

Document downloaded from:

<http://hdl.handle.net/10251/147419>

This paper must be cited as:

Paniagua-Martínez, I.; Mulet Pons, A.; García Alvarado, MÁ.; Benedito Fort, JJ. (2018). Orange juice processing using a continuous flow ultrasound-assisted supercritical CO₂ system: Microbiota inactivation and product quality. *Innovative Food Science & Emerging Technologies*. 47:362-370. <https://doi.org/10.1016/j.ifset.2018.03.024>



The final publication is available at

<https://doi.org/10.1016/j.ifset.2018.03.024>

Copyright Elsevier

Additional Information

1
2 **Orange juice processing using a continuous flow ultrasound-**
3 **assisted supercritical CO₂ system: microbiota inactivation and**
4 **product quality**
5

6
7
8 Paniagua-Martínez, I.^{1,3}, Mulet, A.¹, García-Alvarado, M.A.² Benedito, J.*¹
9

10 1: Food Technology Department. Universitat Politècnica de València. Camino de Vera
11 s/n, Valencia, Spain.

12 2: Chemical and Biochemical Engineering Department. Instituto Tecnológico de
13 Veracruz. Av. Miguel A. de Quevedo 2779. Veracruz, Mexico.

14 3: Universidad Veracruzana, Facultad de Bioanálisis, Carmen Serdán e Iturbide S/N,
15 col. Flores Magón 91700, Veracruz, Veracruz, México.
16
17
18
19
20
21
22
23
24
25
26
27
28

29 *Corresponding author: Grupo de Análisis y Simulación de Procesos Agroalimentarios,
30 Departamento Tecnología de Alimentos, Universitat Politècnica de València, Camí de
31 Vera s/n, E46022, Valencia, Spain. Tel.: +34-96-3879147 Fax:+34-96-3879839.

32 E-mail address: jjbenedi@tal.upv.es (J. Benedito)

33 **Abstract**

34 The feasibility of using supercritical CO₂ assisted by ultrasound (SC-CO₂-HPU)
35 in continuous mode (3.06 min residence time) for the non-thermal pasteurization of
36 orange juice was evaluated. The proposed technology was effective for microbial
37 inactivation; complete inactivation was obtained for *E. coli* and total aerobic mesophilic
38 bacteria while 99.7% reduction for *S.cerevisiae*. Results showed that the SC-CO₂-HPU
39 treatment brought about small changes in the pH, °Brix and titratable acidity of the juice.
40 Furthermore, although SC-CO₂-HPU technology produced a higher browning index
41 (211%) and greater changes in color, it was possible to improve the cloud of juice by
42 173%; what is more, a smaller percentage of phenolic compounds (6.5%) and ascorbic
43 acid (5.5%) was lost compared to the thermally pasteurized juice (10 % decrease in both
44 parameters). Moreover, the antioxidant capacity could be increased (12%) with respect
45 to the natural juice. Therefore, SC-CO₂-HPU technology appears to be effective for
46 microbial pasteurization and the mild process conditions used could lead to an increase
47 in the juice quality.

48

49

50 **Industrial relevance**

51 The demand for high quality processed foods which preserve their natural and fresh-like
52 characteristics has awakened a growing interest in non-thermal technologies. The
53 ultrasound-assisted SC-CO₂ continuous system is an innovative non-thermal technology
54 that could represent a development in the area of emerging technologies. This
55 technology allows high quality products to be obtained by preserving their natural
56 bioactive compound content while maintaining their fresh-like organoleptic
57 characteristics. In fact, food experts working in academia, industry or governmental
58 agencies worldwide foresee that non-thermal emerging technologies will be among the
59 most impactful novel food processing technologies for the next decade in terms of
60 product commercialization.

61

62 **Keywords:** Non-thermal process, supercritical CO₂, ultrasound, continuous regime,
63 orange juice

64

65

66

67

68

69

70 **1. Introduction**

71

72 In recent years, while developed countries have witnessed a rise in the consumption
73 of processed fruit juices, that of fresh citrus fruit has been on the wane (Tiwari, O'Donnell,
74 Muthukumarappan, & Cullen, 2009a). Worldwide, orange juice is a very popular product
75 due to its high nutritional value, its bioactive components, such as phenolic compounds,
76 vitamin C and carotenoids, and its sensory characteristics (Ortuño, Balaban, & Benedito,
77 2014).

78 Despite its low pH, this juice needs to be processed because it is of limited stability
79 due to microbial growth and enzyme activity, which can cause unpleasant organoleptic
80 changes or the degradation of compounds during storage (Fabroni, Amenta, Timpanaro,
81 & Rapisarda, 2010; Ferrentino, Plaza, Ramirez-Rodrigues, Ferrari, & Balaban, 2009;
82 Khandpur, & Gogate, 2016 ; Liu et al, 2010; Zinoviadou et al, 2015).

83 Although thermal pasteurization remains the most commonly-used method for the
84 preservation of juices, there is growing interest in developing alternative techniques. The
85 new techniques are expected to minimize changes in the nutritional and organoleptic
86 characteristics of food, obtaining fresher and richer juices than traditional thermal
87 technology. Two such techniques are high hydrostatic pressure (HHP) and pulsed
88 electric fields (PEF), which result in better quality retention and adequate shelf life;
89 however, they cannot inactivate enzymes, such as PME, well enough to produce a shelf-
90 stable juice, unless they are combined with elevated temperatures. In addition, these
91 new technologies involve high investment and operational costs, which is an important
92 obstacle to their industrial application (Niu et al, 2010; Ozuna, Paniagua-Martínez,
93 Castaño-Tostado, Ozimek, & Amaya-Llano, 2015; Tiwari, Muthukumarappan, O'donnell,
94 & Cullen, 2009b; Vervoort et al., 2011). Moreover, at present, HHP processing consists
95 of batch processes, which limits its use because of its low processing capacity (Damar
96 & Balaban, 2006).

97 For the purposes of processing large volumes of liquid food, such as orange juice, a
98 continuous preservation process is more desirable. This objective can be attained by
99 applying supercritical fluids, a non-thermal preservation technique in which both CO₂ and
100 the product are pumped through the system by high-pressure pumps, mixed and
101 maintained in contact for a period of time (Fabroni et al., 2010; Paniagua-Martínez,
102 Mulet, García-Alvarado, & Benedito, 2016).

103 Supercritical CO₂ (SC-CO₂) has a density close to that of liquids, as well as gas
104 properties like high diffusivity and low viscosity; therefore, it has excellent transport
105 properties. Furthermore, these properties can be controlled by temperature and pressure

106 changes (Calix, Ferrentino, & Balaban, 2008; Niu et al, 2010.; Wimmer & Zarevúcka,
107 2010). Supercritical CO₂ is considered an excellent alternative to solvents because of its
108 non-toxic and non-flammable nature and its relatively low critical pressure and
109 temperature (73.6 bar, 31.0 °C). Moreover, the SC-CO₂ has a lethal effect on bacteria
110 (Garcia González et al., 2007). This effect is directly proportional to the applied pressure,
111 time and temperature. SC-CO₂ acts on bacteria as follows: first, solubilization occurs in
112 the external liquid phase, causing carbonic acid formation (which dissociates into
113 bicarbonate and hydrogen ions); therefore, it increases cell membrane fluidity and
114 permeability, increasing the diffusion of CO₂ into the cell and causing a decrease in
115 intracellular pH. Thus, the inactivation/inhibition of key cellular metabolic enzymes for
116 microorganisms occurs. As a result, a disorder in the electrolyte balance of intracellular
117 constituents is produced and vital constituents of cells and cell membranes are extracted
118 (Fabroni et al., 2010; Garcia González et al., 2007; Kincal et al., 2005; Ortuño, Martínez-
119 Pastor, Mulet, & Benedito, 2013; Paniagua et al., 2016).

120 Despite all the aforementioned advantages of SC-CO₂ inactivation, even the
121 continuous systems require long treatment times and high pressures and temperatures
122 (Fabroni et al., 2010; Kincal et al., 2005) to ensure the safety and stability of food, limiting
123 the efficiency of the inactivation process, compromising the food quality and increasing
124 processing costs. In this sense, there is growing interest in process intensification, with
125 the simultaneous application of different non-thermal technologies, in the search for
126 synergistic effects. One of the techniques that synergistically improves the inactivation
127 mechanisms of SC-CO₂ is high power ultrasound (HPU), which accelerates and
128 improves heat and mass transfer processes (Ortuño et al., 2013, 2014; Paniagua et al.,
129 2016).

130 When high power ultrasound propagates in a liquid, cavitation bubbles are generated
131 by pressure changes. These microbubbles collapse violently in the succeeding
132 compression cycles of a propagated sonic wave. This results in localized high
133 temperatures, pressures and significant shearing effects. Consequently, the intense
134 local energy and high pressure bring about a localized pasteurization effect (without
135 causing significant temperature increases, while shortening processing time and cutting
136 energy consumption) (Abid et al., 2013; Tiwari, Mu hukuma appan, O'Donnell, & Cullen,
137 2008a; Tiwari et al., 2009). Therefore, with the combination of SC-CO₂ and HPU (SC-
138 CO₂-HPU), an increase is produced both in the solubilization rate of SC-CO₂ in the liquid
139 and in the mass transfer due to the vigorous stirring produced by the ultrasonic field.
140 Thereby, a quick saturation of CO₂ in the medium is achieved, as well as the
141 intensification of the inactivation mechanisms. Furthermore, cavitation and agitation
142 produced by the HPU cause cell wall damage, increasing the SC-CO₂ penetration, the

143 intracellular compound extraction and the death of microbial cells. In addition, thermal,
144 chemical and mechanical effects induced by HPU cavitation contribute to enzyme
145 inactivation (Tiwari et al., 2008a). The combined use of use of SC-CO₂ and HPU can be
146 considered as a green processing technique since it can contribute to the reduction of
147 energy and waste, the increase of the product quality and safety and the decrease of the
148 carbon and water footprint (Chemat et al., 2017).

149 Ortuño, Martínez-Pastor, Mulet, and Benedito (2012) reported that by using a batch-
150 mode SC-CO₂ at 350 bar and 36 °C for 25 min, a reduction of 1 log-cycle in *Escherichia*
151 *coli* DH1 (*E. coli*) was obtained in orange juice. However, Kincal et al. (2005) reported
152 that a continuous SC-CO₂ treatment (210 bars, 34.5 °C, 10 min residence time) caused
153 at least a 5 log-cycle reduction in pathogens (*E. coli* O157: H7, *Salmonella Typhimurium*
154 and *Listeria monocytogenes*). Consequently, it can be expected that batch-mode
155 equipment requires a much longer inactivation time compared to continuous SC-CO₂
156 systems. There are a few studies of batch-mode SC-CO₂ intensified using ultrasound
157 (SC-CO₂-HPU); two of them prove the complete inactivation of the *E.coli* and
158 *S.cerevisiae* population in orange juice after 1.5 min (225 bar, 36 °C) and 5 min (350 bar,
159 36 °C) of treatment, respectively (Ortuño et al., 2012, 2013). In order to improve the
160 efficiency of batch SC-CO₂ treatments, a continuous system was developed by Paniagua
161 et al. (2016) who studied the inactivation of *S. cerevisiae* in apple juice, using the
162 continuous flow SC-CO₂-HPU at different juice residence times (3.06-9.2 min),
163 temperatures (31-41 °C) and pressures (100-300 bars). The results demonstrated that
164 the maximum inactivation achieved by the system was 7.8 log-cycles. However, there
165 are no studies covering either the use of this continuous technique (SC-CO₂-HPU) for
166 other types of juices or the effect of the process on the product quality. Therefore, the
167 aim of this study was to determine the effect of SC-CO₂-HPU treatment in a continuous
168 regime on both the inactivation of the microbiota and the quality attributes of orange
169 juice.

170

171 **2. Materials and methods**

172

173 **2.1. Orange juice**

174 Valencia Navel oranges (*Citrus sinensis*) were purchased from a local market and
175 kept at 4 °C for 2 days until juice extraction. Orange juice was obtained by washing,
176 peeling and extracting the fruit juice (Ultra Juicer, Robot Coupe J80, USA). Juice
177 extraction took place just prior to the treatment application; consequently, an extraction
178 was required for each experiment. Each experiment required about 1.5 L of juice, 1 L
179 was used for processing (SC-CO₂-HPU and thermal pasteurization), and 0.5 L served

180 as control. Juices were not inoculated and only the inactivation of the microbiota was
181 considered.

182

183 2.2. SC-CO₂-HPU processing

184 Laboratory continuous regime equipment was designed and built for supercritical CO₂
185 assisted by high power ultrasound (SC-CO₂-HPU) (Figure 1) (Paniagua et al., 2016).

186 The SC-CO₂-HPU process applied to the juice was as follows: first, liquid carbon
187 dioxide was supplied from the tank to the chiller reservoir in which it was compressed to
188 200 bar by means of the injection of pressurized gaseous N₂. The liquid CO₂ was
189 supplied from the bottom of the chiller reservoir (which stores it at -18 °C) to the pump
190 where it was compressed at the target pressure. The equipment was stabilized at the
191 treatment pressure (P) and temperature (T) by flowing SC-CO₂ at a constant flow rate
192 of 5 mL/min. Thereafter, the ultrasound equipment was connected, and once the process
193 conditions (P, T) were attained, the sample to be treated was pumped to the mixing
194 point (7, Fig.1) where it mixed with the SC-CO₂. The mixture went into the sonication
195 vessel (8, Fig. 1), where the HPU was applied. For the experiments with HPU, the power
196 applied during the whole experiment was 40 W±5W ($I=250 \pm 10\text{mA}$; $U=220 \pm 5 \text{V}$,
197 measured with a Digital Power Meter, Yokogawa, Model WT210). Pressure and
198 temperature were kept constant during the experiment. The mixture of juice/SC-CO₂
199 exiting the sonication vessel went into the holding tube (14, Fig. 1) and, finally, into the
200 separation vessel (15, Fig. 1), where it was depressurized and the CO₂ separated from
201 the juice and recirculated to the reservoir (3, Fig. 1). Prior to each experiment, the
202 different sections of the equipment through which the product flows were cleaned and
203 sanitized with disinfectant solution (Delladet VS2, Diversey, Spain) and distilled and
204 autoclaved water. To determine the effect of temperature on both the quality parameters
205 and on the inactivation of the microbiota of orange juice, samples (0.5 L) were treated
206 by SC-CO₂-HPU in a continuous system at 100 bar and different temperatures (31, 36
207 and 41 °C). The pressure and temperature conditions were selected according to
208 Paniagua et al. (2016), taking into account that low pressures reduce the operating costs
209 while maintaining an acceptable microbial inactivation. The flow rate of juice was 25
210 mL/min and the residence time 3.06 min. The process conditions were selected from
211 previous experiments in order to attain adequate inactivation levels. All the experiments
212 were run in triplicate.

213

214 2.3. Heat treatment

215 To evaluate the effect of conventional thermal treatment on the quality parameters
216 and microbiota inactivation of orange juice, the juice was pasteurized (PASC Computer
217 Controlled Laboratory pasteurizer, EDIBON, Spain) at 90 °C for 1 minute. For this
218 purpose, the juice was placed in a feed tank, driven by a pump to a plate heat exchanger,
219 rapidly heated to the desired temperature and taken to the holding tube where it
220 remained throughout the processing time. After the treatment, the juice was cooled
221 rapidly in a water bath (4 °C). Experiments were run in triplicate. Thus, it was possible to
222 compare the SC-CO₂-HPU processing results (quality and microbiology) with those of
223 the conventional heat treatment.

224

225 2.4. Microbiota analysis

226 The viability of *E. coli*, total aerobic mesophilic and *S. cerevisiae* in the orange juice
227 samples was determined by the plate count method to evaluate the effect of both
228 treatments (SC-CO₂-HPU in continuous system and thermal pasteurization) on the
229 microbiota of orange juice. Each sample was serially diluted with sterilized distilled water.
230 100 µL of the appropriate dilution (10⁻¹ and 10⁻²) were plated in triplicate on LB Agar,
231 PCA Agar or YPD Agar plates and incubated for 24 h at 37 °C, 35 °C or 30 °C, for *E.*
232 *coli*, total aerobic mesophilic or *S. cerevisiae*, respectively, before counting. Results were
233 expressed as -log(N/N₀), where N₀ is the initial number of cells in the control sample and
234 N is the number of cells in the sample after the different treatments. When the total
235 microbial inactivation was achieved, results were expressed as log(N₀).

236

237 2.5. Physico-chemical analysis of orange juice

238 All the physico-chemical measurements were taken in triplicate.

239 2.5.1. pH and °BRIX

240 The pH of treated and untreated orange juice samples was measured using a digital
241 pH-meter (pH Crison 25, Spain). Samples were measured in triplicate at room
242 temperature.

243 Soluble solids were measured using a refractometer (Pocket Digital Refractometer
244 Hand-held, Atago, Japan). Measurements were taken in triplicate at room temperature.

245

246

247 2.5.2. Titratable acidity

248 Titratable acidity was measured using the method described by Kincal et al. (2006),
249 using NaOH 0.1 N. Results were obtained in triplicate and expressed as grams of citric
250 acid per 100 mL of juice.

251

252 2.5.3. Phenolic compounds

253 Total phenolic compounds were determined by the method described by Gao,
254 Ohlander, Jeppsson, Björk, and Trajkovski (2000) applying 1:3 dilution factor of the
255 samples. The quantification of the phenolic compounds with respect to a standard curve
256 of gallic acid with concentrations between 110.4 and 552 ppm was performed. Results
257 were expressed as ppm equivalent of gallic acid.

258

259 2.5.4. Antioxidant capacity (FRAP)

260 Antioxidant capacity was assessed by the method described by Pulido, Bravo, and
261 Saura-Calixto (2000) using the FRAP reagent and applying 1:20 dilution factor of the
262 samples. To obtain the results, a calibration curve of Trolox with concentrations between
263 50 and 750 μM was built, plotting the concentration of Trolox versus absorbance at 30
264 minutes. The antioxidant capacity of samples at 30 minutes with the FRAP reagent was
265 expressed as the equivalent Trolox concentration at 30 minutes.

266

267 2.5.5. Browning index

268 Browning index was used to discover the effect of treatments on juice browning. For
269 this purpose, a spectrophotometric method was used after centrifuging and filtering the
270 samples. This method is described by Xu et al. (2011). In the present study, however,
271 the centrifugation time was 10 minutes and the angular velocity 12600 rpm.

272

273 2.5.6. Color

274 Color was measured using a colorimeter (Spectrophotometer CM-2500d, Konica
275 Minolta, Japan) based on the L^* , a^* , b^* color coordinates (Kincal et al., 2006; Ferrentino
276 et al, 2009). Color measurements were taken in triplicate.

277 The total color difference (ΔE) was determined from Equation 1, which indicates the
278 magnitude of the color change after treatment.

279

$$280 \quad \Delta E = [(L-L_0)^2 + (a-a_0)^2 + (b-b_0)^2]^{1/2} \quad (1)$$

281

282 where: L_0 , a_0 and b_0 are the color values of untreated juice and L , a and b those of the
283 treated samples. Differences in perceivable color can be classified as very different (ΔE
284 > 3), different ($1,5 < \Delta E < 3$) and slightly different ($\Delta E < 1,5$) (Tiwari, Muthukumarappan,
285 O'donnell, & Cullen, 2008b).

286

287 2.5.7. Cloud

288 To evaluate the loss of cloud or juice clarification after treatment, a spectrophotometric
289 method was used after sample centrifugation, as described by Ferrentino et al. (2009).

290

291 Absorbance was recorded as the cloud value with distilled water used as blank. The
292 percentage of cloud change was calculated by Equation 2.

293

294

$$295 \quad \text{Percentage cloud change} = \frac{\text{final cloud value} - \text{initial cloud value}}{\text{initial cloud value}} \cdot 100 \quad (2)$$

296

2.5.8. Ascorbic acid

297

298 The ascorbic acid content was measured using the 2, 6 dichloroindophenol titrimetric
299 method (AOAC 967.21). The ascorbic acid reduced the indicator dye, 2, 6
dichloroindophenol, to a colorless solution through oxidation–reduction reactions.

300

301 2.6. Statistical Analysis

302

303 Using the statistical package, Statgraphics Centurion XVI, a multifactorial ANOVA
304 was carried out, and LSD (Least Significant Differences) were identified in order to
305 evaluate the influence of the treatments considered (Ortuño et al., 2012).

306

307 3. Results and discussions

308

309 3.1. Microbiota inactivation after the SC-CO₂-HPU treatment

310

311 The inactivation of the microbiota of orange juice is shown in Table 1. After the SC-
312 CO₂-HPU treatment was applied, the total inactivation of the initial microbial load of *E.coli*
313 and total aerobic mesophilic bacteria was measured at the different temperatures
314 employed. However, the initial population of *S.cerevisiae* could not be completely
315 inactivated, obtaining levels of inactivation of 2.60, 2.24 and 2.19 log cycles at 31, 36
316 and 41 °C, respectively, which corresponds to average reductions of 99.7, 99.4 and
317 99.3%, respectively. The use of different temperatures produced no significant
318 differences ($p > 0.05$) in the level of *S.cerevisiae* inactivation. The difficulty of achieving
319 the complete inactivation of *S. cerevisiae* could be related with its thicker cell wall, which
320 measures 124.8 nm, in comparison with *E.coli*, which is 17.7 nm but also mainly to the
321 different structure of bacteria and yeast (Ortuño et al., 2014). In a similar way to the
322 results of the present study, Ortuño et al. (2014) obtained a reduction of 7 and 4 log units
323 in *E.coli* and *S.cerevisiae*, respectively, starting from the same initial cell concentration,

324 for a treatment of orange juice with SC-CO₂-HPU (225 bar; 31°C; 6 min). In the same
325 way, but using SC-CO₂-HPU in continuous system (200 bar, 36°C), Paniagua-Martínez
326 et al. (2016) obtained reductions of 6.8 log cycles in *S. cerevisiae* in 3.1 min of residence
327 time using apple juice as model medium. Fabroni et al. (2010), studied the effect of
328 continuous SC-CO₂ (130-230 bar, 5.08 L/h juice flow rate, 1.96-3.91 L/h of CO₂ flow rate
329 and 15 min of residence time) on the inactivation of total aerobic mesophilic bacteria and
330 yeast population in blood orange juice, obtaining reductions of 3 log cycles for each type
331 of microorganism.

332 On the other hand, the thermal pasteurization treatment attained the complete
333 inactivation of the assessed microbiota.

334

335 3.2. Effect of the SC-CO₂-HPU treatment on the physico-chemical properties of orange
336 juice

337

338 3.2.1. pH, °Brix, Titratable acidity (TA)

339 The results of pH, °Brix and TA are shown in Table 2. The continuous treatment of
340 orange juice using SC-CO₂-HPU had a non-significant ($p>0.05$) effect on the pH of the
341 juice, similarly to what happens in the case of the thermal treatment (Table 2). No
342 significant ($p<0.05$) differences were observed between the pH of the juice after the SC-
343 CO₂-HPU or the pasteurization treatments. This could be due to the short treatment time
344 and to the low initial pH value of the juice. In this regard, for pH values of 3.7-3.8, the
345 dissociation of the carbonic acid formed by the dissolution of the CO₂ into the juice is
346 difficult, due to the high dissociation constants of carbonic acid and the bicarbonate
347 ($pK_a=6.57$ and $pK_a=10.62$, respectively) (Zhou, Wang, Hu, Wu, & Liao, 2009). Kincal et
348 al. (2006) observed a change of between 0.14% and 0.54% in the pH of orange juice
349 treated with a continuous SC-CO₂ process (380, 720 and 1070 bar; 0.40-1.18 ratio
350 CO₂/juice; 40°C; 10 min). Fabroni et al. (2010) observed an increase in the pH of orange
351 juice of around 1.47% after a treatment with a continuous SC-CO₂ process (230 bar; 5.08
352 L/h juice; 3.91 L/h CO₂; 36°C; 15 min), as well as a percentage of 1.18% after thermal
353 pasteurization. As can be observed from Table 2, the range of pH values of the control
354 samples used in the different experiments comprises the range of the pH of the treated
355 ones; this points to the scarce impact of the treatment on this quality attribute, the natural
356 variability being more noticeable than the possible effect of the treatment.

357

358 In the case of the °Brix, the results obtained showed a slight decrease at 31 °C (-
359 0.81%), 36 °C (-1.74%) and 41 °C (-1.41%), although non-significant ($p<0.05$)
360 differences between the control and processed juice samples were observed for any

361 treatment. Gasperi et al. (2009) studied the use of a batch SC-CO₂ treatment (100 bar;
362 36 °C; 10 min) on apple juice, and found a reduction percentage of 0.85%. Kincal et al.
363 (2006) obtained reductions of approximately 1.80% after a continuous SC-CO₂ treatment
364 (380 bar; 0.40 ratio CO₂/juice; 40°C; 10 min) of orange juice. As happened for the pH,
365 the natural variability of the juice is more important than the possible influence of the
366 treatment on the °Brix.

367

368 Finally, the acidity results showed that, although an average reduction of 4.94% was
369 found for the SC-CO₂-HPU treated samples, the differences were not significant ($p>0.05$)
370 due to the high degree of variability of the natural orange juice and the resulting
371 treatments.

372 In a similar way, both Kincal et al. (2006) and Tiwari et al. (2008a) found non-
373 significant changes in acidity after a continuous SC-CO₂ treatment (720 bar; 0.64 ratio
374 CO₂/juice; 40°C; 10 min) and an ultrasonic process (8.61-22.79 W/cm²; 2-10 min),
375 respectively.

376

377

378 3.2.2. Phenolic compounds and antioxidant capacity

379 The content of phenolic compounds significantly ($p<0.05$) decreased after the
380 continuous SC-CO₂-HPU treatment compared to that of the untreated juice, for all the
381 temperatures studied (-3.54, -3.68 and -4.15% at 31, 36 and 41°C, respectively; Figure
382 2). The differences among the treatments for this parameter were only significant
383 ($p>0.05$) between 31 and 41°C. Moreover, a significant difference ($p<0.05$) in phenolic
384 compounds was found between the SC-CO₂-HPU and pasteurization treatments; thus,
385 while the average decrease in phenolic compounds for SC-CO₂-HPU treatments was of
386 $-3.79\pm 0.9\%$, the thermal pasteurization brought about a decrease of -10%. This greater
387 loss of phenolic compounds could be due to the high degree of degradation of
388 carbohydrates and organic acids during the thermal processing, which could give rise to
389 furfurals and other carbonyl compounds which may form condensation products with
390 polyphenols (Fabroni et al., 2010). The results found in this study coincide with those
391 reported by Fabroni et al. (2010), who found reductions of 5.27% after a continuous SC-
392 CO₂ treatment (130 bar; 5.08 L/h juice; 1.96 L/h CO₂; 36°C; 15 min) of orange juice and
393 9.99% for a conventional thermal pasteurization treatment (90°C, 30 s). Therefore, it
394 seems that the use of HPU, which intensifies the microbial inactivation, do not negatively
395 affect the amount of phenolic compounds in the processed orange juice. Similarly,
396 Rawson et al. (2011a) observed no reduction in the content of phenolic compounds after
397 a HPU treatment (24.1–60 µm; 25-45 °C; 2-10 min) of watermelon juice.

398 The antioxidant capacity results showed a significant ($p < 0.05$) decrease in the
399 samples processed with SC-CO₂-HPU compared to the control samples (Figure 3),
400 except for the treatment at 31 °C in which a significant ($p < 0.05$) increase (12.13%) was
401 obtained. However, between 36 and 41 °C, there were no significant differences, leading
402 the treatments to reductions of 3.68 and 3.96%, respectively. Therefore, the use of
403 temperatures of over 31°C in the SC-CO₂-HPU treatment leads to a greater reduction in
404 the juice antioxidant capacity. On the other hand, thermal pasteurization presented a
405 significantly ($p < 0.05$) greater reduction (-9.07%) in the antioxidant capacity compared to
406 the continuous SC-CO₂-HPU treatments. Fabroni et al. (2010) reported similar results
407 after a continuous treatment with SC-CO₂ (130-230 bar; 5.08 L/h juice; 3.91 L/h CO₂; 36
408 °C; 15 min) of orange juice: the percentages of antioxidant capacity decreased by
409 between 1.39 and 2.53% versus 5.50 and 10.89% for pasteurized juice. However, in the
410 present study, an increase in the antioxidant capacity was observed at 31°C. It has been
411 widely reported that the use of HPU can lead to an increase in the antioxidant capacity
412 of vegetable samples, due to the increased extraction of active compounds. Therefore,
413 two effects could be superimposed in the present study: the increase in the antioxidant
414 capacity due to HPU and the decrease due to the SC-CO₂ treatment and the
415 temperature. In the case of 31°C, the result of these two effects brought about an
416 increase in the antioxidant capacity, since the greater quantity of compounds extracted
417 from juice pulp would compensate for the decrease produced by the SC-CO₂ treatment.

418

419 3.2.3. Browning index and color

420 Tables 3 and 4 show the results obtained for the browning index and color,
421 respectively. The SC-CO₂-HPU treatment of orange juice produced a significant ($p < 0.05$)
422 increase in the browning index when compared with the control sample at every
423 treatment temperature; the higher the temperature, the greater the browning index
424 increase (Table 3). However, the only significant differences found were those between
425 the treatment at 31 °C and the other two temperatures considered. The average change
426 in the browning index for the SC-CO₂-HPU treated samples was of 226%. An even
427 greater difference between the variation of the browning index in treated and untreated
428 samples was observed by Tiwari et al. (2008b) when working on sonicated orange juice
429 (40-100% amplitude), where it increased by 636.8%. Those authors attributed the
430 browning of the samples to the destruction of the pigments, mainly carotenoids,
431 produced by the HPU. One of the main factors contributing to the browning of orange
432 juice is ascorbic acid oxidation, leading to the appearance of reactive carbonyl groups,
433 such as furfural and 5-hydroxymethylfurfural, which can be precursors of non-enzymatic
434 browning (Bharate, & Bharate, 2012; Bull et al., 2004; Yeom, Streaker, Zhang, & Min,

435 2000). In addition, the browning effect could be linked to the decomposition of sugars or
436 caramelization (Vervoort et al., 2011) as well as to the Maillard reactions between
437 reducing sugars and free amino groups, leading to the formation of melanoidins, which
438 are compounds that cause dark browning (Ibarz-Martínez, Pagán, Garza, & Ibarz, 2010).
439 Vervoort et al. (2011) reported that the non-enzymatic browning is accelerated by the
440 temperature and processing time, as observed for the SC-CO₂-HPU treatment (Table 3).

441

442 However, the changes in the browning index are much larger for the SC-CO₂-HPU
443 treatment than for that of the thermal pasteurization. This shows that, although
444 temperature is an influential factor as regards juice browning, the mixing of the juice with
445 SC-CO₂ and/or the application of ultrasound are much more determinant.

446

447 On the other hand, the non-enzymatic browning produced by the treatment with SC-
448 CO₂-HPU was also observed from the results of the color analysis (Table 4). For every
449 treatment, there was a decrease in the L*, a*, b* values. Thus, the decrease in the L*
450 value showed a loss in brightness or increase in darkness which is directly related with
451 juice browning (Tiwari et al., 2008b; Yeom et al., 2000), although it could also be related
452 with the juice cloud, because the reflected light is affected by the cloud (Liu, Hu, Zhao,
453 & Song, 2012). The decrease in a* and b* values showed the color change to tonalities
454 less red and yellow. Considering the ΔE parameters, the greatest color difference was
455 obtained in samples treated with SC-CO₂-HPU at 36 °C, followed by 41 and 31 °C, while
456 a smaller color difference was observed for the pasteurized juice. Moreover, in this case,
457 the ΔE values do not point to a relationship with temperature. Therefore, with ΔE values
458 above 3, the color changes were noticeable for every treatment considered. A similar
459 finding was observed by Fabroni et al, (2010) in orange juice samples after continuous
460 SC-CO₂ treatment (130 bar; 5.08 L/h juice; 1.96-3.91 L/h CO₂; 36 °C; 15 min) and
461 thermal pasteurization (88-91 °C, 30s). These authors also observed a decrease in L*,
462 a*, b* values for both types of treatments, obtaining ΔE values of 7.87-11.89 and 2.88-
463 6.23 for the continuous treatment of SC-CO₂ and thermally pasteurized juice,
464 respectively. The color changes that take place after SC-CO₂-HPU treatment could also
465 be related to the cavitation effect of HPU, which regulates various physical, chemical and
466 biological reactions, between them, carotenoid degradation due to the free radicals
467 formed during the treatment (Abid et al., 2013; Tiwari et al., 2008a).

468

469 3.2.4. Cloud

470 The cloud is related with the particle suspension which is composed of a complex
471 mixture of proteins, pectins, lipids, hemicellulose, cellulose, and other minor components

472 (Niu et al., 2010; Tiwari et al., 2009b). This is an important attribute that positively affects
473 the turbidity, taste, aroma and characteristic color of orange juice. Its loss is mainly
474 attributed to the enzymatic activity of the PME, which causes phase separation in the
475 juice and the resulting loss of cloud (Bull et al., 2004; Polydera, Galanou, Stoforos, &
476 Taoukis, 2004).

477

478 The SC-CO₂-HPU treatment of orange juice significantly ($p < 0.05$) increased the cloud
479 values when compared with the control at every temperature considered (Figure 4),
480 showing that there is a significant ($p < 0.05$) difference between the treatments at 31 and
481 36 °C when compared to that at 41 °C. The average increases in the cloud value were
482 of 195.0, 198.4 and 270.6% at 31, 36 and 41°C, respectively; therefore, the cloud value
483 increased when the treatment temperature rose. In a similar way, after the SC-CO₂
484 treatment (400 bar; 55°C; 10-60 min) of orange juice, Niu et al. (2010) obtained
485 percentages of cloud increase of 91.33-115.48%. Also, Tiwari et al. (2008a), after the
486 ultrasonic treatment of orange juice (8.61-22.79 W/cm²; 2-10 min) obtained increases of
487 63-222%. As can be observed, after the SC-CO₂-HPU treatment of juice, the cloud was
488 preserved and improved. This phenomenon is mainly due to the reduction in the
489 enzymatic activity of the PME, as well as the system depressurization, which
490 homogenizes juice, causing the breakdown or reduction of the colloidal particles in the
491 juice (Kincal et al., 2006; Liu et al., 2012). Another factor that contributed to the increase
492 in the cloud is the HPU effect, which produced the rupture of the linear molecule pectin,
493 reducing its molecular weight (Tiwari et al., 2009b). The cloud values obtained after
494 pasteurization were significantly ($p < 0.05$) lower than those obtained after SC-CO₂-HPU
495 treatment which indicates that the use of this novel SC-CO₂-HPU-based technology
496 could improve some quality attributes of orange juice while reducing the processing time
497 compared to when only SC-CO₂ or HPU is used.

498

499 3.2.5. Ascorbic acid

500 The continuous SC-CO₂-HPU orange juice treatment at the different temperatures
501 considered produced a statistically significant reduction ($p < 0.05$) in ascorbic acid when
502 compared with the control (Figure 5). However, a considerable percentage of ascorbic
503 acid was preserved after treatments, observing reductions of only 4.55, 5.85, and 6.50%,
504 at 31, 36, and 41 °C, respectively. As can be observed, the ascorbic acid loss increased
505 as the temperature rose, although, in the range considered, the only significant ($p < 0.05$)
506 differences that exist are those between the treatment at 31 and that carried out at 41°C.
507 This slight degradation may be due to the formation of free radicals produced by the
508 effect of cavitation generated by the HPU, leading to the oxidation of polar organic

509 compounds, such as ascorbic acid and total phenols; and it may also be due to the
510 thermolysis produced inside bubbles and the subsequent activation of the Maillard
511 reaction (Tiwari et al, 2009a; Rawson et al, 2011b). A significant difference ($p < 0.05$)
512 between the ascorbic acid content of the juice after SC-CO₂-HPU treatment and thermal
513 pasteurization was also observed, with a greater loss of ascorbic acid in the case of the
514 thermal pasteurization (-10.05%). The greater reduction in ascorbic acid in the latter
515 treatment can be explained by the application of a higher processing temperature, since
516 ascorbic acid is a thermolabile nutrient (Sanchez-Moreno et al., 2005) and seems to be
517 more affected by high temperatures than by the use of the combined treatment (SC-CO₂-
518 HPU). Similar behavior was observed by Fabroni et al. (2010) after the continuous SC-
519 CO₂ processing of orange juice, finding a lower reduction of ascorbic acid in SC-CO₂-
520 HPU treated samples (6.37%, 130 bar; 5.08 L / h juice; 1.96 L / h CO₂; 36 ° C; 15 min)
521 compared to the thermally pasteurized ones (88-91 °C, 30s). Also, Tiwari et al. (2009a)
522 obtained reductions of 1.46-5.17% of ascorbic acid after the ultrasonic treatment of
523 orange juice (from 0.33 to 0.88 W / mL; 10.2 min) and a reduction of 7.14% after thermal
524 pasteurization. Despite the decrease in ascorbic acid when using SC-CO₂-HPU, at the
525 lowest temperature, less than half is reduced if compared to the case of thermal
526 pasteurization.

527 In order to apply new processing techniques at industrial level it is important to
528 consider the best process conditions (Boukroufa, Boutekedjiret, Petigny,
529 Rakotomanomana, & Chemat, 2015). In this regard, according to the results of the
530 present study, the use of a low pressure (100 bar), close to the critical pressure of CO₂
531 (72.9 bar), and a low temperature (31°C), which provides the best juice quality, would
532 facilitate the scaling of the process to the juice industry. Nevertheless, since the use of
533 ultrasound in liquid food could lead to degradation of some chemical compounds
534 (Jacotet-Navarro et al., 2016), further research should be conducted to evaluate the
535 influence of ultrasound intensification on possible degradation of bioactive compounds
536 in juices.

537

538

539 **4. Conclusions**

540 The SC-CO₂-HPU continuous treatment was effective for microbial inactivation in
541 orange juice, the effectiveness being dependent on the microbial cell wall thickness. The
542 SC-CO₂-HPU continuous treatment did not affect the pH, °Brix or Titratable acidity of the
543 juice. Moreover, compared with thermal pasteurization, the loss of phenolic compounds
544 was small and the antioxidant capacity could even be increased with respect to the
545 untreated juice. Although the treatment affected the color of the juice, causing an overall

546 darkening, the cloud and, therefore, the stability of the treated juices were greatly
547 improved. The obtained results demonstrated the potential of the continuous SC-CO₂-
548 HPU inactivation technique, the use of mild process conditions leading to an increase in
549 the quality of the product processed using this technique. Moreover, the fact that the
550 proposed technique works in a continuous mode greatly facilitates its industrial
551 implementation.

552

553 **5. Acknowledgements**

554 This work was supported by the PROMETEOII\2014\005 project financed by the
555 Generalitat Valenciana (Conselleria d'Educació, Cultura i Esport, Valencia, Spain). The
556 authors acknowledge the Consejo Nacional de Ciencia y Tecnología (CONACyT) for the
557 scholarship awarded to PhD Student Paniagua-Martínez, I. The authors especially wish
558 to thank Eng. Ramón Peña for his technical assistance in the development of the
559 equipment.

560

561 **6. References**

562

563 Abid, M., Jabbar, S., Wu, T., Hashim, M.M., Hu, B., Lei, S., Zhang, X., & Zeng, X. (2013).
564 Effect of ultrasound on different quality parameters of apple juice. *Ultrasonics*
565 *Sonochemistry*, 20(5), 1182-1187.

566 AOAC International. (2007). Official methods of analysis of the Association of Official
567 Analytical Chemists international. 18th edition, 2nd revision.

568 Boukroufa, M., Boutekedjiret, C., Petigny, L., Rakotomanomana, N., & Chemat, F.
569 (2015). *Ultrasonics Sonochemistry*, 24, 72-79.

570 Bharate, S.S., & Bharate, S.B. (2012). Non-enzymatic browning in citrus juice: chemical
571 markers, their detection and ways to improve product quality. *Journal of Food Science*
572 *and Technology*, 1-18.

573 Bull, M.K., Zerdin, K., Howe, E., Goicoechea, D., Paramanandhan, P., Stockman, R.,
574 Sellahewa, J., Szabo, E.A., Johnson, R.L., & Stewart, C.M. (2004). The effect of high
575 pressure processing on the microbial, physical and chemical properties of Valencia
576 and Navel orange juice. *Innovative Food Science & Emerging Technologies*, 5(2),
577 135-149.

578 Calix, T.F., Ferrentino, G., & Balaban, M.O. (2008). Measurement of high-pressure
579 carbon dioxide solubility in orange juice, apple juice, and model liquid foods. *Journal*
580 *of Food Science*, 73(9), 439-445.

581 Chemat, F., Rombaut, N., Meullemiestre, A., Turk, M., Perino, S., Fabiano-Tixier, A.S.,
582 & Abert-Vian, M. (2017). Review of Green Food Processing techniques. Preservation,

- 583 transformation and extraction. *Innovative Food Science and Emerging Technologies*,
584 41, 357-377.
- 585 Damar, S., & Balaban, M.O. (2006). Review of dense phase CO₂ technology: microbial
586 and enzyme inactivation, and effects on food quality. *Journal of Food Science*, 71(1),
587 1-11.
- 588 Fabroni, S., Amenta, M., Timpanaro, N., & Rapisarda, P. (2010). Supercritical carbon
589 dioxide-treated blood orange juice as a new product in the fresh fruit juice market.
590 *Innovative Food Science & Emerging Technologies*, 11(3), 477-484.
- 591 Ferrentino, G., Plaza, M.L., Ramirez-Rodrigues, M., Ferrari, G., & Balaban, M.O. (2009).
592 Effects of dense phase carbon dioxide pasteurization on the physical and quality
593 attributes of a red grapefruit juice. *Journal of Food Science*, 74(6), 333-341.
- 594 Gao, X., Ohlander, M., Jeppsson, N., Björk, L., & Trajkovski, V. (2000). Changes in
595 antioxidant effects and their relationship to phytonutrients in fruits of sea buckthorn
596 (*Hippophae rhamnoides* L.) during maturation. *Journal of Agricultural and Food
597 Chemistry*, 48(5), 1485-1490.
- 598 Garcia-Gonzalez, L., Geeraerd, A.H., Spilimbergo, S., Elst, K., Van Ginneken, L.,
599 Debevere, J., Van Impe, J.F., & Devlieghere, F. (2007). High pressure carbon dioxide
600 inactivation of microorganisms in foods: the past, the present and the future.
601 *International Journal of Food Microbiology*, 117, 1-28.
- 602 Gasperi, F., Aprea, E., Biasioli, F., Carlin, S., Endrizzi, I., Pirretti, G., & Spilimbergo, S.
603 (2009). Effects of supercritical CO₂ and N₂O pasteurisation on the quality of fresh
604 apple juice. *Food chemistry*, 115(1), 129-136.
- 605 Ibarz-Martínez, R., Pagán, J., Garza, S., & Ibarz, A. (2010). Browning of clarified lemon
606 juices treated at high temperatures. *Scientia Agropecuaria*, 1(1), 7-20.
- 607 Jacotet-Navarro, M., Rombaut, N., Deslis, S., Fabiano-Tixier, A.S., Pierre, F.X., Bilyb,
608 A., & Chemat, F. (2016). Towards a "dry" bio-refinery without solvents or added water
609 using microwaves and ultrasound for total valorization of fruit and vegetable by-
610 products. *Green Chemistry*, 18, 3106–3115.
- 611 Khandpur, P., & Gogate, P.R. (2016). Evaluation of ultrasound based sterilization
612 approaches in terms of shelf life and quality parameters of fruit and vegetable juices.
613 *Ultrasonics Sonochemistry*, 29, 337-353.
- 614 Kincal, D., Hill, W.S., Balaban, M.O., Portier, K.M., Wei, C.I., & Marshall, M.R. (2005). A
615 continuous high pressure carbon dioxide system for microbial reduction in orange
616 juice. *Journal of Food Science*, 70(5), 249-254.
- 617 Kincal, D., Hill, W.S., Balaban, M.O., Portier, K.M., Sims, C.A., Wei, C.I., & Marshall,
618 M.R. (2006). A continuous high-pressure carbon dioxide system for cloud and quality
619 retention in orange juice. *Journal of Food Science*, 71(6), 338-344.

- 620 Liu, X., Gao, Y., Xu, H., Hao, Q., Liu, G., & Wang, Q. (2010). Inactivation of peroxidase
621 and polyphenol oxidase in red beet (*Beta vulgaris* L.) extract with continuous high
622 pressure carbon dioxide. *Food Chemistry*, 119(1), 108-113.
- 623 Liu, Y., Hu, X., Zhao, X., & Song, H. (2012). Combined effect of high pressure carbon
624 dioxide and mild heat treatment on overall quality parameters of watermelon juice.
625 *Innovative Food Science & Emerging Technologies*, 13, 112-119.
- 626 Niu, L., Hu, X., Wu, J., Liao, X., Chen, F., Zhao, G., & Wang, Z. (2010). Effect of dense
627 phase carbon dioxide process on physicochemical properties and flavor compounds
628 of orange juice. *Journal of Food Processing and Preservation*, 34(2), 530-548.
- 629 Ortuño, C., Martínez-Pastor, M.T., Mulet, A., & Benedito, J. (2012). An ultrasound-
630 enhanced system for microbial inactivation using supercritical carbon dioxide.
631 *Innovative Food Science & Emerging Technologies*, 15, 31-37.
- 632 Ortuño, C., Martínez-Pastor, M.T., Mulet, A., & Benedito, J. (2013). Application of high
633 power ultrasound in the supercritical carbon dioxide inactivation of *Saccharomyces*
634 *cerevisiae*. *Food Research International*, 51(2), 474-481.
- 635 Ortuño, C., Balaban, M., & Benedito, J. (2014). Modelling of the inactivation kinetics of
636 *Escherichia coli*, *Saccharomyces cerevisiae* and pectin methylesterase in orange
637 juice treated with ultrasonic-assisted supercritical carbon dioxide. *The Journal of*
638 *Supercritical Fluids*, 90, 18-26.
- 639 Ozuna, C., Paniagua-Martínez, I., Castaño-Tostado, E., Ozimek, L., & Amaya-Llano, S.
640 L. (2015). Innovative applications of high-intensity ultrasound in the development of
641 functional food ingredients: Production of protein hydrolysates and bioactive peptides.
642 *Food Research International*, 77, 685-696.
- 643 Paniagua-Martínez, I., Mulet, A., García-Alvarado, M. A., & Benedito, J. (2016).
644 Ultrasound-assisted supercritical CO₂ treatment in continuous regime: Application in
645 *Saccharomyces cerevisiae* inactivation. *Journal of Food Engineering*, 181, 42-49.
- 646 Polydera, A.C., Galanou, E., Stoforos, N.G., & Taoukis, P.S. (2004). Inactivation kinetics
647 of pectin methylesterase of greek Navel orange juice as a function of high hydrostatic
648 pressure and temperature process conditions. *Journal of Food Engineering*, 62(3),
649 291-298.
- 650 Pulido, R., Bravo, L., & Saura-Calixto, F. (2000). Antioxidant activity of dietary
651 polyphenols as determined by a modified ferric reducing/antioxidant power assay.
652 *Journal of Agricultural and Food Chemistry*, 48(8), 3396-3402.
- 653 Rawson, A., Tiwari, B.K., Patras, A., Brunton, N., Brennan, C., Cullen, P.J., & O'Donnell,
654 C. (2011a). Effect of thermosonication on bioactive compounds in watermelon juice.
655 *Food Research International*, 44(5), 1168-1173.
- 656 Rawson, A., Patras, A., Tiwari, B.K., Noci, F., Koutchma, T., & Brunton, N. (2011b). Effect
657 of thermal and non thermal processing technologies on the bioactive content of exotic
658 fruits and their products: Review of recent advances. *Food Research International*,
659 44(7), 1875-1887.

- 660 Sánchez-Moreno, C., Plaza, L., Elez-Martínez, P., De Ancos, B., Martín-Belloso, O., &
661 Cano, M.P. (2005). Impact of high pressure and pulsed electric fields on bioactive
662 compounds and antioxidant activity of orange juice in comparison with traditional
663 thermal processing. *Journal of Agricultural and Food Chemistry*, 53(11), 4403-4409.
- 664 Tiwari, B.K., Mu hukuma appan, K., O'Donnell, C.P., & Cullen, P.J. (2008a). Effects of
665 sonication on the kinetics of orange juice quality parameters. *Journal of Agricultural
666 and Food Chemistry*, 56(7), 2423-2428.
- 667 Tiwari, B.K., Muthukumarappan, K., O'donnell, C.P., & Cullen, P.J. (2008b). Colour
668 degradation and quality parameters of sonicated orange juice using response surface
669 methodology. *LWT-Food Science and Technology*, 41(10), 1876-1883.
- 670 Tiwari, B.K., O'Donnell, C.P., Muthukumarappan, K., & Cullen, P.J. (2009a). Ascorbic
671 acid degradation kinetics of sonicated orange juice during storage and comparison
672 with thermally pasteurised juice. *LWT-Food Science and Technology*, 42(3), 700-704.
- 673 Tiwari, B.K., Muthukumarappan, K., O'donnell, C.P., & Cullen, P.J. (2009b). Inactivation
674 kinetics of pectin methylesterase and cloud retention in sonicated orange juice.
675 *Innovative Food Science & Emerging Technologies*, 10(2), 166-171.
- 676 Vervoort, L., Van der Plancken, I., Grauwet, T., Timmermans, R.A.H, Mastwijk, H.C.,
677 Matser, A.M., Hendrickx, M.E., & Van Loey, A. (2011). Comparing equivalent thermal,
678 high pressure and pulsed electric field processes for mild pasteurization of orange
679 juice: Part II: Impact on specific chemical and biochemical quality parameters.
680 *Innovative Food Science & Emerging Technologies*, 12(4), 466-477.
- 681 Wimmer, Z., & Zarevúcka, M. (2010). A review on the effects of supercritical carbon
682 dioxide on enzyme activity. *International Journal of Molecular Sciences*, 11(1), 233-
683 253.
- 684 Xu, Z., Zhang, L., Wang, Y., Bi, X., Buckow, R., & Liao, X. (2011). Effects of high pressure
685 CO₂ treatments on microflora, enzymes and some quality attributes of apple juice.
686 *Journal of Food Engineering*, 104(4), 577-584.
- 687 Yeom, H.W., Streaker, C.B., Zhang, Q.H., & Min, D.B. (2000). Effects of pulsed electric
688 fields on the quality of orange juice and comparison with heat pasteurization. *Journal
689 of Agricultural and Food Chemistry*, 48(10), 4597-4605.
- 690 Zhou, L., Wang, Y., Hu, X., Wu, J., & Liao, X. (2009). Effect of high pressure carbon
691 dioxide on the quality of carrot juice. *Innovative Food Science & Emerging
692 Technologies*, 10(3), 321-327.
- 693 Zinoviadou, K.G., Galanakis, C.M., Brnčić, M., Grimi, N., Boussetta, N., Mota, M.J., &
694 Barba, F.J. (2015). Fruit juice sonication: Implications on food safety and
695 physicochemical and nutritional properties. *Food Research International*, 77, 743-
696 752.
- 697

Highlights

- Ultrasound-assisted continuous SC-CO₂ **reduced** the orange juice microbiota
- The treatment produced greater changes in color (**darkening**) than thermal pasteurization.
- The cloud of the treated juice and the phenolic and vitamin contents were improved **by 173%, 6.5% and 5.5%, respectively.**
- The antioxidant capacity of the treated juice increased with respect to the fresh (untreated) one.
- **Ultrasound intensification may improve the acceptance of SC-CO₂ processing by the food industry.**

Figure Captions

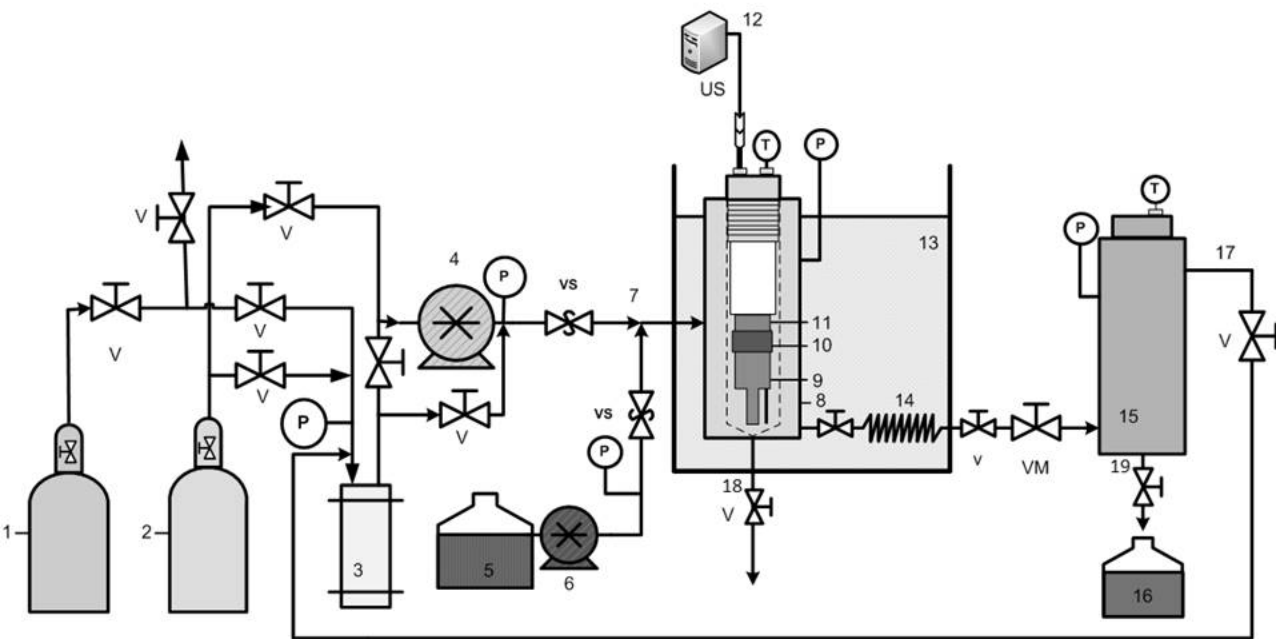
Fig. 1. Supercritical CO₂ continuous treatment system. 1. CO₂ tank; 2. N₂ tank; 3. Chiller reservoir; 4. CO₂ Pump; 5. Liquid reservoir; 6. Liquid Pump; 7. Mixing point; 8. Sonication vessel; 9. Sonotrode; 10. Insulation joint; 11. Ceramics; 12. Power generation unit; 13. Thermostatic bath; 14. Continuous contact tube; 15. Separation vessel; 16. Treated sample; 17. CO₂ Recirculation; 18. Sonication vessel output, 19. Separation vessel output, V. valve; VS. non-return valve; VM. micrometric valve; P. Manometer; T. temperature sensor.

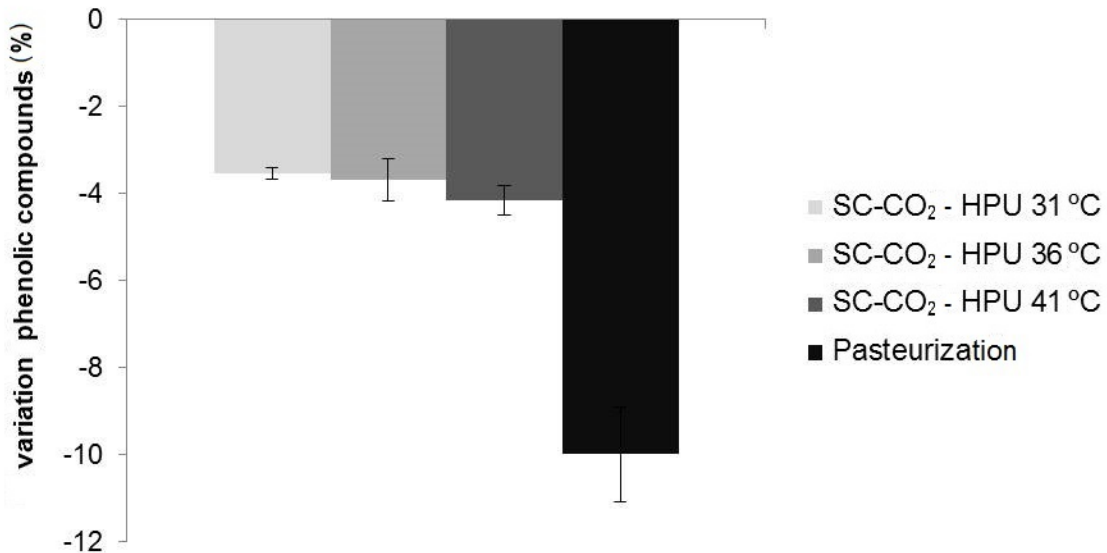
Fig. 2. Loss of phenolic compound content in orange juice after different treatment conditions.

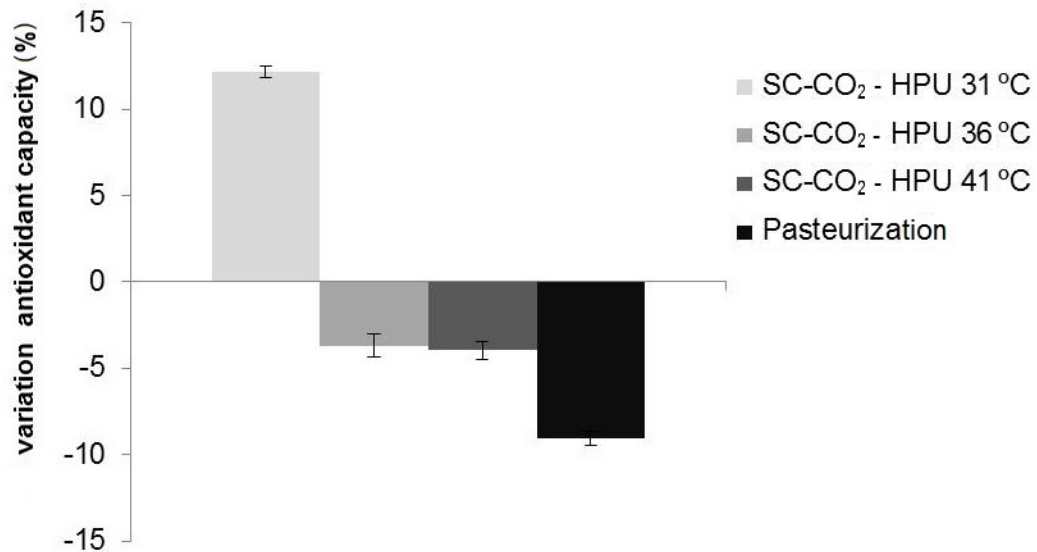
Fig. 3. Percentage variation of antioxidant capacity in orange juice under different treatment conditions.

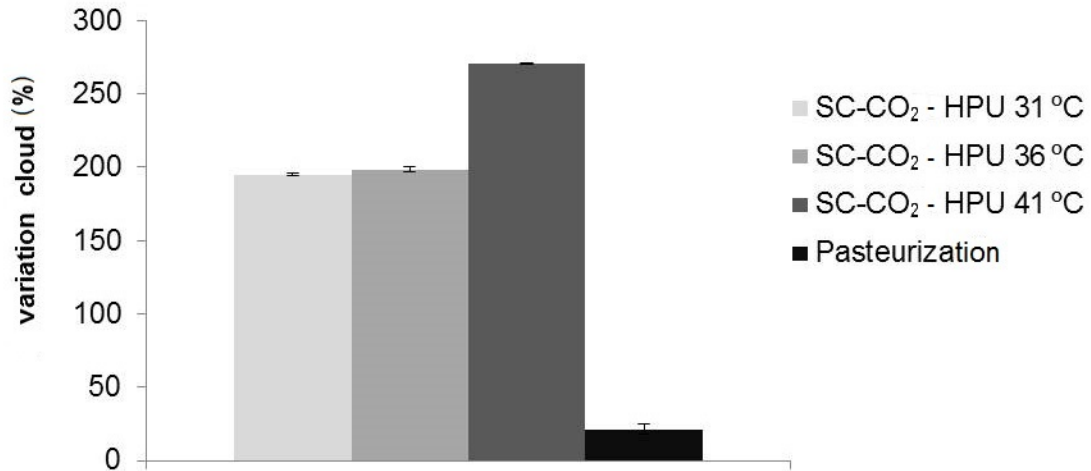
Fig. 4. Percentage variation of cloud in orange juice under different treatment conditions.

Fig. 5. Percentage variation of ascorbic acid in orange juice under different treatment conditions.









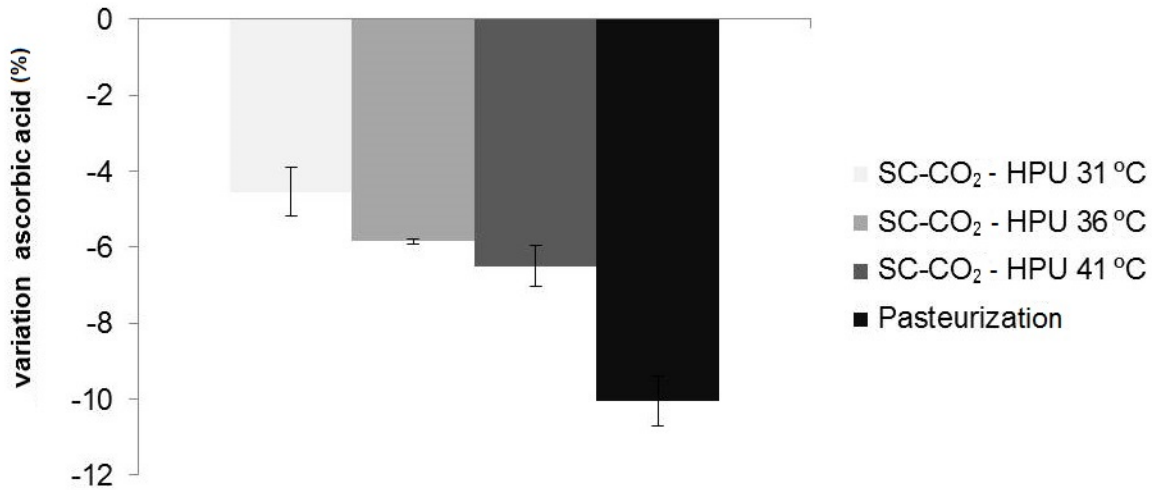


TABLE 1. Inactivation of microbiota in orange juice after SC-CO₂+HPU and thermal pasteurization treatments

| Treatment / conditions | <i>E.coli</i> * | | | <i>S.cerevisiae</i> * | | | Total aerobic mesophilic* | | |
|--|-----------------|----------|--------------------|-----------------------|----------|-----------------------|---------------------------|----------|--------------------|
| | N ₀ | N | Log N ₀ | N ₀ | N | -Log N/N ₀ | N ₀ | N | Log N ₀ |
| SC-CO ₂ -HPU/ 100 bar, 31 °C | 5.75E+03 | 0.00E+00 | 3.47±0.61 | 1.20E+05 | 2.00E+02 | 2.61±0.40 | 1.22E+03 | 0.00E+00 | 2.95±0.43 |
| SC-CO ₂ -HPU/ 100 bar, 36 °C | 1.24E+04 | 0.00E+00 | 3.80±0.63 | 4.53E+04 | 2.55E+02 | 2.24±0.21 | 3.66E+03 | 0.00E+00 | 3.50±0.31 |
| SC-CO ₂ -HPU/ 100 bar, 41 °C | 1.35E+04 | 0.00E+00 | 3.84±0.79 | 1.58E+05 | 1.04E+03 | 2.19±0.02 | 9.65E+03 | 0.00E+00 | 3.95±0.23 |
| Thermal pasteurization/ 90 °C, 1 min | 9.11E+02 | 0.00E+00 | 2.82±0.43 | 6.16E+03 | 0.00E+00 | 3.54±0.54 | 1.03E+03 | 0.00E+00 | 2.88±0.48 |

TABLE 2. pH, °Brix and titratable acidity values of orange juice after SC-CO₂+HPU and thermal pasteurization treatments

| Treatment / conditions | pH | | | °Brix | | | Titratable acidity (g citric acid/100ml)* | | |
|--|-------------------------|-------------------------|------------|--------------------------|---------------------------|-------------|---|---------------------------|-------------|
| | Control | Treated | Variation | Control | Treated | Variation | Control | Treated | Variation |
| SC-CO ₂ -HPU/ 100 bar, 31 °C | 3.58±0.04 ^a | 3.60±0.05 ^a | 0.56±0.01% | 12.23±0.23 ^a | 12.13±0.23 ^a | -0.81±0.3% | 0.874±0.02 ^a | 0.835±0.021 ^{ab} | -4.46±0.15% |
| SC-CO ₂ -HPU/ 100 bar, 36 °C | 3.63±0.03 ^a | 3.67±0.02 ^a | 1.10±0.08% | 11.46±0.06 ^{bc} | 11.26±0.05 ^c | -1.74±0.05% | 0.757±0.05 ^{abc} | 0.718±0.05 ^{bc} | -5.15±0.26% |
| SC-CO ₂ -HPU/ 100 bar, 41 °C | 3.68±0.005 ^a | 3.69±0.005 ^a | 0.27±0.05% | 12.20±0.17 ^a | 12.03±0.11 ^a | -1.39±0.12% | 0.747±0.04 ^{bc} | 0.708±0.04 ^c | -5.22±0.52% |
| Thermal pasteurization/ 90 °C, 1 min | 3.61±0.08 ^a | 3.62±0.09 ^a | 0.28±0.09% | 12.03±0.12 ^a | 11.86 ±0.05 ^{ab} | -1.41±0.23% | 0.836±0.04 ^{ab} | 0.829±0.04 ^{ab} | -0.83±0.11% |

Different letters for the same quality parameter within a row and column indicate significant differences (p <0.05)

TABLE 3. Browning index of orange juice after SC-CO₂+HPU and thermal pasteurization treatments

| Treatment / conditions | Browning Index (A420 nm) | | |
|--|--------------------------|------------------------|--------------|
| | Control | Treated | Variation |
| SC-CO ₂ -HPU/100 bar, 31 °C | 0.21±0.00 ^a | 0.66±0.02 ^b | 216.40±7.85% |
| SC-CO ₂ -HPU/100 bar, 36 °C | 0.23±0.01 ^a | 0.75±0.01 ^c | 228.13±4.49% |
| SC-CO ₂ -HPU/100 bar, 41 °C | 0.22±0.00 ^a | 0.72±0.01 ^c | 233.49±4.03% |
| Pasteurization/90 °C, 1 min | 0.21±0.01 ^a | 0.22±0.01 ^a | 5.38±0.19% |

Different letters within a row and column indicate significant differences (p <0.05)

TABLE 4. Color values of orange juice after SC-CO₂+HPU and thermal pasteurization treatments

| Treatment / conditions | Color* | | | | | | ΔE |
|--|------------|-----------|------------|------------|------------|------------|------------|
| | Control | | | Treated | | | |
| | L* | a* | b* | L* | a* | b* | |
| SC-CO ₂ -HPU/ 100 bar, 31 °C | 33.86±2.04 | 4.61±1.11 | 56.75±3.57 | 29.76±2.88 | 0.805±1.59 | 49.73±4.84 | 8.85±0.60 |
| SC-CO ₂ -HPU/ 100 bar, 36 °C | 40.52±0.96 | 5.54±0.35 | 64.30±2.45 | 33.86±1.73 | 0.98±0.73 | 55.79±2.47 | 11.74±1.25 |
| SC-CO ₂ -HPU/ 100 bar, 41 °C | 33.96±1.39 | 6.37±0.93 | 6.37±0.93 | 30.21±0.67 | 1.44±0.23 | 50.54±1.01 | 8.70±1.86 |
| Pasteurization/ 90 °C, 1 min | 35.00±0.25 | 8.25±1.20 | 58.74±0.26 | 32.42±0.13 | 4.52±0.28 | 54.47±0.18 | 6.27±0.87 |