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Additional Information

1	Morphological and molecular characterization of local varieties, modern cultivars
2	and wild relatives of an emerging vegetable crop, the pepino (Solanum muricatum),
3	provides insight into its diversity, relationships and breeding history
4	
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15 Abstract

16 Availability of standardized morphological and molecular characterization data is essential for the efficient development of breeding programmes in emerging crops. 17 18 Pepino (Solanum muricatum) is an increasingly important vegetatively propagated vegetable crop for which concurrent data on morphological descriptors and molecular 19 20 markers are not available. We evaluated 58 morphological traits, using a collection of 21 14 accessions of pepinos (including local Andean varieties and modern cultivars) and 8 22 of wild relatives, using the IPGRI and COMAV descriptors lists coupled with 20 EST-SSRs from tomato. High morphological diversity was found in both cultivated and wild 23 24 accessions; all morphological traits except three were variable. Cultivated pepino and wild relatives were significantly different for 26 traits. Also, local varieties and modern 25 26 cultivars of pepino were different from each other for 13 morphological traits and were 27 clearly separated in a principal components analysis (PCA). Fourteen of the 20 tomato EST-SSRs were polymorphic, with an average number of alleles per locus of 4.07 and a 28 29 polymorphic information content (PIC) value of 0.4132. This revealed a high degree of 30 transferability from tomato to pepino and wide molecular diversity in the collection. Cultivated materials manifest high levels of observed heterozygosity, suggesting that it 31 32 is related to heterosis for yield associated with heterozygosis. SSR data clearly 33 differentiated cultivated and wild materials. Furthermore, for pepinos, the modern varieties were genetically much less diverse than the traditional local varieties. 34 However, both groups of cultivated material expressed a low degree of genetic 35 36 differentiation. A strong correlation (r=0.673) between morphological and molecular distances was found. Our results provide foundational information for programmes of 37 38 germplasm conservation, and that can be used to enhance breeding for this emerging 39 crop.

40 Keywords: Breeding · Descriptors · Germplasm · Heterozygosity · Solanum muricatum
41 · SSRs

42

43 Introduction

44

Modern breeding programmes in emerging crops are often limited by scanty or nonexistent phenotypic and genetic information, and by small germplasm collections (FAO
2010; Mayes et al. 2012). Complementary studies of morphological and molecular
diversity provide relevant information for identifying sources of variation in breeding
programmes, for establishing relationships among plant materials, as well as a
foundation for promoting breeding and for germplasm conservation (Rao and Hodgkin
2002; Khoury et al. 2010).

The pepino (Solanum muricatum Aiton) is an emerging usually vegetatively 52 propagated vegetable crop native to the Andean region (Anderson et al. 1996). This crop 53 54 is phylogenetically close to tomato (S. lycopersicum L.) and potato (S. tuberosum L.) (Spooner et al. 1993; Särkinen et al. 2013). The pepino is cultivated for its juicy and 55 aromatic fruits. Although the pepino is locally important in the Andean region since 56 long ago (Prohens et al. 1996), in recent decades the increasing interest in exotic fruit 57 markets has promoted increasing interest in pepino cultivation in several countries 58 including New Zealand, Australia, Spain, Turkey, Israel and China (Levy et al. 2006; 59 Yalçin 2010; Rodríguez-Burruezo et al. 2011; Abouelnasr et al. 2014). Nutritionally, 60 61 pepino fruits contain high levels of potassium and vitamin C, and it is low in calories. Furthermore, it offers some properties of medicinal interest, such as antidiabetic, 62 63 antidiuretic and antihypotensive activities (Hsu et al. 2011; Rodríguez-Burruezo et al. 2011; Sudha et al. 2012). 64

Most of the plant material cultivated in the Andean region consists of local 65 varieties that have not been subjected to formal breeding and are adapted to local 66 climatic conditions and preferences for flavour, size and fruit shape and colour 67 (Anderson et al. 1996; Prohens et al. 1996). Local varieties of the pepino are commonly 68 cultivated outdoors in their native range, and they usually have a poor performance 69 when introduced in other regions (where the pepino is cultivated either outdoors or in 70 greenhouses: Prohens et al. 1996; Rodríguez-Burruezo et al. 2011). As a consequence of 71 72 the usually poor performance, several improved cultivars adapted to non-Andean climates and to protected cultivation have been developed in New Zealand, Spain, and 73 Israel (Dawes and Pringle 1984; Simms et al. 1996; Ruiz et al. 1997; Prohens et al. 74 75 2002; Rodríguez-Burruezo et al. 2004a, 2004b; Levy et al. 2006). These materials have been developed using conventional approaches including generating genetically variable 76 77 populations by means of seed propagation of collections from the Andean region or by hybridization between different vegetatively propagated clones in order to exploit 78 79 heterosis (Rodríguez-Burruezo et al. 2011). 80 Wild pepino relatives which, like the domesticated pepino, are included in the section Basarthrum of genus Solanum (Anderson 1975, 1979) represent a genetic 81 resource of interest for pepino breeding (Rodríguez-Burruezo et al., 2003a). Among the 82 wild relatives, the highly variable S. caripense Humb. and Bonpl. ex Dun., as well as S. 83 tabanoense Correll, form part of the primary genepool of pepino. Fully fertile 84 interspecific hybrids and backcross generations to pepino have been obtained among 85 86 these species (Anderson 1979; Rodríguez-Burruezo et al. 2003a, 2011). Other species of interest for pepino breeding include S. trachycarpum Bitter and Sodiro, which grows in 87 dry areas (Anderson 1979), and S. catilliflorum G.J. Anderson, Martine, Prohens and 88 Nuez and S. perlongistvlum G.J. Anderson, Martine, Prohens and Nuez, which are 89

among the most recent species discovered and described for this section (Anderson et al. 90 2006) and that remain to be studied as potential genetic resources for pepino breeding. 91 Given the interests in crop diversity and enhancement, the precise and 92 93 standardized morphological and molecular characterization of the pepino would be of great utility for breeding programmes, for germplasm conservation and for comparison 94 of experimental data of different trials and plant materials (Rao and Hodgkin 2002; 95 Khoury et al. 2010). Fortunately, an internationally accepted list of morphological 96 97 descriptors for the extensive characterization of vegetative, inflorescence and flower, fruit and seed traits of pepino is available (IPGRI and COMAV 2004). However, no 98 99 reports are known to us on the utilization of this list of descriptors for the morphological characterization of pepino collections. Although several studies have been made on 100 phenotypic diversity of pepino, including wild relatives of interest for breeding, they 101 102 have mostly dealt with specific traits of agronomic interest (Rodríguez-Burruezo et al. 103 2003a, 2011; Muñoz et al. 2014)

104 Similarly, few studies have been done on the molecular diversity of collections 105 of cultivated pepino and wild relatives (Anderson et al. 1996; Blanca et al. 2007). The evaluation of the cpDNA-RFLPs polymorphism in the pepino and wild relatives of 106 107 Solanum section Basarthrum revealed that the cultivated pepino was closely related to 108 S. caripense and S. tabanoense (Anderson et al. 1996). A subsequent study using AFLP markers and the sequence variation in the DNA sequence of the nuclear gene 3-109 methylcrotonyl-CoA carboxylase revealed that cultivated pepino is highly diverse and 110 111 showed that this cultigen was genetically differentiated from wild relatives (Blanca et al. 2007). AFLP markers have also been used to evaluate the genetic distances among 112 113 four pepino cultivars as a predictor for heterosis for yield traits (Rodríguez-Burruezo et al. 2003b). However, no studies have been performed with other molecular markers in 114

pepino. Unlike AFLPs, which are dominant (Meudt and Clarke 2007), SSRs are co-115 dominant and particularly valuable because they allow the precise assignment of allelic 116 states and evaluation of the level of heterozygosity of individual pepino clones. 117 118 Furthermore, SSRs (1) have a high reproducibility and therefore are ideal for comparison among different experiments and laboratories, (2) are multiallelic, (3) have 119 locus specificity, (4) are abundant and (5) are randomly distributed throughout the 120 genome (Kalia et al. 2011). For species like the pepino in which no genomic libraries or 121 122 expressed sequence tags (EST) sequences are available, SSRs may be transferred from close relatives, like tomato, in which there has been an abundance of SSRs developed 123 124 (Frary et al. 2005; Suresh et al. 2014). In this respect, EST-SSRs usually offer a greater degree of transferability among species, as transcribed regions have a greater degree of 125 conservation than non-transcribed regions (Kalia et al. 2011). 126

127 The simultaneous study of morphological and molecular diversity of the pepino 128 and wild relatives also provides information on the morphological and molecular 129 variation and relationships of the crop to wild relatives, as well as on the association 130 between morphological and molecular variation. Here, we evaluate the morphological and molecular diversity using standardized descriptors and highly repeatable SSR 131 markers in a collection of local varieties and modern cultivars of pepino, as well as in a 132 set of accessions from wild relatives of interest for breeding. The information obtained 133 will be of interest for breeders and germplasm managers, as well as for understanding 134 the evolution of the crop. 135

136

137 Material and methods

138

139 Plant material

141 We studied a total of 22 accessions, of which six corresponded to local pepino varieties 142 from the Andean region, eight to improved pepino cultivars, and eight to wild relatives 143 (different species) (Table 1). Local varieties originated in Colombia (1), Chile (2), Ecuador (2) and Peru (1). Modern varieties were developed in New Zealand (2), Spain 144 (5) and the United Kingdom (1) as a result of selection and breeding programmes 145 146 (Dawes and Pringle 1984; Simms et al. 1996; Ruiz et al. 1997; Prohens et al. 2002; 147 Rodríguez-Burruezo et al. 2004a, 2004b). Wild relatives were represented by accessions of S. caripense (4), S. catilliflorum (1), S. perlongistylum (1), S. tabanoense (1) and S. 148 trachycarpum (1). The material is part of the germplasm collection of the Instituto de 149 Conservación y Mejora de la Agrodiversidad valenciana (Valencia, Spain). 150 151 Five clonal replicates obtained by in vitro micropropagation (Cavusoglu and 152 Sulusoglu 2013) were used for each of the 22 accessions. Clonal replicates were grown in a glasshouse in Valencia (GPS coordinates: lat. 39° 29' 01'' N, long. 0° 20' 27'' W) 153 154 using a completely randomized design. Rooted plantlets were transplanted to benches 155 filled with quartz sand in January 2014. Plants were spaced 55 cm in the bench, with 115 cm between bench centers. Plants were drip irrigated every 4 h for 5 min. 156 157 Fertilization was applied through the drip irrigation system during the growing cycle. A 158 combination of different fertilizers was used to achieve a final concentration of main ions and cations in the irrigation solution of 11.47 mM NO₃⁻, 1.00 mM NH₄⁺, 1.50 mM 159 H₂PO₄⁻, 6.75 mM K⁺, 3.25 mM Ca²⁺, 2.50 mM Mg²⁺ and 2.82 mM SO₄²⁻. 160 161 Microminerals were supplied by adding the following salts to the irrigation water: 50 μM H₃BO₃, 10 μM FeEDTA, 4.5 μM MnCl₂, 3.8 μM ZnSO₄, 0.3 μM CuSO₄ and 0.1 162 163 µM (NH₄)₆Mo₇O₂₄. Flowers were vibrated mechanically (to approximate the natural bee pollination syndrome of vibratile pollination; Anderson and Symon 1988) twice a week 164

165	to stimulate fruit set. For the self-incompatible wild species S. caripense, S.
166	perlongistylum and S. tabanoense (Mione and Anderson 1992; Anderson et al. 1996),
167	manual pollination using pollen from other plants from each of the species was used in
168	order to ensure fruit set. Phytosanitary treatments against spider mites (Tetranychus
169	urticae Koch.) and whiteflies (Bemisia tabaci Gennadius) were performed when
170	necessary.
171	
172	Morphological and agronomic characterization
173	
174	Individual plants were characterized using 58 primary descriptors (IPGRI and COMAV
175	2004). These descriptors include two plant (P code), seven stem (St code), 12 leaf (L
176	code), three inflorescence (I code), six flower (Fl code), 24 fruit (Fr code), and four seed
177	(Se code) traits. Eighteen traits corresponding to these primary descriptors are
178	quantitative, seven are meristic (traits in which the parts or components are counted)
179	and the other 33 traits are measured in a scale with predetermined values (Table 2).
180	
181	Molecular characterization
182	
183	Genomic DNA was extracted from young leaves of each clone according to the CTAB
184	procedure (Doyle and Doyle, 1987). DNA quality was evaluated on 0.8% agarose gels,
185	dyed with GelRed Nucleic Acid Stain (Biotium, Hayward, CA, USA) and the DNA
186	concentrations estimated using a Nanodrop ND-1000 (Nanodrop Technologies,
187	Wilmington, Delaware, USA) spectrophotometer. Extracted DNA was diluted to a
188	concentration of 20 ng/µL.

189	We used 20 simple sequence repeat (SSR) markers that proved to be
190	polymorphic in tomato (Table 3) and that are distributed throughout the tomato genome
191	(Frary et al. 2005). SSRs were amplified following the M13-tail method described by
192	Schuelke (2000) to facilitate the incorporation of a dye label during PCR.
193	Amplifications were performed in a total volume of 10 ng DNA, 1 mM MgCl ₂ , 0.05 μ M
194	of forward primer, 0.25 μ M of reverse primer, 0.2 μ M of fluorescent-labelled M-13
195	primer, 0.2 mM of dNTPs and 1 unit of <i>Taq</i> polymerase in 1X PCR buffer. PCR
196	amplifications were performed in a Mastercycler ep gradient S thermocycler
197	(Eppendorf, Hamburg, Germany) using the following programme: 1 cycle for 2 min at
198	94 °C, 35 cycles of 15 s at 94°C, 30 s at annealing temperature (Table 3), 45 s at 72 °C,
199	followed by 10 min extensive at 72 °C. SSR alleles were resolved on an ABI PRISM
200	3100 DNA (Applied Biosystems, Carlsbad, California, USA) genetic analyzer using
201	GeneScan 3.7 (Applied Biosystems) software and precisely sized using GeneScan 500
202	LIZ molecular size standards with genotyper 3.7 (Applied Biosystems) software.
203	
204	Data analysis
205	

Range and mean values for the morphological descriptors for the 14 accessions of 206 207 cultivated pepino and for the eight accessions of its wild relatives, as well as for the six 208 local varieties and eight modern cultivars of cultivated pepino, were calculated using 209 average values for each accession. Significance of differences among groups (cultivated pepino vs. wild species, and local varieties vs. modern cultivars) was tested using 210 211 Student's t tests. A principal components analysis (PCA) was performed for standardized morphological data using pairwise Euclidean distances among accessions. 212 Monomorphic traits were excluded from the PCA analysis. 213

214	For the molecular (SSR) data, the number of alleles and of private alleles for
215	each of the groups considered (all accessions, all cultivated accessions, local varieties,
216	modern cultivars, and wild relatives) were calculated. The polymorphism information
217	content (PIC) for each SSR marker was calculated as indicated Botstein et al. (1980).
218	Observed heterozygosity (H_o) was calculated for each accession. Pairwise genetic
219	similarities among accessions were calculated using the codominant genetic distance
220	(Smouse and Peakall 1999). In this context, for a single-locus with four different alleles
221	$(i, j, k \text{ and } l)$ a set of squared distances are defined as $d^2(ii, ii)=0$, $d^2(ij, ij)=0$, $d^2(ii, ij)=1$,
222	$d^2(ij, ik)=1$, $d^2(ij, kl)=2$, $d^2(ii, jk)=3$, and $d^2(ii, jj)=4$. In order to obtain the genetic
223	distance between two accessions, genetic distances are summed across loci under the
224	assumption of independence (Smouse and Peakall 1999). A principal coordinates
225	analysis (PCoA) was performed using pairwise genetic similarities. Total genetic
226	diversity (H_T), among groups genetic diversity (D_{ST}), within groups genetic diversity
227	(H _S), relative magnitude of genetic differentiation (G _{ST}) and standardized G _{ST} (G' _{ST})
228	were calculated according to Nei (1973). Correlations between morphological and
229	molecular distances were investigated with a Mantel (1967) test.
230	
231	Results
232	
233	Morphological characterization
234	
235	A wide morphological diversity was found in the collection (Figure 1). Fifty-five out of
236	the 58 morphological descriptors evaluated were variable in the collections studied. The
237	three morphological traits which were not variable were Fr-Stripes (all clones bore fruits
238	with stripes), Fr-Locules (all clones bore fruits with two locules), and Se-Type (all

clones had seeds with no wings). Furthermore, when considering only the cultivated
materials, Fl-CorollaShape was also monomorphic (all clones had rotate a corolla).

241

242 Differences between cultivated and wild clones

243

244 Significant differences were found between the cultivated pepino and wild 245 relatives for 26 traits (Table 4). On average, the cultivated pepino is less tall than the 246 wild relatives, with significantly lower values for traits related to plant size (P-Size, St-LengthInfl1, St-InternLength or I-LeavesInfl1). The cultivated pepino plants are 247 248 characterized by: more root protuberances at the stem nodes (St-Protuberances), less pubescence (St-Pubescence), fewer divided leaves (L-Type) (i.e., fewer compound, and 249 more simple leaves) and more bifurcated (I-Type) inflorescences than the wild relatives 250 251 (Table 4). Regarding sexual reproduction traits, the cultivated pepino has less style 252 exsertion (FI-StyleExsertion), lower pollen production (FI-PollenProd) and fewer seeds 253 per fruit (Se-SeedsFruit) than wild relatives. Many differences are found for fruit traits; 254 in particular cultivated pepinos are not surprisingly larger (Fr-Length, Fr-Width, Fr-PlacentLength, Fr-PlacentBreadth), have more luminous (Fr-L*), yellow (Fr-b*) and 255 256 glossy (Fr-Glossiness) skin, and more yellow (Fr-FleshColour), and better tasting (Fr-257 Flavour and Fr-OffFlavour) flesh, although with less soluble solids content (Fr-Soluble 258 Solids), than the wild relatives (Table 4). However, the range of variation within cultivated pepinos and related wild species was generally large and overlapped for all 259 260 but six traits, of which three were related to fruit size (Fr-Length, Fr-Width, Fr-PlacentLength), two to fruit taste (Fr-Flavour and Fr-SolubleSolids), and the remaining 261 262 one to the number of seeds per fruit (Se-SeedsFruit) (Table 4).

263

266	Local pepino varieties differed significantly from modern cultivars for 13 traits
267	(Table 5). However, despite the significance of differences in the averages of the two
268	categories of cultivated pepinos for these traits, the range of variation for all traits of
269	local cultivars and modern varieties overlapped. Local varieties, on average, had more
270	pigmented stem and leaves (St-Colour and L-AnthVeins) and shorter internode length
271	(St-InternLength) than modern varieties. Most modern varieties had simple leaves,
272	while local varieties mostly had compound and flat leaves, which resulted in differences
273	among both groups for several leaf shape and type traits (L-LaminaWidth, L.LWRatio,
274	L-Type, L-Leaflets, L-Surface) (Table 5). Modern varieties had, on average, greater
275	pollen production (Fl-PollenProd) and a larger number of seeds (Se-SeedsFruit) than
276	local varieties. Also, fruits of modern varieties were, on average larger and more
277	elongated (Fr-Length and Fr-LW Ratio), and had a higher intensity of green colour (Fr-
278	a*) than local varieties.
279	
280	Principal components analysis
281	
282	The first and second components of the PCA performed with all accessions accounted,

respectively, for 29.7% and 11.8%, of the total variation among accession means. The

284 first component was positively correlated with plant size vigour and growth traits (P-

Size, St-LengthInfl1, St-InternLength, I-LeavesInfl1), high pollen and seed production

286 (Fl-PollenProd and Se-SeedsFruit), and with fruits having off-flavour (Fr-OffFlavour)

and high soluble solids content (Fr-SolubleSolids), and negatively with the density of

288 root protuberances in the stem nodes (St-Protuberances), convex leaf surface (L-

289	Surface), multiparous inflorescences (I-Type), fruit size traits (Fr-Length, Fr-Width, Fr-
290	PlacentLength, and Fr-PlacentBreadth), fruit glossiness (Fr-Glosiness), fruit flesh with
291	no chlorophyll (Fr-FleshColour), and sweet flavour (Fr-Flavour) (Table 6). The second
292	principal component was positively correlated with anthocyanin pigmentation of plant
293	parts (St-Anthocyanins, St-Colour, L-PetioleColour, and L-AnthVeins), compound
294	leaves (L-LaminaWidth, L-Type and L-Leaflets), greater number of flowers per
295	inflorescence (I-NFlowers), more luminous (Fr-L*), less green (Fr-a*), mottled (Fr-
296	Mottling), and fasciated (Fr-Fasciation) fruits, and negatively with dropping (L-
297	Attitude), elongated (L-LWRatio) and convex (L-Surface) leaves, pigmented flowers
298	(Fl-CorollaColour) and obovoid fruits (Fr-WidestPart) (Table 6).
299	The projection of the accessions on a two-dimensional PCA plot showed that the
300	first component clearly separates wild accessions in the right part (i.e., positive values)
301	and cultivated pepino in the left part (i.e., negative values) of the graph (Figure 2). No
302	overlap was found for the first component values between cultivated pepino and wild
303	relatives. The second component clearly separates local varieties and modern cultivars
304	of cultivated pepino, so that the former plot in the upper part (i.e., positive values) of the
305	graph, while the latter plot in the lower part (i.e., negative values) (Figure 2). This
306	second component also separates the different wild species from each other. The highest
307	values belong to S. caripense, followed by the group of the morphologically similar S.
308	perlongistylum and S. catilliflorum, then by S. tabanoense, and finally by S.
309	trachycarpum (Figure 2). The PCA plot also shows that the groups of local varieties of
310	pepino and modern varieties show a considerable degree of dispersion in the PCA
311	graph. Although the four accessions of the wild S. caripense plot in the same section of
312	the PCA graph, they are distinct for the second component (Figure 2). Interestingly, the

313	local varieties originating in Chile (CH and OV) and Colombia (Co) plot close to most
314	of the modern varieties developed in Spain (SL, SR, Tu and Va) (Figure 2).
315	
316	Molecular characterization
317	
318	Out of the 20 tomato SSRs tested, 14 were found to be polymorphic. The six other SSRs
319	either did not amplify (SSR13, SSR51 and SSR136) or were monomorphic (SSR38,
320	SSR150 and SSR248).
321	
322	SSR characterization
323	
324	The 14 polymorphic SSRs amplified 57 alleles, with an average of 4.07
325	alleles/locus and a range between 2 and 8 in the collection (Table 7). When considering
326	cultivated accessions only, two of the SSRs (SSR14 and SSR66) were monomorphic,
327	and the average number of alleles per locus was 2.5, with a range between 1 and 6. The
328	number of alleles for each SSR locus for the local varieties of cultivated pepino was
329	identical to that found for all pepino accessions, except for locus SSR20, in which five
330	alleles were found instead of six (Table 7). As a result, the average number of alleles per
331	locus was very similar to that obtained for all the cultivated accessions. Modern
332	varieties have many fewer alleles per locus, with an average of 1.29, and polymorphism
333	was only found for four SSR loci, in which only two alleles were detected (Table 7).
334	For wild relatives, all SSR loci were polymorphic, except locus SSR578. The average
335	number of alleles per locus was 3.0, with up to 5 alleles being detected for loci SSR45
336	and SSR306 (Table 7). No SSR was found to be specific and universal to cultivated or

wild accessions. The average value for the *PIC* parameter of the 14 polymorphic SSRs

was of 0.4132, with a range for individual SSR loci between 0.0499 (SSR66) and
0.7021 (SSR306) (Table 7).

340	The mean value for observed heterozygosity (H_o) was 0.149, with a range
341	between 0 and 0.333 (Table 8). All the alleles were homozygous for the accessions of
342	the modern pepino cultivar, Sweet Round. Similarly, the wild accessions P-80 (S.
343	catilliflorum), P-62 (S. perlongistylum) and E-257 (S. tabanoense) were homozygous.
344	When considering average values, local varieties of cultivated pepino had the highest H_d
345	value (0.193) , while the wild relatives had the lowest (0.117) .
346	
347	Principal coordinates analysis
348	

The first and second principal coordinates of the PCoA analysis performed with 349 350 SSR data account for 26.0% and 10.6% of the total variation, respectively. The first principal coordinate clearly separated cultivated (right part of the graph) and wild (left 351 352 part of the graph) accessions (Figure 3). As occurred with the PCA for morphological 353 data, no overlap was found for the first coordinate values between cultivated pepino and wild relatives. With the exception of accession 37A, which showed highly negative 354 values for the second principal coordinate, all cultivated pepino accessions had positive 355 356 or moderately negative values for the second component (Figure 3). Regarding wild 357 relatives, the second principal coordinate clearly separated two groups of wild relatives, one formed by S. caripense and S. tabanoense, with positive values for the second 358 coordinate, and another one formed by S. catilliflorum, S. perlongistylum and S. 359 360 trachycarpum, with negative values. All modern varieties clustered together in the same 361 area of the PCoA plot, while local varieties were more dispersed (Figure 3).

362

365	Total diversity (H_T) of the collection had a value of H_T =0.458, with the cultivated
366	pepino having a H_T =0.237 and wild relatives a H_T =0.458 (Table 9). The among-groups
367	diversity (D_{ST}) between cultivated pepino and wild relatives had a value of D_{ST} =0.107,
368	resulting in a relative magnitude of genetic differentiation (G_{ST}) value of G_{ST} =0.274 and
369	a standardized G_{ST} value (G'_{ST}) of $G'_{ST}=0.430$ (Table 9). When comparing the local
370	varieties and modern cultivars of pepino, the total diversity of local varieties was much
371	higher (H_T =0.336) than that of modern varieties (H_T =0.096), with the among groups
372	diversity being relatively very low (D_{ST} =0.021), resulting in low values of G_{ST} (0.047)
373	and G'_{ST} (0.089) (Table 9).
374	
375	Correlation between morphological and genetic distances
376	
376 377	Correlations obtained from the Mantel test between the matrices of morphological and
	Correlations obtained from the Mantel test between the matrices of morphological and genetic distances were high (r=0.673). The graphical representation of the relationships
377	
377 378	genetic distances were high (r=0.673). The graphical representation of the relationships
377 378 379	genetic distances were high (r=0.673). The graphical representation of the relationships between morphological and genetic distances shows that for both distances the values
377 378 379 380	genetic distances were high (r=0.673). The graphical representation of the relationships between morphological and genetic distances shows that for both distances the values between local varieties are generally higher than those of modern varieties (Figure 4).
377 378 379 380 381	genetic distances were high (r=0.673). The graphical representation of the relationships between morphological and genetic distances shows that for both distances the values between local varieties are generally higher than those of modern varieties (Figure 4). For the wild species, there was a wide range of morphological and genetic distances,
377 378 379 380 381 382	genetic distances were high (r=0.673). The graphical representation of the relationships between morphological and genetic distances shows that for both distances the values between local varieties are generally higher than those of modern varieties (Figure 4). For the wild species, there was a wide range of morphological and genetic distances, with the lowest values for both distances being between <i>S. caripense</i> accessions. When
377 378 379 380 381 382 383	genetic distances were high (r=0.673). The graphical representation of the relationships between morphological and genetic distances shows that for both distances the values between local varieties are generally higher than those of modern varieties (Figure 4). For the wild species, there was a wide range of morphological and genetic distances, with the lowest values for both distances being between <i>S. caripense</i> accessions. When comparing accessions of local varieties and modern cultivars of the pepino, it became

distances between cultivated (local varieties and modern cultivars) and wild accessionswere high (Figure 4).

389

390 Discussion

391

392 A combination of morphological and molecular data provides relevant complementary and synergistic information of great interest for plant breeders and for germplasm 393 394 curators, in particular for those working with emerging crops (Rao and Hodgkin 2002; Khoury et al. 2010; Rodríguez-Burruezo et al. 2011; Yildiz 2014). In the case of the 395 396 pepino, a standardized morphological descriptors list is available (IPGRI and COMAV 2004), but the descriptors previously have not been validated or used for the 397 398 characterization of a diverse germplasm collection of pepino. We have demonstrated 399 that most of the IPGRI and COMAV (2004) descriptors used are variable (95% for the 400 whole collection and 93% for cultivated pepino). This allows the acquisition of multiple 401 characterization (i.e., phenomics) data of agronomic interest in the pepino and wild 402 relatives for a precise morphological description. Among the few non-variable traits, some are of relevance for the taxonomic discrimination, like the type of seed (Se-Type), 403 404 which is specific for discrimination between the species used here and other wild 405 relatives of Solanum section Basarhtrum (Anderson 1979), or in the case of the 406 cultivated pepino, the corolla shape (Fl-CorollaShape) which is rotate, while in the wild 407 S. tabanoense is stellate (Anderson 1975). 408 Regarding molecular data, SSR markers are preferred to other molecular markers for the standardized characterization of germplasm (Ghislain et al. 2009; 409 410 Vilanova et al., 2014) as, among other properties, they are highly repeatable, co-

411 dominant, and allow an adequate discrimination among closely related materials (Kalia

2011). Because there are no SSR markers available for the pepino, we tested tomato 412 413 EST-SSRs for transferability, given that the pepino and tomato are phylogenetically 414 close relatives (Spooner et al. 1993; Särkinen et al. 2013), indicated conclusion 415 supported as well by the viable somatic hybrids between the two species that have produced flowers and fruits (Sakomoto and Taguchi 1991). Our results show that a 416 large proportion (70%) of tomato EST-SSRs are transferrable and polymorphic in the 417 418 pepino collection studied. Furthermore, considerable SSR variation has been detected in 419 the collections of pepino and wild relatives studied, with an average number of alleles and PIC values almost as high as the values obtained for a highly variable tomato 420 421 germplasm collection that included wild relatives (Frary et al. 2005). This indicates that the large set of SSRs available in tomato (Frary et al. 2005; Suresh et al. 2014) 422 423 represents a genomic tool of interest for pepino characterization and breeding, as well as 424 for mapping and synteny studies.

425 The morphological characterization results reveal that the pepino and its close 426 wild relatives are notably variable but clearly distinct, with significant differences for 427 average values for almost one half of the descriptors evaluated and a clear separation in the PCA analysis. The domestication syndrome in the case of the pepino includes larger 428 429 fruits and very variable for fruit shape (i.e., the organ for which it is cultivated – 430 illustrating one of Darwin's conclusions about domesticates: the greatest variation in cultivated plants will be in that feature for which they are cultivated) that are more 431 luminous, glossy and yellow and more compact plants (Anderson et al. 1996; Prohens et 432 433 al. 1996). However, we have also found important changes in reproductive traits, like an increased number of root protuberances at the nodes (that facilitate vegetative 434 435 reproduction), shorter styles (that facilitate selfing), a reduction in pollen production (that may accompany the selfing syndrome, or vegetative reproduction) and fewer seeds 436

per fruit. The fact that pepino is vegetatively propagated probably favoured the selection 437 438 of parthenocarpic materials (Prohens et al. 1998), which means that traits that promote effective sexual reproduction are released from selection. Cultivated pepinos also offer 439 440 a better perceived flavour, probably resulting for a selection for lower acidity and lack of off-flavour (Prohens et al. 2005). But, pepino cultigens also have a lower content in 441 soluble solids content (Rodríguez-Burruezo et al. 2003a), which is undesirable for 442 443 producing sweet tasting fruits, obviously highly desirable in the marketplace 444 (Rodríguez-Burruezo et al. 2011). As in other crops, selection for yield may have brought a reduction in the concentration of sugars due to the "dilution effect" associated 445 446 to high yields (Davis 2009). However, it has been demonstrated that it is possible to obtain backcrosses resembling the cultivated pepino with interspecific hybrids derived 447 from S. caripense and S. tabanoense. Such hybrids have high yield and soluble solids 448 449 content levels higher than those of the cultivated recurrent parent, suggesting that these 450 wild species contain genes not present in the cultivated species that can be useful for 451 improving the soluble solids content of pepino (Rodríguez-Burruezo et al. 2003a, 2011). 452 The local varieties and modern cultivars of pepinos also differ by a number of significant morphological differences, and, as a consequence, theycluster in different 453 454 areas of the PCA diagram. Breeding for higher yield and fruit typologies adapted to 455 markets has resulted in modern varieties with larger and more elongated fruits. The elongated fruits may be constitute a selection for shipping: they pack better in layers in 456 boxes, which may result in fewer bruises than in round fruits. Also, modern varieties 457 458 have a higher production of pollen and higher number of seeds per fruit, probably as a result of selection for higher yield under conditions that may not favour expression of 459 460 parthenocarpy. Oddly, and surprisingly, although markets favor golden yellow fruits (Rodríguez-Burruezo et al. 2011), modern varieties have a greener (a* parameter) skin 461

colouration than local varieties. In tomato, enhancing chloroplast development in the 462 463 fruit increases sugar contents in fruit (Cocaliadis et al. 2014), and if the same occurs in pepino this might be the underlying reason for which breeders have unconsciously 464 465 selected for fruits with a greener skin. However, this hypothesis remains to be tested. The high morphological diversity observed in the collections studied is matched 466 467 by high levels of molecular diversity. A high level of molecular diversity was already 468 observed for AFLP and DNA sequence of a nuclear gene (Blanca et al. 2007). The EST-469 SSR markers evaluated are scattered over the genome of tomato and may constitute a good representation of different regions of the genome of pepinos as well, if the high 470 471 degree of synteny exists between the two closely related crops (Peters et al. 2012). The results reveal that cultivated pepino clones manifest a considerable heterozygosis, which 472 is expected as a high degree of heterozygosis is associated with heterosis for yield 473 474 (Rodríguez-Burruezo et al. 2003b). Heterozygosis for DNA sequence data had already 475 been observed by Blanca et al. (2007) in some pepino clones and wild relatives. In the 476 case of modern varieties, despite the lower heterozygosity compared to local varieties, 477 the level of observed heterozygosis has been similar to that of local varieties. This may be taken as evidence that breeders have selected for highly heterozygous individuals in 478 the modern breeding programs. The Sweet Round variety, which has been the only 479 480 modern cultivar homozygous for the 14 loci scored must be heterozygous for other loci as it does not breed true (Ruiz et al. 1997). With the exception of S. caripense, wild 481 relatives present low levels of observed heterozygosity. This is probably caused by the 482 483 fact that many populations of wild species of *Basarthrum* other than the widespread S. caripense are composed of few individuals (Anderson 1975, 1979), which favours 484 485 fixation of alleles, even despite the self-incompatibility of some of these species, like S. perlongistylum and S. tabanoense (Mione and Anderson 1992; Anderson et al. 1996). 486

Wild relatives show greater molecular diversity than the cultivated pepinos 487 488 (Blanca et al. 2007). In addition the genetic differentiation between the cultivated and wild materials was quite high (G_{ST} =0.274 and G'_{ST} =0.430), indicating that wild relatives 489 490 contain a large diversity that is not represented in the genetic background of the cultivated pepino. This suggests that wild relatives constitute an important source of 491 492 variation for pepino breeding (Rodríguez-Burruezo et al. 2003a; Blanca et al. 2007). 493 Local varieties of pepino show much greater genetic diversity than modern varieties, but 494 their differentiation was very low (G_{ST} =0.047 and G'_{ST} =0.089), indicating that the genetic diversity of the modern varieties is mostly present in the local varieties. This is 495 496 expected as modern varieties have been derived by selection of segregating generations derived from local varieties (Dawes and Pringle 1984; Simms et al. 1996; Ruiz et al. 497 498 1997; Prohens et al. 2002; Rodríguez-Burruezo et al. 2004a, 2004b; Levy et al. 2006). 499 Also, in contrast to tomato (Lin et al. 2014), no modern pepino cultivars have been 500 released incorporating artificially introgressed traits from wild relatives, which increases 501 genetic diversity of modern cultivars. The low diversity present in the modern varieties 502 indicates that, as occurred in many crops (Cooper et al. 2001), a genetic bottleneck has taken place during the selection and hybridization programmes performed by breeders. 503 504 Our data confirm the information provided by breeders (Dawes and Pringle 1984; 505 Simms et al. 1996; Ruiz et al. 1997; Prohens et al. 2002; Rodríguez-Burruezo et al. 506 2004a, 2004b; Levy et al. 2006) indicating that they have mostly used local varieties 507 from the peripheral southern (Chile) range of distribution of pepino, where the diversity 508 is much lower than in the center of diversity of the crop in Ecuador, southern Colombia and northern Peru (Anderson et al. 1996; Blanca et al. 2007). In fact in the PCoA 509 510 analysis, the local varieties closest to the modern varieties cluster are those from Chile.

Thus, different results might be expected with different selections of pepino cultivarsand (particularly) with different *S. caripense* wild collections.

513

514 Conclusions

515

The characterization using the IPGRI and COMAV (2004) morphological descriptors 516 list and tomato SSRs molecular markers (Frary et al. 2005) has revealed a large 517 518 variation in the collection studied. These characterization tools will allow the identification of new sources of morphological and genetic variation in pepino and wild 519 520 relatives, the study of diversity and establishment of the relationships in pepino and wild relatives. Cultivated pepino and wild relatives display high morphological and 521 molecular diversity, but the two groups are clearly differentiated from each other. 522 523 Modern cultivars are notably morphological different from local varieties, and are much 524 less variable at the molecular level indicating the existence of a genetic bottleneck 525 during the modern breeding history of this crop. All of these data are of relevance for 526 modern and efficient pepino breeding based on phenotypic and molecular marker selection as well as for the management and conservation of pepino germplasm 527 collections. 528 529 530 References 531 532 Abouelnasr H, Li YY, Zhang ZY, Liu JY, Li SF, Li W, Yu JL, McBeath JH, Han CG (2014) First report of Potato virus H on Solanum muricatum in China. Plant Dis 98:1016 533 534 Anderson GJ (1975) The variation and evolution of selected species of Solanum section Basarthrum. Brittonia 27:209-222 535

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- **Table 1** Plant materials used for the study of morphological and molecular (SSR)
- variation in a germplasm collection of local varieties and modern cultivars of cultivated
- 670 pepino (*S. muricatum*) and wild relatives (other species of *Solanum* section
- 671 *Basarthrum*).

Accession	Code	Species	Origin ^a
Pepino local variet	ies		
37-A	37	S. muricatum	Ecuador (Azuay)
Col-1	Co	S. muricatum	Colombia
CH2-22	СН	S. muricatum	Chile
OV-8	OV	S. muricatum	Chile (Limarí)
PT-154	PT	S. muricatum	Peru
RP-1	RP	S. muricatum	Ecuador
Pepino modern cul	tivars		
El Camino	EC	S. muricatum	New Zealand
Kawi	Ka	S. muricatum	New Zealand
Puzol	Pu	S. muricatum	Spain
Quito	Qu	S. muricatum	United Kingdom
Sweet Long	SL	S. muricatum	Spain
Sweet Round	SR	S. muricatum	Spain
Turia	Tu	S. muricatum	Spain
Valencia	Va	S. muricatum	Spain
Wild relatives			
BIRM/S 1034	c 1	S. caripense	Ecuador
E-7	c2	S. caripense	Ecuador (Pichincha)
EC-40	c3	S. caripense	Ecuador (Loja)
QL-013	c4	S. caripense	Ecuador (Cayambe)
P-80	ct	S. catilliflorum	Peru (Abancay)
P-62	pe	S. perlongistylum	Peru (La Mar)
E-257	ta	S. tabanoense	Ecuador (Loja)
E-34	tr	S. trachycarpum	Ecuador (Cotopaxi)

^aOrigin refers to the country and province (when known) of the collection in the case of

wild relatives and local varieties of pepinos, and to the country where the modern

674 cultivar of the pepino was developed.

675 **Table 2** Morphological and agronomic descriptors used for the characterization of

676 cultivated pepino (*S. muricatum*) and wild relatives. Full details on each descriptor can

Descriptor	Code	Range (scale) / units
Pla	nt descriptors (P)	
Plant size	P-Size	1-9 (3=small; 7=large)
Vigour of the plant	P-Vigour	1-9 (3=weak; 7=strong)
Ster	m descriptors (St)	
Stem length at first inflorescence	St-LengthInfl1	cm
Degree of ramification	St-Ramification	1-9 (3=low; 7=high)
Intensity of anthocyanin of shoot tip	St-Anthocyanin	0-9 (0=absent; 7=strong)
Root protuberances at the node	St-Protuberances	0-9 (0=absent; 7=many)
Stem pubescence density	St-Pubescence	0-9 (0=glabrous; 7=dense)
Stem colour	St-Colour	1-5 (1=green; 5=dark purple
Internode length	St-InternLength	cm
Lea	af descriptors (L)	
Petiole length	L-PetioleLength	mm
Petiole colour	L-PetioleColour	1-5 (1=green; 5=dark purple
Foliage density	L-Density	1-9 (3=sparse; 7=dense)
Leaf attitude	L-Attitude	1-3 (1=semi-erect;
		3=dropping)
Leaf lamina length	L-LaminaLength	cm
Leaf lamina width	L-LaminaWidth	cm
Leaf blade length/width ratio	L-LWRatio	
Type of leaves	L-Type	1-2 (1=simple; 2=compound
Number of leaflets	L-Leaflets	
Leaf colour	L-Colour	1-5 (1=light green; 5=purple
Anthocyanin coloration of leaf veins	L-AnthVeins	1-9 (3=green; 7=purple)
Leaf surface attitude	L-Surface	1-9 (3=flat; 7=very convex)
Inflore	scence descriptors ((I)
Number of leaves from ground to firs	tI-LeavesInfl1	
inflorescence		
Inflorescence type	I-Type	1-3 (1=generally uniparous;
		3=generally multiparous)
Number of flowers per inflorescence	I-NFlowers	
	van daganintang (El)	
Flow	ver descriptors (Fl)	

be consulted elsewhere (IPGRI and COMAV 2004).

Corolla colour	Fl-CorollaColour	1-6 (1=white; 6=purple)
Sepal length	Fl-SepalLength	mm
Stamen length	Fl-StamenLength	mm
Style exsertion beyond anther cone	Fl-StyleExsertion	
Pollen production	Fl-PollenProd	0-9 (0=none; 7=high)
-	it descriptors (Fr)	
Number of fruits per infructescence	Fr-FruitInfruct	
Number of fruits per plant	Fr-FruitPlant	
Fruit size uniformity	Fr-Uniformity	1-9 (3=low; 7=high)
Fruit length	Fr-Length	cm
Fruit width	Fr-Width	cm
Position of the widest part of the fruit	Fr-WidestPart	1-9 (3=less than ¹ / ₄ way from
1		base to tip; 7=more than $\frac{1}{2}$
		way from base to tip)
Fruit length/width ratio	Fr-LWRatio	
Fruit primary colour L* parameter	Fr-L*	
Fruit primary colour a* parameter	Fr-a*	
Fruit primary colour b* parameter	Fr-b*	
Fruit stripes	Fr-Stripes	0-1 (0=absent; 1=present)
Fruit mottling	Fr-Mottling	0-1 (0=absent; 1=present)
Fruit surface covered by additional	Fr-AddColour	1-3 (1=less than 10%;
colour		3=between 30 and 50%)
Fruit epidermis glossiness	Fr-Glossiness	3-7 (3=dull; 7=bright)
Number of locules per fruit	Fr-Locules	
Inner placental area length	Fr-PlacentLength	cm
Inner placental area breadth	Fr-PlacentBreadth	ı cm
Inner placental length/breadth ratio	Fr-PlacentLBRatie	0
Fruit flesh colour	Fr-FleshColour	1-8 (1=dark green; 8=salmon)
Fruit flavour	Fr-Flavour	1-9 (3=acidic; 9=sweet)
Presence of bitter off-flavour	Fr-OffFlavour	0-9 (0=absent; 7=strong)
Fruit cracking	Fr-Cracking	0-9 (0=absent; 9=severe)
Fruit fasciation	Fr-Fasciation	0-9 (0=absent; 9=severe)
Fruit soluble solids content	Fr-SolubleSolids	%
See	d descriptors (Se)	
Number of seeds per fruit	Se-SeedsFruit	
Seed colour	Se-Colour	1-7 (1=white; 7=black)
Seed diameter	Se-Diameter	1-3 (1=small (<1.5 mm);
		3=large (>2.5 mm))
Type of seed	Se-Type	1-3 (1=not winged; 3=winged)

679	Table 3 EST-SSR tomato markers used in the present study along with their repeat
680	motif, annealing temperature, expected size, and linkage group in which they map in the

681 tomato genetic	map (Frary et al. 2005).
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SSR locus	Repeat motif	Annealing	Expected size	Linkage
		temperature		group
SSR13	(AAG) ₆	50	102	5
SSR14	(ATA)9	55	166	3
SSR20	(GAA) ₈	50	157	12
SSR38	(TCT) ₈	55	237	8
SSR43	(TAC) ₇	55	237	4
SSR45	(AAT) ₁₄	50	246	7
SSR51	(ACAA) ₆	50	148	1
SSR52	(AAC)9	50	202	7
SSR66	$(ATA)_8$	50	185	2
SSR80	(TTTCAA)2(GTACAA)2(CAA)7	50	186	11
SSR111	$(TC)_6(TCTG)_6$	50	188	3
SSR128	(CAG) ₆ (CAA) ₃ (CAG) ₇	50	123	6
SSR136	(CAG) ₇	50	149	11
SSR150	(CTT) ₇	50	217	1
SSR248	(TA) ₂₁	55	251	10
SSR285	$(TTAT)_2(AT)_6$	55	276	7
SSR306	(ATT)7	55	258	4
SSR578	(AAC) ₆ (ATC) ₅	55	294	6
SSR590	$(TC)_{6}(AC)_{4}$	55	161	5
SSR593	(TAC) ₇	55	295	4

683	Table 4 Mean	and range for th	e morphological	descriptors for	which significant
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- 684 differences were found between accessions of the cultivated pepino (*S. muricatum*) and
- 685 its wild relatives.

	Cultivat	ed species	Wild r	elatives	
Descriptor ^a	Mean	Range	Mean	Range	Prob. t
N		14		8	
P-Size	4.9	3.4-6.6	6.5	5.0-7.0	0.0014
St-LengthInfl1	51.9	43-71	101.8	63-144	< 0.000
St-Protuberances	4.6	3.0-7.0	2.3	0.0-3.0	0.0002
St-Pubescence	2.7	0.0-3.0	4.7	0.0-7.0	0.0067
St-InternLength	5.3	4.2-6.0	7.5	4.3-9.3	0.0001
L-LaminaLength	31.7	25-37	26.9	20-34	0.0180
L-LWRatio	1.8	1.0-3.0	1.2	0.8-2.2	0.0469
L-Type	1.4	1.0-2.0	1.9	1.0-2.0	0.0077
L-Surface	4.7	3.0-7.0	3.4	3.0-5.0	0.0026
I-LeavesInfl1	11.6	8-17	16.8	13-19	0.0001
I-Type	2.6	1.0-3.0	1.4	1.0-3.0	0.0008
Fl-StyleExsertion	2.8	1.4-3.9	3.9	1.3-5.2	0.0223
Fl-PollenProd	3.4	0.0-5.4	5.7	5.0-7.0	0.0007
Fr-Uniformity	5.0	3.0-6.2	5.9	5.0-7.0	0.0249
Fr-Length	9.1	4.8-15.4	2.9	1.7-4.6	< 0.000
Fr-Width	7.2	4.1-11.1	2.7	1.8-3.6	< 0.000
Fr-L*	60.3	51-65	54.7	40-63	0.0495
Fr-b*	23.6	17-29	18.7	8-24	0.0241
Fr-Glossiness	4.5	3.0-5.7	3.3	3.0-5.0	0.0039
Fr-PlacentLength	5.1	2.2-9.7	1.4	0.6-2.1	0.0012
Fr-PlacentBreadth	0.68	0.2-1.8	0.15	0.1-0.2	0.0020
Fr-FleshColour	5.2	3.0-6.7	2.8	2.0-4.0	0.0001
Fr-Flavour	5.8	5.0-7.0	2.0	1.0-3.0	< 0.000
Fr-OffFlavour	0.66	0.0-3.0	2.75	0.0-5.0	0.0037
Fr-SolubleSolids	6.6	4.9-7.7	9.7	7.8-11.4	< 0.000
Se-SeedsFruit	0.26	0.0-0.7	3.08	1.0-4.0	< 0.000

^aSee Table 2 for a full definition of the descriptors.

687	Table 5 Mean	and range for the	e morphological	descriptors for	which significant
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688 differences were found between local varieties and modern cultivars of cultivated

	Local	varieties	Modern cultivars	cultivars	Prob. t
Descriptor ^a	Mean	Range	Mean	Range	
N		6		8	
St-Colour	3.1	2.0-4.0	2.2	2.0-3.4	0.0195
St-InternLength	4.8	4.2-5.4	5.6	5.1-6.0	0.0024
L-LaminaWidth	24.6	16-34	16.5	11-31	0.0354
L-LWRatio	1.3	1.0-1.7	2.3	1.1-3.0	0.0082
L-Type	1.7	1.3-2.0	1.2	1.0-1.5	0.0057
L-Leaflets	3.0	1.0-5.0	1.6	1.0-3.0	0.0269
L-AnthVeins	4.3	3.0-5.0	3.3	3.0-3.8	0.0196
L-Surface	4.1	3.0-5.4	5.2	4.6-7.0	0.0478
Fl-PollenProd	2.5	0.0-4.6	4.1	3.0-5.4	0.0402
Fr-Length	6.8	4.8-7.9	10.8	6.6-15.5	0.0185
Fr-LWRatio	1.0	0.7-1.8	1.6	0.9-2.2	0.0382
Fr-a*	-3.3	-6.11.3	-6.4	-11.73.1	0.0393
Se-SeedsFruit	0.10	0.0-0.4	0.38	0.0-0.7	0.0223

689 pepino (*S. muricatum*).

^aSee Table 2 for a full definition of the descriptors.

691

Table 6 Correlation coefficients between morphological descriptors and the two first

694 components (29.7% and 11.8% of the total variance explained by the first and second

695 principal components, respectively) for accessions evaluated of the cultivated pepino

Descriptor ^a	First principal component	Second principal component
P-Size	0.172	
St-LengthInfl1	0.225	
St-Anthocyanins		0.181
St-Protuberances	-0.178	
St-Colour		0.227
St-InternLength	0.188	
L-PetioleColour		0.280
L-Attitude		-0.190
L-LaminaWidth		0.270
L-LWRatio		-0.235
L-Type		0.201
L-Leaflets		0.249
L-AnthVeins		0.155
L-Surface	-0.178	-0.180
I-LeavesInfl1	0.199	
I-Type	-0.159	
I-NFlowers		0.217
Fl-CorollaShape	0.198	
Fl-CorollaColour		-0.185
Fl-PollenProd	0.163	
Fr-Length	-0.196	
Fr-Width	-0.220	
Fr-WidestPart		-0.160
Fr-L*		0.152
Fr-a*		0.259
Fr-Mottling		0.164
Fr-Glossiness	-0.183	
Fr-PlacentLength	-0.178	
Fr-PlacentBreadth	-0.172	
Fr-FleshColour	-0.186	
Fr-Flavour	-0.224	
Fr-OffFlavour	0.159	
Fr-Fasciation		0.247
Fr-SolubleSolids	0.184	
Se-SeedsFruit	0.211	

and wild relatives. Only those correlations with absolute values ≥ 0.15 have been listed.

^aSee Table 2 for a full definition of the descriptors.

Table 7 SSR markers successfully amplified and polymorphic in the collection of

699 cultivated pepino and wild relatives evaluated, number of alleles per SSR locus of each

⁷⁰⁰ of the groups considered and PIC value.

	Number of alleles					
SSR locus	All	All cultivated	Cultivated	Cultivated	Wild	PIC
	accessions	accessions	local	modern	relatives	
	(n=22)	(n=14)	varieties	cultivars	(n=8)	
			(n=6)	(n=8)		
SSR14	3	1	1	1	3	0.3360
SSR20	8	6	5	2	3	0.6134
SSR43	4	3	3	1	2	0.2604
SSR45	6	2	2	1	5	0.3665
SSR52	3	2	2	1	2	0.4156
SSR66	2	1	1	1	2	0.0499
SSR80	2	2	2	2	2	0.3715
SSR111	4	2	2	1	4	0.4297
SSR128	5	2	2	1	4	0.3079
SSR285	4	3	3	1	3	0.5188
SSR306	6	4	4	1	5	0.7021
SSR578	2	2	2	2	1	0.3693
SSR590	4	3	3	2	2	0.5774
SSR593	4	2	2	1	4	0.4669
Mean	4.07	2.50	2.43	1.29	3.00	0.4132

703	Table 8 Observed heterozygosity (H _o) for the polymorphic SSR loci in each of the
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accessions of cultivated pepino (*S. muricatum*) and wild relatives evaluated, and mean

705	values (±SE) for the	cultivated pepino	local varieties,	, modern cultiva	s and for wild
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relatives.

Accesion	Ho
Pepino local varieties	
37-A	0.154
Col-1	0.231
CH2-22	0.214
OV-8	0.167
PT-154	0.091
RP-1	0.300
Mean local varieties	0.193 ± 0.029
Pepino improved cultivars	
El Camino	0.154
Kawi	0.333
Puzol	0.154
Quito	0.143
Sweet Long	0.154
Sweet Round	0
Turia	0.077
Valencia	0.167
Mean improved cultivars	0.148 ± 0.033
Wild relatives	
BIRM/S 1034 (S. caripense)	0.154
E-7 (S. caripense)	0.250
EC-40 (S. caripense)	0.154
QL-013 (S. caripense)	0.286
P-80 (S. catilliflorum)	0
P-62 (S. perlongistylum)	0
E-257 (S. tabanoense)	0
E-34 (S. trachycarpum)	0.091
Mean wild relatives	0.117±0.040

708	Table 9 Total genetic diversity (H _T), among groups genetic diversity (D _{ST}), within	1
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groups genetic diversity (H_S), relative magnitude of genetic differentiation (G_{ST}) and

standardized G_{ST} (G'_{ST}) (Nei, 1973) estimated from data for the cultivated pepino (S.

711 *muricatum*) and wild relatives accessions.

Sample	H_{T}	D_{ST}	Hs	G _{ST}	G'st
size					
22	0.458	0.107	0.350	0.274	0.430
14	0.237				
8	0.401				
14	0.237	0.021	0.216	0.047	0.089
6	0.336				
8	0.096				
	size 22 14 8 14 6	size 22 0.458 14 0.237 8 0.401 14 0.237 6 0.336	size 22 0.458 0.107 14 0.237 8 0.401 14 0.237 0.021 6 0.336	size 22 0.458 0.107 0.350 14 0.237 8 0.401 14 0.237 0.021 0.216 6 0.336	size 22 0.458 0.107 0.350 0.274 14 0.237 8 0.401 14 0.237 0.021 0.216 0.047 6 0.336



- **Fig. 1** Diversity in fruit size, shape and colour in the cultivated pepino and wild
- relatives collection studied. Fruits of wild species are indicated by white arrows.

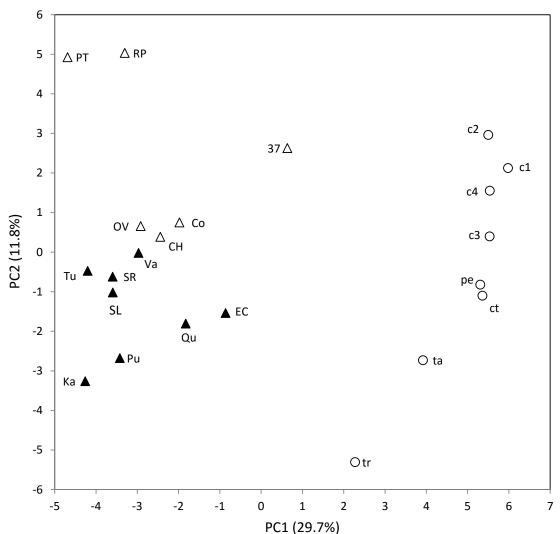


Fig. 2 Principal components analysis (PCA) similarities based on 55 variable
morphological descriptors among 22 accessions of local varieties (open triangle),
modern cultivars (solid triangle) of cultivated pepino and wild relatives (open circle).
First (PC1) and second (PC2) principal components account for 29.7% and 11.8% of the
total variation, respectively.

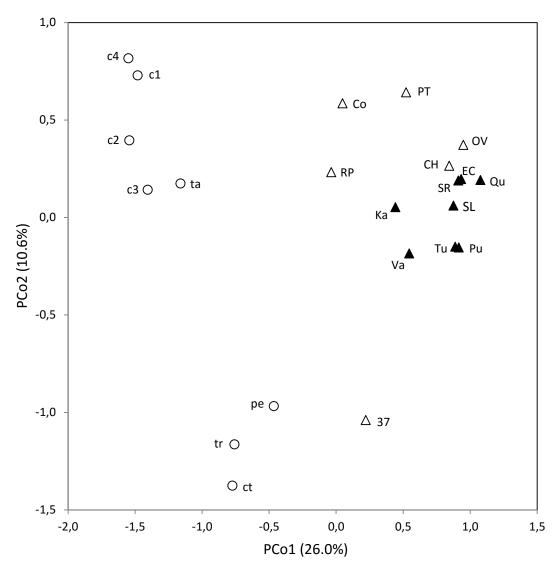


Fig. 3 Principal coordinates analysis (PCoA) similarities based on 14 polymorphic ESTSSRs among 22 accessions of local varieties (open triangle) and modern cultivars (solid
triangle) of cultivated pepino and wild relatives (open circle). First (PC1) and second
(PC2) principal coordinates account for 26.0% and 10.6% of the total variation,
respectively.

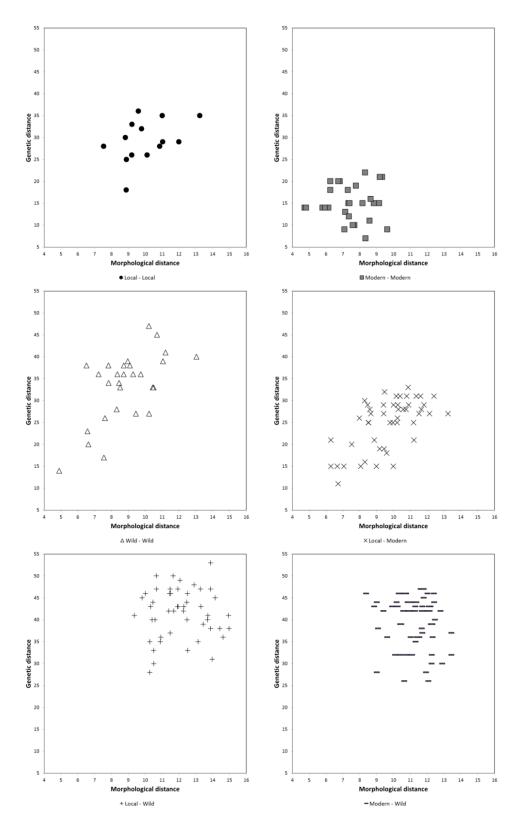


Fig. 4 Relationships between morphological and molecular distances among pairs of
accessions of pepino and wild relatives. Distances between pairs of accessions are
represented for each combination of groups: Local and local (solid circle; above left);
modern and modern (grey square; above right); wild and wild (white triangle; center
left); local and modern (× cross; center right); local and wild (+ cross; below left); and,