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Prado, P.; Baeta, M.; Mestre, E.; Solis, MA.; Sanhauja, I.; Gairin, I.; Camps-Castellà, J....
(2024). Trophic role and predatory interactions between blue crab, *Callinectes sapidus*, and native species in open waters of the Ebro Delta. *Estuarine Coastal and Shelf Science*. 298.
<https://doi.org/10.1016/j.ecss.2024.108638>



The final publication is available at

<https://doi.org/10.1016/j.ecss.2024.108638>

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Additional Information

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2 Trophic role and predatory interactions between the

3 blue crab, *Callinectes sapidus*, and native species in

4 open waters of the Ebro Delta

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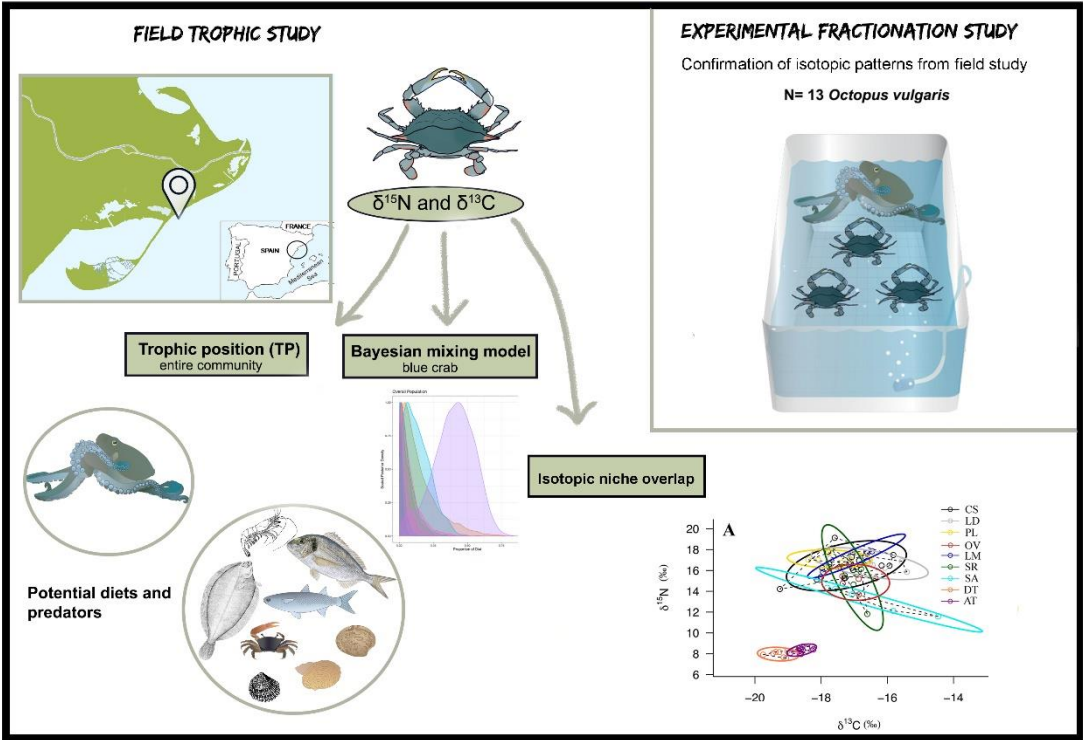
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ABSTRACT

The Ebro Delta has become a major blue crab (*Callinectes sapidus*) fishery area in the NW Mediterranean, but there is limited information on factors controlling the abundance of populations in open waters, a crucial habitat for ovigerous females. Here, we use a stable isotope approach ($\delta^{15}\text{N}$ and $\delta^{13}\text{C}$), to assess blue crab trophic position, the potential consumption of food items using mixing models, and the isotopic niche overlap with local commercial species. For *Octopus vulgaris*, a potential blue crab key predator, a trophic enrichment experiment was also conducted to further assess predation control in wild populations. The blue crab showed 1.6 times higher trophic position than in other habitats of the Ebro Delta, and similar to that found in the harbor crab, *Liocarcinus depurator*, and several predatory fish. Additionally, the isotopic niche of blue crabs showed overlaps from 46.2 to 14.9% with native predators, and mixing models also suggest even dietary contributions throughout the food web. For *O. vulgaris*, field results showed a trophic position of 3.93, lower than that of blue crab, and lower $\delta^{15}\text{N}$ signatures were also obtained in a captivity experiment drawing negative fractionation (-1.1‰). We conclude that high dietary contribution of animal prey might provide a high protein diet that could be crucial for allowing the maintenance of a large local population, but the overall functional trophic similarity could also disfavor local native species. The similarity between experimental fractionation and field differences between predator and prey (-1.6‰) suggests that predation of blue crab is possible, but further research is needed to clarify the metabolic routes involved in octopus $\delta^{15}\text{N}$ fractionation.

Keywords: Stable isotope analyses; isotopic niche overlap; MixSiar; ^{15}N fractionation; *Octopus vulgaris*

1. Introduction

The blue crab *Callinectes sapidus* Rathbun 1896 is a member of the Portunidae native to the Western Atlantic from Uruguay to Nova Scotia (Mancinelli et al. 2017a). The species was recorded in Europe for the first time in 1901 on the Atlantic coast of France (Bouvier 1901), and then appeared in the Mediterranean Sea in 1948 probably introduced by ballast waters (Serbetis 1959; Zenetos et al. 2018). Since then, it has notoriously expanded throughout the entire basin and has become one of the 100 worst invasive species despite its commercial interest (Zenetos et al. 2005). In Spain, it was first detected in the Tancada Lagoon in the Ebro Delta, Catalonia in 2012 (Castejón and Guerao 2013), and since then, it has rapidly spread towards the Atlantic Sea reaching the Southern coast of Portugal (López and Rodon 2018; Vasconcelos et al. 2019).

The species is a voracious generalist consumer capable of using a wide range of animal resources including mollusks, fish, crustaceans, and vegetal (plants and algae) and detrital material in the sediment, depending on availability and the stage of ontogenic development, and local availability (Laughlin 1982; Rosas et al. 1994; Prado et al. 2021). Hence, it can affect benthic communities at multiple trophic levels, altering the functioning and biodiversity of ecosystems, and impacting native fisheries in invaded areas (Zenetos et al. 2005; Nehring 2011; Mancinelli et al. 2017a). For instance, in the Ebro Delta, blue crab has been indicated to prey on locally abundant mollusk species, decreasing the effective size of populations until they become very rare and there is a forced trophic shift to towards other less nutritional resources such as vegetal and detrital material (Prado et al. 2021). Trophic positions (hereafter TP) of blue crab in sites already depleted from animal prey could be lower than 2.6 (Prado et al. 2021), whereas estimates in other Mediterranean regions using the same methodology can reach values as high as 4.5 (Mancinelli et al. 2016; Aslan and Polito 2021).

Poor nutritional conditions may have negative consequences for long-term population traits including survival and growth rates, age at first maturity, and/ or fecundity (review by Metcalfe and Monaghan 2001). Blue crab juveniles when fed on lower levels of dietary protein show lower

growth rates and molting frequency than treatments featuring higher protein contents (Millikin et al. 1980). However, the Ebro Delta is the only area in the Spanish Mediterranean where abundances are large enough to sustain a targeted fishery that may reach over 2 tons of captures per day (López and Rodon 2018; López 2020). Hence, fishing captures also suggest a locally important reproductive output for the species. Ovigerous females, usually gather on the Ebro River mouth and open sea in front of the Ebro Delta (López and Rodon 2018; López 2020), an area that remains unexplored for the trophic structure and resource use of *C. sapidus*. The area hosts a diverse fishery of other commercial species, particularly benthic predatory and omnivorous fish that could overlap the functional trophic niche of blue crab, and behave either as potential predators, food resource, and/ or as an active competitor (see Laughlin 1982; Guillory and Elliot 2001), depending on the size. For instance, the harbor crab, *Liocarcinus depurator*, has been reported to feed mainly on other decapod crustacea followed by small fishes and small cephalopods (Abelló and Cartes 1987). Common predatory flatfish such as *Pegusa lascaris*, *Scophthalmus rhombus*, and *Solea senegalensis* might already undergo certain overlap in their trophic niche which is mostly comprised of different types of crustaceans, bivalves, polychaetes and fish (Cabral et al. 2002). Sparid fishes such as *Lithognathus mormyrus*, *Pagellus erythrinus*, and *Sparus aurata*, are also common opportunistic predators feeding on benthic invertebrates, mainly crustaceans, polychaetes, and bivalve mollusks (Kallianiotis et al. 2005; Taieb et al. 2013) that may be affected by the progressive spread of blue crab. Hence, comparative evaluation of the isotopic niche overlap and TP of these species relative to the blue crab can also provide an adequate proxy to assess functional trophic similarity.

Top-down control of blue crab has been extensively studied in native ecosystems (e.g., Heck and Coen 1995; Guillory and Prejean 2001; Hayes et al. 2022), but is still an unexplored ground in invaded Mediterranean regions. In the comprehensive review conducted by Guillory and Elliot (2001), authors provide a list of 93 species capable of preying on all blue crab life stages, although very few would be capable of preying on adult crabs of commercial size (ca. ≥ 150 g

WW) and/ or are present in a large quantity in the open waters of the Ebro Delta. Among potential candidates, the common octopus *Octopus vulgaris*, usually reaching sizes of 3-4 kg and locally abundant, appear as a good model predator based on general knowledge of decapods being a favorite prey item of octopuses (e.g., Guillory and Elliot 2001; Leite et al. 2009; Pech-Puch et al. 2016) and on local fishermen reports of attacks to blue crab fishing traps (personal communication to P. Prado).

In this context, the main aim of this study was to determine the isotopic niche width, the TP, and the contribution of major prey items to adult female blue crab diet in the open sea of the Ebro Delta, an area that is of key importance for hatching success (females are nearly the exclusive sex in the area) in order to obtain a comprehensive picture of resource uses across the territory (provided only for males in estuarine habitats by Prado et al. 2022) that could help to assess the overall fitness of the population. A second aim was to assess the isotopic niche overlap and compare the TP of the blue crab with other commercial marine species in order to evaluate the potential effect of its introduction in the local food-web structure. A final objective was also to assess the possible role of *O. vulgaris*, in blue crab predation using a controlled manipulative experiment to assess isotopic fractionation. *O. vulgaris* has been indicated as an active predator of both the native green crab, *Carcinus mediterraneus* (Fiorito et al. 1990) and the blue crab in natural (fishermen report to P. Prado) and captivity conditions (P. Prado, personal observation). Hence, information about the specific patterns of fractionation could shed some light on the importance of trophic relationships.

2. Materials and methods

2.1. Study sites, sample collection and processing

The study was conducted in coastal waters ca. 12-16 km south from the Ebro River mouth, an area that is commonly fished for blue crab and other commercial species and features typical Mediterranean marine salinity ranges (ca. 36-37 ppt; Fig. 1). Due to enhanced salinity, the local

sex composition of blue crab in this area is extremely biased towards adult females which constitute the bulk of the captures (López and Rodon 2018).

Fieldwork was conducted in five different dates from mid-October 2020 to late February 2021 to ensure that captures were representative of the abundance of local species. In particular, adult females were collected in a ca. 1.5-month period (two of the sampling occasions), from the beginning of December 2020 to mid-January 2021. Sampling was conducted at ca. 1-10 m depth, using a small-scale fishing boat equipped with three different fishing gears to collect benthic organisms: (1) Mechanized clam dredges (typically 3-4 per boat) were used to collect available infauna such as bivalves, and crustaceans, and some fishes at 1 to 5 m depth; (2) gill nets were used to collect *Callinectes sapidus* and other locally abundant fishes at 1 to 10 m depth; and (3) basket traps were used to capture *Octopus vulgaris* at 5 to 10 m depth.

After collection, blue crabs and other captured species were separately placed within aerated ice coolers and transported to the lab. For each species, a minimum of N= 3 individuals was considered in the study. Individuals of similar sizes were also selected to minimize possible changes in the diet due to age-dependent energy requirements, and except for octopus (ca. 2000 g WW), and mullet (ca. 500 g WW based on feasible predation according to previous observations by P. Prado) featured sizes that were similar or smaller to that of blue crab, in order to allow for possible predation. Individuals of crustaceans: *C. sapidus* (females, n= 25, 250-300 g WW), *Penaeus kerathurus* (n= 3), and *Liocarcinus depurator* (n= 10); bivalves: *Acanthocardia tuberculata* (n= 14), *Donax trunculus* (n= 7), and *Macra stultorum* (n= 3); Octopoda: *Octopus vulgaris* (n= 5); and fishes: *Pegusa lascaris* (n= 5), *Trachinus draco* (n= 3), *Scophthalmus rhombus* (n= 6), *Sparus aurata* (n= 5), *Lithognathus mormyrus* (n= 5), *Pagellus erythrinus* (n= 3), *Mugil cephalus* (n= 4), *Bothus podas* (n= 3) and *Sciaena umbra* (n= 3) were obtained and processed for stable isotope analyses (SIA).

Fish specimens were filleted for the extraction of the entire musculature and rinsed with ultrapure water to prevent contamination with other tissues. For blue crab and harbor crab (*L.*

depurator) a piece of the right claw musculature was extracted, whereas *P. kerathurus* were peeled to obtain the tail muscle, and bivalves were rinsed of sediments and processed as whole individuals without the valves. For octopus, a tentacle fragment was dissected. All tissue samples were dried at 60 °C until constant weight and reduced to powder with a mortar and pestle, which were carefully rinsed with alcohol after each sample to remove any remains.

No plant or macroalgae sources were considered because no consistent material was recovered in the study area during the surveys. Sediments were not included either because within the study area they feature an extremely low organic content (0.011% N and 0.26% C) and were considered as an unlikely food source for blue crab.

2.2. Stable isotope analyses

Samples were analyzed with a Flash 112 IRMS delta C series EA Thermo Finnigan mass spectrometer connected to an elemental analyzer for the determination of C and N contents at the isotopic ratio mass spectrometry facility in the University of Barcelona - Scientific and Technological Center (CCiT-UB). Isotope ratios in samples were calculated from linear calibration curves constructed with standard reference materials of known composition and a blank correction. The difference in isotopic composition between the sample and reference material is determined by the equation:

$$\delta_{\text{sample-standard}} = [(R_{\text{sample}} - R_{\text{standard}}) / R_{\text{standard}}] \times 1000$$

where R_{sample} is the $^{13}\text{C}/^{12}\text{C}$ or $^{15}\text{N}/^{14}\text{N}$ in the sample; R_{standard} is the $^{13}\text{C}/^{12}\text{C}$ or $^{15}\text{N}/^{14}\text{N}$ in the calibration material and $\delta_{\text{sample-standard}}$ is the difference in isotopic composition of the sample relative to that of the reference. The reproducibility of the stable isotope measurements was ~0.1‰. The possible variability in $\delta^{13}\text{C}$ signatures induced by the presence of lipids, which are typically depleted in ^{13}C , were normalized for crustaceans, mollusks, and fish using the equations provided by Post et al. (2007). The threshold for the application of this correction was an increase in corrected signatures of at least 0.1‰ (Prado et al. 2012).

2.3. Isotopic niche of blue crab and native marine species

We used the package SIBER (Stable Isotope Bayesian Ellipses in R) (Jackson et al. 2011) to analyze the structure of the food web. Standard ellipses corrected for small sample size (SEA_c) were used to represent each species in the isotopic space (only those with N ≥ 5 were included). The Bayesian estimate of the standard ellipse and its area (SEA_b) was calculated for each investigated species as a measure of their isotopic niche width or resource use area. SEA_b in units of area (‰²), captures all the same properties as SEA_c and the Total Area (TA) of the convex polygon, but it is unbiased with respect to sample size and exhibits more uncertainty with smaller sample size (Jackson et al. 2011). This Bayesian method based on Markov-Chain Monte Carlo (MCMC) simulations uses posteriori randomly replicated sequences in the 95% confidence interval for the distributions of both stable isotopes to correct the bivariate ellipses (δ¹³C and δ¹⁵N). The magnitude of isotopic overlap (‰²) –as a measure of resource partitioning over time– was obtained from SEA_b estimations and expressed as a % overlap of the blue crab isotopic niche over that of the other species and vice versa.

2.4. Estimates of trophic position

The trophic position (TP) of blue crab and other local predators (octopus, fishes, and crustaceans) was calculated according to the methodology and equation proposed by Vander Zanden et al. (1997):

$$TP_{\delta^{15}N} = (\delta^{15}N_{Consumer} - \delta^{15}N_{Baseline}) / \Delta^{15}N + \lambda$$

Where δ¹⁵N_{consumer} is the nitrogen isotopic signature of the consumer species, δ¹⁵N_{Baseline} is the mean isotopic signature of available filter feeding bivalves (*A. tuberculata*, *D. trunculus*, and *M. stultorum*) which are well-known primary consumers (primary producers= trophic level 1,

primary consumers= trophic level 2, and so on). $\Delta^{15}\text{N}$ is the 3.4‰ $\delta^{15}\text{N}$ isotopic enrichment considered across trophic levels (Post 2002), and λ the trophic level of the baseline indicator, respectively. Bivalves, and particularly the Mediterranean mussel, have been commonly used as the food web baseline in previous stable isotope studies (Mancinelli et al. 2013, 2016, 2017b; Carrozzo et al. 2014; Aslan and Polito 2021; Prado et al. 2021) and therefore provide a robust baseline for comparative purposes with other locations, and with previous studies. In the present study, the mean of the 3 collected bivalve species was used for calculations since there were no significant differences in $\delta^{15}\text{N}$ among them.

2.5. Isotope mixing models

To assess dietary contributions to female blue crab diets ($N=25$; $N \geq 10$ is recommended to prevent large bias and high dispersion; see Brett 2014) in the open sea of the Ebro Delta, the MixSIAR Bayesian mixing model (V.3.1) was implemented as an open-source R package (Stock et al. 2018), using a generalist prior distribution. The model was first outlined by Moore and Semmens (2008) and modified later to improve the robustness of results. Prior to the development of mixing models, an isospace plot was produced to visually inspect that consumer $\delta^{15}\text{N}$ and $\delta^{13}\text{C}$ values fell within the prey polygon in isospace (Stock and Semmens 2016). MixSIAR uses stable isotope signatures with their standard error (SE), and tissue-diet discrimination factors as input variables to estimate the probability distributions of each food item to a mixture within 95% confidence intervals and accounts for uncertainty associated with multiple sources. The estimated median contribution (i.e., the 50% percentile) represents the median source contribution value for each source and is usually given along with standard deviation (SD) and/or confidence intervals for comparative purposes.

Models were run considering two different possible fractionations. First, the 0.4‰ $\Delta^{13}\text{C}$ and the 3.4‰ $\Delta^{15}\text{N}$ values derived from literature data on non-herbivorous aquatic consumers (Post 2002) that have been largely used in other stable isotope works in invaded Mediterranean

regions (Mancinelli et al. 2013, 2016, 2017b; Carrozzo et al. 2014; Aslan and Polito 2021; Prado et al. 2021) and associated SD values (1.3‰ for $\delta^{13}\text{C}$ and 1‰ for $\delta^{15}\text{N}$; Post 2002). Second, the $0.6 \pm 0.7\text{‰}$ $\Delta^{13}\text{C}$ and $1.6 \pm 0.6\text{‰}$ $\Delta^{15}\text{N}$ values obtained for blue crab feeding with clams and black sea bass obtained by McCann and Jensen (2018). Large octopus (1-2 kg) were not included in the runs since this size was experimentally demonstrated to be a highly effective predator of blue crabs.

The Markov Chain Monte Carlo (MCMC) in MixSIAR, a method for obtaining information and estimating posterior distributions in Bayesian inference, was set as follows: chain length 100,000; burn-in 50,000; thin 50; and number of chains 3. With these settings, model selection was based on the adequacy of default diagnostic tests in MixSIAR: for the Gelman–Rubin diagnostic convergence conditions < 1.05 in all cases, and for the Geweke diagnostic, the number of variables falling outside the ± 1.96 range in the first and last part of a Markov chain being $< 5\%$ for the three chains.

2.6. Experimental feeding of *Octopus vulgaris* with blue crab

A total of $N = 13$ individuals of *O. vulgaris* were obtained alive in late March from the outer sea of the Ebro Delta by local fishermen in La Ràpita (DG051201-331/2021) and transported to IRTA facilities. Individuals were first weighted to the nearest 0.01 kg within a mesh bag using a CH-KERN tension dynamometer and then transferred to a large, 3000 L tank, equipped with PVC hides, and seawater at ca. 17-20 °C, $\geq 80\%$ O_2 saturation, and 36 ppt for 2 months (late March to late May 2022). Each animal was first adapted to captivity with dead blue crab (48 h) and then fed daily with 1-2 live individuals bought from fishermen at the lower Ebro River (Deltebre) and remains removed to avoid decomposition.

By the end of May, animals were weighted again to assess biomass incorporation prior SIA. Then, a small fragment of a tentacle from each octopus was dissected and animals returned to the open sea. For blue crabs, samples ($n = 12$) were obtained throughout the 2-month feeding

period (i.e., 1-2 individuals per week). Samples of muscle from octopus and blue crabs were processed as indicated previously in section 2.1.

2.7. Data analyses

Differences in the composition of $\delta^{13}\text{C}$ (normalized) and $\delta^{15}\text{N}$ signatures, and TP of blue crabs and potential resources were investigated with a one-way nested ANOVA (Species fixed factor) and SNK post-hoc analysis. ANOVA assumptions of normality (Chi-square test) and homogeneity of variances (Cochran's test) were not always achieved by transformation and in those instances the significance level was fixed to $p < 0.01$ to minimize the risk of making a Type I error (Underwood 1997). All ANOVAs were performed using Statistica v.12 software.

Comparisons of isotopic signatures between blue crabs from the lower Ebro River (used during the feeding experiment to determine stable isotope fractionation) and those from the sea (trophic guild evaluation) were assessed with a two-tailed t-test for two independent samples with equal variance in order to establish possible salinity effects that could affect interpretation of isotopic patterns of octopuses in captivity and under natural conditions.

3. Results

3.1. Isotopic signatures, TP and diet composition of blue crabs in the open sea of the Ebro Delta

Female blue crabs from the open sea featured mean isotopic signatures of -17.2 ± 0.15 for $\delta^{13}\text{C}$ and 16.4 ± 0.2 for $\delta^{15}\text{N}$. This resulted in a considerably high TP of 4.40 ± 0.06 using $\delta^{15}\text{N}$ of bivalves as baseline for primary consumers. In fact, the TP of blue crabs from the open sea was also 1.4 and 1.6 times higher than that found in other habitats of the Ebro Delta, such as the Alfacs Bay and the Tancada Lagoon, respectively (Table 1).

The MixSiar model showed good performance (no variables higher than 1.05 in the Gelman-Rubin diagnostic and adequate chain convergence in the Geweke test) using 0.4 and 3.4‰ as

fractionation values for $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$, respectively. Results evidenced that blue crab exerts a relatively even consumption of available resources, although with slightly higher contribution of bivalve species (5.9 to 6.4%) and *P. kerathurus* (6.3%) than from the fish community (2.8 to 5.9%). Yet, the total contributions explained by the model (50% percentile) was 80.8%, suggesting the possible contribution of other not considered resources (Table 2). For the second run based on $0.6 \pm 0.7\text{‰}$ $\Delta^{13}\text{C}$ and $1.6 \pm 0.6\text{‰}$ $\Delta^{15}\text{N}$ values, the model did not fit Geweke test and was discarded.

3.2. Comparison of blue crab isotopic niche and TP with those of other local marine species

Patterns of $\delta^{13}\text{C}$ showed significant differences across the food web ($\text{df}= 16$, $\text{MS}= 9.6$, $F= 25.5$, $p< 0.0001$), with higher values in predatory fishes and crustaceans (-18.13 to -16.4), followed by bivalves (-19.3 to -18.6), and lowest values in the fish *M. cephalus* (-21.9 ± 0.4 ; see Fig. 2). There were also significant differences in patterns of $\delta^{15}\text{N}$ among species ($\text{df}= 16$, $\text{MS}= 75.4$, $F= 104.0$, $p< 0.0001$), with highest values in the fishes *L. mormyrus*, *P. lascaris*, and *P. eritrinus* (17.8 to 16.7), and in the crustaceans *L. depurator* (16.6 ± 0.2) and *C. sapidus* (16.4 ± 0.2). Intermediate signatures were observed in the fishes *S. rhombus*, *M. cephalus*, *T. draco*, *B. podas*, *S. umbra*, and *S. senegalensis* (15.3 to 12.5) and in octopus (14.8 ± 0.3). Significant lower groupings were also observed for *P. kerathurus* (10.8 ± 0.3) and bivalves (8.4 to 7.9) (Fig. 2).

The standard ellipse area corrected for small sample size (SEA_c) and the Bayesian standard ellipse area (SEA_b) providing an indication of isotopic niche width showed highest sizes for blue crab, *S. aurata* and *S. rhombus*, intermediate for harbor crab, *P. lascaris*, *L. mormyrus*, and *O. vulgaris*, and lowest for bivalves, which appear as a clearly distinctive trophic group (Fig. 3A-B; Table 3). SEA_b credibility intervals for each species showed higher ranges for *S. aurata*, and *S. rhombus* and lowest for bivalves (Fig 3B). Blue crab showed niche overlaps of 46.2% of its SEA_b with harbor crab, 36.9% with *L. mormyrus*, 33.7% with *O. vulgaris*, 30.6% with *S. rhombus*, 29.2% with *P. lascaris*, and 14.9% with *S. aurata*. No overlap was found with bivalve species (Fig. 3A).

The TP of consumers showed significant differences across the food-web ($df= 13$, $MS= 1.57$, $F= 19.77$, $p< 0.0001$) with values ranging from 4.67 ± 0.11 in *L. mormyrus* to 2.73 ± 0.08 in *P. kerathurus* (Table 1). The harbor crab, *L. depurator* (4.46 ± 0.06) and other well-known benthic fish predators such as *P. erythrinus* (4.48 ± 0.05) and *P. lascaris* (4.59 ± 0.05) showed TPs very similar to that of blue crab. In contrast, the TP of octopus was slightly but significantly lower (3.93 ± 0.09) (Table 1).

3.3. Experimental feeding of *Octopus vulgaris* with blue crab

Octopus easily captured and eagerly consumed blue crab individuals, provided in daily rations (Fig. 4A-C). The average weight increment of octopus during the captivity period was 816.3 ± 121 g WW, from a mean initial size of 2243.3 ± 114.3 to a final weight of 3059.6 ± 129.2 g WW.

Individuals of *C. sapidus* from the lower Ebro River used for feeding octopus had a $\delta^{13}C$ of -21.2 ± 0.1 , which is 3.8‰ lower than those from the open sea ($df= 35$, $t= -17.27$; $p<0.0001$). Similarly, values for $\delta^{15}N$ (13.3 ± 0.3) were also 3.1‰ lower compared to individuals from the sea ($df= 35$, $t= -9.14$; $p<0.0001$). Feeding in these crabs from the Ebro River for two months resulted on octopus' signatures of -19.4 ± 0.2 for $\delta^{13}C$, and 12.2 ± 0.1 for $\delta^{15}N$ (Fig. 2). Hence, experimental results suggest 1.8‰ enrichment for $\delta^{13}C$ and negative values for $\delta^{15}N$ (by -1.1‰), which is coherent with the ca. 1.5‰ higher $\delta^{15}N$ for *C. sapidus* observed in the field, whereas for $\delta^{13}C$ the values were only 0.2‰ higher.

4. Discussion

Our results evince a high blue crab trophic position (TP= 4.4) in open waters of the Ebro Delta (coherent with an average total animal contribution of $>80\%$), very similar to that found for other potential resource competitors of similar size. This situation contrasts with considerably lower TPs found in Ebro Delta Bays and the Tancada lagoon (TP= 2.8), coherent with an average total animal contribution of ca. 57% (Prado et al. 2022); and even more reduced values suggested for

other habitats with sparser animal resources such as drainage channels, the Encanyissada lagoon, and upriver sites. All values observed in the Ebro Delta are, however, within the range reported across invaded Mediterranean locations (Aslan and Polito 2021; Mancinelli et al. 2016, 2017b). The trophic position of organisms is greatly dependent on environmental factors. For instance, blue crabs are active and feeding during warm summer months while most are buried in cold winter months (Hines 2007). Reduced food intake and starvation could be a source of enrichment in $\delta^{15}\text{N}$ as indicated animals are using their own reserves during the fasting period (Oelbermann and Scheu 2002; Haubert et al. 2005; Prado et al. 2021). Alternatively, enhanced consumption of readily available plant material (not available in our study site), and a parallel decrease in cannibalism have also been proposed to account for more reduced trophic levels of blue crabs in winter than in summer, although results were not consistent across sampling locations (Mancinelli et al. 2017b). Given the opportunistic behavior of the blue crab, differences in resource availability across habitat types are, however, expected to be the main source of variability in stable isotopes signatures (Mancinelli et al. 2016, 2017b). In the open sea of the Ebro Delta the high availability of preferred animal items such as mollusks, crustaceans, and fish (Laughlin 1982; Rosas et al. 1994), might enhance TP compared to other estuarine habitats where vegetal material and/or detritus are abundant (Prado et al. 2022).

Trophic position arising from variation in consumer diet has been associated to consumer fitness and linked to evolution via natural selection (Moosmann et al. 2021). Blue crab individuals exclusively fed with an animal diet have shown lower mortality, and enhanced lipid storage and reproductive efforts compared to individuals fed vegetal diets (Belgrad and Griffen 2016a). In turn, population fitness has been indicated a linear relationship with $\log_{10}$ population size, starting at minimum sizes of at least 2000 individuals –to provide adequate levels of buffer against extinction resulting from environmental stochasticity–, and with no apparent maximum asymptote (Reed 2005). Hence, more optimal nutritional conditions in certain areas –particularly those concentrating ovigerous females–, could potentially support enhanced recruitment

success (Phillips 2002) and allow for the development of a large local population capable of sustaining a high-pressure target fishery (López and Rodon 2018; López 2020). Considering the pervasive occupation of all Ebro Delta habitats by blue crab (Prado et al. 2020; Clavero et al. 2022), local measures in the open sea area (e.g., increased local capture of gravid females) could benefit population control at the large-scale.

Blue crab showed a large overlap of nearly half its isotopic niche with the harbor crab, *L. depurator*, and lower but also important overlap occurred with octopus, and 4 species of benthic fish (*L. mormyrus*, *S. rhombus*, *P. lascaris*, and *S. aurata*), pointing that they partly share trophic resources. This overlap concurred with very similar values of TP between blue crab (4.4) and harbor crab (4.46), octopus (3.93) as well as with three of the previous fish species (*L. mormyrus*, *S. rhombus* and *P. lascaris*: 4.06 to 4.67), suggesting that, for the size of organisms investigated, they feature a similar trophic role (see also Mancinelli et al. 2016). Food competition or eating native species of concern has been indicated as a mechanism promoting the success of invasive crustaceans (Weis 2009), and might partly account for the reported vulnerability to the spread of blue crab by other decapods such as *Carcinus aestuarii* (Taybi and Mabrouki 2020; Clavero et al. 2022). Our results suggest that it might also be the case of the harbor crab, a species that is typically a scavenger and a carnivore (Abelló and Cartes 1987) and features a more similar pattern of locomotor activity to blue crab than fish, which can further influence feeding (Moller and Naylor 1980). For benthic fish, very few studies have addressed the impacts of the spread of blue crabs at the population level, but species decreases reported in Ebro Delta estuarine habitats (eel, sandsmelt, toothcarp, and grey mullet) are more arguably driven by predation than by resource competition (Clavero et al. 2022). Conversely, the relatively large isotopic niche overlap between blue crab and octopus –presumably capable of preying on large crabs– requires further investigation, since the assessment of TP assumes the same patterns of isotopic fractionation.

Results of the MixSiar model suggests that the blue crab is preying on all the available benthic resources, including other predatory fish species and crustaceans which constituted the bulk of the local community. This is consistent with pervasive impacts reported at multiple trophic levels (Zenetos et al. 2005; Nehring 2011; Mancinelli et al. 2017a), with possible outcomes depending on predator-prey size variability (Weitz and Levin 2006), the relative abundance of populations, and/ or the predator hunting mode and prey behavior (Belgrad and Griffen 2016b). In the particular case of the harbor crab, the species attains a maximum size of only about 4 cm (carapace width) but can be found at much lower depths of up to ca. 400 m (Abelló et al. 1990), being this partly segregated distribution a possible key aspect for the local persistence of both species. Besides, given their similar isotopic signature, the possible effect of juvenile cannibalism cannot be discarded (up to ca. 5% of the diet), since it has been pointed to as an important factor contributing to population dynamics (Bromilow and Lipcius 2017).

For bivalves, although they are expected to be a preferred food source for the blue crab (Laughlin 1982) a multifactorial decline on the Catalan Coast has been proposed (Baeta et al. 2018) that might account for the fact that they were only consumed at a slightly higher rate than other resources (ca. 5.9 to 6.4% vs. 2.8 to 6.3%). Also, the abundance of other infauna, which are usually a common prey of benthic predatory fish and crabs, such as polychaetes (Laughlin 1982; Whitehouse et al. 2017) was very low and could not be incorporated in the model, although occasional predation might account for some part of the unexplained variation.

The TP of blue crab was very close to other investigated species and results do not show any clear evidence of possible predation control. Unexpectedly, *O. vulgaris*, showed a TP below that of blue crab (3.93 vs. 4.40). Additional fractionation experiments drew a similar outcome: higher $\delta^{15}\text{N}$ in blue crab (13.3 ‰) than in octopus (12.2 ‰), resulting in a negative enrichment of - 1.13‰. Since animals had significantly increased in weight by an average of 816 g WW during the 2-month captivity period, it was arbitrarily considered that they had reached the stage of isotopic equilibrium (see for instance Miller 2006; Prado et al. 2012 for similar experimental

periods). However, $\Delta^{13}\text{C}$ was much higher than that observed in the field (1.8 vs. 0.2‰), suggesting that the use of riverine crabs (lower Ebro River) with lower $\delta^{13}\text{C}$ signatures (Prado et al. 2014, 2022) could have slowed the process. In contrast, $\Delta^{15}\text{N}$, was similar to the -1.6‰ difference observed in the open sea suggesting that predation of blue crab is possible both in captivity and in the natural media (Fiorito et al. 1990; Grisley et al. 1999; this study). Junqueira de Souza Dantas et al. (2020), used a $\Delta^{15}\text{N}$ of 3.3‰ to assess the diet of *O. insularis* with a mixing model, and results pointed to a single dietary resource, the limpet, *Lottia noronhensis*, whereas digestive contents and midden piles observations clearly evidenced crustaceans as the main diet item (by ca. 90-100%). Yet, *Sepia officinalis* fed with a shrimp diet has shown $\Delta^{15}\text{N}$ of ca. 3.3‰ (Hobson and Cherel 2006), suggesting large intragroup variability. Particularly low values of $\Delta^{15}\text{N}$ were reported for mollusks and crustaceans featuring ammonia excretion (Vanderklift and Ponsard 2003). Most mollusks species appear to have limited capacities for interconversion and biosynthesis of amino acids (Bishop 1983) that could have implications for fractionation rates. Additionally, amino acid oxidases of cephalopods, appear to differ from others found in vertebrates (Dixon and Kenworthy, 1967).

Reduced $\Delta^{15}\text{N}$ has been reported for consumers feeding on high-quality food or on diets with a high nitrogen content (Oelbermann and Scheu 2002; Haubert et al. 2005; Prado et al. 2012). Extreme $\Delta^{15}\text{N}$ patterns, ranging from +2.85 to -24.96‰ have been found in laboratory-reared black fly larvae (*Simulium vittatum* IS-7) fed isotopically distinct diets (Overmyer et al. 2008). Among factors potentially accounting for negative fractionation, authors found that molts and feces were enriched in $\delta^{15}\text{N}$ compared to the larvae (by 1.2 to 7.1‰ and up to 143‰, respectively), although other effects such as high metabolism and growth rate, and/ or differential assimilation of nutrients could not be discarded. Further, these patterns might also point to possible homeostatic efforts of consumers, as indicated for nauplii of *Acartia tonsa* (Aberle and Malzahn 2007).

5. Conclusions

Blue crab in open waters displayed a very high TP, comparable to that found in the harbor crab, *L. depurator*, and several predatory fish species. This contrast with TPs found in other habitats of the Ebro Delta where animal prey has become rare (Prado et al. 2022), but agrees with findings pointing to important rates of isotopic niche overlap with native predators, and with dietary modelling suggesting generalist predation. Functional trophic similarity with native species could potentially cause changes in the biodiversity and the functioning of the entire ecosystem (Finke and Denno 2005). In turn, high nutritional status might have positive effects in population fitness (Phillips 2002) and help to maintain local fishing rates. Besides humans, octopus may be one of the few species capable of exerting an effective predation on adult crabs (Fiorito et al., 1990; Grisley et al., 1999; this study). However, preliminary results for $\Delta^{15}\text{N}$ point to unusual metabolic pathways causing a slight depletion of the heavy isotope. Further research is necessary to clarify the metabolic routes involved in octopus $\delta^{15}\text{N}$ fractionation.

Acknowledgements

Financial support for fieldwork and SIA was provided by the Departament d'Agricultura, Ramaderia, Pesca i Alimentació through the BivalCat project (Ref. CAT 152CAT) to M. Ballesteros. P. Prado was contracted under the INIA-CCAA research program for postdoctoral incorporation from the Spanish National Institute for Agricultural and Food Research and Technology (INIA). Stabbing of *O. vulgaris* was supported by the Spanish Government (Ministry of Science and Technology) under the ECESIS project (PID2020-118476RR-C21) to P. Prado and S. Falco. Authors are also grateful to C. Alcaraz for assessment with script preparation and interpretation of SIBER results, and to K.B. Andree, for a double comprehensive review and correction of English throughout the ms. We would also like to thank the detailed review conducted by two anonymous referees that has been of great help for improving the overall quality of the ms.

REFERENCES

- Abelló, P., Pertierra, J.P., Reid, D.G., 1990. Sexual size dimorphism, relative growth and handedness in *Liocarcinus depurator* and *Macropipus tuberculatus* (Brachyura: Portunidae) Sci. Mar. 54, 195–202.
- Aberle, N., Malzahn, A.M., 2007. Interspecific and nutrient-dependent variations in stable isotope fractionation: experimental studies simulating pelagic multitrophic systems. Oecologia 154, 291–303.
- Abelló, P., Cartes, J., 1987., Observaciones sobre la alimentación *Liocarcinus depurator* (L.) (Brachyura: Portunidae) en el Mar Catalán. Invest. Pesq. 51 (Suppl 1), 413–419.
- Aslan, H., Polito, M.J., 2021. Trophic ecology of the Atlantic blue crab *Callinectes sapidus* as an invasive non-native species in the Aegean Sea. Biol. Inv. 23, 2289–2304.
- Baeta, M., Breton, F., Ubach, R., Ariza, E., 2018. A socio-ecological approach to the declining Catalan clam fisheries. Ocean Coast. Manag. 154, 143–154.
- Belgrad, B.A., Griffen, B.D., 2016a. The influence of diet composition on fitness of the blue crab *Callinectes Sapidus*. PLoS ONE 11 (1): e0145481.
- Belgrad, B.A., Griffen, B.D., 2016b. Predator-prey interactions mediated by prey personality and predator hunting mode. Proceed. Royal Soc. B: Biol. Sci. 283 (1828), 20160408.
- Bishop, S.H., Ellis, L.L., Burcham, J.M., 1983. Amino acid metabolism in molluscs. In: Hochachka, P.W. (Ed.), 6- Metabolic biochemistry and molecular biomechanics. Academic Press. pp. 243–327.
- Bouvier, E.L., 1901. Sur un *Callinectes sapidus* M. Rathbun trouvé à Rocheford. Bull. Mus. Hist. Nat. (Paris) 7, 16–17.
- Brett, M.T., 2014. Resource polygon geometry predicts Bayesian stable isotope mixing model bias. Mar. Ecol. Prog Ser. 514, 1–12.
- Bromilow, A.M., Lipcius, R.N., 2017. Mechanisms governing ontogenetic habitat shifts: role of trade-offs, predation, and cannibalism for the blue crab. Mar. Ecol. Prog. Ser. 584, 145–159.

489 Cabral, H.N., Lopes, M., Loeper, R., 2002. Trophic niche overlap between flatfishes in a nursery
490 area on the Portuguese coast. *Sci. Mar.* 66 (3), 293–300.

491 Carrozzo, L., Potenza, L., Carlino, P., Costantini, M.L. Rossi, L., Mancinelli, G., 2014. Seasonal
492 abundance and trophic position of the Atlantic blue crab *Callinectes sapidus* Rathbun 1896 in
493 a Mediterranean coastal habitat. *Rend. Lincei Sci. Fis. Nat.* 25, 201–208.

494 Castejón, D., Guerao, G., 2013. A new record of the American blue crab, *Callinectes sapidus*
495 Rathbun, 1896 (Decapoda: Brachyura: Portunidae), from the Mediterranean coast of the
496 Iberian Peninsula. *BiolInvasions Rec.* 2 (2), 141–143.

497 Clavero, M., Franch, N., Bernardo-Madrid, R., López, V., Abelló, P., Queral, J.M., Mancinelli, G.
498 (2022). Severe, rapid and widespread impacts of an Atlantic blue crab invasion. *Mar. Poll. Bull.*
499 176, 113479.

500 De Souza Dantas, R.J., Leite, T. S., De Albuquerque, C. Q. 2020. Assessing the diet of octopuses:
501 traditional techniques and the stable isotopes approach. *J. Mollusc Stud.* 86 (3), 210–218.

502 Dixon, M., Kenworthy, P., 1967. D-aspartate oxidase of kidney. *BBA-Enzymology*, 146(1), 54–76.

503 Finke, D.L., Denno, R.F., 2005. Predator diversity and the functioning of ecosystems: the role of
504 intraguild predation in dampening trophic cascades. *Ecol. Lett.* 8 (12), 1299–1306.

505 Fiorito, G., von Planta, C., Scotto, P., 1990. Problem solving ability of *Octopus vulgaris* lamarck
506 (Mollusca, Cephalopoda). *Behav. Neural Biol.* 53 (2), 217–230.

507 Grisley, M.S., Boyle, P.R., Pierce, G.J., Key, L.N., 1999. Factors affecting prey handling in lesser
508 octopus (*Eledone cirrhosa*) feeding on crabs (*Carcinus maenas*). *J. Mar. Biol. Assoc. UK* 79(6),
509 1085–1090.

510 Guillory, V., Elliot, M., 2001. A review of blue crab predators. In: Guillory, V., Perry, H., Vanderkooy,
511 S. (Eds.), *Proceedings of the Blue Crab Mortality Symposium*. Gulf States Marine Fisheries
512 Commission, Ocean Springs, Mississippi, vol 90, pp 69–83.

513 Guillory, V., Prejean, P., 2001. Red drum predation on blue crabs. In: Guillory, V., Perry, H.,
 514 VanderKooy, S. (Eds.), Proceedings of the blue crab mortality symposium. Gulf States Marine
 515 Fisheries Commission, Ocean Springs, Mississippi, vol 90, pp. 93–104.
 516 Haubert, D., Langel, R., Scheu, S., Ruess, L., 2005. Effects of food quality, starvation and life stage
 517 on stable isotope fractionation in Collembola. *Pedobiologia* 49, 229–237.
 518 Hayes, C.T., Alford, S.B., Belgrad, B.A., Correia, K.M., Darnell, M.Z., Furman, B.T., ... Darnell, K.M.,
 519 2022. Regional variation in seagrass complexity drives blue crab *Callinectes sapidus* mortality
 520 and growth across the northern Gulf of Mexico. *Mar. Ecol. Progr. Ser.* 693: 141–155.
 521 Heck Jr., K.L., Coen, L.D., 1995. Predation and the abundance of juvenile blue crabs: a comparison
 522 of selected east and gulf coast (USA) studies. *Bull. Mar. Sci.* 57(3): 877–883.
 523 Hines, A.H., 2007. Ecology of juvenile and adult blue crabs. In: Kennedy, V.S., Cronin, L.E. (Eds.),
 524 The blue crab: *Callinectes sapidus*. Maryland Sea Grant College, pp. 565–650.
 525 Hobson, K. A., Cherel, Y., 2006. Isotopic reconstruction of marine food webs using cephalopod
 526 beaks: new insight from captively raised *Sepia officinalis*. *Can. J. Zool.* 84(5), 766–770.
 527 Jackson, A. L., Inger, R., Parnell, A.C., Bearhop, S., 2011. Comparing isotopic niche widths among
 528 and within communities: SIBER–Stable Isotope Bayesian Ellipses in R. *J. Anim. Ecol.* 80(3),
 529 595–602.
 530 Kallianiotis, A., Torre, M., Argyri, A., 2005. Age, growth, mortality, reproduction and feeding
 531 habits of the striped seabream, *Lithognathus mormyrus* (Pisces: Sparidae) in the coastal
 532 waters of the Thracian Sea, Greece. *Sci. Mar.* 69 (3), 391–404.
 533 Laughlin, R.A. 1982. Feeding habits of the blue crab, *Callinectes sapidus* Rathbun, in the
 534 Apalachicola estuary, Florida. *Bull. Mar. Sci.* 32 (4), 807–822.
 535 Leite, T.S., Haimovici, M., Mather, J., 2009. *Octopus insularis* (Octopodidae), evidences of a
 536 specialized predator and a time-minimizing hunter. *Mar. Biol.* 156, 2355–2367.
 537 López, V. 2020. Seguiment del cranc blau (*Callinectes sapidus*) al Delta de l'Ebre. Monverte
 538 Estudis Ambientals, Amposta, pp. 1–127.

539 López, V., Rodon, J., 2018. Diagnosi i situació actual del cranc blau (*Callinectes sapidus*) al delta
 540 de l'Ebre, Direcció General de Pesca i Afers Marítims, Generalitat de Catalunya, pp. 1–86.
 541 [https://agricultura.gencat.cat/ca/ambits/pesca/gestio-pesquera/especies-interes-](https://agricultura.gencat.cat/ca/ambits/pesca/gestio-pesquera/especies-interes-pesquer/crustacis/cranc-blau)
 542 [pesquer/crustacis/cranc-blau](https://agricultura.gencat.cat/ca/ambits/pesca/gestio-pesquera/especies-interes-pesquer/crustacis/cranc-blau)
 543 Mancinelli, G., Chainho, P., Cilenti, L., Falco, S., Kapisir, K., Katselis, G., Ribeiro, F., 2017a. The
 544 Atlantic blue crab *Callinectes sapidus* in southern European coastal waters: distribution,
 545 impact and prospective invasion management strategies. Mar. Pollut. Bull. 119 (1), 5–11.
 546 Mancinelli, G., Raho, D., Zotti, M., Alujević, K., Guerra, M.T., Vizzini, S., 2017b. Trophic flexibility
 547 of the Atlantic blue crab *Callinectes sapidus* in invaded coastal systems of the Apulia region
 548 (SE Italy): A stable isotope analysis. Estuar. Coastal Shelf Sci. 198, 421–431.
 549 Mancinelli, G., Glamuzina, B., Petrić, M., Carrozzo, L., Glamuzina, L., Zotti, M., Raho, D., Vizzini, S.,
 550 2016. The trophic position of the Atlantic blue crab *Callinectes sapidus* Rathbun 1896 in the
 551 food web of Parila Lagoon (South Eastern Adriatic Croatia): A first assessment using stable
 552 isotopes. Mediterr. Mar. Sci. 17, 634–643.
 553 Mancinelli, G., Carrozzo, L., Marini, G., Costantini, M.L., Rossi, L., Pinna, M., 2013. Occurrence of
 554 the Atlantic blue crab *Callinectes sapidus* Rathbun, 1896 in two Mediterranean coastal
 555 habitats: Temporary visitor or permanent resident? Estuar. Coastal Shelf Sci. 135, 46–56.
 556 McCann, M.J., Jensen, O.P., 2018. Laboratory experiments to determine trophic enrichment
 557 factors of stable isotope and fatty acid biomarkers in the blue crab *Callinectes sapidus*. Texas:
 558 Texas A&M University–Corpus Christi. <https://doi.org/10.7266/1.7697>, vol. 7266, pp. N7697.
 559 Metcalfe, N.B., Monaghan, P., 2001. Compensation for a bad start: grow now, pay later?. Trends
 560 Ecol. Evol. 16 (5), 254–260.
 561 Millikin, M.R., Biddle, G.N., Siewicki, T.C., Fortner, A.R., Fair, P.H. 1980. Effects of various levels of
 562 dietary protein on survival, molting frequency and growth of juvenile blue crabs (*Callinectes*
 563 *sapidus*). Aquaculture 19 (2), 149–161.

564 Miller, T.W., 2006. Tissue-specific response of $\delta^{15}\text{N}$ in adult Pacific herring (*Clupea pallasii*)
 565 following an isotopic shift in diet. Environ. Biol. Fishes 76, 177–189.

566 Moller, T.H., Naylor, E., 1980. Environmental influence on locomotor activity in *Nephrops*
 567 *norvegicus* (Crustacea: Decapoda). J. Mar. Biol. Assoc. UK. 60(1), 103–113.

568 Moore, J.W., Semmens, B.X., 2008. Incorporating uncertainty and prior information into stable
 569 isotope mixing models. Ecol. Lett. 11 (5), 470–480.

570 Moosmann, M., Cuenca-Cambronero, M., De Lisle, S., Greenway, R., Hudson, C.M., Lürig, M.D.,
 571 Matthews, B., 2021. On the evolution of trophic position. Ecol. Lett. 24 (12), 2549–2562.

572 Nehring, S., 2011. Invasion history and success of the American blue crab *Callinectes sapidus* in
 573 European and adjacent waters. In: Galil, B.S., Clark, P.F., Carlton, J.T. (Eds.), In the wrong place—
 574 alien marine crustaceans: distribution, biology and impacts, Invading Nature-Springer Series
 575 6, Berlin, pp. 607–624.

576 Oelbermann, K., Scheu S., 2002. Stable isotope enrichment ($\delta^{15}\text{N}$ and $\delta^{13}\text{C}$) in a generalist
 577 predator (*Pardosa lugubris*, Araneae: Lycosidae): effects of prey quality. Oecologia 130, 337–
 578 344.

579 Overmyer, J.P., MacNeil, M.A., Fisk, A.T., 2008. Fractionation and metabolic turnover of carbon
 580 and nitrogen stable isotopes in black fly larvae. Rapid Commun. Mass Spectrom., 22(5), 694–
 581 700.

582 Phillips, N.E., 2002. Effects of nutrition-mediated larval condition on juvenile performance in a
 583 marine mussel. Ecology 83 (9), 2562–2574.

584 Pech-Puch, D., Cruz-López, H., Canche-Ek, C., Campos-Espinosa, G., García, E., Mascaro, M.,
 585 Rosas, C., Chávez-Velasco, D. Rodríguez-Morales, S., 2016. Chemical tools of *Octopus maya*
 586 during crab predation are also active on conspecifics. PLoS One 11(2), e0148922.

587 Post, D.M., Layman, C.A., Arrington, D.A., Takimoto, G., Quattrochi, J., Montana, C.G., 2007.
 588 Getting to the fat of the matter: models, methods and assumptions for dealing with lipids in
 589 stable isotope analyses. Oecologia 152 (1), 179–189.

590 Prado, P., Ibáñez, C., Chen, L., Caiola, N., 2022. Feeding habits and short-term mobility patterns
 591 of blue crab, *Callinectes sapidus*, across invaded habitats of the Ebro Delta subjected to
 592 contrasting salinity. *Estuar. Coasts* 45 (3), 839–855.

593 Prado, P., Cabanes, P., Hernandis, S., García-March, R., Tena, J., 2021. Stable isotope analyses
 594 reveal major nutritional deficiencies in captive vs. field juvenile individuals of *Pinna nobilis*.
 595 *Mar. Environ. Res.* 168, 105304.

596 Prado, P., Peñas, A., Ibáñez, C., Cabanes, P., Jornet, L., Álvarez, N., Caiola, N., 2020. Prey size and
 597 species preferences in the invasive blue crab, *Callinectes sapidus*: Potential effects in marine
 598 and freshwater ecosystems. *Estuar. Coastal Shelf Sci.* 245, 106997.

599 Prado, P., Vergara, C., Caiola, N., Ibáñez, C., 2014. Influence of salinity regime on the food-web
 600 structure and feeding ecology of fish species from Mediterranean coastal lagoons. *Estuar.*
 601 *Coastal Shelf Sci.* 139, 1–10.

602 Prado, P., Carmichael, R.H., Watts, S.A., Cebrian, J., Heck Jr, K.L., 2012. Diet-dependent $\delta^{13}\text{C}$ and
 603 $\delta^{15}\text{N}$ fractionation among sea urchin *Lytechinus variegatus* tissues: implications for food web
 604 models. *Mar. Ecol. Prog. Ser.* 462, 175–190.

605 Reed, D.H., 2005. Relationship between population size and fitness. *Conserv. Biol.* 19.2, 563–568.

606 Rosas, C., Lazaro-Chavez, E., Bückle-Ramirez, F., 1994. Feeding habits and food niche
 607 segregation of *Callinectes sapidus*, *C. rathbunae*, and *C. similis* in a subtropical coastal lagoon
 608 of the Gulf of Mexico. *J. Crust. Biol.* 14 (2), 371–382.

609 Serbetis, C., 1959. Un nouveau crustacé comestible en mer Egeé *Callinectes sapidus* Rath.
 610 (Decapod brach.). *Proceedings GFCM* 5, 505–507.

611 Stock, B.C., Jackson, A.L., Ward, E.J., Parnell, A.C., Phillips, D.L., Semmens, B.X., 2018. Analyzing
 612 mixing systems using a new generation of Bayesian tracer mixing models. *PeerJ* 6, e5096.

613 Stock, B.C. Semmens, B.X. 2016. MixSIAR GUI User Manual. Version 3.1.
 614 <https://github.com/brianstock/MixSIAR/>.<https://doi.org/10.5281/zenodo.47719>.

615 Taieb, A.H., Sley, A., Ghorbel, M., Jarboui, O., 2013. Feeding habits of *Sparus aurata* (Sparidae)
616 from the Gulf of Gabes (central Mediterranean). Cah. Biol. Mar. 54, 263–270.

617 Taybi, A.F., Mabrouki, Y., 2020. The American blue crab *Callinectes sapidus* Rathbun, 1896
618 (Crustacea: Decapoda: Portunidae) is rapidly expanding through the Mediterranean coast of
619 Morocco. Thalassas 36(2), 267–271.

620 Underwood, A.J., 1997. Experiments in ecology: their logical design and interpretation using
621 analysis of variance. Cambridge University Press, Cambridge.

622 Vanderklift, M.A., Ponsard, S., 2003. Sources of variation in consumer–diet $\delta^{15}\text{N}$ enrichment: a
623 meta-analysis. Oecologia 136, 169–182.

624 Vander Zanden, M.J., Cabana, G., Rasmussen, J.B., 1997. Comparing the trophic position of littoral
625 fish estimated using stable nitrogen isotopes ($\delta^{15}\text{N}$) and dietary data. Can. J. Fish. Aquat. Sci.
626 54: 1142–1158.

627 Vasconcelos, P., Carvalho, A.N., Piló, D., Pereira, F., Encarnaç o, J., Gaspar, M.B., Teod sio, M.A.,
628 2019. Recent and consecutive records of the Atlantic blue crab (*Callinectes sapidus* Rathbun,
629 1896): rapid westward expansion and confirmed establishment along the Southern Coast of
630 Portugal. Thalassas 35 (2), 485–494.

631 Weis, J.S., 2010. The role of behavior in the success of invasive crustaceans. Mar. Fresh. Behav.
632 Physiol. 43(2), 83–98.

633 Weitz, J.S., Levin, S.A., 2006. Size and scaling of predator–prey dynamics. Ecol. Lett. 9 (5), 548–
634 557.

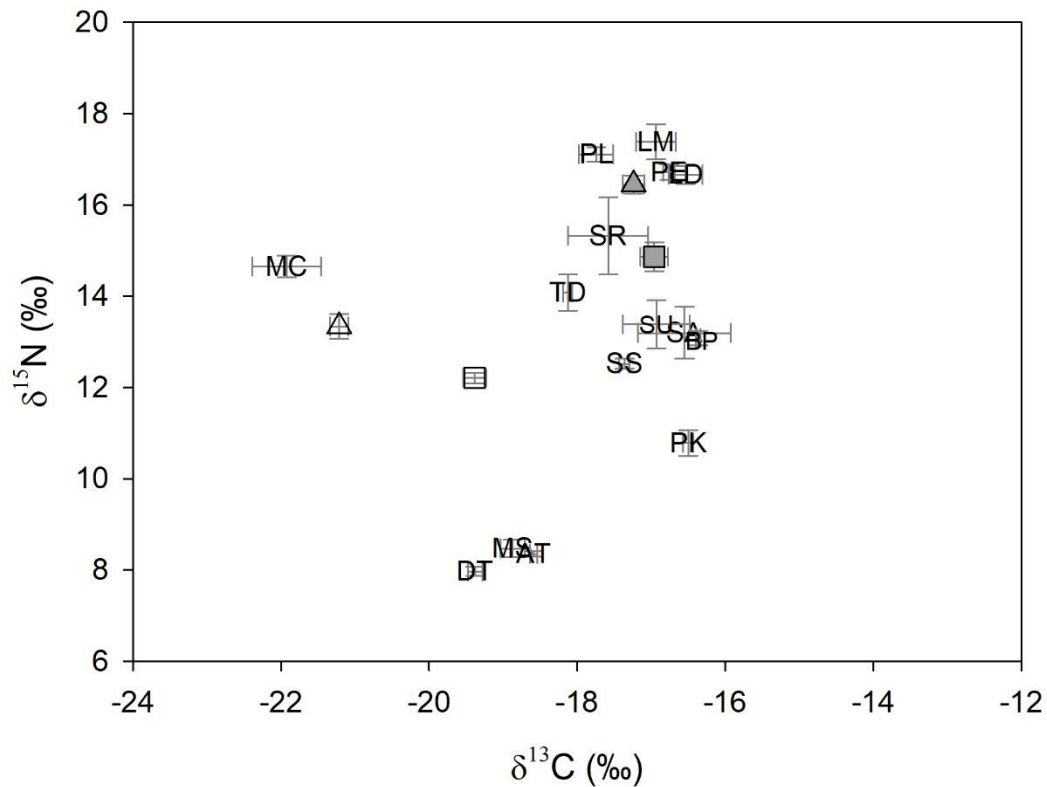
635 Zenetos, A., Corsini-Foka, M., Crocetta, F., Gerovasileiou, V., Karachle, P.K., Simboura, N., Tsiamis,
636 K., Pancucci–Papadopoulou, M.A., 2018. Deep cleaning of alien and cryptogenic species
637 records in the Greek Seas (2018 update). Manag. Biol. Invasions 9 (3), 209–226.

638 Zenetos, A.,  inar, M.E., Pancucci–Papadopoulou, M.A., Harmelin, J.G., Furnari, G., Andaloro, F.,
639 Bellou, N., Streftaris, N., Zibrowius, H., 2005. Annotated list of marine alien species in the
640 Mediterranean with records of the worst invasive species. Mediterr. Mar. Sci. 6 (2), 63–118.

Fig. 1. Map of the Ebro Delta showing the position of the study area (in blue from ca. 40°39'21.23"N; 0°47'51.58"E to 40°37'38.38"N; 0°44'41.01"E) at ca. 12-16 km south from the Ebro River mouth, and the lower section of the Ebro River where crabs were bought to local fishermen for the enrichment experiment.



648 **Fig. 2.** Mean values of $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ of blue crab (triangles) and octopus (squares) from the
 649 Ebro Delta open sea (grey) and the vivarium experiment (white). Values for these items are:
 650 blue crab field $-17.2 \pm 0.1 \delta^{13}\text{C}$ and $16.4 \pm 0.2 \delta^{15}\text{N}$ (N= 25); octopuses' field $-17 \pm 0.2 \delta^{13}\text{C}$ and
 651 $14.9 \pm 0.3 \delta^{15}\text{N}$ (N= 5); blue crab vivarium $-21.2 \pm 0.1 \delta^{13}\text{C}$ and $13.3 \pm 0.3 \delta^{15}\text{N}$ (N= 12); and
 652 octopuses' vivarium $-19.2 \pm 0.2 \delta^{13}\text{C}$ and $12.2 \pm 0.1 \delta^{15}\text{N}$ (N= 13). Other species are indicated
 653 with letters of genus and species names: *Liocarcinus depurator* (LD, N= 10), *Penaeus kerathurus*
 654 (PK, N= 3), *Pegusa lascaris* (PL, N= 5), *Lithognathus mormyrus* (LM, N= 5), *Trachinus draco* (TD,
 655 N= 4), *Pagellus erythrinus* (PE, N= 4), *Scophthalmus rhombus* (SR, N= 6), *Solea senegalensis* (SS,
 656 N= 3), *Mugil cephalus* (MC, N= 4), *Sparus aurata* (SA, N= 5), *Bothus podas* (BP, N= 3), *Sciaena*
 657 *umbra* (SU, N= 3), *Donax trunculus* (DT, N= 7), *Macrura stultorum* (MS, N= 3), *Acanthocardia*
 658 *tuberculata* (AT, N= 14). Error bars are SE.



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Fig 3. Isotopic niches of *Callinectes sapidus* (CS), and other investigated species (LD: *L. depurator* PL: *P. lascaris*, LM: *L. mormyrus*, SR: *S. rhombus*, SA: *Sparus aurata*, DT: *D. trunculus*, AT: *A. tuberculata*). A: Points are individual $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ measurements, and colored solid lines the standard ellipse areas corrected for small sample size (SEAc). B: Boxplots of the SIBER Bayesian ellipse areas (SEA_b in ‰^2) for the different species. 50, 75 and 95% credibility intervals are indicated, and dots represent the modes of the Bayesian distributions. The cross indicates the standard ellipse areas computed using the frequentist algorithm for small samples sizes (SEAc).

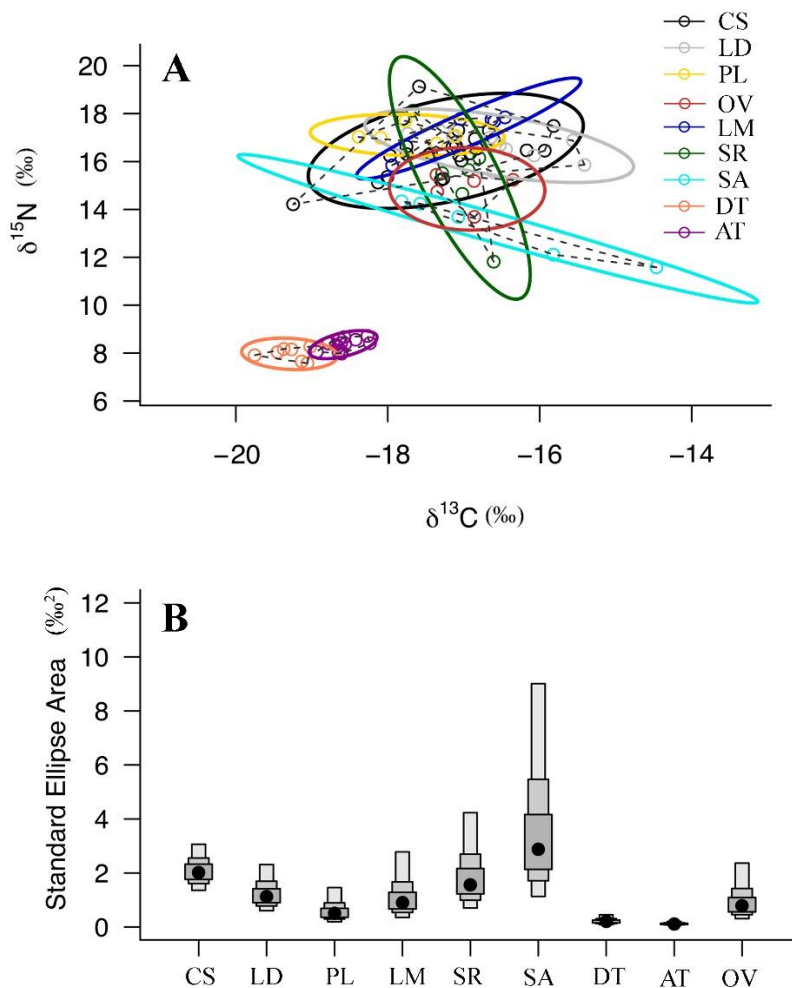


Fig. 4. A: Octopus capturing one blue crab during the stabling period for the evaluation of isotopic enrichment. B: Empty blue crab shells after being consumed by *Octopus vulgaris*.

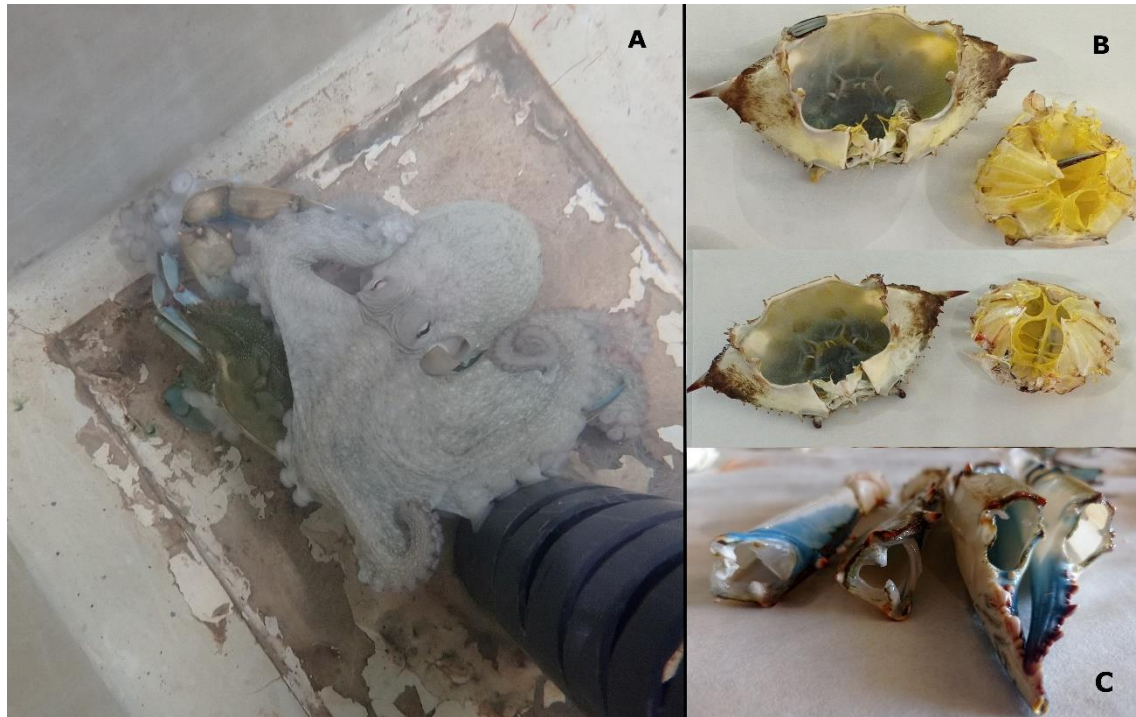


Table 1. Results of Trophic Position (TP) for the different predators investigated in this study. For *C. sapidus*, TP values found in other habitats of the Ebro Delta are also shown (¹Prado et al. 2021) for further comparison.

Animal group	Species	TP
<i>Cephalopoda</i>	<i>Octopus vulgaris</i>	3.93 ± 0.09
<i>Crustaceans</i>	<i>Callinectes sapidus</i> (open sea)	4.40 ± 0.06
	<i>C. sapidus</i> (Alfacs Bay) ¹	3.06 ± 0.03
	<i>C. sapidus</i> (Tancada lagoon) ¹	2.64 ± 0.21
	<i>Penaeus kerathurus</i>	2.73 ± 0.08
	<i>Liocarcinus depurator</i>	4.46 ± 0.06
<i>Fish</i>	<i>Pagellus erythrinus</i>	4.48 ± 0.05
	<i>Lithognathus mormyrus</i>	4.67 ± 0.11
	<i>Sparus aurata</i>	3.44 ± 0.17
	<i>Trachinus draco</i>	3.70 ± 0.12
	<i>Sciaena umbra</i>	3.41 ± 0.05
	<i>Mugil cephalus</i>	3.87 ± 0.07
	<i>Pegusa lascaris</i>	4.59 ± 0.05
	<i>Scophthalmus rhombus</i>	4.06 ± 0.25
	<i>Solea senegalensis</i>	3.24 ± 0.03
	<i>Bothus podas</i>	3.50 ± 0.16

Table 2. MixSiar results (mean, SD and 2.5, 5, 25, 50, 75, 95, and 97.5 percentiles) showing relative contributions of food resources to blue crab diet. Median values are indicated in **bold**. We used the $\Delta^{15}\text{N}$ of 3.4 and a $\Delta^{13}\text{C}$ of 0.4 given for non-herbivorous aquatic consumers (Post 2002), and previously used in blue crab (Mancinelli et al. 2016, 2017a, b; Prado et al. 2021).

		Mean	SD	Percentiles						
				2.5%	5%	25%	50%	75%	95%	97.5%
Bivalves	<i>Acanthocardia tuberculata</i>	0.073	0.056	0.002	0.005	0.028	0.062	0.108	0.180	0.205
	<i>Macra stultorum</i>	0.072	0.056	0.002	0.004	0.026	0.059	0.107	0.180	0.202
	<i>Donax trunculus</i>	0.074	0.053	0.003	0.006	0.031	0.064	0.107	0.176	0.197
Crustaceans	<i>Penaeus kerathurus</i>	0.076	0.059	0.003	0.006	0.030	0.063	0.110	0.194	0.222
	<i>Liocarcinus depurator</i>	0.067	0.055	0.002	0.004	0.023	0.053	0.096	0.179	0.207
Fish	<i>Pagellus erythrinus</i>	0.065	0.052	0.003	0.005	0.024	0.053	0.095	0.169	0.197
	<i>Lithognathus mormyrus</i>	0.060	0.051	0.001	0.003	0.020	0.048	0.088	0.162	0.186
	<i>Sparus aurata</i>	0.072	0.059	0.003	0.005	0.026	0.058	0.104	0.187	0.218
	<i>Trachinus draco</i>	0.072	0.061	0.002	0.004	0.025	0.055	0.103	0.192	0.225
	<i>Sciaena umbra</i>	0.073	0.060	0.002	0.005	0.027	0.058	0.106	0.192	0.222
	<i>Mugil cephalus</i>	0.045	0.033	0.002	0.004	0.017	0.037	0.064	0.108	0.123
	<i>Pegusa lascaris</i>	0.064	0.051	0.002	0.005	0.023	0.053	0.091	0.164	0.190
	<i>Scophthalmus rhombus</i>	0.038	0.035	0.001	0.002	0.012	0.028	0.053	0.107	0.135
	<i>Solea senegalensis</i>	0.074	0.064	0.001	0.003	0.023	0.058	0.107	0.203	0.237
	<i>Bothus podas</i>	0.074	0.062	0.002	0.005	0.026	0.059	0.108	0.193	0.228

Table 3. Summary table of isotopic niche metrics provided by the SIBER package. TA: total area of the convex polygon; SEA_c : standard ellipse area corrected for small sample size; SEA_b : the estimated Bayesian modes of the standard ellipse area.

	CS	LD	PL	LM	SR	SA	DT	AT	OV
TA	7.78	2.79	0.61	0.53	1.88	1.15	0.27	0.29	0.99
SEA_c	2.14	1.32	0.77	0.77	1.85	1.50	0.25	0.12	1.23
SEA_b	1.97	1.11	0.47	0.84	1.60	3.15	0.19	0.10	0.79