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Additional Information

1     **EVALUATION OF EVOH-COATED PP FILMS WITH OREGANO**  
2     **ESSENTIAL OIL AND CITRAL TO IMPROVE THE SHELF-LIFE**  
3             **OF PACKAGED SALAD**

4  
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7  
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12  
13 **ABSTRACT**

14  
15 The aim of this study was to improve the present packaging of salad by combining modified  
16 atmosphere packaging with a new antimicrobial active bag consisting of PP/EVOH film with  
17 oregano essential oil or citral, with the purpose of extending shelf-life and reducing possible  
18 microbiological risks. The (O<sub>2</sub>) and CO<sub>2</sub> barrier properties of PP/EVOH, mechanical properties  
19 (Young's modulus, tensile strength and elongation at break) were determined and compared  
20 with those of standard PP films. Antimicrobial tests were carried out for enterobacteria, total  
21 aerobic counts, yeasts and moulds, and lactic acid bacteria and psychrotrophic bacteria, and the  
22 effect of the release of the antimicrobial agent on the sensory characteristics of the salads was  
23 also studied. The application of the EVOH coating results in an increase in the tensile resistance  
24 of the PP films and a reduction in the elongation at break. The results showed that  
25 microorganism counts bacteria decreased especially at the beginning of the storage period. OEO  
26 and CITRAL samples had reductions of 1.38 log and 2.13 log respectively against  
27 enterobacterias, about 2 log against yeasts and moulds. The total aerobic counts reduced 1.08  
28 log with OEO and 1.23 log with CITRAL and the reduction of lactic acid bacteria and  
29 psychrotrophic was about 2 log.. Citral-based films appeared to be more effective than materials  
30 containing oregano essential oil in reducing spoilage flora during storage time. Sensory studies  
31 also showed that the package with citral was the most accepted by customers at the end of the  
32 shelf life

33  
34 **Keywords:** *minimally processed salads, citral, oregano essential oil, antimicrobial packaging,*  
35 *mechanical properties, EVOH.*

36

37 **1. INTRODUCTION**

38 Consumer demand for healthy, fresh-like, easy-to-prepare products coupled with consumer  
39 lifestyles changes have led to the development of a variety of novel products, of which  
40 minimally processed salads are an example. Cut vegetables are more susceptible to chemical  
41 and microbiological deterioration because during the cutting, cells are destroyed and exudates  
42 rich in minerals, sugar, vitamins, and other compounds are released. These nutrients and storage  
43 conditions may allow growth of microorganisms (Froeder, et al., 2007). During the last decades  
44 food borne outbreaks associated with consumption of raw vegetables have been increasing and  
45 green leafy vegetables seem to be the most frequently implicated products (Santos, et al., 2012).

46 In order to extend the shelf life of minimally processed products, various technologies are  
47 available including well implemented equilibrium modified atmosphere packaging (MAP)  
48 (Carrasco, Perez-Rodriguez, Valero, Garcia-Gimeno, & Zurera, 2008; Oms-Oliu, Martinez,  
49 Soliva-Fortuny, & Martin-Belloso, 2008) and more recently antimicrobial active packaging. The  
50 combination of active antimicrobial packaging and MAP is a possible alternative to enhance the  
51 quality, safety and shelf-life of products of this kind, in which antimicrobial agents added  
52 directly to the packaging material are released exerting their activity on the packaged product  
53 (Sanchez-Gonzalez, Chafer, Hernandez, Chiralt, & Gonzalez-Martinez, 2011).

54 In a previous work (Muriel-Galet, et al., 2012), oregano essential oil and citral were  
55 incorporated in an ethylene-vinyl alcohol copolymer (EVOH) matrix and a preliminary study on  
56 the antimicrobial activity of the films and on the sensory attributes of salad was reported.  
57 Results showed that the antimicrobial activity increased with concentration of the antimicrobial  
58 agent in the film but also increased the sensory perception of off-flavors. EVOH materials have  
59 been used recently as matrices for the development of active packaging systems, where the  
60 polymer triggers their activity on exposure to a humid environment (the food product) (Lopez-  
61 de-Dicastillo, Alonso, Catala, Gavara, & Hernandez-Munoz, 2010a; Lopez-de-Dicastillo,  
62 Catala, Gavara, & Hernandez-Munoz, 2011; Lopez-de-Dicastillo, Gallur, Catala, Gavara, &  
63 Hernandez-Munoz, 2010b)

64 In this work, active EVOH coated polypropylene (PP) films were manufactured using gravure  
65 printing technologies and used for modified atmosphere packaging of fresh produce in a  
66 processing plant. The aim of this study was to improve the stability of a mixed vegetable salad  
67 with the new antimicrobial active packaging that released oregano essential oil or citral  
68 combined with MAP and to characterize the films and the antimicrobial and sensory effects on  
69 salad.

70

## 71 2. MATERIALS AND METHODS

### 72 2.1 Chemicals and reagents

73

74 Ethylene vinyl alcohol copolymer with a 29% ethylene molar content (EVOH) was kindly  
75 provided by The Nippon Synthetic Chemical Company (Osaka, Japan). Oregano essential oil  
76 and citral were purchased from Sigma (Madrid, Spain). Polypropylene films were provided by  
77 Envaflex (Utebo, Zaragoza, Spain). “*Four season salad*” (a mixture of cut iceberg lettuce, red  
78 cabbage and shredded carrot) was purchased from Verdifresh (Ribarroja, Valencia, Spain).  
79 Water was obtained from a Milli-Q Plus purification system (Millipore, Molsheim, France).

80

### 81 2.2 Film preparation

82

83 A coating technology based on gravure printing was used to produce the antimicrobial material  
84 at Envaflex at a production speed of 60 m/min. Based on the results of previous work (Virginia  
85 Muriel-Galet & Hernández-Muñoz, 2012), the antimicrobial films were produced by  
86 incorporating a known concentration (7.5 % w/w) of oregano essential oil (OEO) or citral  
87 (CITRAL) in the EVOH coating solution. Polypropylene films were treated with corona and a  
88 primer based on polyethylene imine (Mica Corp., Kansas City, USA) was applied to improve  
89 adherence.. EVOH coating was applied on the PP films, keeping an area uncoated to maintain  
90 sealability for the manufacture of the salad bags. Specifically, of the 1140 cm<sup>2</sup> of film that was  
91 used to manufacture a bag, only 750 cm<sup>2</sup> of surface was coated. Control films were obtained  
92 without active agents (CONTROL). Thicknesses of the films were measured at 10 points of  
93 samples. The thickness of the coating was calculated from the difference in thickness between  
94 coated and uncoated areas.

95

### 96 2.3 Packaging and storage

97

98 “*Four season salad*” was packed in individual bags manufactured with the various films with  
99 (OEO and CITRAL) and without agents (CONTROL) as shown in Figure 1. The packaging was  
100 done at Verdifresh (Ribarroja, Valencia, Spain) with a MULTIVAC A-NG 85021 packaging  
101 unit, and with an atmospheric composition of 12% CO<sub>2</sub> and 4% O<sub>2</sub>. Bags of “*four season*  
102 *salad*” were stored for 5 days at 4°C plus 3 days at 8°C, simulating commercial conditions of  
103 production, transport and commercialization (Lopez-Galvez, Gil, Truchado, Selma, & Allende,  
104 2010).

105

106

**107 2.4 Identification and quantification of volatile compounds**

108

109 The equilibrium concentration of carvacrol (main component of oregano essential oil) and citral  
110 in the polymer samples was determined by thermal desorption and GC analysis using a thermal-  
111 desorber (890/891 model, Dynatherm Analytical Inst., Supelco, Bellafonte, PA, USA)  
112 connected in series to the gas chromatograph (Lopez-Carballo, Cava, Lagaron, Catala, &  
113 Gavara, 2005). In brief, a portion of the tested film (ca. 1cm<sup>2</sup>) was placed in the desorption cell  
114 and heated at 210 °C for 7 min. A He gas stream carried the desorbed compounds to the GC  
115 through a transfer line heated at 230 °C. The GC was equipped with a 30 m, 0.53 mm, 2.65 µm  
116 Agilent HP-1 semicapillary column. The chromatographic conditions were: He as the carrier  
117 gas, 210 °C and 260 °C injector and detector temperatures, 7 min at 45 °C, heating ramp to  
118 220 °C at 18 °C/min, and 12 min more at 220 °C. At the end of the desorption process the  
119 sample was weighed with a 0.1 mg precision balance (Voyager V11140 model, Ohaus,  
120 Switzerland). Calibration was done by injecting known amounts of carvacrol or citral in the  
121 desorption cell.

122

**123 2.5 Oxygen and carbon dioxide permeance**

124

125 The measurements of O<sub>2</sub> and CO<sub>2</sub> permeance through the film samples were carried out using  
126 isostatic methods (Lopez-de-Dicastillo, et al., 2010b). The O<sub>2</sub> permeation rates of the materials  
127 were determined at 0 and 90% RH and 23°C using an OXTRAN Model 2/21 ML Mocon  
128 (Lippke, Neuwied, Germany). The CO<sub>2</sub> permeation rates of the materials were determined at 0  
129 and 90% RH and 23°C using an assembly that makes use of a gas chromatograph as detector.

130 The evolution of headspace atmosphere composition during storage was carried out by  
131 withdrawal of 20 mL gas samples and injection on a Checkmaster gas analyzer (Lippke,  
132 Germany).

133

**134 2.6 Mechanical properties**

135

136 A Mecmesin model MultiTest 1-I universal machine (Landes Poli Ibérica, S.L., Barcelona,  
137 Spain) equipped with a 100 N static load cell was used to evaluate the maximum tensile strength  
138 ( $\sigma_m$ ), percentage of elongation at break ( $\epsilon_b$ ) and Young's modulus ( $E$ ) of the films according to  
139 ASTM standard method D882 (ASTM, 2009). The films were conditioned at 50% RH and room

140 temperature for at least 1 week prior to testing. Film samples were cut into strips 2.54 cm wide  
141 and 10 cm long. The grip separation was set at 5.08 cm and the cross-head speed at 25 mm·min<sup>-1</sup>.  
142 At least 6 replicates from each sample were tested. The tensile properties were calculated  
143 from the plot of stress (tensile force/initial cross-section area) versus strain (elongation as a  
144 fraction of the original length).

145

## 146 **2.7 Antimicrobial activity of films on microflora of minimally processed salad**

147

148 The antimicrobial activity of the films was tested *in vivo*. Analyses were performed on the 1<sup>st</sup>, 4<sup>th</sup>  
149 and 8<sup>th</sup> days after packaging.. 25-g samples of the packaged salad were weighed in sterile  
150 Stomacher bags, diluted with 25 mL of peptone water (Scharlab, Barcelona, Spain) and  
151 homogenized in Stomacher for 3 min (IUL S.L., Barcelona). Ten-fold dilution series were made  
152 in the same saline solution for plating.

153 The enumeration of particular microbial groups was performed by using the following media  
154 (Scharlab, Barcelona, Spain) and culture conditions: a) Nutrient Agar for total aerobic plate  
155 count, pour-plated and incubated at 30 °C for 48 h, and also for total aerobic psychrotropic  
156 count, pour-plated and incubated at 10 °C for 10 days; b) MEA for yeasts and moulds, pour-  
157 plated and incubated at 25 °C for 5 days; c) VRBD agar for total enterobacteria, pour-plated,  
158 overlaid with the same medium and incubated aerobically at 37 °C for 48 h; d) MRS agar for  
159 lactic acid bacteria, pour-plated and incubated at 25 °C for 5 days. The counts were performed  
160 in triplicate.

161

## 162 **2.8 Sensory analyses**

163

164 A group of 51 panellists, consumers of ready-to-eat fresh products with no previous experience,  
165 evaluated the smell, visual appearance, texture and general acceptability of the samples by  
166 paired comparison tests (ISO 5495-2005). A total of three pairs of samples were evaluated. Each  
167 pair consisted of two samples. The first pair were samples packaged with CONTROL and OEO  
168 films, the second pair were CONTROL and CITRAL, and the third pair were OEO and  
169 CITRAL.

170 Three pairs of samples were evaluated in each session and a total of two sessions was  
171 performed: on the 1<sup>st</sup> day, to determine the initial impact of the aromatic agent, and on the 8<sup>th</sup>  
172 day after packaging, to verify the quality and acceptability at the end of storage.. The tests were  
173 done in a standardized test room (ISO 8589-2007). Samples of “*four season salad*” were placed

174 in PP/EVOH bags identified by three-digit codes. The Williams method for constructing such  
175 designs was applied to the sensory studies (MacFie, Bratchell, Greenhoff, & Vallis, 1989).

176

## 177 **2.9 Statistical analysis**

178

179 Statistical analysis of the data was performed by one-way ANOVA study. The SPSS® computer  
180 program (SPSS Inc., Chicago, IL, USA) was used. Differences in pairs of mean values were  
181 evaluated by the Tukey-b test for a confidence interval of 95%. Data are represented as mean  $\pm$   
182 standard deviation. The design of the sensory evaluation and the data analysis were carried out  
183 with Compusense Five software (release 5.5, Ontario, Canada).

184

## 185 **3. RESULTS AND DISCUSSION**

186

187 PP/EVOH films containing oregano essential oil (OEO) or citral (CITRAL) were prepared as  
188 described in the experimental section. PP/EVOH film without antimicrobial agent was also  
189 obtained as control (CONTROL). The EVOH coatings had a thickness of  $1.15 \pm 0.05 \mu\text{m}$ .

190 One of the major components of oregano essential oil is carvacrol (72.46%), which is  
191 responsible for its antimicrobial properties (Burt, 2004; Charai, Mosaddak, & Faid, 1996).  
192 Because of this, carvacrol was the only component of the OEO monitored in this study.. Films  
193 with 7.5% oregano essential oil had a final carvacrol concentration of  $3.5 \pm 0.2\%$ . These results  
194 show a carvacrol loss of about 35%, which can be attributed to evaporation during the drying  
195 process. The films with citral had higher losses owing to its higher volatility. The final  
196 concentration was  $2.6 \pm 0.2\%$ .

197 The film samples obtained were stored in desiccators with silica gel. In these dry storage  
198 conditions, the concentration of the agents was maintained constant for 3 months. EVOH is  
199 characterized by its excellent gas and organic vapour barrier properties when dry, and therefore  
200 the release of carvacrol or citral is highly impeded in these storage conditions (Aucejo, Catala,  
201 & Gavara, 2000; Lopez-de-Dicastillo, et al., 2010b).

202

### 203 **3.1. Oxygen and carbon dioxide permeance**

204

205 Permeance ( $\wp$ ) to ( $\text{O}_2$ ) and ( $\text{CO}_2$ ) was evaluated under two relative humidity conditions: 0%  
206 and 90%. Table 1 shows the values for all the samples. As can be seen, the incorporation of the  
207 coating results in an increase in barrier which is very considerable in dry conditions. The



208 permeance to both O<sub>2</sub> and CO<sub>2</sub> is reduced by more than 100-fold. This effect is to be expected,  
 209 owing to the well-known high barrier properties of EVOH. Nevertheless, the effect is severely  
 210 reduced by humidity because of water plasticization of EVOH. The permeance decrease caused  
 211 by the coating process on the PP film was 1.5-fold for O<sub>2</sub> and 4-fold for CO<sub>2</sub> in the coated area.  
 212 Permeance values for the materials with the agents could not be measured because of the  
 213 sensitivity of the Oxtran sensor to volatile organic compounds and their interference with the  
 214 chromatographic detector used in the assays with CO<sub>2</sub>. Effect of the agents in the barrier  
 215 properties of the coatings should be negligible since, plasticization should not be expected from  
 216 the addition of apolar substances to a polar matrix and agents with double functionality which  
 217 could promote chain cross-linking are not present.

218 Considering that only part of the bag surface was coated, the effect of the coating on the  
 219 transmission rate was even lower. An estimation of the O<sub>2</sub> (OER) and CO<sub>2</sub> exchange rates  
 220 (CO<sub>2</sub>ER) for the entire bag at the beginning of storage can be obtained by the use of the  
 221 permeation equation for a bag constituted by two different materials, considering the permeance  
 222 values for each film ( $\rho_i$ ), the film surface ( $A_i$ ) and the pressure gradient ( $\Delta p_i$ ):

$$223 \quad OER = \sum_{i=1}^n (\rho_i(O_2) \cdot A_i) \cdot \Delta p(O_2); \quad CO_2ER = \sum_{i=1}^n (\rho_i(CO_2) \cdot A_i) \cdot \Delta p(CO_2)$$

$$224 \quad OER(\text{coated}) = 2.3 \cdot 10^{-10} \frac{m^3}{s}; \quad OER(\text{CONTROL}) = 2.9 \cdot 10^{-10} \frac{m^3}{s}$$

$$225 \quad CO_2ER(\text{coated}) = 6.5 \cdot 10^{-10} \frac{m^3}{s}; \quad CO_2ER(\text{CONTROL}) = 1.3 \cdot 10^{-9} \frac{m^3}{s}$$

227 In technological units, the gas exchange rates for a finished bag are: 25 and 20 cc/day of O<sub>2</sub> and  
 228 112 and 56 cc/day of CO<sub>2</sub> for the CONTROL and coated sample, respectively.

229 Table 2 shows the evolution of the headspace composition for the three samples tested. In all  
 230 cases, the O<sub>2</sub> was rapidly consumed as the product respired and there was a corresponding  
 231 increase in CO<sub>2</sub> levels. The presence of the EVOH coating in the active packaging did not  
 232 significantly influence the headspace composition during the storage period. EVOH generally  
 233 acts as a high barrier material to gases when dry. However, when in contact with fresh  
 234 vegetables, the EVOH layer becomes humid and the barrier properties are dramatically reduced.  
 235 Thus the EVOH layer becomes permeable and the whole package behaves similarly to the  
 236 reference one. Although the differences were not significant, it appears that the packages  
 237 containing CITRAL as active agent had a lower CO<sub>2</sub> content than the other two samples, as if  
 238 the CITRAL had an effect on the reduction in the respiration rate of the salad.

239

### 240 3.2. Mechanical properties



241

242 The mechanical parameters ( $\sigma_m$ ,  $E$  and  $\epsilon_b$ ) of the CONTROL and treated films conditioned at  
243 50% RH are shown in Table 3. As can be seen, the application of the EVOH coating results in  
244 an increase in the tensile resistance of the PP films and a reduction in the elongation at break.  
245 This effect can be explained as a consequence of the different morphology of the two materials.  
246 PP is a material with a glass transition temperature below room temperature, whilst EVOH is a  
247 semicrystalline glassy copolymer, therefore more rigid and more fragile than PP. No effect was  
248 observed as a result of incorporation of the agent in the EVOH layer and the corona-treated. The  
249 effect of the coating application would be severely diminished in humid conditions as a  
250 consequence of the plasticization of the EVOH coating. Unfortunately, no measurement could  
251 be carried out in these environmental conditions (90%RH and room temperature) with the  
252 equipment used.

### 253 **3.3 Antimicrobial activity of active films on microflora of minimally processed salads**

254

255 The samples were subjected to microbiological analysis on the 1<sup>st</sup>, 4<sup>th</sup> and 8<sup>th</sup> days of  
256 refrigerated storage. The results of microbiological counts of minimally processed salad  
257 samples are shown in Table 4.

258 On the 1<sup>st</sup> day of storage, the enterobacteria counts for “*four season salad*” packaged in the  
259 active samples showed a significant decrease in comparison with the control sample. OEO and  
260 CITRAL samples had reductions of 1.38 log and 2.13 log respectively. With storage time, there  
261 was no measurable effect. It should be noted that high levels of enterobacteria are common in  
262 raw vegetables and it is reported that they are not used to indicate the microbiological quality of  
263 fresh vegetables (Little, Roberts, Youngs, & Louvois, 1999).

264 The total aerobic counts were reduced only on the 1<sup>st</sup> day of storage; OEO films reduced count  
265 by 1.08 log and CITRAL films 1.23 log. The aerobic population increased to 5 log in all cases  
266 on the 4<sup>th</sup> day and to 6 log in all cases at the end of storage.

267 Table 4 also shows the antifungal effectiveness of the active films. The active compounds  
268 appear to be most effective on day 1. OEO and CITRAL samples had a reduction of about 2 log.  
269 At the end of the storage period, the coatings produced an inhibition of about 0.5 log. The role  
270 of yeasts and moulds in the spoilage of vegetables is not well studied, although they have been  
271 implicated in the spoilage of fermented vegetable products (Fleet, 1992)(Abadias, Usall,  
272 Anguera, Solson, & Vinas, 2008) and could cause off-odours and visual defects in minimally  
273 processed vegetables. Some authors (Tournas, 2005) have pointed out the possible health  
274 problems associated with the presence of moulds in fruits and vegetables. Their growth could be  
275 related with the quality of the salad and acceptability by the consumer.

276 The reduction of lactic acid bacteria was more significant for the films containing CITRAL,  
277 about 2 log on the 1<sup>st</sup> day of storage, and about 1 log reduction was observed on the 4<sup>th</sup> and 8<sup>th</sup>  
278 days. OEO films showed an inhibition of 0.5 log on day 1, which was just maintained during  
279 storage.

280 With respect to psychrotrophic bacteria that are able to grow at refrigerator temperature, the  
281 samples packaged with the OEO and CITRAL films showed an inhibition of approximately 2  
282 log at the beginning of storage at 4 °C. However, there was no measurable effect after 8 days of  
283 storage, because psychrotrophic bacteria counts increased at 8 °C and they reached a stationary  
284 phase before the day of analysis (data not shown).

285 It can be concluded that these active films provided inhibition of microflora bacteria in the first  
286 stages of the storage period, when the concentration of the agent in the package headspace was  
287 higher and the storage temperature lower. The intrinsic properties of the food (fat/protein/water  
288 content, antioxidants, preservatives, pH, salt and other additives) and the extrinsic determinants  
289 (temperature, packaging in vacuum/gas/air, characteristics of microorganisms) can also  
290 influence bacterial sensitivity (Tassou, Drosinos, & Nychas, 1995). The antimicrobial activity of  
291 essential oil benefits from a decrease in storage temperature and the low fat content in salads. It  
292 is generally supposed that the high levels of fat in foodstuffs protect the bacteria from the action  
293 of the EO (Pandit & Shelef, 1994; Tassou, et al., 1995), therefore these factors could contribute  
294 to the successful results observed with essential oils (Burt, 2004).

295 CITRAL, which appears to be more effective than OEO, has already been successfully used for  
296 the inhibition of yeast growth and lactic acid bacteria in fruit-based salads (Belletti, Lanciotti,  
297 Patrignani, & Gardini, 2008). CITRAL can also be used to prolong the shelf-life of “*four*  
298 *season salads*”. In this study with real foodstuffs, essential oil produced lower effects than in  
299 another test carried out “*in vitro*” in our laboratory (Muriel-Galet, et al., 2012), which can be  
300 attributed to partial sorption of the agent in the food, with a subsequent reduction in  
301 concentration in the vapour phase. A reduction in the antimicrobial activity of essential oils due  
302 to the nature of the food matrix has been documented (Belletti, et al., 2008; Burt, 2004;  
303 Lanciotti, et al., 2004; Muriel-Galet, et al., 2012). Therefore, higher concentrations of plant  
304 essential oils are generally required when added to food (Gutierrez, Barry-Ryan, & Bourke,  
305 2008). However, the application of essential oils in food may be limited by the changes they  
306 produce in the organoleptic and textural quality of the food or interactions of these compounds  
307 with food components (Devlieghere, et al., 2004).

308

### 309 3.4. Sensory analyses

310

311 The organoleptic effect of oils is one of the most important factors that limit their application to  
312 real food products as antimicrobial agents, a property widely described in *in vitro* tests (Belletti,  
313 et al., 2008). Their use may have a significant sensory impact that could result in non-  
314 acceptance by the consumer. For this reason, sensory analyses were carried out on the “*four*  
315 *season salads*” packaged with the films developed.

316 The effects of the addition of different kinds of agents on the sensory properties of smell  
317 acceptability, visual appearance, texture and general acceptability were studied. On the 1<sup>st</sup> day  
318 of storage, the assessors perceived significant differences in odour acceptability, depending on  
319 the agents (OEO or CITRAL), in comparison with the CONTROL samples, as shown in Figures  
320 2–4. In terms of visual appearance, no significant differences were observed between the  
321 samples. This can be explained by the fact that 1 storage day is not enough to cause  
322 deterioration of the sample.

323 On the 8<sup>th</sup> day of storage, no significant differences were recorded for smell, visual appearance,  
324 texture and general acceptability when the control was compared with OEO or CITRAL. Only  
325 differences between oregano essential oil and citral were found by the judges. The samples with  
326 CITRAL were clearly preferred on day 8 of storage.

327 OEO and CITRAL resulted in a pleasant flavour, compatible with final products. This is a  
328 prerequisite for the use of essential oils in foods for antimicrobial purposes.

329

#### 330 4. CONCLUSIONS

331

332 In conclusion, the use of active packaging based on the release of volatile antimicrobials  
333 combined with modified atmosphere packaging is an alternative technology to improve the  
334 microbiological and sensory stability of salad. The addition of the active EVOH coating did not  
335 substantially affect the functional properties of the packaging film, since neither barrier nor  
336 mechanical properties had unacceptable values. Nevertheless, the packaged product did produce  
337 improved microbial stability when active films were used, especially with regard to growth  
338 inhibition of enterobacteria, lactic acid and psychrotrophic bacteria, and yeasts and moulds at  
339 the beginning of storage. The sensory studies showed that the active agent release resulted in a  
340 non-typical smell which led the judges to prefer the control samples, whereas after long-term  
341 storage the samples in active bags were preferred, especially the one containing citral. The  
342 results of the present work demonstrated that packaging of minimally processed salad in MAP

343 bags manufactured with PP/EVOH with essential oils is an appropriate way to reach a balance  
344 between the demand for microbial safety and consumer acceptability.

345

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350

351

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354 **REFERENCES**

355

- 356 Abadias, M., Usall, J., Anguera, M., Solson, C., & Vinas, I. (2008). Microbiological quality of  
357 fresh, minimally-processed fruit and vegetables, and sprouts from retail  
358 establishments. *International Journal of Food Microbiology*, *123*(1-2), 121-129.
- 359 ASTM, D. (2009). Standard test method for tensile properties of thin plastic sheeting. In. In  
360 American Society for Testing Materials.
- 361 Aucejo, S., Catala, R., & Gavara, R. (2000). Interactions between water and EVOH food  
362 packaging films. *Food Science and Technology International/Ciencia y Tecnologia de*  
363 *Alimentos Internacional*, *6*(2), 159-164.
- 364 Belletti, N., Lanciotti, R., Patrignani, F., & Gardini, F. (2008). Antimicrobial efficacy of citron  
365 essential oil on spoilage and pathogenic microorganisms in fruit-based salads. *Journal*  
366 *of Food Science*, *73*(7), M331-M338.
- 367 Burt, S. (2004). Essential oils: their antibacterial properties and potential applications in foods -  
368 a review. *International Journal of Food Microbiology*, *94*(3), 223-253.
- 369 Carrasco, E., Perez-Rodriguez, F., Valero, A., Garcia-Gimeno, R. M., & Zurera, G. (2008). Growth  
370 of *Listeria monocytogenes* on shredded, ready-to-eat iceberg lettuce. *Food Control*,  
371 *19*(5), 487-494.
- 372 Charai, M., Mosaddak, M., & Faid, M. (1996). Chemical composition and antimicrobial activities  
373 of two aromatic plants: *Origanum majorana* L. and *O. compactum* Benth. *Journal of*  
374 *Essential Oil Research*, *8*(6), 657-664.
- 375 Devlieghere, F., Francois, K., Vereecken, K. M., Geeraerd, A. H., Impe, J. F. v., & Debevere, J.  
376 (2004). Effect of chemicals on the microbial evolution in foods. *Journal of Food*  
377 *Protection*, *67*(9), 1977-1990.
- 378 Fleet, G. (1992). Spoilage yeasts. *CRC Critical Reviews in Biotechnology*, *12*(1/2), 1-44.
- 379 Froeder, H., Martins, C. G., Oliveira de Souza, K. L., Landgraf, M., Franco, B. D. G. M., & Destro,  
380 M. T. (2007). Minimally processed vegetable salads: microbial quality evaluation.  
381 *Journal of Food Protection*, *70*(5), 1277-1280.
- 382 Gutierrez, J., Barry-Ryan, C., & Bourke, P. (2008). The antimicrobial efficacy of plant essential  
383 oil combinations and interactions with food ingredients. *International Journal of Food*  
384 *Microbiology*, *124*(1), 91-97.
- 385 Lanciotti, R., Gianotti, A., Patrignani, F., Belletti, N., Guerzoni, M. E., & Gardini, F. (2004). Use of  
386 natural aroma compounds to improve shelf-life and safety of minimally processed  
387 fruits. *Trends in Food Science & Technology*, *15*(3-4), 201-208.
- 388 Little, C., Roberts, D., Youngs, E., & Louvois, J. d. (1999). Microbiological quality of retail  
389 imported unprepared whole lettuces: a PHLS Food Working Group study. *Journal of*  
390 *Food Protection*, *62*(4), 325-328.
- 391 Lopez-Carballo, G., Cava, D., Lagaron, J. M., Catala, R., & Gavara, R. (2005). Characterization of  
392 the interaction between two food aroma components, alpha-pinene and ethyl  
393 butyrate, and ethylene-vinyl alcohol copolymer (EVOH) packaging films as a function of  
394 environmental humidity. *Journal of Agricultural and Food Chemistry*, *53*(18), 7212-  
395 7216.
- 396 Lopez-de-Dicastillo, C., Alonso, J. M., Catala, R., Gavara, R., & Hernandez-Munoz, P. (2010a).  
397 Improving the Antioxidant Protection of Packaged Food by Incorporating Natural  
398 Flavonoids into Ethylene-Vinyl Alcohol Copolymer (EVOH) Films. *Journal of Agricultural*  
399 *and Food Chemistry*, *58*(20), 10958-10964.
- 400 Lopez-de-Dicastillo, C., Catala, R., Gavara, R., & Hernandez-Munoz, P. (2011). Food applications  
401 of active packaging EVOH films containing cyclodextrins for the preferential scavenging  
402 of undesirable compounds. *Journal of Food Engineering*, *104*(3), 380-386.

- 403 Lopez-de-Dicastillo, C., Gallur, M., Catala, R., Gavara, R., & Hernandez-Munoz, P. (2010b).  
404 Immobilization of beta-cyclodextrin in ethylene-vinyl alcohol copolymer for active food  
405 packaging applications. *Journal of Membrane Science*, 353(1-2), 184-191.
- 406 Lopez-Galvez, F., Gil, M. I., Truchado, P., Selma, M. V., & Allende, A. (2010). Cross-  
407 contamination of fresh-cut lettuce after a short-term exposure during pre-washing  
408 cannot be controlled after subsequent washing with chlorine dioxide or sodium  
409 hypochlorite. *Food Microbiology*, 27(2), 199-204.
- 410 MacFie, H. J., Bratchell, N., Greenhoff, K., & Vallis, L. V. (1989). Designs to balance the effect of  
411 order of presentation and first-order carry-over effects in hall tests. *Journal of Sensory  
412 Studies*, 4(2), 129-148.
- 413 Muriel-Galet, V., Cerisuelo, J. P., López-Carballo, G., Lara, M., Gavara, R., & Hernández-Muñoz,  
414 P. (2012). Development of antimicrobial films for microbiological control of packaged  
415 salad *International Journal of Food Microbiology*.
- 416 Oms-Oliu, G., Martinez, R., Soliva-Fortuny, R., & Martin-Belloso, O. (2008). Effect of  
417 superatmospheric and low oxygen modified atmospheres on shelf-life extension of  
418 fresh-cut melon. *Food Control*, 19(2), 191-199.
- 419 Pandit, V. A., & Shelef, L. A. (1994). SENSITIVITY OF LISTERIA-MONOCYTOGENES TO ROSEMARY  
420 (ROSMARINUS-OFFICINALIS L). *Food Microbiology*, 11(1), 57-63.
- 421 Sanchez-Gonzalez, L., Chafer, M., Hernandez, M., Chiralt, A., & Gonzalez-Martinez, C. (2011).  
422 Antimicrobial activity of polysaccharide films containing essential oils. *Food Control*,  
423 22(8), 1302-1310.
- 424 Santos, M. I., Cavaco, A., Gouveia, J., Novais, M. R., Nogueira, P. J., Pedroso, L., & Ferreira, M.  
425 (2012). Evaluation of minimally processed salads commercialized in Portugal. *Food  
426 Control*, 23(1), 275-281.
- 427 Tassou, C. C., Drosinos, E. H., & Nychas, G. J. E. (1995). EFFECTS OF ESSENTIAL OIL FROM MINT  
428 (MENTHA-PIPERITA) ON SALMONELLA-ENTERITIDIS AND LISTERIA-MONOCYTOGENES  
429 IN MODEL FOOD SYSTEMS AT 4-DEGREES AND 10-DEGREES-C. *Journal of Applied  
430 Bacteriology*, 78(6), 593-600.
- 431 Tournas, V. H. (2005). Moulds and yeasts in fresh and minimally processed vegetables, and  
432 sprouts. *International Journal of Food Microbiology*, 99(1), 71-77.
- 433 Virginia Muriel-Galet, J. P. C., Gracia López-Carballo, Marta Lara,, & Hernández-Muñoz, R. G. a.  
434 P. (2012). Development of antimicrobial films for microbiological control of packaged  
435 salad. *International Journal of Food Microbiology*.
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438

439 **LEGENDS TO FIGURES**

440

441 **Figure 1.** “*Four season salad*” bags in PP/EVOH films with oregano essential oil (A) and citral  
442 (B) in active modified atmosphere.

443

444 **Figure 2.** Sensory evaluation of the differences between the smell, appearance, texture and  
445 general preference of CONTROL and OEO on the 1<sup>st</sup> (A) and 8<sup>th</sup> (B) day of storage. The dashed  
446 line indicates the minimum number of responses for which the difference is significant.

447

448 **Figure 3.** Sensory evaluation of the differences between the smell, appearance, texture and  
449 general preference of CONTROL and CITRAL on the 1<sup>st</sup> (A) and 8<sup>th</sup> (B) day of storage. The  
450 dashed line indicates the minimum number of responses for which the difference is significant.

451

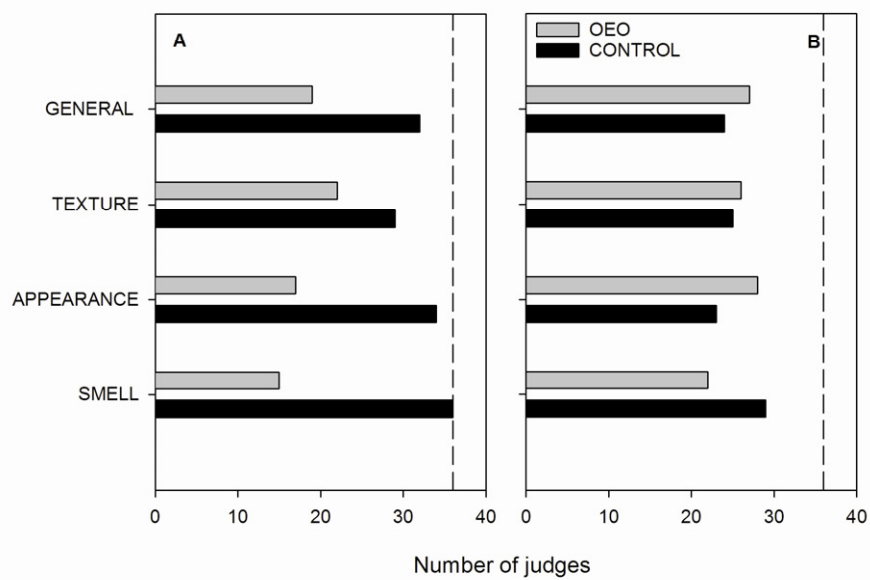
452 **Figure 4.** Sensory evaluation of the differences between the smell, appearance, texture and  
453 general preference of OEO and CITRAL on the 1<sup>st</sup> (A) and 8<sup>th</sup> (B) day of storage. The dashed  
454 line indicates the minimum number of responses for which the difference is significant

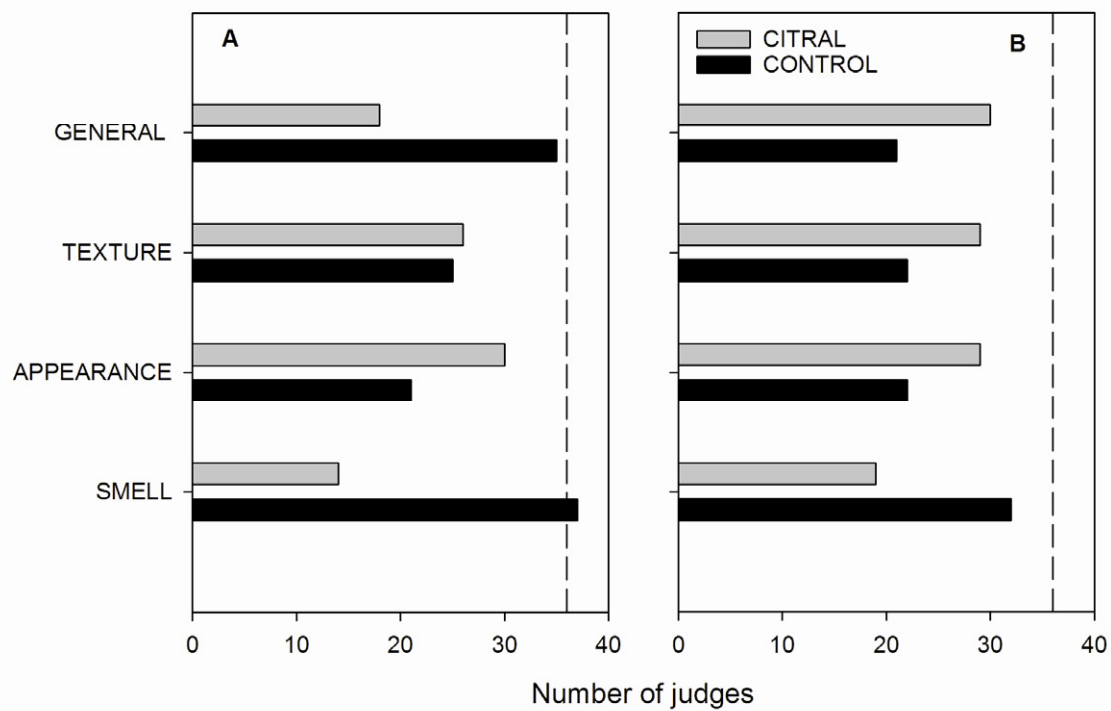
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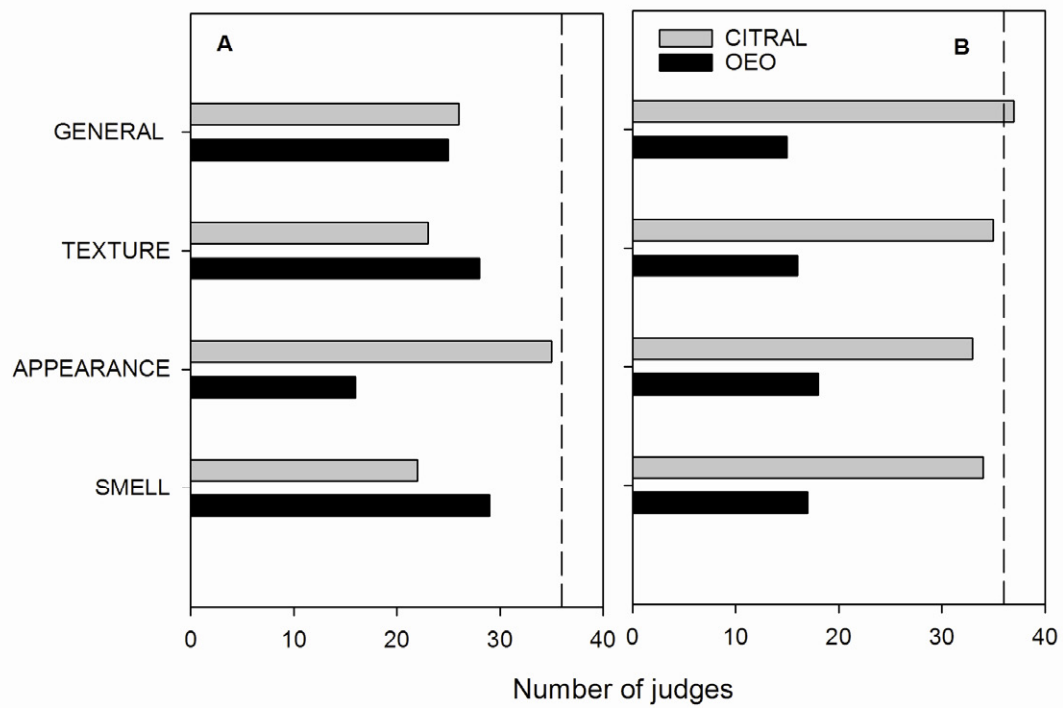
**HIGHLIGHTS**

- Development of antimicrobial film of PP/EVOH with oregano essential oil or citral.
- Active coating did not affect the functional properties of the packaging film.
- Antimicrobial films improved microbial stability at the beginning of storage.
- Consumers preferred salad packaged in active bags with citral at the end of storage.





ACCEPTED





ACCEPTED MANUSCRIPT

**Table 1.** Values of O<sub>2</sub> and CO<sub>2</sub> permeance of PP films and PP/EVOH.

	Thickness ( $\mu\text{m}$ )	$10^{14} \cdot \text{O}_2$ permeance ( $\text{m}^3/\text{m}^2 \cdot \text{s} \cdot \text{Pa}$ )		$10^{14} \cdot \text{CO}_2$ permeance ( $\text{m}^3/\text{m}^2 \cdot \text{s} \cdot \text{Pa}$ )	
		0% RH	90% RH	0% RH	90% RH
		<b>PP</b>	30	$13.54 \pm 0.15$	$15.06 \pm 0.20$
<b>PP+EVOH</b>	30/1.15	$0.085 \pm 0.003$	$10.17 \pm 0.08$	$0.78 \pm 0.02$	$24.67 \pm 0.22$

**Table 2.** Atmospheric composition (% O<sub>2</sub> and CO<sub>2</sub>) of headspace in mixed salad packages manufactured with the control film and the active films at different storage times.

	Day 1		Day 4		Day 8	
	O <sub>2</sub>	CO <sub>2</sub>	O <sub>2</sub>	CO <sub>2</sub>	O <sub>2</sub>	CO <sub>2</sub>
<b>CONTROL</b>	7.3 ± 1.3a	6.1 ± 1.4a	0.16 ± 0.08a	10.8 ± 1.4b	0.03 ± 0.01a	15.1 ± 1.1b
<b>OEO</b>	5.5 ± 1.5a	6.8 ± 1.3a	0.06 ± 0.03a	11.4 ± 1.7b	0.02 ± 0.01a	15.5 ± 1.4b
<b>CITRAL</b>	6.5 ± 1.5a	6.3 ± 1.0a	0.25 ± 0.12a	9.0 ± 1.0a	0.02 ± 0.01a	11.8 ± 1.2a

a, b: the letters are indicative of significance



**Table 4.** Enumeration of microbial population in mixed salad packages: enterobacteria, total aerobic, yeasts and moulds, lactic acid bacteria and psychrotrophic bacteria on 1<sup>st</sup>, 4<sup>th</sup> and 8<sup>th</sup> days of storage.

ENTEROBACTERIA								
Coating	Day 1			Day 4			Day 8	
	Log	Reduction		Log	Reduction		Log	Reduction
CONTROL	4.98 ± 0.65c			4.00 ± 0.00			4.87 ± 0.02	
OEO	3.60 ± 0.38b	-1.38		4.01 ± 0.14	-		4.73 ± 0.38	-
CITRAL	2.85 ± 0.09a	-2.13		3.96 ± 0.50	-		4.90 ± 0.03	-
TOTAL AEROBIC								
CONTROL	5.18 ± 0.36 c			5.13 ± 0.02			6.17 ± 0.03	
OEO	4.10 ± 0.15 b	-1.08		5.03 ± 0.23	-0.10		5.89 ± 0.18	-
CITRAL	3.95 ± 0.11 a	-1.23		5.23 ± 0.07	-		6.05 ± 0.07	-
YEASTS AND MOULDS								
CONTROL	5.54 ± 0.12 c			4.28 ± 0.03 b			6.05 ± 0.16 b	
OEO	3.76 ± 0.25 b	-1.77		3.88 ± 0.11 a	-0.40		5.36 ± 0.32 a	-0.69
CITRAL	3.52 ± 0.53 a	-2.02		3.68 ± 0.00 a	-0.60		5.41 ± 0.04 a	-0.64
LACTIC ACID BACTERIA								
CONTROL	4.20 ± 0.14 c			4.05 ± 0.05 c			6.24±0.08 c	
OEO	3.53 ± 0.51 b	-0.67		3.71 ± 0.28 b	-0.34		5.80±0.20 b	-0.44
CITRAL	2.00 ± 0.00 a	-2.20		3.13 ± 0.16 a	-0.91		5.05±0.15 a	-1.19
PSYCHROTROPHIC								
CONTROL	5.74 ± 0.55 c			5.11 ± 0.00 b			6.11±0.06 b	
OEO	4.00 ± 0.01 b	-1.75		4.95 ± 0.28 a	-0.16		5.88±0.12 a	-0.23
CITRAL	3.74 ± 0.24 a	-2.01		4.93 ± 0.11 a	-0.18		5.82±0.60 a	-0.28

a,b,c: the letters are indicative of significance

Table 3. Mechanical properties of films fested at 50% RH.

Sample	$\sigma_m$ (MPa)	$\epsilon_b$ (%)	$E$ (MPa)
PP	125.65 ± 5.76 a	356.65 ± 29.74 b	658.48 ± 41.40 a
CONTROL	205.67 ± 42.56 b	303.76 ± 38.14 a	1729.02 ± 90.55 b
OEO	211.89 ± 10.32 b	274.44 ± 27.88 a	1746.98 ± 35.65 b
CITRAL	213.64 ± 9.29 b	281.51 ± 26.29 a	1754.90 ± 82.94 b

$\sigma_m$ : Maximum tensile strength.  $\epsilon_b$ : Elongation at break.  $E$ : Young's modulus. Reported values are means of 6 replicates ± standard deviation.

a, b: the letters are indicative of significance