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Additional Information

1       **Modeling and optimization of the E-beam treatment of chicken steaks**  
2       **and hamburgers, considering food safety, shelf-life, and sensory quality.**

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11  
12       **ABSTRACT**

13       The present work was carried out to model the effect of E-beam treatment on the safety,  
14       shelf-life and sensory attributes of two poultry products, steaks and hamburgers, and to  
15       optimize the radiation treatment. The inactivation of *Salmonella* spp. by means of different  
16       radiation doses was modeled using a first order kinetics. The shelf-life was studied by  
17       periodically counting the bacterial number in samples. **For the modelling of experimental data,**  
18       **only the exponential phase of growth was taking into account.** The effect of the radiation dose  
19       on the sensory attributes (appearance, odor and flavor) and instrumental color ( $L^*$ ,  $a^*$  and  $b^*$   
20       parameters) was modeled using the Gompertz function and the Activation-Inactivation or  
21       linear models. The optimization of the radiation dose was carried out by maximizing the  
22       sensory scores of samples and minimizing the instrumental color changes. The safety and the  
23       shelf-life of samples were assured by introducing constraints in the optimization problem. In  
24       the case of hamburgers, the optimum calculated dose was 2.04 kGy, which guarantees the  
25       safety of the product and provides the best combination of sensory and instrumental  
26       attributes. As regards the steaks, the optimum assessed dose was 1.11 kGy, significantly lower  
27       than for hamburgers.

28       **KEYWORDS:** radiation, *Salmonella*, appearance, color, objective function, constraint,

## 30 1. INTRODUCTION

31 Nowadays, the sale of food products in individual or domestic portions is very common. In the  
32 meat industry, it is possible to differentiate two categories of these presentations: that  
33 corresponding with convenience, ready-to-eat (RTE) foods, including different kinds of cooked  
34 or dry-cured meats and dry fermented sausages, and small pieces of ready-to-cook (RTC) meat  
35 (steaks, hamburgers, etc.). This last category is one of the most common products in the  
36 poultry meat industry. To prepare these products, it is necessary to carry out several  
37 operations, such as cutting, mixing or shaping, which increase the risk of cross-contamination,  
38 the most important contaminant organisms in this case being *Salmonella* spp. and  
39 *Campylobacter jejuni* (Nachamkin, 2007; Cutter et al., 2012). Despite efforts by the industry to  
40 remove *Salmonella* from meat and poultry carcass, it is not possible to avoid the meat  
41 contamination from both the animals during slaughtering and the industrial equipment, which  
42 are the main sources of *Salmonella* contamination, not only in the case of the carcasses but  
43 also the small meat pieces (Lillard, 1990; Domingues et al., 2002; Ruban et al., 2010; Panisello  
44 et al., 2000; Cutter et al., 2012). Consequently, there is a need to improve microbial control so  
45 as to minimize the contamination of the final product, in order both to reduce the incidence of  
46 foodborne pathogens and to extend the shelf-life of the products.

47 Heat treatment is the technique most commonly used to reduce the microbial load of many  
48 foods. However, this technique cannot be applied to fresh meat due to the associated changes  
49 in the product characteristics (flavor, odor, color, appearance or texture). Therefore, to reduce  
50 the bacterial number in meat products, including pathogens, different non-thermal sanitizing  
51 treatments have been proposed. Of them, high hydrostatic pressure (Lakshmanan and  
52 Dalgaard, 2004), oscillating magnetic fields (Barbosa-Cánovas et al. 1998), light pulses (Hierro  
53 et al., 2012) or E-beam radiation (Cabeza et al., 2007; Garcia-Márquez et al., 2012) are sound  
54 alternatives.

55 Irradiation is known as an effective way to eliminate foodborne pathogens, such as *Listeria*  
56 *monocytogenes*, *Salmonella* spp., *Yersinia enterocolitica* or *Escherichia coli* O157:H7 (Cabeza et  
57 al., 2007, 2009; Schilling et al., 2009). However, it has been reported that this technology has  
58 limited application for meat, since an excessive level of irradiation can produce changes in the  
59 sensory properties of treated products, which could significantly affect the consumer  
60 acceptance (Arthur et al., 2005; Lee and Ahn, 2005). In this regard, the odor of excessively  
61 irradiated meat has been described as being like rotten egg, cooked meat, hot culture  
62 medium, sulphur, alcohol, acetic acid, liver-like serummy, and bloody (Brewer, 2009; Cabeza et  
63 al., 2007). Therefore, the adjustment of the radiation dose is a critical point; it must be set to a

64 level that allows an adequate level of microbial inactivation to be achieved (food safety), while  
65 at the same time minimizing the changes in treated meat to avoid consumer rejection and, an  
66 additional, valuable feature, to extend the shelf-life.

67 The mathematical modelling of the influence of the radiation dose on the microbial load  
68 reduction and the sensory properties retention of E-beam treated products, permits the  
69 quantification of effects and, in a later step, the optimization of treatments (Benedito et al.,  
70 2011). For that purpose, it is necessary to consider models that define the inactivation of  
71 microorganisms, the change in the sensory properties and the growth of spoilage  
72 microorganisms after treatments in order to estimate the product shelf-life. The model  
73 proposed by Bigelow (1921) is probably one of the most commonly used to describe microbial  
74 thermal inactivation. This traditional first-order model was developed as a way of quantifying  
75 the effect of the duration of the heat treatment on the reduction of the viable organisms. A  
76 similar model was used, replacing the time variable by the radiation dose, to assess the  
77 influence of this factor on the inactivation of *L. monocytogenes* in vacuum-packaged cooked  
78 ham (Benedito et al., 2011). Other models that have also been used, not only to define  
79 microorganism or enzyme inactivation, but also the kinetic changes of quality attributes, are  
80 the Gompertz function (Ding et al. 2010) or the Activation-Inactivation model (Soysal, 2008).

81 The shelf-life of products treated by means of any preservation technology is a factor which it  
82 is very important to take into account in order to achieve process optimization and, in the case  
83 of meat, it is linked to the growth of microorganisms that produce the spoilage of food. In this  
84 regard, of the models used to define the microbial growth, Hill's model or the modified  
85 Gompertz equation (Huang, 2010) are worth mentioning. However, the classical linear models  
86 have also been used because of their simplicity and the acceptable way they fit to numerous  
87 different cases (McKellar and Lu, 2004).

88 Then, the optimization of the E-beam treatment using the modeling of the radiation effects on  
89 the microbial and quality factors of the treated products may be an interesting tool for process  
90 design. Although this type of optimization has barely been reported, it has been developed for  
91 vacuum-packaged cooked ham (Benedito et al., 2011). However, as far as the authors know,  
92 this strategy has not been applied to raw poultry meat. Therefore, the goal of this work was to  
93 model the effect of E-beam on the safety, shelf-life and sensory attributes of two chicken  
94 products (steaks and hamburgers) and to optimize the radiation treatment, maximizing the  
95 product quality while guaranteeing its microbial safety and a reasonable shelf-life.

## 96 **2. MATERIALS AND METHODS**

97 **2.1. Microorganisms**

98 The *Campylobacter jejuni* is a very sensitive microorganism to ionizing radiation being the  
99 reported decimal reductions (D-value) lower than 0.2 kGy (Clavero et al., 1994; Verde et al.,  
100 2004). For this reason, the target organism selected for this study was the other main  
101 pathogenic bacteria present in poultry meat, i.e. *Salmonella* spp., namely *Salmonella enterica*  
102 serovar Enteritidis (CECT4300) and *Salmonella enterica* serovar Typhimurium (CECT 443). The  
103 strains were maintained by freezing (-40 °C) in trypticase soy broth (TSB; Difco), adding 10 %  
104 glycerol as the cryogenic agent. Fresh cultures were prepared for each experiment by  
105 removing a piece of frozen culture from vials and inoculating it into 9 mL of TSB, then  
106 incubating it at 32 °C for 24h. The cultures were then centrifuged (at 4 °C) and the pellet  
107 suspended in beakers with 50 mL sterile saline solution, which yielded a bacterial load of  
108 approximately 10<sup>8</sup> cells/mL that were used to contaminate meat samples.

109 **2.2. Sample preparation and radiation treatment**

110 Chicken breasts, immediately separated from the carcass, were obtained in a local market and  
111 transported to the laboratory at 4 °C. Several breasts were cut into steaks (8-12 g, 3 mm  
112 thickness). To establish the death kinetics of *Salmonellae*, a batch of these steaks was  
113 contaminated by immersion for one minute in the bacterial suspension obtained as previously  
114 described. The contaminated (kinetics studies) and uncontaminated (for the determination of  
115 color, sensory characteristics and shelf-life) samples were packed in a plastic bag of low gas  
116 permeability (diffusion coefficient of 35 cm<sup>3</sup> 24 h<sup>-1</sup>m<sup>-2</sup> bar to oxygen and 150 cm<sup>3</sup> 24 h<sup>-1</sup>m<sup>-2</sup> bar  
117 to carbon dioxide) and thermo-sealed. Other set of breasts was chopped (hamburger samples)  
118 using a domestic mincing machine and divided in two batches. In order to mimic the deep  
119 cross contamination produced in the industry during chopping operation, a set was  
120 contaminated with a bacterial suspension (bs) at the ratio 1ml bs/20 g meat previously to be  
121 minced. Then, aliquots of 20 g (contaminated and no contaminated batches) were placed into  
122 Petri dishes (0.5 cm diameter and 0.5 cm height). Once hamburgers were ready, they were  
123 handled as described for steaks. The samples were transported (less than 1 h) in insulated  
124 polystyrene boxes (<5°C) to the irradiation plant (IOSNISIOS sterilization S.A., Tarancón,  
125 Cuenca, Spain) and irradiated under an electron beam radiation source, which operates at 10  
126 MeV. The radiation doses employed were 0, 1, 2, 3, and 4 kGy. The actual dose absorbed by  
127 the samples was determined by assessing the absorbance of cellulose triacetate dosimeters  
128 (STM, American Society of Testing and Materials, 2000) simultaneously irradiated with the  
129 samples. The experiments were carried out in triplicate at room temperature (18 - 20 °C) and,

130 after treatment, the samples were transferred to the laboratory and stored at 4 °C until use.  
131 During treatment, the product temperature increased by less than 2 °C.

### 132 **2.3. Microbial analyses**

133 To count the number of total viable organisms in hamburgers and steaks, 10 g of sample were  
134 homogenized with 90 mL of a sterile saline solution in a Stomacher bag for two minutes.  
135 Counts were determined on the surface of plates with TSB and using a spiral plate system  
136 (model Eddy Jet, IUL Instrument, Barcelona, Spain). Since the fresh poultry only presents a very  
137 complex microbiota dominated by the indigenous microbiota, a selective medium VRBG (violet  
138 red bile glucose agar, Oxoid) was used for *Salmonellae* enumeration in order to avoid the  
139 count of background microbiota. Previously, it was observed that this strategy did not affect  
140 the growth of the survival cells (Cambero et al., 2011). In every case, plates were incubated at  
141 32 °C for 24-36 h. Colonies were enumerated with an automatic counter (CounterMat Flash  
142 model, IUL Instrument, Barcelona, Spain).

### 143 **2.4. Sensory analyses**

144 Independent triangular, rank order and descriptive test trials were carried out on three  
145 sensory characteristics of meat samples, appearance, odor and flavor, in order to determine  
146 the possible influence of the radiation dose (0, 1, 2, 3 and 4kGy) on meat quality. The samples  
147 were evaluated by a panel of twenty tasters (ten women and ten men). The panelists were  
148 previously trained and familiarized with the terms necessary to describe the sensory  
149 characteristics of the chicken steaks and hamburgers, such as general appearance (color and  
150 brightness), odor (richness and intensity), off-odor (absence), taste (richness of taste notes,  
151 off-taste absence, tenderness, juiciness and after-taste intensity) as well as the expected off-  
152 sensory features resulting from the E-beam treatment (off-odors and taste, such as hot culture  
153 medium, sulfuric, metallic, scalded feather, burnt feather, pungent pepper). The evaluations  
154 were performed in individual booths built according to the International Standards  
155 Organization DP66.58 criteria (ISO, 1981a). The evaluation was carried out between meals,  
156 after breakfast, and before the midday meal. To reduce fatigue, panel members performed  
157 three sessions per day with a minimum break of 1 h between sessions. Three independent  
158 tests were performed to evaluate appearance, odor and flavor.

159 Prior to sensory analysis, the samples were removed from the refrigerator and maintained in a  
160 temperature-controlled room at 10 °C for no more than 1 h. For the appearance test, closed  
161 thermo-sealed bags with the steak or hamburger samples were supplied to the panelist, thus  
162 avoiding direct contact with the sample. For the odor test, the bags with samples were opened

163 just before the test. To evaluate the flavor, samples of around 5 g were cooked in a domestic  
164 microwave (800 W) at 30 % of the total power for 60 s. Afterwards, they were maintained (2-5  
165 min) at 75 °C in an infrared oven until the test. The tasters received unsalted crackers and  
166 water at room temperature to cleanse the palate between samples. White fluorescent light  
167 was used during appearance analysis. The odor and flavor of the samples were evaluated  
168 under red light conditions.

169 The sensory analyses were carried out just after treatment and after 5 days of storage at 4 °C.  
170 The triangle test (ISO, 1981b) was performed by the forced choice option, in which the tasters  
171 must choose the sample that, in their opinion, is different. All the possible combinations of  
172 untreated and irradiated samples were tested.

173 For the rank order test, five samples (untreated and treated) were supplied for each test and  
174 the panelists were instructed to rank samples in order of preference, according to the  
175 proximity of the sensory characteristic (appearance, odor or flavor) of the analyzed sample to  
176 the fresh product. For this, a 5-point preference scale (in which 1 corresponded to the lowest  
177 and 5 to the highest) was used. No repetitions were allowed. Results of the rank order test  
178 were used to obtain the sum of ranks, which corresponds to the sum of the scores of sample  
179 preference (the sum of the products of the value given for each sample on a 5-point scale  
180 multiplied by the number of times that each sample was allocated this specific score) for a  
181 specific sensory characteristic. The significance level of data was determined by the Friedman  
182 rank addition following the model proposed by Joanes (1985) and the tables for multiple  
183 comparison procedures for the analysis of ranked data (Christensen et al., 2006). The sum of  
184 the ranks, as quantitative values of the sensory evaluation, was used in the modeling,  
185 statistical analysis, and optimization of the irradiation process.

186 Panelists were also asked to provide information regarding specific characteristics (color,  
187 brightness, odor, taste, texture, any off-sensory feature) of the chicken samples by following a  
188 profile descriptive analysis (ISO, 2003). They were also asked to qualify the intensity of these  
189 sensations with the following terms: negligible or very slight, slight, moderate and strong.

## 190 **2.5. Instrumental color determination**

191 The color of the sample surface was measured using a tristimulus colorimeter (Minolta Chroma  
192 Meter CR300, Minolta Corporation, NJ) provided with a 10° standard observer and a D65  
193 standard illuminant. In each sample, the L, a and b coordinates (CIE-Lab) were measured in  
194 quadruplicate. The measurements were taken the same day of the irradiation treatment and

195 after 5 days of storage at 4 °C and in both cases after 4-5 minutes after opening the bags  
196 containing the samples.

## 197 **2.6. Shelf-life determination**

198 To assess the shelf-life, irradiated and non-irradiated samples were removed from the bags  
199 and the total viable counts were determined. From a microbiological point of view, the end of  
200 shelf-life was established when this exceeded the value of  $5 \times 10^7$  cfu/g. Analyses were  
201 performed just after E-beam treatment (0 days) and at various times during storage (2, 4, 6, 8  
202 and 11 days) until the end of the shelf-life. The shelf-life was also assessed by means of  
203 sensory analysis (odor and visual appearance).

## 204 **2.7. Estimation of the food safety objective (FSO)**

205 For Salmonellae, the “zero tolerance” (absence in 25 g of product) is a universal criterion and,  
206 therefore, it has been adopted in most microbiological regulations. This level may be  
207 considered as equivalent to a FSO of 4 cells/100 g ( $\log_{10} = -1.39$ ). Previously, it has been  
208 reported that, according to the zero tolerance criterion, a treatment of 1.5 kGy warrants safe  
209 ready-to-eat intermediate moisture foods (dry-cured ham, beef or smoked tuna), which  
210 accounted for a Salmonellae reduction of 2.3 D. The water activity ( $a_w$ ) of these products is  
211 lower than 0.88 (Cambero et al., 2012) and, therefore, *Salmonellae* are unable to grow.  
212 However, since the value of  $a_w$  is higher (about 0.985) in fresh poultry, the growth of  
213 *Salmonellae* is possible, particularly if a temperature abuse (for example, a temperature  
214 increase up to 8 - 10 °C) is produced during distribution or even at home. This fact and the  
215 reasoning used previously for sanitizing mayonnaise potato salad (Cambero et al. 2012), since  
216 the egg is also a product in which the *Salmonellae* acquire importance, was followed in this  
217 case to increase the degree of decimal reduction to 2.7 D to achieve the FSO.

## 218 **2.8. Modeling inactivation of *Salmonella* spp. by E-beam radiation**

219 To describe the influence of the radiation dose on the inactivation of *S. Enteritidis* and *S.*  
220 *Typhimurium*, a first order kinetics model was used, replacing the time variable by the  
221 treatment dose ( $d_{irr}$ ) (Equation 1), according to Benedito et al. (2011).

$$222 \quad \text{Log}(N_{d_{irr}}) = \text{Log}(N_{d_0}) - k \cdot d_{irr} \quad (1)$$

223 Where  $\log(N_{d_{irr}})$  and  $\log(N_{d_0})$  are the decimal logarithm of the number of microorganisms  
224 (cfu/g) after a radiation dose of  $d_{irr}$  and without treatment,  $d_0$ , respectively, and  $k$  is the rate  
225 constant ( $\text{kGy}^{-1}$ ).

## 226 **2.9. Modeling the E-beam radiation effect on sensory properties**



227 To describe the effect of radiation on the sensory attributes measured by panelists, several  
 228 inactivation models were assessed. However, only those which most accurately fitted the data  
 229 are included. Specifically, the Gompertz Function (**Equation 2**) and the Activation-Inactivation  
 230 model (**Equation 3**) (Mckellar and Lu, 2004) were adapted, replacing the time of treatment by  
 231 the irradiation dose:

$$232 \quad \text{Log} \left( \frac{S_{d_{irr}}}{S_{d_0}} \right) = C \cdot \exp(-\exp(A + B \cdot d_{irr})) - C \cdot \exp(-\exp(A)) \quad (2)$$

$$233 \quad \text{Log} \left( \frac{S_{d_{irr}}}{S_{d_0}} \right) = \left[ \frac{d_{irr}}{k_1 + k_2 \cdot d_{irr}} \left( 1 - \log(1 + \exp(k_3 \cdot d_{irr}))^m \right) \right] \quad (3)$$

234 where  $S_{d_{irr}}$  and  $S_{d_0}$  are the scores assigned by panelists for the appearance, odor or flavor of  
 235 samples irradiated using different doses ( $d_{irr}$ ) or without irradiation ( $d_0$ ). The  $A$ ,  $B$  and  $C$   
 236 variables correspond with the model parameters of Gompertz Function and the  $k_1$ ,  $k_2$ ,  $k_3$  and  $m$   
 237 correspond with those of the Activation-Inactivation model. The models were fitted to the  
 238 data of sensory parameters corresponding to the same day that irradiation took place (0 days)  
 239 and after 5 days of storage at 4 °C.

#### 240 **2.10. Modeling the influence of E-beam radiation on instrumental color.**

241 From the experimentally measured CIE-Lab coordinates ( $a^*$  and  $b^*$ ), the chroma was calculated  
 242 (**Equation 4**).

$$243 \quad \text{Chroma} = \sqrt{a^{*2} + b^{*2}} \quad (4)$$

244 The influence of the radiation dose on chroma was described by a linear relationship (**Equation**  
 245 **5**).

$$246 \quad \text{Chroma} = p + q \cdot d_{irr} \quad (5)$$

#### 247 **2.11. Modeling the E-beam radiation effect on the product shelf-life.**

248 Throughout storage, the changes in the total microorganism count in hamburgers and steaks,  
 249 radiated or not, were modeled through a first order kinetics model. The meat could be  
 250 considered as a biological reactor where organisms are in the exponential growth phase.  
 251 Under these conditions, the number of microorganisms during storage could be estimated  
 252 from a balance of biomass (**Equation 6**).

$$253 \quad \frac{d(M_s X)}{dt} = \mu M_s \quad (6)$$

254 where

255  $M_s$  represents the mass of meat sample (g),  $X$  the number of microorganisms per mass unit  
256 (ufc/g),  $t$  the storage time (days) and  $\mu$  the rate of microorganism growth. Considering only the  
257 exponential growth phase,  $\mu$  is the constant maximum growth rate. **Equation 7** is obtained by  
258 integrating **Equation 6**.

$$259 \quad \ln\left(\frac{X_t}{X_0}\right) = \mu \cdot t \quad (7)$$

260 where  $X_0$  and  $X_t$  stand for the number of microorganisms at the beginning of storage and after  
261 time  $t$ , respectively.

### 262 **2.12. Model fitting and statistical analyses of experimental data**

263 The parameters of the models included in **Equations 1, 5 and 7** were identified by linear  
264 regression using the Microsoft Excel 2007™ software. In the case of **Equations 2 and 3**, the  
265 model parameters were identified by minimizing the sum of the square differences between  
266 the experimental and calculated data of sensory properties using Solver tool from Microsoft  
267 Excel™.

268 In order to assess the ability of these models to fit the experimental data, the percentage of  
269 explained variance (*%var*, **Equation 8**) and the mean relative error (*MRE*, **Equation 9**) were  
270 computed (Lipson and Sheth, 1973).

271

$$272 \quad \% \text{ var} = \left(1 - \frac{S_{se}^2}{S_s^2}\right) \quad (8)$$

273

$$274 \quad MRE = \frac{100}{N} \sum_{i=1}^N \frac{|v_{ei} - v_{ci}|}{v_{ei}} \quad (9)$$

275

276 where  $S_s^2$  and  $S_{se}^2$  are the variance of the sample and the estimation, respectively,  $N$  the  
277 number of experimental data and  $v_{ei}$  and  $v_{ci}$  the values of the experimental and calculated  
278 variables, respectively.

279 On the other hand, the multifactor ANOVA and the LSD (Least Significant Difference) intervals  
280 (Statgraphics Plus 5.1; Statistical Graphics Corp.) were calculated to evaluate the significance  
281 of the differences between the experimental measurements carried out on samples treated  
282 with different radiation doses.

### 283 **2.13. Optimization of the radiation process**

284 The main objective of the study was to optimize the radiation dose in order to achieve the  
285 inactivation of *Salmonellae*, and then to reach a safe level, with minimum changes in the  
286 quality factors of poultry. Then, after modeling the influence of irradiation on both  
287 microbiological and quality factors, a mathematical optimization approach was performed. For  
288 that, an objective function was defined, the decision variables selected and some restraints  
289 were considered. The highest quality of meat was linked with the highest sensory property  
290 scores (appearance, odor and flavor) and the smallest color changes (instrumentally  
291 measured). Therefore, the objective function chosen was the ratio between the sum of scores  
292 for the sensory parameters divided by the color changes and the optimization algorithm was  
293 set to maximize this function. The restraints considered were those related with both the food  
294 safety and the shelf-life of the product. For food safety, it was considered that the irradiation  
295 treatment must produce a 2.7D reduction of *Salmonella* spp., according to the FSO defined in  
296 **Section 2.8**. On the other hand, the shelf-life was considered to be expired when the bacterial  
297 counts reached the level  $5 \times 10^7$ cfu/g. The decision variable was the radiation dose. The  
298 optimization problem was solved using the Generalized Reduced Gradient method available in  
299 the Solver tool of Microsoft Excel™.

### 300 **3. RESULTS AND DISCUSSION**

#### 301 **3.1. Influence of radiation on the inactivation of *Salmonella* spp.**

302 As expected, the radiation of chicken samples led to a reduction in the load of *Salmonellae*  
303 proportional to the intensity of the treatment. As can be observed in **Table 1**, the first order  
304 kinetics model proposed in **Equation 1** was adequate for describing the effect of the radiation  
305 dose on the lethal power of E-beam. Thus, the percentage of explained variance (%var) of the  
306 model was above 98% for both *S. Enteritidis* and *S. Typhimurium*. For the latter, the inverse of  
307 the identified  $k$  (**Table 1**), the decimal reduction dose ( $D$ ), was 0.41 kGy in steaks and 0.52 kGy  
308 in hamburgers, and this difference was statistically significant ( $p < 0.05$ ). These data coincide  
309 with what is reported in literature being then the microbial inactivation rate dependent on the  
310 way the meat has been processed (Radomyski et al., 1994). In any event, the  $D$  values obtained  
311 are in the reported range. Thus, Thayer et al. (1990) reported a value of 0.53 kGy for  
312 mechanically deboned poultry, Grant and Patterson (1991) observed values from 0.40 to 0.44  
313 kGy for pork and more recently, a figure of 0.53 kGy has been reported for both dry fermented  
314 sausages (Cabeza et al., 2009) and dry cured ham (Cambero et al., 2012).

315 The  $D$ -values observed for *S. Enteritidis*, 0.37 kGy and 0.39 kGy in steaks and hamburgers  
316 respectively, were significantly lower ( $p < 0.05$ ) than those found for *S. Typhimurium*. Moreover,

317 the differences between steaks and hamburgers were not significant (**Table 1**) for this serovar.  
318 In this case, the values found in the literature were more variable, ranging from 0.77 kGy in  
319 mechanically deboned chicken to 0.17 kGy in growth media (Thayer et al., 1990), although  
320 similar values to those obtained in the present study have also been reported, e.g. 0.37 kGy in  
321 poultry meat (Mulder et al., 1977) or 0.41 kGy in dry fermented sausages (Cabeza et al., 2009).  
322 From these results, it may be concluded that *S. Enteritidis* was significantly more sensitive to  
323 radiation than *S. Typhimurium* and, therefore, the latter may be considered as the target  
324 microorganism in poultry steaks and hamburgers in order to ensure the product safety.

### 325 **3.2. Effect of irradiation on meat quality**

#### 326 **3.2.1. Sensory attributes**

327 In the triangular analysis performed just after radiation treatment, no significant differences  
328 ( $p > 0.05$ ) were detected in the sensory attributes of hamburgers when samples of 0 and 1 kGy  
329 were compared (**Table 2**). All the other combinations of sample treatments (both for chicken  
330 steaks and hamburgers) exhibited differences in, at least, one sensory characteristic. Many  
331 combinations showed differences in two or three sensory attributes (**Table 2**). In general,  
332 significant differences ( $p < 0.05$ ) in appearance and odor were found between the untreated  
333 and radiated samples. No significant differences ( $p > 0.05$ ) were detected between samples  
334 treated at 3 and 4 kGy (Table 2). The results of the microbiological analysis indicated that the  
335 untreated chicken products were not suitable for consumption after 5 days of refrigerated  
336 storage ( $> 5 \times 10^7$  cfu/g), which agrees fully with the flavor analysis performed. Significant  
337 differences ( $p < 0.05$ ) in flavor were detected when 1 kGy treated samples were compared  
338 with those subjected to doses of over 2 kGy.

339 The results of the sensory analysis obtained by means of the rank order test on the chicken  
340 products are shown in **Table 3**. As in the case of the triangular test, the panelists' scores  
341 showed that there were significant differences ( $p < 0.05$ ) in appearance, odor and flavor  
342 between samples irradiated with different doses, not only for hamburger samples but also for  
343 steaks (**Table 3**). Storage for 5 days at 4 °C also affected the sensory attributes. Thus, in the  
344 case of their appearance, the hamburger samples treated with doses above 2 kGy showed  
345 higher scores than non-radiated ones (**Table 3**), even after 5 days of storage. The appearance  
346 descriptive analysis of samples showed that the yellowish aspect of fresh hamburgers became  
347 pink-purple; the higher the radiation dose, the more intense the pink. In this sense, the  
348 samples irradiated at 3 or 4 kGy showed a similar pattern to that described for pork or turkey  
349 hamburgers. Nam and Ahn (2002) attributed the increase in the pink and red color of

350 irradiated meat to the formation of a carbon monoxide-myoglobin complex induced by the  
351 production of carbon monoxide and reducing conditions during radiation treatment. Samples  
352 treated at 2 kGy retained the appearance during storage but the non-radiated samples (0 kGy)  
353 and those treated with 1 kGy became pale and grayish.

354 On the contrary, for the steak samples (**Table 3**), the higher score obtained for samples  
355 irradiated above 2 kGy became, after storage, lower than that of non-radiated steaks. The 2  
356 kGy treated samples exhibited a pink color and a high degree of brightness that, despite not  
357 being the conventional color of chicken steaks, was associated with a higher quality fresh  
358 meat. However, after 5 days of storage, the color of the samples treated above 3 kGy became  
359 pale, yellowish and grayish.

360 The dose applied also affected the odor just after radiation (0 days), reducing the scores for  
361 both steaks and hamburgers as the radiation dose increased (**Table 3**). However, after 5 days  
362 of storage, samples radiated at doses over 2 kGy scored better than non-radiated ones, and  
363 the odor differences between radiation doses decreased. The odor of samples radiated at 1  
364 and 2 kGy was described as "scalded feather", while at 3 and 4 kGy it was defined as  
365 "irradiated" and "sulfured", which could make the commercialization of this product difficult.  
366 However, the intensity of this odor decreased after the storage time, achieving a final value  
367 which is adequate for consumption. Similar findings concerning the elimination of radiation  
368 off-odors during storage have been reported (Nam and Ahn, 2003; Brewer, 2009; García-  
369 Márquez et al., 2012), which has been explained by the dissipation of some volatiles (e.g.,  
370 dimethyl disulfide, dimethyl trisulfide, S-methyl ester) aroused during radiation (Du et al.,  
371 2002).

372 As regards the flavor, the results after radiation (0 days) were similar to those found for odor  
373 (**Table 3**). Just after treatment, steaks and hamburgers treated at doses higher than 2 kGy  
374 showed lower scores than non-treated samples. In the descriptive analyses, irradiated samples  
375 (particularly those treated at doses over 2 kGy) were judged less juicy and they presented a  
376 very slight taint of "burnt" or "hot culture medium" and a slightly astringent aftertaste,  
377 although they were considered acceptable for sale. It should be emphasized that the samples  
378 were cooked before carrying out the test, which can favor the dissipation of off-odors during  
379 cooking (Hashim et al., 1995). In general, after storage, panelists' scores of the samples treated  
380 with 1, 2, 3 or 4 kGy only showed non-significant differences (**Table 3**). Several authors have  
381 reported (Du and Ahn, 2002; Du et al., 2002) that post-radiation storage could allow the flavor  
382 to return to levels close to those of the untreated products as the volatile compounds are lost.

383 The adapted Gompertz Function (**Equation 2**) and the Activation-Inactivation model (**Equation**  
384 **3**) were fitted to the scores attributed by panelists to the appearance, odor and flavor  
385 characteristics of chicken-breast steaks and hamburgers. **Table 4** shows the identified model  
386 parameters, only including the model that best fitted each sensory attribute. Thus, the  
387 Gompertz function was the model that best fitted the appearance data (**Table 4, Figure 1**). In  
388 the case of odor and flavor, the Activation-Inactivation model achieved the highest values of  
389 percentage of explained variance (**Table 4**). Flavor is defined as the combined chemical  
390 sensations of taste and smell. Consequently, odor and flavor behaved in a similar way just after  
391 irradiation (0 days) and after 5 days of storage at 4 °C. The model was able to describe the  
392 influence of the applied irradiation dose on the odor of meat samples (**Figure 2**). The data  
393 concerning flavor was found to follow a similar tendency to that shown in **Figure 2**.

394 The fact that the radiation dose just after treatment had a different effect from the one after  
395 5, days of storage should be highlighted. After treatment, the tendency exhibited by the model  
396 between the doses applied and the odor or flavor scores was negative (**Figure 2**); the lowest  
397 scores were achieved as the applied radiation dose rose. This pattern changed for the samples  
398 stored for 5 days. In this case, the increase in the radiation dose led to an increase in the  
399 scores, but once a maximum was reached, the increase in the radiation dose brought about a  
400 decrease in the scores. Therefore, this behavior showed the existence of an optimum radiation  
401 dose, which provided the best flavor and odor after 5 days of storage.

### 402 **3.2.2. Instrumentally determined color**

403 The experimental results obtained from the CIE Lab analysis are shown in **Table 5**. The steak  
404 samples had lower  $a^*$ ,  $b^*$  and  $L^*$  values than the hamburger samples, probably due to the fact  
405 that the surface directly exposed to the air is much higher for the latter meat product. No clear  
406 trend was identified between the radiation dose or storage time and the  $L^*$  coordinate in  
407 hamburger samples. In steak samples, the radiation produced a decrease in the  $L^*$  value  
408 compared to the non-radiated samples, but no relationship was identified between  $L^*$  and the  
409 radiation dose applied. The  $L^*$  values of the untreated steaks after 5 days of storage at 4 °C  
410 were lower than those obtained just after the E-beam treatment, which could be related to  
411 the loss of the surface water. However, the brightness of the radiated samples was not  
412 significantly ( $p < 0.05$ ) affected by storage.

413 With regard to the  $a^*$  and  $b^*$  parameters, the radiation affected both coordinates. Then, the  
414 values measured just after radiation rose as a larger radiation dose was applied (**Table 5**),  
415 which means that both the hamburger and the steak samples became pink in colour. Thus, the

416 instrumental measurements of color were consistent with the results obtained in the sensory  
417 analyses (**Tables 2 and 3**) and the color changes observed by other authors (Nam and Ahn,  
418 2002) in meat and meat products after radiation. Some authors have speculated that when the  
419 myoglobin is primarily in the MbFe<sup>3+</sup> form, the radiation produces an increase in both the  $a^*$   
420 (Giroux et al., 2001; Satterlee et al., 1972) and  $b^*$  (Brewer, 2009) values in all meat species.

421 After 5 days of storage, the values of  $b^*$  for steaks were lower than those observed just after  
422 radiation, but the relationship between  $b^*$  and the radiation dose was maintained. Similar  
423 behavior has been observed in fresh pork loin (Garcia-Marquez et al., 2012). However, in the  
424 case of the hamburgers, no significant differences were detected in the  $b^*$  values associated  
425 with storage.

426 Therefore, it being difficult to identify a clear influence on the  $L^*$  coordinate, the  $a^*$  and  
427  $b^*$  parameters were the main ones affected by the E-beam treatment. For this reason, the  
428 evaluation of the global changes in sample color induced by radiation was carried out by  
429 chroma (**Equation 4**). As can be observed in **Figures 3A and 3B**, the relationship between  
430 radiation dose and chroma followed a linear trend. **Equation 5** was an adequate means of  
431 describing this relationship (**Table 6**), as can be observed in **Figures 3A and 3B** and from the  
432 mean relative error value obtained, in all cases lower than 5.26 %. The low value of the  
433 percentage of explained variance obtained for the samples stored for 5 days can be attributed  
434 to the wide experimental variability of the data.

435 As may be seen in **Table 6**, the  $y$ -intercept,  $p$ , was higher in hamburgers than in steaks  
436 according to the higher values of  $a^*$  and  $b^*$  measured in the former product. The slope of the  
437 linear relationship,  $q$ , of the just radiated hamburgers was almost twice as high as that  
438 obtained for steaks, which indicates that the color of hamburger was more sensitive than that  
439 of steaks to the increase in radiation. However after 5 days of storage, the identified slopes for  
440 hamburgers and steaks were quite similar.

### 441 **3.3. Modeling of microorganism growth in E-beam radiated samples.**

442 Compared with the non-radiated samples, radiation slowed down the growth of the surviving  
443 microorganisms (**Figure 4**). From **Table 7** (columns  $\mu$  regardless of dose), it may be observed  
444 that the proposed model (**Equation 7**) was found to be an adequate method of describing  
445 microbial growth, the percentage of explained variance being higher than 97 % and the mean  
446 relative error lower than 7.5%.

447 The higher the dose applied, the lower the kinetic parameters of the model ( $\mu$  regardless of  
448 dose), which shows how the microorganisms' growth rate was affected by radiation. Radiation

449 was found not only to have an effect on the initial inactivation of bacteria as shown in **Section**  
450 **3.1**, but also on their survival and multiplication rate, leading to a delay in meat spoilage. This  
451 relationship, between  $\mu$  and the radiation dose, was also linear for both the hamburger  
452 (**Equation 10**) and steak (**Equation 11**) samples.

$$453 \quad \mu = -0.196 \cdot d_{irr} + 1.716 ; \quad r^2 = 0.96 \quad (10)$$

$$454 \quad \mu = -0.276 \cdot d_{irr} + 1.529 ; \quad r^2 = 0.93 \quad (11)$$

455 Therefore, from **Equations 10** and **11** it was possible to estimate the  $\mu$  parameter by only  
456 taking the applied radiation dose into account. While the y-intercept was quite similar for both  
457 products (similar growth when no irradiation was applied), the negative slope for steak  
458 samples was 40 % higher than those of the hamburger samples. This indicates that the  
459 increase in the irradiation dose was more effective at slowing down the microorganisms'  
460 growth in steaks than in hamburgers.

461 For the purposes of obtaining a single equation that predicted the growth of microorganisms  
462 at any irradiation dose, **Equations 10** and **11** were combined with **Equation 7**, yielding  
463 **Equation 12** for hamburgers and **Equation 13** for steaks.

$$464 \quad \ln\left(\frac{X_t}{X_0}\right) = (-0.196 \cdot d_{irr} + 1.716) \cdot t \quad (12)$$

$$465 \quad \ln\left(\frac{X_t}{X_0}\right) = (-0.276 \cdot d_{irr} + 1.529) \cdot t \quad (13)$$

466 As can be observed from the %var and MRE (%) (**Table 7**, columns  $\mu$  dependent on dose), the  
467 fit of the model proposed in **Equations 12** and **13** is slightly poorer than the fit provided by  
468 **Equation 7**, where  $\mu$  was fitted for each radiation considered. However, **Equations 12** and **13**  
469 allowed the post-radiation microorganism growth to be predicted in the interval from 0 to 4  
470 kGy for both steaks and hamburgers, only considering the radiation dose as a factor.  
471 Accordingly, this model was used to estimate the shelf-life of meat samples in the optimization  
472 procedure.

#### 473 **3.4. Process optimization.**

474 The aim of the defined optimization problem was to find the radiation dose that provided the  
475 best panelists' scores for appearance and odor and reduced the instrumentally measured color  
476 change in samples after 5 days of storage. The radiation effect on the appearance scores was  
477 described using the Gompertz function, the influence it had on the flavor scores by means of  
478 the Activation-Inactivation model (parameters shown in **Table 4**) and how it affected color



479 changes using chroma (**Table 7**). Therefore, the objective function to be maximized is shown in  
480 **Equation 14**.

$$481 \quad OF = \frac{[C \cdot \exp(-\exp(A + B \cdot d_{irr})) - C \cdot \exp(-\exp(A))] + \left[ \frac{d_{irr}}{k_1 + k_2 \cdot d_{irr}} (1 - \log(1 + \exp(k_3 \cdot d_{irr}))^m) \right]}{p + q \cdot d_{irr}} \quad (14)$$

482  
483 Two constraints to the objective function were considered: first, the radiation dose applied  
484 must produce a log reduction of Salmonellae of 2.7 *D* (**Section 2.8**) and, secondly, during the 5-  
485 day shelf-life of both products, steaks and hamburgers, the bacterial count must be under 5 x  
486 10<sup>7</sup>cfu/g. The effect of the irradiation dose on Salmonellae was modeled through **Equation 1**  
487 using the estimated parameters shown in **Table 1** and the influence on the bacterial growth by  
488 **Equations 12 and 13**, for hamburgers and steaks, respectively and the estimated parameters in  
489 **Table 7**.

490 The optimization results obtained for hamburger samples showed that the optimum radiation  
491 dose was 2.04 kGy. This value was the result of combining the changes in the appearance, odor  
492 and chroma with the increase in radiation dose (**Figure 5A**). The appearance factor exhibited  
493 an increasing trend while odor showed a maximum value at a dose of around 1.1 kGy,  
494 subsequently decreasing the more dose was applied. The chroma value also increased linearly  
495 with the radiation dose applied, which means that the color difference between radiated and  
496 non-radiated samples increased in line with the radiation dose. For that reason, this factor was  
497 placed in the denominator in the objective function. As a result, the objective function showed  
498 a maximum value, representing the optimum irradiation dose which provides the best  
499 combination of sensory and instrumental attributes as considered by **Equation 14**. This  
500 estimated optimum dose of 2.04 kGy was enough to ensure that the microorganism will grow  
501 less (1.2 x 10<sup>6</sup>cfu/g) than the limit considered (5 x 10<sup>7</sup>cfu/g) throughout the shelf-life period in  
502 question (5 days). Furthermore, it was also enough to exceed the FSO for salmonella (2.7*D*),  
503 since the treatment will produce a reduction of 3.95*D* (**Figure 5A**).

504 As for the steaks, the evolution of appearance score (decreased as the irradiation dose  
505 increased) differ from those found in the case of hamburgers, which affected the evolution of  
506 the objective function (**Figure 5B**). Thus, the maximum value of the objective function was  
507 reached at a lower radiation dose, i.e., 0.95 kGy than the obtained for hamburgers. This means  
508 that, above this dose, the sensory features and instrumental color parameters of radiated  
509 samples were worse than the optimum and this fact would negatively affect the consumer  
510 acceptance. This level of dose guarantees that the 5-day shelf-life constraint is achieved, since

511 the number of microorganisms predicted by the model will be  $8.4 \cdot 10^5$ cfu/g, almost 2 log units  
512 lower than  $5 \times 10^7$ cfu/g. However, the Salmonellae number will be reduced by  $2.3D$ , a value  
513 lower than the constraint ( $2.7D$ ), which, in turn, means that it will not be possible to reach the  
514 FSO. As a consequence, it will be necessary to increase the radiation dose in order to achieve  
515 the safety goal. The application of the model provided an optimum radiation dose of 1.11 kGy,  
516 which, as can be observed in **Figure 5B**, was the minimum value necessary to achieve the FSO  
517 for Salmonellae. Therefore, the proposed optimization methodology allowed us to find the  
518 radiation dose that provided the best values of the selected quality factors while, at the same  
519 time, achieving the necessary microbial safety and stability of the product.

#### 520 **4. CONCLUSIONS**

521 The models proposed were an adequate means of describing the effect of a radiation dose on  
522 both the sensory (appearance, odor and flavor) and instrumental (color) attributes of the two  
523 raw chicken products (steaks and hamburgers). In the same way, the influence of radiation on  
524 the inactivation of *Salmonella* spp. and the shelf-life of radiated samples was properly  
525 predicted. From this mathematical modeling and by applying an optimization procedure to the  
526 defined objective function, it was possible to identify an optimum radiation dose that provided  
527 the best quality attribute values but also guaranteed the food's safety and stability  
528 requirements. This procedure can help the process management by permitting objective  
529 decision-making to be adopted.

530

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535

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641

**Table 1.** First order kinetics modeling (Equation 1) of the *Salmonella* spp. inactivation in chicken meat as a function of the radiation dose

Product	Microorganisms	Log ( $N_{d_0}$ )	$k$ (kGy <sup>-1</sup> )	% var	MRE (%)
Steaks	<i>S. Enteritidis</i>	8.23±0.28	2.67±0.15	98.26	12.14
	<i>S. Typhimurium</i>	8.28±0.16	2.43±0.09	99.25	2.99
Hamburgers	<i>S. Enteritidis</i>	9.06±0.24	2.60±0.14	98.55	9.66
	<i>S. Typhimurium</i>	8.79±0.14	1.94±0.07	99.15	4.34

$\log(N_{d_0})$  decimal logarithm of the number of microorganisms (cfu/g) before treatment;  $k$  is the rate constant; %var is the percentage of explained variance by the model; MRE(%) is the mean relative error of the model

**Table 2.** Sensory characteristics of untreated and E-beam radiated chicken breast products (hamburgers and steaks) showing significant differences ( $P < 0.05$ ) in the triangular test just after treatment (0) and after 5 days of storage at 4 °C.

Dose (kGy)	0 <sup>a</sup>	1	2	3	4	
0 <sup>a</sup>		O <sub>5</sub>	A <sub>0</sub> , O <sub>0</sub> , F <sub>0</sub> , A <sub>5</sub> , O <sub>5</sub> ,	A <sub>0</sub> , O <sub>0</sub> , F <sub>0</sub> , A <sub>5</sub> , O <sub>5</sub>	A <sub>0</sub> , O <sub>0</sub> , F <sub>0</sub> , A <sub>5</sub> , O <sub>5</sub>	<b>Hamburgers</b>
1	A <sub>0</sub> , O <sub>0</sub> , O <sub>5</sub>			A <sub>0</sub> , O <sub>0</sub> , F <sub>0</sub> , A <sub>5</sub> , O <sub>5</sub>	A <sub>0</sub> , O <sub>0</sub> , F <sub>0</sub> , A <sub>5</sub> , O <sub>5</sub> , F <sub>5</sub>	
2	A <sub>0</sub> , O <sub>0</sub> , O <sub>5</sub>	O <sub>0</sub>		O <sub>5</sub>	A <sub>0</sub> , O <sub>0</sub> , F <sub>0</sub> , O <sub>5</sub>	
3	A <sub>0</sub> , O <sub>0</sub> , F <sub>0</sub> , O <sub>5</sub>	A <sub>0</sub> , O <sub>0</sub> , F <sub>0</sub>	O <sub>0</sub> , F <sub>0</sub>			
4	A <sub>0</sub> , O <sub>0</sub> , F <sub>0</sub> , O <sub>5</sub>	A <sub>0</sub> , O <sub>0</sub> , F <sub>0</sub> , O <sub>5</sub>	O <sub>0</sub> , F <sub>0</sub> , O <sub>5</sub>			
<b>Steaks</b>						

A<sub>0</sub>, O<sub>0</sub>, F<sub>0</sub>, A<sub>5</sub>, O<sub>5</sub>, F<sub>5</sub>: Significant differences ( $p < 0.05$ ) in appearance (A), odor (O) and flavor (F) after 0 (0) or 5 (5) days of storage at 4 °C

<sup>a</sup> In non-radiated samples, the flavor test was not performed since they were spoiled after 5 days of storage.

**Table 3.** Sensory evaluation by means of the rank order test of untreated and E-beam radiated chicken breast products (hamburgers and steaks) just after treatment (0) and after 5 days of storage at 4 °C.

Dose (kGy)	Hamburgers						Steaks					
	Appearance		Odor		Flavor		Appearance		Odor		Flavor	
	Storage days											
	0	5	0	5	0	5	0	5	0	5	0	5
0	22c $\alpha$	31b $\alpha$	92a $\alpha$	20b $\beta$	92a	NP	34c $\beta$	69a $\alpha$	98a $\alpha$	20c $\beta$	88a	NP
1	43bc $\alpha$	48b $\alpha$	82a $\alpha$	87a $\alpha$	74a $\alpha$	89a $\alpha$	32b $\beta$	65a $\alpha$	78ab $\alpha$	80a $\alpha$	82a $\alpha$	74a $\alpha$
2	62ab $\alpha$	57ab $\alpha$	53b $\beta$	76a $\alpha$	66a $\alpha$	76ab $\alpha$	68a $\alpha$	58a $\alpha$	60b $\alpha$	79a $\alpha$	67a $\alpha$	65a $\alpha$
3	84a $\alpha$	84a $\alpha$	44b $\beta$	74a $\alpha$	38b $\beta$	66b $\alpha$	88a $\alpha$	57a $\beta$	32c $\beta$	70ab $\alpha$	35b $\beta$	66a $\alpha$
4	89a $\alpha$	80a $\alpha$	29b $\alpha$	43b $\alpha$	30b $\alpha$	49b $\alpha$	78a $\alpha$	51a $\beta$	32c $\alpha$	51b $\alpha$	28b $\beta$	75a $\alpha$

Different Latin characters in the same column (a,b,c) or Greek characters in the same row ( $\alpha$ ,  $\beta$ ) for each sensory feature of each chicken product indicates significant differences ( $p < 0.05$ ).

Final scoring =  $(N_1 \times 1) + (N_2 \times 2) + (N_3 \times 3) + (N_4 \times 4) + (N_5 \times 5)$ , where  $N_1$ ,  $N_2$ ,  $N_3$ ,  $N_4$  and  $N_5$  are the number of panelists that ranked the sample in position 1 (the lowest), 2, 3, 4 or 5 (the highest) in the rank order test.

NP, Flavor test was not performed in non-radiated samples since they were spoiled after 5 days of storage. In this case, only the flavor of the irradiated samples was evaluated [Final scoring =  $(N_1 \times 1) + (N_2 \times 2) + (N_3 \times 3) + (N_4 \times 4)$ ].



**Table 4.** Parameters of the models used for describing the effect of radiation on the appearance, odor and flavor of chicken breast products (hamburgers and steaks) after 0 and 5 days of storage at 4 °C.

Attribute	Model	Parameters	Hamburgers		Steaks	
			0 days	5 days	0 days	5 days
Appearance	Gomperzt	A	0.23	0.29	27.01	0.34
		B	-0.89	-0.75	-13.14	-0.24
		C	0.90	0.63	0.39	-0.38
		%var	0.99	0.93	0.98	0.93
Odor	Activation-Inactivation	k <sub>1</sub>	22.33	1.66	55.26	0.32
		k <sub>2</sub>	55.82	-4.80	44.18	0.31
		k <sub>3</sub>	47.98	3.47	48.31	0.67
		m	0.94	-2.72	0.77	0.91
		%var	0.97	0.88	0.88	0.99
Flavor	Activation-Inactivation	k <sub>1</sub>	47.98	0.31	11.57	1·10 <sup>-10</sup>
		k <sub>2</sub>	52.79	0.09	81.60	1·10 <sup>-6</sup>
		k <sub>3</sub>	5.55	0.91	29.85	1.63
		m	0.53	1.06	1.10	1.80
		%var	0.94	0.99	0.92	0.97

A, B and C and k<sub>1</sub>, k<sub>2</sub>, k<sub>3</sub> and m are the parameters of the models relating the effect of the radiation dose on the sensory scores (appearance, odor and flavor). Equations 2 and 3.

**Table 5.** CIE-Lab analysis of untreated and E-beam radiated chicken breast products (hamburgers and steaks) just after treatment (0) and after 5 days of storage at 4 °C.

Chicken breast product	Dose (kGy)	$L^*$		$a^*$		$b^*$	
		Days of storage					
		0	5	0	5	0	5
Hamburger	0	62.5±0.6 a, $\alpha$	62.0±0.8 a, $\alpha$	13.6±0.8 a, $\alpha$	14.7±0.5 a, $\alpha$	8.9±0.7 a, $\alpha$	8.9±0.5 a, $\alpha$
	1	62.2±0.6 a, $\alpha$	62.4±0.6 a, $\alpha$	15.4±0.7 a, $\alpha$	15.5±0.6 a, $\alpha$	9.3±0.7 a, $\alpha$	9.6±0.6 a, $\alpha$
	2	60.7±0.7 a, $\alpha$	62.4±0.9 a, $\alpha$	16.3±0.8 ab, $\alpha$	15.1±0.9 a, $\alpha$	10.2±0.8 ab, $\alpha$	9.0±0.9 a, $\alpha$
	3	63.3±0.7 a, $\alpha$	61.0±0.9 a, $\beta$	17.3±0.7 b, $\alpha$	15.9±0.7 ab, $\beta$	12.0±0.8 b, $\alpha$	9.6±0.4 a, $\beta$
	4	61.6±0.7 a, $\alpha$	62.0±0.8 a, $\alpha$	18.3±0.8 b, $\alpha$	17.4±0.8 b, $\alpha$	12.2±0.8 b, $\alpha$	11.7±0.6 b, $\alpha$
Steak	0	59.2±0.8 a, $\alpha$	57.6±0.6 a, $\beta$	3.2±0.3 a, $\alpha$	2.7±0.5 a, $\alpha$	3.0±0.3 a, $\alpha$	2.3±0.3 a, $\beta$
	1	55.6±0.9 b, $\alpha$	56.4±0.7 ab, $\alpha$	4.0±0.4 a, $\alpha$	3.3±0.4 ab, $\alpha$	3.7±0.3 a, $\alpha$	3.0±0.4 b, $\beta$
	2	53.5±0.8 b, $\alpha$	55.0±0.9 b, $\alpha$	4.3±0.3 ab, $\alpha$	3.9±0.2 b, $\alpha$	4.0±0.6 ab, $\alpha$	3.5±0.5 b, $\beta$
	3	53.5±0.9 b, $\alpha$	55.0±0.9 b, $\alpha$	5.3±0.3 b, $\alpha$	3.7±0.4 b, $\beta$	4.4±0.3 b, $\alpha$	3.6±0.4 b, $\beta$
	4	54.0±0.7 b, $\alpha$	55.0±0.9 b, $\alpha$	5.2±0.3 b, $\alpha$	5.4±0.6 c, $\alpha$	5.1±0.4 b, $\alpha$	4.9±0.3 c, $\alpha$

$L^*$ : lightness,  $a^*$ : redness,  $b^*$ : yellowness.

Different Latin characters in the same column (a,b,c) or Greek characters in the same row ( $\alpha$ ,  $\beta$ ) for each parameter of each chicken product indicates significant differences ( $p < 0.05$ ).

**Table 6.** Linear relationships between the radiation dose and chroma in samples of chicken breast products (hamburgers and steaks).

Meat sample	Days after radiation	p	q	% var	MRE (%)
Hamburger	0	16.393	1.457	99.14	0.74
	5	16.935	0.787	71.26	3.25
Steak	0	4.508	0.738	97.37	2.02
	5	3.506	0.823	89.09	5.26

$p$  and  $q$  are the parameters of the model relating the effect of the radiation dose on chroma (Equation 5); %var is the percentage of explained variance by the model; MRE(%) is the mean relative error of the model

**Table 7.** Parameters of first order kinetics model for microorganism growth in samples of chicken breast products (hamburgers and steaks), treated with different E-beam doses.

Dose (kGy)	Hamburgers						Steaks					
	$\mu$ regardless of dose			$\mu$ dependent on dose			$\mu$ regardless of dose			$\mu$ dependent on dose		
	$\mu$	% var	MRE (%)	$\mu$	% var	MRE (%)	$\mu$	% var	MRE (%)	$\mu$	% var	MRE (%)
<b>0</b>	1.81	99.01	2.81	1.72	98.13	3.20	1.57	99.09	2.31	1.57	98.85	2.53
<b>1</b>	1.45	97.78	4.53	1.50	97.31	4.42	1.08	97.59	4.59	1.22	92.06	7.79
<b>2</b>	1.35	94.63	7.39	1.31	94.41	7.63	1.01	97.01	3.81	0.96	96.39	3.85
<b>3</b>	1.12	98.37	2.87	1.12	98.37	2.87	0.84	98.47	2.20	0.69	87.39	5.63
<b>4</b>	1.01	97.96	3.44	0.92	95.49	4.94	0.31	97.22	1.05	0.41	63.30	5.23

$\mu$  the rate of microorganism growth; %var is the percentage of explained variance by the model; MRE(%) is the mean relative error of the model