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Additional Information

1 **Inactivation of the microbiota and effect on the quality**
2 **attributes of pineapple juice using a continuous flow**
3 **ultrasound-assisted supercritical CO₂ system**

4
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30 **Abstract**

31 Supercritical carbon dioxide (SC-CO₂) inactivation technology represents a promising
32 non-thermal processing method, as it causes minimum impact on the nutritional food
33 properties. The aim of this study was to analyze the combined effect of SC-CO₂ and
34 high power ultrasound (HPU) on the inactivation of natural microbiota and the quality
35 attributes of pineapple juice treated in a continuous flow system. Different juice
36 residence times (3.06-4.6 min), at 100 bar and 31.5 °C, were used. The results
37 indicated that the microbiota inactivation was complete and the differences obtained in
38 the quality attributes (2.2 % for pH, 4.8 % for °Brix, 2% for Vitamin C) were minimal.
39 During storage, microorganisms were not able to recover and the vitamin C decrease
40 could be limited to 8.2% after 4 weeks. The results demonstrated that the SC-CO₂-HPU
41 technique could be an excellent alternative for the cold pasteurization of pineapple
42 juice.

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49 **Keywords:** Non-thermal process; supercritical CO₂; ultrasound; pineapple juice; quality
50 attributes

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68 INTRODUCTION

69 Pineapple is an important tropical fruit largely consumed in the form of processed
70 products, such as juices (Costa et al., 2013). Pineapple, and consequently its juice, is
71 one of the fruits with the highest content of antioxidant and phenolic compounds. Some
72 of the phenolic compounds existing in pineapple juice are S-sinapyl-L-cysteine, N-γ-L-
73 glutamyl-S-sinapyl-L-cysteine, S-sinapyl glutathione, and p-coumaric acid. Pineapple
74 juice also contains phytosterols, such as ergostanol and stigmastanol (Ng and Hupé,
75 1998). These phytosterols lower cholesterol by reducing its absorption. Vitamin C, a
76 water-soluble vitamin abundant in pineapple juice, plays an important role in human
77 nutrition due to its high antioxidant activity. Thus, it reduces the risk of heart disease by
78 preventing the oxidation of low-density lipoprotein (LDL) cholesterol. Pineapple juice is
79 appreciated for its very pleasing aroma and flavor. It is generally drinkable in single-
80 strength, reconstituted or concentrated forms, and can be mixed with other juices to
81 develop new flavors for beverages and other products due to its strong acid flavor (De
82 Carvalho et al., 2008). Conventional thermal treatments of foods, such as
83 pasteurization, have been the first-choice method for the purposes of extending the
84 shelf-life of fruit juices. However, heating processes can affect the freshness and
85 quality of food products, which leads to consumer rejection. Non-enzymatic browning
86 reactions and pigment destruction have been found to be major causes of such
87 problems (Rattanathanarerk et al., 2005), in addition, the loss of organic acids, such as
88 vitamin C, and the decrease in phenolic compounds cause a reduction in product
89 quality (Gómez et al., 2011).

90

91 The demand for high quality processed foods, which preserve their natural and
92 fresh-like characteristics, has led to the development of non-thermal processing
93 techniques, as an alternative to conventional heat treatments (Char et al., 2010). New
94 technologies applied to foods are first and foremost of concern because of safety
95 implications. Then, as data accumulate and microbial safety can be ensured to a
96 satisfactory level, other concerns are addressed. These include quality attributes,
97 which involve the physical (color, viscosity, particle size, Brix, etc) and chemical (pH,
98 flavor volatiles and composition) properties of processed products. Enzyme activity,
99 nutritional quality and shelf life, including sensory properties, are also addressed
100 (Balaban and Ferrentino, 2012).

101 Supercritical carbon dioxide (SC-CO₂) processing is a non-thermal process capable
102 of inactivating microorganisms, at relatively moderate pressures (7.3 MPa) and at
103 temperatures (31.1 °C) low enough to avoid the thermal effects of traditional methods

104 (Benedito et al., 2015). Beyond the critical point of CO₂, the differences between liquid
105 and gaseous CO₂ no longer exist in the newly formed supercritical fluid phase, in which
106 its viscosity is lower than in the liquid state and its density and dissolving power are
107 higher than in the gaseous state. Therefore, the use of SC-CO₂ for sterilization
108 purposes is considered to be more effective than the use of CO₂ in its subcritical state
109 (Balaban and Duong, 2014; Ortuño et al., 2014). Moreover, CO₂ can be used in the
110 food industry because it is non-toxic, non-flammable, inexpensive, and GRAS status
111 (Balaban and Duong, 2014; Calvo and Torres, 2010). Although SC-CO₂ technology
112 represents an excellent non-thermal processing method, high pressures, high
113 temperatures and long treatment times are required to guarantee the safety and
114 stability of the food, especially in batch systems. In these systems, coupling with other
115 technologies, such as high-power ultrasound (HPU), might be necessary in order to
116 obtain the required lethality at shorter processing times and lower treatment intensities.
117 As a means of improving the efficiency of batch SC-CO₂ treatments, a continuous
118 system was developed by Paniagua et al. (2016), who studied the inactivation of *S.*
119 *cerevisiae* in apple juice, using the continuous flow SC-CO₂-HPU at different juice
120 residence times (3.06-9.2 min), temperatures (31-41 °C) and pressures (100-300 bars).
121 The results demonstrated that the maximum inactivation achieved by the system was
122 7.8 log-cycles. However, there is no report in the literature addressing the use of this
123 continuous technique (SC-CO₂-HPU) for other kinds of juices, nor any reference to the
124 effect of the process on the processed product quality and microbiota after the
125 treatment and subsequent refrigerated storage. Therefore, the aim of this study was to
126 determine the effect of the SC-CO₂-HPU treatment in continuous regime on both the
127 inactivation of the microbiota and the quality attributes of pineapple juice both after the
128 treatment and during refrigerated storage.

129

130 **MATERIALS AND METHODS**

131

132 **Pineapple juice**

133

134 Pineapples (*Ananas comosus L.*) were purchased from a local market and kept at 4
135 °C. Pineapple juice was obtained by washing, peeling and extracting the fruit juice
136 (Ultra Juicer, Robot Coupe J80, USA). Juice extraction took place just prior to
137 processing; consequently, an extraction was required for each experiment. Each
138 experiment required about 1.5 L of juice, 1 L was used for processing (SC-CO₂-HPU),
139 and 0.5 L served as control.

140

141

142 **SC-CO₂-HPU processing**

143

144 Laboratory continuous regime equipment was designed and built for high-power
145 ultrasound-assisted supercritical CO₂ treatment (Figure 1) (Paniagua et al., 2016).

146 The SC-CO₂-HPU process applied to the juice was as follows: first, liquid carbon
147 dioxide was supplied from the tank to the chiller reservoir. The liquid CO₂ was supplied
148 from the bottom of the chiller reservoir (which stores it at -18 °C) to the pump where it
149 was compressed at the targeted pressure. Initially, the equipment was stabilized at the
150 treatment pressure (100 bar) and temperature (31.5°C) only with SC-CO₂ at a constant
151 flow rate of 5 mL/min. Then, the ultrasound equipment was connected, and, once the
152 process conditions were attained, the sample to be treated was pumped to the mixing
153 point (7, Fig.1) where it mixed with the SC-CO₂. The mixture went into the sonication
154 vessel (8, Fig. 1), where the HPU was applied. For the experiments with HPU, the
155 power applied during the whole experiment was 40 W±5W (I=250 ±10mA; U=220 ±5 V,
156 measured with a Digital Power Meter, Yokogawa, Model WT210). Pressure and
157 temperature were kept constant during the experiment. The mixture of juice/SC-CO₂
158 exiting the treatment vessel went into the holding tube (14, Fig. 1) and, finally, into the
159 separation vessel (15, Fig. 1), where it was depressurized and the CO₂ separated from
160 the juice and recirculated to the reservoir (3, Fig. 1). Prior to each experiment, the
161 different sections of the equipment that the product flows through were cleaned and
162 sanitized with disinfectant solution (Delladet VS2, Diversey, Spain), and distilled and
163 autoclaved water. To determine the effect of the residence time on the quality
164 parameters and inactivation of the microbiota of pineapple juice, two residence times
165 (3.06 and 4.6 min) were considered. These residence times were chosen according to
166 our previous work with apple juice (Paniagua et al., 2016)., where it was observed that
167 4.6 min were enough to obtain a 6 logs reduction of *S. cerevisiae*. Residence time was
168 calculated using Eq. (1).

169

$$\tau_{TOT} = \frac{V_{SeV} + V_{SoV}}{q + q_{CO_2}} \quad (1)$$

170

171 Where (V_{SoV}) was the volume in the sonication vessel (40 mL) and the holding
172 tube volume (V_{SeV}) was 52 mL, the juice flows (q) were 15 and 25 mL/min and the
173 SC-CO₂ flow (q_{CO_2}) was 5 mL/min.

173

174 The process conditions: pressure (100 bar), temperature (31.5°C) and total
residence time (3.06 and 4.6 min) were selected from previous experiments in order to

175 attain acceptable microbial inactivation levels (Paniagua et al., 2016). Before and after
176 the treatment, the natural microbiota and the quality attributes of pineapple juice were
177 analyzed.

178 [insert Figure 1]

179 **Microbiota analysis**

180

181 The viability of mesophilic viable bacteria (MVC), yeast and *E. coli* in the juices were
182 determined by plate count. Each sample was serially diluted with sterilized distilled
183 water. 100 μ L of the appropriate dilution (10^{-1} and 10^{-2}) were plated in triplicate on LB
184 Agar, PCA Agar or YPD Agar plates and incubated for 24 h at 37 °C, 35 °C or 30 °C,
185 for *E. coli*, MVC and yeast, respectively, before counting. Results were expressed as -
186 $\log(N/N_0)$, where N_0 is the initial number of cells in the control sample and N is the
187 number of cells in the sample after the different treatments. However, in the case of
188 total inactivation, the results were expressed as $\log(N_0)$.

189

190 **pH and °Brix**

191

192 The pH of treated (TJ) and control (CJ) pineapple juice samples was measured
193 using a digital pH-meter (pH Crison 25, Spain). Soluble solids were measured using a
194 refractometer (Pocket Digital Refractometer Hand-held, Atago, Japan). Samples were
195 measured in triplicate at room temperature.

196

197 **Ascorbic acid (Vit. C)**

198

199 The ascorbic acid content of TJ and CJ samples was measured using the 2, 6
200 dichloroindophenol titrimetric method (AOAC 967.21). The ascorbic acid reduced the
201 indicator dye, 2, 6 dichloroindophenol, to a colorless solution through oxidation–
202 reduction reactions. The measurements were taken in triplicate.

203

204 **Storage of treated samples**

205

206 0.5 L of the control and treated samples were stored in glass vials and refrigerated
207 at 4°C for 4 weeks. The samples were analyzed to examine the characteristics of the
208 juice during the storage and to compare the behavior of the microbiota and vitamin C in
209 the control and treated samples. The analyses were performed at weeks 0, 1, 2, 3 and
210 4.

211

212

213 **Data analysis**

214

215 Using the statistical package, Statgraphics Centurion XVI, a multifactorial ANOVA
216 was carried out, and Tukey's test ($\alpha = 0.05$) was performed to calculate the mean
217 differences in order to evaluate the influence of the treatments used.

218

219 **RESULTS AND DISCUSSION**

220

221 ***Microbiota inactivation after the SC-CO₂-HPU treatment***

222

223 The results of the microbiota inactivation in pineapple juice are shown in Table 1.
224 When the SC-CO₂-HPU continuous treatment was applied, the total inactivation of the
225 initial microbial load of MVC, yeast and *E.coli* was obtained at the two residence times
226 employed. According to Garcia-Gonzalez et al. (2007), the fundamental step in
227 microbial inactivation by means of SC-CO₂ is its contact with the cell membrane and
228 the consequent physico-chemical modifications. The mechanisms involved in the
229 microbial inactivation using SC-CO₂ include the solubilization of CO₂ into the medium
230 where the cells are suspended, an intracellular pH decrease, key enzyme
231 inactivation/cellular metabolism inhibition due to intracellular pH lowering, a direct
232 inhibitory effect of molecular CO₂ and HCO₃⁻ on the microbial metabolism, a
233 disordering of the intracellular electrolyte balance and the removal of vital constituents
234 from cells and cell membranes.

235 [insert Table 1]

236 As for the inactivation rate, it has been seen to increase alongside temperature,
237 pressure and exposure time, where the temperature and pressure tend to act
238 synergistically on each other (Erkmen, 2003; Lin et al., 1992). The inactivation rate is
239 dependent on the initial number of cells, the type of bacterial species and the kind of
240 suspended materials. On the other hand, the organic compounds (such as
241 carbohydrates, fats and others) present in the media may increase the resistance of
242 bacteria to SC-CO₂ treatment (Balaban and Duong, 2014; Benedito et al., 2015).

243

244 In SC-CO₂-HPU treatments, the acceleration of the solubilization rate of SC-CO₂
245 into the liquid and the increase in the mass transfer due to the vigorous agitation
246 produced by the ultrasonic field would permit the rapid saturation of CO₂ in the
247 medium, which might accelerate the inactivation mechanisms (Gao et al., 2009). The
248 other possible mechanism is the cavitation produced by HPU in the liquid phase

249 (Gogate et al., 2011). Cavitation refers to the formation, growth, and implosion of tiny
250 bubbles of CO₂ or water vapor in a liquid when ultrasounds travels through it.
251 Cavitation has been proven to cause cracked or damaged cell walls, which enhances
252 the penetration of SC-CO₂ inside the cells, changing the cellular equilibrium and
253 facilitating the extraction of intracellular compounds, thus accelerating the death of the
254 microbial cells. Ortuño et al. (2014) observed that after the SC-CO₂-HPU treatment, the
255 cell wall and cell membrane were totally disrupted, thus easing the disintegration of the
256 cytoplasm and the inactivation of cells. The damage caused by the treatment was
257 serious enough to prevent a possible regrowth of cells. In the present research,
258 although the conditions of the treatment were mild (31.5 °C, 100 bar), the inactivation
259 was complete. This is due to the combined effect of both technologies that allows the
260 mechanisms explained above to obtain satisfactory results. To date, there is only one
261 work covering the use of a continuous SC-CO₂-HPU system for microbial inactivation
262 (Paniagua et al., 2016). In this previous work, apple juice was treated and only the
263 inactivation of inoculated *S. Cerevisiae* was considered. However, in the present paper
264 it has been proved that this novel technology can be used to inactivate the juice
265 microbiota, which is the real contamination in food products and its removal is a
266 condition to exploit the technology at industrial level.

267

268 **Effect of the SC-CO₂-HPU treatment on the quality attributes of pineapple juice**

269

270 The results obtained for the effect of the treatments on the quality attributes of
271 pineapple juice are shown in Table 2. The treatment caused a significant ($p < 0.05$)
272 increase in the pH values between the CJ and TJ (2.22% and 3.61% increase for 3.06
273 and 4.6 min respectively). In the case of the °Brix, results showed a slight but
274 significant ($p < 0.05$) decrease between the CJ and TJ (4.8% for both residence times).
275 As regards the pH, there was a significant effect of the treatment time on the variation
276 produced by the treatment; the longer the residence time, the greater the variation.
277 However, no effect of residence time was observed for the °Brix. Arreola et al. (1991)
278 measured the pH and °Brix of Valencia orange juice treated with SC-CO₂ at 7–34 MPa,
279 35–60°C and for 15–180 min in a batch system. They showed that there was no
280 significant ($p < 0.01$) difference between the pH and °Brix of the original juice and the
281 SC-CO₂ treated one. Kincal (2000) used a continuous-flow SC-CO₂ system for orange
282 juice treatment under pressures of 38, 72 and 107 MPa, CO₂ juice ratios of 0.40 to 1.18
283 and a residence time of 10 min. This author found no significant ($p > 0.05$) changes in
284 pH and °Brix after the treatment. Moreover, Fabroni et al. (2010) studied the orange

285 juice treated with supercritical carbon dioxide, under different treatment conditions
286 (130, 230 bar, 36 ± 1 °C, 0.385, 0.770 gCO₂/gjuice; residence time was 15 min) and no
287 statistically significant ($p > 0.05$) differences were found between the pH and °Brix of the
288 treated and control samples. However, according to the research carried out by
289 Bermúdez-Aguirre and Barbosa-Cánovas (2012), the application of thermo-sonication
290 (24 kHz, 400 W) promoted significant changes ($p < 0.05$) in the pH of the three juices
291 tested (pineapple, grape and cranberry). The main changes observed in pH by
292 Bermúdez-Aguirre and Barbosa-Cánovas (2012) were attributed to the formation of
293 certain chemical products (nitrite, hydrogen peroxide and nitrate) during the ultrasonic
294 application. Therefore, it seems that the change in the pH observed in the present
295 study brought about by the continuous SC-CO₂-HPU treatment could mainly be due to
296 the effect of ultrasound, or the combination of both techniques (SC-CO₂-HPU), rather
297 than for the single use of SC-CO₂. However, although a change in pH is observed in
298 the present study, the final values of treated juices are within the range of pH values for
299 natural pineapple juice.

300 [insert Table 2]

301 As to vitamin C, the results indicated a moderate, but significant ($p < 0.05$), decrease
302 between the CJ and TJ (Table 2). Moreover, a significant ($p < 0.05$) effect of the
303 residence time was observed, the vitamin loss only being 1.97% for a residence time of
304 3.06 min and 5.9% for 4.6 min. Fabroni et al. (2010) observed that the average vitamin
305 C content was reduced by 6.5% after the SC-CO₂ treatment (230 bar, 36 ± 1 °C, 5.08L/h
306 juice flow rate, 0.770 gCO₂/gjuice), but remained unchanged, with respect to the
307 untreated juice, after the treatments at lower pressure, regardless of the amount of CO₂
308 employed. Ascorbic acid degradation is characterized by simultaneous aerobic and
309 anaerobic reactions, the aerobic degradation being the fastest one (Ahrne et al., 1996).
310 As air content is limited in the SC-CO₂ treatment, the observed vitamin degradation
311 could be due to the effect of ultrasound in the process. Adekunte et al. (2010) observed
312 a degradation of vitamin C (32.5%) using ultrasounds (61 μm for 10 min). Rawson et al.
313 (2011) observed a reduction in ascorbic acid in watermelon juice when using
314 thermosonication. The reduction was variable, but reached 50% when the maximum
315 temperature (45°C), amplitude (61 μm) and time (10 min) were employed. This
316 behavior is mainly due to sonochemical reactions and the extreme physical conditions
317 which occur during sonication. It is known that hydrogen ions (H⁺), free radicals (O[•],
318 OH[•], HO₂[•]) and hydrogen peroxide (H₂O₂) are formed during the sonolysis of the water
319 molecules (Feril & Kondo, 2004; Pétrier et al., 2007) present in juice samples. The
320 ascorbic acid degradation during ultrasonic processing could be related to oxidation
321 reactions, promoted by the interaction with free radicals formed during sonication.

322 Hydroxyl radicals produced by cavitation may be involved in the degradation of
323 ascorbic acid. Sonication can be related to advanced oxidative processes, since both
324 pathways are associated with the production and use of hydroxyl radicals (Adekunte et
325 al., 2010). Although ultrasounds were employed in the present research, the ascorbic
326 acid degradation was minimal due to the short residence times used, the low oxygen
327 content and, possibly, to the lower cavitation energy found in a supercritical medium.
328 Another important factor in this type of juice is the effect of the treatment on its phenolic
329 content. Although previous works have shown no reduction of phenolic compounds
330 when using only SC-CO₂ (Del Pozo et al., 2006), its combination with HPU could affect
331 its content and should be matter of future research.

332

333 **Microbiota changes during refrigerated storage**

334

335 Microbial counts were performed every week for a storage period of 4 weeks at
336 4°C, chosen to emulate the temperature found during retail and household refrigerated
337 storage. The initial microbial load of control juice was 1.4 x 10⁴ CFU/mL of MVC, 5.8 x
338 10³ CFU/mL of yeast and 6.9 x 10³ of *E. coli*. After 4 weeks, the counts for the control
339 juice reached values of 9.06 x 10⁶ CFU/mL, 2.46 x 10⁶ CFU/mL and 1.17 x 10⁶
340 CFU/mL for MVC, yeast and *E. coli*, respectively. In the case of the processed juices,
341 the treatment produced an initial microbial count of zero and after the 4 weeks of
342 storage, no growth was observed. Other authors that have conducted SC-CO₂
343 inactivation treatments have found that, although just after the treatment no microbial
344 growth is found, bacterial cells are able to recover and growth appears during product
345 storage (Ortuño et al., 2014). However, in the present study, the combined treatment
346 (SC-CO₂-HPU) causes critical damage to the cells, which are not able to recover and,
347 therefore, no growth is observed during refrigerated storage. Although the non-
348 recovery of microorganisms after a SC-CO₂+HPU batch treatment was previously
349 proved for *E. coli* and *S. cerevisiae* (Ortuño et al., 2014), this is the first work that
350 demonstrates the non-viability of natural microbiota after a SC-CO₂+HPU continuous
351 treatment.

352

353 **Stability of vitamin C during refrigerated storage**

354

355 The results for vitamin C obtained during the 4 weeks of refrigerated storage are
356 shown in Table 3. At the beginning of the storage, the content of vitamin C in the
357 treated juice was 33.4 and 34.8 ppm for 3.06 and 4.6 min of residence time,
358 respectively. This value was slightly lower than that found in control (untreated)

359 pineapple juice (35.51 ppm Vit. C / 100 mL). The vitamin C content of the 3 different
360 samples studied significantly ($p < 0.05$) decreased during the 4 weeks of storage. The
361 greatest decrease corresponded to the SC-CO₂-HPU treated sample with a residence
362 time of 3.06 min (19.5% variation) and the smallest to the control juice (6%). No
363 change in the vitamin content was observed after 2 weeks of refrigerated storage in the
364 case of the SC-CO₂-HPU treated samples with a residence time of 4.6 min; this shows
365 that, although the initial inactivation is greater for this residence time, it provides better
366 results during storage. Choi et al. (2002), studied ascorbic acid retention in blood
367 orange juice during refrigerated storage. These authors observed that ascorbic acid
368 decreased gradually as storage time progressed; more than 50% was lost within 3
369 weeks of storage and was completely degraded after 5 weeks of storage in the case of
370 the natural juice. The decrease in vitamin C content during storage was also observed
371 by Klimczak et al. (2007), who found that 19 % of the ascorbic acid in natural orange
372 juice degraded while refrigerated for 6 weeks. Piljac-Zegarac et al. (2009) studied the
373 ascorbic acid degradation of six dark fruit juices (blackcurrant, cranberry, blueberry,
374 pomegranate, strawberry and cherry). The juice samples were separated into two
375 groups, with strawberry, blackcurrant and cherry exhibiting high vitamin C content and
376 cranberry, pomegranate, and blueberry exhibiting moderate-low vitamin C content.
377 During refrigerated storage, the vitamin C content in blueberry juice dropped to zero
378 after 7 days of storage, and reduced to 50% of the initial value within the first 74 h.
379 Vitamin C showed better stability in the other juices; in the cases of pomegranate and
380 cranberry, it dropped to zero after 9 days in refrigerated storage, while it showed a
381 gradual, but steady, decline in cherry and strawberry juices. By day 28, the vitamin C
382 content in strawberry and cherry juices dropped to 58% and 35% of the initial values,
383 respectively. According to the literature, the decrease in the vitamin C content in the
384 juice during storage is dependent on the storage conditions, such as temperature,
385 oxygen and light access. Oxygen is usually mainly responsible for the loss in vitamin C
386 during storage. In this regard, the significant vitamin C reduction in the present study
387 might be due to the presence of oxygen in the head-space of the glass bottle. Vitamin
388 C retention has been used as an indicator of fruit juice shelf-life. It has been accepted
389 that the shelf-life of fruit juice could be determined by a 50% loss or the half-life of the
390 vitamin C (Odriozola-Serrano et al., 2008; Laorko et al., 2013). Therefore, according to
391 Table 3, none of the juices reached their shelf-life after the 4 weeks of refrigerated
392 storage. Moreover, the lower reduction of vitamin C during storage (4 weeks) of the
393 juice treated with a residence time of 4.6 min (8.16% reduction), indicates that this
394 treatment would provide a higher quality of the juice, compared to the juice treated for
395 3.06 min.

396

397 [insert Table 3]

398

399

400 **CONCLUSIONS**

401

402 The results demonstrated the potential of the continuous SC-CO₂-HPU inactivation
403 technique. The microbiota was completely inactivated using mild process conditions.
404 The changes in the different quality attributes provoked by the treatment are minimal
405 and the final values are within normal ranges for natural pineapple juice. On the other
406 hand, the storage results showed that no microbial growth/recovery is observed and
407 that minimal reductions in vitamin C are found. **Further studies should consider the**
408 **effect of the treatment on the sensory attributes, especially flavor, of the processed**
409 **juice.** Thus, the use of mild process conditions could lead to an increase in the quality
410 of the product treated using this technique.

411

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413

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415 in the development of the equipment.

416

417 **DECLARATION OF CONFLICTING INTERESTS**

418

419 The authors declare that there is no conflict of interest.

420

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427

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Highlights

- Ultrasound-assisted supercritical CO₂ treatment completely inactivated juice microorganisms
- By shortening residence time, juice quality characteristics were improved
- Microorganisms were not able to recover during refrigerated storage
- Changes in vitamin C during storage were minimal
- The innovative technology presented employs mild process conditions

Fig. 1. Supercritical CO₂ continuous treatment system. 1. CO₂ tank; 2. N₂ tank; 3. Chiller reservoir; 4. CO₂ Pump; 5. Liquid reservoir; 6. Liquid Pump; 7. Mixing point; 8. Sonication vessel; 9. Sonotrode; 10. Insulation joint; 11. Ceramics; 12. Power generation unit; 13. Thermostatic bath; 14. Continuous contact tube; 15. Separation vessel; 16. Treated sample; 17. CO₂ Recirculation; 18. Sonication vessel output, 19. Separation vessel output, V. valve; VS. non-return valve; VM. micrometric valve; P. Manometer; T. temperature sensor.

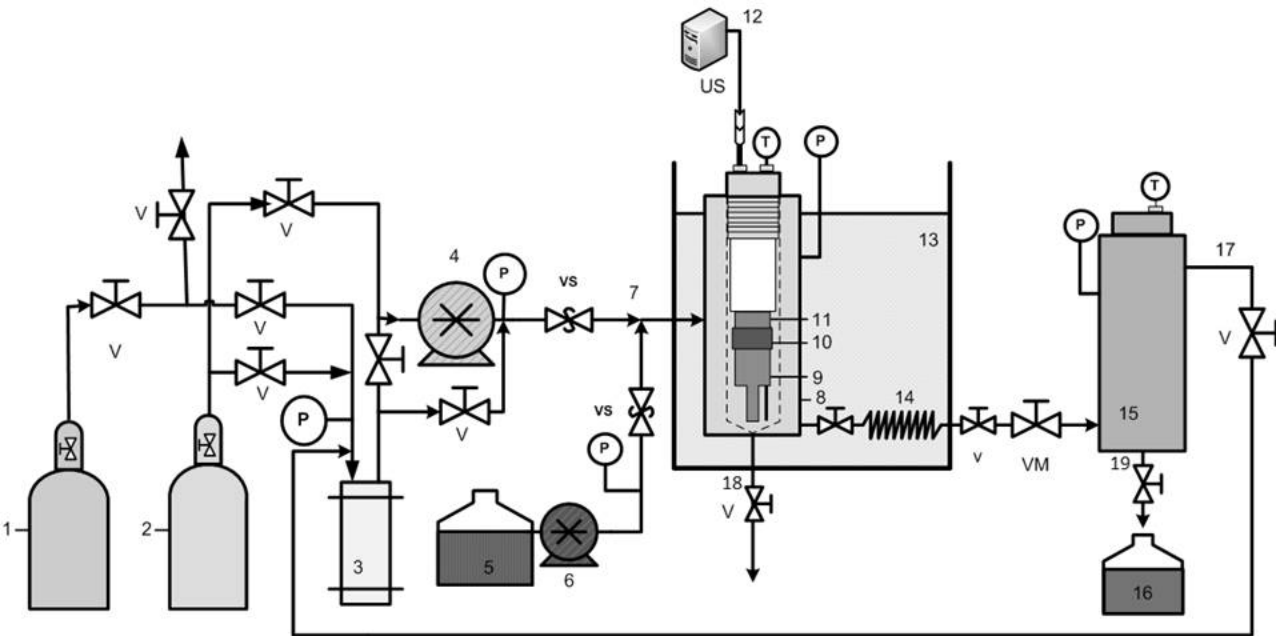


Table 1. Inactivation of the microbiota in pineapple juice at different residence times (3.06 and 4.6 min)

		Treatment/Conditions	
		100 bar, 31 °C 3.06 min	100 bar, 31 °C 4.6 min
MVC	N ₀ (CFU/mL)	1.40x10 ⁴	1.40x10 ⁴
	N (CFU/mL)	0	0
	log (N ₀)	4.15	4.15
Yeast	N ₀ (CFU/mL)	5.80x10 ³	5.80x10 ³
	N (CFU/mL)	0	0
	log (N ₀)	3.76	3.76
<i>E. coli</i>	N ₀ (CFU/mL)	6.90x10 ³	6.90x10 ³
	N (CFU/mL)	0	0
	log (N ₀)	3.84	3.84

MVC: Mesophilic viable bacteria

Table 2. Effect of the SC-CO₂-HPU continuous treatment on pH, °Brix, and Vit. C for different juice residence times (3.06 and 4.6 min).

		Treatment/Conditions	
		100 bar, 31 °C 3.06 min	100 bar, 31 °C 4.6 min
pH	Control (CJ)	3.6±0.01 ^c	3.6±0.01 ^c
	Treated (TJ)	3.68±0.01 ^a	3.73±0.01 ^b
	Variation	2.22%	3.61%
° Brix	Control (CJ)	12.5±0.11 ^c	12.5±0.11 ^c
	Treated (TJ)	11.90±0.10 ^a	11.93±0.09 ^a
	Variation	-4.80%	-4.80%
Vit. C (ppm)	Control (CJ)	35.50±0.18 ^c	35.5±0.18 ^c
	Treated (TJ)	34.82±0.07 ^a	33.45±0.13 ^b
	Variation	-1.97%	-5.90%

Different letters indicate significant differences (p <0.05)

Table 3. Analysis of vitamin C during refrigerated storage (4 weeks) of SC-CO₂-HPU treated (at two different residence times) and control pineapple juices

Week	Control		Treated (residence time 3.06 min)		Treated (residence time 4.6 min)	
	ppm Vit. C	% of Variation	ppm Vit. C	% of Variation	ppm Vit.C	% of Variation
0	35.51± 0.01 ^a		33.4±0.10 ^a		34.8±0.22 ^a	
1	35.51± 0.01 ^a	0	31.81±0.08 ^b	-4.76	34.8±0.15 ^a	0
2	34.8± 0.09 ^b	-1.99	30.2±0.12 ^c	-9.58	34.8±0.08 ^a	0
3	34.51±0.05 ^c	-2.81	29.5±0.17 ^d	-11.67	33.38±0.05 ^b	-4.08
4	33.38±0.04 ^d	-5.99	26.9±1.02 ^e	-19.46	31.96±0.09 ^c	-8.16

Different letters within a column indicate significant differences (p <0.05)