), 49–57. doi:10.1016/j.jfoodeng.2013.11.016

522 Sánchez, A. J., Albarracín, W., Grau, R., Ricolfe, C., & Barat, J. M. (2008).
523 Control of ham salting by using image segmentation. *Food Control*,
524 19(2), 135–142. doi:10.1016/j.foodcont.2007.02.012

525 Schölkopf, B., Smola, A. J., Williamson, R. C., & Bartlett, P. L. (2000). New
526 Support Vector Algorithms. *Neural Computation*, 12(5), 1207–1245.
527 doi:10.1162/089976600300015565

528 Sun, C., Berman, M., Coward, D., & Osborne, B. (2007). Thickness
529 measurement and crease detection of wheat grains using stereo
530 vision. *Pattern Recognition Letters*, 28(12), 1501–1508.
531 doi:10.1016/j.patrec.2007.03.008

532 Tello, J., & Ibáñez, J. (2014). Evaluation of indexes for the quantitative and
533 objective estimation of grapevine bunch compactness. *Vitis* - , 53 (1),
534 pp. 9-16. *Journal of Grapevine Research*, 53, 9–16.

535 Udomkun, P., Nagle, M., Mahayothee, B., & Müller, J. (2014). Laser-based
536 imaging system for non-invasive monitoring of quality changes of
537 papaya during drying. *Food Control*, 42(0), 225–233.
538 doi:10.1016/j.foodcont.2014.02.010

539 Verdú, S., Ivorra, E., Sánchez, A. J., Girón, J., Barat, J. M., & Grau, R. (2013).
540 Comparison of TOF and SL techniques for in-line measurement of food
541 item volume using animal and vegetable tissues. *Food Control*, 33(1),
542 221–226. doi:10.1016/j.foodcont.2013.02.031

543 Whalley, J., & Shanmuganathan, S. (2013). Applications of image processing
544 in viticulture: A review.

545 Woodford, O., Pham, M.-T., Maki, A., Perbet, F., & Stenger, B. (2014).
546 Demisting the Hough Transform for 3D Shape Recognition and
547 Registration. *International Journal of Computer Vision*, 106(3), 332–
548 341. doi:10.1007/s11263-013-0623-2

Descriptors	Morphological (measured by hand)	3D (automatic measures)	Units
Cluster length	Non-destructive	Non-destructive	$m \cdot 10^{-2}$
Cluster width	Non-destructive	Non-destructive	$m \cdot 10^{-2}$
Cluster volume	Non-destructive	Non-destructive	$m^3 \cdot 10^{-3}$
Cluster weight	Non-destructive	Estimated	$kg \cdot 10^{-3}$
Berry size	Non-destructive	Non-destructive	$m^3 \cdot 10^{-6}$
Number of berries per cluster	Destructive	Estimated	
Seeds per berry	Destructive	Occluded	
Pedicle length	Destructive	Occluded	$m \cdot 10^{-2}$
Rachis weight	Destructive	Occluded	$kg \cdot 10^{-3}$
First to seventh rachis node length	Destructive	Occluded	$m \cdot 10^{-2}$
Ramifications per cluster	Destructive	Occluded	
Concavity measure	Not possible	Non-destructive	
Intersection between berries	Not possible	Non-destructive	$m^3 \cdot 10^{-6}$
Number of berries per area	Not possible	Non-destructive	

Table 1 Morphological and 3D descriptors

Compactness descriptors	Rho
CI-12: B. weight/B. length ²	0.530**
Concavity measure	-0.710**
Intersection between berries	0.569**
Number of berries per area	-0.205*

Table 2 Kendall tau results for compactness descriptors **p<0.01 *p<0.05

	Number of SVs	RMSEC	RMSECV	RMSEP	R² Cal	R² CV	R² Pred
Compactness	20	0.920	1.139	0.817	0.886	0.826	0.808
Berry size	16	0.214	0.227	0.180	0.842	0.820	0.830
Cluster volume	36	28.204	46.945	56.798	0.944	0.845	0.822
Berry weight	42	19.549	36.672	44.656	0.966	0.880	0.830
Number of berries	23	13.307	23.821	23.791	0.947	0.826	0.714

Table 3 SVM results of the cluster components

Figure 01

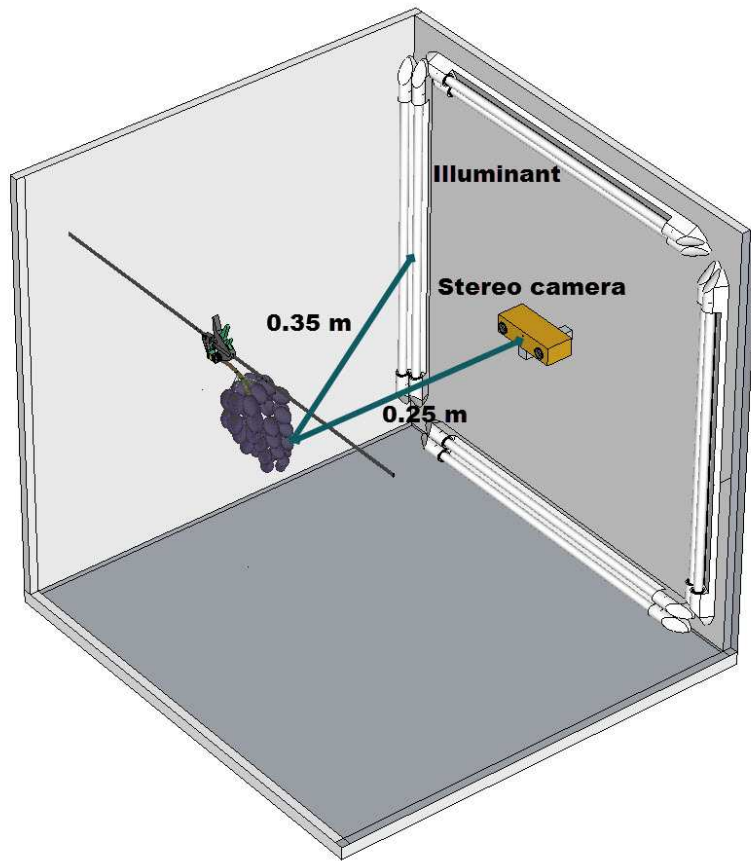


Figure 02

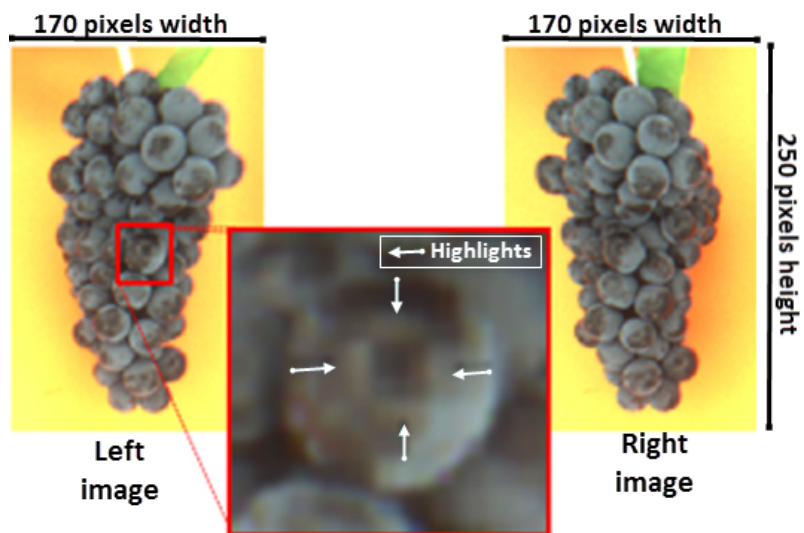


Figure 03

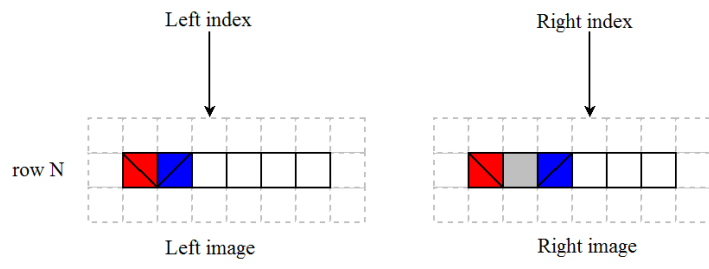


Figure 04

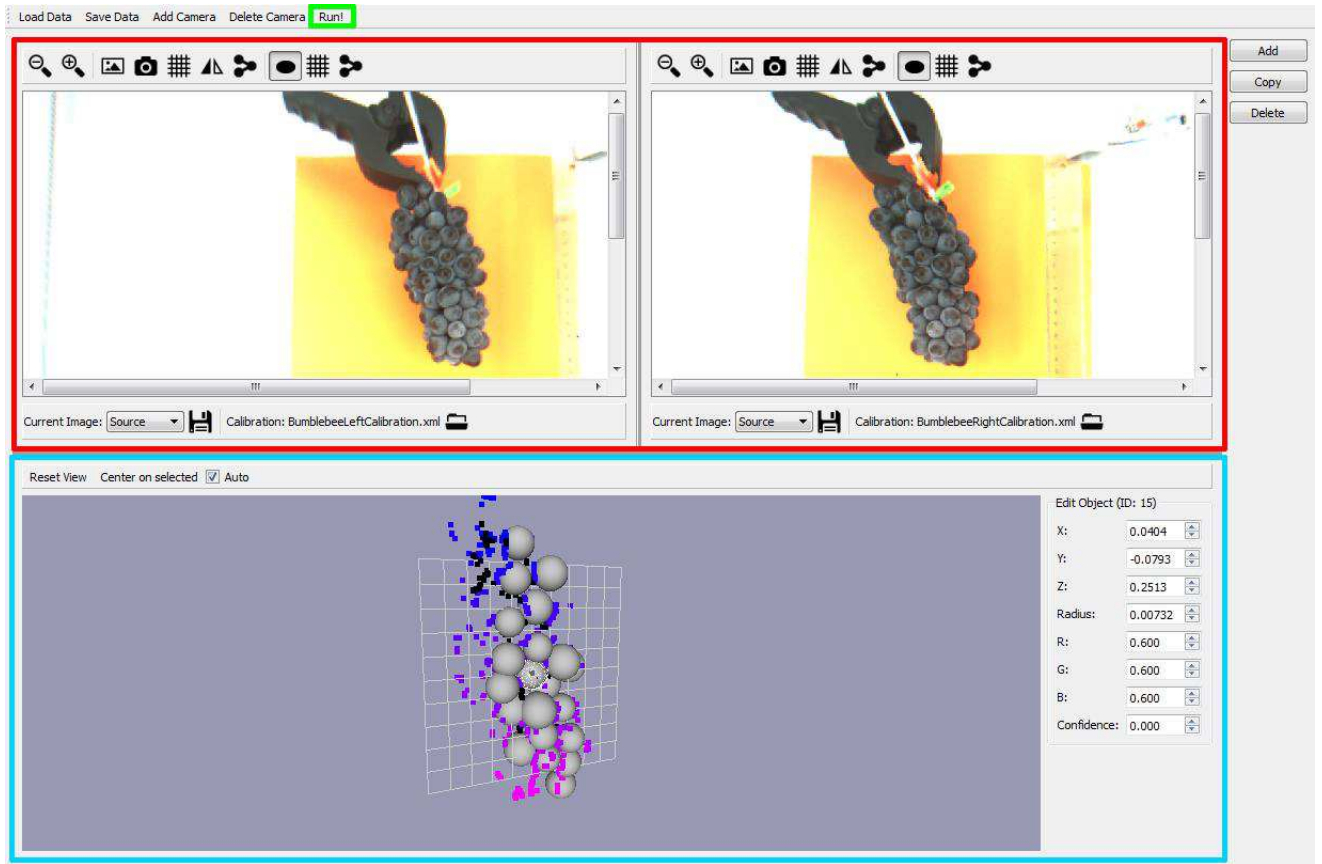


Figure 05

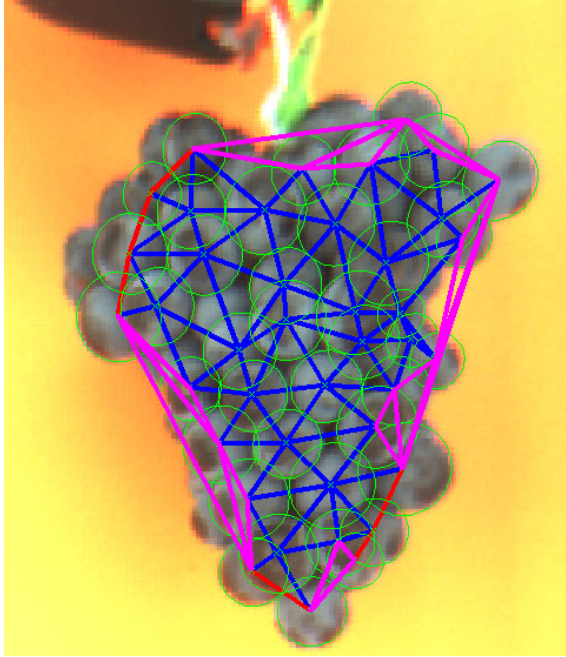


Figure 06



Figure 07

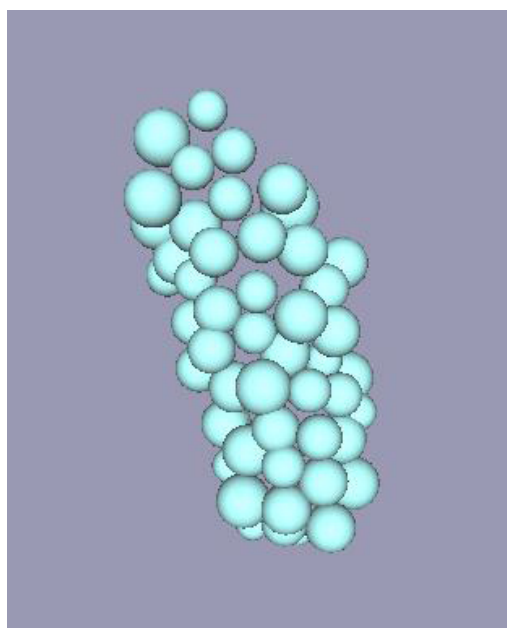


Figure 08

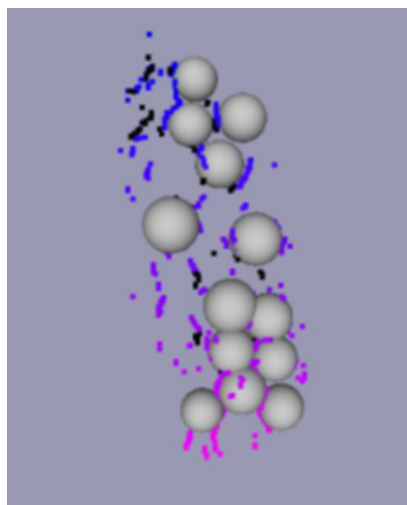


Figure 09

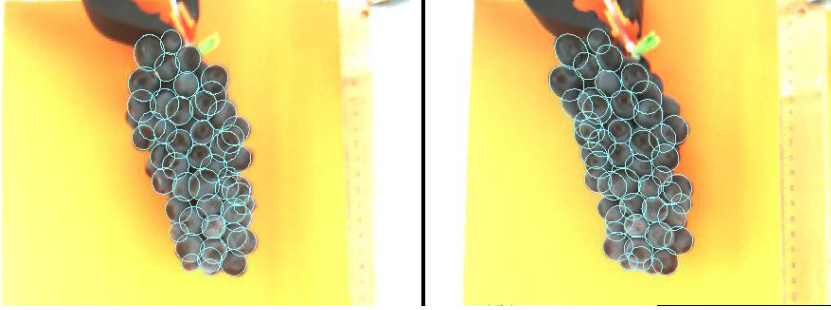


Figure 10



Figure 11

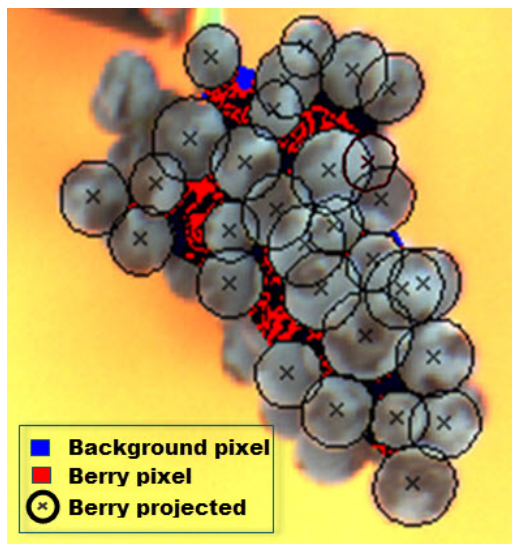


Figure 12

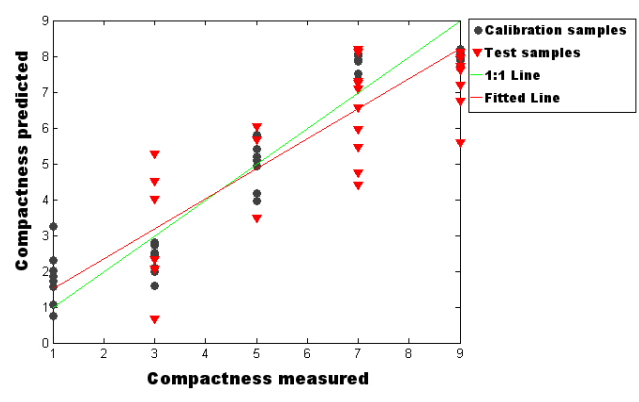


Fig. 01 Image acquisition set-up.

Fig. 02 Images captured with low resolution and highlights (Bobal sample 6)

Fig. 03 A node example. A pair of features with the same color represents a matching pair. The gray color represents an occluded feature.

Fig. 04 3D reconstruction tool interface. The 2D viewports are marked in red. The 3D viewport is marked in blue.

Fig. 05 Convex hull created by Delaunay triangulation.

Fig. 06 Region of interest selected for pixel classification.

Fig. 07 3D model refined by hand.

Fig. 08 Berries detected using Hough transform. Color points are the 3D points extracted from the vertical edges. Black points represent 3D points between berries.

Fig. 09 Checking the 3D model on the rectified images.

Fig. 10 Image segmentation of sample 2 Cabernet Franc (Compactness class 3).

Fig. 11 Image segmentation of sample 2 Cabernet Franc (Compactness class 7).

Fig. 12 SVM compactness results.

Table 1 Morphological and 3D descriptors

Table 2 Kendall tau results for compactness descriptors **p<0.01 *p<0.05

Table 3 SVM results of the cluster components