Document downloaded from:

http://hdl.handle.net/10251/77345

This paper must be cited as:

Gao, F.; Calatayud Lorente, V.; García-Breijo, F.; Reig Armiñana, J.; Feng, Z. (2016). Effects of elevated ozone on physiological, anatomical and ultrastructural characteristics of four common urban tree species in China. Ecological Indicators. 67:367-379. doi:10.1016/j.ecolind.2016.03.012.



The final publication is available at

https://dx.doi.org/10.1016/j.ecolind.2016.03.012

Copyright Elsevier

Additional Information

- 1 Effects of elevated ozone on physiological, anatomical and ultrastructural
- 2 characteristics of four common urban tree species in China

- 4 Feng Gao<sup>a,1</sup>, Vicent Calatayud<sup>a,b,1</sup>, Francisco García-Breijo<sup>c,d</sup>, José Reig-Armiñana<sup>c</sup>,
- 5 Zhaozhong Feng<sup>a\*</sup>

6

- <sup>a</sup>State Key Laboratory of Urban and Regional Ecology, Research Center for
- 8 Eco-Environmental Sciences, Chinese Academy of Sciences, Shuangqing Road 18,
- 9 Haidian District, Beijing 100085, China
- <sup>b</sup>Fundación CEAM, c/Charles R. Darwin 14, Parque Tecnológico, 46980 Paterna,
- 11 Valencia, Spain
- <sup>c</sup>Laboratorio de Anatomía e Histología Vegetal "Julio Iranzo", Jardín Botánico,
- Universitat de València, c/Quart, 80, 46008 Valencia, Spain.
- de Ecosistemas Agroforestales. ETSIAMN. Universidad Politécnica
- de Valencia. Camino de Vera s/n, 46022-Valencia, Spain.

16

- \*Corresponding author: Zhaozhong Feng. E-mail: fzz@rcees.ac.cn, Tel: +86-10-
- 19 62943823, Fax: +86-10-62943822;
- 20 <sup>1</sup>Both authors contributed equally to this work.

# Abstract

•	,

21

Fast urbanization has led to ozone (O<sub>3</sub>) being the main pollutant in summer in most of 23 China. To assess future ground-level O<sub>3</sub> effects on the service of urban greening 24 species and clarify the underlying mechanism of O<sub>3</sub> damage, four common urban 25 26 greening species of Ailanthus altissima (AA), Fraxinus chinensis (FC), Platanus orientalis (PO) and Robinia pseudoacacia (RP) were exposed to non-filtered air (NF) 27 28 and to elevated O<sub>3</sub> (E-O<sub>3</sub>) in open-top chambers. E-O<sub>3</sub> induced visible injury in all species as well as microscopic alterations such as collapse of the palisade parenchyma 29 cells, callose accumulation, or chloroplast and mitochondrial accelerated senescence. 30 31 E-O<sub>3</sub> significantly reduced light-saturated  $CO_2$  assimilation ( $A_{sat}$ ), the maximum activity of Rubisco ( $Vc_{\text{max}}$ ), the maximum electron transport rate ( $J_{\text{max}}$ ), and 32 33 fluorescence parameters such as the quantum yield of noncyclic electron transport  $(\phi_{PSII})$ , and the quenching of photochemical efficiency of PSII (qP). It also increased 34 total antioxidant capacity, phenolics and ascorbate contents. No significant interaction 35 between O<sub>3</sub> and species was found in photosynthetic performance and antioxidant 36 37 systems, suggesting that the four species selected were sensitive to O<sub>3</sub>. Of all four species, AA was the most sensitive species due to a combination of earlier injury 38 39 onset, anatomical features, lower antioxidant and higher stomatal conductance. The sensitivity of tree species to O<sub>3</sub> is a factor to be considered for urban greening. Ozone 40 may affect important urban forest ecosystem services by reducing CO<sub>2</sub> assimilation. 41

42

43

44

# **Keywords**

Antioxidant system, Ozone, Photosynthesis, Ultrastructure, Urban greening species

45

46

# Capsule

47 Ground-level ozone negatively affects common greening tree species in China

#### 48 Abbreviations

- 49 A = photosynthetic rate
- AA = Ailanthus altissima
- AOT40 = accumulated hourly  $O_3$  concentration over a threshold of 40 ppb during
- 52 daytime
- AsA = ascorbate
- 54  $A_{\text{sat}} = \text{light-saturated photosynthesis}$
- 55 BVOC = biogenic volatile organic compounds
- $C_a$  = ambient  $CO_2$  concentration
- Car = carotenoid
- 58 Chl = chlorophyll
- 59  $C_i$  = intercellular CO<sub>2</sub> concentration
- 60  $F_{\rm v}'/F_{\rm m}' = {\rm actual\ photochemical\ efficiency\ of\ PSII\ in\ the\ saturated\ light}$
- FC = Fraxinus chinensis
- 62  $\phi_{PSII}$  = the quantum yield of noncyclic electron transport.
- 63  $g_s$  = stomatal conductance
- 64  $J_{\text{max}}$  = the maximum rate of electron transport
- LM = light microscopy
- 66 LMA = leaf mass per area
- 67  $L_{\rm s}$  = stomatal limitation to photosynthesis
- 68  $O_3 = ozone$
- 69 OTC = open-top chambers
- 70 PBS = phosphate buffered saline solution
- 71 PO = Platanus orientalis
- RP = Robinia pseudoacacia
- 73 TEM = Transmission Electron Microscopy
- 74 qP = quenching of photochemical efficiency of PSII
- 75  $Vc_{\text{max}}$  = the maximum carboxylation efficiency
- 76 WUE = water use efficiency

#### 1. Introduction

78

Tropospheric ozone (O<sub>3</sub>) levels are of great concern as this pollutant affects human 79 health, ecosystem services and food security besides being a greenhouse gas (The 80 Royal Society, 2008; IPCC, 2013). For sensitive plants, high O<sub>3</sub> concentration is 81 known to induce visible injury, impair photosynthesis, produce reductions in growth 82 and yield, and alter plant interactions with pests and diseases (Krupa et al., 2000). 83 Projected changes of the annual daily mean maximum eight-hour (DM8H) 84 85 surface O<sub>3</sub> concentrations are expected to be in the range of 2 to 8 ppb, -3 to 8 ppb, and -7 to 9 ppb for the 2020s, the 2050s, and the 2090s in summertime for the whole 86 East Asia (Lee et al., 2015). Ozone concentrations in China are rising at a higher rate 87 than in other countries because O<sub>3</sub> precursors (mainly NO<sub>2</sub>) have steadily increased at 88 annual growth rate of 5% caused by its fast industrialization and urbanization (Wang 89 and Mauzerall, 2004; Feng et al., 2015a). In Beijing, the monthly average of peak O<sub>3</sub> 90 concentrations is currently 100 ppb in July, while the AOT40 (accumulated hourly O<sub>3</sub> 91 concentration over a threshold of 40 ppb during daytime) from June to August is 29 92 93 ppm.h (Feng et al., 2015b; Yuan et al., 2015). The yearly average of O<sub>3</sub> concentration in Beijing cities reaches more than 60 ppb during May 2014 to April 2015 (Chen et al., 94 2015), and short-term projected emissions suggest that O<sub>3</sub> concentration will further 95 increase (Yamaji et al. 2008). Therefore, current O<sub>3</sub> levels are, and future O<sub>3</sub> levels are 96 expected to be by far above the threshold value of 40 ppb and AOT40 critical level of 97 5 ppm.h, which have been established to protect sensitive plants against O<sub>3</sub> (LRTAP, 98 99 2010). Cities are characterized by higher levels of pollutant emissions, energy 100 101 consumption and higher temperatures (heat island effect) than surrounding areas. One 102 of the multiple environmental benefits of vegetation is the improvement in air quality 103 (Nowak et al., 2014). In the central part of Beijing, the removal of pollutants by trees was quantified to be 1261.4 tons in 2002, mostly particles (61%), with O<sub>3</sub> accounting 104 105 for 20% (Yang et al., 2005). However, biogenic volatile organic compounds (BVOCs) emitted from vegetation such as isoprene and monoterpenes are precursors of O<sub>3</sub> (The 106

Royal Society, 2008), so the contribution of urban trees to O<sub>3</sub> formation can even 107 offset their removal capacity (Yang et al., 2005). Therefore, it is important to select 108 109 plants with low BVOC emission rates, high pollutant removal capacity and also tolerant to air pollutants when planting trees in cities. The present paper focuses on 110 the latter aspect. 111 In Beijing, O<sub>3</sub> concentrations are high enough to induce visible injury in 112 sensitive species and cultivars, including several ornamental trees (Feng et al., 2014). 113 114 These symptoms are observed in large gardens or urban forests in parks inside the city, or in crop areas or tree plantation surrounding the city rather than in streets where 115 high NO traffic emissions locally scavenge O<sub>3</sub> (due to the titration effect). Ozone 116 effect on plants depends both on the O<sub>3</sub> dose entering the plant through the stomata 117 which is directly related to water vapor stomatal conductance  $(g_s)$ , and also on their 118 defense ability to cope with oxidative stress (Matyssek et al., 2007; Paoletti et al., 119 2008). On the other hand, leaf functional traits are considered to play a role in O<sub>3</sub> 120 sensitivity, e.g. plants with higher leaf mass per area (LMA), or higher thickness or 121 122 density of mesophyll tissues being tolerant to O<sub>3</sub> (Bussotti, 2008; Zhang et al., 2012). However, information on the effects of current and predicted future O<sub>3</sub> levels on 123 urban greening species is still very scarce in China. In the present study, we exposed 124 four commonly planted urban greening tree species to elevated O<sub>3</sub> level which is the 125 representative of a future scenario by 2050 on the basis of annual increase rate of 0.73 126 ppb/year at Shangdianzi station observation nearby Beijing city (Dr. XB Xu, personal 127 communication) and an increase rate of 0.5-2% at a global scale (Vingarzan, 2004). 128 The four species are the tree of heaven (Ailanthus altissima (Mill.) Swingle, AA), the 129 130 Chinese ash (Fraxinus chinensis Roxb., FC), the American sycamore (Platanus orientalis L., PO) and the black locust (Robinia pseudoacacia L., RP). Three of them 131 (AA, FC, RP) are regarded as O<sub>3</sub> sensitive. This study tests the following two 132 hypotheses: (1) O<sub>3</sub> sensitivity differs among investigated species, considering 133 134 anatomical and ultrastructural changes, photosynthetic performance and antioxidant

systems; (2) plants with a higher stomatal conductance, lower antioxidant capacity 135 and thinner leaves are more sensitive to  $O_3$ . 136 137 2. Materials and methods 138 2.1. Plant materials 139 One-year-old seedlings of AA, FC, PO and RP were obtained from a commercial 140 nursery near the experimental site. Bare rooted seedlings were planted in 20 L circular 141 142 plastic pots on 31 March 2013 and grown at ambient field condition. Pots were filled with native light loamy soil (pH 7.96, Organic C 14.7 g/kg; total N 1.64 g/kg, 143 available P 6.59 mg/kg, available K 139.8 mg/kg) randomly selected from a nearby 144 farmland. Plants with similar height and basal diameter were selected. Ten days 145 before O<sub>3</sub> fumigation, they were pre-adapted to open-top chamber (OTC, octagonal 146 base, 12.5 m<sup>2</sup> of growth space with a diameter of 4 m, and 3.0 m in height). All plants 147 were watered at field capacity at 1-3 day intervals to avoid water stress. Solid, 148 slow-release fertilizer (N/P/K = 17,17,17) was applied at a rate of 300 kg ha<sup>-1</sup> to each 149 150 plant at July during the experiment. 151 2.2.  $O_3$  treatments 152 The experiment was carried out at Changping (40°19′N, 116°13′E), Northwest 153 Beijing. The area has a semi-humid continental climate, with a yearly precipitation of 154 550 mm, and an annual mean temperature of 11.8 °C. Plants were exposed to two O<sub>3</sub> 155 treatments in OTCs for four and a half months (from 1 June to 15 October): 156 non-filtered ambient air (NF, averaged O<sub>3</sub> concentration of 42 ppb from 09:00 to 157 18:00), and NF supplied with 40 ppb of O<sub>3</sub> (E-O<sub>3</sub>, averaged O<sub>3</sub> concentration of 69 158 ppb from 09:00 to 18:00). The four species and two O<sub>3</sub> treatments were selected for 159 the present study from a wider investigation involving a total of 10 species and six 160 different O<sub>3</sub> regimes in six OTCs. Positional effects were avoided by changing plant 161 positions within each OTC weekly, and by switching them randomly among six OTCs 162 monthly (Feng et al., 2011a). For each O<sub>3</sub> treatment, 4-6 plant replicates were used for

each species. Ozone was generated from pure oxygen using an O<sub>3</sub> generator (HY003, 164 Chuangcheng Co., Jinan, China), mixed with ambient air and then piped into OTCs 165 through a PVC tube (11 cm in diameter) using a fan (1.1 kW, 1080 Pa, 19 m<sup>3</sup> min<sup>-1</sup>, 166 CZR, Fengda, China). The flow rate of pure oxygen was regulated by mass flow 167 controllers so as to achieve the target O<sub>3</sub> concentration at the top of the canopy in the 168 fumigation treatments. An O<sub>3</sub> analyzer (Model 49i-Thermo, USA) was used to 169 continuously monitor O<sub>3</sub> concentrations inside the OTCs via a Teflon solenoid valve 170 171 switch system connected to a set of Teflon tubes (4 mm in diameter), which collected air from sampling points at approximately 10 cm above the plant canopy in each 172 chamber. The monitors were calibrated by a 49i-PS calibrator (Thermo Scientific, 173 USA) before the experiment and once a month during the experiment. The daily 174 maximum fumigation period was 9 h (from 09:00 to 18:00) through a fan running 175 when there was no rain, fog, mist, or dew, according to the protocols in free air O<sub>3</sub> 176 concentration enrichment system (Feng et al., 2011b). The monthly ambient O<sub>3</sub> 177 concentration (from 09:00 to 18:00) in the open field was ranged from 43 ppb 178 179 (September) to 69 ppb (June), with the highest one hour peak being 153 ppb (at 16 h on 20 September). 180 181 2.3. Visible injury 182 Visible injury was assessed weekly in all plants (4-6 per O<sub>3</sub> treatment). The 183 percentage of injured leaves (for PO, with simple leaves) or leaflets (for the rest of 184 species, with composite leaves) per plant was scored, in order to classify each plant 185 according to the following classes: 0, no leaves injured;  $1, \le 1\%$  leaves injured; 2, >1%186  $-\le10\%$  leaves injured; 3, >10%  $-\le50\%$  leaves injured; 4, >50% leaves injured. 187 188 2.4. Leaf traits 189 Twenty-five asymptomatic mature leaves from NF plants were collected studying the 190 leaf traits using a scanner and ImageJ software (Gao et al., 2011). LMA was 191

calculated as dry mass (mg) / leaf surface area (cm<sup>2</sup>). Leaf dry mass was determined

by oven-drying leaves at 60 °C for 40 h until steady weight. To estimate the leaf traits, 193 paraffin-embedded sections of six leaves or leaflets (five sections per leaf or leaflet, 194 and five measurements made per section) were examined under the microscope 195 following the methods described below. 196 197 198 2.5. Microscopy examinations To study the effects of  $O_3$ , six symptomatic and six asymptomatic leaves or leaflets 199 200 from the upper canopy layer (6-8th fully expanded leaves from the top) of three plants for each species were collected in August from E-O<sub>3</sub> and NF plants, respectively, 201 before the onset of senescence. Samples were fixed in 2% Karnovsky fixative for 8 h 202 at 4°C, then washed three times for 15 min with 0.01 M PBS (pH 7.4). For 203 microscopy examinations, leaf portions were submitted to freeze-cut, paraffin- and 204 resin-embedded sections. For freeze-cut sections (~30 µm), a freezing microtome 205 (CM 1325; Leica, Germany) was used, paraffin-embedded sections (~10 µm) were 206 cut with an Anglia Scientific microtome, and Spurr's resin-embedded samplers were 207 208 cut with a diamond knife (DIATOME Histo 45°) and an ultramicrotome (Ultratome Nova LKB Bromma) (~1.5 µm). Freeze-cut sections were observed under 209 epifluorescence BV (autofluorescence) or stained with aniline blue and observed 210 under epifluorescence UV to detect callose depositions. For fluorescence microscopy, 211 an Olympus U-ULS 100 HG epifluorescence system with U-MWU (excitation filter 212 330–385 nm, dichroic mirror 400 nm, barrier filter 420 nm) and U-MWBV (excitation 213 filter 400-440 nm, dichroic mirror 455 nm, barrier filter 475 nm) cubes was used. 214 Paraffin-embedded sections were stained with safranin-fast green, or with trichromic 215 216 FSA for observation of the different structures and for identifying their composition. Semi-thin sections were stained with toluidine blue. Pectinaceous drops were detected 217 with this stain. All light microscope (LM) observations were carried out by an 218 Olympus Provis AX 70 fluorescence microscope equipped with an Infinity 2-3C 219 Lumenera® digital camera and analyzed with "Infinity Analyze" Software v.6.4.1. 220

For transmission electron microscopy (TEM), samples were fixed as LM, then

222 washed three times with 0.02 M PBS (pH 7.4) for 15 min and fixed again with 2% osmium tetroxide (OsO<sub>4</sub>) in 0.01 M PBS (pH 7.4) for 2 h at room temperature. After 223 washing in buffer, samples were dehydrated and embedded as indicated for LM. 224 Ultrathin sections 80 nm thick were made with a diamond knife (mod. DIATOME 225 ultra 458; DIATOME, Hatfield, PA, USA), mounted on copper grids of 100 mesh, 226 and then stained with 10 % uranyl acetate and 0.1% lead citrate using the 'Synaptek 227 Grid-Stick Kit' (EMS; 228 229 http://www.ems-diasum.com/microscopy/technical/datasheet/71175.aspx ). Sections were observed at 80 kV under the JEOL JEM-1010 microscope (JEOL USA Inc., 230 Peabody, MA, USA). Images were obtained using an Olympus MegaView III camera 231 and processed by Olympus analysis getIT software (Olympus Corp., Japan). In total, 232 40–50 root segments were examined through TEM. 233 234 2.6. Leaf gas exchange and chlorophyll a fluorescence measurements 235 Two upper canopy leaves (6-8th fully expanded leaves from the top) from each plant, 236 237 i.e. eight leaves from four plants for each species, were selected randomly. Gas exchange and chlorophyll a fluorescence were determined simultaneously from 9:00 238 to 11:00 at both August and September using a LiCor-6400 photosynthesis system 239 (LICOR, Lincoln, NE, USA) fitted with a 6400-40 leaf chamber fluorometer (LCF). 240 During the measurements, photosynthetic active radiation (PAR) was set at 1200 241  $\mu$ mol m<sup>-2</sup> s<sup>-1</sup>, CO<sub>2</sub> levels at 380 ppm, block temperature at 32  $\pm$  0.5 °C and relative 242 humidity between 50% and 70%. Fluorescence parameters include actual 243 photochemical efficiency of PSII in the saturated light  $(F_v'/F_m')$ , quenching of 244 photochemical efficiency of PSII (qP), and the quantum yield of noncyclic electron 245 transport ( $\phi_{PSII}$ ). Water Use Efficiency (WUE) was calculated as the ratio between 246 light-saturated photosyntehsis rate ( $A_{sat}$ ) and transpiration rate. 247 During the experiment, photosynthesis and intercellular  $CO_2$  concentration (A/Ci) 248 curves were performed in four plants (one leaf per plant, selected from the 6-8th fully 249 expanded leaves from the top) for either O<sub>3</sub> treatment, using the automatic program in 250

the LiCor-6400 photosynthesis system. When the stomatal conductance  $(g_s)$  reached 251 equilibrium, subsequent measurements were made following step-wise changes in 252 reference CO<sub>2</sub> concentration: 380, 300, 200, 100, 50, 400, 600, 900, 1200, 1500 ppm, 253 under PAR of 1200  $\mu$ mol m<sup>-2</sup> s<sup>-1</sup>, block temperature of 32  $\pm$  0.5 °C and relative 254 humidity of 50-70%. The maximum carboxylation efficiency ( $Vc_{max}$ ) and the 255 maximum rate of electron transport  $(J_{\text{max}})$  were derived by iteratively fitting curves to 256 A/Ci response data according to the program of Sharkey et al. (2007). Stomatal 257 258 limitation ( $L_s$ ) was calculated as  $L_s = 1 - A_{Ca380}/A_{Ci380}$ , where  $A_{Ca380}$  and  $A_{Ci380}$  represent net  $CO_2$  assimilation rate at ambient  $CO_2$  concentration (Ca) of 380 ppm and at Ci = 259 380 ppm, respectively. 260 261 2.7 Pigment and antioxidant content 262 Those leaves (6-8th fully expanded leaves from the top) for photosynthesis 263 measurements were used again for pigment and antioxidant assays in both NF and 264 E-O<sub>3</sub> plants in September. Leaf discs (~10 mm diameter) in two leaves per plant were 265 266 plunged from 4-5 plants of each species between 11:30 and 12:30, wrapped tinfoil, frozen in liquid nitrogen, and stored at -80°C until physiological and biochemical 267 analysis. 268 269 2.7.1. Pigments content 270 The pigment content of two leaf discs (~10 mm diameter) from two leaves per plant 271 in either O<sub>3</sub> treatment was extracted with 4 ml 95% ethanol in the dark for 72 h at 4°C. 272 The absorbance of leaf pigment extracts was measured at 646 nm, 663 nm and 470 273 274 nm. Total chlorophyll (Chl) and carotenoid (Car) contents were calculated according to the specific absorption coefficients provided by Lichtenthaler (1987). 275

- 277 2.7.2. Ascorbate (AsA) content
- 278 Two leaf discs from two leaves per plant (~30 mg) were homogenized in a pre-chilled
- mortar and extracted in 2 ml ice-cold 6% (w/v) trichloroacetic acid, and then

centrifugated at 13,000 g for 5 min at 4°C. Reduced and total ascorbate (AsA) contents were determined following the protocol of Gillespie and Ainsworth (2007).

282

280

281

- 283 2.7.3. Total antioxidant capacity (TAC) and total phenolics content
- For total antioxidant capacity (TAC), 2 ml cold 70% (v/v) ethanol was added to leaf
- 285 disc samples (~ 30 mg) and homogenized in darkness. The mixture was centrifuged at
- 286 13,000 g for 10 min. Subsequently, 0.1 ml of supernatant was taken for the ferric
- reducing antioxidant power (FRAP) assay to express the total antioxidant activity. The
- 288 principle of the assay was conducted following the method of Benzie and Strain
- 289 (1996), expressed as  $Fe^{3+}$  equivalents (mmol  $Fe^{2+}/g$  fresh mass).
- For phenolics content, 2 ml cold 95% (v/v) methanol was added to leaf disc
- samples (~ 30 mg). The mixture was incubated for 48 h in darkness, then centrifuged
- 292 at 13,000 g for 10 min in darkness. 0.1 ml of supernatant was taken for total phenolics
- analysis, using Folin-Ciocalteau reagent as described by Ainsworth & Gillespie
- 294 (2007), and expressed as mg of gallic acid equivalents per unit of fresh mass.

- 296 2.8. Statistical analyses
- In this study, each plant was treated as the statistical unit, i.e. plant means were firstly
- obtained and statistical analyses were applied subsequently. In order to test the effects
- of O<sub>3</sub>, species, sampling date and their interactions, the data of each dependent
- variable was subjected to the analysis of variance with a Mixed Lineal Model using
- JMP software (SAS Institute, USA). Except reduced ASC, TAC and qP with square
- root transformation, all variables in the original data passed the homogeneity test of
- variance. In case of significant interactions between  $O_3$  and species, differences
- among species in combination with O<sub>3</sub> treatment were tested with an ANOVA
- followed by Tukey's HSD. Otherwise, Student *t*-test was applied to compare the
- difference between  $O_3$  treatments for any variable at any sampling date.  $P \le 0.05$  was
- considered statistically significant. Data shown in Tables and Figures are means  $\pm$  SE
- 308 (n = 4-6, excluding Table 1 with n = 6-25).

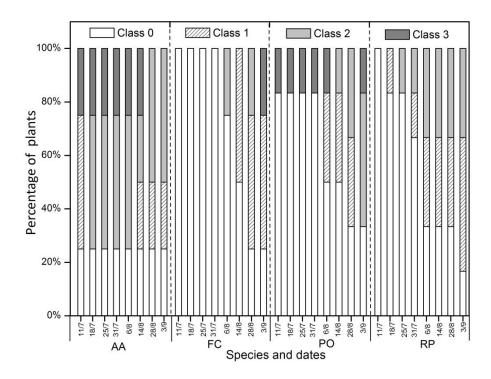
# 3. Results 309 3.1. Leaf traits 310 Three species (AA, FC and RP) have composite leaves, and the other one (PO) has 311 312 large single leaves. LMA was on average higher in AA and FC, and lower in PO and RP. AA was the species showing the smallest palisade parenchyma. Upper epidermis 313 was thicker in AA and PO than in the other two species, and mesophyll thickness 314 ranked as FC and PO > RP and AA (Table 1). 315 316 317

Table 1. Leaf traits of the four species. For AA, FC and RP with composite leaves, measurements are based on leaflets. Different letter indicates significant difference between species.

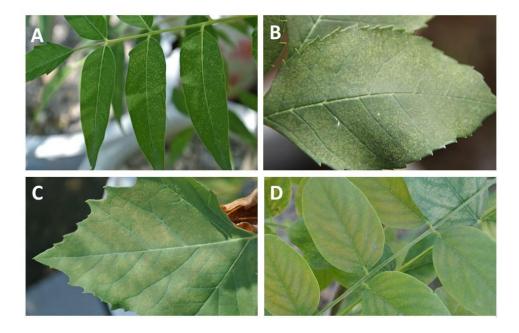
	Units	AA	FC	PO	RP
Length	cm	$16.7 \pm 0.3a$	$10.1 \pm 0.2b$	$15.9 \pm 0.7a$	$5.3 \pm 0.1c$
Width	cm	$4.7 \pm 0.1b$	$4.5 \pm 0.1b$	$18.6 \pm 1.4a$	$2.3 \pm 0.0c$
Perimeter	cm	$38.1 \pm 0.7b$	$23.3 \pm 0.4c$	$82.0 \pm 5.8a$	$12.6 \pm 0.1$ d
Leaf area	$cm^2$	$56.2 \pm 1.8b$	$30.8 \pm 1.1c$	$231.7 \pm 21.4a$	$10.1 \pm 0.2d$
Leaf mass per area	mg cm <sup>-2</sup>	$9.1 \pm 0.9a$	$8.8 \pm 0.2a$	$5.6 \pm 0.2b$	$5.0 \pm 0.3$ b
Upper epidermis	μm	$19.5 \pm 0.2a$	$10.2\pm0.2c$	$20.3 \pm 0.6a$	$13.4 \pm 0.4b$
Lower epidermis	μm	$10.1 \pm 0.2b$	$10.4\pm0.2b$	$15.0 \pm 0.5a$	$11.3 \pm 0.3b$
Palisade parenchyma					
thickness	μm	$49.0 \pm 1.6c$	$68.5 \pm 0.9a$	$71.2 \pm 2.1a$	$60.8 \pm 0.7$ b
Spongy parenchyma					
thickness	μm iickness		$72.7 \pm 2.5a$	68.4±1.9a	$43.7 \pm 1.2b$
M 1 H 4 ' 1	s μm	$99.0 \pm 2.7$ b	141.3 ±	120 6 . 2 6	104.4 ±
Mesophyll thickness			3.2a	$139.6 \pm 3.6a$	1.2b

# 3.2. Visible injury

No visible injury was observed in any of the plants in NF treatment (control). The onset of O<sub>3</sub> symptoms in E-O<sub>3</sub> was at the following dates: AA at 04/07/2013, PO at 11/07/2013, RP at 18/07/2013 and FC at 06/08/2013, and the corresponding AOT40 values were 12.8, 14.2, 16.2, 23.2 ppm. h, respectively. From Fig. 1, AA was the species showing symptoms earliest, and was also the most severely affected from the beginning, although towards the end of the experiment the percentage of injured leaves was reduced due to the development of new, not injured leaves. Photos of E-O<sub>3</sub>-induced visible injury are provided in Fig. 2.



**Figure 1**. Percentage of plants belonging to different classes. Classes: 0, no leaves injured; 1,  $\leq$ 1% leaves injured; 2, >1%  $-\leq$ 10% leaves injured; 3, >10%  $-\leq$ 50% leaves injured.



**Figure 2**. Visible symptoms in (**A**) *Ailanthus altissima*. (**B**) *Fraxinus chinensis*. (**C**) *Platanus orientalis*. (**D**) *Robinia pseudoacacia*.

341

342

343

344

345

346

347

348

349

350

351

352

353

354

355

356

357

358

359

360

361

362

363

364

365

366

367

368

3.3 Anatomical and ultrastructural changes induced by E-O<sub>3</sub>

LM and TEM studies showed the effects of O<sub>3</sub> as well as plant responses against oxidative stress. Palisade parenchyma cells were in general the most affected part of the mesophyll (AA > PO > FC > RP; see Fig. 3). The middle lamella of the cell wall was degraded (Figs 4A, 5B) and small pectinaceous drops produced by its degradation were observed (Figs. 4A, 5A, 5E, 5F, 6A). Cell walls were progressively altered (Figs. 3C,3F, 4B, 4C, 5B, 5C, 7C, 7E), and fluorescence LM (Fig. 3A) and TEM (Figs. 5A, 5B, 6B) showed that callose was accumulated between the membrane and cell wall, especially in AA and PO. Inside the cells, vacuole content becomes altered. In AA and RP, vacuolar content of the affected cells became denser by the accumulation of tannins (Figs. 3B, 3H), and in PO, the normal cells of which have vacuoles with abundant tannic content, a gradual coagulation of tannins in the affected cells was observed (Figs. 5E, 5F), as well as a large accumulations of crystals in their cytoplasm (Figs. 5D, 5D, 6C). Finally, tonoplast broke and cells lost turgor (Fig. 7B). In a later stage, cells collapsed (Figs. 3, 4A, 4C, 5B, 5E, 5F, 7E) leading to an increase of intercellular spaces (AA, PO > FC > RP). Chloroplasts were also strongly affected in all species, increasing electrodense material and changing their shape. Accumulation of big starch grains (Figs. 4A, 5B, 5C, 7A), abundant plastoglobuli (Figs. 4A, 7A, 7B, 7C, 7E), and lipid-protein bodies (especially in AA, Figs. 4A, 4B, 4D) were also observed. Finally, thylakoid membranes were partly or totally disaggregated (Fig. 7B). Mitochondria also experienced degradation processes. In some cases, the accumulation of lipid droplets similar to plastoglobuli was observed, especially in AA (Fig. 4D). Similar but much less conspicuous changes were observed in the spongy parenchyma. Upper and lower epidermises were not distinctly affected in any of the species with an exception of AA and RP, in which some epidermal cells can collapse (Figs. 3A, 3G, 4C). In some cases, chloroplast of the guard cells of stomata were also affected, as indicated by large starch accumulations (Fig. 7A). In the vascular bundles, xylem was never affected but

phloem cells can experience changes in shape and callose deposition may increase in the sieve tubes (Figs. 3H, 6E, 7D).

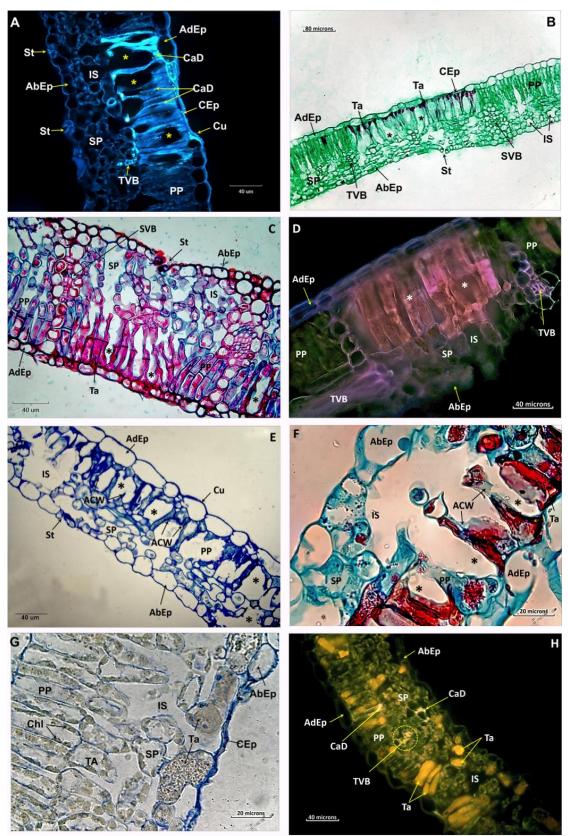


Figure 3. (A) Fluorescence micrograph (UV) of a cross section of Ailanthus

altissima leaf stained with aniline blue showing an affected area (asterisks) with numerous collapsed cells and with callose depositions (CaD) on its cell walls. (B) Micrograph of a cross section of an Ailanthus altissima affected leaf stained with safranin-fast green showing numerous collapsed cells (asterisks) with tannic (Ta) contents inside. (C) Micrograph of a cross section of a Fraxinus chinesis affected leaf stained with trichromic FSA showing numerous collapsed cells (asterisks) with tannic (Ta) contents inside. (D) Autofluorescence micrograph (BV) of a cross section of Fraxinus chinensis leaf showing an affected area (asterisks) of palisade parenchyma (PP) without chlorophylls. (E) Micrograph of a semithin cross section of a *Platanus* orientalis affected leaf stained with toluidine blue. Numerous collapsed cells (asterisks) leaving between them large intercellular spaces (IS) are observed. (F) Micrograph of a cross section of a Platanus orientalis affected leaf stained with trichromic FSA showing collapsed cells (asterisks) with affected cell wall (ACW) and tannic (Ta) contents inside. (G) Micrograph of a semithin cross section of a Robinia pseudoacacia affected leaf stained with toluidine blue. A collapsed epidermis (CEp) is observed. (H) Autofluorescence micrograph (BV) of a cross section of a Robinia pseudoacacia affected leaf. Numerous cells filled with tannin content (Ta) are observed. Others abbreviations. AbEp: abaxial epidermis; AdEp: Adaxial epidermis; Cu: cuticle; TVB: tertiary vascular bundle; SP: spongy parenchyma; St: stomata; **SVB**: secondary vascular bundle.

373

374

375

376

377

378

379

380

381

382

383

384

385

386

387

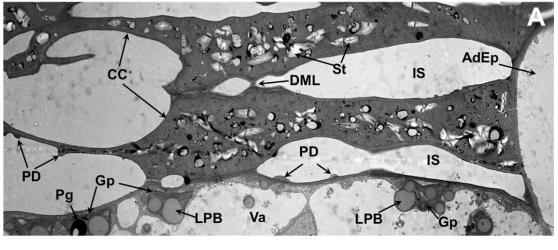
388

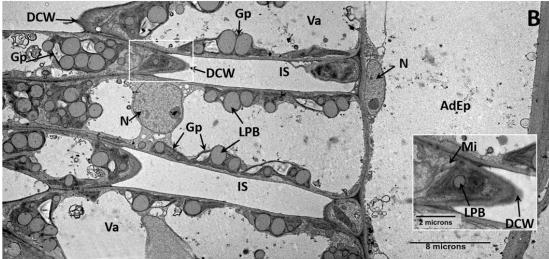
389

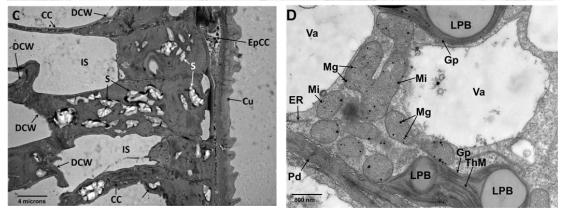
390

391

392







**Figure 4.** Details of TEM micrographs of cross sections of *Ailanthus altissima* leaves.

(A) Two collapsed cells (CC) of palisade parenchyma with great intercellular spaces

(IS). Pectinacious drops (PD) and degraded middle lamella (DML) are observed.

(B) Cells of palisade parenchyma with abundant gerontoplasts (Gp) filled with abundant lipid-protein bodies (LPB), degraded cell wall (DCW) and great intercellular spaces (IS). The area marked with a white frame is shown magnified at

the bottom right of the image. (C) Collapsed cells (CC) of palisade parenchyma and adaxial epidermis (EpCC) showing a degraded cell wall (DCW). (D) Detail of a cell with several mitochondria (Mi) containing some mitoglobuli (Mg). Other abbreviations. AdEp: adaxial epidermis; N: nucleus; Pd: plasmodesma; Pg: plastoglobuli; St: starch; ThM: thylakoidal membranes; Va: Vacuole.

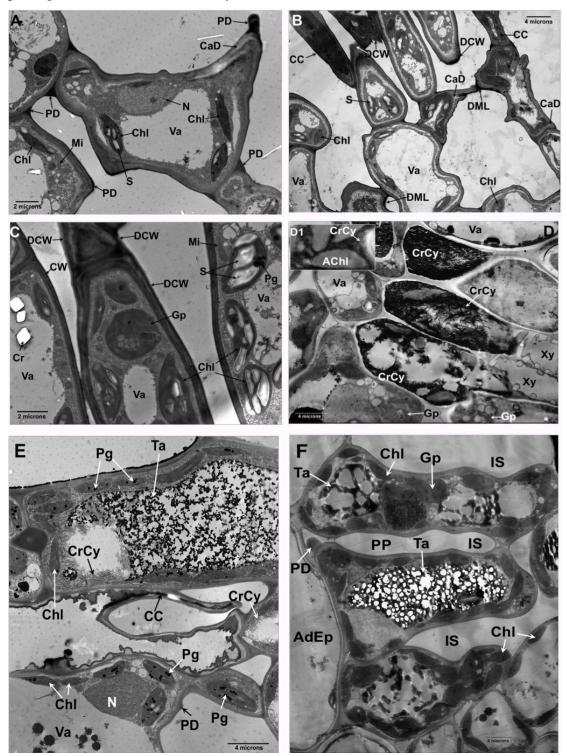


Figure 5. A to B: Details of TEM micrographs of cross sections of *Fraxinus chinensis* leaves. **D** to **F**: Details of TEM micrographs of cross sections of *Platanus orientalis* leaves. (A) Spongy parenchyma cell showing an altered medium lamella. Some pertinacious drops (PD) and callose depositions (CaD) between the cell membrane and cell wall are observed. (B) Cells of spongy and palisade parenchyma showing different alterations. Some of them are collapsed (CC). (C) Detail of palisade parenchyma cells. Degraded cell wall (DCW) and gerontoplasts (Gp) are observed. (D) Cells showing a crystalized cytoplasm (CrCy). In (D1) a detail of this crystallization is observed. (E) Affected palisade parenchyma cells. Collapsed cells (CC), vacuoles with tannins (Ta), crystalized cytoplasm (CrCy) and numerous chloroplasts (Chl) with plastoglobuli (Pg) are observed. (F) Cells of palisade parenchyma showing varying degrees of decomposition of tannins (Ta) within the vacuoles (Va). Other abbreviations. AChl: Altered chloroplast; AdEp: adaxial epidermis; Cr: (calcium oxalate) crystal; DML: degraded medium lamella; N: nucleus; S: starch; Xy: xylem.

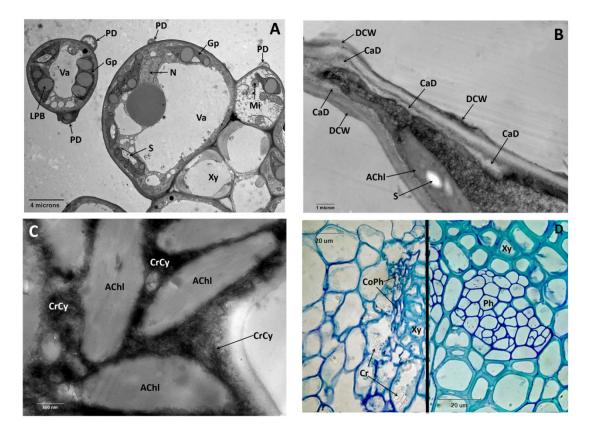
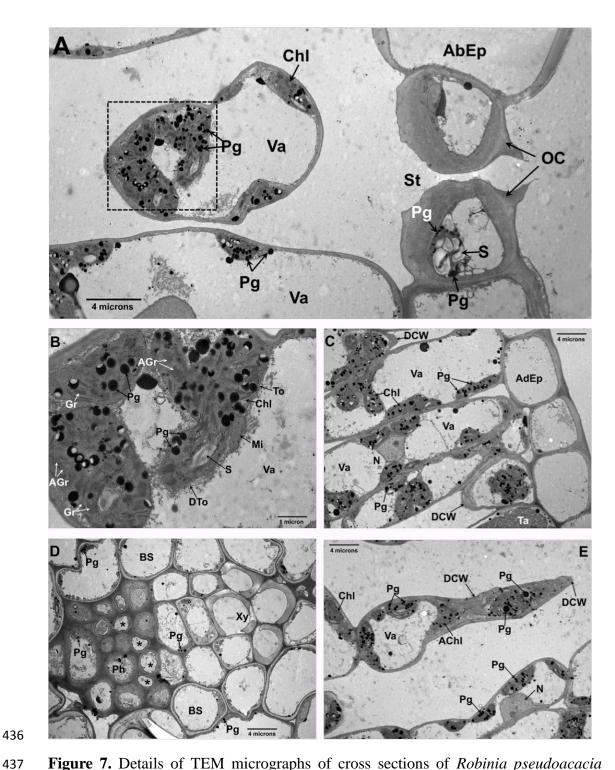


Figure 6. (A). TEM micrograph of cross sections of *Platanus orientalis* leaf. Detail of spongy parenchyma cells showing pertinacious drops (PD). (B). TEM micrograph of cross sections of *Platanus orientalis* leaf. Callose depositions (CaD) between the cell membrane and cell wall are observed. (C). TEM micrograph of cross sections of *Platanus orientalis* leaf. Cell showing a crystalized cytoplasm (CrCy). (D). Micrograph of a cross section of a *Fraxinus chinesis* affected leaf (left) and a control leaf (right) stained with toluidine blue. The affected leaf shows a collapsed phloem (CoPh). Other abbreviations. AChl: altered chloroplast; Cr: crystals; CrCy: crystalized cytoplasm; DCW: degraded cell wall; Gp: gerontoplast; LPB: lipid-protein body; Mi: mitochondria; N: nucleus; Ph: Phloem; S: starch Va: Vacuole; Xy: xylem.



**Figure 7.** Details of TEM micrographs of cross sections of *Robinia pseudoacacia* leaves. **(A)** Detail of a stoma **(ST)** with affected chloroplast and spongy parenchyma. Chloroplast **(Chl)** in cells of spongy parenchyma are affected and contain numerous plastoglobuli **(Pg)**. The area marked with the dashed square is shown in Figure **(B)**. **(B)** Detail of affected chloroplasts containing numerous plastoglobuli and many grana disorganized **(AGr)**. The tonoplast **(To)** is disintegrated into many areas **(DTo)**. **(C)** 

Palisade parenchyma cells containing chloroplasts with numerous plastoglobuli and degraded cell walls (**DCW**) in some areas. (**D**) Detail of a tertiary vascular bundle with the affected phloem. Callose depositions (**asterisks**) are observed in some phloem (**Ph**) vessels. (**E**) Cells affected in the spongy parenchyma. Their cell walls are degraded (**DCW**) and contain chloroplasts with numerous plastoglobuli (**Pg**). Other abbreviations. **AbEp**: abaxial epidermis; **AdEp**: Adaxial epidermis; **BS**: Bundle sheath; **Gr**: grana; **Mi**: mitochondria; **N**: nucleus; **OC**: occlusive cells; **S**: starch; **Ta**: tannins; **Va**: vacuole; **Xy**: xylem.

3.4. Leaf gas exchange and chlorophyll a fluorescence

Effects of E-O<sub>3</sub> on gas exchange and fluorescence parameters were significant for  $A_{\text{sat}}$ ,  $\phi_{PSII}$ , and qP (Table 2). There were no significant interactions between species and O<sub>3</sub>, suggesting the responses to E-O<sub>3</sub> were the same among all species (Table 2). Effects on  $A_{\text{sat}}$  increased with  $O_3$  exposure, as indicated that there was a significant reduction in A<sub>sat</sub> by 27%, 21%, 17%, and 31% for AA, FC, PO and RP, respectively, in September (Tables 2 and 3). For the same month  $\phi_{PSII}$  was significantly reduced by 13% and 21% in PO and RP, respectively (Table 3). C<sub>i</sub> and WUE were not significantly affected by  $O_3$ , while  $g_s$  was significantly reduced in AA and RP in September. Notably, there was significant difference among species in  $g_s$  (Table 2), as shown by highest  $g_s$  values in AA (0.15 mol m<sup>-2</sup> s<sup>-1</sup>) and lowest in RP (0.07 mol m<sup>-2</sup> s<sup>-1</sup>), with FC and PO showing intermediate values (0.13 mol m<sup>-2</sup> s<sup>-1</sup>). 

**Table 2**. Analysis of variance of the effects of  $O_3$ , species and sampling date, and their interactions on gas exchange and chlorophyll a fluorescence parameters, pigment and antioxidant contents.

						O <sub>3</sub> ×Date
	$O_3$	Species	O <sub>3</sub> ×Species	Date	$O_3 \times Date$	×Species
$A_{sat}$	0.0002	< 0.0001	0.9983	0.2049	0.0096	0.8472
$g_s$	0.5411	0.0002	0.5788	0.8176	0.0096	0.2809
$C_{\rm i}$	0.7889	< 0.0001	0.2548	0.7265	0.3888	0.3006
WUE	0.9437	0.1959	0.5503	< 0.0001	0.3258	0.3105
<i>F</i> v'/ <i>F</i> m'	0.6297	0.0002	0.0773	0.1689	0.3076	0.8374
$\phi$ PSII	0.0427	< 0.0001	0.5481	< 0.0001	0.4844	0.7984
qP	0.0033	0.0065	0.6313	< 0.0001	0.7380	0.3661
$Vc_{max}$	< 0.0001	0.4209	0.5318	0.0337	0.8005	0.2668
$J_{\it max}$	< 0.0001	0.0896	0.3419	0.1311	0.9928	0.3479
$L_s$	0.0006	0.2542	0.4295	0.2473	0.0148	0.9018
TAC	< 0.0001	< 0.0001	0.0965			
Phenolics	< 0.0001	< 0.0001	0.7386			
Total AsA	< 0.0001	< 0.0001	0.0908			
Reduced AsA	< 0.0001	< 0.0001	0.0799			
Chl a	0.0825	0.0991	0.8684			
Chl b	0.081	0.0403	0.9558			
Chl a+b	0.0815	0.0822	0.891			
Car	0.0715	0.0183	0.7867			
Chla/Chlb	0.2442	0.0200	0.816			

**Table 3**. Gas exchange and chlorophyll a fluorescence variables in plants grown in non-filtered air (NF) and in elevated  $O_3$  (E- $O_3$ ) treatments in August and September. Different letter indicates significant difference between  $O_3$  treatments.

			$A_{\rm sat}$	$g_{\rm s}$	$C_{ m i}$	WUE	$F_{\rm v}$ '/ $F_{\rm m}$ '	$\phi_{ m PSII}$	qP
						(µmol CO <sub>2</sub>			
			$(\mu mol m^{-2} s^{-1})$	$(\text{mol m}^{-2} \text{ s}^{-1})$	(ppm)	/mmol H <sub>2</sub> O)			
Aug.	AA	NF	$11.33 \pm 1.28$	$0.14 \pm 0.03$	$236.32 \pm 15.13$	$3.79 \pm 0.33$	$0.38 \pm 0.03$	$0.22 \pm 0.03$	$0.56 \pm 0.03$
		$E-O_3$	$11.33 \pm 1.05$	$0.23 \pm 0.07$	$276.32 \pm 27.95$	$2.84 \pm 0.62$	$0.46 \pm 0.01$	$0.22 \pm 0.02$	$0.47 \pm 0.05$
	FC	NF	$12.79 \pm 1.30$	$0.12 \pm 0.02$	$205.87 \pm 13.83$	$3.60 \pm 0.26$	$0.44 \pm 0.01$	$0.28 \pm 0.01$	$0.62 \pm 0.01$
		$E-O_3$	$12.07 \pm 0.61$	$0.12 \pm 0.02$	$206.27 \pm 22.41$	$3.86 \pm 0.47$	$0.46 \pm 0.04$	$0.24 \pm 0.02$	$0.53 \pm 0.05$
	PO	NF	$8.91 \pm 0.82$	$0.11 \pm 0.01$	$242.29 \pm 22.64$	$3.67 \pm 0.55$	$0.45 \pm 0.01$	$0.22 \pm 0.00$	$0.48 \pm 0.02$
		$E-O_3$	$7.98 \pm 0.89$	$0.14 \pm 0.02$	$284.54 \pm 19.85$	$2.56 \pm 0.50$	$0.45 \pm 0.02$	$0.22 \pm 0.02$	$0.49 \pm 0.04$
	RP	NF	$7.99 \pm 0.52$	$0.08 \pm 0.01$	$214.51 \pm 7.13$	$3.34 \pm 0.18$	$0.36 \pm 0.02$	$0.19 \pm 0.01$	$0.52 \pm 0.02$
		$E-O_3$	$7.53 \pm 1.27$	$0.06 \pm 0.02$	$156.39 \pm 39.80$	$4.22 \pm 0.71$	$0.37 \pm 0.02$	$0.18 \pm 0.03$	$0.48 \pm 0.06$
Sep.	AA	NF	$12.47 \pm 0.58a$	$0.15 \pm 0.02a$	$236.36 \pm 14.53$	$6.11 \pm 0.79$	$0.40 \pm 0.02$	$0.14 \pm 0.01$	$0.36 \pm 0.01$
		$E-O_3$	$9.14 \pm 0.08b$	$0.09 \pm 0.01b$	$221.73 \pm 16.98$	$6.35 \pm 0.71$	$0.48 \pm 0.08$	$0.14 \pm 0.00$	$0.33 \pm 0.02$
	FC	NF	$13.32 \pm 0.77a$	$0.17 \pm 0.02$	$252.58 \pm 14.25$	$5.54 \pm 0.28$	$0.50 \pm 0.03$	$0.20 \pm 0.01$	$0.40\pm0.01$

	$E-O_3$	$10.54 \pm 0.65$ b	$0.12 \pm 0.01$	$244.34 \pm 13.96$	$5.96 \pm 0.33$	$0.44 \pm 0.02$	$0.17 \pm 0.01$	$0.40 \pm 0.03$
PO	NF	$12.38 \pm 0.10a$	$0.16 \pm 0.01$	$258.47 \pm 4.21$	$4.92\pm0.23$	$0.51 \pm 0.05$	$0.17\pm0.00a$	$0.35 \pm 0.03$
	$E-O_3$	$10.23\pm0.18b$	$0.13 \pm 0.02$	$249.55 \pm 17.47$	$5.36 \pm 0.45$	$0.46 \pm 0.01$	$0.14 \pm 0.00b$	$0.32 \pm 0.01$
RP	NF	$9.55 \pm 0.10a$	$0.09\pm0.01a$	$172.74 \pm 18.89$	$5.79 \pm 0.52$	$0.38 \pm 0.00$	$0.17 \pm 0.00a$	$0.44 \pm 0.01$
	$E-O_3$	$6.61 \pm 0.09b$	$0.06 \pm 0.01b$	$157.95 \pm 17.30$	$5.76 \pm 0.60$	$0.38 \pm 0.03$	$0.13 \pm 0.01b$	$0.36 \pm 0.03$

Ozone also induced significant reductions in carboxylation parameters across two measurements and four species (Table 2), but for the gas exchange and fluorescence parameters, responses were similar between species as there were no significant interactions between species and  $O_3$  (Table 2). In August,  $Vc_{max}$  was significantly reduced by 32% in PO and in September by 42% and 37% in AA and RP, respectively.  $J_{max}$  was reduced by 27% and 36% in FC and PO in August, and by 19% for FC in September (Fig. 8). E- $O_3$  also significantly reduced the stomatal limitation ( $L_8$ ), which increased significantly with time (Table 2): reductions were significant for PO by 20% in August, for AA, FC and PO by 48%, 43% and 59%, respectively, in September (Fig. 8). No significant difference between species was found in carboxylation parameters and  $L_8$ .

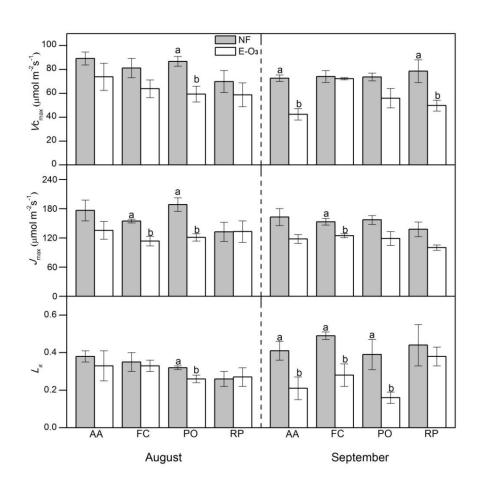


Figure 8. Carboxylation-related variables in plants grown in non-filtered air (NF) and

in elevated  $O_3$  (E- $O_3$ ) treatment in August and September. Different letter indicates significant difference between  $O_3$  treatments.

# 3.5. Pigment contents

Chlorophyll b (Chl b) and carotenoid (Car) contents as well as the Chl a / Chl b ratio differed significantly between species (Table 2), as indicated by the highest Chl b and Car in FC and the lowest in PO, but both of them did not have significant difference from AA and RP (Table 4). Although mean values in chlorophyll and carotenoid contents were always lower in the E-O<sub>3</sub> treatment, these changes were not statistically significant at the measuring time (Tables 2 and 4).

**Table 4**. The Chlorophyll (Chl) and carotenoid (Car) contents in plants grown in non-filtered air (NF) and in elevated O<sub>3</sub> (E-O<sub>3</sub>) treatments in September.

		Chl <i>a</i>	Chl <i>b</i>	Chla+b	Car	Chla/Chlb
		$(x10 \text{ mg m}^{-2})$				
AA	NF	$17.98 \pm 1.89$	$4.46 \pm 0.37$	$22.43 \pm 2.26$	$8.67 \pm 0.59$	$4.02 \pm 0.11$
	$E-O_3$	$13.23 \pm 2.16$	$3.31 \pm 0.55$	$16.54 \pm 2.71$	$6.46 \pm 0.98$	$4.00 \pm 0.06$
FC	NF	$19.84 \pm 2.09$	$5.52 \pm 0.64$	$25.36 \pm 2.72$	$9.72 \pm 0.83$	$3.62 \pm 0.07$
	$E-O_3$	$18.69 \pm 3.29$	$5.05 \pm 0.97$	$23.73 \pm 4.26$	$9.12 \pm 1.23$	$3.76 \pm 0.11$
PO	NF	$14.61 \pm 1.07$	$3.81\pm0.27$	$18.42 \pm 1.32$	$6.72 \pm 0.52$	$3.84 \pm 0.10$
	$E-O_3$	$12.63 \pm 1.42$	$3.17 \pm 0.38$	$15.80 \pm 1.79$	$6.12 \pm 0.59$	$4.00 \pm 0.08$
RP	NF	$17.51 \pm 2.97$	$4.49\pm0.82$	$22.00 \pm 3.79$	$8.10\pm1.18$	$3.94 \pm 0.05$
	$E-O_3$	$14.39 \pm 1.75$	$3.65 \pm 0.53$	$18.05 \pm 2.28$	$6.88 \pm 0.81$	$4.00 \pm 0.15$

#### 3.6. Antioxidants

At the end of the growing season, plants in E-O<sub>3</sub> significantly increased their contents in antioxidant compounds (TAC, phenolics, total and reduced AsA) with regard to control plants (Table 2). Responses of different species to O<sub>3</sub> were similar for all tested antioxidant variables as indicated by no significant interaction (Table 2). Across

O<sub>3</sub> treatments, there was significant difference between species in TAC, phenolics and reduce AsA contents (Table 2). Of all, PO had the largest antioxidant capacity, RP was the lowest, and FC and AA ranked in the middle level. RP was the species with lower constitutive levels of antioxidants, but it showed the strongest responses to O<sub>3</sub>, with increases of 67% in TAC, 67% in total phenolics, 85% in total AsA, and 65% in reduced AsA in E-O<sub>3</sub> plants (Fig. 9). On the contrary, AA did not experience significant changes in TAC and in reduced AsA after O<sub>3</sub> exposure (Fig. 9).

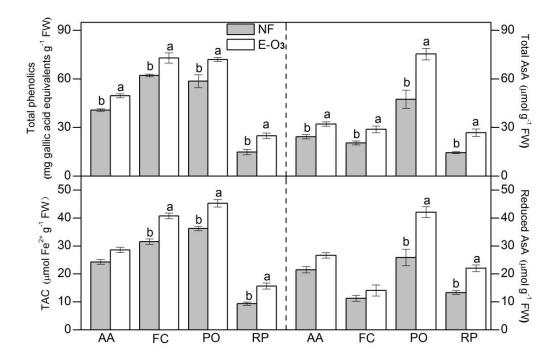


Figure 9. Total phenolics, total antioxidant capacity (TAC), total and reduced ascorbic acid (AsA) contents in plants grown in non-filtered air (NF) and in elevated  $O_3$  (E- $O_3$ ) treatments in September. Different letter indicates significant difference between  $O_3$  treatments.

# 4. Discussion

E-O<sub>3</sub> induced visible injury in the four investigated species with more severe effects on AA, as 75% of the plants were affected before middle July. AA, FC and RP were among the tree species showing symptoms in the field (Feng et al., 2014). AA can be considered a good bio-indicator tree in Beijing area, because it was the tree species

showing symptoms at more sites and the symptoms were observed already in July with AOT40 of 12.8 ppm.h. Symptoms of these 3 tree species in E-O<sub>3</sub> were consistent with those observed in the field, supporting the validation of field observations in Beijing area.

523

524

525

526

527

528

529

530

531

532

533

534

535

536

537

538

539

540

541

542

543

544

545

546

547

548

549

550

551

Many of the anatomical and ultrastructural changes induced by E-O<sub>3</sub> are hypersensitive-like response (HR), in which oxidative burst (OB) plays a central role (Rao et al., 2000). Both are considered to be plant defence reactions against O<sub>3</sub> as well as other biotic and abiotic stresses (Sanderman et al., 1998; Vollenweider et al, 2003). OB triggers programmed death, leading to a cell collapse as observed in stippling areas in AA and PO. As shown in the present study, palisade parenchyma is usually the most affected tissue by O<sub>3</sub> (Günthardt and Vollenweider, 2007; Paoletti et al., 2009). Our observations suggest that reactive oxygen species (ROS) degrades the pectins of the middle lamella inducing a separation of the mesophyll cells, and increasing intercellular spaces. Hydrogen bounds between cellulose molecules and other cell wall components may also be affected, finally softening the cell which becomes deformed. Pectinaceous and polysaccharidic extrusions also result from the breakdown of the middle lamella. These projections have been reported in several species exposed to O<sub>3</sub> in fumigation chambers or in the field (e.g., Calatayud et al., 2011; Vollenweider et al., 2003), but are not specific of O<sub>3</sub> as they can be induced by other stress factors such as e.g. heavy metals (Günthard-Goerg and Vollenweider, 2007). Progressive degradation of the cell walls combined with the loss of turgor finally leads to cell collapse.

Callose accumulation is a defence response to abiotic and biotic factors such as wounding, desiccation, metal toxicity or insect attacks, and it has been postulated that it might act as a physical barrier against microbial and fungal attack (Stone and Clarke, 1992), isolating affected tissues from healthy cells. Ozone is considered to be an abiotic elicitor inducing responses similar to plant defence responses to pathogen attacks (Sandermann et al., 1998). Callose accumulation after O<sub>3</sub> exposure has been reported in different species (Gravano et al., 2003; Bussotti et al., 2005; García-Breijo

et al., 2008; Calatayud et al., 2011). Phenolic metabolites are suggested to play a protective role against oxidative stress as antioxidants (Kangasjärvi et al., 1994). In the four species, E-O<sub>3</sub> increased accumulation of phenolic compounds. In the vacuoles of AA and PO, rich in condensed tannins, these compounds changed their even distribution by a coagulated aspect most likely due to oxidative processes (Vollenweider et al., 2003). Changes in vacuole tannins due to O<sub>3</sub> have been observed in the mastic plant (Reig-Armiñana et al. 2004). In RP, less rich in tannins, an accumulation of phenolic compounds is observed. Previous microscopy studies have shown that accumulation of phenolic compounds including tannins and anthocyanins can be induced by O<sub>3</sub> in some species (Vollenweider et al., 2003; Bussotti et al., 2005). At the ultrastructural level, changes in organelles that may be considered as an acceleration of the natural foliar senescence process can be observed. Foliar senescence is characterized by a decline in whole leaf gas exchange and protein levels, leaf yellowing, and the chloroplast-to-gerontoplast transition. Ultrastructurally, gerontoplast development is seen primarily as a progressive unstacking of grana, a loss of thylakoid membranes and a massive accumulation of plastoglobuli and other lipid-protein inclusions (Harris & Schaefer, 1981). In our study, ultrastructural changes in the chloroplast included an increase in electrodense material, accumulation of starch, plastoglobuli and lipid-protein bodies, thylakoid degradation, membrane disruption and changes in shape. Starch accumulation may be related to a difficulty in sucrose transport outside the chloroplasts and to other tissues (Calatayud et al., 2011). Landolt et al. (1997) found enhanced content of soluble sugars in leaves of O<sub>3</sub>-exposed birch plants, and in the same species, Matyssek et al. (1992) observed accumulation of starch along veins, which suggested a reduction of carbon export from source leaves. This can be related with altered cell membranes and impaired phloem loading induced by O<sub>3</sub> (Grantz, 2003). In O<sub>3</sub>-injured leaves, phloem cells can experience evident changes in shape, which may obviously impair their functionality

552

553

554

555

556

557

558

559

560

561

562

563

564

565

566

567

568

569

570

571

572

573

574

575

576

577

578

579

580

and affect sucrose translocation (Calatayud et al., 2011). On the other hand, an

increase in plastoglobuli can originate from the lipid-soluble degradation products from the thylakoid membranes (Matile, 1992; Kivimäenpäa et al., 2010). Overall, the four species show partly similar anatomical and ultrastructural changes that are related both to O<sub>3</sub> damage to different cell components, defence responses and accelerated senescence. Notably, changes were more evident in AA, with many palisade parenchyma cells collapsed leading to a large intercellular spaces, distinct callose dipositions, denser vacuolar content due to tannin accumulation, and more altered chloroplasts (with accumulation of abundant lipid-protein bodies). On the contrary, much less evident changes were observed in RP, with FC and PO being intermediate. The damage to chloroplast functionality has led to reduced photosynthesis (Table 3). Investment in defence may, on the other hand, reduce carbon availability for plant growth.

Compared to control, changes in leaf gas exchange, chlorophyll a fluorescence and carboxylation parameters by E-O<sub>3</sub> showed similar responses in all species, as indicated by no significant interactions between O<sub>3</sub> and species. Ozone induced significant declines in  $A_{\text{sat}}$  and in chlorophyll a fluorescence parameters ( $\phi_{\text{PSII}}$  and qP). This decline in photosynthetic CO<sub>2</sub> assimilation was associated with significant reductions in  $Vc_{\text{max}}$  and  $J_{\text{max}}$ , suggesting that biochemical limitations play an early and primary role in the decline of CO<sub>2</sub> assimilation by O<sub>3</sub> (e.g., Calatayud et al., 2010; Feng et al., 2011b; Cho et al., 2011). Changes in fluorescence parameters under steady-state illumination may reflect a down-regulation process for adjusting the production of reductive power and chemical energy to a lower demand by the Calvin-Benson cycle (Calatayud et al., 2007). In a complementary study conducted with the same plants but restricted to only three of the species (AA, FC and PO), E-O<sub>3</sub> increased stomatal sluggishness (i.e. slowed stomatal response) with FC and PO being the most and less affected species, respectively (Hoshika et al., 2014). Increased sluggishness may be related to accelerated leaf senescence in the cell physiological processes (Paoletti et al., 2009), although the underlying mechanisms are still under investigation (Hoshika et al., 2014). In the present study, chloroplasts of the guard

cells are sometimes altered, which could partly impair the normal performance of these cells.

610

611

612

613

614

615

616

617

618

619

620

621

622

623

624

625

626

627

628

629

630

631

632

633

634

635

636

637

638

E-O<sub>3</sub> significantly increased total and reduced AsA contents in the leaves. Activation of the AsA synthesis is a defence response against oxidative stress, as AsA is a central metabolite in plant antioxidant system. It serves as a chemical scavenger to protect plants by reducing free radicals, and also as a substrate of extracellular enzymes such as the ascorbate peroxidase (APX) which detoxify peroxides; therefore propagation of oxidative signaling diminishes (Burkey et al., 2006; Dizengremel et al., 2013). Besides, all species in the present study showed an increase in phenolics contents. Similar results have been previously reported in other tree species (Oksanen et al., 2013). Phenolic metabolites probably increase O<sub>3</sub> tolerance of plants due to its effectiveness as radicals and ROS scavengers (Grace, 2005; Langebartels et al., 2002). Increased TAC levels are consistent with increased phenolics content as phenolic compounds have been found to be the major contributors to the antioxidant properties in extracts of different parts of 30 plants (e.g. Dudonné et al., 2009). In the present study, however, all four species showed a similar response to O<sub>3</sub> in antioxidant levels. Besides the increases in AsA, phenols or TAC, the different species were not able to counteract photosynthesis impairment, cellular and tissue damage, and visible injury, suggesting that the possible contribution of these physiological responses was insufficient to offset the high levels of oxidative stress.

Considering visible injury, anatomical and ultrastructural responses, AA is the most sensitive of all species as it developed symptoms the earliest and the percentage of injuried leaves was the highest from the beginning. Anatomical and ultrastructural alterations were also the severest in this species. On the other side, any of the RP plants reached the higher injury classes, and visible injury was much less conspicuous than the other species, with anatomical and ultrastructural changes being also moderate. FC and PO were intermediate, with some of the leaves strongly affected by a marked yellow or brown stippling. Several studies have shown that LMA is well correlated with O<sub>3</sub> sensitivity of the species: plants with higher LMA are, in general,

more O<sub>3</sub> tolerant, partly due to the fact that they have a higher chlorophyll and nitrogen content per area unit, supporting a more efficient photosynthesis activity, which can better feed detoxification processes (Bussotti, 2008; Zhang et al., 2012). Our results are not in line with such a hypothesis, probably because the investigated species have a narrow LMA range (5.0-9.1 mg cm<sup>-2</sup>). LMA is better related with O<sub>3</sub> tolerance when a larger range among species is considered (e.g., comparing deciduous with evergreen species, Calatayud et al., 2011). The higher sensitivity in AA can rather be explained as a combination of factors including a thinner palisade parenchyma layer, less active antioxidant responses to  $O_3$ , and the highest  $g_s$ . The palisade parenchyma is the most photosynthetically active tissue, so a reduced thickness may imply that less apoplast surface is available for defense reactions (Dizengremel et al., 2013). The less active antioxidant responses in combination with higher  $g_s$  (related to a higher  $O_3$  uptake), would make this species more prone to oxidative imbalances. These features are consistent with the pioneering, fast-growing strategy of this species. On the other hand, although RP is also a sensitive species, its lower  $g_s$  and, probably also, its stronger antioxidant responses could better withstand O<sub>3</sub> stress, besides the fact of having lower constitutive antioxidant levels. Besides showing more or less marked macro- and microscopic responses, it is noteworthy that photosynthesis was also impaired in all the species, with CO<sub>2</sub> assimilation being reduced. On the long run, such reductions are expected to affect CO<sub>2</sub> fixation as plant biomass, one of the services of urban forests. In the present study, however, only one growing season was covered and the effects on biomass were negligible (data not shown).

662

663

664

665

666

667

639

640

641

642

643

644

645

646

647

648

649

650

651

652

653

654

655

656

657

658

659

660

661

#### 5. Conclusions

The investigated four species are sensitive to  $O_3$ , as indicated by an increase in foliar damage, accumulation of phenolic compounds in the leaf tissues, degradation of the cell walls and organelles such as the chloroplasts, changes in antioxidant levels and reduced photosynthesis rate. These changes might also alter plant interactions with

other abiotic and biotic stresses (Karnosky et al., 2005; Bussotti et al., 2008). The current study suggests that sensitivity to O<sub>3</sub> should be taken into account for urban tree plantations in areas at risk of high ozone concentrations, avoiding very sensitive species such as AA. This is especially relevant for urban forests and large tree plantations in parks or in green belts around cities rather than for trees planted in streets, where O<sub>3</sub> is locally scavenged by reacting with NO traffic emissions. Further studies of O<sub>3</sub> exposure under controlled or semi-controlled conditions involving many urban trees are still needed in order to support decision making for tree plantation in ozone polluted cities of China. Other additional factors to be considered in urban greening species in relation to air pollution are their rate of BVOC emission and capacity to remove O<sub>3</sub> and particulate matter (Paoletti et al., 2009; Calfapietra et al., 2013).

# Acknowledgements

This study has been funded by the Hundred Talents Program, Chinese Academy of Sciences and State Key Laboratory of Urban and Regional Ecology. Collaboration between RCEES and Fundación CEAM has been possible thanks to project AMIS (Fate and Impact of Atmospheric Pollutants, PIRSES-GA-2011-295132), and by the Chinese Academy of Sciences Visiting Professorships for Senior International Scientists (Grant Number: 2013T2Z0009). VC also acknowledges the support of PROMETEOII/2014/038 project (Generalitat Valenciana, G.V.), and FGB and JRA that of PROMETEOII2013/021 (G.V.), and CGL2012-40058-C02-01/02 (MINECO).

#### 6. References

Ainsworth, E.A., Gillespie, K.M., 2007. Estimation of total phenolic content and other oxidation substrates in plant tissues using Folin-Ciocalteu reagent. Nature Protocols 2, 875-877.

We thank Mr. Yulong Zhang for the experimental management.

- Benzie, I.F.F, Strain, J.J., 1996. The ferric reducing ability of plasma (FRAP) as a
- measure of 'antioxidant power': the FRAP assay. Analytical Biochemistry 239,
- 698 70-76.
- Burkey, K.O., Neufeld, H.S., Souza, L., Chappelka, A.H., Davison, A.W., 2006.
- Seasonal profiles of leaf ascorbic acid content and redox state in ozone-sensitive
- wildflowers. Environmental Pollution 143, 427-434.
- Bussotti, F. 2008. Functional leaf traits, plant communities and acclimation processes
- in relation to oxidative stress in trees: a critical overview. Global Change Biology
- 704 14, 2727–2739.
- Bussotti, F., Agati, G., Desotgiu, R., Matteini, P., Tani, C., 2005. Ozone foliar
- symptoms in woody plant species assessed with ultrastructural and fluorescence
- analysis. New Phytologist 166, 941–955.
- Calatayud V., Cerveró J., Sanz M.J., 2007. Foliar, physiologial and growth responses
- of four maple species exposed to ozone. Water, Air, and Soil Pollution 185,
- 710 239-254.
- 711 Calatayud V., Marco F., Cerveró J., Sánchez-Peña G., Sanz M.J., 2010. Contrasting
- ozone sensitivity in related evergreen and deciduous shrubs. Environment Pollution
- 713 158, 3580-3587.
- Calatavud, V., García-Breijo, F.J., Cervero, J., Reig-Armiñana, J., Sanz, M.J., 2011.
- Physiological, anatomical and biomass partitioning responses to ozone in the
- Mediterranean endemic plant *Lamottea dianae*. Ecotoxicology and Environmental
- 717 Safety 74,1131-1138.
- Calfapietra, C., Fares, S., Manes, F., Morani, A., Sgrigna, G., Loreto, F., 2013. Role of
- biogenic volatile organic compounds (BVOC) emitted by urban trees on ozone
- concentration in cities: a review. Environmental Pollution 183.71-80.
- 721 Chen, W., Tang, H.Z., Zhao, H.M., 2015. Diurnal, weekly and monthly spatial
- variations of air pollutants and air quality of Beijing. Atmospheric Environment
- 723 119, 21-34.
- 724 Cho K., Tiwari S., Agrawal S., Torres N., Agrawal M., Sarkar A., et al., 2011.
- 725 Tropospheric ozone and plants: absorption, responses, and consequences, in:

- Whitacre D.W. (Eds.), Reviews of Environmental Contamination and Toxicology
- 727 Volume 212. Springer, PP. 61-111.
- Dizengremel, P., Jolivet, Y., Tuzet, A. Ranieri, A., Le Thiec, D., 2013. Integrative
- leaf-level ozone phytotoxic ozone dose assessment for forest risk modeling, in:
- Matyssek, R., Clarke, N., Cudlin, P., Mikkelsen T.N., Tuovinen, J.P., Wieser, G.,
- Paoletti, E., (Eds.), Climate Change, Air Pollution and Global Challenges.
- Developments in Environmental Sciences 13. Elsevier, Netherlands, pp. 267-288.
- Dudonne, S., Vitrac, X., Coutiere, P., Woillez, M., Mérillon, J.M., 2009. Comparative
- study of antioxidant properties and total phenolic content of 30 plant extracts of
- industrial interest using DPPH, ABTS, FRAP, SOD, and ORAC assays. Journal of
- Agricultural and Food Chemistry 57, 1768-1774.
- Feng, Z.Z., Hu, E.Z., Wang, X.K., Jiang, L.J., Liu, X.J., 2015a. Ground-Level O<sub>3</sub>
- pollution and its impacts on food crops in China: a review. Environmental Pollution
- 739 199, 42-48.
- Feng, Z.Z., Liu, X.J., Zhang, F.S., 2015b. Air pollution affects food security in China:
- taking ozone as an example. Frontier Agricultural. Sciences and Engineer 2(2),
- 742 152-158.
- Feng, Z.Z., Niu, J.F., Zhang, W.W., Wang, X.K., Yao, F.F., Tian, Y., 2011a. Effects of
- ozone exposure on Sub-Tropical evergreen Cinnamomum Camphora seedlings
- grown in different nitrogen loads. Trees 25, 617-625.
- Feng, Z.Z., Pang, J., Kobayashi, K., Zhu, J., Ort, D.R., 2011b. Differential responses
- in two varieties of winter wheat to elevated ozone concentration under fully
- open-air field conditions. Global Change Biology 17, 580-591.
- Feng, Z.Z., Sun, J.S., Wan, W.X., Hu, E.Z., Calatayud, V., 2014. Evidence of
- videspread ozone-induced visible injury on plants in Beijing, China.
- 751 Environmental Pollution 193, 296-301.
- Gao, J.C., Guo, G.J., Guo, Y.M., Wang, X.X., Du, Y.C., 2011. Measuring plant leaf
- area by scanner and ImageJ software. China Vegetables 2, 73-77.

- García-Breijo, F.J., Reig-Armiñana, J., Bautista-Peris, B., Calatayud, V., Cerveró, J.,
- Sanz, M.J., 2008. Effets anatomiques de l'ozone sur le térébinthe (Pistacia
- 756 *terebinthus*). Forêt Méditerranéenne 29, 13–22.
- 757 Gillespie, K.M., Ainsworth, E.A., 2007. Measurement of reduced, oxidized and total
- ascorbate content in plants. Nature Protocols 2, 871-874.
- 759 Grace, S.C., 2005. Phenolics as antioxidants. In: Smirnoff, N. (Eds.), Antioxidants
- and Reactive Oxygen Species in Plants. Blackwell Publishing Ltd, Oxford, UK, pp.
- 761 141–168.
- 762 Grantz, D., 2003. Ozone impacts on cotton: Towards an integrated mechanism.
- Environmental Pollution 126, 331-344.
- Gravano, E., Giulietti, V., Desotgiu, R., Bussotti, F., Grossoni, P., Gerosa, G., et al.,
- 765 2003. Foliar response of an *Ailanthus altissima* clone in two sites with different
- levels of ozone-pollution. Environmental Pollution 121, 137–146.
- Günthardt-Goerg, M.S., Vollenweider, P., 2007. Linking stress with macroscopic and
- microscopic leaf response in trees: New diagnostic perspectives. Environmental
- 769 Pollution 147, 467-488.
- Harris, J.B., Schaefer, V.G., 1981. Some correlated events in aging leaf tissues of tree,
- tomato and tobacco. Botanical Gazette, 43-54.
- Hoshika, Y., Carriero, G., Feng, Z., Zhang, Y., Paoletti, E., 2014. Determinants of
- stomatal sluggishness in ozone-exposed deciduous tree species. Science of the Total
- 774 Environment 481, 453-458.
- 775 IPCC, 2013. Intergovernmental Panel on Climate Change. Fifth Assessment Report.
- http://www.ipcc.ch/report/ar5/index.shtml.
- 777 Kangasjärvi, J., Talvinen, J., Utriainen, M., Karjalainen, R., 1994. Plant defense
- systems induced by ozone: commissioned review. Plant Cell & Environment 17,
- 779 783–794.
- 780 Karnosky D., Pregitzer K. S., Zak D. R., Kubiske M. E., Hendrey G., Weinstein D.,
- Nosal M., Percy, K.E., 2005. Scaling ozone responses of forest trees to the
- ecosystem level in a changing climate. Plant, Cell and Environment, 965–981.

- 783 Kivimäenpää, M., Sutinen, S., Calatayud, V., Sanz, M.J., 2010. Visible and
- microscopic needle alterations of mature Aleppo Pine (Pinus halepensis) trees
- growing on an ozone gradient in eastern Spain. Tree Physiology 30, 541-554.
- Krupa, S., McGrath, M.T., Andersen, C.P., Booker, F., Burkey, K.O., Chappelka, A.H.,
- et al., 2000. Ambient ozone and plant health. Plant Disease 85, 4-12.
- Landolt, W., Günthardt-Goerg, M.S., Pfenninger, I., Einig, W., Hampp, R., Maurer, S.,
- et al., 1997. Effect of fertilization on ozone-induced changes in the metabolism of
- birch (*Betula pendula*) leaves. New Phytologist 137, 389–397.
- Langebartels, C., Schraudner, M., Heller, W., Ernst, D., Sandermann, H., 2002.
- Oxidative stress and defense reactions in plants exposed to air pollutants and UV-B
- radiation, In: Inze', D., Van Montagu, M. (Eds.), Oxidative Stress in Plants. Taylor
- 794 & Francis, London, UK, pp.105–135.
- 795 Lee, J.B., Cha, J.S., Hong, S.C., Choi, J.Y., Myoung, J.S., Park, R.J., et al., 2015.
- Projections of summertime ozone concentration over East Asia under multiple
- 797 IPCC SRES emission scenarios. Atmospheric Environment 106, 335-346.
- Lichtenthaler, H.K., 1987. Chlorophylls and carotenoids: pigments of photosynthetic
- biomembranes. Methods in Enzymology 148, 350-382.
- LRTAP Convention, 2010. Manual on Methodologies and Criteria for Modelling and
- Mapping Critical Loads & Levels and Air Pollution Effects, Risks and Trends. In:
- Chapter 3: Mapping Critical Levels for Vegetation.
- Matile, P., 1992. Chloroplast senescence. In: Baker, N.R., Thomas H. (Eds.), Crop
- Photosynthesis: Spatial and Temporal Determinants. Elsevier Ltd., pp. 413-440.
- Matyssek, R., Bytnerowicz, A., Karlsson, P.E., Paoletti, E., Sanz, M., Schaub, M., et
- al. 2007. Promoting the O<sub>3</sub> flux concept for European forest trees. Environmental
- 807 Pollution, 146, 587–607.
- Matyssek, R., Günthardt-Goerg, M.S., Saurer, M., Keller, T., 1992. Seasonal growth,
- $\delta^{13}$ C in leaves and stem, and phloem structure of birch (*Betula pendula*) under low
- ozone concentrations. Trees 6, 69–76.

- Nowak, D.J., Hirabayashi, S., Bodine, A., Greenfield, E.G, 2014. Tree and forest
- effects on air quality and human health in the United States. Environmental
- 813 Pollution 193, 119-129.
- Oksanen, E., Keski-Saari, S., Kontunen-Soppela, S., Keinänen, M., 2013.
- Metabolomics and transcriptomics increase our understanding about defence
- responses and genotypic differences of northern deciduous trees to elevating ozone,
- 817 CO<sub>2</sub> and climate warming. Climate Change, Air Pollution and Global Challenges.
- Developments in Environmental Science 13, 309-326.
- Paoletti, E., Contran, N., Bernasconi, P., Günthardt-Goerg, M.S., Vollenweider, P.,
- 820 2009. Structural and physiological responses to ozone in Manna ash (Fraxinus
- Ornus L.) leaves of seedlings and mature trees under controlled and ambient
- conditions. Science of the Total Environment 407, 1631-1643.
- Paoletti, E., Ranieri, A., Lauteri, M. 2008. Moving toward effective ozone flux
- assessment. Environmental Pollution, 156, 16–19.
- Rao, M.V., Koch, J.R., Davis, K.R., 2000. Ozone: A Tool for Probing Programmed
- Cell Death in Plants, In: Lam E., Fukuda H., Greenberg J. (Eds.), Programmed Cell
- Death in Higher Plants. Springer, pp.101-114.
- Reig-Armiñana, J., Calatayud, V., Cerveró, J., Garcia-Breijo, F., Ibars, A., Sanz, M.,
- 829 2004. Effects of ozone on the foliar histology of the mastic plant (*Pistacia*
- 830 *Lentiscus* L.). Environmental Pollution 132, 321-331.
- 831 Sandermann, H., Ernst, D., Heller, W., Langebartels, C., 1998. Ozone: An abiotic
- elicitor of plant defence reactions. Trends in Plant Science 3, 47-50.
- 833 Sharkey, T.D., Bernacchi, C.J., Farquhar, G.D., Singsaas, E.L., 2007. Fitting
- photosynthetic carbon dioxide response curves for C3 leaves. Plant, Cell &
- 835 Environment 30, 1035-1040.
- Stone, B.A., Clarke, A.E. ,1992. Chemistry and biology of 1, 3-[beta]-glucans. La
- Trobe University Press Melbourne.
- The Royal Society, 2008. Ground-level ozone in the 21st century: future trends,
- impacts and policy implications. Science Policy Report 15/08. The Royal Society,
- 840 London.

- Valkama, E., Koricheva, J., Oksanen, E., 2007. Effects of elevated O<sub>3</sub>, alone and in
- combination with elevated CO<sub>2</sub>, on tree leaf chemistry and insect herbivore
- performance: a meta-analysis. Global Change Biology 13, 184–201.
- Vingarzan, R., 2004. A review of surface ozone background levels and trends.
- Atmospheric Environment 38, 3431-3442.
- Vollenweider, P., Ottiger, M., Günthardt-Goerg, M., 2003. Validation of leaf ozone
- 847 symptoms in natural vegetation using microscopical methods. Environmental
- 848 Pollution 124, 101-118.
- Wang, X.P., Mauzerall, D.L., 2004. Characterizing distributions of surface ozone and
- its impact on grain production in China, Japan and south Korea: 1990 and 2020.
- Atmospheric Environment 38, 4383-4402.
- Yamaji, K., Ohara, T., Uno, I., Kurokawa, J.I., Pochanart, P., Akimoto, H., 2008.
- Future prediction of surface ozone over east Asia using models-3 community
- multiscale air quality modeling system and regional emission inventory in Asia.
- Journal of Geophysical Research Atmospheres 113, 2156-2202.
- Yang, J., McBride, J., Zhou, J.X., Sun, Z.Y., 2005. The urban forest in Beijing and its
- role in air pollution reduction. Urban Forestry & Urban Greening 3, 65-78.
- Yuan, X.Y., Calatayud, V., Jiang, L.J., Manning, W.J., Hayes, F., Tian, Y., Feng, Z.Z.,
- 2015. Assessing the effects of ambient ozone in China on snap bean genotypes by
- using ethylenediurea (EDU). Environmental Pollution 205, 199-208.
- Zhang, W.W., Feng, Z.Z., Wang, X.K., Niu, J.F., 2012. Responses of native
- broadleaved woody species to elevated ozone in subtropical China. Environmental
- 863 Pollution 163, 149-157.