Document downloaded from:

http://hdl.handle.net/10251/141951

This paper must be cited as:

Ferrandiz Manglano, ML.; Nacher-Juan, J.; Alcaraz Tormo, MJ. (06-2). Nrf2 as a therapeutic target for rheumatic diseases. Biochemical Pharmacology. 152:338-346. https://doi.org/10.1016/j.bcp.2018.04.010



The final publication is available at http://doi.org/10.1016/j.bcp.2018.04.010

Copyright Elsevier

Additional Information

### Accepted Manuscript

### Commentary

Nrf2 as a therapeutic target for rheumatic diseases

María Luisa Ferrándiz, Josep Nacher-Juan, Maria José Alcaraz

PII:	S0006-2952(18)30153-9
DOI:	https://doi.org/10.1016/j.bcp.2018.04.010
Reference:	BCP 13119
To appear in:	Biochemical Pharmacology
Received Date:	15 February 2018
Accepted Date:	11 April 2018



Please cite this article as: M.L. Ferrándiz, J. Nacher-Juan, M.J. Alcaraz, Nrf2 as a therapeutic target for rheumatic diseases, *Biochemical Pharmacology* (2018), doi: https://doi.org/10.1016/j.bcp.2018.04.010

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

### Nrf2 as a therapeutic target for rheumatic diseases

María Luisa Ferrándiz, Josep Nacher-Juan and Maria José Alcaraz

Instituto Interuniversitario de Investigación de Reconocimiento Molecular y Desarrollo Tecnológico (IDM), Universitat Politècnica de València, Universitat de València, Av. Vicent A. Estellés s/n, 46100 Burjasot, Valencia, Spain

MAN

E-mail: luisa.ferrandiz@uv.es, jojuana3@uv.es, maria.j.alcaraz@uv.es

Correspondence: María Luisa Ferrándiz, luisa.ferrandiz@uv.es

#### Abstract

Nuclear factor (erythroid-derived 2)-like 2 (Nrf2) is a master regulator of cellular protective processes. Rheumatic diseases are chronic conditions characterized by inflammation, pain, tissue damage and limitations in function. Main examples are rheumatoid arthritis, systemic lupus erythematosus, osteoarthritis and osteoporosis. Their high prevalence constitutes a major health problem with an important social and economic impact. A wide range of evidence indicates that Nrf2 may control different mechanisms involved in the physiopathology of rheumatic conditions. Therefore, the appropriate expression and balance of Nrf2 is necessary for regulation of oxidative stress, inflammation, immune responses, and cartilage and bone metabolism. Numerous studies have demonstrated that Nrf2 deficiency aggravates the disease in experimental models while Nrf2 activation results in immunoregulatory and anti-inflammatory effects. These reports reinforce the increasing interest in the pharmacologic regulation of Nrf2 and its potential applications. Nevertheless, a majority of Nrf2 inducers are electrophilic molecules which may present off-target effects. In recent years, novel strategies have been sought to modulate the Nrf2 pathway which has emerged as a therapeutic target in rheumatic conditions.

**Keywords**: Nrf2, rheumatic conditions, rheumatoid arthritis, systemic lupus erythematosus, osteoarthritis, osteoporosis

#### 1. Introduction

The nuclear factor (erythroid-derived 2)-like 2 (Nrf2) pathway is involved in the regulation of many antioxidant, anti-inflammatory and cell survival genes. Activation of this signaling pathway which contributes to detoxification and protective processes has encouraged a wealth of studies on its potential health benefits and therapeutic applications. Nrf2 is a CNC-bZIP (Cap'n'collar-basic region leucine zipper) transcription factor [1]. In unstressed conditions, Nrf2 binds to the cytosolic inhibitor Kelch-like ECH-associated protein 1 (Keap1) an adaptor protein to the cullin-3 E3 ubiquitin ligase complex [2], which targets Nrf2 for proteasomal degradation. Nrf2 contains two Keap1 binding motifs, ETGE and DLG. Keap1 is a main regulator of Nrf2 and a sensor of oxidative and xenobiotic stress. Under basal intracellular redox conditions, Keap1 drives Nrf2 regulation but in the presence of cellular stress. Nrf2 can be regulated by both Keap1-dependent and -independent mechanisms [3]. Nrf2 accumulates in the nucleus and binds to the antioxidant-response element (ARE) sites in the promoter of target genes as a heterodimer with a small musculoaponeurotic fibrosarcoma (sMaf) protein (Figure 1). In addition, Nrf2 has been reported to form heterodimers with proteins such as c-Jun, activating transcription factor 4, and others depending on cell type and stimuli [4,5]. Genes targeted by Nrf2 include genes involved in the synthesis and conjugation of glutathione, heme and iron metabolism, drug metabolism and transport, as well as antioxidant proteins, enzymes and transcription factors. Nrf2 is essential for the transcriptional induction of phase II detoxifying enzymes and antioxidant proteins which represent a main defense mechanism. For instance, glutathione reductase, glutathione S-transferase, y-glutamylcysteine synthetase,

NAD(P)H:quinone oxidoreductase-1 (NQO1) and heme oxygenase-1 (HO-1) (reviewed in [6]).

Oxidants and electrophiles modify Keap1 cysteine residues which causes a conformational change in this protein leading to cessation of Nrf2 polyubiquitination. Then, Nrf2 translocates to the nucleus to initiate the transcription of target genes [7,8]. Phosphorylation mediated by glycogen synthase kinase 3 (GSK-3) creates a recognition motif for the E3 ligase adapter  $\beta$ -transducin repeat containing E3 ubiquitin protein ligase ( $\beta$ -TrCP) leading to an alternative pathway for ubiquitin-dependent proteasomal degradation of Nrf2 [3].

Nrf2 can be regulated by acetylation by p300/cAMP response elementbinding protein (CREB)-binding protein (CBP) [9]. Thus, acetylation of Nrf2 results in binding to the ARE and activation of gene transcription, whereas deacetylation releases it leading to transcriptional termination and nuclear export [10]. Nrf2 acetylation is determined by the relative activities of histone acetyl transferases and histone deacetylases (HDACs) [9]. Bach proteins dimerize with sMaf proteins and these complexes compete with Nrf2-sMaf. In particular, Bach1 plays significant roles by activating and repressing transcriptional activities to regulate the oxidative stress response and suppress HO-1 [11].

Rheumatic diseases are chronic conditions affecting the musculoskeletal system. Arthritis and related illnesses cause inflammation, changes in the joints, pain and limitations in motion and function. These conditions have a profound effect on work capacity and quality of life of affected people. In the industrialized world rheumatic diseases affect more individuals than any other disease group (European League Against Rheumatism: www.eular.org, and American College

of Rheumatology: www.acr.org) and constitute a major health problem with an important social and economic impact. The burden related to these conditions is expected to increase in the near future as the population ages.

In recent years, several lines of evidence have supported the notion that Nrf2 plays a regulatory role not only in oxidative stress, but also in inflammation, immunity and cartilage and bone metabolism. The results of many *in vitro* and *in vivo* studies have led to propose that Nrf2 activation may control different processes and mediators involved in the physiopathology of rheumatic conditions. Some relevant examples can be rheumatoid arthritis (RA), systemic lupus erythematosus (SLE), osteoarthritis (OA) and osteoporosis. The aim of this Commentary is to focus on Nrf2 as a new therapeutic target in these conditions.

### 2. Nrf2 regulation of inflammatory and immune responses

A wide range of evidence indicates that Nrf2 plays an important role in the regulation of inflammation as well as in innate and adaptive immune responses. The control of redox activity by Nrf2 and antioxidant downstream targets may play a role in the activation of the NLR family, pyrin domain containing 3 (NLRP3) inflammasome. NLRP3 is involved in homeostasis and tissue repair although its dysregulation contributes to inflammatory and degenerative diseases [12]. The activation of the cytosolic inflammasome complex results in inflammatory caspase activation leading to the secretion of interleukin(IL)-1β and IL-18 mainly in monocytes/macrophages and dendritic

cells (DCs) [13]. Increased NLRP3 inflammasome activity is a key feature of both autoinflammatory and autoimmune diseases [14] and Nrf2 has recently been proposed as a potential target for the therapeutic modulation of NLRP3associated diseases [15].

Nevertheless, Nrf2 deficiency in mice results in defective activation of NLRP3 and absent in melanoma 2 (AIM2) inflammasomes in bone marrow macrophages after treatment with a variety of stimuli including monosodium urate and non crystalline agents [16]. In contrast, it has been reported that Nrf2 is an inhibitor of NLRP3 expression at the transcriptional level and thus, Nrf2 activation inhibited lipopolysaccharide-induced NLRP3 production in THP1 cells [15]. These conflicting findings clearly emphasize the need of studying the precise molecular mechanisms involved in Nrf2 interaction with inflammasomes.

Nrf2 not only inhibits inflammation through redox control but also downregulates pro-inflammatory cytokines, chemokines, adhesion molecules and enzymes. Nrf2 and HO-1 have shown the ability to control the migration of inflammatory cells, a key process in the development of chronic inflammatory conditions. The inhibition of adhesion molecules and matrix metalloproteinase (MMP) expression can mediate these anti-inflammatory effects [17,18].

Nrf2 activity is essential to control cellular mechanisms contributing to the resolution of the inflammatory process. For this purpose, Nrf2 interplays with nuclear factor- $\kappa$ B (NF- $\kappa$ B) through multiple molecular interactions (reviewed in [19]). Phosphorylation of NF- $\kappa$ B inhibitor (I $\kappa$ B $\alpha$ ) by I $\kappa$ B kinase (IKK $\beta$ ) leads to I $\kappa$ B $\alpha$  degradation which results in nuclear translocation and DNA binding of NF- $\kappa$ B. Hydroperoxides can regulate NF- $\kappa$ B activation by several mechanisms. For instance, in basal redox conditions, Keap1 is responsible for IKK $\beta$  ubiquitination

and degradation but in the presence of oxidants, Keap1 is inhibited and NF- $\kappa$ B can be activated [20]. Nrf2 decreases NF- $\kappa$ B activation by interacting with Keap1. In addition, Nrf2 can interact with the sMaf protein MafK. This protein enhances the acetylation of p65 and thus the DNA-binding activity of NF- $\kappa$ B. Therefore, Nrf2 may maintain low levels of MafK avoiding excessive p65 acetylation [21].

Nrf2 contains several  $\kappa$ B sites in its proximal promoter, which are subjected to regulation by p50 and p65. In turn, NF- $\kappa$ B may play a dual role in the regulation of Nrf2 activity. NF- $\kappa$ B activation induces Nrf2 expression in cells such as human monocytes and acute myeloid leukemia cells, leading to enhanced activation of Nrf2-dependent antioxidant defense responses but it can also inhibit Nrf2 by several mechanisms such as the competition for the transcriptional co-activator p300/CBP. In addition, NF- $\kappa$ B increases the recruitment of HDAC3 to the ARE region and thus Nrf2 transcriptional activation is prevented [18].

Nrf2 activation promotes the resolution of inflammation through the induction of prostaglandin (PG) D synthase expression in macrophages leading to the rapid production of PGD<sub>2</sub>/15-deoxy-delta(12,14)-prostaglandin J<sub>2</sub> (15d-PGJ<sub>2</sub>) which sustains a positive feedback loop to limit the inflammatory response. It has been shown that 15d-PGJ<sub>2</sub> activates Nrf2 leading to the induction of CD36 and HO-1 in macrophages to promote efferocytosis and the resolution of inflammation [22].

Nrf2 deficiency induces autoimmune phenotypes in certain strains of mice and increases susceptibility to the development of autoimmune diseases [23] whereas Nrf2 activation is associated with the attenuation of these

conditions [17,24]. In the presence of Nrf2 dysregulation, oxidative tissue damage and apoptosis could increase the production of autoantigens leading to activation of T cells and production of autoantibodies by B cells. In addition, loss of phase II enzymes causes an elevation in the steady state of reactive intermediates by failing to remove them. This can promote the activation of immune cells. As Nrf2 is a master regulator of cellular responses against environmental stresses [8], it is likely that Nrf2 activation can protect against environmental factors contributing to autoimmune pathogenesis.

Nrf2-mediated regulation of autoimmune function can involve the suppression of pro-inflammatory T helper(Th)1 and Th17 cell responses, and the enhancement of anti-inflammatory Th2, regulatory T cells (Treg) and regulatory B cells functions (Figure 2). In addition, Nrf2 may control the differentiation and function of DCs and macrophages. Nrf2 deficiency alters the function and phenotype of DCs with increased co-stimulatory molecule expression and enhanced antigen-specific T cell stimulatory capacity in immature cells [25].

The influence of Nrf2 on T cell function is complex. Disruption of Nrf2 limits glutathione availability leading to the inhibition of antigen-induced CD8+ T cell proliferation and function [26]. In contrast, Nrf2 activation inhibits secretion of the Th1 cytokines, interferon  $\gamma$  (IFN- $\gamma$ ) and tumor necrosis factor  $\alpha$  (TNF $\alpha$ ), promotes early production of IL-2 [27] and skews CD4+ T cells toward Th2 differentiation [28]. Recently, clustered regularly interspaced short palindromic repeats (CRISPR) technology has been used for *ex vivo* Keap1 editing in primary human T cells in order to achieve Nrf2 activation and enhanced antiinflammatory and immunosuppressive functions [29].

The inhibition of transcription of inflammatory mediators may contribute to the therapeutic effects of Nrf2 activation in autoimmune diseases. It also appears that HO-1 induction may contribute to the immunosuppressive ability of Nrf2 activation. It is known that HO-1 and CO reduce the capacity of antigenpresenting cells, such as DCs and macrophages to recognize pathogenassociated molecular patterns thus suppressing both antigen presentation and the production of pro-inflammatory cytokines [19]. HO-1 activity was also found to modulate the proliferative capacity of T cells, the effector functions of T cells and natural killer cells, and the suppressive functions of Treg [30].

The immunoregulatory effects of Nrf2 have been classically related to the control of oxidative stress and phase II enzymes, glutathione levels or NF- $\kappa$ B activation [19]. Nevertheless, some data suggest the contribution of other mechanisms such as the p38-CREB/activating transcription factor 1 (ATF1) signaling axis in DCs [25] or the disruption of RNA polymerase II recruitment which results in the inhibition of IL-6 and IL-1 $\beta$  transcription in macrophages [17].

### 3. Rheumatoid arthritis

RA is a chronic autoimmune disease characterized by synovial hyperplasia, immune cell infiltration and degradation of cartilage and bone. There are activation and migration of neutrophils, macrophages, and lymphocytes which result in the increased production of pro-inflammatory mediators such as oxidants, eicosanoids, cytokines (IL-17, TNF $\alpha$ , IFN- $\gamma$ , IL-6, and IL-1 $\beta$ ) and catabolic enzymes, with hyperproliferation of synovial fibroblasts

[31]. This results in joint swelling and progressive destruction of cartilage and bone. Excessive oxidant generation may contribute to the pathogenesis of RA. In fact, RA patients show a marked increase in lipid peroxidation, protein oxidation and DNA damage associated to a reduced activity of the antioxidant defense system, which may contribute to tissue damage and the perpetuation of disease [32]. As a response to oxidative stress, Nrf2 expression is activated in synovial cells from RA patients and also in joints of antibody-induced arthritic mice although this response is not enough to counteract arthritis progression [33]. In fact, enhanced gene expression of Nrf2/HO-1 in a subgroup of RA patients has been related to more severe disease state with an underlying lack of apoptosis in synovial fibroblasts, macrophages, lymphocytes, and other cells that may contribute to the persistence of RA [34].

Nrf2 deficiency enhances joint alterations in experimental RA models. In K/BxN serum transfer arthritis and antibody-induced arthritis, Nrf2 deletion accelerates the incidence and aggravates the disease, with important inflammation and lesions [33,35]. Nrf2 deficiency dramatically upregulates oxidative stress, cell migration, cyclooxygenase-2 (COX-2) and inducible nitric oxide synthase expression, the pro-inflammatory cytokines TNF $\alpha$  and IL-6, as well as the chemokine CXCL-1. In addition, we showed that Nrf2 may be a protective factor for bone metabolism in the presence of arthritis [35]. On the contrary, Nrf2 activation and HO-1 induction exert anti-inflammatory and antioxidant effects in animal models of RA and in human RA synovial fibroblasts [36,37].

Many molecules have shown anti-inflammatory and immunoregulatory properties via the oxidative stress-responsive transcription factor Nrf2. There is

a variety of Nrf2 inducers, most of which are electrophilic and react with cysteine thiols of Keap1. In particular, Cys151, Cys273, and Cys288 play an important role in Nrf2 activation. Therefore, modification of thiol moieties in Keap1 leads to disturbance of the structure and the decline of ubiquitin ligase activity [38]. As a main example, sulforaphane (SFN, Figure 3) can exert immunoregulatory effects leading to the inhibition of T cell proliferation and the production of IL-17 and TNF $\alpha$  by RA CD4+ T cells [39]. This compound is also able to polarize pro-inflammatory M1 macrophages into anti-inflammatory M2 cells [40]. In cultured human synoviocytes, SFN induced apoptosis by modulating the expression of Bcl-2/Bax, p53, and pAkt and inhibited inflammation [39]. Intraperitoneal administration of SFN to mice reduced the clinical severity of collagen-induced arthritis, anti-collagen II antibody levels, T cell responses and the production of IL-17, TNF $\alpha$ , IL-6, and IFN- $\gamma$  by lymph node cells and spleen cells [39]. Part of the anti-inflammatory effects of SFN would be dependent on Nrf2 activation which indirectly inhibits NF-κB via HO-1mediated CO production but part of them are independent of Nrf2 as SFN can directly inhibit NF-kB [41].

The Nrf2 activator and immunoregulatory agent dimethyl fumarate (DMF) is used in systemic sclerosis and severe plaque psoriasis. It has been reported that DMF partly exerts its anti-inflammatory effects via inflammasome inhibition [42]. Furthermore, blockade of T cell activation by DMF may be related to binding specific cysteine residues in protein kinase C (PKC) $\theta$  which is a key kinase in T cell signaling [43], and DMF and monomethylfumarate (MMF) may activate the hydroxycarboxylic acid receptor 2 (HCAR2) which results in downregulation of NF- $\kappa$ B [44].

Many studies have reported the effects of polyphenols, and mainly of the green tea's active ingredient, epigallocatechin 3-gallate in preclinical models of RA as well as *in vitro*, in cartilage and bone protection and synovial fibroblast regulation. This compound induces Nrf2 and HO-1 in different cell types while decreasing NF-kB activity and the production of inflammatory and extracellular matrix degradative mediators [45]. The activation of synovial fibroblasts and the production of inflammatory cytokines are crucially involved in the pathogenesis of RA. It is interesting to note that calycosin, an isoflavone from the Chinese medicinal herb Radix Astragali, has been reported to downregulate pro-inflammatory cytokines and COX-2 via p62-Nrf2-HO-1 induction in RA synovial fibroblasts [46].

Antirheumatic gold(I)-containing compounds stimulate the antioxidative stress response through activation of Nrf2/sMaf leading to the upregulation of HO-1 and  $\gamma$ -glutamylcysteine synthetase [47]. Nrf2-HO-1 activation also mediates the anti-inflammatory effects of H<sub>2</sub>S and related compounds which are able to modify by sulfydrylation the cysteine residue of Keap1. For instance, the endogenous H<sub>2</sub>S modulator S-propargyl-cysteine is able to reduce the generation of inflammatory mediators, oxidants, and MMP-9 as well as the cell invasive activity in rheumatoid fibroblast-like synoviocytes MH7A. *In vivo*, this compound ameliorated the severity of arthritis in the adjuvant model in rats [37].

Oxidized phospholipids able to regulate antioxidant gene expression via Nrf2 signaling *in vivo* have been detected under chronic inflammatory conditions. Interestingly, epoxycyclopentenone derivatives have been shown to activate Nrf2 leading to inhibition of pro-inflammatory cytokines and chemokines in myeloid cells. These effects were similar to those of the pro-resolving lipid

mediator 15d-PGJ2 which has been reported to interact with the nuclear hormone receptor peroxisome proliferator-activated receptor- $\gamma$  (PPAR- $\gamma$ ) as well as with Nrf2. Of note, 15d-PGJ<sub>2</sub> was active in PPAR- $\gamma$ -deficient cells but not in Nrf2-deficient cells implying that the anti-inflammatory activity of 15d-PGJ<sub>2</sub> was mediated via Nrf2 rather than PPAR- $\gamma$  [48]. 15d-PGJ<sub>2</sub> forms an adduct to Keap1 and disrupts Nrf2 ubiquitination, leading to the accumulation of Nrf2 in the nucleus. In addition, 15d-PGJ<sub>2</sub> ameliorated adjuvant-induced arthritis with suppression of pannus formation and mononuclear cell infiltration [49].

Polyunsaturated fatty acids are able to inhibit inflammation in macrophages through the induction of Nrf2 [50]. It is interesting to note that eicosapentaenoic acid and docosahexaenoic acid inhibit inflammatory cartilage degradation. The last compound also ameliorates disease activity in patients with RA and enhances plasma levels of pro-resolving maresin/resolvin precursors [51].

### 4. Systemic lupus erythematosus

An increased production of oxidative stress may contribute to immune cell death and autoimmunity in SLE. Necrosis secondary to deregulated cell death and removal processes results in the generation of autoantigens and formation of immune complexes which induce inflammation and tissue damage in organs such as the kidney, skin and joints. SLE patients show alterations in repair mechanisms of oxidative DNA damage, high serum levels of oxidized proteins, apoCIII, oxidized phospholipids and autoantibodies to oxidatively modified lipoproteins [52].

Nrf2 polymorphisms have not been associated to lupus susceptibility although the Nrf2 -653 G/A polymorphism is related to the risk of nephritis among Mexican childhood-onset female SLE patients [53]. Increased levels of Nrf2, NQO1 and 8-oxo-7,8-dihydro-2'-deoxyguanosine were observed in glomeruli from human lupus nephritis. Nrf2 is also induced in other types of nephritis and may result from immune-complex deposition [54].

Female mice deficient in Nrf2 develop with age a multi-organ autoimmune disorder similar to SLE with increased DNA oxidation and lipid peroxidation, splenocyte apoptosis, presence of antibodies against dsDNA and the Smith antigen, and tissue damage (vasculitis, glomerulonephritis, hepatitis, and myocarditis) [55]. According to the genetic background of mice, there are differences in the age necessary to develop these changes [23]. Although oxidative damage due to Nrf2 deficiency is present in both male and female mice, only female mice show progression to SLE suggesting that genderspecific factors are involved in breaking of immune tolerance to self antigens [55]. In addition to deficiency in phase 2 detoxification enzymes and antioxidant genes in hepatic and lymphoid cells which results in oxidative damage to tissues, Nrf2 knock-out results in enhanced proliferative responses of CD4+ T cells, altered CD4+/CD8+ ratios and promotion of Th17 cells differentiation and function [56]. Nrf2 deletion has also been shown to promote Th17 differentiation and function during lupus nephritis development by regulating the suppressor of cytokine signaling 3 (SOCS3)/signal transducer and activator of transcription (STAT) 3 pathway and IL-1ß [56].

Nrf2 inducers inhibit the development of disease in animal models such as pristane-induced lupus nephritis and spontaneous lupus in MRL/lpr mice.

Therefore, induction of Nrf2 by SFN protects renal cells from developing lupus nephritis by downregulation of oxidative stress and inhibition of NF-KB and extracellular matrix deposition [57]. Interestingly, fumaric acid esters have been used as systemic combination therapy in the treatment of severe, extensive and recalcitrant cutaneous manifestation of SLE. They were well-tolerated and showed excellent efficacy and a steroid-sparing effect [58], supporting the interest of this approach in the therapy of lupus. As another example, epigallocatechin-3-gallate prevents lupus nephritis development via the upregulation of the Nrf2 antioxidant pathway, which inhibits NLRP3 inflammasome activation [59]. NAS

### 5. Osteoarthritis

OA is characterized by a progressive cartilage degradation associated with hypertrophic differentiation of chondrocytes, synovitis and alterations in subchondral bone and periarticular tissues. Long-time exposure to a low-grade chronic inflammation concomitant with a failure in oxidant-antioxidant balance has an important impact on the pathogenesis of disease. Catabolic and proinflammatory mediators are produced by the inflamed synovium leading to excess production of proteolytic enzymes responsible for cartilage breakdown [60].

Mitochondrial dysfunction and oxidative damage are involved in the pathogenesis of OA. Oxidative stress is involved in the production of inflammatory and catabolic mediators and also contributes to joint degradation

by mechanisms such as the reduction in extracellular matrix synthesis, the induction of chondrocyte apoptosis and MMP activation [61]. Therefore, the control of oxidative stress and chronic inflammation by Nrf2 would result in protective effects against joint alterations in OA (Figure 4). Furthermore, as revised in the osteoporosis section, Nrf2 is an important factor to regulate the balance between osteoclast-driven bone resorption and osteoblast-driven remodeling which may play a role in the control of bone metabolism in OA.

The appropriate expression and balance of Nrf2 is necessary for normal chondrogenesis and regulation of cartilage metabolism. In fact, sustained overexpression of Nrf2 can inhibit chondral differentiation markers as collagen II, collagen X and osteopontin [62], but downregulation of Nrf2 could result in inhibition of chondrogenesis through apoptotic cell death [63]. In agreement with these findings, Nrf2 and glutathione transferase A4-4 expression is significantly lower in OA cartilage from humans and mice in comparison to normal controls [64].

The control of excessive oxidative stress and pro-inflammatory and catabolic mediators may sustain a protective role of Nrf2 in OA. In this context, Nrf2 knock-out mice display more severe cartilage damage in both the monoiodoacetate (MIA) and the surgical destabilization of medial meniscus (DMM) models of OA [65]. Induction of HO-1 by Nrf2 activation can also play a role in its anti-inflammatory and chondroprotective effects as HO-1 is able to reduce NF-κB activity and inflammatory and degradative mediators in OA chondrocytes, synoviocytes and osteoblasts [66-68].

Recent investigations have shown that induction of Nrf2 by protandim (a commercial dietary supplement composed of five antioxidant phytochemicals)

and 6-gingerol mediates their protective effects in OA chondrocytes, where both drugs were able to attenuate the production of oxidative stress and inflammatory mediators as well as 6-hydroxynonenal-induced cell death. In addition, protandim administration to mice significantly reduced joint destruction in the DMM [64]. Piceatannol is another Nrf2 inducer with protective effects in this model of OA that also inhibits the production of inflammatory mediators, MMP-13 and aggrecanase-2 in OA chondrocytes stimulated with IL-1 $\beta$  [69]. Recently, the natural flavonoid wogonin has been reported to exert anti-inflammatory and protective effects in human OA chondrocytes and cartilage explants [70]). Wogonin modulates the oxidant-mediated activation of Nrf2 signaling axis and also disrupts Keap 1/Nrf2 interaction by blocking the binding site of Nrf2 in Keap 1 protein.

Nrf2 acetylation, mediated by histone acetyltransferase/HDAC enhances its transcriptional ability and the expression of downstream targets. Therefore, inhibition of HDAC results in Nrf2 activation. This is the mechanism of action of trichostatin A, a pan-HDAC inhibitor which protects against cartilage degradation and inflammation in the MIA and DMM models of OA through the induction of Nrf2 in joint tissues [65]. Besides trichostatin A, other HDAC inhibitors such as sodium butyrate and vorinostat have been shown to reduce inflammatory responses and the upregulation of MMPs and aggrecanase 2 in human OA chondrocytes. As a result, HDAC inhibitors have demonstrated protective effects against cartilage degradation through mechanisms such as Nrf2 activation and the inhibition of NF-κB and MAPK [71].

Thus, strategies aimed at stimulating antioxidant gene expression, as HO-1 and NQO-1, through Nrf2 activation in aging cartilage may hold promise

for OA therapy [72]. Nevertheless, it should be taken into account that some Nrf2 inducers can protect against cartilage degradation by other mechanisms. For instance, SFN inhibited cytokine-induced MMP expression in human articular chondrocytes and fibroblast-like synovial cells independently of Nrf2 and HDAC activity [73].

#### 6. Osteoporosis

Formation and maintenance of bone tissues are regulated by two main mechanisms, bone formation by osteoblasts and bone resorption by osteoclasts. Many factors can disturb bone remodeling and break the balance between bone resorption and formation, which contribute to osteoporosis. Activation of immune cells in chronic inflammation results in an excessive production of bone-resorbing cytokines which are major stimulators of osteoclastogenesis. Elevated oxidative stress contributes to alterations in bone metabolism. As a consequence, there is systemic or local bone loss associated with osteoporosis, RA, etc. [74].

Nrf2 is one of the transcription factors responsible for the regulation of differentiation and function of osteoblasts and osteoclasts in normal bone metabolism. The results from different studies indicate that Nrf2 is indispensable for normal bone microarchitecture and suggest a role for this transcription factor in the maintenance of bone integrity in pathological situations.

Nrf2 plays an essential role in bone regeneration as Nrf2 knock-out mice show impaired fracture healing [75]. The Keap1/Nrf2 axis regulates receptor activator of nuclear factor κ-B ligand (RANKL)-dependent osteoclastogenesis via expression of enzymes such as HO-1, γ-glutamylcysteine synthetase, and glucose-6-phosphate dehydrogenase which modulate intracellular oxidative stress signaling [76]. Nrf2 deficiency promotes osteoclast differentiation mediated by increased oxidants production and activation of MAPKs and nuclear factor of activated T-cells 1 leading to bone resorption [77]. These animals show enhanced RANKL and osteoclast numbers accompanied by a decrease in osteoblast mineralization which increase their susceptibility to radiation-induced bone loss [78].

However, the effects on osteoblast metabolism are more complex. A high level of oxidative stress exerts negative effects on osteoblast metabolism and results in cell damage which suggests a protective role for Nrf2. Nevertheless, there are conflicting reports as Nrf2 regulates the antioxidant endogenous response and bone accrual differently depending on factors such as sex and age [79]. Stable overexpression of Nrf2 exerts negative effects on MC3T3 osteoblastic cells differentiation through inhibition of Runx2-dependent transcriptional activity [80]. In experiments using Nrf2 knock-out mice, it was concluded that Nrf2 exerts inhibitory effects on osteoblastic and osteoblastic differentiation with a higher effect on osteoblasts, in 9-week old mice [81]. In contrast, it has been reported that female Nrf2 knock-out mice exhibit a marked deficit in postnatal bone acquisition, by 3 weeks, related to a low osteoblast number and increased oxidants production which might impair early osteoblastogenesis and lead to the failure of bone acquisition [82]. In line with

these results, we have shown that Nrf2 deficiency in female mice leads to increased oxidative stress, bone turnover and bone resorption as a result of the predominance of osteoclastic activity over osteoblastic activity [83].

In addition, data from a model of osteoporosis in ovariectomized mice suggest a role for Nrf2 in bone anabolic effects, as an increase in HDAC2 by miR-455-3p inhibited the activation of Nrf2/ARE leading to increased oxidative stress and inhibition of osteoblast growth exacerbating osteoporosis [84].

Nrf2 inducers may be useful as inhibitors of bone destruction. As an example, DMF is able to inhibit RANKL-mediated osteoclastogenesis and attenuate bone destruction in lipopolysaccharide-treated mice [85]. SFN, epigallocatechin gallate [86], carnosic acid [87], caffeic acid phenethyl ester [88] and an ETGE-peptide [89] can also inhibit osteoclastogenesis through Nrf2 activation while mangiferin protects osteoblasts from oxidative stress by this mechanism [90].

Activation of Nrf2 has been suggested as a therapeutic target to avoid glucocorticoid-induced osteoporosis. Thus, indole-3-carbinol, a natural product found in broadly consumed plants of the *Brassica* genus [91], alpinumisoflavone [92] and icariside II [93] block oxidants overproduction and osteoblast apoptosis induced by dexamethasone through Nrf2 induction.

Another strategy to control excessive bone resorption may be the induction of Bach1 nuclear export which activates Nrf2-dependent antioxidant enzyme expression leading to the attenuation of osteoclastogenesis [94].

#### 7. Perspectives

Our understanding of the significance of Nrf2 activation for the control of human disease has so far been greatly hampered by the complexity of signaling pathways and biological responses modified by this transcription target as exemplified by the knowledge of Nrf2 interactome, regulome and fine-tuned regulatory loop [95]. As a consequence, there are discrepancies with regard to the reported biological effects of Nrf2 modulation although most results primarily reveal an anti-inflammatory effect.

At present, the possible crosslink between the Nrf2-ARE pathway and NLRP3 inflammasome is not well understood. Further in-depth biochemical analysis about Nrf2 and inflammasome interactions should provide the mechanistic insights necessary to establish the potential interest of Nrf2 in inflammasome-related diseases.

The regulation of Nrf2 opens up new therapeutic opportunities for the treatment of rheumatic conditions. Classic Nrf2 activators mimic the endogenous process of Nrf2 activation by covalent modification of cysteine groups in Keap1. At the same time, they can exert non-specific effects through the covalent modification of nucleophilic groups in proteins. Many types of post-translational oxidation reactions are thus possible leading to protein changes in conformation and activity in a wide range of kinases, phosphatases, transcription factors, transporters and cytoskeletal proteins [96]. As a consequence, significant side effects can be caused by these drugs. In this regard, less electrophilic derivatives such as MMF and monoethyl fumarate may be safer drugs than DMF [97]. Therefore, chemical modification of classic Nrf2

activators can lead to the development of potential therapeutic agents based on the Nrf2 pathway.

Another crucial aspect is the need for efficient control of intensity and duration of Nrf2 activation as it has been demonstrated that high-level and/or long-term activation of Nrf2 and its target genes, which are associated with the stress response in a normal physiological context, may result in deleterious effects such as growth of cancer cells and chemoresistance [38]. These observations support the importance of approaches for inducible/transient expression of cytoprotective enzymes which can be achieved with some drugs such as synthetic triterpenoids [98].

Other strategies focus on non-canonical mechanisms of Nrf2 activation. Nrf2 interaction with Keap1 can be disrupted by protein-protein interaction inhibitors such as peptide antagonists. Therefore, the peptide DEETGE-CAL-Tat has been shown to effectively activate Nrf2 and protect against cerebral ischemia in a pre-clinical model [99]. In addition, small-molecule Keap1–Nrf2 protein-protein interaction inhibitors have been designed [100].

Activation of Nrf2 can be achieved by Bach1 gene knockout which has shown immunoregulatory and protective effects in different disease models suggesting that drugs binding Bach1 may be a novel strategy to enhance Nrf2 activity [96] which may be useful in the control of autoimmune diseases.

There is a need of selective Nrf2 activation in target cells thus avoiding indiscriminated activation throughout the body and possible toxicity. In this respect, a number of pro-drugs able to be converted into Nrf2-activating molecules in the presence of oxidative stress or specific enzymes are under study [96]. In addition, the application of new technologies such as CRISPR

have the potential for clinical translation and can help to understand the mechanisms involved in Nrf2 effects in rheumatoid conditions.

These attractive strategies related to the Nrf2 pathway need to be validated to allow the development of new therapeutic agents. Activation of Nrf2 may also lead to adjuvant drugs thus helping improve cellular responses to other treatments. More studies are needed for a deep understanding of Nrf2 mechanisms and effects as a necessary step before entering clinical trials of rheumatic diseases.

On the other hand, the development of biomarkers provides objective parameters for diagnostic or prognostic purposes. Clinically validated biomarkers are needed in rheumatic diseases largely in early phases of disease. Nrf2 activation and expression of its target genes have been considered as biomarkers of oxidative stress in different pathological states such as cancer or multiple sclerosis. However, the potential of this pathway to become a biomarker for rheumatic diseases is not known and it should be explored.

**Acknowledgements**: This work has been funded by grant SAF2017-85806-R (MINECO, FEDER, Spain).

Conflict of interests: there is no conflict of interest to declare.

#### 8. References

- [1] Itoh K, Igarashi K, Hayashi N, Nishizawa M, Yamamoto M. Cloning and characterization of a novel erythroid cell-derived CNC family transcription factor heterodimerizing with the small Maf family proteins. Mol Cell Biol 1995;15:4184-93.
- [2] Villeneuve NF, Lau A, Zhang DD. Regulation of the Nrf2-Keap1 antioxidant response by the ubiquitin proteasome system: an insight into cullin-ring ubiquitin ligases. Antioxid Redox Signal 2010;13:1699-712.
- [3] Tebay LE, Robertson H, Durant ST, Vitale SR, Penning TM, Dinkova-Kostova AT, Hayes JD. Mechanisms of activation of the transcription factor Nrf2 by redox stressors, nutrient cues, and energy status and the pathways through which it attenuates degenerative disease. Free Radic Biol Med 2015;88, Part B:108-46.
- [4] He CH, Gong P, Hu B, Stewart D, Choi ME, Choi AM, Alam J. Identification of activating transcription factor 4 (ATF4) as an Nrf2-interacting protein. Implication for heme oxygenase-1 gene regulation. J Biol Chem 2001;276:20858-65.
- [5] Venugopal R and Jaiswal AK. Nrf2 and Nrf1 in association with Jun proteins regulate antioxidant response element-mediated expression and coordinated induction of genes encoding detoxifying enzymes. Oncogene 1998;17:3145-56.
- [6] Suzuki T, Motohashi H, Yamamoto M. Toward clinical application of the Keap1-Nrf2 pathway. Trends Pharmacol Sci 2013;34:340-6.
- [7] Dinkova-Kostova AT, Kostov RV, Canning P. Keap1, the cysteine-based mammalian intracellular sensor for electrophiles and oxidants. Arch Biochem Biophys 2017;617:84-93.
- [8] Suzuki T and Yamamoto M. Stress-sensing mechanisms and the physiological roles of the Keap1-Nrf2 system during cellular stress. J Biol Chem 2017;292:16817-24.

- [9] Sun Z, Chin YE, Zhang DD. Acetylation of Nrf2 by p300/CBP augments promoterspecific DNA binding of Nrf2 during the antioxidant response. Mol Cell Biol 2009;29:2658-72.
- [10] Kawai Y, Garduno L, Theodore M, Yang J, Arinze IJ. Acetylation-deacetylation of the transcription factor Nrf2 (nuclear factor erythroid 2-related factor 2) regulates its transcriptional activity and nucleocytoplasmic localization. J Biol Chem 2011;286:7629-40.
- [11] Davudian S, Mansoori B, Shajari N, Mohammadi A, Baradaran B. BACH1, the master regulator gene: A novel candidate target for cancer therapy. Gene 2016;588:30-7.
- [12] Afonina IS, Zhong Z, Karin M, Beyaert R. Limiting inflammation-the negative regulation of NF-[kappa]B and the NLRP3 inflammasome. Nat Immunol 2017;18:861-9.
- [13] Netea MG, Van de Veerdonk FL, van der Meer JW, Dinarello CA, Joosten LA. Inflammasome-independent regulation of IL-1-family cytokines. Annu Rev Immunol 2015;33:49-77.
- [14] Zhong Z, Sanchez-Lopez E, Karin M. Autophagy, NLRP3 inflammasome and autoinflammatory/immune diseases. Clin Exp Rheumatol 2016;34:12-6.
- [15] Liu X, Zhang X, Ding Y, Zhou W, Tao L, Lu P, Wang Y, Hu R. Nuclear factor E2-related factor-2 negatively regulates NLRP3 inflammasome activity by inhibiting reactive oxygen species-induced NLRP3 priming. Antioxid Redox Signal 2017;26:28-43.
- [16] Zhao C, Gillette DD, Li X, Zhang Z, Wen H. Nuclear factor E2-related factor-2 (Nrf2) is required for NLRP3 and AIM2 inflammasome activation. J Biol Chem 2014;289:17020-9.
- [17] Kobayashi EH, Suzuki T, Funayama R, Nagashima T, Hayashi M, Sekine H, Tanaka N, Moriguchi T, Motohashi H, Nakayama K, Yamamoto M. Nrf2 suppresses macrophage inflammatory response by blocking proinflammatory cytokine transcription. Nat Commun 2016;7:11624.

- [18] Ahmed SM, Luo L, Namani A, Wang XJ, Tang X. Nrf2 signaling pathway: Pivotal roles in inflammation. Biochim Biophys Acta 2017;1863:585-97.
- [19] Wardyn JD, Ponsford AH, Sanderson CM. Dissecting molecular cross-talk between Nrf2 and NF-KB response pathways. Biochem Soc Transact 2015;43:621-6.
- [20] Lee DF, Kuo HP, Liu M, Chou CK, Xia W, Du Y, Shen J, Chen CT, Huo L, Hsu MC, Li CW, Ding Q, Liao TL, Lai CC, Lin AC, Chang YH, Tsai SF, Li LY, Hung MC. KEAP1 E3 ligasemediated downregulation of NF-kappaB signaling by targeting IKKbeta. Mol Cell 2009;36:131-40.
- [21] Hwang YJ, Lee EW, Song J, Kim HR, Jun YC, Hwang KA. MafK positively regulates NFkappaB activity by enhancing CBP-mediated p65 acetylation. Sci Rep 2013;3:3242.
- [22] Kim W, Lee HN, Jang JH, Kim SH, Lee YH, Han YI, Ngo HK, Choi Y, Joe Y, Chung HT, Chen Y, Cha YN, Surh YJ. 15-Deoxy-(delta)12,14-prostaglandin J2 exerts pro-resolving effects through Nrf2-induced expression of CD36 and heme oxygenase-1. Antioxid Redox Signal 2017;27:1412-31.
- [23] Yoh K, Itoh K, Enomoto A, Hirayama A, Yamaguchi N, Kobayashi M, Morito N, Koyama A, Yamamoto M, Takahashi S. Nrf2-deficient female mice develop lupus-like autoimmune nephritis. Kidney Int 2001;60:1343-53.
- [24] Suzuki T, Murakami S, Biswal SS, Sakaguchi S, Harigae H, Yamamoto M, Motohashi H.
   Systemic activation of NRF2 alleviates lethal autoimmune inflammation in scurfy mice.
   Mol Cell Biol 2017;37, e00063-17.
- [25] Al-Huseini LM, Aw Yeang HX, Sethu S, Alhumeed N, Hamdam JM, Tingle Y, Djouhri L, Kitteringham N, Park BK, Goldring CE, Sathish JG. Nuclear factor-erythroid 2 (NF-E2) p45-related factor-2 (Nrf2) modulates dendritic cell immune function through regulation of p38 MAPK-cAMP-responsive element binding protein/activating transcription factor 1 signaling. J Biol Chem 2013;288:22281-8.

- [26] Sha LK, Sha W, Kuchler L, Daiber A, Giegerich AK, Weigert A, Knape T, Snodgrass R, Schroder K, Brandes RP, Brune B, von KA. Loss of Nrf2 in bone marrow-derived macrophages impairs antigen-driven CD8(+) T cell function by limiting GSH and Cys availability. Free Radic Biol Med 2015;83:77-88.
- [27] Zagorski JW, Turley AE, Freeborn RA, VanDenBerg KR, Dover HE, Kardell BR, Liby KT, Rockwell CE. Differential effects of the Nrf2 activators tBHQ and CDDO-Im on the early events of T cell activation. Biochem Pharmacol 2018;147:67-76.
- [28] Rockwell CE, Zhang M, Fields PE, Klaassen CD. Th2 skewing by activation of Nrf2 in CD4(+) T cells. J Immunol 2012;188:1630-7.
- [29] Noel S, Lee SA, Sadasivam M, Hamad ARA, Rabb H. KEAP1 editing using CRISPR/Cas9 for therapeutic NRF2 activation in primary human T lymphocytes. J Immunol 2018;200:1929-36.
- [30] George JF, Braun A, Brusko TM, Joseph R, Bolisetty S, Wasserfall CH, Atkinson MA, Agarwal A, Kapturczak MH. Suppression by CD4+CD25+ regulatory T cells is dependent on expression of heme oxygenase-1 in antigen-presenting cells. Am J Pathol 2008;173:154-60.
- [31] Roberts CA, Dickinson AK, Taams LS. The interplay between monocytes/macrophages and CD4+ T cell subsets in rheumatoid arthritis. Frontiers in Immunology 2015;6:571.
- [32] Mateen S, Moin S, Khan AQ, Zafar A, Fatima N. Increased reactive oxygen species formation and oxidative stress in rheumatoid arthritis. PLoS One 2016;11:e0152925.
- [33] Wruck CJ, Fragoulis A, Gurzynski A, Brandenburg LO, Kan YW, Chan K, Hassenpflug J, Freitag-Wolf S, Varoga D, Lippross S, Pufe T. Role of oxidative stress in rheumatoid arthritis: insights from the Nrf2-knockout mice. Ann Rheum Dis 2011;70:844-50.
- [34] van Wietmarschen H, Yuan K, Lu C, Gao P, Wang J, Xiao C, Yan X, Wang M, Schroen J, Lu A, Xu G, van der Greef J. Systems biology guided by Chinese medicine reveals new markers for sub-typing rheumatoid arthritis patients. J Clin Rheumatol 2009;15:330-7.

- [35] Maicas N, Ferrandiz ML, Brines R, Ibanez L, Cuadrado A, Koenders MI, van den Berg WB, Alcaraz MJ. Deficiency of Nrf2 accelerates the effector phase of arthritis and aggravates joint disease. Antioxid Redox Signal 2011;15:889-901.
- [36] Devesa I, Ferrandiz ML, Terencio MC, Joosten LA, van den Berg WB, Alcaraz MJ. Influence of heme oxygenase 1 modulation on the progression of murine collageninduced arthritis. Arthritis Rheum 2005;52:3230-8.
- [37] Wu WJ, Jia WW, Liu XH, Pan LL, Zhang QY, Yang D, Shen XY, Liu L, Zhu YZ. S-propargylcysteine attenuates inflammatory response in rheumatoid arthritis by modulating the Nrf2-ARE signaling pathway. Redox Biol 2016;10:157-67.
- [38] Taguchi K, Maher JM, Suzuki T, Kawatani Y, Motohashi H, Yamamoto M. Genetic analysis of cytoprotective functions supported by graded expression of Keap1. Mol Cell Biol 2010;30:3016-26.
- [39] Kong JS, Yoo SA, Kim HS, Kim HA, Yea K, Ryu SH, Chung YJ, Cho CS, Kim WU. Inhibition of synovial hyperplasia, rheumatoid T cell activation, and experimental arthritis in mice by sulforaphane, a naturally occurring isothiocyanate. Arthritis Rheum 2010;62:159-70.
- [40] Pal S and Konkimalla VB. Sulforaphane regulates phenotypic and functional switching of both induced and spontaneously differentiating human monocytes. Int Immunopharmacol 2016;35:85-98.
- [41] Fragoulis A, Laufs J, Muller S, Soppa U, Siegl S, Reiss LK, Tohidnezhad M, Rosen C, Tenbrock K, Varoga D, Lippross S, Pufe T, Wruck CJ. Sulforaphane has opposing effects on TNF-alpha stimulated and unstimulated synoviocytes. Arthritis Res Ther 2012;14:R220.
- [42] Garstkiewicz M, Strittmatter GE, Grossi S, Sand J, Fenini G, Werner S, French LE, BeerHD. Opposing effects of Nrf2 and Nrf2-activating compounds on the NLRP3

inflammasome independent of Nrf2-mediated gene expression. Eur J Immunol 2017;47:806-17.

- [43] Blewett MM, Xie J, Zaro BW, Backus KM, Altman A, Teijaro JR, Cravatt BF. Chemical proteomic map of dimethyl fumarate-sensitive cysteines in primary human T cells. Sci Signal 2016;9:rs10.
- [44] Parodi B, Rossi S, Morando S, Cordano C, Bragoni A, Motta C, Usai C, Wipke BT, Scannevin RH, Mancardi GL, Centonze D, Kerlero de RN, Uccelli A. Fumarates modulate microglia activation through a novel HCAR2 signaling pathway and rescue synaptic dysregulation in inflamed CNS. Acta Neuropathol 2015;130:279-95.
- [45] Ahmed S. Green tea polyphenol epigallocatechin 3-gallate in arthritis: progress and promise. Arthritis Res Ther 2010;12:208.
- [46] Su X, Huang Q, Chen J, Wang M, Pan H, Wang R, Zhou H, Zhou Z, Liu J, Yang F, Li T, Liu L. Calycosin suppresses expression of pro-inflammatory cytokines via the activation of p62/Nrf2-linked heme oxygenase 1 in rheumatoid arthritis synovial fibroblasts. Pharmacol Res 2016;113, Part A:695-704.
- [47] Kataoka K, Handa H, Nishizawa M. Induction of cellular antioxidative stress genes through heterodimeric transcription factor Nrf2/small Maf by antirheumatic gold(I) compounds. J Biol Chem 2001;276:34074-81.
- [48] Bretscher P, Egger J, Shamshiev A, Trotzmuller M, Kofeler H, Carreira EM, Kopf M, Freigang S. Phospholipid oxidation generates potent anti-inflammatory lipid mediators that mimic structurally related pro-resolving eicosanoids by activating Nrf2. EMBO Mol Med 2015;7:593-607.
- [49] Kawahito Y, Kondo M, Tsubouchi Y, Hashiramoto A, Bishop-Bailey D, Inoue K, Kohno M, Yamada R, Hla T, Sano H. 15-deoxy-delta(12,14)-PGJ(2) induces synoviocyte apoptosis and suppresses adjuvant-induced arthritis in rats. J Clin Invest 2000;106:189-97.

- [50] Wang H, Khor TO, Saw CL, Lin W, Wu T, Huang Y, Kong AN. Role of Nrf2 in suppressing LPS-induced inflammation in mouse peritoneal macrophages by polyunsaturated fatty acids docosahexaenoic acid and eicosapentaenoic acid. Mol Pharm 2010;7:2185-93.
- [51] Dawczynski C, Dittrich M, Neumann T, Goetze K, Welzel A, Oelzner P, Volker S, Schaible AM, Troisi F, Thomas L, Pace S, Koeberle A, Werz O, Schlattmann P, Lorkowski S, Jahreis G. Docosahexaenoic acid in the treatment of rheumatoid arthritis: A doubleblind, placebo-controlled, randomized cross-over study with microalgae vs. sunflower oil. Clin Nutr 2018;37:494-504.
- [52] Frostegard J, Svenungsson E, Wu R, Gunnarsson I, Lundberg IE, Klareskog L, Horkko S, Witztum JL. Lipid peroxidation is enhanced in patients with systemic lupus erythematosus and is associated with arterial and renal disease manifestations. Arthritis Rheum 2005;52:192-200.
- [53] Cordova EJ, Velazquez-Cruz R, Centeno F, Baca V, Orozco L. The NRF2 gene variant, -653G/A, is associated with nephritis in childhood-onset systemic lupus erythematosus. Lupus 2010;19:1237-42.
- [54] Jiang T, Tian F, Zheng H, Whitman SA, Lin Y, Zhang Z, Zhang N, Zhang DD. Nrf2 suppresses lupus nephritis through inhibition of oxidative injury and the NF-kappaBmediated inflammatory response. Kidney Int 2014;85:333-43.
- [55] Li J, Stein TD, Johnson JA. Genetic dissection of systemic autoimmune disease in Nrf2deficient mice. Physiol Genomics 2004;18:261-72.
- [56] Zhao M, Chen H, Ding Q, Xu X, Yu B, Huang Z. Nuclear Factor Erythroid 2-related Factor
  2 deficiency exacerbates lupus nephritis in B6/lpr mice by regulating Th17 Cell function. Sci Rep 2016;6:38619.
- [57] Ebihara S, Tajima H, Ono M. Nuclear factor erythroid 2-related factor 2 is a critical target for the treatment of glucocorticoid-resistant lupus nephritis. Arthritis Res Ther 2016;18:139.

- [58] Saracino AM and Orteu CH. Severe recalcitrant cutaneous manifestations in systemic lupus erythematosus successfully treated with fumaric acid esters. Br J Dermatol 2017;176:472-80.
- [59] Tsai PY, Ka SM, Chang JM, Chen HC, Shui HA, Li CY, Hua KF, Chang WL, Huang JJ, Yang SS, Chen A. Epigallocatechin-3-gallate prevents lupus nephritis development in mice via enhancing the Nrf2 antioxidant pathway and inhibiting NLRP3 inflammasome activation. Free Radic Biol Med 2011;51:744-54.
- [60] Portal-Nuñez S, Esbrit P, Alcaraz MJ, Largo R. Oxidative stress, autophagy, epigenetic changes and regulation by miRNAs as potential therapeutic targets in osteoarthritis. Biochem Pharmacol 2016;108:1-10.
- [61] Henrotin YE, Bruckner P, Pujol JP. The role of reactive oxygen species in homeostasis and degradation of cartilage. Osteoarthritis Cartilage 2003;11:747-55.
- [62] Hinoi E, Takarada T, Fujimori S, Wang L, Iemata M, Uno K, Yoneda Y. Nuclear factor E2 p45-related factor 2 negatively regulates chondrogenesis. Bone 2007;40:337-44.
- [63] Khan NM, Ahmad I, Haqqi TM. Nrf2/ARE pathway attenuates oxidative and apoptotic response in human osteoarthritis chondrocytes by activating ERK1/2/ELK1-P70S6K-P90RSK signaling axis. Free Radic Biol Med 2018;116:159-171.
- [64] Abusarah J, Benabdoune H, Shi Q, Lussier B, Martel-Pelletier J, Malo M, Fernandes JC, de Souza FP, Fahmi H, Benderdour M. Elucidating the role of protandim and 6-gingerol in protection against osteoarthritis. J Cell Biochem 2017;118:1003-13.
- [65] Cai D, Yin S, Yang J, Jiang Q, Cao W. Histone deacetylase inhibition activates Nrf2 and protects against osteoarthritis. Arthritis Res Ther 2015;17:269.
- [66] Clerigues V, Murphy CL, Guillen MI, Alcaraz MJ. Haem oxygenase-1 induction reverses the actions of interleukin-1beta on hypoxia-inducible transcription factors and human chondrocyte metabolism in hypoxia. Clin Sci (Lond) 2013;125:99-108.

- [67] Garcia-Arnandis I, Guillen MI, Castejon MA, Gomar F, Alcaraz MJ. Heme oxygenase-1 down-regulates high mobility group box 1 and matrix metalloproteinases in osteoarthritic synoviocytes. Rheumatology 2010;49:854-61.
- [68] Clerigues V, Guillen MI, Castejon MA, Gomar F, Mirabet V, Alcaraz MJ. Heme oxygenase-1 mediates protective effects on inflammatory, catabolic and senescence responses induced by interleukin-1beta in osteoarthritic osteoblasts. Biochem Pharmacol 2012;83:395-405.
- [69] Tang Q, Feng Z, Tong M, Xu J, Zheng G, Shen L, Shang P, Zhang Y, Liu H. Piceatannol inhibits the IL-1beta-induced inflammatory response in human osteoarthritic chondrocytes and ameliorates osteoarthritis in mice by activating Nrf2. Food Funct 2017;8:3926-37.
- [70] Khan NM, Haseeb A, Ansari MY, Devarapalli P, Haynie S, Haqqi TM. Wogonin, a plant derived small molecule, exerts potent anti-inflammatory and chondroprotective effects through the activation of ROS/ERK/Nrf2 signaling pathways in human Osteoarthritis chondrocytes. Free Radic Biol Med 2017;106:288-301.
- [71] Khan NM and Haqqi TM. Epigenetics in osteoarthritis: Potential of HDAC inhibitors as therapeutics. Pharmacol Res 2018;128:73-9.
- [72] Collins JA, Diekman BO, Loeser RF. Targeting aging for disease modification in osteoarthritis. Curr Opin Rheumatol 2018;30:101-7.
- [73] Davidson RK, Jupp O, de FR, Kay CD, Culley KL, Norton R, Driscoll C, Vincent TL, Donell ST, Bao Y, Clark IM. Sulforaphane represses matrix-degrading proteases and protects cartilage from destruction in vitro and in vivo. Arthritis Rheum 2013;65:3130-40.
- [74] Pietschmann P, Mechtcheriakova D, Meshcheryakova A, Foger-Samwald U, Ellinger I. Immunology of osteoporosis: a mini-review. Gerontology 2016;62:128-37.

- [75] Lippross S, Beckmann R, Streubesand N, Ayub F, Tohidnezhad M, Campbell G, Kan YW, Horst F, Sonmez TT, Varoga D, Lichte P, Jahr H, Pufe T, Wruck CJ. Nrf2 deficiency impairs fracture healing in mice. Calcif Tissue Int 2014;95:349-61.
- [76] Kanzaki H, Shinohara F, Kajiya M, Kodama T. The Keap1/Nrf2 protein axis plays a role in osteoclast differentiation by regulating intracellular reactive oxygen species signaling. J Biol Chem 2013;288:23009-20.
- [77] Hyeon S, Lee H, Yang Y, Jeong W. Nrf2 deficiency induces oxidative stress and promotes RANKL-induced osteoclast differentiation. Free Radic Biol Med 2013;65C:789-99.
- [78] Rana T, Schultz MA, Freeman ML, Biswas S. Loss of Nrf2 accelerates Ionizing radiationinduced bone loss by upregulating RANKL. Free Radic Biol Med 2012;53:2298-307.
- [79] Pellegrini GG, Cregor M, McAndrews K, Morales CC, McCabe LD, McCabe GP, Peacock M, Burr D, Weaver C, Bellido T. Nrf2 regulates mass accrual and the antioxidant endogenous response in bone differently depending on the sex and age. PLoS One 2017;12:e0171161.
- [80] Hinoi E, Fujimori S, Wang L, Hojo H, Uno K, Yoneda Y. Nrf2 negatively regulates osteoblast differentiation via interfering with Runx2-dependent transcriptional activation. J Biol Chem 2006;281:18015-24.
- [81] Park CK, Lee Y, Kim KH, Lee ZH, Joo M, Kim HH. Nrf2 is a novel regulator of bone acquisition. Bone 2014;63:36-46.
- [82] Kim JH, Singhal V, Biswal S, Thimmulappa RK, DiGirolamo DJ. Nrf2 is required for normal postnatal bone acquisition in mice. Bone Res 2014;2:14033.
- [83] Ibañez L, Ferrandiz ML, Brines R, Guede D, Cuadrado A, Alcaraz MJ. Effects of Nrf2 deficiency on bone microarchitecture in an experimental model of osteoporosis . Oxid Med Cell Longev 2014;2014:726590.

- [84] Zhang S, Wu W, Jiao G, Li C, Liu H. MiR-455-3p activates Nrf2/ARE signaling via HDAC2 and protects osteoblasts from oxidative stress. Int J Biol Macromol 2018;107:2094-101.
- [85] Yamaguchi Y, Kanzaki H, Katsumata Y, Itohiya K, Fukaya S, Miyamoto Y, Narimiya T, Wada S, Nakamura Y. Dimethyl fumarate inhibits osteoclasts via attenuation of reactive oxygen species signalling by augmented antioxidation. J Cell Mol Med 2018;22:1138-47.
- [86] Kanzaki H, Shinohara F, Itohiya-Kasuya K, Ishikawa M, Nakamura Y. Nrf2 activation attenuates both orthodontic tooth movement and relapse. J Dent Res 2015;94:787-94.
- [87] Thummuri D, Naidu VGM, Chaudhari P. Carnosic acid attenuates RANKL-induced oxidative stress and osteoclastogenesis via induction of Nrf2 and suppression of NFkappaB and MAPK signalling. J Mol Med (Berl) 2017;95:1065-76.
- [88] Ha J, Choi HS, Lee Y, Lee ZH, Kim HH. Caffeic acid phenethyl ester inhibits osteoclastogenesis by suppressing NF kappaB and downregulating NFATc1 and c-Fos. Int Immunopharmacol 2009;9:774-80.
- [89] Kanzaki H, Shinohara F, Kajiya M, Fukaya S, Miyamoto Y, Nakamura Y. Nuclear Nrf2 induction by protein transduction attenuates osteoclastogenesis. Free Radic Biol Med 2014;77:239-48.
- [90] Xia G, Li X, Zhu X, Yin X, Ding H, Qiao Y. Mangiferin protects osteoblast against oxidative damage by modulation of ERK5/Nrf2 signaling. Biochem Biophys Res Commun 2017;491:807-13.
- [91] Lin H, Gao X, Chen G, Sun J, Chu J, Jing K, Li P, Zeng R, Wei B. Indole-3-carbinol as inhibitors of glucocorticoid-induced apoptosis in osteoblastic cells through blocking ROS-mediated Nrf2 pathway. Biochem Biophys Res Commun 2015;460:422-7.

- [92] Wang Y, Liu J, Pang Q, Tao D. Alpinumisoflavone protects against glucocorticoidinduced osteoporosis through suppressing the apoptosis of osteoblastic and osteocytic cells. Biomed Pharmacother 2017;96:993-9.
- [93] Liu W, Mao L, Ji F, Chen F, Wang S, Xie Y. Icariside II activates EGFR-Akt-Nrf2 signaling and protects osteoblasts from dexamethasone. Oncotarget 2017;8:2594-603.
- [94] Kanzaki H, Shinohara F, Itohiya K, Yamaguchi Y, Katsumata Y, Matsuzawa M, Fukaya S, Miyamoto Y, Wada S, Nakamura Y. RANKL induces Bach1 nuclear import and attenuates Nrf2-mediated antioxidant enzymes, thereby augmenting intracellular reactive oxygen species signaling and osteoclastogenesis in mice. FASEB J 2017;31:781-92.
- [95] Papp D, Lenti K, Modos D, Fazekas D, Dul Z, Turei D, Foldvari-Nagy L, Nussinov R, Csermely P, Korcsmaros T. The NRF2-related interactome and regulome contain multifunctional proteins and fine-tuned autoregulatory loops. FEBS Lett 2012;586:1795-802.
- [96] Wani R, Nagata A, Murray BW. Protein redox chemistry: post-translational cysteine modifications that regulate signal transduction and drug pharmacology. Front Pharmacol 2014;5:224.
- [97] Satoh T and Lipton S. Recent advances in understanding NRF2 as a druggable target: development of pro-electrophilic and non-covalent NRF2 activators to overcome systemic side effects of electrophilic drugs like dimethyl fumarate. F1000Res 2017;6:2138.
- [98] Probst BL, McCauley L, Trevino I, Wigley WC, Ferguson DA. Cancer cell growth is differentially affected by constitutive activation of NRF2 by KEAP1 deletion and pharmacological activation of NRF2 by the synthetic triterpenoid, RTA 405. PLoS One 2015;10:e0135257.

- [99] Tu J, Zhang X, Zhu Y, Dai Y, Li N, Yang F, Zhang Q, Brann DW, Wang R. Cell-permeable peptide targeting the Nrf2-Keap1 interaction: A potential novel therapy for global cerebral ischemia. J Neurosci 2015;35:14727-39.
- [100] Jiang ZY, Lu MC, You QD. Discovery and development of Kelch-like ECH-associated protein 1. Nuclear Factor Erythroid 2-Related Factor 2 (KEAP1:NRF2) protein-protein interaction inhibitors: achievements, challenges, and future directions. J Med Chem 2016;59:10837-58.

### LEGENDS TO FIGURES

**Figure 1**. Nrf2 activation: a) basal conditions; b) activation by electrophiles or oxidative stress; c) nuclear translocation and gene transcription. ARE, antioxidant response element; GST, glutathione-S-transferase; HO-1, heme oxygenase-1; Keap-1, Kelch-like ECH-associated protein 1; MAPK, mitogen-activated protein kinase; NQO1, NAD(P)H:quinone oxidoreductase-1; Pl3K phosphoinositide 3-kinase; PKC, protein kinase C; sMaf, small musculoaponeurotic fibrosarcoma; Ub, ubiquitin.

**Figure 2**. Regulation of the immune response by Nrf2. In blue: enhancement; in red: inhibition (DC, dendritic cell; NK, natural killer cell; Th, T helper cell; Treg, T regulatory cell.

Figure 3. Structures of representative Nrf2 inducers.

**Figure 4**. Nrf2 effects in joint cells. In blue: enhancement; in red: inhibition. MMPs, matrix metalloproteinases; NO, nitric oxide; PGs, prostaglandins; PMN, polymorphonuclear leukocytes; RANKL, receptor activator of nuclear factor-κB ligand.









