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Additional Information

1 **A new methodology to assess the performance of AOPs in**
2 **complex samples: Application to the degradation of phenolic**
3 **compounds by O₃ and O₃/UVA-vis**

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12

13

14 **Abstract**

15

16 A methodology combining experimental design methodology, liquid chromatography,
17 excitation emission matrixes (EEM) and bioassays has been applied to study the
18 performance of O₃ and O₃/UVA-vis in the treatment of a mixture of eight phenolic
19 pollutants. An experimental design methodology based on Doehlert matrixes was
20 employed to determine the effect of pH (between 3 and 12), ozone dosage (0.2-1.0 g/h)
21 and initial concentration of the pollutants (1-6 mg/L each). The following conclusions
22 were obtained: a) acidic pH and low O₃ dosage resulted in an inefficient process, b)
23 increasing pH and O₃ amount produced an enhancement of the reaction, and c)

24 interaction of basic pH and high amounts of ozone decreased the efficiency of the
25 process. The combination of O₃/UVA-vis was able to enhance ozonation in those
26 experimental regions where this reagent was less efficient, namely low pH and low
27 ozone dosages. The application of EEM-PARAFAC showed four components,
28 corresponding to the parent pollutants and three different groups of reaction product and
29 its evolution with time. Bioassays indicated important detoxification (from 100% to less
30 than 30% after 1 min of treatment with initial pollutant concentration of 6 mg/L, pH = 9
31 and ozone dosage of 0.8 g/h) according to the studied methods (*D. magna* and *P.*
32 *subcapitata*). Also estrogenic activity and dioxin-like behavior were significantly
33 decreased.

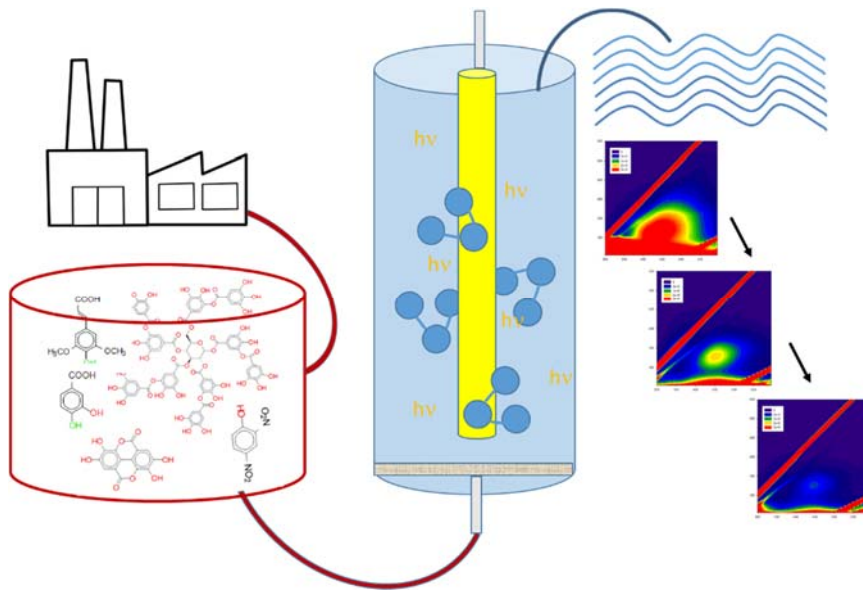
34

35 **Keywords**

36 Ozone, UV-VIS, phenolic compounds, Doehlert matrix, toxicity, excitation emission
37 matrix fluorimetry

38

39 **Graphical abstract**



40

41

42 **Highlights**

43 UV-A-vis light improves the efficiency of the ozonation at acid pH and low O₃
 44 concentration.

45 EEM is a good technique to detect intermediates in the oxidation process.

46 PARAFAC is able to decompose the EEM into more simple components, allowing
 47 monitoring of the generated intermediates.

48 This process resulted in a decrease in estrogenic and dioxine-like properties and
 49 detoxification (standard bioassays).

50

51 **1. Introduction**

52 Food processing industry (e.g. wine, olive mill or cork boiling) produces highly polluted
 53 effluents, containing sugars, tannins and (poly)phenolic compounds, among other
 54 substances at different concentrations. For instance, high concentrations of polyphenols

55 (ca. 1300 mg/L) can be found in the wastewater originated in the baking process of cork
56 (Mendoza et al., 2004). Since some of those compounds are refractory to classical
57 treatments, development of alternative techniques to deal with these effluents seems
58 necessary. In this context, advanced oxidation processes (AOPs) have received great
59 attention in the last two decades, and promising results have been obtained in most
60 cases (Vilar et al., 2009).

61

62 One of the AOPs that have been considered is ozonation. This process does not only
63 involve direct reaction of molecular ozone with the pollutant, but also an indirect
64 mechanism involving hydroxyl radicals (Dodd et al, 2009; Beltran, 2003). The
65 mechanism is pH dependent: while in acidic medium attack of molecular ozone is
66 predominating, at basic pH decomposition of O₃ into highly reactive ·OH is favoured
67 (Ikehata and El Din, 2004, Ikehata, 2006). These differences in the prevailing
68 mechanism explains the higher efficiency of ozonation at high pH values (Luis et al,
69 2011): reaction with molecular ozone is selective towards some moieties such as double
70 bonds or aromatic rings, while hydroxyl radicals are unselective and, in general, more
71 reactive (Legrini et al, 1993). This pH dependence is even more evident in the case of
72 mineralization of organic matter, that is a goal more difficult to be achieved due to the
73 formation of by-products that, in turn, should be removed (Ribeiro et al, 2015); in fact,
74 high pH is needed to reach significant mineralization of effluents (Beltran, 2003).
75 Alternative ways to generate hydroxyl radicals from ozone is the combination of this
76 gas with other substances such as TiO₂ (Rivas et al. 2012, Tichonovas et al., 2017),
77 Fe(III) (Quiñones, 2015), zero-valent zinc (Zhang et al. 2015), hydrogen peroxide
78 (Mizuno et al., 2011).

79

80 Also irradiation can be employed to generate $\cdot\text{OH}$ from ozone (Kuo, 1999). In fact,
81 decomposition of O_3 in water under UVC irradiation has been widely studied. The
82 mechanism of the process is complex and depends on different parameters, such as the
83 pH (Chávez, 2016), or the presence of radical scavengers in the reaction medium
84 (Garoma, 2005). In general, UV irradiation and ozonation show an important synergetic
85 effect, both in pollutants removal and in mineralization of the organics present in the
86 effluent (Orge et al., 2017, Lovato et al., 2017), what is useful in the treatment of
87 effluents having high concentration of organic matter (Collivignarelli et al., 2004). The
88 higher efficiency of O_3 /irradiation has been reported to reduce the amount of ozone that
89 is required and hence, the energy consumption (Quiñones et al., 2015).

90

91 The UVA fraction can also be employed for this purpose, although the efficiency is
92 lower than UVC, as the absorption of ozone in this interval is very low (see Oh et al.,
93 2016 for the spectrum of ozone in aqueous solution) . However, this approach is
94 interesting as sunlight can be employed as irradiation source (Marquez et al, 2014;
95 Quiñones et al., 2015) and it has been reported to enhance the process in the conditions
96 more disfavoured for ozonation, namely acidic pH (Chavez et al., 2016). In fact,
97 recent studies have shown that irradiation results in an enhancement of the process
98 efficiency, although the intensity of light has scarce influence on the oxidation rate
99 (Siqueira and Amaral, 2015).

100

101 With this background, this work has two major goals that are: a) to study the use of O_3
102 and its combination with simulated sunlight in the removal of phenolic components
103 commonly found in food processing effluents and b) to develop a reliable methodology

104 to monitor the performance of AOPs in complex samples. For this purpose, a mixture of
105 six phenolic compounds (2,4-dinitrophenol, gallic acid, protocatechuic acid, vanillic
106 acid, syringic acid and sinapic acid) and two polyphenols (tannic acid and elagic acid,
107 were chosen; this group of pollutants has already be employed to check the effect of
108 photo-Fenton in these type of effluents (García Ballesteros et al., 2016). Ozonation was
109 performed with and without irradiation with a xenon lamp under different conditions
110 and experimental design methodology based on Doehlert matrices was used to
111 determine the effect of experimental conditions (Ferreira et al., 2004); this methodology
112 has been commonly employed for this purpose in wastewater treatment processes
113 (Gomis et al., 2015). In order to follow the effect of ozonation on the mixture of
114 phenolic compounds, excitation emission matrices (EEM) have been employed. This
115 technique is based on the measurement of bi-dimensional plots in which the
116 fluorescence emission spectra of the sample is plotted versus the wavelength of the
117 excitation source (Chen et al., 2003). It provides interesting structural information on
118 the organic matter present in solution and it increasingly used to analyse the behaviour
119 of complex samples as it provides alternative information of fluorescent by-products to
120 chromatography if the aim of the work is not to establish a detail degradation path has to
121 be established. (Liu et al, 2015 and 2016).

122 In this work, parallel factor analysis (PARAFAC) has been employed to gain further
123 insight into the information provided by EEM. This mathematical procedure allows
124 decomposing the EEM into different components, which can be assigned to
125 chromophores or groups of substances; hence, major trends in decomposition of
126 pollutants in complex mixtures can be recognized (García-Ballesteros et al., 2016;
127 Mangalgi et al., 2017). Finally, a battery of bioassays have been employed to monitor
128 detoxification of the solution along the treatment.

129

130 **2. Experimental**

131 2.1 Reagents, microorganisms and cell lines

132 2,4-dinitrophenol, tannic acid, elagic acid, gallic acid, protocatechuic acid, vanillic acid,
133 syringic acid and sinapic acid were purchased from Sigma-Aldrich and used as
134 received. Ultra-pure water (Mili-Q) was used to prepare all solutions.

135 *P. subcapitata* and *ephippia* (dormant eggs) of *D. magna*, employed for the bioassays
136 were supplied by ECOtest S.L. (Valencia, Spain). Two recombinant strains of
137 *Saccharomyces cerevisiae* were employed: BY4741 strain to detect estrogenic activity
138 (ER strain) and YCM4 strain to monitor aryl hydrocarbon receptor agonist activity
139 (AhR strain); they were kindly provided by Dr. Benjamin Piña from IDAEA-CSIC,
140 Barcelona, Spain.

141 Medium constituents employed in bioassays were supplied by Difco (Basel,
142 Switzerland). Prototrophic markers, standards (17- β -estradiol (E2) and β -
143 naphthoflavone (β -NF)) and enzymatic substrate (4-methylumbelliferone β -D-
144 galactopyranoside and β -mercaptoethanol) were purchased form Sigma-Aldrich. Assay
145 buffer (Triton X-100) and Y-PER (Yeast Protein Extraction Reagent) were supplied by
146 Fisher Chemical and Thermo Scientific, respectively.

147

148 2.2. Reaction conditions

149 Ozonation tests were performed in a cylindrical glass reactor with 1 L capacity which
150 has been described elsewhere (Amat et al., 2003). Ozone was supplied by a generator
151 (Ozogas, T.R.C.E. 4000), able to produce 8 g/h when fed with oxygen. The ozone

152 production was tuned to the desired experimental conditions, with a production between
153 0.2 and 1 g/h. UVA-vis irradiation tests were performed by means of a xenon lamp
154 (Heraeus TXE 150), which solar spectrum closely matches the solar one (see
155 <http://peschl-ultraviolet.com> for spectrum), which was placed axially inside the
156 cylindrical reactor. The lamp, which power was 220 W, was inside a Pyrex glass
157 envelope that cuts off radiation below 300 nm. The system was refrigerated by water. In
158 all cases, the reactor was loaded with 250 mL and the gas flow was 10 NL/min.

159

160 2.3. Chemical analysis

161 The concentration of the six pollutants with low molecular weight (2,4-dinitrophenol,
162 gallic acid, protocatechuic acid, vanillic acid, syringic acid and sinapic acid) was
163 determined by liquid chromatography (Perkin Elmer model Flexiar UPLC FX-10). A
164 Bronwnlee Analytical column (DB-C18) was employed as stationary phase. The eluent
165 consisted in a linear gradient of acetonitrile (A) and an aqueous solution of formic acid
166 (0.1% w/w) (B); the composition of the eluent changed from 5% of A to 50% of A in
167 8.5 min and the flow rate was 0.3 mL min⁻¹. Detection wavelength was 230 nm. The
168 identification and quantification of the pollutants were carried out by comparison with
169 standards. Samples were filtered through polypropylene filters (VWR, 0.45 µm) before
170 analysis.

171 The fluorescence EEM of the samples were recorded using a modular fluorimeter
172 QuantaMaster (PTI) by a subsequent emission scanning from 300 to 600 nm at 5 nm
173 increments by varying the excitation wavelength from 250 to 550 nm at 5 nm
174 increments.

175

176 2.4. Statistic analysis

177 The effect of three operational parameters (pH, ozone doses and pollutants
178 concentration) on the ozonation and combination ozone/UVA-Vis process was studied
179 using an experimental design methodology. As phenolic concentration varies depending
180 the effluent, the range 8-48 mg/L was chosen to be high enough to obtain reliable
181 kinetics by HPLC analysis and not as high to saturate the response of EEM or toxicity
182 experiment concentrations; ozone dosage was also tuned in a low domain to avoid too
183 fast reactions and to allow and accurate sampling . For this purpose, a Doehlert matrix
184 was used in order to select the experimental points required to obtain a three-
185 dimensional response surface; this methodology was chosen because it minimizes the
186 number of experiments required to obtain the n-dimensional surface response (Ferreira
187 et al., 2004). To this end, a total of 15 experiment were chosen: 13 combinations of the
188 three variables studied and two replicates of the central point (see Table 1 for detailed
189 experimental conditions). The software Statgraphics Centurion XVI was used for
190 response surface model fitting by means of the least squares method. The treatment time
191 required to decrease in an 80% the sum of the concentration of the 6 pollutants that
192 could be analysed via liquid chromatography ($t_{80\%}$), was used as response. This type of
193 parameters have been used in previous papers for statistical analysis as it provides a
194 more “visual” information of the time required to achieve the desired result when
195 compared with with pseudo-first order kinetic constants (Gomis et al., 2015, García-
196 Ballesteros et al., 2017; Caram et al., 2018). In particular $t_{80\%}$ was chosen in order to
197 ensure that a significant degradation of the pollutants occurred (80%) and to avoid the
198 asymptotic behaviour of the concentration vs time plot observed at values close to 100%
199 degradation and that results in important errors in the determination of the required time
200 of treatment.

201

202 2.5 Bioassays

203 Bioassays employed in this work have been described in detail elsewhere (García-
204 Ballesteros et al., 2016). Toxicity tests based on *P. subcapitata* were performed
205 according to ISO 8692:2012. Briefly, microplates were filled with the test dilutions and
206 controls, and inoculated with 10^4 cells/mL being the total volume of each well 300 μ L.
207 Microplates were incubated at 23 °C under continuous illumination (6000-10000 lux),
208 saturated with 2% CO₂ and stirred at 120 rpm. The algal growth was measured by *in-vivo*
209 chlorophyll fluorescence every 24 h, (until 72 h) using a microplate reader (Tecan Infinite
210 M200).

211

212 Assays based on the inhibition of the mobility of *D. magna* (ISO 6341:2012) used 24-h old
213 daphnids hatched from the ephippia. Five individuals were placed in 15 mL recipients
214 containing 10 mL of test dilution and corresponding controls. The assays were conducted in
215 the dark at a constant temperature of 21 °C.

216

217 For the RYA (Recombinant Yeast Assay), yeast cells were grown overnight in a synthetic
218 medium. Assays were performed in a polypropylene microplate (NUNC™, Roskilde,
219 Denmark). After plates incubation (6 h at 30 °C at 120 rpm in orbital shaker), were added
220 sequentially Y-PER (30 °C, 30 min) and then, enzymatic substrate. After centrifugation,
221 β -galactosidase activity was monitored using a spectrofluorometer (TECAN Infinite
222 M2000). The relative activity of β -galactosidase was calculated from recorded activity of
223 positive and negative control. Results were calculated as E2 equivalents (EEQ)
224 (estrogenic activity) or β -NF (BNFEQ) (dioxin-like activity), defined as the concentration
225 (ng/L) of standard that should be present to account for the observed response.

226
227
228
229

3. Results and discussion

230 3.1. Ozonation of the phenolic compounds

231 The mixture of all eight phenols and polyphenols was treated by means of O₃ under the
232 following experimental conditions (initial concentration of each pollutant = 3.5 mg/L;
233 O₃ dose = 0.2 g/h and pH = 7.5). Only six of them could be analysed by liquid
234 chromatography (tannic and ellagic acids were not detected, most probably due to their
235 high molecular weight). Plots of the relative concentration of all six pollutants (C/Co,
236 where C is the concentration at the sampling time and Co the initial concentration of
237 each pollutant) vs time were obtained (Figure 1a). In general, very fast elimination of
238 the studied pollutants was observed (complete removal in a few minutes in all cases),
239 being the 2,4-dinitrophenol the more reluctant to the treatment (ca. 20 min of
240 ozonation).

241

242 In order to gain further insight into the effect of ozone on the mixture of phenols and
243 polyphenols, a set of experiments was carried out in order to determine the effect of pH,
244 ozone dose and initial pollutants concentration. Based on these data, the t_{80%} was
245 calculated in each experiment by interpolation from the time profile of the relative
246 concentration of the six detected pollutants (Table 1). A three-dimensional full quadratic
247 response surface model was obtained (Eq 1), where O₃-D is the ozone dose given in g/h
248 and [P] is the pollutants concentration (mg/L); t_{80%} is expressed in minutes:

249

$$250 \quad t_{80\%} = 12.36 - 13.31 \cdot O_3\text{-D} - 1.86 \cdot \text{pH} + 0.83 \cdot [P] + 4.32 \cdot O_3\text{-D}^2 + 1.05 \cdot O_3\text{-D} \cdot \text{pH} - 0.55 \cdot$$
$$251 \quad O_3\text{-D} \cdot [P] + 0.07 \cdot \text{pH}^2 - 0.03 \cdot \text{pH} \cdot [P] - 0.015 \cdot [P]^2 \quad (\text{Equation 1})$$

252

253 The value of the determination coefficient was high ($R^2 = 0.92$), indicating good
254 correspondences between experimental data and calculated values of $t_{80\%}$. Pareto charts,
255 (Figure 2a), indicate that, only pH, its quadratic term (pH^2), interaction of pH with O_3
256 and O_3 were significant; Taking this into account, equations can be reformulated as
257 follows:

258 $t_{80\%} = 12.36 - 13.31 \cdot O_3-D - 1.86 \cdot pH + 1.05 \cdot O_3-D \cdot pH + 0.07 \cdot pH^2$ (Equation 2)

259

260 In order to better analyse the results, two-dimensional contour plots can be obtained
261 from the three-dimensional curve given by the model by fixing one of the variables at
262 the desired value (Figure 3). First, pH was fixed at an acidic ($pH = 5$) and a basic value
263 ($pH = 10$). At low pH, time required to remove the pollutants decreases with increasing
264 ozone dosages; on the other hand at basic pH the reaction is always faster and the effect
265 of ozone dosage is not so evident. This result might be due to the different mechanisms
266 operating at both media: while the attack of molecular ozone is predominating at low
267 pH, decomposition of this reagent into more reactive hydroxyl radicals occurs at $pH =$
268 10; furthermore, ozone decomposition at basic pH also favours its transfer into the
269 liquid phase increasing the utilization rate of ozone. For this reasons, when operating
270 the most efficient mechanism, less ozone is needed. When looking at the surface
271 obtained at $pH = 10$, two regions can be identified where reaction rate is lower: a) at
272 low ozone amount and high concentration of pollutants, most probably due to the lack
273 of oxidizing agent, and b) at high ozone amounts, when recombination of radicals might
274 decrease the efficiency of ozonation (Chavez, 2016), or because hydroxyl radical

275 participates in the chain decomposition cycle of ozone when this reagent is found at
276 high concentrations.

277

278 Another plot was obtained by fixing the concentration of pollutants at the central point
279 (3.5 mg/L) in order to check the effect of pH and ozone dosage (Figure 4). Similar
280 trends were also observed: a) best results were observed at basic pH because of the
281 prevalence of radical-mediated mechanism; b) an strong interaction between ozone
282 dosage and pH was observed (increasing pH required lower ozone dosages to reach the
283 same % of degradation) and c) a slight decrease in efficiency was observed at highest
284 ozone amounts at basic media.

285

286 Finally, ozone dosage was fixed at a low (0.3 g/h) and a high value (0.8 g/h). The effect
287 of pH was predominating, being the reaction faster at basic media. However, some
288 differences could be observed: a) at low ozone dosage, a pH increase resulted in an
289 acceleration of the process in all the studied domain, and pollutant concentration had a
290 slight influence on the process (higher pH was required to reach the same reaction rate
291 at increasing pollutant concentration) and b) on the contrary, at high ozone dosage, the
292 effect of pollutant concentration is not significant and pH also plays a lower role,
293 decreasing the efficiency at high pHs, again in this case because of the detrimental
294 effect of hydroxyl radicals, via recombination or alteration of the ozone cycle.

295

296 As a consequence, of these data, the following conclusions can be obtained: a) at acidic
297 pH and low ozone dosage, the reaction is inefficient, b) increasing both parameters,
298 represents an enhancement in the process, and c) interaction of basic pH and high

299 amounts of ozone decreases the efficiency of the process. Finally, it is worthy to
300 indicate that being the amount of ozone between 0.2 and 1 g/h and the $t_{80\%}$ between 1
301 and 2 min in most cases, the amount of ozone supplied to the reactor can be estimated in
302 3-30 mg; in the central point, 10 mg where needed, that accounted for ca. 40 mg/L.
303 However, this value should not be overemphasised as may strongly depend in the
304 characteristics of the reactor.

305

306 3.2. Combination of ozonation and UV-vis irradiation

307

308 The combination of O_3 and UV-VIS was also studied; although the overlap between the
309 light emitted by the xenon lamp (and that was not cut off by the Pyrex filter, namely $\lambda >$
310 300 nm) and the absorption of ozone in water is very low, in some cases slight increases
311 in the efficiency of the process might be of interest (always excluding highly coloured
312 effluents). Time resolved profiles of C/C_0 were obtained (see Figure 1b for an
313 example). Again in this case, Doehlert matrixes were used, based in the same
314 experimental points used in the O_3 experiment, being $t_{80\%}$ the response parameter (see
315 Table 1): An equation was obtained (eq. 3) with a regression coefficient of 0.99.

316

$$317 \quad t_{80\%} = 3.22 - 0.78 \cdot O_3 \cdot D - 0.46 \cdot pH + 0.38 \cdot [P] - 0.96 \cdot O_3 \cdot D^2 - 0.06 \cdot O_3 \cdot D \cdot pH - 0.03 \cdot O_3 \cdot$$
$$318 \quad D \cdot [P] + 0.03 \cdot pH^2 - 0.06 \cdot pH \cdot [P] - 0.0 \cdot [P]^2 \quad (\text{eq. 3})$$

319

320 Pareto charts showed that the same parameters as in O₃ were found, except that for
321 O₃/UVA-vis also pollutant concentration became significant (Figure 2b). Hence,
322 equation 3 can be reformulated as equation 4

323

$$324 \quad t_{80\%} = 3.22 - 0.78 \cdot O_3\text{-D} - 0.46 \cdot \text{pH} + 0.38 \cdot [\text{P}] - 0.06 \cdot O_3\text{-D} \cdot \text{pH} + 0.03 \cdot \text{pH}^2 \quad (\text{eq. 4})$$

325

326 Figure 4 shows the bi-dimensional contour plots obtained by fixing all three variables in
327 a mid-value. When [P] = 3,5 mg/L, the pH was the predominating factor, although at
328 basic media increasing O₃ dose resulted in a process enhancement. When pH value was
329 fixed, highest efficiency was observed at high O₃ dose and low [P]. Finally, for O₃ dose
330 = 0.6 g/h, highest reaction rates were observed at basic pH and high pollutants
331 concentration.

332

333 However, it seems more interesting to see the synergic effect of O₃ and simulated
334 sunlight. To clarify this point, the subtraction of t_{80%} obtained by O₃ minus t_{80%}
335 obtained with O₃/UVA-Vis was calculated (Figure 5). Negative values indicate that
336 UVA-vis is able to enhance the process. In general this happens at low pH values and
337 low ozone dosages, indicating that this combination is meaningful in those regions
338 where ozonation is inefficient.

339

340 3.3. Fluorescence measurements

341

342 As two of the compounds could not be detected, and analysis and identification of
343 intermediates is difficult in such a complex mixture, using alternative analytical seems
344 interesting. An attractive possibility is employing the fluorescence emission-excitation
345 matrixes. They consist of bi-dimensional plots of the emission of a sample at a given
346 wavelength (λ_{em}) when excited at another wavelength (λ_{exc}). According to the regions of
347 the matrix where signals can be found, some information on the nature of the organics
348 present in the samples can be obtained (García-Ballesteros et al., 2017). Excitation
349 emission matrices (EEMs) were recorded for both processes (O_3 and O_3 /UVA-VIS
350 combination, Figure 6 and 7 respectively) in the central point conditions at different
351 treatments time. The EEM for each individual pollutant can be found elsewhere
352 (García-Ballesteros et al., 2016). A constant decrease in the fluorescence emission can
353 be observed in the area where $\lambda_{ex} < 300$ nm and $\lambda_{em} < 400$ nm. In sharp contrast, there
354 was a significant increase in the signal in the region 300 nm $< \lambda_{ex} < 350$ nm and 420 nm
355 $< \lambda_{em} < 470$ nm, to reach a maximum value in the sample taken after 1 or 2 min of
356 reaction. Then, there was also some decrease in this signal, which became very weak
357 beyond 8 min of irradiation. These trends were clearer in the O_3 /irradiation
358 combination. This behaviour is an indicator of the generation of intermediates along the
359 process, which are responsible for the signal with maximum at ca. $\lambda_{ex} = 325$ nm and λ_{em}
360 $= 450$ nm; they might be associated to quinones/hydroquinones, which are expected to
361 be intermediates in the oxidation of aromatic rings and whose signal is compatible with
362 that of the EEM (Cory et al., 2005).

363

364 In order to gain further insight into the information provided by the EEM, PARAFAC
365 was applied to a set of experiments consisting in all the matrixes obtained in the O_3 and
366 O_3 /UV-vis. This mathematical procedure is able to decompose the EEM into more

367 simple components. Models with 3-8 components were tested and results were more
368 consistent with that considering 4 components (C1-C4); info. The evolution of the score
369 of each component vs. time can be found in supporting Figure 8 a and b for O₃ and
370 O₃/UV-vis combination, showing in both cases similar trends. For the samples recorded
371 at t = 0 min, the highest score was reached by C4 and beyond this point, a continuous
372 decrease of this component can be observed; hence, C4 might be associated with the
373 parent phenolic compounds, that are degraded in the early stages of the process (5-8
374 min). On the other hand, C2 and C3 suffer an initial increase in the score, followed by a
375 decrease, which is compatible with associating these components with intermediates
376 formed in the oxidative process. Finally, C1 follows a continuous increase throughout
377 the process, and might represent more oxidized by-products.

378

379 3.3. Bioassays

380

381 As complete mineralization of the sample could not be achieved and formation of
382 reaction by-products is evident according to the results shown in the previous sections,
383 monitoring changes in the toxicity of the mixture along the treatment seems meaningful.
384 As done with the photo-Fenton process (García-Ballesteros et al., 2016) different
385 bioassays were performed in order to monitor changes in toxicity. The chosen
386 experimental conditions were: a initial concentration of 6 mg/L of each pollutant, pH =
387 9 and ozone dosage of 0.8 g/h (corresponding to point 4 in the experimental plot). They
388 were chosen in order to ensure relatively fast reaction and, at the same time, to have an
389 important concentration of intermediates in the solution. In fact, all the phenolic
390 compounds were removed after 10 min of treatment, but 70 % of the DOC still

391 remained after 30 min of treatment. Results are shown in Table 2. The initial sample
392 was highly toxic, as according to both, *D. magna* and *P. subcapitata* experiments, 100
393 % toxicity was measured; in the initial stages of the process, a fast detoxification was
394 recorded and beyond 5 min of treatment measured values were systematically below
395 30%, what indicated that samples were scarcely toxic. A similar behaviour was
396 observed for the estrogenic and dioxin-like properties, as the initial sample gave values
397 equivalent to more than 11000 ng/L of dioxine and 75 ng/L of E2 (for sample at t = 1
398 min, as the untreated sample was too toxic to measure estrogenic activity). Ozonation
399 resulted in a fast decrease in both parameters and the estrogenic activity was completely
400 removed after 30 min of ozonation and the dioxin-like behaviour decreased three order
401 of magnitude.

402

403 **4. Conclusions**

404 In this paper a methodology combining experimental design methodology, liquid
405 chromatography, excitation emission matrixes (EEM) and bioassays has been tested and
406 a satisfactory assessment of the decontamination process has been achieved, even when
407 pollutants that cannot be analysed by HPLC are present in the initial sample.

408 Complementary information has been obtained according to the different analytical
409 methods: HPLC analyses have been employed to monitor the removal of the parent
410 phenolic pollutants and, based on these data, experimental design methodology is useful
411 to determine favourable experimental conditions for ozonation, as well as to evaluate
412 the convenience of combining O₃ and UVA-vis irradiation.

413

414 The scarce mineralization reached indicate formation of reaction by-products. EEM
415 combined with PARAFAC analysis is a good technique to gain further insight into the
416 nature of these intermediates, as well as for those pollutants that cannot be detected by
417 HPLC analyses. Finally, a battery of bioassays allows having a reliable monitoring of
418 the toxicity of the samples along the chemical oxidative processes.

419

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426 **References**

427 Amat, A.M., Arques, A., Beneyto, H., García, A., Miranda, M.A. Seguí, S., 2003.
428 Ozonisation coupled with biological degradation for treatment of phenolic pollutants: a
429 mechanistically based study. *Chemosphere* 53, 79–86.

430 Beltrán, F.J., 2003. *Ozone Reaction Kinetics for Water and Wastewater Systems* (Boca
431 Raton, FL: Lewis Publishers).

432 Caram, B., García-Ballesteros, S., Santos Juanes, L., Arques, A., García Einschlag, F.S.,
433 2018. Humic-like substances for the treatment of scarcely soluble pollutants by mild
434 photo-Fenton process. *Chemosphere*, 198, 139-146.

435 Chávez, A.M., Rey, A., Beltrán F.J., Álvarez P.M., 2016. Solar photo-ozonation: A
436 novel treatment method for the degradation of water pollutants. *J. of Hazardous*
437 *Materials* 317, 36–43.

438 Chen, W., Westerhoff, P., Leenheer, J.A., Booksh, K. 2003. Fluorescence
439 excitation–emission matrix regional integration to quantify spectra for dissolved organic
440 matter. *Environ. Sci. Technol.* 37, 5701–5710.

441 Collivignarelli C1., Sorlini S., 2004. AOPs with ozone and UV radiation in drinking
442 water: contaminants removal and effects on disinfection byproducts formation. *Water*
443 *Sci Technol.* 49(4), 51-6.

444 Cory, R.M., Mcknight, D.M., 2005. Fluorescence spectroscopy reveals ubiquitous
445 presence of oxidized and reduced quinones in dissolved organic matter. *Environ.Sci.*
446 *Technol.* 39, 8142-8149.

447 Dodd, M. C., Kohler, H. E., Von Gunten, U., 2009. Oxidation of antibiotic compounds
448 by ozone and hydroxyl radical: elimination of biological activity during aqueous
449 ozonation processes. *Environ. Sci. Technol.*, 43, 2498.

450 Ferreira, S.L.C., dos Santos, W. N. L., Quintella, C.M., Neto, B. B., Bosque-Sendra,
451 J.M., 2004. Doehlert matrix: a chemometric tool for analytical chemistry – review.
452 *Talanta* 63, 1061-1067.

453 García-Ballesteros, S., Mora, M., Vicente, R., Sabater, C., Castillo, M.A., Arques A.,
454 Amat, A.M., 2016. Gaining further insight into photo-Fenton treatment of phenolic
455 compounds commonly found in food processing industry. *Chem Eng. J.* 288, 126-136.

456 García Ballesteros, S., Constante, M., Vicente, R., Mora, M., Amat, A.M., Arques, A.,
457 Carlos, L., García Einschlag, F. S., 2017. Humic-like substances from urban waste as

458 auxiliaries for photo-Fenton treatment: a fluorescence EEM-PARAFAC study.
459 Photochem. Photobiol. Sci., 16, 38-45.

460 Garoma, T., Gurol, M.D., 2005. Modeling Aqueous Ozone/UV Process Using Oxalic
461 Acid as Probe Chemical. Environ. Sci. Technol. 39, 7964-7969.

462 Gomis, J., Carlos, L., Bianco Prevot, A., Teixeira, A. C. S. C., Mora, M., Amat, A.M.,
463 Vicente, R., Arques, A., 2015. Bio-based substances from urban waste as auxiliaries for
464 solar photo-Fenton treatment under mild conditions: optimization of operational
465 variables. Catal. Today, 240, 39-45.

466 Ikehata, K. and El Din, M.G. , 2004. Degradation of recalcitrant surfactants in
467 wastewater by ozonation and advanced oxidation process. A review. Ozone Sci. Eng.
468 26, 327-343.

469 Ikehata, K., Naghashkar, N.J. and El Din, M.G. , 2006. Degradation of aqueous
470 pharmaceuticals by ozonation and advanced oxidation process. A review. Ozone Sci.
471 Eng. 28, 3537-414.

472 Kuo, W.S., 1999. Synergistic effects of combination of photolysis and ozonation on
473 destruction of chlorophenols in water. Chemosphere, 39, 1853-1860.

474 Legrini, O., Oliveros, E., and Braun, A.M., 1993. Photochemical Processes for Water-
475 Treatment, Chem. Rev., 93(2):671–698.

476 Liu, C., Tang, X., Kim, J., Korshin, V., 2015. Formation of aldehydes and carboxylic
477 acids in ozonated surface water and wastewater: A clear relationship with fluorescence
478 changes. Chemosphere 125, 182–190.

479 Liu, C., Li, P., Tang, X., Korshin, V., 2016. Ozonation effects on emerging
480 micropollutants and effluent organic matter in wastewater: characterization using

481 changes of three-dimensional HP-SEC and EEM fluorescence data. *Environ Sci Pollut*
482 *Res.* 23:20567–20579.

483 Lovato, M.E., Fiasconaro, M. L., Martín, C. A., 2017. Degradation and toxicity
484 depletion of RB19 anthraquinone dye in water by ozone-based technologies. *Water*
485 *Science and Technology*, 75(4), 813-822.

486 Luis, P., Saquib, M., Vinckier, C., Van Der Bruggen, B., 2011. Effect of membrane
487 filtration on ozonation efficiency for removal of atrazine from surface water. *Ind.*
488 *Eng. Chem. Res.* 50, 8686-8692.

489 Mangalgi, K.P., Timko, S.A., Gonsior, M. Blaney, L., 2017. PARAFAC Modeling of
490 Irradiation- and Oxidation-Induced Changes in Fluorescent Dissolved Organic Matter
491 Extracted from Poultry Litter. *Environ. Sci. Technol.*, 51 (14), 8036–8047.

492 Márquez, G., Rodriguez, E.M., Beltrán, F.J., Álvarez P.M., 2014. Solar photocatalytic
493 ozonation of a mixture of pharmaceutical compounds in water. *Chemosphere* 113, 71–
494 78.

495 Mendonça, E., Pereira, P., Martins, A., Anselmo, A.M., 2004. Fungal and Detoxification
496 of Cork Boiling Wastewaters. *Engin Life Sci.* 4 (2), 144-149.

497 Mizuno, T., Ohara, S., Nishimura, F., Tsuno, H., 2011. O₃/H₂O₂ process for both
498 removal algal-derived compounds and control of bromate ion formation. *Ozone Sci.*
499 *and Eng.*, 33, 121-135.

500 Oh, J.S., Yajima, H., Hashida, K., Ono, T., Ishijima, T., Serizawa, I., Furuta, H., Hatta,
501 A., 2016. In-situ UV Absorption spectroscopy for observing dissolved ozone in water. *J.*
502 *Photopolymer Sci. Technol.*, 29, 427-432.

503 Orge, C.A., Pereira, M.F.R., Faria J.L., 2017. Photocatalytic-assisted ozone degradation
504 of metolachlor aqueous solution. Chem. Eng. J., 318, 247-253.
505 [http://peschl-
ultraviolet.com/cms/upload/bilder_produktsseiten/1_photochemie/1_praeparative_photoc
hemie/mpds-basic/bilderset/spektren/txe150_en.jpg](http://peschl-
506 ultraviolet.com/cms/upload/bilder_produktsseiten/1_photochemie/1_praeparative_photoc
507 hemie/mpds-basic/bilderset/spektren/txe150_en.jpg)

508 Quiñones, D.H., Álvarez, P.M., Rey, A., Contreras, S., Beltrán. F.J., 2015. Application
509 of solar photocatalytic ozonation for the degradation of emerging contaminants in water
510 in a pilot plant. Chemical Engineering Journal 260, 399–410.

511 Ribeiro, A.R., Nunes, O.C., Pereira, M.F.R., Silva, A.M.T., 2015. An overview on the
512 advanced oxidation process applied for the treatment of water pollutants defined in the
513 recently launched Directive 2013/39/EU. Environmental International, 75, 33-51.

514 Rivas, L., Beltran F.J., Encinas, 2012. A. Removal of emergent contaminants:
515 Integration of ozone and photocatalysis. J. of Env. Management, 100, 10-15.

516 Siqueira F., Amaral, L., 2015. Degradation of caffeine by advanced oxidative processes:
517 O₃ and O₃/UV. Ozone Sci. & Eng., 37, 379-384.

518 Tichonovas, M., Krugly, E., Jankunaite, D., Racys, V., Martuzevicius, D., 2017. Ozone-
519 UV-catalysis based advanced oxidation process for wastewater treatment.
520 Environmental Sci. Pollut. Res., 224, 17584-17597.

521 Vilar, V.J.P., Maldonado, M.I., Oller, I., Malato, S., Boaventura, R.A.R., 2009. Solar
522 treatment of cork boiling and bleaching watewaters in a pilot plant. Water Research. 43
523 (16), 4050-4062.

524 Zhang, J., Wu, Y., Qin, C., Liu, L., Lan, Y., 2015. Rapid degradation of aniline in
525 aqueous solution by ozone in the presence of zero-valent zinc. *Chemosphere*, 141, 258-
526 264.

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529

530 Table 1: Experimental points and response (t_{80}) used in the experimental design
 531 methodology based on Doehlert matrix)

Experiment number	[O ₃] (g/h)	pH	[Pollutant] (mg/L)	$t_{80\% \text{ O}_3}$ (min)	$t_{80\% \text{ O}_3/\text{UVA-VIS}}$ (min)
1	0.6	7.5	3.5	1.54	1.65
2	1	7.5	3.5	1.29	0.91
3	0.8	12	3.5	0.95	1.03
4	0.8	9	6	1	1.06
5	0.2	7.5	3.5	2.34	1.73
6	0.4	3	3.5	6.26	3.10
7	0.4	6	1	1.53	1.72
8	0.8	3	3.5	2.44	2.97
9	0.8	6	1	0.99	1.38
10	0.4	12	3.5	0.97	1.37
11	0.6	10.5	1	0.85	1.57
12	0.4	9	6	1.38	1.53
13	0.6	4.5	6	2.93	3.15
14	0.6	7.5	3.5	0.93	1.42
15	0.6	7.5	3.5	0.9	1.35

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533

534 Table 2: Toxicities measured by a battery of bioassays for samples taken after different
535 ozonation periods (initial concentration of 6 mg/L, pH = 9 and ozone dosage of 0.8 g/h).

536

t (min)	Toxicity (%)		Estrogenic activity	dioxin-like behaviour
	<i>D. magna</i>	<i>P. subcapitata</i>	E2 (ng/L)	(ng/L)
0	100	100	Very toxic	11342
1	5	30	75.7	14464
2	50	20	11.4	22.5
5	30	25	45.3	82.4
10	20	26	7.3	166.2
30	25	3	--	107.5

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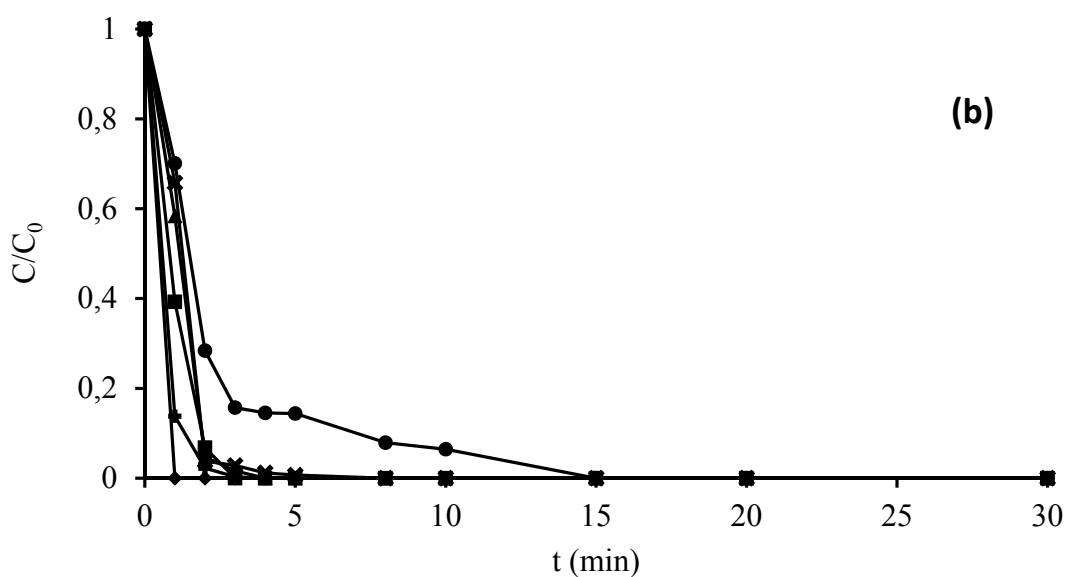
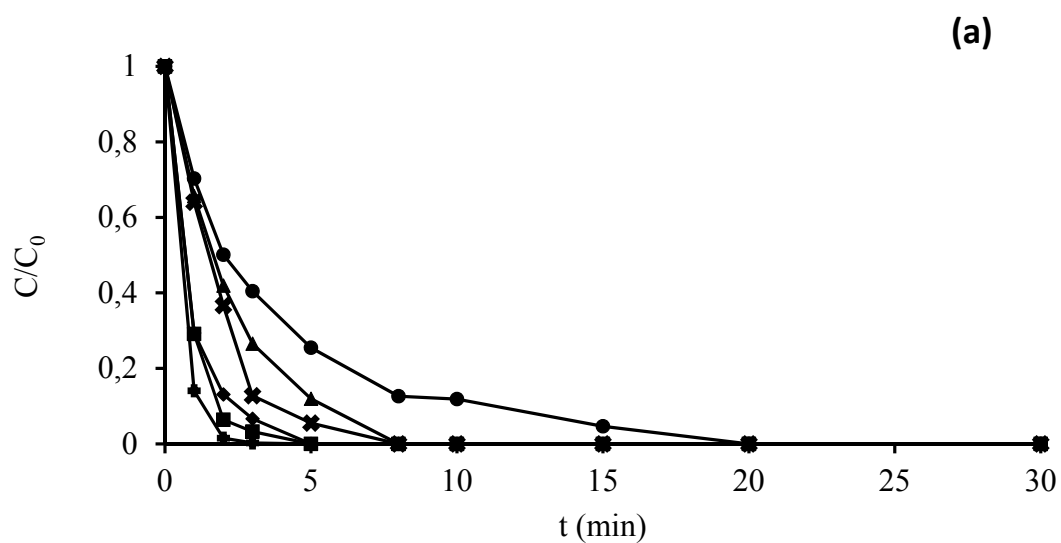
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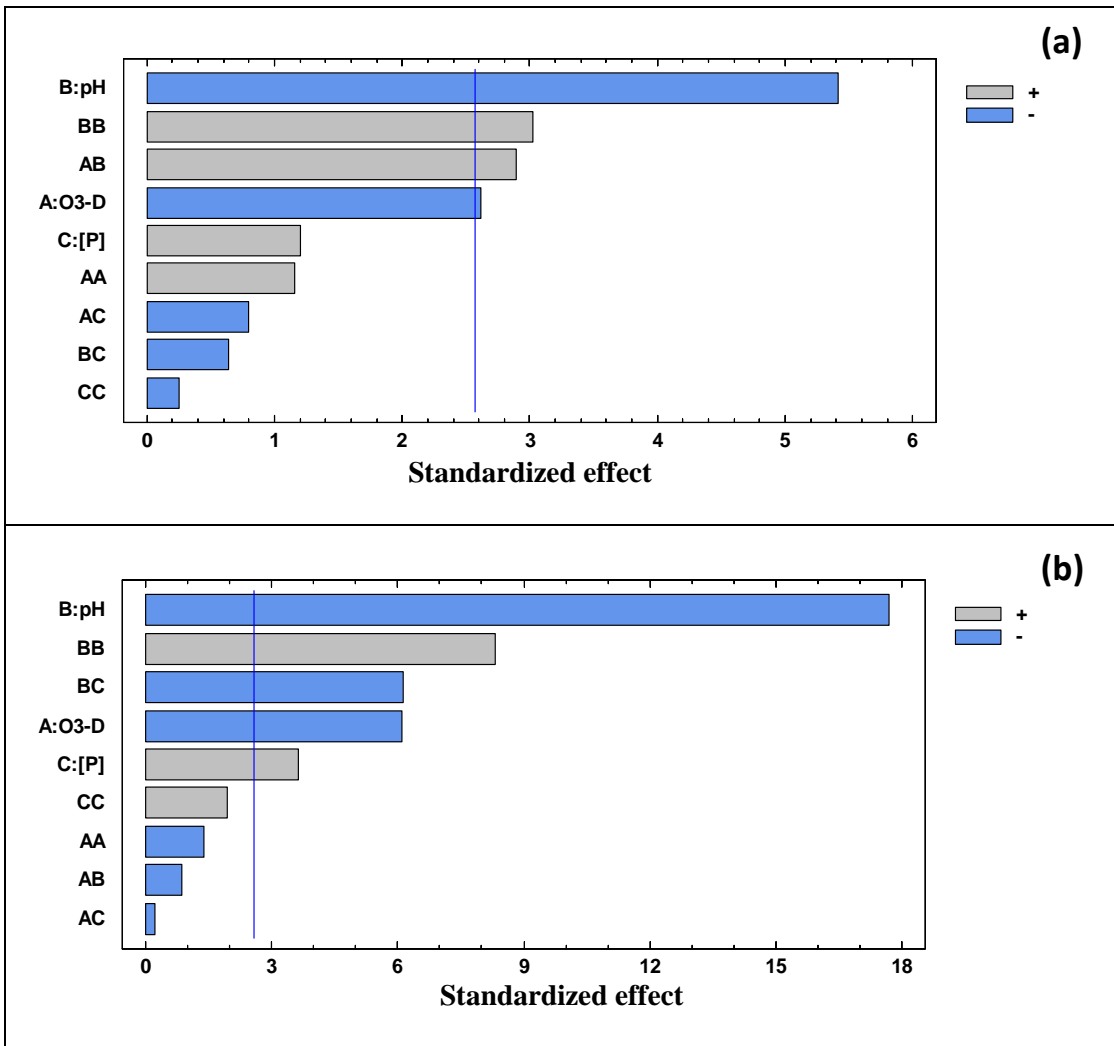
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546 Figure 1: Ozonisation (a) and O₃/UVA-VIS combination (b) for the removal of a
 547 mixture of phenolic compounds: initial concentration of each pollutant = 3.5 mg/L; O₃
 548 dose = 0.2 g/h and pH = 7.5. A plot of the relative concentration of each pollutant vs.
 549 time is given: gallic acid (◆); protocatechuic acid (■); vanillic acid (▲); sinapic acid
 550 (+); syringic acid (×) and 2,4-dinitrophenol (●).

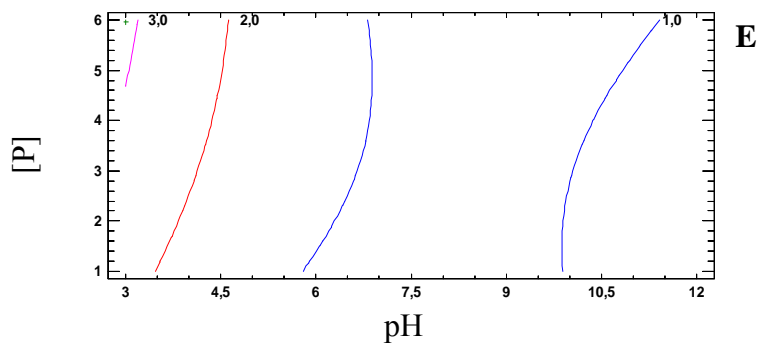
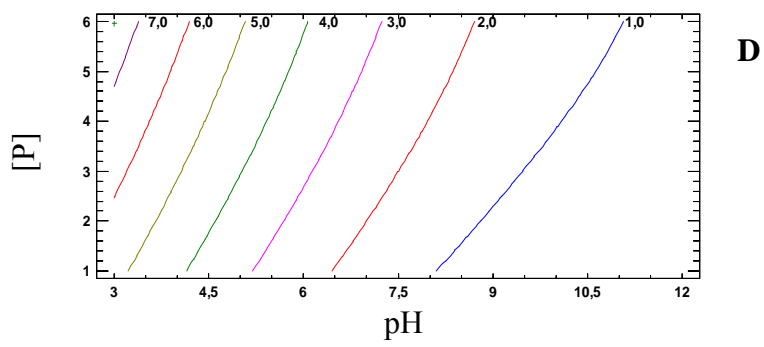
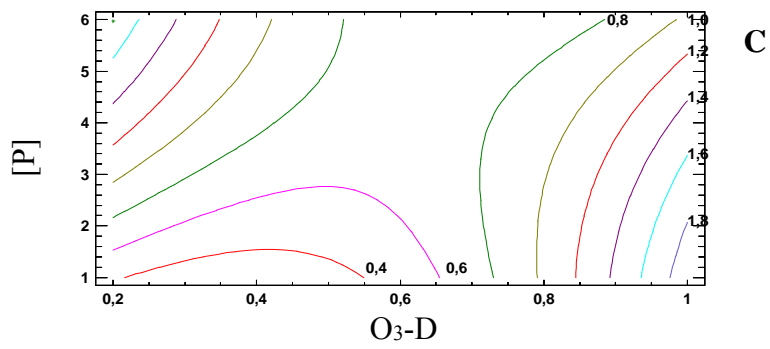
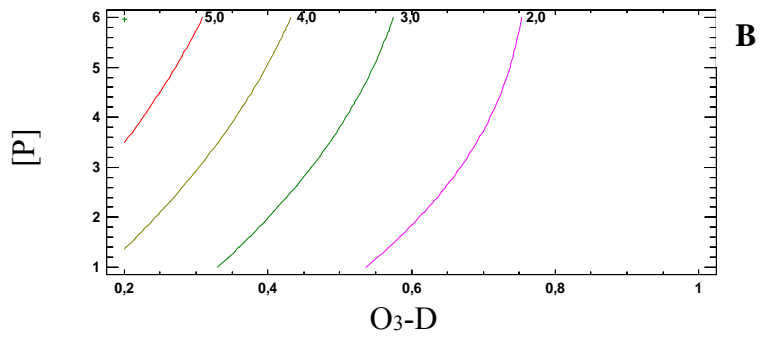
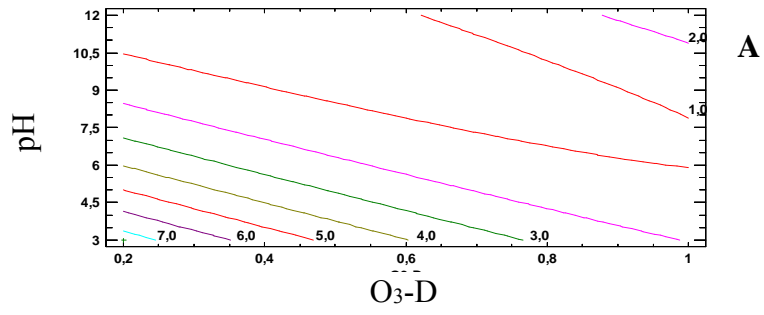
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553 Figure 2: Pareto charts for the response $t_{80\%}$ (min) obtained for the ozonisation (a) and
 554 the O₃/UVA-VIS combination (b) for the degradation of a mixture of six phenolic
 555 compounds.

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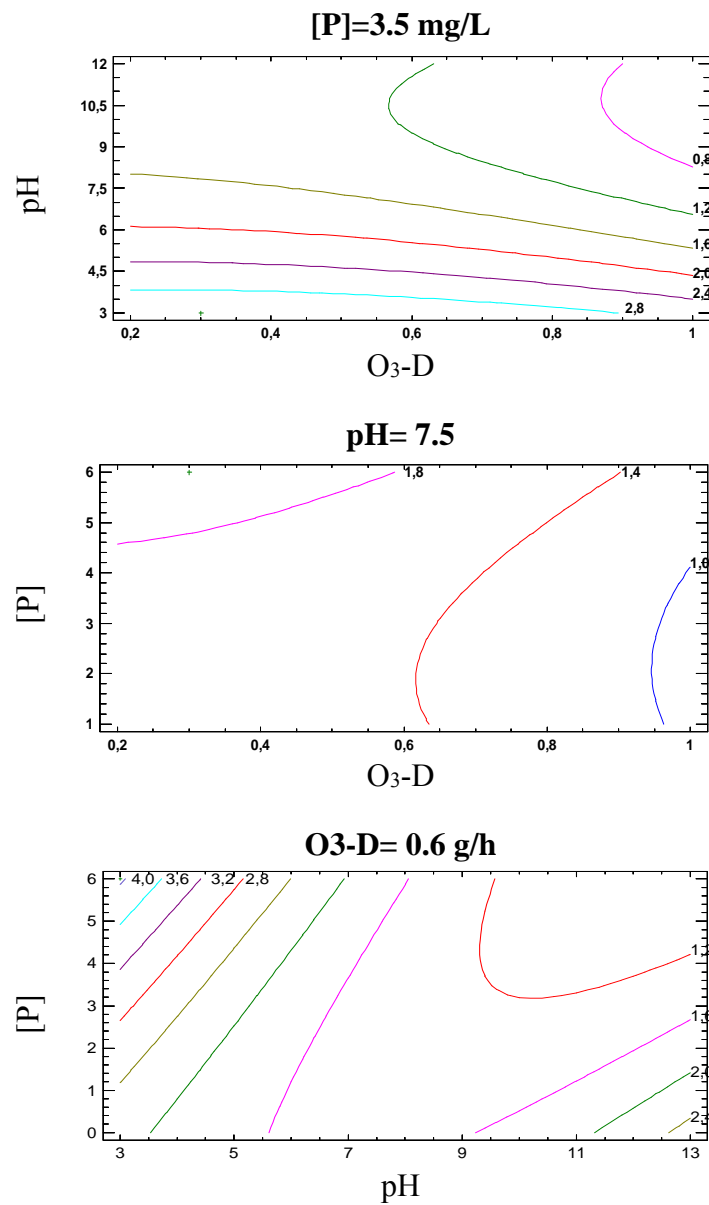
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558 Figure 3: Two-dimensional contour plots for $t_{80\%}$ (min) obtained for the removal of a
559 mixture of six phenolic compounds by ozonisation by fixing one parameter at a desired
560 value. A) $[P] = 3.5$ mg/L, B) pH = 5, C) pH = 10, D) $O_3-D = 0.3$ g/h, E) $O_3-D = 0.8$ g/h.

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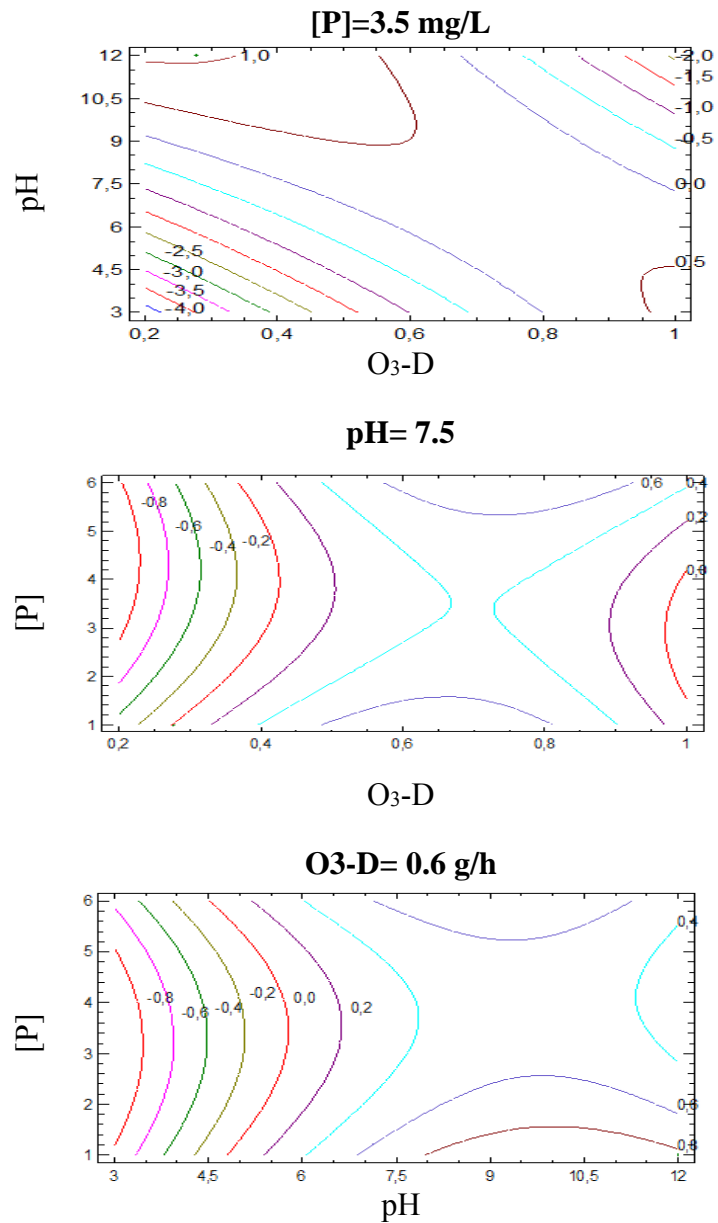
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565 Figure 4. Contour plots obtained for the removal of a mixture of six phenolic
566 compounds by O₃/UVA-Vis combination. Value of t_{80%} (min) at fixed values in the
567 midpoint of the three different factors (O₃-D, pH and [P] employed in the Doehlert
568 design.

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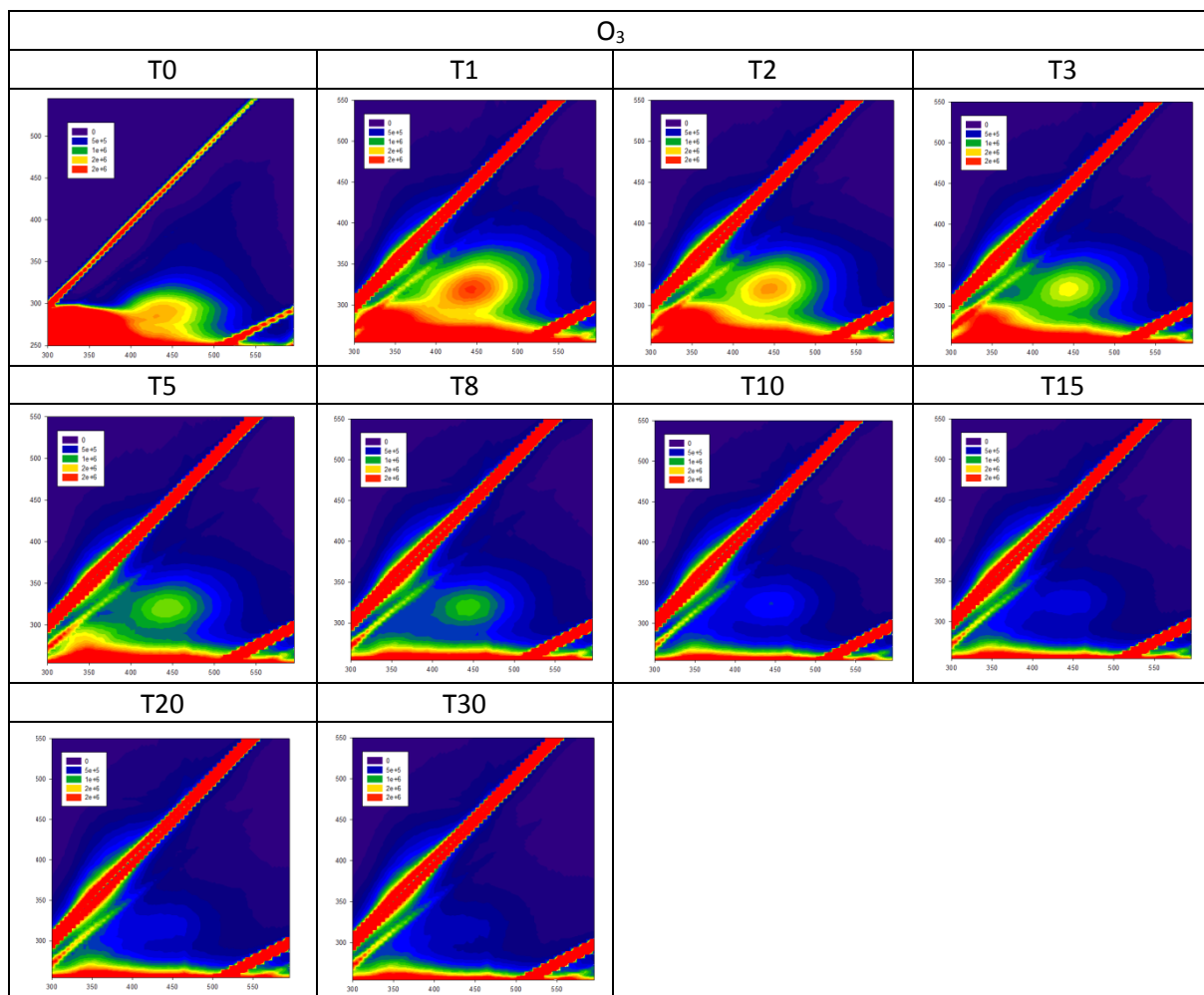
571 Figure 5. Contour plots obtained for the difference of $t_{80\%}$ in the combination O₃/UVA-
 572 Vis and O₃. Negative values show that UVA-vis is able to enhance the process.

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578 Figure 6: EEMs obtained for the removal of a mixture of six phenolic compounds by
 579 O₃ at different treatments times. Figures shows the excitation area (Y-axis) between 250
 580 and 550 nm, the emission area (X-axis) between 300 and 600 nm and the fluorescence
 581 intensity in the 0 and 2 x 10⁶ range.

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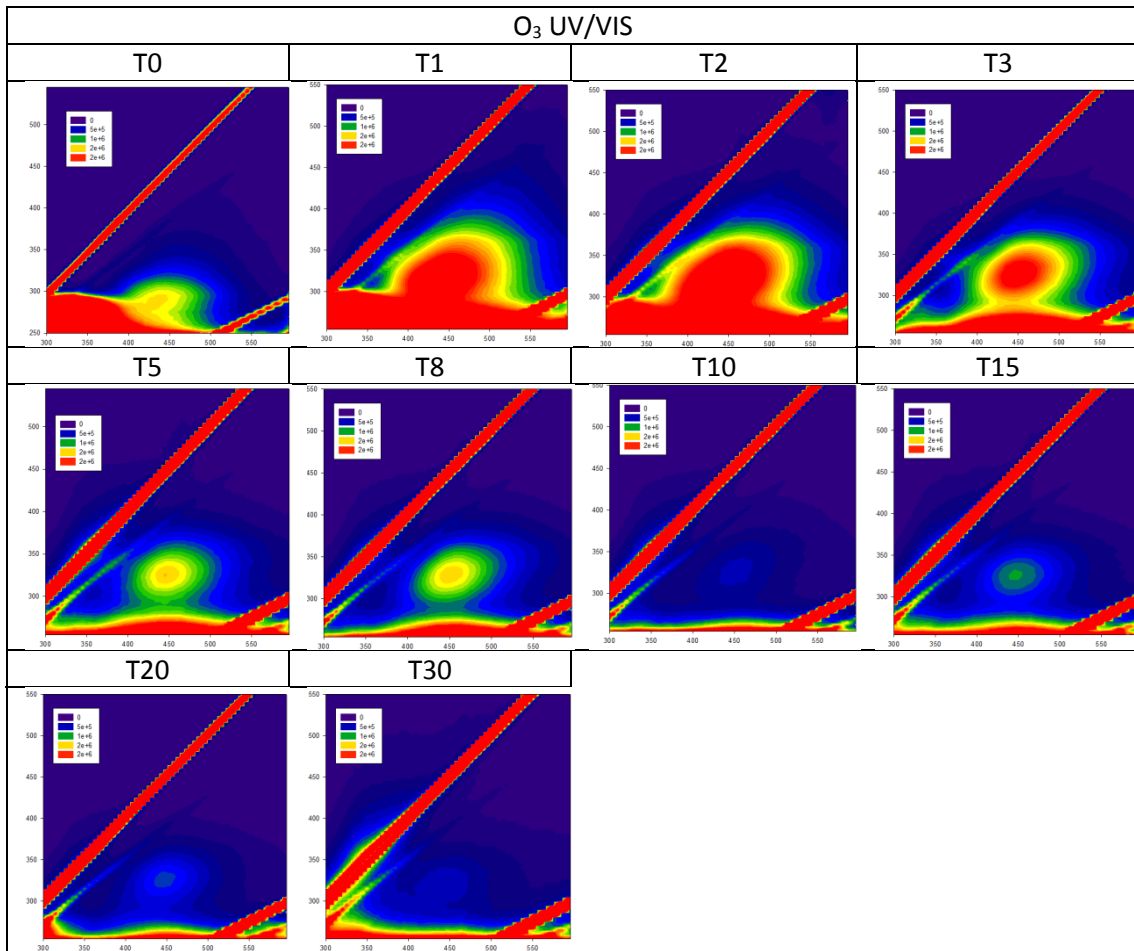
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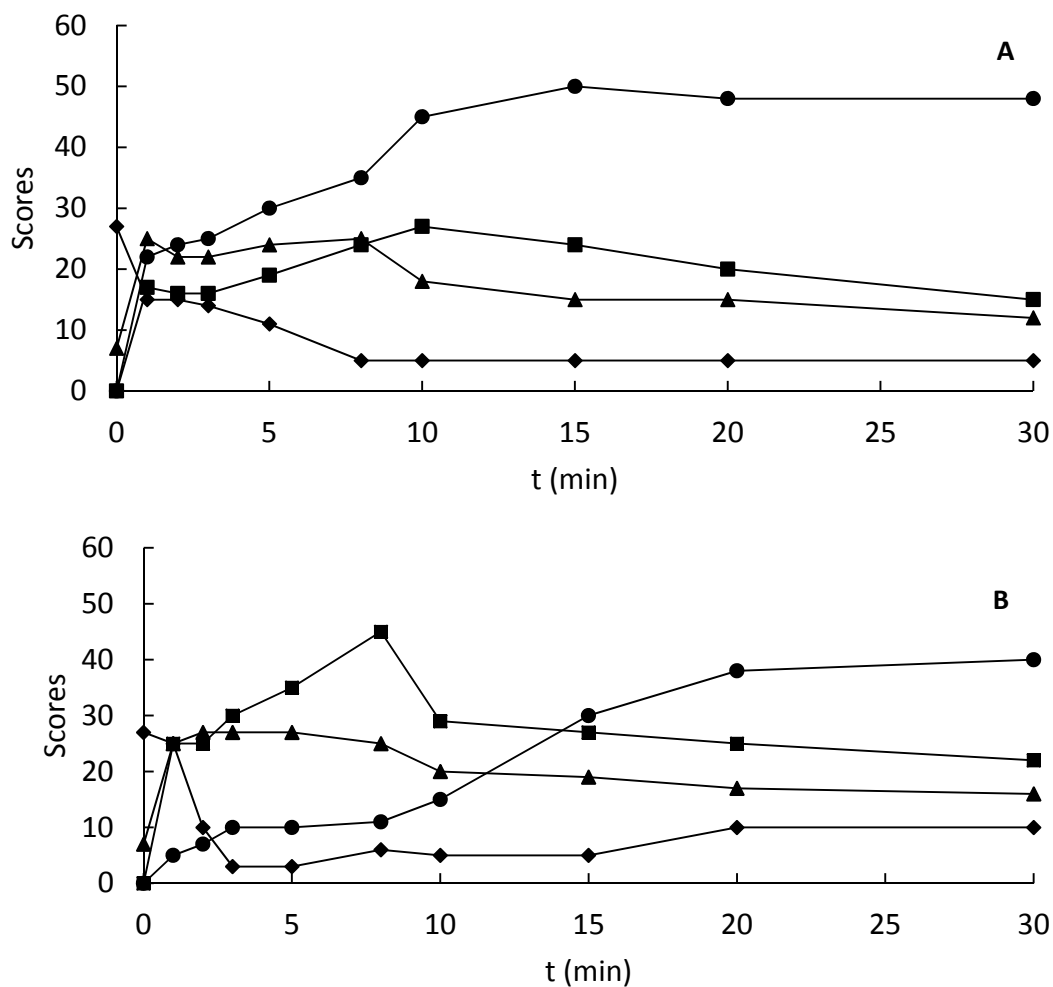


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593 Figure 7: EEMs obtained for the removal of a mixture of six phenolic compounds by
594 O₃/UVA-VIS combination at different treatments times. Figures shows the excitation
595 area (Y-axis) between 250 and 550 nm, the emission area (X-axis) between 300 and 600
596 nm and the fluorescence intensity in the 0 and 2 x 10⁶ range.

597



598 Figure 8. Time resolved plot of the scores reached by each component of the
 599 PARAFAC analysis of the EEM matrixes obtained in the O₃ (A) and O₃/UVA-VIS (B)
 600 treatment of a mixture of eight phenolic compounds The components are represented
 601 by: C1 (●), C2(■), C3 (▲) and C4 (◆)

602