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Additional Information

1 **Microbial system for identification of antibiotic residues in milk**

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24 **ABSTRACT**

25
26 The aim of this study was to evaluate the ResScreen[®] microbiological system for
27 the identification of antibiotic residues in milk. This microbiological system consists of two
28 methods, the "BT" (betalactams and tetracyclines) and "BS" (betalactams and sulfamides)
29 bioassays, containing spores of *G. stearotherophilus* subsp. *calidolactis*, culture media
30 and indicators (acid-base and redox). The detection limits of 29 antimicrobial agents were
31 calculated using a logistic regression model.

32 Both methods detect residues of penicillin-G, ampicillin, amoxicillin, cloxacillin,
33 oxacillin, cephalexin, cefoperazone and ceftiofur[®] at levels close to their Maximum
34 Residue Limits (MRL). The "BT" bioassay also presents good sensitivity to tetracycline
35 and oxytetracycline residues, whereas the "BS" bioassay detects sulfadiazine,
36 sulfamethoxazole and sulfathiazole residues in milk.

37 The simultaneous use of both bioassays identifies betalactam, tetracycline and
38 sulfamide residues in milk. Neomycin, tylosin and lincomycin residues can also be
39 detected, but these molecules are positive with the "BT" and "BS" bioassays, e.g.,
40 betalactams, given the microorganisms' sensitivity to these molecules.

41 Key words: screening test, microbiological inhibition system, betalactams, tetracyclines,
42 sulfamides, milk.

INTRODUCTION

47

48

49 The presence of certain antibiotic residues in milk is a potential risk for consumers
50 because they may be toxic and dangerous for human health, and may potentially cause
51 antimicrobial resistance⁽¹⁻²⁾ and technological problems during dairy product manufacturing
52 ⁽³⁻⁵⁾.

53 For this purpose, several commercially available tests have been developed for the
54 swift, precise detection of the presence of antibiotic residues in milk ⁽⁶⁻⁷⁾. Many screening
55 tests are based on the inhibition of microorganism growth by the presence of drug residues.
56 Among the most widely used microorganisms, we find *Geobacillus stearothermophilus*
57 subsp. *calidolactis* in the following tests: Delvotest^{®(8)}, BRT[®] AiM⁽⁹⁾, Eclipse^{®(10)} and
58 Charm[®] AIM-96⁽¹¹⁾.

59 These methods can nonspecifically detect the “presence” or “absence” of antibiotic
60 residues in milk. To identify β -lactam or sulfonamide compounds however, “positive” and
61 “doubtful” samples are tested using penicillinase and p-aminobenzoic acid (PABA)
62 solutions. Thus, antibiotic residues can be classified into betalactam antibiotics or
63 sulfamides⁽¹²⁾.

64 However, the penicillinase and PABA methods do not suffice to identify other
65 antimicrobial agents such as tetracyclines. So, when Yamaki *et al.*⁽¹³⁾ investigated 2686
66 samples of ewe’s milk, 47 samples were found to be positive with the Delvotest "SP" test.
67 When using penicillinase and PABA methods, only 29.8% of the samples were identified
68 as containing betalactam residues, while the remaining milk samples (70.2%) remained

69 unidentified. These authors suggested that this methodology is insufficient for a complete
70 identification of milk antibiotic residues.

71 In order to identify a higher number of antibiotic groups, Althaus and Nagel⁽¹⁴⁾
72 proposed to use a microbiological system which not only complies with the International
73 Standardization Organization guidelines⁽¹⁵⁾, but also identifies betalactam, tetracycline and
74 sulfonamide residues.

75 This microbiological system consists of two methods, the "BT" (betalactams and
76 tetracyclines) and "BS" (betalactams and sulfamides) bioassays, containing spores of *G.*
77 *stearothermophilus* subsp. *calidolactis*, culture media and indicators (acid-base and redox).
78 Moreover, this system includes synergistic components that improve the sensitivity of
79 tetracycline⁽¹⁶⁾ and sulfamide⁽¹⁷⁾ residues in milk.

80 Thus, the objective of this research was to evaluate the ResScreen[®] system for the
81 identification of antimicrobial agent residues in milk by means of studying detection limits.

82

83 MATERIALS AND METHODS

84

85 Animals and milk samples

86 The animals came from cattle herds of Las Colonias (Santa Fe, Argentina). For this
87 study, milk samples corresponding to the morning machine milking session (6 am) of 16
88 cows were collected in the 60-90 day postpartum period. The animals received no
89 pharmacological treatment throughout the sampling period⁽¹⁸⁾.

90 The chemical composition and pH values of the selected samples were normal for
91 bovine milk, with low somatic cell counts (SCC < 400000 cells ml⁻¹) and an acceptable
92 bacterial count for cow's milk (CFU < 100000 cfu ml⁻¹).

93

94 **Antimicrobial solutions and spiked samples**

95 The drugs used for the preparation of antimicrobial solutions were stored and
96 handled according to the manufacturers' instructions before use. All the dilutions were
97 prepared in 100 ml volumetric flasks at the time the analyses were carried out in order to
98 avoid the possibility of unstable solutions.

99 Antimicrobial solutions were prepared using antimicrobial-free milk⁽¹⁸⁾, as
100 determined by the Delvotest[®]. The final drug concentrations in milk ($\mu\text{g l}^{-1}$) were achieved
101 after serial dilutions so that the volume of the antimicrobial agent solution did not exceed 1
102 % of the volume of the final solution to be analyzed⁽¹⁸⁾.

103

104 **ResScreen[®] test**

105 The system consists of two microbial bioassays using *Geobacillus*
106 *stearothermophilus* subsp. *calidolactis* C-953 spores. The microbiological method is based
107 on growth inhibition of bacteria-test when milk containing residues of antibiotics.

108 The BT bioassay (Betalactams and Tetracyclines) is composed of a culture medium
109 containing spores of thermophilic microorganism, chloramphenicol and bromocresol purple
110 indicator⁽¹⁶⁾. If the milk sample is free of antibiotics and allows bacteria-test growth and
111 changes in color of the acid base indicator (purple to yellow). Otherwise the test will
112 remain the same color.

113 Moreover, the BS bioassay (Beta-lactams and Sulfonamides) use a medium
114 inoculated with a microorganism spore suspension, brilliant black indicator, toluidine blue

115 and trimethoprim⁽¹⁷⁾. So, the absence of antibiotic residues in milk causes bacteria-test
116 growth, producing a color change of indicators from black to amber.

117 The ResScreen[®] system was carried out according to the manufacturer's
118 instructions. Thus, 50 µl milk sample was added to individual plates of the "BT" and "BS"
119 ResScreen[®] methods. Plates were incubated in a water bath at 64±1 °C for 3 ("BT"
120 ResScreen[®]) and 4 hours ("BS" ResScreen[®]) until the color change of the negative samples
121 had taken place.

122 Visual interpretation was performed independently by 3 trained persons, and was
123 assessed visually as "negative" and "positive"; "doubtful" qualifications were interpreted as
124 positive⁽¹⁹⁾.

125

126 **Detection limits and cross specificity**

127 **Detection limits:** The following substances (Sigma Chemical Co, St. Louis, MO) were used
128 to determine the ResScreen[®] system detection limits:

129 - Ten betalactams: amoxicillin, ampicillin, cloxacillin, oxacillin, penicillin "G", cefadroxil,
130 cephalixin, cefoperazone, cefuroxime and ceftiofur[®].

131 - Four sulfonamides: sulfadiazine, sulfadimethoxine, sulfamethoxazole and sulfathiazole.

132 - Three tetracyclines: chlortetracycline, oxytetracycline and tetracycline.

133 The detection limits of the antimicrobial agents were established according to the
134 Codex Alimentarius guidelines⁽¹⁸⁾. For this purpose, 12 concentrations were prepared with
135 different levels of each drug. For each concentration, 16 replicates were prepared using
136 antibiotic-free milk samples.

137 **Cross specificity:** The Codex Alimentarius guidelines⁽¹⁸⁾ were used to calculate the
138 detection limits of the following antibiotics (Sigma Chemical Co, St. Louis, MO):

139 - Four aminoglycosides: gentamycin, kanamycin, neomycin and streptomycin.

140 - Four macrolides: erythromycin, lincomycin, tylosin and spiramycin.

141 - Four quinolones: ciprofloxacin, norfloxacin, enrofloxacin and marbofloxacin.

142

143 **Statistical analysis**

144 The results were obtained by following the SAS[®] Logistic procedure⁽²⁰⁾. A logistic
145 regression model was also done to calculate the detection limits, as follows:

$$146 \quad L_{ij} = \text{logit} [P_{ij}] = \beta_0 + \beta_1 [A]_i + \varepsilon_{ij}$$

147 where: L_{ij} = lineal logistic model; $[P_{ij}] = \text{logit} [P_p/(1-P_p)]$: the probability of “positive”
148 response / probability of “negative” response); β_0, β_1 = coefficients estimated for the logistic
149 regression models; $[A]_i$ = antimicrobial concentration. ε_{ij} = residual error. The concordance
150 coefficient⁽²⁰⁾ was applied as the rank correlation between the observed responses and the
151 predicted probabilities.

152 The detection limit of the visual interpretation of the ResScreen[®] system was
153 estimated as the concentrations at which 95 % of the results were assessed as “positive” or
154 “doubtful”^(19,21).

155

156 **RESULTS AND DISCUSSION**

157 **Detection limits**

158 The results of applying the logistic regression model to the positive relative
159 frequency of the “BT” and “BS” ResScreen[®] system for the different antimicrobial agents
160 assayed are shown in Table 1.

161 The concordance coefficients obtained by applying the logistic model were high,
162 between 89.2 % for oxytetracycline (“BT” ResScreen[®]) and 99.4 % for tetracycline (“BS”
163 ResScreen[®]), demonstrating the correct adjustment achieved by the logistic model.

164 The " β_1 " coefficient represents the sensitivity of *G. stearothermophilus* to the
165 antibiotics studied. This parameter reached higher values for penicillin antibiotics
166 (amoxicillin, ampicillin, cloxacillin, oxacillin and penicillin "G") than for the rest of the
167 antimicrobial agents assayed, demonstrating the sensitivity of *G. stearothermophilus* to
168 detect the residues of these antimicrobials.

169 The " β_1 " coefficients values of cephalosporins (cefadroxil, cephalexin,
170 cefoperazone, ceftiofur[®] and cefuroxime) were similar to those calculated for tetracyclines
171 (“BT” ResScreen[®]) and sulfamides (“BS” ResScreen[®]). In contrast, the " β_1 " parameter of
172 tetracyclines (“BS” ResScreen[®]) and sulfonamides (“BT” ResScreen[®]) were very low,
173 showing low sensitivity for detection purposes.

174 Figures 1 and 2 show the effect of penicillin and cephalosporin concentrations on
175 the visual interpretations of the ResScreen[®] system, as well as the curves constructed by the
176 logistic model (“ β_0 “ and “ β_1 “ coefficients, Table 1). The concentrations of ampicillin,
177 amoxicillin, oxacillin and penicillin "G" (high “ β_1 “ coefficient values) underwent a slight
178 increase to produce 100 % positive results, whereas the concentrations of cephalosporins
179 (Figure 2) had to undergo greater increments to obtain positive results in both methods
180 (lower “ β_1 “ coefficient values).

181 The dose-response curves for tetracyclines (“BT” ResScreen[®], Figure 3) and
182 sulfonamides (“BS” ResScreen[®], Figure 4) showed adequate sensitivity to detect the
183 residues belonging to both antibiotic groups. Conversely, high concentrations of
184 tetracyclines (“BS” ResScreen[®], Figure 3) and sulfonamides (“BT” ResScreen[®], Figure 4)
185 were needed given the low “ β_1 ” coefficients values (Table 1).

186 The detection limits of the ResScreen[®] system calculated by means of logistic
187 regression models for betalactams, tetracyclines and sulfonamides are shown in Table 2.

188 Amoxycillin, ampicillin, cloxacillin, oxacillin, penicillin “G”, cephalexin,
189 cefoperazone and ceftiofur[®] showed similar detection limits (Table 2) for the ResScreen[®]
190 system to their respective Maximum Residue Limits (MRLs).

191 For betalactam antibiotics, other microbiological methods such as BRT[®] AiM⁽²²⁻²³⁾,
192 Charm^{®(24)}, Delvotest[®] “SP”^(22, 25), Eclipse[®] 100ov⁽²⁶⁾ have similar detection limits to the
193 ResScreen[®] system.

194 With regard to tetracyclines, Table 2 indicates how the “BT” ResScreen[®] method
195 presented detection limits near at the MRLs, unlike the “BS” method which required higher
196 concentrations of these antibiotics for them to be detected.

197 The detection limits calculated for the three tetracyclines with the “BT” ResScreen[®]
198 method were lower than those reported by other authors with the BRT[®] AiM⁽²²⁻²³⁾, Charm[®]
199 AIM-96⁽²⁴⁾, Delvotest[®] “SP”^(22,25) and Eclipse[®] 100ov⁽²⁶⁾ methods due to improved
200 sensitivity from adding chloramphenicol to the culture medium⁽¹⁶⁾.

201 Also, Table 2 indicates how the detection limits of sulfonamides for “BS”
202 ResScreen[®] approached their MRLs, while the “BT” method was not sensitive enough to
203 detect these drugs in milk (detection limits higher than 12000 $\mu\text{g l}^{-1}$).

204 The “BS” ResScreen[®] method detection limits of sulfonamides were slightly higher
205 than those observed for BRT[®] AiM⁽²⁷⁾, although other authors have reported higher
206 detection limits for BRT[®] AiM⁽²²⁻²³⁾, Delvotest[®] “SP”^(22,25) and Eclipse[®] 100⁽²⁶⁾.

207

208 **Cross specificity**

209 The detection limits calculated by the logistic model for other antimicrobial agents
210 (aminoglycosides, macrolides and quinolones) with the ResScreen[®] system are provided in
211 Table 3. Of all these antibiotics, only neomycin, lincomycin and tylosin residues were
212 detected by the ResScreen[®] system at levels approaching their MRLs.

213 Various authors have indicated similar detection limits to those calculated in Table
214 3 by other methods using *G. stearothermophilus* subsp. *calidolactis*, such as BRT[®] AIM⁽²²⁻
215 ²³⁾, Charm[®] AIM-96⁽²⁴⁾ and Delvotest[®] “SP”^(22,25) indicating good sensitivity to these three
216 antibiotics (neomycin, lincomycin, tylosin) and a low detection capacity for the rest of
217 antimicrobials.

218

219 **Identification of antibiotic residues by the ResScreen[®] system**

220 Table 4 summarizes the results of Table 2 and Table 3 by collectively and simply
221 presenting the interpretation of the results of both bioassays.

222 Milk samples that led to changes in color of both bioassays indicate the absence of
223 antimicrobials (or substances that were not detected by this system). Beta-lactam antibiotics
224 were identified by the persistence of both methods’ original colors. The fact that the
225 original color of the “BT” bioassay remained and the original color of the “BS” bioassay
226 changed denotes the presence of tetracycline residues. Conversely, milk samples that have

227 sulfamides brought about a change in the color of the “BT” method but maintained the
228 color of the “BS” method.

229 Finally, those milk samples containing neomycin, lincomycin or tylosin residues
230 were detected by the ResScreen[®] system, but were identified as beta-lactams because the
231 "BT" and "BS" bioassays were sensitive enough to detect such substances (Table 3). The
232 difficulty owing to the cross specificity of the ResScreen[®] system could be resolved by
233 implementing subsequent tests with penicillinase and cephalosporinase enzymes.

234

235

CONCLUSIONS

236

237 To summarize, the ResScreen[®] system uses only two bioassays and provides a
238 simple, economical solution to identify residues in milk. Moreover, this microbiological
239 system identifies a larger number of antibiotic families (beta-lactams, tetracyclines and
240 sulfamides) compared with current penicillinase and p-aminobenzoic acid methodologies
241 (beta-lactams and sulfamides).

242 In the future, new bioassays can be incorporated into the ResScreen[®] system in
243 order to increase its identification capacity to other antibiotic groups (macrolides,
244 aminoglycosides or quinolones).

245

246

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247

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251

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- 331

332 **Table 1.** Summary of the logistic regression model parameters of antibiotics in milk for the
 333 ResScreen[®] system
 334

Antibiotics	ResScreen [®] “BT”		ResScreen [®] “BS”	
	Logit = $\beta_0 + \beta_1*[A]$	C	Logit = $\beta_0 + \beta_1*[A]$	C
<i>Betalactams</i>				
Amoxicillin	Logit = -11.3966 + 1.5185*[A]	96.7	Logit = -15.8159 + 3.7160*[A]	97.7
Ampicillin	Logit = -14.7862 + 2.3659*[A]	98.5	Logit = -21.6358 + 6.8009*[A]	99.1
Cloxacillin	Logit = -13.1755 + 0.3835*[A]	97.9	Logit = -10.9673 + 0.3371*[A]	97.5
Oxacillin	Logit = -18.4151 + 1.2483*[A]	98.6	Logit = -22.3155 + 1.5409*[A]	98.9
Penicillin “G”	Logit = -16.1514 + 6.1636*[A]	98.9	Logit = -22.5024 + 8.1827*[A]	99.4
Cefadroxil	Logit = -7.9435 + 0.0683*[A]	95.7	Logit = -16.0260 + 0.0970*[A]	97.5
Cephalexin	Logit = -10.0512 + 0.1313*[A]	97.4	Logit = -9.9664 + 0.0767*[A]	97.0
Cefoperazone	Logit = -11.0985 + 0.2277*[A]	98.3	Logit = -12.6755 + 0.1669*[A]	97.9
Ceftiofur [®]	Logit = -12.1451 + 0.1438*[A]	98.8	Logit = -6.7069 + 0.0841*[A]	94.3
Cefuroxime	Logit = -13.07 + 0.3282*[A]	98.9	Logit = -20.0044 + 0.1321*[A]	99.6
<i>Tetracyclines</i>				
Clortetracycline	Logit = -9.4066 + 0.0556*[A]	90.7	Logit = -10.1408 + 0.0036*[A]	97.7
Oxytetracycline	Logit = -10.8242 + 0.0933*[A]	89.2	Logit = -9.9616 + 0.0153*[A]	97.0
Tetracycline	Logit = -9.0156 + 0.0627*[A]	89.8	Logit = -26.5938 + 0.0309*[A]	99.4
<i>Sulfonamides</i>				
Sulfadiazine	Logit = -8.2241 + 0.0002*[A]	95.0	Logit = -22.089 + 0.1525*[A]	91.0
Sulfadimethoxine	Logit = -18.8281 + 0.0018*[A]	98.7	Logit = -11.9029 + 0.0577*[A]	90.0
Sulfamethoxazole	Logit = -16.7196 + 0.0015*[A]	97.9	Logit = -11.0868 + 0.1167*[A]	89.3
Sulfathiazole	Logit = -20.2747 + 0.0017*[A]	98.6	Logit = -9.0399 + 0.1246*[A]	89.3

335 β_0, β_1 = coefficients estimated for the logistic regression models; [A]: antimicrobial concentrations; C:
 336 concordance coefficients.

337 **Table 2.** The ResScreen[®] system detection limits ($\mu\text{g l}^{-1}$) for antibiotics in milk
 338

Antibiotics	ResScreen [®]		BRT [®] AIM		Delvotest [®] SP		Charm [®] AIM	Eclipse [®] 100ov	MRLs ^a
	BT	BS	Charm y Ruth (1993) ⁽²²⁾	Heeschen <i>et al.</i> (1995) ⁽²³⁾	Charm y Ruth (1993) ⁽²²⁾	Althaus <i>et al.</i> (2002) ^{(25)*}	Linage <i>et al.</i> (2007) ^{(24)*}	Montero <i>et al.</i> (2005) ^{(26)*}	
<i>Beta-lactams</i>									
Amoxicillin	8	5	5	---	10	5	---	7	4
Ampicillin	7	4	10	5	10	3	6	---	4
Cloxacillin	42	40	100	35	50	23	42	68	30
Oxacillin	17	16	---	---	---	---	---	28	30
Penicillin "G"	3	3	10	1.5	2.5	1.4	4	5	4
Cefadroxil	159	190	---	---	---	63	---	86	---
Cephalexin	99	160	---	---	---	68	202	115	100
Cefoperazone	62	94	---	---	---	41	82	110	50
Ceftiofur [®]	105	115	100	---	50	59	107	---	100
Cefuroxime	42	170	---	---	---	41	---	85	---
<i>Tetracyclines</i>									
Clortetracycline	275	3600	>1000	---	420	---	3989	1500	100
Oxytetracycline	150	850	1000	---	200	420	501	560	100
Tetracycline	158	720	1000	450	420	450	257	480	100
<i>Sulfonamides</i>									
Sulfadiazine	49000	164	1000	100-1000	>1000	260	---	---	100
Sulfadimethoxine	12000	260	100	100-1000	>1000	---	119	170	100
Sulfamethoxazole	14000	120	---	---	---	110	---	---	100
Sulfathiazole	13000	100	1000	100-1000	>1000	---	151	250	100

339 ^aMRLs ($\mu\text{g l}^{-1}$), EU maximum residue limits, * . Decision limits in ewe milk.
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341 **Table 3.** The ResScreen[®] system detection limits ($\mu\text{g l}^{-1}$) for other antimicrobials in milk
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Other antimicrobials	ResScreen [®]		BRT [®] AIM		Delvotest [®] SP		Charm [®] AIM	Eclipse [®] 100ov	MRLs ^a
	BT	BS	Charm y Ruth (1993) ⁽²²⁾	Heeschen <i>et al.</i> (1995) ⁽²³⁾	Charm y Ruth (1993) ⁽²²⁾	Althaus <i>et al.</i> (2002) ^{(25)*}	Linage <i>et al.</i> (2007) ^{(24)*}	Montero <i>et al.</i> (2005) ^{(26)*}	
<i>Aminoglycosides</i>									
Gentamycin	320	530	>500	---	150	1200	382	3140	100
Kanamycin	5600	6200	---	---	---	---	---	18700	150
Neomycin	600	1200	>500	300	150	3300	1084	9100	1500
Streptomycin	2300	3600	>1000	---	>1000	10000	3593	10100	200
<i>Macrolides</i>									
Erythromycin	210	190	1000	2250	400	980	522	750	40
Lincomycin	150	220	---	---	---	---	---	---	150
Tylosin	74	50	50	---	100	120	51	230	50
Spiramycin	3400	2600	---	---	---	---	1346	18100	200
<i>Quinolones</i>									
Ciprofloxacin	1750	1710	---	---	---	---	---	5100	100
Enrofloxacin	2000	2300	---	---	---	---	46000	4000	100
Marbofloxacin	2700	4400	---	---	---	---	---	---	75
Norfloxacin	7100	6800	---	---	---	---	---	9500	---

343 ^aMRLs ($\mu\text{g l}^{-1}$), EU maximum residue limits, * : Decision limits in ewe milk.

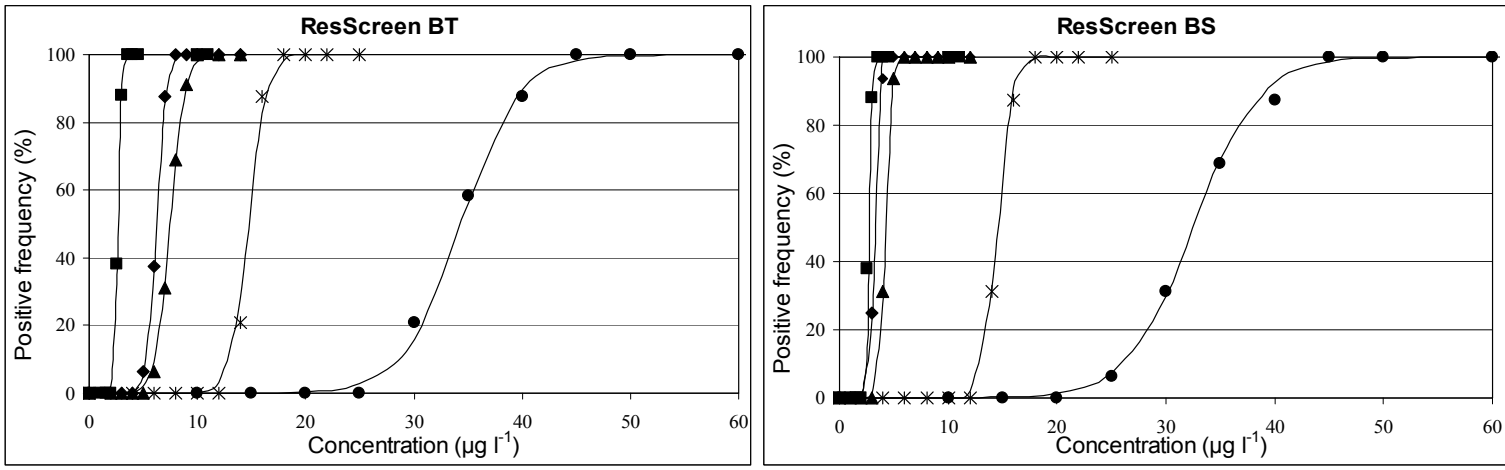
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Table 4. Interpretation of the ResScreen[®] system results

Antibiotics	Results	
	ResScreen [®] “BT”	ResScreen [®] “BS”
Absence (or not detect)	-	-
Betalactams	+(*)	+(*)
Tetracyclines	+	-
Sulfamides	-	+

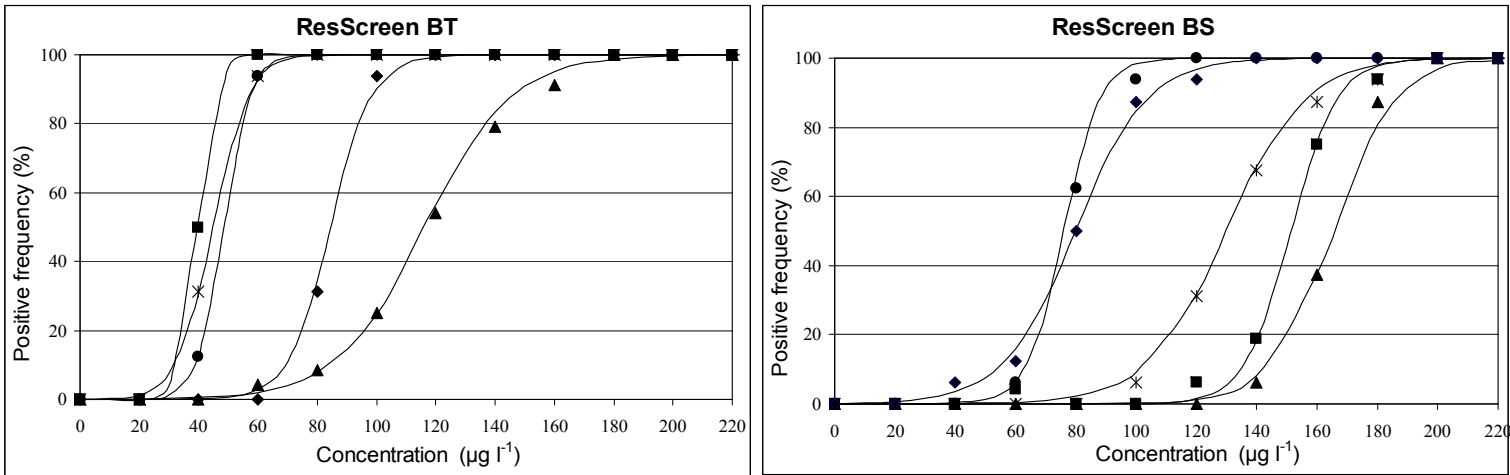
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(*) Interference due to neomycin, lincomycin and tylosin.



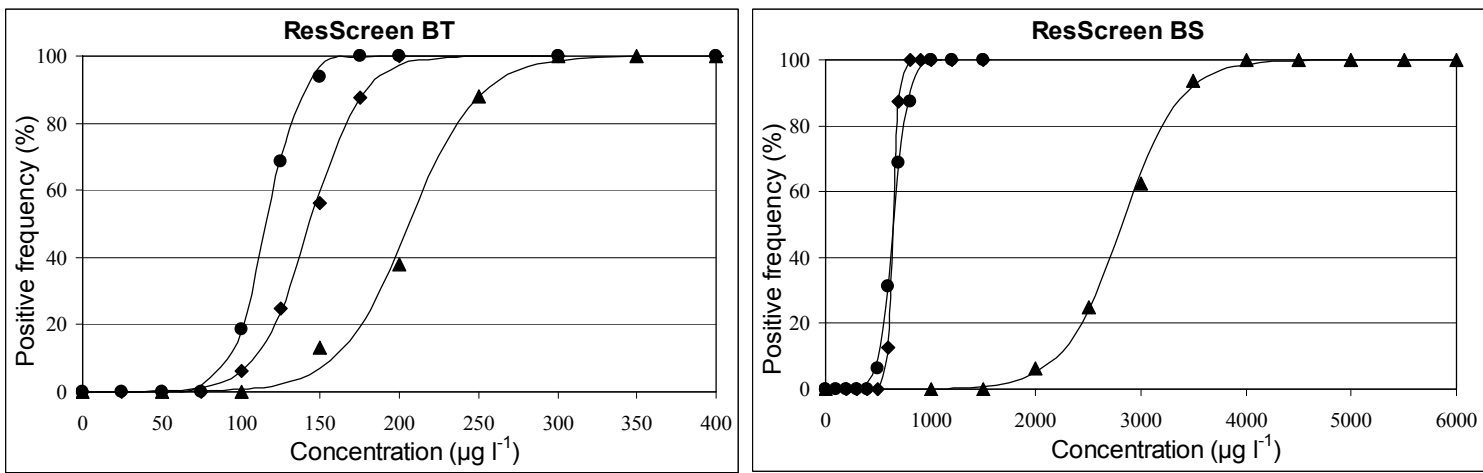
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Fig. 1 Dose-response curves for different penicillin concentrations in milk analyzed by the Rescreen[®] system (▲: amoxycillin, ◆: ampicillin, ●: cloxacillin, X: oxacilina, ■: penicillin “G”).



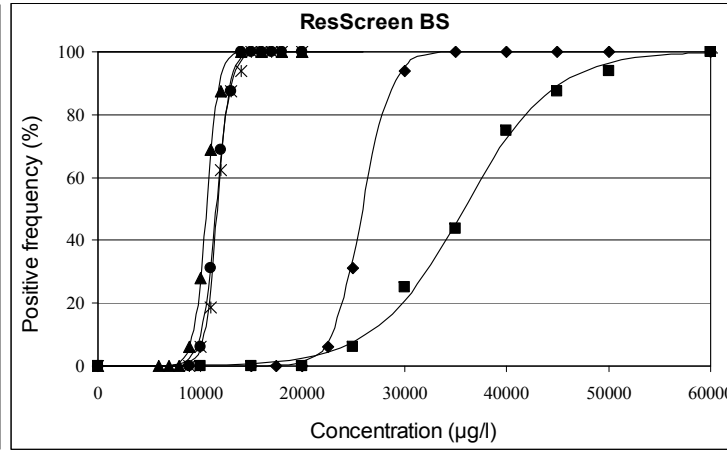
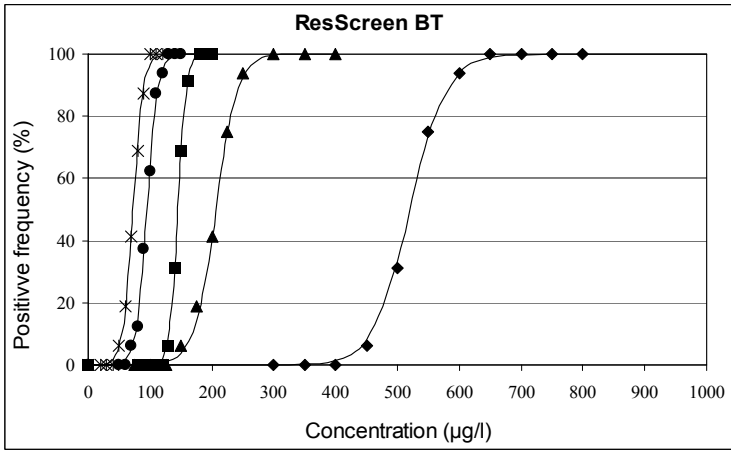
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Fig. 2 Dose-response curves for different cephalosporin concentrations in milk analyzed by the Rescreen[®] system (▲: cefadroxil, X: cephalexin, ●: cefoperazone, ◆: ceftiofur[®], ■: cefuroxime).



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Fig. 3 Dose-response curves for different tetracycline concentrations in milk analyzed by the Rescreen[®] system (▲: chlortetracycline, ●: oxytetracycline, ◆: tetracycline).



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Fig. 4 Dose-response curves for different sulphamide concentrations in milk analyzed by the Rescreen[®] system (■: sulfadiazine, ▲: sulfadimethoxine, ◆: sulfamethazine, ●: sulfamethoxazole, X: sulfatiazole).