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Freshwater Biology

Flow regulation increases food chain length through omnivory mechanisms in a

Mediterranean river network

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Keywords: Dams, Hydrologic alteration, Food webs, Stable isotope analysis, Trophic position.

1 Summary

2	1.	Dams fragment river systems worldwide, and Mediterranean-climate rivers, characterized by
3		highly-seasonal hydrographs and adapted biotas, are particularly impacted by flow
4		regulation. Whereas the effects of flow regulation on hydrology, sediment transport and
5		biodiversity have long been examined, responses at the food-web level remain understudied.
6	2.	Environmental variation is a key control of food-web structure. Thus, we predicted that flow
7		regulation would impact food chain length (FCL) via changes in the flow variation regime,
8		and we tested this prediction in a set of flow-unregulated- to completely regulated reaches in
9		a Mediterranean river basin.
10	3.	In each reach we characterized flow variation, together with two other putative controls of
11		FCL (productivity and habitat size). We combined community data with carbon and nitrogen
12		stable isotopes to estimate food chain length, and Bayesian mixing models allowed
13		estimating dietary proportions of consumers.
14	4.	Flow variation was paramount in controlling FCL in the studied river network, and this same
15		control largely explained the degree of top predators' omnivory. Thus, omnivory
16		mechanisms were the main proximate structural mechanism allowing shifts in food-web
17		structure and linking disturbance regimes to FCL.
18	5.	Our results suggest that flow regulation in Mediterranean rivers may impact food-web
19		structure even when no significant changes in community composition are observed. If
20		highly-variable Mediterranean streams become increasingly muted due to flow regulation,
21		stable conditions could enhance intraguild predation and thus lengthen riverine food chains.
22	K	ywords: Dams, Dynamic stability hypothesis, Hydrologic alteration, Omnivory mechanisms,
23		verine fish, Stream ecology.

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24 Introduction

Dams fragment large river systems across the globe (Nilsson et al. 2005), and effects are remarkable in rivers of Mediterranean climates, owing to the high number of dams located in Mediterranean-climate regions (Nilsson et al. 2005) and to biota in these rivers being adapted to highly-seasonal hydrographs (Gasith & Resh 1999). Dams impact natural flow and temperature regimes (Lehmkuhl 1972; Poff et al. 1997, 2007), sediment transport (Ibàñez, Prat & Canicio 1996; Topping, Rubin & Vierra Jr 2000), and biodiversity both via filtering of life histories (Bunn & Arthington 2002) as well as by facilitation of invasion (Johnson, Olden & Vander Zanden 2008). However, research on flow regulation impacts at the food-web level is still relatively recent, and so far has largely focused on high-order, stable river systems (e.g., Hoeinghaus, Winemiller & Agostinho 2008; Cross et al. 2011, 2013). Because smaller rivers are intrinsically more hydrologically variable (Sabo *et al.* 2010) and discharge variation is a key driver of food-web structure (Power, Dietrich & Finlay 1996; Wootton, Parker & Power 1996; Sabo et al. 2010), dams in highly-seasonal Mediterranean streams may strongly alter food-web structure.

Food chain length (FCL), or the number of trophic transfers between basal resources and top predators, is a key measure of food-web structure. It has long attracted ecological research (Elton 1927; Lindeman 1942), much of which has occurred in freshwater ecosystems (e.g., Vander Zanden et al. 2007; McHugh, McIntosh & Jellyman 2010; Sabo et al. 2010; Schriever & Williams 2013). Despite the variety of implications of FCL, still some controversy exists on which are the mechanisms controlling FCL (reviewed in Post 2002a). Three main non-mutually-exclusive hypotheses compete: (i) The productivity or resource availability hypothesis is based on the energy losses that occur with each trophic transfer, and suggests that more productive

systems should allow longer food chains than less productive ones (Pimm 1982; Schoener 1989).
(ii) The disturbance or dynamic stability hypothesis is based on the fact that in model systems
longer food chains are more vulnerable and take longer to recover than shorter ones, which
implies that food chains should be shorter in highly variable environments (Pimm & Lawton
1977). Finally, (iii) the ecosystem size hypothesis suggests that because larger ecosystems are
often more heterogeneous in habitat, they should host more compartmentalized food webs and
hence allow for longer food chains than smaller ecosystems (Post, Pace & Hairston 2000). Dams
increase water residence time in the impoundment, mute high flows downstream, and increase
inter-flood interval (Poff *et al.* 2007; Sabater 2008; Aristi *et al.* 2014). Thus, the disturbance
hypothesis of FCL should come into play when assessing food-web structure responses to flow
regulation.

Within the food web, variation in FCL can only be explained by (i) the addition or removal of top predators, which changes the identity of the apical predator (additive mechanisms); (ii) by the addition or removal of primary and secondary consumers, which changes the trophic position of top predators (insertion mechanisms); or (iii) by changes in the degree of trophic omnivory by predators (omnivory mechanisms) (Figure 1, adapted from Post & Takimoto 2007). A change in the incidence or identity of the apical predator (i.e., additive mechanism) is probably the most evident and widely-discussed proximate mechanism for variation in FCL (Schoener 1989; Post et al. 2000). In contrast, insertion and omnivory mechanisms have seldom been considered (but see Townsend et al. 1998). These mechanisms assume that FCL may change even if the apical predator remains constant, and thus both may be assessed by measuring changes in the trophic position of predators (Post 2002a). Within this context, a few studies have suggested that foraging shifts of predators (Denno & Fagan 2003)

and variation in the availability or diversity of consumers (McHugh *et al.* 2010; Young *et al.*2013) may cause changes in the vertical structure of food webs. Since omnivory is widespread in
nature (Polis 1991; Denno & Fagan 2003; Diehl 2003; Thompson *et al.* 2007), omnivory
mechanisms may be important controls of FCL across small spatial scales (i.e., when
biogeographical effects are minimal), for instance across reaches differing in flow regulation in a
single river network.

Here we asked how flow regulation affects FCL in a Mediterranean river network, and which proximate structural mechanisms (sensu Post & Takimoto 2007) may explain eventual shifts in food-web structure. To this end we selected 9 stream reaches that were regulated to different extents by several reservoirs and weirs, and characterized the potential environmental controls of FCL (disturbance, productivity, habitat size). We predicted that hydrologic disturbance would be the main control of FCL, with regulation reducing flow variation and thus lengthening food chains (as previously suggested by McHugh et al. 2010; Sabo et al. 2010). We also tested the strength of evidence supporting additive, insertion, and omnivory mechanisms mediating FCL responses to the environment. According to theory, disturbance differentially impairs the higher levels of food chains (Pimm & Lawton 1977). Thus, when top predators in Mediterranean-climate rivers (usually fishes) recolonize after hydrologic disturbance (floods and droughts), they may be forced to feed disproportionally more on low trophic levels in order to meet their energetic demands. By feeding relatively more on basal resources and less on primary and secondary consumers, their trophic position should decrease relative to that held in hydrologically stable, consumer-rich reaches (Figure 1). If this is true, diet shifts of top predators could be an important mechanism linking flow regulation to FCL in rivers, and evidence on this

92 could be obtained by comparing energy flow food webs (as opposed to connectance food webs)93 across sites that differ in flow regulation regimes and FCL.

94 Methods

95 <u>Study site</u>

Field sampling was conducted during spring and summer 2012 in the Siurana River catchment (NE Iberian Peninsula). The Siurana catchment ranges from 23 to 1000 m.a.s.l. and has an extension of 615 km^2 down to its river mouth, where it reaches the Ebro River (Figure 2). Its land use is characterized by coniferous and evergreen oak forests (45 %), shrublands (24 %) and arable rain-fed lands (22 %). The climate is Mediterranean, with precipitation averaging 590 mm vear⁻¹ (concentrated from October to April). The heaviest rainfall usually occurs in autumn and as a result, there are intense flood episodes during autumn in these water courses, which have either permanent flow regimes with low baseflows or temporary flow regimes. We selected 9 reaches from the two main watercourses in the catchment (Siurana and Montsant), with the last reach being located at the confluence (Figure 2). Flows in these reaches were either unregulated or regulated to different extents by three reservoirs, a weir and a bypass (Figure 2). Each study reach (100 m long, ranging between 3.4 and 9.8 m in width) was centered on a pool and contained at least 2 riffles, one upstream and one downstream of the pool. In this setting, all reaches could potentially share the same faunal pool and could be influenced by similar environmental variables (Table S1). We validated this assumption visually and numerically, with multivariate methods: First, a Principal Components Analysis (PCA) ordered reaches according to their physico-chemical parameters, and reaches did not display any directional pattern (Figure S1). Second, permutation-based models (PERMANOVA and Mantel-type tests) combining physico-chemical variables and biota showed that physico-chemical variation did not

1 2		
3 4	115	significantly differ between sections, and did not significantly influence the algae, invertebrate,
5 6	116	or vertebrate compositions (see details in Table S2).
7 8 9	117	Characterizing disturbance, productivity, and habitat size
10 11	118	In order to understand how flow regulation may control FCL in our basin, we first
12 13 14	119	characterized the putative controls of FCL in the light of existing theory (disturbance,
15 16	120	productivity, and size hypotheses).
17 18	121	To define disturbance, following McHugh et al. (2010) we combined several
19 20 21	122	hydrological and geomorphological measures (Table 1). First, we implemented a distributed
22 23	123	hydrological model (TETIS; Francés, Velez & Vélez 2007) to characterize hydrology at each
24 25 26	124	reach. The model was supplied with topographical, geological, soil, and land use information,
27 28	125	considering the presence of the aforementioned reservoirs and diversions. The model was
29 30	126	calibrated and validated using the built-in automatic procedure (Nash-Sutcliffe efficiency = 0.67;
31 32 33	127	see Figure S2 for details). The validation data set consisted of 13 years of daily streamflow data
34 35	128	at the inflow of the Siurana Reservoir. After calibration, the model was run for the whole
36 37 38	129	catchment and delivered daily flow series for the 15 years prior to the study (from 1/1/1998 to
39 40	130	30/06/2012) in each of the 9 reaches. We then analyzed those data using the Indicators of
41 42	131	Hydrologic Alteration (IHA; Richter et al. 1996). With the IHA software v. 7.1 (The Nature
43 44 45	132	Conservancy, Charlottesville, VA, U.S.A.) we computed, for each reach during the entire period,
46 47	133	the number of days with small floods (2-year return interval) and with extreme low flows
48 49	134	(defined as the 10 th percentile of daily flows). These metrics (hereafter, <i>high-</i> and <i>low flows</i>)
50 51 52	135	reflect river rises that overtop the main channel, and drought conditions that may be stressful for
53 54	136	organisms, respectively. Second, bed sediment samples were obtained from exposed gravel bars
55 56 57 58 59 60	137	with low vegetation cover. The surface sediments were characterized using the pebble count

method (Wolman 1954). We then elaborated cumulative grain-size distributions and derived the ratio between the median surface and subsurface grain-sizes (D_{50s} and D_{50ss} respectively) as a measure of bed armoring. We combined the flow metrics, the degree of bed armoring, drainage area (as it mediates the stochastic component of flow variation, sensu Sabo & Post 2008; Sabo et al. 2010) and the number of hydraulic infrastructures regulating streamflow in a Principal Components Analysis (Table S3). Replicating the approach of McHugh et al. (2010), we used the scores of the first axis of this PCA (Table S3) as an integrative "disturbance index" (hereafter, DIST).

We addressed the productivity hypothesis in two complementary ways: by studying ecosystem metabolism (hereafter, *MET*) and by quantifying the availability of basal resources that support the stream food web (hereafter, *RES*). We studied the stream metabolism because this is one of the most integrative measures of river ecosystem functioning, it is meaningful across all types and sizes of streams and rivers, and it indicates energy and material fluxes through ecosystems. Stream ecosystem metabolism rates – gross primary production (GPP) and ecosystem respiration (ER) – can be measured fairly accurately in all types of lotic systems (Tank et al. 2010) using the open-channel technique (Odum 1956; Reichert, Uehlinger & Acuña 2009). In particular, ecosystem flux (i.e., the sum of GPP and ER in absolute values) was used as MET, as an indicator of the total flux of energy in the ecosystem (see detailed methods and measurements in Figure S3).

To characterize RES we mapped the streambed by identifying *in situ* the different patches of biofilm types, macrophytes, and mosses present in each reach. We used a 30×30 cm viewer to estimate the relative % cover of each patch every 30 cm, along eleven transverse transects evenly spaced (i.e., every ten meters) in each reach. When present, we also considered coarse

particulate organic matter (CPOM) of terrestrial origin, such as decaying leaves from riparian trees and herbaceous riparian vegetation; fine particulate organic matter (FPOM); and emerged substrata. Samples of each patch were collected at each reach. Biofilms were differentiated by their color, mucilage presence, and the presence of macrocolonies. We randomly selected five stones from the stream bottom for each of the most representative biofilm patches at each reach. and scraped the stones with a knife to fully detach the algal communities (five 2-10 cm^2 replicates/reach). Samples of macrophytes and mosses were collected using a metallic 900 cm² frame (3 replicates/reach). CPOM and FPOM were collected using a sediment corer (314 cm², 3 replicates/reach). Samples were frozen (-20°C) and processed to obtain ash free dry weight estimates (AFDW, in g/m^2). We finally obtained the AFDW of basal resources per reach by scaling up the AFDW of each patch with the percent cover obtained from the streambed mapping and the previous reach size estimates.

Finally, we characterized habitat size (hereafter, SIZE) as the wetted cross-sectional perimeter (after McHugh et al. 2010). Cross-sectional perimeter was measured in the field at each of the previously-defined eleven transverse transects, and the reach average provided a measure of habitat size that was independent of flow variation (unlike drainage area; Sabo & Post 2008; Sabo et al. 2010). Topographic data were obtained through RTK-GPS surveying. A Leica GS15 Real Time Kinematic GPS system was employed, with topographic 3D data quality (position and elevation) being lower than 0.05 m. Field measures were done in spring and summer to consider seasonal variation.

181 <u>Food-web sampling and sample processing</u>

Potential basal resources included the different biofilm types, macrophytes and
bryophytes, benthic CPOM and FPOM, and suspended particulate organic matter (SPOM). We

collected two composite samples per biofilm type for taxonomy (for diatom and for non-diatom algae respectively, preserved in 4 % formaldehyde), and three samples per biofilm type for SIA (frozen to -20 °C in the field), from the same previously selected stones (2-10 cm²/sample). Up to 400 diatom valves per slide were identified to species level using Nomarski differential interference contrast at 1000x (Nikon Eclipse 80*i*, Japan), whereas non-diatom algae were identified after counting 50 random fields per aliquot. Macrophytes and bryophytes (three samples per patch) were collected and divided into two subsamples for species identification and SIA respectively. Subsamples for taxonomic identification were preserved in zip-lock bags at 4°C and were identified upon arrival in the laboratory. CPOM and FPOM were collected using the same methods previously described for basal resources sampling, and SPOM by filtering stream water through precombusted Whatman GF/F filters (three replicates per reach). These samples were frozen (-20 °C) in the field.

We collected 8 Surber replicates (area 625 cm^2 ; mesh aperture $250 \mu\text{m}$) per reach for benthic macroinvertebrates, covering two different riffle areas and one pool at each reach. These samples were preserved in 70 % ethanol and all individuals were sorted from debris in the laboratory. Taxa were identified to the lowest possible taxonomic level (mostly to species level; Diptera to genus/subfamily level; Annelida to subclass level) and counted. A further qualitative sample across all habitats was collected for SIA using a dip-net (20 cm diameter, mesh aperture $500 \,\mu\text{m}$), and sorted in the field. Individuals were kept for 24 h in filtered stream water to allow for gut clearance, and were subsequently frozen (-20 °C).

Finally, aquatic vertebrates were sampled by depletion electrofishing of the entire 100 m
survey stretch with a SmithRoot backpack engine (200–350 V, 2–3 A fully rectified triphasic
DC). We used stop-nets at both ends of the stretch, and three runs were completed moving

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upstream and sweeping from one side of the stream to the other. All fishes and adult reptiles and
amphibians were identified, counted and measured (length and body mass). In each reach, up to
3 individuals per species and cohort were euthanized and frozen (-20 °C) in the field for SIA.

210 <u>Stable isotope analysis</u>
211 SIA samples were thawed in the laboratory, mollusks were removed from their shells and

all three types of samples (basal resources, macroinvertebrates, and vertebrates) were dried at 212 55 °C until a constant weight was reached. Then, samples were ground using a mortar and pestle. 213 For basal resources, three replicates (one for each sample, 1 - 3 mg each) were loaded into tin 214 capsules. In most cases macroinvertebrate samples consisted of several individuals from the 215 same species and reach (three replicates of \approx 500 µg each). For larger taxa (e.g., Coleoptera, 216 Heteroptera), each replicate came from a different individual. For vertebrate samples, three 217 replicates of muscle tissue were analyzed for each individual ($\approx 500 \,\mu g$ each). This delivered a 218 total of 819 samples. Carbon (C) and nitrogen (N) content and stable isotopes of C and N were 219 analyzed on a Flash 1112 elemental analyzer connected to a Delta C isotopic ratio mass 220 221 spectrometer with Conflo III interphase (Thermo Scientific, Inc.). Stable isotope data were expressed as the relative difference between ratios (R) of samples and standards (PeeDee 222 Belemnite limestone for δ^{13} C and atmospheric nitrogen for δ^{15} N), and expressed in per mille 223 delta notation [i.e., $\delta^{15}N = (R_{sample}/R_{standard}) - 1) \times 1000$]. Analytical precision from multiple runs 224 was 0.1 ‰ for δ^{13} C and 0.2 ‰ for δ^{15} N. 225

226 FCL and dietary proportions

227 On the one hand, we measured FCL following the maximum trophic position (MTP) 228 convention, as follows. First, we visualized δ^{15} N and δ^{13} C isotopic data and grouped samples 229 based on their signals, resulting in six trophic compartments: POM (CPOM+FPOM+SPOM),

1 2				
2 3 4	230	biofilm and algae, macrophytes, primary consumers, secondary consumers, and top predators		
5 6	231	(consumers were classified after Doadrio 2002; Pleguezuelos, Márquez & Lizana 2002; Tachet		
7 8 9	232	<i>et al.</i> 2002). Second, we assessed trophic positions of consumers by comparing their $\delta^{15}N$ to the		
10 11	233	mean δ^{15} N of all basal resources in that reach (assuming a 3.4 % fractionation factor, after Post		
12 13	234	2002b). FCL was then obtained as the maximum trophic position of the top predator species in a		
14 15 16	235	given reach, defined as:		
17 18	236	$\max\{\tau_i i \in T\} $ (Equation 1)		
19 20				
21 22	237	where τ_i is the trophic position of species <i>i</i> from <i>T</i> , the set of putative top predators present in that		
23 24	238	river reach.		
25 26 27	239	On the other hand, dietary proportions were estimated using 'SIAR', an R package that		
28 29	240	solves mixing models for isotopic data within a Bayesian framework (Parnell et al. 2010). SIAR		
30 31	241	estimates probability distributions of resource contributions to a primary consumer diet by		
 32 33 34 242 accounting for variation and uncertainties in the input data. In this study we followed the 				
35 36	243	fractionation factors published in Post (2002b), i.e., 1.3 ± 0.4 % for δ^{13} C and 3.4 ± 1.0 % for		
37 38 39 244 δ^{15} N. Given the potential effects that variability in fractionation factors can have on				
39 40 41	245	stable-isotope mixing models (Bond & Diamond 2011), we validated that inferences on dietary		
42 43	246	proportions did not change if we used one of the lowest fractionation factors for δ^{15} N reported in		
44 45	247	the literature (i.e., 2.54‰; Vanderklift & Ponsard 2003). We also corrected fractionation factors		
46 47 48	248	to account for the number of trophic levels between the consumer and the resources (Phillips <i>et</i>		
49 50	249	<i>al.</i> 2014). Because we had found no visible trace of calcium carbonate on basal resources		
51 52				
53 54	250	samples, we did not acidify them. δ^{13} C data showed however that basal resources were slightly		
55 56	251	enriched relative to primary consumers, with mean δ^{13} C per site ranging from -28.8 to -20.2 ‰		
57 58	252	(study-wide mean: -24.6 ‰) for basal resources, and from -33.4 to -27.9 ‰ (study-wide		
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253	mean: -30.8 ‰) for primary consumers. For secondary consumers, mean δ^{13} C per site ranged
254	from -31.8 to -27.4 ‰ (study-wide mean: -29.7 ‰), and for top predators, from -30.5 to -25.2 ‰
255	(study-wide mean: -28.3 ‰). We ran three different mixing models: (i) one considering the top
256	predators' assemblage as the consumer group, and the 5 lower trophic compartments as potential
257	sources; (ii) one considering the secondary consumers' assemblage as the consumer group, and
258	the 4 immediately-lower trophic compartments as potential sources; and (iii) another one
259	focusing on the most widespread top predator in the study, Barbus sp. (following the same layout
260	as for the top predators' assemblage model). All modeled feeding links were ecologically
261	reasonable according to the literature (Doadrio 2002; Pleguezuelos, Márquez & Lizana 2002;
262	Tachet et al. 2002) and to occasional gut content analyses of collected fish (results not shown).
263	The last model, focusing on <i>Barbus</i> sp., allowed testing if inferences depended on the level
264	ecological organization being studied (i.e., assemblage vs. one particular species).

265 <u>Data analysis</u>

First, we qualitatively assessed variation in realized FCL, in the species identity of the 266 predator holding the highest Maximum Trophic Position (MTP), and in the MTP's of each 267 268 putative top predator, across reaches. We also estimated trophic diversity in each reach by quantifying the isotopic niche space occupied by predators (Layman et al. 2007, 2012). To this 269 end we first visualized δ^{15} N- δ^{13} C predators' isotopic data with convex hulls (i.e., the total area 270 of a convex polygon encompassing all species at a site). Then, we fitted a Standard Ellipse Area 271 (SEA) to each food web. Unlike convex hulls, SEA's are robust to variation in sample size. Here 272 we estimated SEA's via Bayesian inference (following Jackson et al. 2011) to allow for a more 273 robust comparison among local food webs differing in number of species, samples, and hence 274 variability in the input isotopic data. We made 10,000 posterior draws at each site, and plotted 275

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the distribution of posterior ellipses to test for patterns in estimated trophic diversity across
reaches. These analyses were made using 'SIBER' (Stable Isotope Bayesian Ellipses in R), a
subset of functions of the 'SIAR' package (Parnell *et al.* 2010) in R (R Development Core Team
2015) that allows for analysis of data in isotope-space (as opposed to diet-space).
Second, we tested to what extent each putative environmental control explained variation
in FCL. To this end we used linear regressions with DIST, MET, RES and SIZE as explanatory
variables and FCL as a response variable.

Third, we applied an information-theoretic model-selection approach (Burnham & 283 Anderson 2002) to test the weight of evidence supporting each hypothesized environmental 284 control on the proximate structural mechanisms that may allow variation in FCL (i.e., additive, 285 *insertion*, and *omnivory*, Figure 1). We followed a step-wise selection procedure, and models 286 were ranked according to their support, as indicated by Akaike's Information Criterion corrected 287 for small sample size, AICc. We explored additive mechanisms (Figure 1.i) by testing if any 288 289 environmental control was significantly related to differences among reaches in top predator composition (i.e., in the identities of the top-predator assemblage), and in top predator structure 290 (i.e., in the relative abundances of the top-predator species). We used the Bray-Curtis coefficient 291 292 to describe similarities in structure, and the Sorensen coefficient to describe similarities in composition. We addressed insertion mechanisms (Figure 1.ii) similarly, by relating similarities 293 in non-top-predator taxa among these reaches to the set of potential environmental controls. In 294 particular, we performed two additional distance-based linear models; relating putative controls 295 to similarity matrices of invertebrate composition (Sorensen coefficient) and structure (Bray-296 Curtis coefficient). Finally, in order to test for omnivory mechanisms (Figure 1.iii) we used the 297 dietary proportions obtained from the Bayesian mixing models. In this case, we tested if any 298

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3 4	299	environmental control influenced the diets of the top predators' assemblage, of the secondary
5 6 7	300	consumers' assemblage, and of Barbus (thus adding three additional models). If a particular
7 8 9	301	structural mechanism responded significantly to a FCL environmental control, it could
10 11 12	302	potentially mediate environment-FCL relationships.
13 14	303	Last, because variation in predator body size could also influence FCL (i.e., bigger
15 16 17	304	predators can eat bigger prey), we tested if Barbus size distributions changed across reaches, and
18 19	305	if Barbus body size (fork length) correlated with its trophic position.
20 21 22 23	306	Results
24 25 26	307	Food-web overview
27 28 29	308	Macrophytes and bryophytes largely dominated basal resources biomass, with 7
30 31	309	macrophyte and 2 bryophyte species that combined accounted for 75.9 % (\pm 11.9 SE) of the total
32 33 34	310	resource biomass. Biofilm and algae accounted in turn for 5.5 % (\pm 2.3 SE), with 118 taxa of
35 36	311	diatoms and 52 of non-diatom algae and cyanobacteria. A total of 89 macroinvertebrate taxa
37 38	312	were detected, of which 41 were primary consumers and 48 were predators. We also captured 10
39 40 41	313	species of aquatic vertebrates, including 6 putative top predators: the viperine water snake Natrix
42 43	314	maura, and the fishes Anguilla anguilla, Barbus sp. [cf. graellsii x haasi], Gobius lozanoi, Salmo
44 45 46	315	trutta, and Squalius laietanus.
47 48 49	316	Reaches hosted 1 - 4 top predator species and FCL ranged from 1.9 to 4.0 (Table 2,
50 51	317	Figure 3). Natrix maura and Barbus sp. presented a very high MTP range (>1 trophic level),
52 53 54	318	Barbus sp. and Anguilla anguilla were the species that most often set the FCL (in 3 reaches
54 55 56	319	each), and Barbus was the top predator most commonly found, present everywhere except in the
 57 58 59 320 fishless reach M1 (Table 2). The biggest leap in FCL occurred, in both Siurana and Montsa 59 		

rivers, between the most upstream reach (unregulated) and the reach just downstream of the first dam: between reaches M1 and M2 the food chain lengthened 1.5 trophic levels, and between reaches S1 and S2, 0.9 trophic levels (Table 2, Figure 3). Interestingly, FCL was not positively correlated with trophic diversity (Spearman's Rho = -0.06; Figure 3). This indicates that downstream increases in FCL were not driven by an increase in the diversity of trophic pathways used by predators. Instead, trophic diversity decreased non-significantly between reaches M1 and M2; and significantly (as indicated by the non-overlap of 95% credible intervals) between reaches S1 and S2, almost halving (Figure 3). FCL environmental controls Reaches differed in disturbance (DIST), ecosystem metabolism (MET), standing basal resources (RES), and wetted perimeter (SIZE), but showed low covariation among these variables (Figure S4). Hydrological disturbance (DIST) was the most evident FCL environmental control (Figure 4). Lowest hydrological disturbance (DIST) values represent relatively more regulated reaches, larger in drainage area and less affected by high and low flows (Table S3).

Thus, as expected, regulation increased FCL.

336 FCL proximate structural mechanisms

The set of distance-based linear models testing the influence of environmental controls on the three proximate structural mechanisms of FCL provided complementary information (Table 3). Out of the seven models run (two testing for additive, two testing for insertion, and three testing for omnivory mechanisms; Table 3), four showed statistically-significant associations, of which three selected DIST (either alone or combined with another control) as the best predictor. DIST had no effect on additive mechanisms, and had a significant but small effect in insertion

mechanisms (Table 3). In contrast, omnivory mechanisms of the top predators' assemblage were largely explained by DIST; and omnivory mechanisms of *Barbus*, by a combination of DIST and SIZE (Table 3). When comparing dietary proportions across the study reaches, notable patterns were observed for top predators in general and for *Barbus* in particular (Figure 5); the most stable reaches (i.e., those exhibiting longer food chains) hosted top predators that fed more on secondary consumers and less on basal resources (POM, biofilm and algae), whereas the opposite was true in the relatively more disturbed, short-FCL reaches (Figure 5b). Similarly, primary and secondary consumers contributed most to *Barbus* diets in the low bookend of the disturbance gradient (Figure 5c). We found no apparent relationship between *Barbus* individual trophic position and body size, despite the wide range of trophic positions (2.2-3.6) and sizes considered (55-320 mm; Figure 6). Therefore, FCL was not a function of bigger Barbus in some reaches.

355 Discussion

Regulation dampened discharge variation by muting high and low flows, and this unnatural hydrologic stability controlled food chain length (FCL) in a Mediterranean river network. Omnivory mechanisms of top predators allowed food webs to shift from short- to long-FCL configurations, with hydrologic stability increasing intraguild predation (and hence FCL). This finding implies that even if no species colonize or become locally-extirpated as a consequence of flow regulation, food-web structure may change via shifts in the dominant energy pathways. These results may help better understanding the impacts of flow regulation on structure and function of highly-seasonal streams. Free-flowing streams in Mediterranean-climate regions typically exhibit strong intra- and inter-annual variability (Gasith & Resh 1999: Bonada & Resh 2013), hosting biotas with adaptive suites of traits (Bonada, Doledec & Statzner

2007). Dams in Mediterranean rivers lower biodiversity (Clavero, Blanco-Garrido & Prenda
2004) and here we provide evidence that they also impact riverine food-web structure (as
suggested by Power *et al.* 1996).

369 <u>Flow regulation and FCL controls</u>

The observed FCL average $(3.2 \pm 0.2; \text{ mean} \pm \text{SE})$ was slightly lower than the global FCL average reported for streams (i.e., 3.5; Vander Zanden *et al.* 2007). FCL variation across fish-bearing sites in our study (1.7 trophic levels) was higher than the variation between the fishless site and the lowest-FCL fish-bearing site (0.4 trophic levels). Therefore, although dispersal barriers probably prevented fish being part of the local food web in a particular reach (sensu Power & Dietrich 2002), hydrological disturbance shortened food chains consistently, regardless of which group or species held the Maximum Trophic Position in each reach.

We confirmed the prediction that flow variation would be a major determinant of FCL in our basin, with productivity (tested through resource availability and ecosystem metabolism) and habitat size playing secondary roles. The disturbance (or dynamical stability) hypothesis has been supported in a variety freshwater ecosystems, including artificial phytotelmata (Jenkins, Kitching & Pimm 1992) and ponds (Schneider 1997; Schriever & Williams 2013). However, in running waters results have been mixed: some studies did not find relevant effects of bed disturbance (Townsend *et al.* 1998) or hydrological disturbance on FCL (Walters & Post 2008), whereas others reported negative effects (McHugh et al. 2010; Sabo et al. 2010). Differences in methods could cause these inconsistent results, as could the difficulty of disentangling confounding variables (e.g., across different streams or catchments) in large-scale field studies. Our results agree with theory, and with a large-scale field study that reported FCL decreasing with hydrologic variability across 36 American rivers, with drainage area mediating flow

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variation (Sabo *et al.* 2010). In our study regulation scaled flow variation, and equivalent
patterns to those observed by Sabo *et al.* (2010) were found in a single river basin.

The effect of regulation on food-web structure via flow variation may be paramount, but 391 392 our results cannot exclude that regulation may also affect food-web structure via the productivity and the ecosystem size hypotheses, or via the interaction between these two (coined as the 393 productive-space hypothesis; Schoener 1989). In this vein, dams increase the river's capacity to 394 store and process materials and energy in downstream reaches (Aristi et al. 2014; von Schiller et 395 al. 2015), and energy supply has been identified as a significant influence of food-web structure 396 in streams (Thompson & Townsend 2005). Because the effects of productivity and resource 397 availability only seem to be apparent when very low bookends of these controls are considered 398 (Post 2002a), the productivity hypothesis may come into play only if regulation fundamentally 399 expands the range of resource availability and/or GPP and ER. This did not seem to be the case 400 in this study (Figure S3), and our negative result supports previous tests of the effects of GPP or 401 standing crop biomass of resources on riverine FCL (McHugh et al. 2010; Sabo et al. 2010). We 402 403 also acknowledge the short gradient in habitat size considered in our study, which may contribute to explain why this control did not influence FCL. Habitat size effects have been 404 corroborated in well-delimited ecosystems such as lakes (Vander Zanden et al. 1999; Post et al. 405 2000) and ponds (Doi et al. 2009), whereas studies in running waters have been less conclusive 406 (but see Thompson & Townsend 2005; McHugh et al. 2010). A positive association between 407 habitat size and FCL would have probably emerged by means of species-area relationships (Holt 408 et al. 1999) if we had expanded the size gradient by including higher-order reaches. However, 409 such expansion (and hence the habitat size - FCL association) would have not reflected the 410 411 impact of flow regulation by dams, which was the aim of our study.

412 <u>Omnivory mechanisms link flow regulation to FCL</u>

Flow variation also controlled omnivory mechanisms, with top predators in unregulated reaches feeding relatively more on several trophic levels. This pattern matches our prediction and provides empirical evidence of omnivory mechanisms connecting flow variation to FCL in streams. Although this link had been previously proposed (Parker & Huryn 2006; Post & Takimoto 2007; McHugh *et al.* 2010; Takimoto & Post 2013), it had not been yet tested, to our knowledge, in the context of flow regulation.

A previous study had shown that hydrologic disturbance may shorten arctic stream food chains by limiting the contribution of secondary consumers to predator fish diets (Parker & Huryn 2006). Similarly, Lepidodactylus geckos had higher proportions of predators in their stomach contents when found on high- than on low-productivity islets, with productivity and predator size also influencing FCL across the set of islets (Young et al. 2013). Because in nature prey tend to be smaller than their predators (Woodward *et al.* 2005), variation in predator body size could be actually driving omnivory and hence FCL patterns (Hoeinghaus *et al.* 2008). However, when we tested this hypothesis we found no apparent relationship between these variables. Therefore, even if flow variation regimes could partly explain top predators' size distributions across our reaches (as in Jellyman, McHugh & McIntosh 2014), changes in their trophic positions likely reflected "forced" diet shifts due to disturbance-driven scarcity of invertebrates, rather than ontogenetic diet shifts.

431 <u>Concluding remarks</u>

Changes in FCL may present relevant implications in terms of pollutant bioaccumulation
by animals (Cabana & Rasmussen 1994), nutrient recycling (McIntyre *et al.* 2007), and carbon
exchange between freshwater ecosystems and the atmosphere (Schindler *et al.* 1997). It is

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therefore important to understand the links between increasing hydrologic alteration in rivers and FCL. It had been previously suggested that the ultimate effects of river regulation on FCL may depend on whether regulation increases the proportion of "protected" consumers (e.g., predator-resistant grazers), which may subsequently disrupt energy flow to higher trophic levels and thus reduce the abundance of intraguild-predators (Wootton et al. 1996). In our study we found no evidence of "protected" consumers subsuming the effects of flow regulation (confirmed by occasional gut content analyses of collected fish). Instead, effects rippled through the food web via omnivory mechanisms—ultimately affecting FCL. Our design tested for flow-regulation effects in a semi-controlled manner (i.e., in a single small basin and pool of organisms), but this was at the expense of sacrificing replication. Further research should ask whether food chain lengthening is a general or a contingent consequence of flow regulation; whether these effects are consistent intra- and inter-annually; and whether omnivory mechanisms are universal or replaced by additions/insertions when larger spatial scales (and hence potential predator extirpations due to riverine habitat fragmentation) are considered. Given the multitude of dams already present or planned across the globe, it is crucial that we advance our understanding on their impacts at the higher levels of biological organization.

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Aristi I., Arroita M., Larrañaga A., Ponsatí L., Sabater S., Schiller D., et al. (2014) Flow

regulation by dams affects ecosystem metabolism in Mediterranean rivers. Freshwater

4 6

References

Biology **59**, 1816–1829.

462 463 464	Bonada N., Doledec S. & Statzner B. (2007) Taxonomic and biological trait differences of stream macroinvertebrate communities between mediterranean and temperate regions: implications for future climatic scenarios. <i>Global Change Biology</i> 13, 1658–1671.
465 466	Bonada N. & Resh V.H. (2013) Mediterranean-climate streams and rivers: geographically separated but ecologically comparable freshwater systems. <i>Hydrobiologia</i> 719 , 1–29.
467 468	Bond A.L. & Diamond A.W. (2011) Recent Bayesian stable-isotope mixing models are highly sensitive to variation in discrimination factors. <i>Ecological Applications</i> 21 , 1017–1023.
469 470	Bunn S.E. & Arthington A.H. (2002) Basic principles and ecological consequences of altered flow regimes for aquatic biodiversity. <i>Environmental Management</i> 30 , 492–507.
471 472 473 474	Bunte K. & Abt S.R. (2001) Sampling surface and subsurface particle-size distributions in wadable gravel- and cobble-bed streams for analyses in sediment transport, hydraulics, and streambed monitoring. US Department of Agriculture, Forest Service, Rocky Mountain Research Station Fort Collins, Colorado.
475 476	Burnham K.P. & Anderson D.R. (2002) <i>Model selection and multimodel inference: a practical information-theoretic approach</i> , 2nd edn. Springer, New York.
477 478	Cabana G. & Rasmussen J.B. (1994) Modelling food chain structure and contaminant bioaccumulation using stable nitrogen isotopes. <i>Nature</i> 372 , 255–257.
479 480 481	Clavero M., Blanco-Garrido F. & Prenda J. (2004) Fish fauna in Iberian Mediterranean river basins: biodiversity, introduced species and damming impacts. <i>Aquatic Conservation:</i> <i>Marine and Freshwater Ecosystems</i> 14, 575–585.
482 483 484	 Cross W.F., Baxter C. V, Donner K.C., Rosi-Marshall E.J., Kennedy T.A., Hall Jr R.O., <i>et al.</i> (2011) Ecosystem ecology meets adaptive management: food web response to a controlled flood on the Colorado River, Glen Canyon. <i>Ecological Applications</i> 21, 2016–2033.
485 486 487	Cross W.F., Baxter C. V, Rosi-Marshall E.J., Hall Jr R.O., Kennedy T.A., Donner K.C., <i>et al.</i> (2013) Food-web dynamics in a large river discontinuum. <i>Ecological Monographs</i> 83 , 311–337.
488 489	Denno R.F. & Fagan W.F. (2003) Might nitrogen limitation promote omnivory among carnivorous arthropods? <i>Ecology</i> 84, 2522–2531.
490 491	Diehl S. (2003) The evolution and maintenance of omnivory: dynamic constraints and the role of food quality. <i>Ecology</i> 84 , 2557–2567.
492 493	Doadrio I. (2002) <i>Atlas y libro rojo de los peces continentales de España</i> . Dirección General de Conservación de la Naturaleza: Museo Nacional de Ciencias Naturales, Madrid.
494 495	Doi H., Chang KH., Ando T., Ninomiya I., Imai H. & Nakano S. (2009) Resource availability and ecosystem size predict food-chain length in pond ecosystems. <i>Oikos</i> 118 , 138–144.
496	Elton C.S. (1927) Animal ecology. Macmillan Co., New York.
	22

2		
3 4 5	497 498	Francés F., Velez J.I. & Vélez J.J. (2007) Split-parameter structure for the automatic calibration of distributed hydrological models. <i>Journal of Hydrology</i> 332, 226–240.
6 7 8 9	499 500 501	 Gasith A. & Resh V.H. (1999) Streams in Mediterranean climate regions: abiotic influences and biotic responses to predictable seasonal events. <i>Annual Review of Ecology and Systematics</i> 30, 51–81.
10 11 12 13	502 503 504	Hoeinghaus D.J., Winemiller K.O. & Agostinho A.A. (2008) Hydrogeomorphology and river impoundment affect food-chain length of diverse Neotropical food webs. <i>Oikos</i> 117, 984– 995.
14 15 16	505 506	Holt R.D., Lawton J.H., Polis G.A. & Martinez N.D. (1999) Trophic rank and the species-area relationship. <i>Ecology</i> 80 , 1495–1504.
17 18 19 20	507 508 509	Ibàñez C., Prat N. & Canicio A. (1996) Changes in the hydrology and sediment transport produced by large dams on the lower Ebro river and its estuary. <i>Regulated Rivers: Research & Management</i> 12, 51–62.
20 21 22 23 24	510 511 512	Jackson A.L., Inger R., Parnell A.C. & Bearhop S. (2011) Comparing isotopic niche widths among and within communities: SIBER – Stable Isotope Bayesian Ellipses in R. <i>Journal of</i> <i>Animal Ecology</i> 80 , 595–602.
25 26	513 514	Jellyman P.G., McHugh P.A. & McIntosh A.R. (2014) Increases in disturbance and reductions in habitat size interact to suppress predator body size. <i>Global Change Biology</i> 20 , 1550–1558.
27 28 29	515 516	Jenkins B., Kitching R.L. & Pimm S.L. (1992) Productivity, disturbance and food web structure at a local spatial scale in experimental container habitats. <i>Oikos</i> 65, 249–255.
30 31 32 33	517 518 519	Johnson P.T.J., Olden J.D. & Vander Zanden M.J. (2008) Dam invaders: impoundments facilitate biological invasions into freshwaters. <i>Frontiers in Ecology and the Environment</i> 6 , 357–363.
34 35 36 37	520 521 522	Layman C.A., Araujo M.S., Boucek R., Hammerschlag-Peyer C.M., Harrison E., Jud Z.R., <i>et al.</i> (2012) Applying stable isotopes to examine food-web structure: an overview of analytical tools. <i>Biological Reviews</i> 87, 545–562.
38 39 40	523 524	Layman C.A., Arrington D.A., Montaña C.G. & Post D.M. (2007) Can stable isotope ratios provide for community-wide measures of trophic structure? <i>Ecology</i> 88 , 42–48.
41 42 43	525 526	Lehmkuhl D.M. (1972) Change in thermal regime as a cause of reduction of benthic fauna downstream of a reservoir. <i>Journal of the Fisheries Board of Canada</i> 29 , 1329–1332.
44	527	Lindeman R.L. (1942) The trophic-dynamic aspect of ecology. <i>Ecology</i> 23, 399–417.
45 46 47	528 529	McHugh P.A., McIntosh A.R. & Jellyman P.G. (2010) Dual influences of ecosystem size and disturbance on food chain length in streams. <i>Ecology Letters</i> 13 , 881–890.
48 49 50 51	530 531 532	McIntyre P.B., Jones L.E., Flecker A.S. & Vanni M.J. (2007) Fish extinctions alter nutrient recycling in tropical freshwaters. <i>Proceedings of the National Academy of Sciences</i> 104, 4461–4466.
52 53 54	533 534	Nilsson C., Reidy C.A., Dynesius M. & Revenga C. (2005) Fragmentation and flow regulation of the world's large river systems. <i>Science</i> 308 , 405–408.
55 56 57 58 59 60	535 536	Odum H.T. (1956) Primary production in flowing waters. <i>Limnology and Oceanography</i> 1 , 102–117.

3 4 5	537 538	5					
6 7 8	539 540	Parnell A.C., Inger R., Bearhop S. & Jackson A.L. (2010) Source partitioning using stable isotopes: coping with too much variation. <i>PLOS one</i> 5 , e9672.					
9 10 11 12 13	541 542 543	Phillips D.L., Inger R., Bearhop S., Jackson A.L., Moore J.W., Parnell A.C., <i>et al.</i> (2014) Best practices for use of stable isotope mixing models in food-web studies. <i>Canadian Journal</i> <i>Zoology</i> 92, 823–835.					
	544	Pimm S.L. (1982) Food webs. Chapman and Hall, London.					
14 15 16	545 546	Pimm S.L. & Lawton J.H. (1977) Number of trophic levels in ecological communities. <i>Nature</i> 268 , 329–331.	?				
17 18 19 20	547 548 549	Pleguezuelos J.M., Márquez R. & Lizana M. (2002) <i>Atlas y libro rojo de los anfibios y reptiles de España</i> . Dirección General de la Conservación de la Naturaleza - Asociación Herpetológica Española, Madrid.	5				
21 22 23	550 551	Poff N.L., Allan J.D., Bain M.B., Karr J.R., Prestegaard K.L., Richter B.D., <i>et al.</i> (1997) The natural flow regime. <i>BioScience</i> 47, 769–784.					
24 25 26 27	552 553 554	Poff N.L., Olden J.D., Merritt D.M. & Pepin D.M. (2007) Homogenization of regional river dynamics by dams and global biodiversity implications. <i>Proceedings of the National Academy of Sciences</i> 104 , 5732–5737.					
28 29 30	555 556	Polis G.A. (1991) Complex trophic interactions in deserts: an empirical critique of food-web theory. <i>American Naturalist</i> 138 , 123–155.					
31 32	557 558	Post D.M. (2002a) The long and short of food-chain length. <i>Trends in Ecology & Evolution</i> 17 269–277.	Ι,				
33 34 35	559 560	Post D.M. (2002b) Using stable isotopes to estimate trophic position: models, methods, and assumptions. <i>Ecology</i> 83 , 703–718.					
36 37 38	561 562	Post D.M., Pace M.L. & Hairston N.G. (2000) Ecosystem size determines food-chain length in lakes. <i>Nature</i> 405 , 1047–1049.	1				
39 40 41	563 564	Post D.M. & Takimoto G. (2007) Proximate structural mechanisms for variation in food-chain length. <i>Oikos</i> 116 , 775–782.	l				
41 42 43 44 45 46 47 48 49 50 51 52 53 45 56 57 58	565 566	Power M.E. & Dietrich W.E. (2002) Food webs in river networks. <i>Ecological Research</i> 17, 45 471.	1–				
	567 568 569	Power M.E., Dietrich W.E. & Finlay J.C. (1996) Dams and downstream aquatic biodiversity: potential food web consequences of hydrologic and geomorphic change. <i>Environmental management</i> 20 , 887–895.					
	570 571	R Development Core Team (2015) R: a language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria.	-				
	572 573 574	Reichert P., Uehlinger U. & Acuña V. (2009) Estimating stream metabolism from oxygen concentrations: Effect of spatial heterogeneity. <i>Journal of Geophysical Research:</i> <i>Biogeosciences (2005–2012)</i> 114.					
	575 576	Richter B.D., Baumgartner J. V, Powell J. & Braun D.P. (1996) A method for assessing hydrologic alteration within ecosystems. <i>Conservation Biology</i> 10 , 1163–1174.					
59 60			24				

1 2			
3 4 5	577 578	Sabater S. (2008) Alterations of the global water cycle and their effects on river structure, function and services. <i>Freshwater Reviews</i> 1, 75–88.	
6 7 8	579 580	Sabo J.L. & Post D.M. (2008) Quantifying periodic, stochastic, and catastrophic environmental variation. <i>Ecological Monographs</i> 78 , 19–40.	
9 10 11 12 13 14	581 582	Sabo J.L.L., Finlay J.C.C., Kennedy T. & Post D.M.M. (2010) The role of discharge variation in scaling of drainage area and food chain length in rivers. <i>Science</i> 330 , 965–967.	
	583 584 585	von Schiller D., Aristi I., Ponsatí L., Arroita M., Acuña V., Elosegi A., <i>et al.</i> (2015) Regulation causes nitrogen cycling discontinuities in Mediterranean rivers. <i>Science of The Total Environment, doi:10.1016/j.scitotenv.2015.07.017</i> .	
15 16 17 18	586 587 588	Schindler D.E., Carpenter S.R., Cole J.J., Kitchell J.F. & Pace M.L. (1997) Influence of food web structure on carbon exchange between lakes and the atmosphere. <i>Science</i> 277, 248– 251.	
19 20 21	589 590	Schneider D.W. (1997) Predation and food web structure along a habitat duration gradient. <i>Oecologia</i> 110 , 567–575.	
22 23 24 25 26 27 28 29 30 31 32 33 34 35 36	591 592	Schoener T.W. (1989) Food webs from the small to the large: the Robert H. MacArthur award lecture. <i>Ecology</i> 70 , 1559–1589.	
	593 594	Schriever T.A. & Williams D.D. (2013) Influence of pond hydroperiod, size, and community richness on food-chain length. <i>Freshwater Science</i> 32 , 964–975.	
	595 596	Tachet H., Richoux P., Bournaud M. & Usseglio-Polatera P. (2002) Invertebrés d'eau douce, 2nd corrected ed. CNRS éditions, Paris.	
	597 598	Takimoto G. & Post D. (2013) Environmental determinants of food-chain length: a meta- analysis. <i>Ecological Research</i> 28 , 675–681.	
	599 600 601	Tank J.L., Rosi-Marshall E.J., Griffiths N.A., Entrekin S.A. & Stephen M.L. (2010) A review of allochthonous organic matter dynamics and metabolism in streams. <i>Journal of the North American Benthological Society</i> 29 , 118–146.	
37 38 39	602 603	Thompson R.M., Hemberg M., Starzomski B.M. & Shurin J.B. (2007) Trophic levels and trophic tangles: the prevalence of omnivory in real food webs. <i>Ecology</i> 88 , 612–617.	2
40 41 42	604 605	Thompson R.M. & Townsend C.R. (2005) Energy availability, spatial heterogeneity and ecosystem size predict food-web structure in streams. <i>Oikos</i> 108 , 137–148.	
42 43 44 45 46	606 607 608	Topping D.J., Rubin D.M. & Vierra Jr L.E. (2000) Colorado River sediment transport 1. Natural sediment supply limitation and the influence of Glen Canyon Dam. <i>Water Resources Research</i> 36 , 515–542.	
47 48 49	609 610	Townsend, Thompson, McIntosh, Kilroy, Edwards & Scarsbrook (1998) Disturbance, resource supply, and food-web architecture in streams. <i>Ecology Letters</i> 1, 200–209.	
50 51	611 612	Vanderklift M.A. & Ponsard S. (2003) Sources of variation in consumer-diet δ15N enrichment: a meta-analysis. <i>Oecologia</i> 136 , 169–182.	ł
52 53 54	613 614	Walters A.W. & Post D.M. (2008) An experimental disturbance alters fish size structure but not food chain length in streams. <i>Ecology</i> 89 , 3261–3267.	
55 56 57	615 616	Wolman M.G. (1954) A method of sampling coarse river-bed material. American Geophysical Union Transactions 35, 951–956.	
58 59 60		25	5

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- 3 4 5	617 618	Woodward G., Ebenman B., Emmerson M., Montoya J.M., Olesen J.M., Valido A., <i>et al.</i> (2005)Body size in ecological networks. <i>Trends in ecology & evolution</i> 20, 402–409.
6 7 8	619 620	Wootton J.T., Parker M.S. & Power M.E. (1996) Effects of disturbance on river food webs. <i>Science</i> 273 , 1558–1561.
9 10 11 12	621 622 623	Young H.S., McCauley D.J., Dunbar R.B., Hutson M.S., Ter-Kuile A.M. & Dirzo R. (2013) The roles of productivity and ecosystem size in determining food chain length in tropical terrestrial ecosystems. <i>Ecology</i> 94 , 692–701.
13 14	624 625	Vander Zanden M.J., Fetzer W.W., Jake Vander Zanden M. & W. Fetzer W. (2007) Global patterns of aquatic food chain length. <i>Oikos</i> 116, 1378–1388.
15 16 17	626 627	Vander Zanden M.J., Shuter B.J., Lester N. & Rasmussen J.B. (1999) Patterns of food chain length in lakes: a stable isotope study. <i>The American Naturalist</i> 154, 406–416.
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2 3 4 5	629	Supporting Information	
6 7 8	630	Table S1. Summary of the physico-chemical variables considered.	
9 10 11	631	Table S2. Permutation-based models combining data on physico-chemical variables and biota	ì.
12 13 14	632	Table S3. Details of the Principal Component Analysis (PCA) combining hydrological and	
15 16 17	633	geomorphological data.	
18 19 20	634	Figure S1. Principal Components Analysis (PCA) ordering the 9 studied reaches according to)
21 22 23	635	their physico-chemical characteristics.	
23 24 25 26	636	Figure S2. Observed (Q obs) versus simulated flow (Q sim) at the Siurana reservoir.	
27 28 29	637	Figure S3. Ecosystem metabolism (MET) across the studied reaches.	
$\begin{array}{c} 30\\ 31\\ 32\\ 33\\ 34\\ 35\\ 36\\ 37\\ 38\\ 39\\ 40\\ 41\\ 42\\ 43\\ 44\\ 45\\ 46\\ 47\\ 48\\ 49\\ 50\\ 51\\ 52\\ 53\\ 54\\ 55\\ 56\\ 57\\ 58\\ 59\\ \end{array}$	638	Figure S4. Scatter plots showing the correlations among the hypothesized environmental	
	639	controls of FCL (<i>DIST, MET, RES, SIZE</i>).	
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640 Tables

Table 1. Site characteristics and data used to compute the multivariate stability index (DIST).

Drainage area and hydraulic infrastructure control the stochastic component of flow variation

643 (after Sabo *et al.* 2010). The number of low- and high-flow days was obtained after Richer *et al.*

(1996), using mean daily flow series from 1/1/1998 to 30/06/2012 (see text for details). The

degree of bed armoring is the ratio between the median surface (D_{50s}) and the median subsurface

646 (D_{50ss}) material (Bunte & Abt 2001) and indicates absence of sediment supply from upstream.

647	M = Montsant river; S = Siurana	river; CON = confluence of Montsant and Siurana rivers.

Reach	ch UTM Drainage (x, y) area (km ²)		Regulation	Low flows	High flows	Bed armoring	
M1	824850, 4582650	40.66	- 6	50	56	3.15	
M2	816050, 4578550	97.58	Margalef reservoir	26	22	4.92	
M3	814850, 4577650	113.1	Margalef reservoir	35	22	4.03	
M4	810850, 4573050	141.4	Margalef reservoir	24	28	2.98	
S1	830350, 4574750	35.64	-	68	56	5.08	
S2	827750, 4573750	60.96	Siurana reservoir	2	13	8.02	
S 3	826050, 4572150	88.15	Siurana reservoir	2	30	4.98	
S 4	818050, 4569050	140.2	Siurana reservoir, Riuedecanyes weir and bypass	18	32	4.43	
CON	814150, 4564150	421.6	Margalef reservoir, Vilella reservoir, Siurana reservoir, Riudecanyes weir and bypass	13	40	2.56	

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648	Table 2. Maximum trophic position (MTP) of the putative top predator species in each reach.
649	The value of the species holding the absolute MTP in each reach (i.e., the one setting FCL) is
650	shown in italics. $\Delta_{MTP} = MTP$ range across reaches. M = Montsant river; S = Siurana river; CON
651	= confluence of Montsant and Siurana rivers.

Top predator	M1	M2	M3	M4	S1	S2	S3	S4	CON	$\Delta_{\rm MTP}$
Natrix maura	1.93			3.14		3.26		3.57	3.48	1.64
(reptile)										
Anguilla anguilla		3.43	3.47						4.00	0.57
(fish)										
Barbus sp.		3.31	3.35	3.38	2.34	3.23	3.43	3.49	3.64	1.30
(fish)										
Gobius lozanoi		3.17	3.05	3.15	6					0.12
(fish)										
Salmo trutta		3.36				Q				-
(fish)										
Squalius laietanus					2.15			3.64	3.37	1.49

Table 3. Set of distance-based linear models testing the influence of environmental controls on the proximate structural mechanisms (additive, insertion, and omnivory mechanisms) of FCL variation (DIST = disturbance, MET = ecosystem metabolism, RES = basal resource availability, *SIZE* = wetted perimeter; see text for details). A step-wise selection procedure based on 9,999 permutations allowed identifying the environmental controls (Var.) that minimized AICc for each of the 7 models. The test statistic (Pseudo-F) and P-value of the selected controls are shown for each best model, together with the marginal percentage of explained variance after entering the variable (Prop.) and the corresponding degrees of freedom (df) of the model. In all cases except for *Barbus* sp. dietary proportions, the best model involved only one control. Statistically-significant control-mechanism associations are shown in bold.

Test	Response matrix	Var	df	AIC _c	SS (trace)	Pseudo-F	Р	Prop.
tive nisms	Top predators composition	RES	7	65.58	5188	5.377	0.036	0.434
Additive mechanisms	Top predators structure	RES	7	71.57	3830	2.041	0.065	0.226
Insertion mechanisms	Invertebrate composition	MET	7	67.78	2050	1.664	0.097	0.192
	Invertebrate structure	DIST	7	70.85	2966	1.713	0.044	0.197
Omnivory mechanisms	Top predators dietary proportions	DIST	7	52.31	1898	8.600	0.002	0.551
	Secondary consumers dietary proportions	RES	7	51.35	366.1	1.846	0.213	0.209
	Barbus sp.	SIZE	6	46.05	823.9	4.352	0.048	0.420
	dietary proportions	DIST	5	43.02	749.6	9.703	0.024	0.382

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Figure legends

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Figure 1. Schematization of the proximate structural mechanisms that allow FCL variation

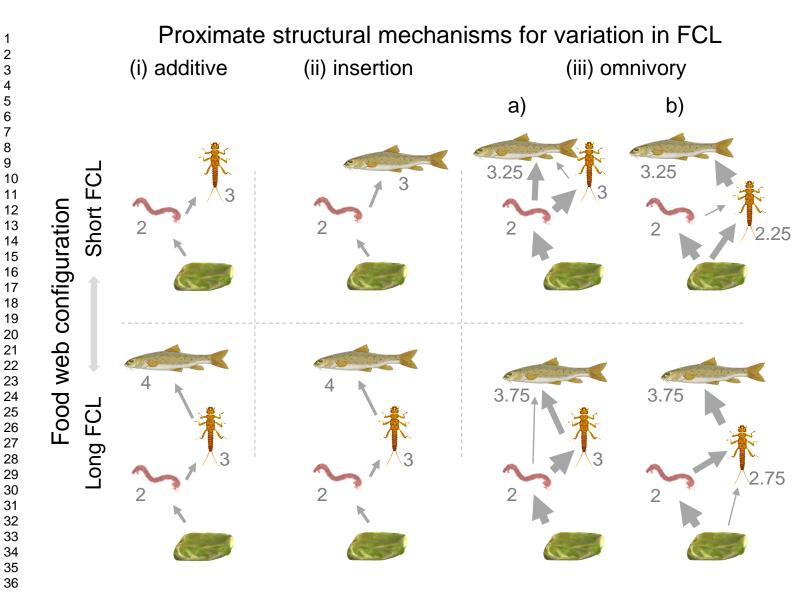
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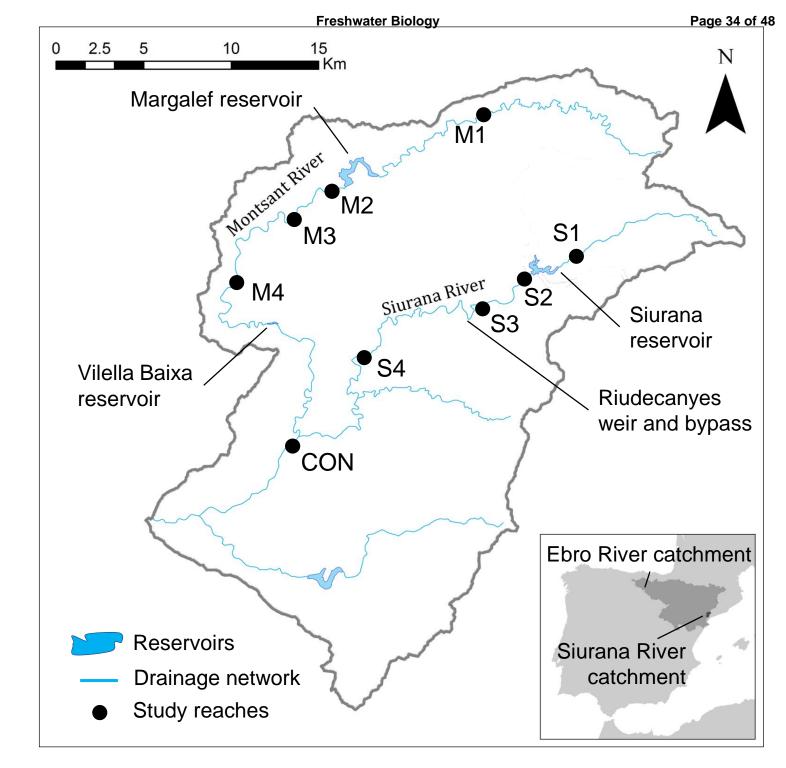
001	Figure 1 . Senemalization of the proximate structural meenanisms that anoth 1 eD variation
665	(adapted from Post & Takimoto 2007). Stones with green algae represent basal resources,
666	Orthocladiinae non-biting midges represent primary consumers, Isoperla stoneflies represent
667	secondary consumers, and Catalonian barbels Barbus haasi represent top predators. The three
668	mechanisms may explain transitions between shorter (~3-3.25) and longer (~3.75-4) food-chain
669	configurations (numbers represent trophic positions). For (i) additive and (ii) insertion
670	mechanisms, constant energy fluxes are assumed between short- and long-FCL food web
671	configurations (connectance food webs). In contrast, (iii) omnivory mechanisms are represented
672	in energy-flow food webs, where relatively thicker lines represent stronger fluxes. Omnivory
673	mechanisms can be exerted by top predators (iii.a) or by secondary consumers (iii.b).
674	Figure 2. Study site map showing the drainage area, the main hydraulic infrastructures that
675	regulate streamflow (Margalef, Siurana and Vilella Baixa reservoirs, Riudecanyes weir and
676	bypass), and the location of the 9 studied reaches. M = Montsant river; S = Siurana river; CON =
677	confluence of Montsant and Siurana rivers. See details in Table 1.

Figure 3. Variation in food-chain length and in trophic diversity (bivariate δ^{13} C - δ^{15} N isotopic space) across reaches. Trophic diversity was estimated using Bayesian Standard Ellipse Areas (SEA), with the 'siber.ellipses' function of the 'SIAR' R-package (Parnell *et al.* 2010). 10,000 posterior draws were made in each case, and the corresponding credible intervals are shown for each local food web. Only predators were included (*N* = 462).

Figure 4. Relationships between the hypothesized environmental controls and FCL (DIST =disturbance, MET = ecosystem metabolism, RES = basal resource availability, SIZE = habitat

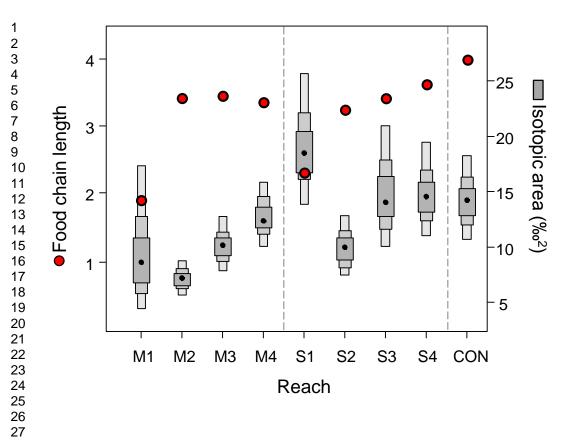
size; see text for details). \mathbf{r}^2 and *P*-values of the linear regressions between each control (explanatory variables) and FCL (response variable) are shown. DIST was the only significant regression (P < 0.05) and it is indicated by a solid line. Figure 5. Effects of the main environmental control on *omnivory*. The dietary proportions of (a) the secondary consumers' assemblage, (b) the top predators' assemblage, and (c) *Barbus*, are represented across the hydrological disturbance gradient (DIST). Each color represents a different trophic compartment being consumed: orange triangles = POM; green diamonds = biofilm + green algae; blue diamonds = macrophytes; grev circles = primary consumers; black circles = secondary consumers (only considered in **b** and **c**). \mathbf{r}^2 and *P*-values of the linear regressions are shown, and significant regressions (P < 0.05) are indicated by solid lines. Figure 6. (a) Relationship between *Barbus* sp. individual fork length and its individual trophic position. The r^2 and *P*-value of the linear regression between these variables are shown. (b) Size distribution of this taxon across the studied reaches. Sizes differed significantly overall (GLM, $F_{7,304} = 54.202, P < 0.001$). Letters show which reaches share a similar size distribution (as identified by Scheffé post-hoc tests).



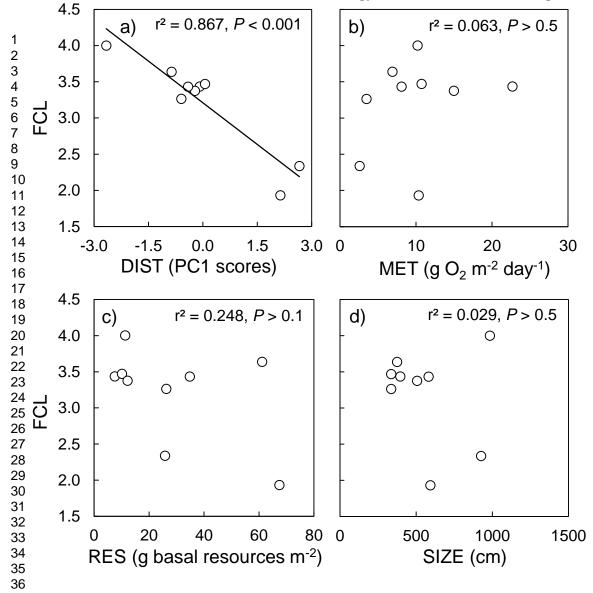


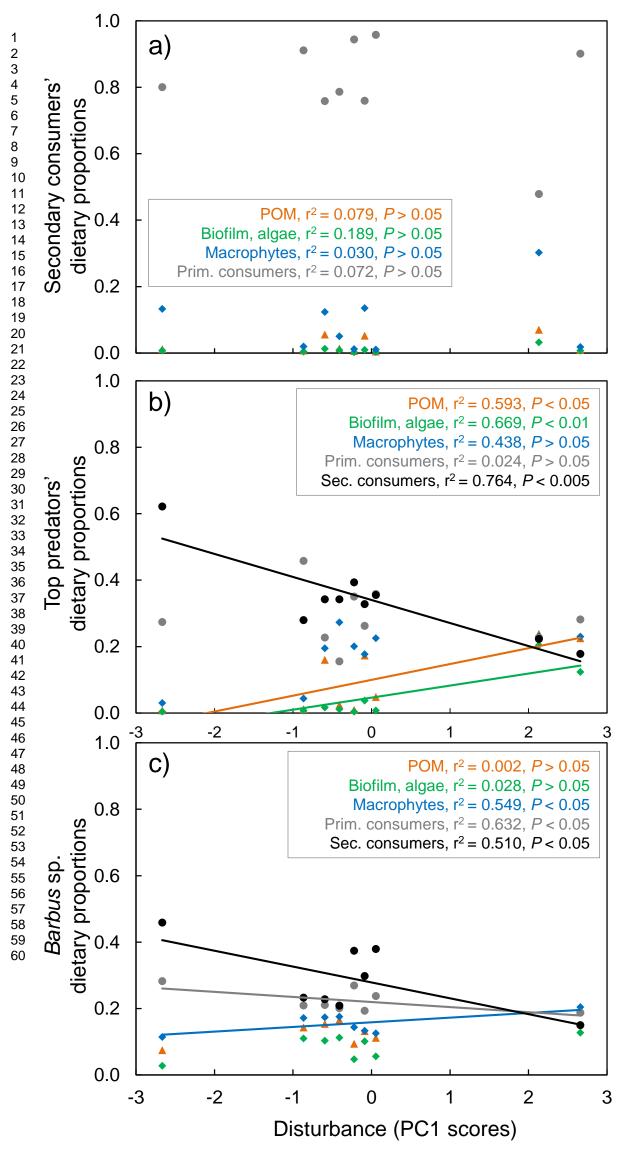
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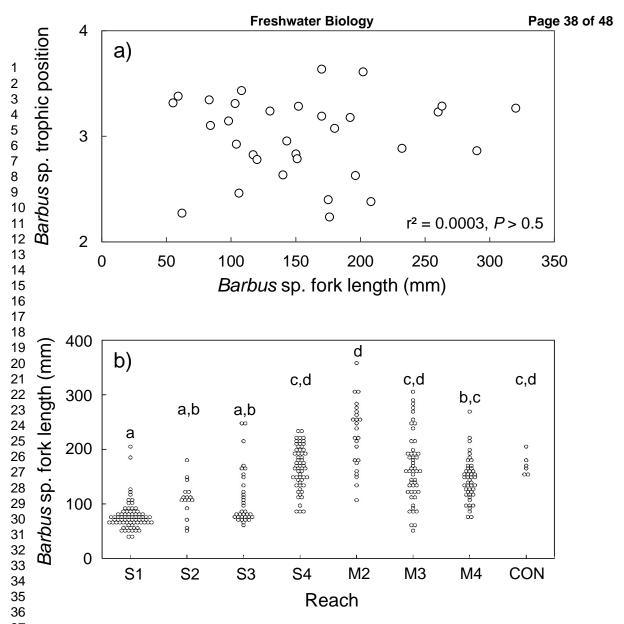
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Supporting Information

Flow regulation increases food chain length through omnivory mechanisms in a Mediterranean river network

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Table S1. *Summary of the physico-chemical variables considered.* Water temperature (Temp), dissolved oxygen (O₂) and conductivity (Cond) were determined in situ using field meters (Hach, Loveland, USA). Water samples for nutrients were collected in triplicate from each reach, at approximately 10 cm below the water surface. These samples were filtered in situ through combusted and pre-weighed glass fiber filters (Whatman, UK) using portable filtration devices connected to a vacuum pressure pump (Millipore, USA). Samples were placed in rinsed polyethylene bottles and frozen (-20°C) in the field until analysis. Total dissolved phosphorus (TDP) was measured as the concentration of phosphate (PO₄³⁻) after acid digestion in a Selecta Presoclave-II 30L autoclave (JP Selecta, Spain). The concentrations of ammonium (NH₄⁺), nitrate (NO₃⁻) and nitrite (NO₂⁻) were determined colorimetrically using an Alliance-AMS Smartchem 140 (AMS, France). The concentrations of total dissolved nitrogen (TDN) and dissolved organic carbon (DOC) were measured on a Shimadzu TOC-V CSH coupled to a TNM-1 module (Shimadzu Corporation, Japan). Values are 3-replicate averages. M = Montsant river; S = Siurana river; CON = confluence of Montsant and Siurana rivers.

Reach	M1	M2	M3	M4	S1	S2	S3	S4	CON
Ktati		1,12	1010	171 1			50	51	con
Temp (°C)	11.9	13.7	13.4	13.1	19.3	11.2	15.1	20.7	19.5
$O_2 (mg L^{-1})$	9.6	12.1	10.6	7.8	9.1	11.4	9.0	6.2	11.0
Cond (µS cm ⁻¹)	365.7	412.0	432.0	485.0	691.7	581.0	924.0	711.0	547.0
$PO_4^{3+} (mg L^{-1})$	0.013	0.008	0.008	0.005	0.008	0.008	0.026	0.007	0.006
TDP (mg L ⁻¹)	0.030	0.013	0.010	0.016	0.012	0.013	0.043	0.017	0.022
$NH_4^+ (mg L^{-1})$	0.005	0.001	0.001	0.001	0.003	0.015	0.009	0.003	0.001
$NO_3^- (mg L^{-1})$	0.002	0.533	0.377	0.028	0.054	0.069	0.440	0.002	0.015
$NO_2^- (mg L^{-1})$	0.003	0.006	0.006	0.005	0.003	0.008	0.006	0.003	0.005
TDN (mg L ⁻¹)	0.158	0.686	0.442	0.177	0.221	0.300	0.455	0.170	0.165
DOC (mg L ⁻¹)	1.747	3.030	2.364	1.687	1.256	2.545	1.625	1.687	1.518

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 Table S2. Permutation-based models combining data on physico-chemical variables and biota.
 A)
 Results of the permutational MANOVA (PERMANOVA) employed to explore physico-chemical variation among upstream, downstream, and confluence sections, and within the downstream section. In order to account for the spatially-autocorrelated structure of the study, *reach* factor (random, 9 levels: each reach) was nested within section factor (fixed, 5 levels: M1, M2-M3-M4, S1, S2-S3-S4, CON). Physico-chemical conditions did not significantly differ between sections, despite variation across the reaches located downstream of the main reservoirs. B) Results of the RELATE and BIO-ENV routines combining the environmental matrix with the algae, invertebrate and vertebrate community matrices. The RELATE procedure (*Rho* and *Sign* columns) is a Mantel-type test that contrasts the null hypothesis of no significant link between two similarity matrices by permuting the sample labels from one of the similarity matrices and recalculating the matches (the rank correlation, *Rho*) many times (999 permutations; Sign = significance level of sample statistic expressed as a percentage) (Clarke & Gorley 2006). Subsequently, the BIO-ENV routine (column Variables selected) allowed identifying the subset of variables that maximize the rank correlation (Clarke & Gorley 2006). Note that physico-chemical variation did not significantly influence algae, invertebrate, or vertebrate community composition (Sign > 5% in all 3 cases). Furthermore, the variables maximizing the rank correlation presented very limited predictive power (Rho < 0.35 in all 3 cases). All data were $\log_{10}(X+1)$ transformed. Euclidean distances were used for the physicochemical matrix, and Bray-Curtis coefficient for the algae, invertebrate and vertebrate matrices.

A)	df	SS	MS	Pseudo-F	P(perm)
Section	4	3.341	0.835	1.229	0.333
Reach(Section)	4	2.718	0.680	83.57	0.001
B)	Rho	Sign	Variables selected		
Algae community	0.317	66%	NO ₂		
Invertebrate community	0.347	45%	Cond		
Vertebrate community	0.315	47%	TDN, Temp, Cond		

Table S3. Details of the Principal Component Analysis (PCA) combining hydrological and

geomorphological data. PC1 was selected as our multivariate index of stability (DIST).

Principal Component Analysis (PCA):

PC	Eigenvalues	%Variation	Cum.%
1	2.51	50.2	50.2
2	1.94	38.7	88.9
3	0.32	6.4	95.3
4	0.187	3.7	99.1
5	0.0474	0.9	100.0

Variable	PC1	PC2	PC3	PC4	PC5
Area	-0.497	0.410	-0.262	-0.228	0.681
Regulation	-0.590	0.192	-0.277	-0.176	-0.712
Armoring	0.058	-0.648	-0.739	-0.143	0.100
Droughts	0.522	0.321	-0.107	-0.775	-0.112
Floods	0.359	0.521	-0.545	0.544	-0.079

Eigenvectors (Coefficients in the linear combinations of variables making up PC's):

combination.

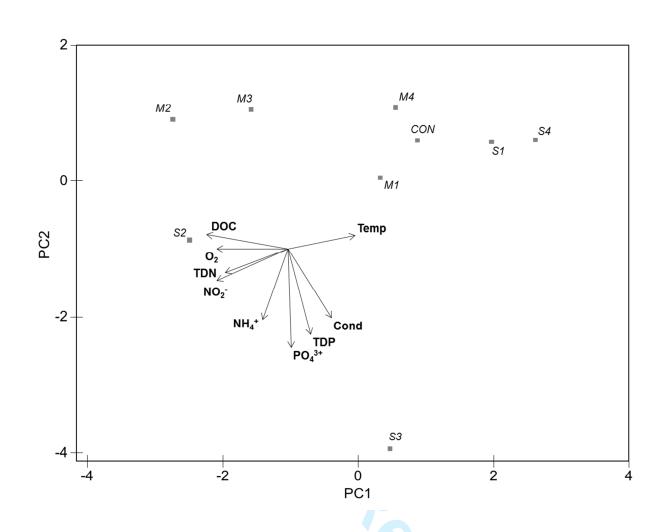


Figure S1. *Principal Components Analysis (PCA) ordering the 9 studied reaches according to their physico-chemical characteristics.* See table S1 for data and units, methods outline, and variable codes. Data was $log_{10}(X+1)$ transformed and normalized, and the variable NO_3^- was excluded from this analysis due to high correlation ($\rho > 0.95$) with TDN. PC1 axis explains 39.3 % of the variation, PC2 explains 28.3%. M = Montsant river; S = Siurana river; CON = confluence of Montsant and Siurana rivers.

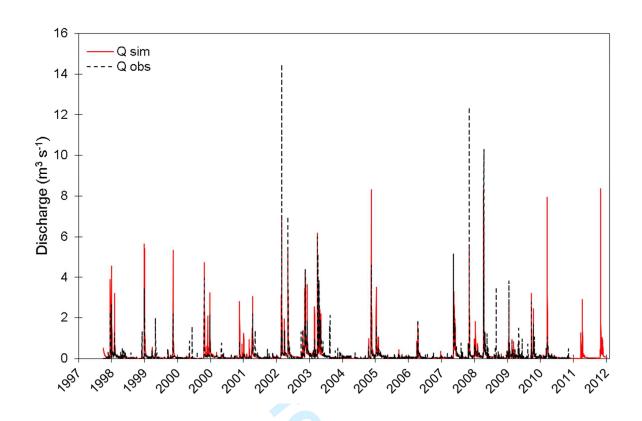


Figure S2. *Observed (Q obs) versus simulated flow (Q sim) at the Siurana reservoir.* The TETIS model includes an automatic calibration tool, based on the SCE-UA algorithm (Duan, Sorooshian & Gupta 1992). The model is calibrated by adjusting up to 9 coefficients (i.e., correction factors). Each correction factor multiplies a parameter map, allowing calibrating the model without losing the spatial structure of the parameter, and reducing drastically the number of parameters to be calibrated. We included an additive stochastic error term (Vrugt *et al.* 2002) to account for measurement errors. TETIS is a dynamic conceptual model well-suited to describe the hydrological cycle in Mediterranean watersheds (Medici *et al.* 2008). For more information on this model, see Francés *et al.* (2007), Vélez *et al.* (2009), and Bussi *et al.* (2013).

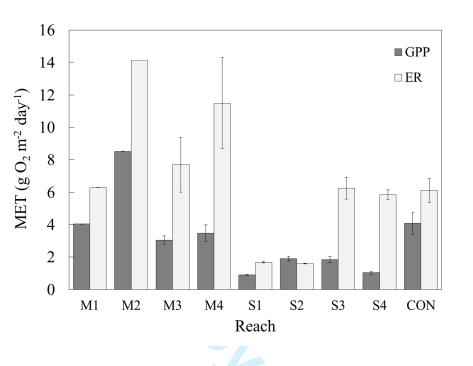
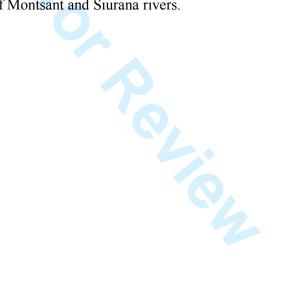


Figure S3. *Ecosystem metabolism (MET) across the studied reaches*. MET was calculated as the sum of gross primary production (GPP, in dark gray) and ecosystem respiration (ER, in white). Error bars represent standard deviations. Ecosystem metabolism was estimated in the 9 study reaches by means of the two-station technique (Odum 1956; Marzolf, Mulholland & Steinman 1994; Reichert, Uehlinger & Acuña 2009). At all stations, dissolved oxygen and temperature were recorded in 10-min intervals during 4 days with optical dissolved oxygen probes YSI 6150 connected to YSI 600 OMS V2 multiparameter sonde (YSI Inc., Yellow Springs, Ohio, USA). The probes were deployed in the thalweg of the stream, about 5 cm below the water surface. Before deployment, the dissolved oxygen sensors were calibrated according to the manufacturer's manual. After the field measurements, sonde-to-sonde variability was determined by simultaneously immersing the probes in a thermo-regulated and aerated water bath (\pm 0.1 °C). The temperature of the water bath was successively adjusted to 20, 18, 16, 14, 12, 10, 8 and 6 °C and dissolved oxygen recorded every 30 s. Saturation concentration of dissolved oxygen was calculated using recorded temperatures and barometric pressure from a meteorological station of the Catalan Meteorological Service (XEMA). Deviations from the calculated saturation concentrations were determined and used to correct the

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field dissolved oxygen records. Estimates of reaeration coefficients were based on the decline of dissolved O₂ concentration after dusk (Hornberger & Kelly 1975). Daily reaeration coefficients were the means of values from the nights before and after the day of interest. Net metabolism rate (NM, expressed as g $O_2 \text{ m}^{-2} \text{ min}^{-1}$) was calculated from the dissolved O_2 concentration, water temperature, atmospheric pressure, gas-exchange rate, and flow velocity using the open-channel two-station techniques originally developed by (Odum 1956) and further modified by (Reichert *et al.* 2009). Three daily metabolic parameters were based on NM: net ecosystem metabolism (NEM), ecosystem respiration (ER), and gross primary production (GPP). NEM was calculated as the sum of NM over 24 h, ER as the sum of NM during the dark period and respiration rates during the light period (calculated as the linear interpolation between the NMs of sunrise and sunset of the nights before and after the day of interest), and GPP as the difference between NEM and ER. M = Montsant river; S = Siurana river; CON = confluence of Montsant and Siurana rivers.



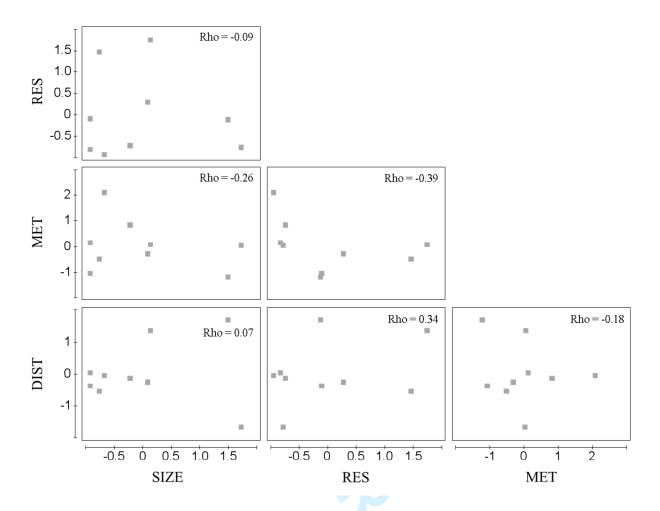


Figure S4. Scatter plots showing the correlations among the hypothesized environmental controls

of FCL (DIST, MET, RES, SIZE). Spearman's ranks correlations (Rho) are shown.

References for Supporting Information

- Bussi G., Rodríguez-Lloveras X., Francés F., Benito G., Sánchez-Moya Y. & Sopeña A. (2013) Sediment yield model implementation based on check dam infill stratigraphy in a semiarid Mediterranean catchment. *Hydrology and Earth System Sciences* **17**, 3339–3354.
- Clarke K.R. & Gorley R.N. (2006) *Primer version 6: user manual/tutorial Primer-E*. Plymouth Marine Laboratory, Plymouth, UK.
- Duan Q., Sorooshian S. & Gupta V. (1992) Effective and efficient global optimization for conceptual rainfall-runoff models. *Water resources research* **28**, 1015–1031.
- Francés F., Velez J.I. & Vélez J.J. (2007) Split-parameter structure for the automatic calibration of distributed hydrological models. *Journal of Hydrology* **332**, 226–240.
- Hornberger G.M. & Kelly M.G. (1975) Atmospheric reaeration in a river using productivity analysis. *Journal of the Environmental Engineering Division* **101**, 729–739.
- Marzolf E.R., Mulholland P.J. & Steinman A.D. (1994) Improvements to the diurnal upstreamdownstream dissolved oxygen change technique for determining whole-stream metabolism in small streams. *Canadian Journal of Fisheries and Aquatic Sciences* **51**, 1591–1599.
- Medici C., Butturini A., Bernal S., Vázquez E., Sabater F., Vélez J.I., *et al.* (2008) Modelling the non-linear hydrological behaviour of a small Mediterranean forested catchment. *Hydrological Processes* **22**, 3814–3828.

Odum H.T. (1956) Primary production in flowing waters. *Limnology and Oceanography* 1, 102–117.

- Reichert P., Uehlinger U. & Acuña V. (2009) Estimating stream metabolism from oxygen concentrations: Effect of spatial heterogeneity. *Journal of Geophysical Research: Biogeosciences (2005–2012)* **114**.
- Vélez J.J., Puricelli M., López Unzu F. & Francés F. (2009) Parameter extrapolation to ungauged basins with a hydrological distributed model in a regional framework. *Hydrology and Earth System Sciences* **13**, 229–246.
- Vrugt J.A., Bouten W., Gupta H. V. & Sorooshian S. (2002) Toward improved identifiability of hydrologic model parameters: The information content of experimental data. *Water resources research* 38, 1–13.