Document downloaded from:

http://hdl.handle.net/10251/182179

This paper must be cited as:

Alvim, C.; Bes-Piá, M.; Mendoza Roca, JA. (2021). An innovative approach to the application of ultrasounds to remove polyethylene microspheres from activated sludge. Separation and Purification Technology. 264:1-8. https://doi.org/10.1016/j.seppur.2021.118429



The final publication is available at https://doi.org/10.1016/j.seppur.2021.118429

Copyright Elsevier

Additional Information

1	An innovative approach to the application of ultrasounds to remove polyethylene
2	microspheres from activated sludge
3	
4	Authors – C.Bretas Alvim, M.A Bes-Piá, J.A. Mendoza-Roca
5	Instituto de Seguridad Industrial, Radiofísica y Medioambiental, Universitat Politècnica
6	de València, Camino de Vera, s/n, Valencia 46022, Spain
7	* Corresponding author: claal@posgrado.upv.es
8	
9	Keywords – Microplastic, Polyethylene, Activated sludge, Ultrasound
10	
11	Abstract

12 During the secondary treatment of Wastewater Treatment Plants (WWTP) previous 13 studies have reported that microplastics (MPs), which correspond to plastic fragments 14 smaller than 5mm, are mostly retained in the activated sludge. Since the sludge can be applied as a fertilizer in agricultural land, the presence of MPs in this matrix can lead to 15 soil contamination. In this study, the application of ultrasounds at a frequency of 80 kHz 16 was tested to remove polyethylene (PE) microspheres from activated sludge samples in 17 order to reduce their concentration in this organic matrix. The application of ultrasounds 18 at a power of 50 W and 8 min 45 sec promoted the removal of $48 \pm 4\%$ of MPs from the 19 20 mixed liquor (2,820 \pm 51 mg/L of TSS). However, under the same operating conditions, $12 \pm 4\%$ of MPs were recovery from the concentrated mixed liquor (9,395 ± 164 mg/L of 21 22 TSS). For the concentrated sludge, a higher power and a longer sonication time (80 kHz; 23 90 W; 30 min) was necessary to improve the MP recovery. Under these conditions $38 \pm$ 4% of MPs were removed from the sludge. This is the first study applying ultrasounds to remove MPs from activated sludge and the results indicate that ultrasonication can be a suitable instrument in minimizing the concentration of PE microspheres in activated sludge, and thereby minimizing soil contamination when biosolids are applied to agriculture.

29

30 **1. Introduction**

A great interest has been shown in the applications of ultrasounds (US) in wastewater 31 treatment plants (WWTPs) since the beginning of this century. Focusing on municipal 32 33 wastewater management, the US irradiation has been studied for four main purposes: elimination of compounds that are not biodegraded in the biological treatment, 34 disinfection, reduction of sludge production and enhancement of methane production in 35 anaerobic digesters [1-12]. Cavitation generated by US can concentrate energy in 36 microreactors leading to the removal of a wide range of organic compounds like 37 38 surfactants [1] and polycyclic aromatic hydrocarbons, among others [2]. The removal 39 mechanism is based on the collapse of the bubbles created by the cavitation phenomenon, which causes changes in the liquid phase leading to oxidation or to breaking of the 40 41 chemical bonds and pyrolysis [3]. US can be applied to assist other techniques such as electrochemical treatment [4] or oxidation processes with ozone or hydrogen peroxide 42 [5]. US can be also used for water disinfection as reported by [6], also in combination 43 with conventional oxidation techniques like chlorination [7]. In addition, the combination 44 of US and chlorine reduces the concentration of disinfection by-products as 45 46 trichloromethane and trichloroacetic acid, which are compounds of great concern for the environment [8]. 47

Concerning to the applications focused on the sludge, US has been applied to produce 48 49 cell lysis in activated sludge reactors. In this way, the sludge production is reduced, with the consequent saving in the sludge management. In this way, Lin et al. [9] combined 50 oxidation with chlorine dioxide and US irradiation to achieve the reduction of the sludge 51 52 production and Romero-Pareja et al. [10] assisted the OSA (Oxic-settling-anaerobic) process with US for optimize the reduction of the sludge production. Finally, the 53 application of US for the enhancement of methane production is a well-known application 54 that is based on the effect of US on dissolving organic matter of the waste sludge, which 55 means that the hydrolysis phase of the anaerobic digestion is accelerated, and methane 56 57 production is enhanced [11,12].

58 The aforementioned applications have been hardly implemented in WWTPs due to the 59 investment and operating costs. However, the increasingly concern about microplastics (MPs) and emerging compounds can modify this scenario. MPs are plastic particles 60 smaller than 5mm, that can be classified as primary (microfibers and those intentionally 61 62 added in the formulation of products) and secondary (originated from the fragmentation of larger plastics) [13,14]. Despite the small dimensions of MPs, their numerous harmful 63 effects to living organisms have been reported. For instance, animals are not able to 64 65 distinguish between a plastic fragment and suitable food, and its ingestion can lead to blockage of the animals' trachea, causing starvation and even death [15]. The harmful 66 effects of MPs on the human body are still unclear. However, due to their hydrophobic 67 surface and high surface area, MPs can act as a vector for large quantities of persistent 68 organic contaminants [16,17,18] which can increase the risk of cancer in humans [18] and 69 70 can also result in endocrine dysregulation [17].

Previous studies have also reported the high retention of MPs in the activated sludgeduring the secondary treatment [19,20,21]. During the secondary treatment, the

microorganisms can use the MPs' surface as support for their growth and the formation 73 74 of complex structures of microorganisms on the MPs can lead to the reduction of the 75 MPs' buoyancy and thereby promote their accumulation in the sludge [22-25]. Since this biological material, also named 'biosolid', can be applied as a fertilizer on agricultural 76 77 land, the retention of MPs in this organic matrix can lead to soil contamination [26,27]. Nizzetto et al. [28] estimated that between 63,000-430,000 and 44,000-300,000 tons per 78 year of MPs are added to farmlands in Europe and North-America, respectively. In 79 Australia it was estimated that 2,800-19,000 tons per year of MPs enter agroecosystems 80 due to the application of biosolids [29]. The impacts of the presence of MPs in soils still 81 82 have many gaps, but recent research has proved that these small particles can alter the 83 germination and growth processes by preventing the uptake of water by plants due to physical blockage [30]. Furthermore, plants can also uptake these particles [31,32]. A 84 85 recent study by Oliveri Conti et al. [32] reports the presence of MPs in edible fruits and vegetables. Therefore, MPs can be ingested by humans when contaminated plant matter 86 is consumed. Due to their capacity to adsorb and leach toxic substances, such as 87 Bisphenol A (BPA) and polycyclic aromatic hydrocarbons (PAHs) [18,33], the ingestion 88 of MPs can lead to health issues for humans and animals. 89

90 There is still no process to remove the MPs from the activated sludge to minimize terrestrial contamination. Therefore, this study is focused on using the US process to 91 92 remove PE microspheres retained in the activated sludge flocs. With the use of ultrasounds, the sludge floc could be disrupted, releasing the MPs to the liquid phase. 93 94 Once the MPs were displaced to the liquid phase, these particles could be collected and 95 removed from the sludge. As a result, the concentration of MPs in biosolids and the terrestrial contamination would be reduced. Furthermore, to determine the capacity of the 96 US process to remove MPs from sludge with different concentrations of suspended solids, 97

98 samples from the aeration basin (mixed liquor) and from the secondary settling99 (concentrated mixed liquor also named secondary sludge), were studied.

100

101 **2.** Materials and methods

102

2.1. PE Microspheres and mixed liquor

In this study, the mixed liquor was collected from the aeration basin of a WWTP located 103 104 in Comunitat Valenciana (Spain). The mixed liquor (also named activated sludge) was used in a sequencing batch reactor (SBR), which was monitored for five days. Sludge 105 106 characteristics were presented in Table 3. The SBR was operated with a hydraulic retention time of 24 h, and 3 cycles per day. In each cycle 2 L of feed entered the system 107 and 2 L of effluent (supernatant) were withdrawn (Table 1). During the test, the SBR was 108 fed with a synthetic feed. The synthetic feed was prepared with peptone, meat extract and 109 110 K₂HPO₄ (supplied by Panreac) diluted in tap water, in proportions that guaranteed 500 111 mg/L of COD and nutrients (nitrogen and phosphorous) following the ratio of COD:N:P 112 of 100:5:1, as reported by Ferrer-Polonio et al. [34]. Regarding to the PE microsphere dosage, a concentration of 253MP per liter of feed was prepared. The MPs used in this 113 research was polyethylene microspheres (from COSPHERIC) with a diameter of 150-180 114 115 µm. PE microspheres concentration was calculated considering the diameter of the microspheres and the density of the polymer (around 1 g/cm^3). The volume of one sphere 116 was estimated from the diameter value and the mass from the estimated volume and the 117 118 density. Based on these calculations, the mass required to provide a concentration of 253 MP per liter was added in the feed. This method was already applied in a previous study 119 120 [24].

121

Table 1 -	SBR o	perational	conditions	and	parameters
-----------	-------	------------	------------	-----	------------

Cycles/day	3
Hydraulic retention time (HRT)	24h
Filling and aerobic reaction	бh
Sedimentation	90min
Draw and idle	30min

To measure the concentration of the MPs in the mixed liquor, 50mL of sample were chemically digested with 100 mL of hydrogen peroxide (35% wt) (AcrosOrganics) for 4 hours at $60 \pm 2^{\circ}$ C. The efficiency of this method was reported in a previous study [35]. After that, the sample was passed through a 150µm aperture sieve, and the retained material was rinsed with distilled water and filtered on glass fiber filter (1 µm). The number of microspheres was counted visually in a stereomicroscope (LEICA MZ APO).

131

132 2.2. Mechanism of extraction process via US

133 When MPs are dispersed in the activated sludge, these particles may have their surface 134 colonized by microbial communities, which increases their density and promotes their settlement in the sludge (Fig. 1.a) [19,23,24]. When the sludge is submitted to US, the 135 136 cavitation process promotes the rupture of the sludge floc [36], and consequently MPs are 137 released to the liquid phase (supernatant). Therefore, this study aims to evaluate different operational conditions to remove via US the PE microspheres retained in the activated 138 sludge floc. As colloids are generated from the disruption of the sludge floc, they will 139 140 remain in suspension due to their low density, which increases the supernatant turbidity 141 (Fig. 1.b) [36,37].

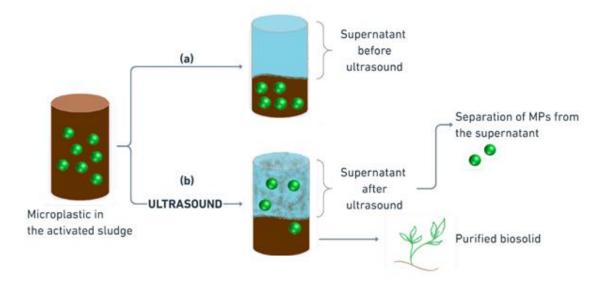


Fig. 1. Extraction of PE microspheres from activated sludge via US

- 144
- 145
- 146

147 **2.3.** Mixed liquor – Extraction of MPs by ultrasound

The ultrasound tests for the mixed liquor were performed at a frequency of 80kHz, and 148 varying the power input (30 W, 60 W and 90 W) and sonication time (5, 10 and 15 149 150 minutes). For each test, 100mL of sample were collected and poured into a beaker 151 (250mL) for the extraction process, including an initial sample that was not submitted to US. The US tests were operated under sweep mode controlling the temperature of the 152 mixed liquor. Since the cavitation could result in a heat increase, it was key to maintain 153 the temperature at $25 \pm 5^{\circ}$ C. At the end of the US, the samples were settled for 90 min 154 and 25 mL of supernatant was extracted and filtered through a 1 µm glass fiber filter. The 155 experiments were replicated to check their reproducibility. The MPs that were attached 156 157 on the beaker wall were carefully collected using high precision tweezers. The mixed 158 liquor after ultrasonication was centrifuged at 12,000 rpm during 15 min at 4 °C to extract the soluble microbial products (SMP). The supernatant was filtered through a 0.45 µm 159 160 cellulose acetate filter for SCOD, SMP (SMPp - proteins and SMPc - carbohydrates)

analysis. SMPp were measured by BCA method [38] and SMPc by anthrone method [39].
The SCOD was measured with a kit of the provider Merck Milipore.

- 163
- 164

2.3.1. Experimental Design

An experimental design was carried out with the software Statgraphics Centurion XVII 165 166 to predict the optimum point of maximum recovery of MPs and minimum turbidity of the 167 supernatant when the mixed liquor was submitted to US. Specifically, a three-level factorial design with response surface method (RSM) was performed. For this, the 168 independent variables were power input (W) and sonication time (min) (Table 2), whereas 169 170 the response variables were the percentage of microspheres recovered from the mixed 171 liquor and turbidity. In this work, 18 experimental runs were performed and executed in 172 two blocs. In addition, Statgraphics Centurion XVII was used for data analysis. Thus, 173 based on the results of RSM it was possible to better visualize the behavior of a single 174 variable response under the operational conditions proposed, and the interaction of the 175 variables in the optimum condition to obtain the highest percentual removal of MPs from 176 the mixed liquor. The analysis of variance (ANOVA) was applied in the experimental 177 models to assess if the independent variables had significance, and to determinate the 178 degree of the correlation between the predicted and real responses.

- 179
- 180

Table 2 – Values of variables for the experimental design.

Independent variables		Coded va	lues		
	-1	0	1		
(Factor_A) Time (min)	5	10	15		
(Factor_B) Power (W)	30	60	90		

183

2.4. Concentrated mixed liquor

Concentrated mixed liquor (simulating sludge from secondary settling) was achieved concentrating the mixed liquor 3 times to reach a concentration of around 9,000mg/L. US tests for this sample were performed using a frequency of 80 kHz and a power of 90 W. Longer US times, compared to the tests with mixed liquor, were applied (30 min and 60 min). The temperature was also controlled and maintained at $25 \pm 5^{\circ}$ C. 100 mL of concentrated mixed liquor were used for each experiment, which were replicated to evaluate the reproducibility of the method.

Unlike the procedure applied for the mixed liquor (Section 2.3), the supernatant of the concentrated mixed liquor was not drained due to the small column of water in this sample. Therefore, after US, the sample was left to stand for 90 min and then the MPs were carefully removed from the sample surface with high precision tweezers and placed in other beaker with distilled water. This final sample was filtered through a glass fiber filter, similarly to the process adopted in section 2.3.

197

198 **2.5. Sonochemical effects**

199 The sonochemical effects, which correspond to the production of hydroxyl radicals due to the abrupt collapse of the microbubbles, were determined by KI dosimetry method 200 (Weissler reaction). This method is based on the oxidation of I⁻ when free radicals are 201 202 generated during the ultrasonication, and the final product is the ion I_3 , which can be measured at wavelength of 355 nm. A solution of KI 1% was prepared for the tests. 203 204 Quantification of sonochemical effects was carry out applying the same conditions used for the activated sludge. 100 mL of KI solution were placed in a 250 mL beaker and 205 exposed to ultrasonication. The power values used were 30 W, 60 W and 90 W. An aliquot 206

of KI solution was collected at the beginning of the test (initial sample) and after 5 min, 10 min, 15 min and 20 min of reaction, and its absorbance was measure at λ =355 nm by DR6000 (Hach) spectrophotometer. As suggested by Wang et al. [40], the concentration of free radicals (HO•) in the system is related to the absorption of the KI solution after US treatment and can be estimated by Eq.1.

212

213
$$C(\text{HO} \bullet) = \frac{2A}{\varepsilon b} \times 10^6 \times 2 \tag{1}$$

214

215 Where the concentration of hydroxyl radical C(HO•) is obtained in μ mol/L, A is the 216 absorption of the KI solution after US, ϵ is the molar absorptivity of I_3^- (26,303 217 dm³mol⁻¹cm⁻¹) and b is the thickness of quartz cuvette (10 mm).

218

219

9 **2.6.** Particle size distribution of MPs after ultrasonication

Since MPs could be downsized to nanoplastics (NPs) due to water shear forces [41] the 220 221 particle size distribution of the supernatant was determined by Dynamic Light Scattering 222 (Zetasizer Nano ZS90) to assess the eventual fragmentation of PE microspheres into 223 smaller fragments. The experiments were performed with distilled water because the high 224 presence of suspended solids from the activated sludge after US could make the analysis 225 unfeasible. After the US, the sample was passed through a 150 µm aperture sieve to remove the initial MPs size. The removal of the initial MPs of 150 µm was required since 226 227 the equipment identified particles in a range size of 0.3 nm and 5 µm and bigger particles could interfere in the results. Moreover, the analysis of the distilled water blank was also 228 required to minimize the influence of contamination. 229

230

231 **3. Results**

3.1. Microspheres concentration in the mixed liquor

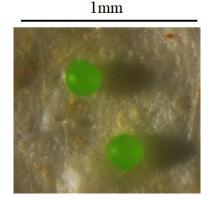
When the supernatant of the initial mixed liquor (without US application) was collected, 233 only $2 \pm 0.5\%$ of the microspheres were recovered, i.e. $98 \pm 0.5\%$ were accumulated in 234 235 the sludge (Fig. 2). Due to the high deposition of MPs in the biomass the final concentration of microspheres in the mixed liquor was 1,400 MP/L (Table 3). The 236 237 concentrated mixed liquor had a TSS concentration of 9,395 ± 164mg/L and a 238 concentration of microspheres approximately 3 times higher than the mixed liquor (4,480 MP/L). Likewise, for this sample $2 \pm 0.5\%$ of MPs were recovered from the supernatant 239 240 before the US.

- 241
- 242

Table 3 – Sludge samples characterization (mean±S.D)

Parameter		Sample	
	Mixed liquor	Concentrated mixed liquor	
TSS (mg/L)	2,820 ± 51	9,395 ± 164	
VSS/TSS	0.86	0.81	
MPs/L	1,400	4,480	

243



244

245

Fig.2. PE microspheres removed from the activated sludge

3.2. Influence of power and time on MP extraction by ultrasounds

248

3.2.1. Results of the experiment design for mixed liquor

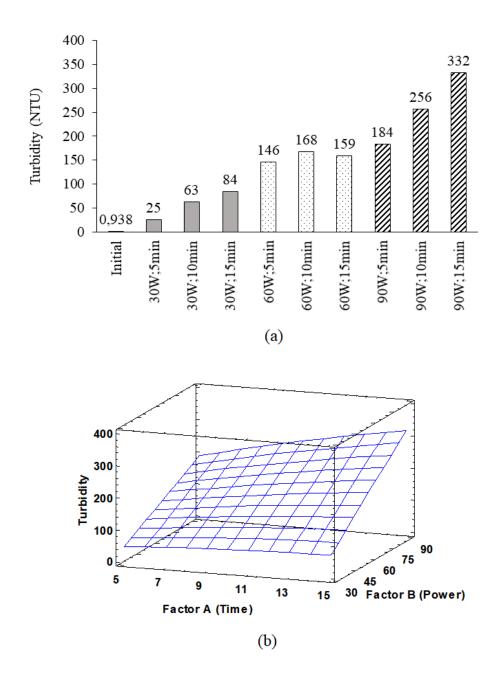
249 In the Fig.3.a, it was observed that the increase in the power and the exposure time 250 contributed to the increase of the supernatant turbidity, reaching a maximum value of 332 NTU when the sludge was exposed to 90 W and 15 min. The increase of the turbidity 251 252 indicates that the sludge flocs were disrupted by the hydro-mechanical effects of the US, 253 leading to a higher concentration of suspended solids in the supernatant. The contribution 254 of the power and the sonication time to the disintegration of the sludge floc and therefore 255 the rise of the turbidity, is clearly noted with the surface response (Fig.3.b). The following 256 equation (Eq.2) corresponds to the regression model established according to the results of turbidity response in the experimental design, where A is the time and B the power. 257

258

259 $Turbidity(NTU) = -34,44 + 1.78A + 1.70B - 0.23A^2 + 0.17AB - 0.00092B^2$ 260 (Eq.2)

261

The analysis of variance for the regression model also corroborated that the variables A and B and the interactive model terms (AB) are significant (p-Value < 0.05), at the 95% confidence level, for the increment in the turbidity value. Moreover, a high squared regression coefficient (R^2) and adjusted squared regression coefficient (R^2_{adj}) were achieved (0.95 and 0.93, respectively), indicating a great correlation between the predicted and real responses.



269

Fig.3. Turbidity of the supernatant after US tests at different operating conditions (a);
Response surface plot for turbidity (b)

The percentage of recovery of MPs as a function of the power and time is represented in Fig.4.a. In general terms, the higher time and power, the more MPs were recovered from the mixed liquor supernatant. For 90 W and 15 min the turbidity was so high that it made impossible the separation of the supernatant and the MPs. Because of that, the value for

this operational condition of MPs recovery was registered as null. Appling 60 W and 15 277 min, the MPs recovery decreased compared to 10 min, which does not follow the 278 279 aforementioned trend. In order to better understand the relation between the recovery of PE microspheres and the sludge disintegration, the Fig.4.b shows that the percentage of 280 microspheres recovered from the mixed liquor can be directly related to the degree of 281 sludge floc disintegration (express in terms of turbidity). As the rupture of the sludge is 282 accompanied by increased turbidity in the supernatant, the lower recovery of PE 283 284 microspheres under 60 W and 15 min compared with the recovery under 60W and 10 min can be justified due to the lower sludge disintegration in the first operational condition 285 286 (lower turbidity in the supernatant).

287

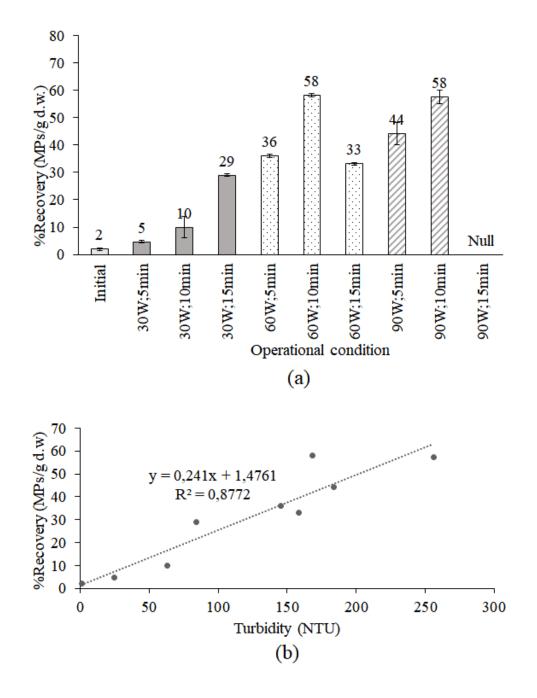


Fig.4. Recovery of MPs after US tests at different operating conditions (a); Recovery of
MPs in turbidity function (b)

289

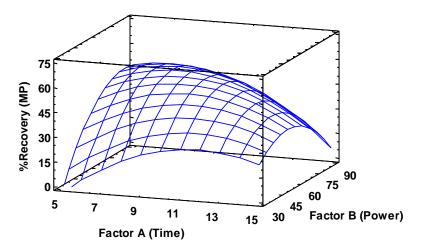
The following equation (Eq.3) corresponds to the regression model established according to the results of percentual MPs recovery response in the experimental design. The analysis of variance conduced for the regression model, showed that the factor B, the quadratic models (A² and B²), and the interactive model terms (AB), were all significative
(p-Value< 0.05), at the 95% confidence level.

298

299 %*Recovery* (*MP*) = $-164.22 + 19.35A + 3.82B - 0.66A^2 - 0.11AB - 0.019B^2$ 300 (Eq.3)

301

The squared regression coefficient (R^2) and the adjusted squared regression coefficient 302 (R^{2}_{adi}) (0.82 and 0.73, respectively) indicate that there is a correlation between the 303 predicted and real responses. The recovery of MPs from mixed liquor was influenced by 304 305 power and time (Fig.5), but the extent of this dependence should be further evaluated in 306 future studies to improve the extraction process and reach a higher correlation between 307 the model and real responses. The contribution of the frequency can also be an important parameter to understand better the role of US in the recovery of MPs from activated 308 309 sludge.



310

Fig.5. Response surface plot for %Recovery (MP)

3.2.2. Mixed liquor disintegration by ultrasonication

313 As stated above, the sonication power and time are parameters that played an important 314 role in the degree of sludge disintegration. Thus, the higher power input, the more 315 effective the sound pressure was, and therefore, a greater disintegration of the sludge was 316 obtained. Similarly, with a longer ultrasonication time, the rupture of the sludge floc and 317 cell membranes was more effective. In section 3.2.1 turbidity was reported as a 318 measurement of sludge disintegration. In this section, results are focused on reporting the 319 quality of the supernatant (final effluent from WWTP) in terms of SCOD and of its main components (SMP). 320

According to the SCOD results (Fig.6.a), it was observed an increment in SCOD value 321 with the increase of the power input. In addition, an increment in SCOD with 10 minutes 322 323 of ultrasonication, compared to 5min, was also observed for all experiments. The increase 324 in SCOD value is related to the organic matter released into water due to the 325 microorganism's cell lysis. As the microorganism's cells are mostly composed by protein, 326 their lysis result in more soluble microbial proteins (SMPp) released in the water comparing with soluble microbial carbohydrates (SMPc) [42,43]. The increase in SMPp 327 concentration was observed after all the US experiments (Fig.6.b), suggesting that the 328 hydro-mechanical shear forces, during the US, were sufficient to promote the rupture of 329 330 the sludge floc.

331

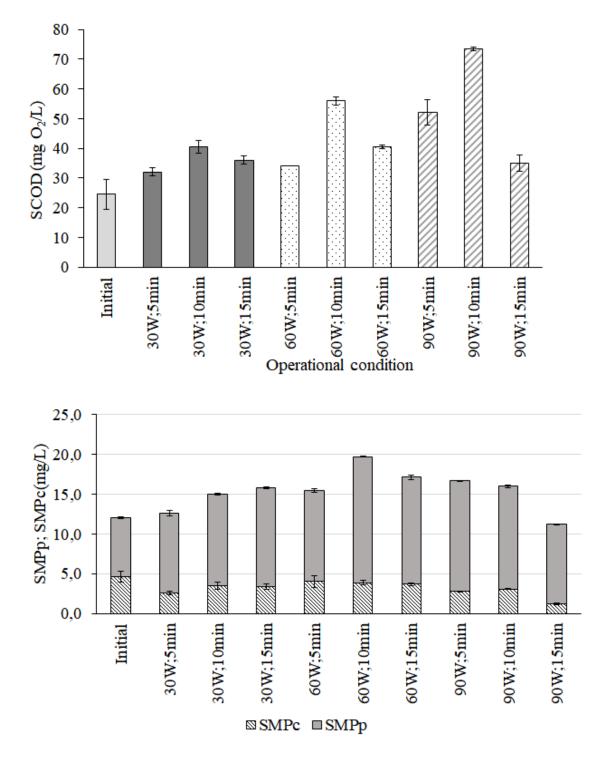
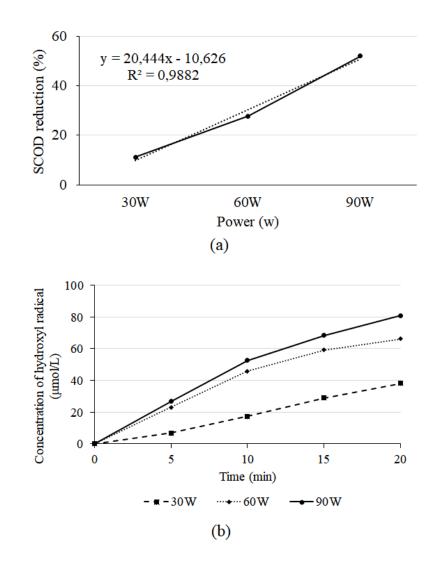


Fig.6. Mixed liquor disintegration in terms of SCOD released concentration (a) and carbohydrates and proteins concentrations (b)

However, for all powers tested it was observed that, when the time was increased from 336 337 10 minutes up to 15 minutes, a decrease of SCOD occurred. This reduction can be justified by the sonochemical effects. In this process, due to the acoustic cavitation, 338 microbubbles are formed and violently collapsed, resulting in high temperature and 339 pressure points. The result of this mechanism is the sonochemical effect, which results in 340 the generation of hydroxyl radicals [44]. Hydroxyl radicals have a high oxidant capacity 341 and react with organic species fast and in a nonspecific way [45]. The reaction of •OH 342 with the extracellular polymeric substances (EPS) - present in the sludge floc - can also 343 promote its rupture and the release of soluble organics as SMP, or even its mineralization 344 345 into CO₂ and H₂O [42]. When the mixed liquor was submitted to 15 minutes of US at any 346 tested power, the oxidation of soluble organic matter due to the sonochemical effects, was more evident, resulting in a final value of SCOD lower than 10min. Analyzing the SMPc 347 348 results after the ultrasonication, a reduction in its concentration was observed compared to the initial value. This can be another evidence of the oxidation of soluble organic matter 349 by the generated hydroxyl radicals. Based on these results, it was established a hypothesis 350 that a maximum cell lysis was achieved at 10min. After that, cell lysis still occurred, but 351 352 the mineralization of SCOD seemed to be more intense. Wang et al. [45] reported that the 353 sludge disintegration during the ultrasonication is performed by both hydro-mechanical shear forces and oxidizing effect. To mask the oxidizing effect of radical •OH, the authors 354 proposed the addition of NaHCO₃ in the sludge before the US. Al-Juboori et al. [46] 355 356 investigated the removal of dissolved organic carbon (DOC) present in Narda lagoon (Autralia) applying a 20khz frequency, power input of 93W and 143W and treatment time 357 358 of 5 and 15min. The authors used real water, which contains a high concentration of microbes, to assess the influence of the microbe's presence in the ultrasonication process. 359

360 They concluded that the release of microbial cell lysis products and the reduction of DOC361 occurred simultaneously when applying US to water with a high microbial load.

362 With the application of 30W there was a decrease of SCOD in 12% between 10 and 363 15min. For 60W and 90W a reduction of 28% and 58% were obtained, respectively 364 (Fig.7.a). The explanation of the relation between the reduction of SCOD and the power input is that the power intensity is linked to the size of the cavitation bubbles, the 365 366 temperature, and the internal pressure of the bubbles during their collapse. Increasing the power intensity results in a greater number of collapsed bubbles, higher temperature and 367 greater sonochemical effects [47]. This may be the reason why the increase in the power 368 369 produced a proportional SCOD reduction between 10 and 15min test. The results of the sonochemical effects measured at 5, 10, 15 and 20 min, were plotted in Fig.7(b) in order 370 to assess the hydroxyl radical production in the system. As expected, the sonochemical 371 effects, increased with both exposition time and power applied. 372



373

Fig.7. SCOD reduction with power at 10min and 15min of US (a); Production of free
radicals in function of time and power (b)

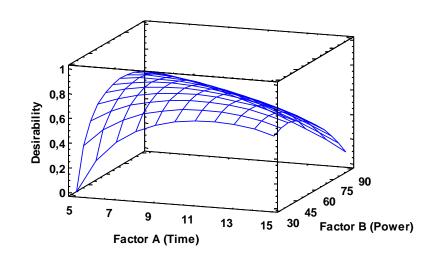
377

3.2.3. Optimized extraction model

From the results obtained in experimental design (section 3.2.1), the optimum point of maximum recovery of MPs and minimum turbidity of the supernatant was predicted. According to the response surface established by the software (Fig.8), a test time of 8 min 45 sec at a power of 54 W was ideal. These operational conditions would theoretically result in a recovery percentage of 50 MPs and a turbidity of 134 NTU (Table 4). Due to the power adjustment possibilities in the US equipment, it was not possible to apply 54

W. Therefore, 50 W was established as the power for the optimal point test. According to 384 the established model, the expected response for the percentage of MPs recovery was 385 50% and for the turbidity 136 NTU. When carrying out the extraction process, $48 \pm 4\%$ 386 of MPs were recovered and a turbidity of 179 ± 6 NTU was produced. Moreover, an 387 increase in SCOD was observed ($45 \pm 3 \text{ mgO}_2/\text{L}$ after US). The results obtained under 388 the optimal condition showed a high correlation between the expected values and the real 389 390 ones, and therefore, it showed a relevant step in the possible application of US to remove 391 MPs from activated sludge.

392



393

394

Fig.8. Response surface plot for optimization

396

Table 4 - Ultrasound optimized operational conditions for mixed liquor.

Factor	Value	Parameter	
Factor_A (min)	8 min 45 sec	%Recovery (MPs)	Maximize
Factor_B (W)	54	Turbidity (NTU)	Minimize
	Predicted response	Real response	

%Recovery (MPs)	50	48±4
Turbidity (NTU)	136	179±6

398

399

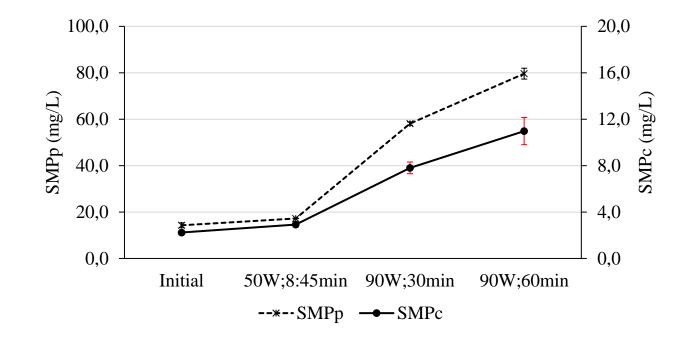
3.2.4. Concentrated mixed liquor

400 The extraction of MPs from WWTP sludge may be influenced by the concentration of 401 total solids (TS) in the sludge. A high concentration of TS can absorb acoustic energy and 402 reduce the efficiency of the sludge floc disintegration [48], and consequently decrease the 403 release of MPs. Applying the optimum point established in section 3.2.3. for the mixed liquor, $48 \pm 4\%$ of PE microspheres were recovered by the supernatant. However, when 404 405 the same experimental conditions were applied to the concentrated mixed liquor, only 12 406 \pm 4% recovery was achieved. The high concentration of TS, compromised the cavitation 407 process, making difficult to detach the MPs retained in the sludge. However, submitting 408 the sludge to a higher power and time (90 W; 30 min) the recovery of MPs reached $38 \pm$ 409 4% (Table 5).

411 Table 5 – MPs recovery (%) and SCOD concentrations achieved at different ultrasound
412 operational conditions for concentrated mixed liquor.

%Recovery (MPs)	SCOD (mg/L)
2 ± 1	39 ± 1
12 ± 4	61 ± 4
38 ± 4	188 ± 6
29 ± 1	310 ± 4
	2 ± 1 12 ± 4 38 ± 4

On the contrary, upon being subjected to US, the SCOD value increased instead of 414 415 decreasing due to the sludge fragmentation, that caused a release of more organic products. In addition, a longer US time (comparing the same power of 90 W) resulted in 416 a greater disintegration of the sludge. Although the US process produces free radicals 417 (extremely oxidizing), results suggested that the sludge disintegration process by hydro-418 419 mechanical was higher than the sonochemical effects. Tian et al. [49] studied the 420 ultrasonication effects in the activated sludge (16.2-17.2 g/L of TS), under 20 kHz frequency, and they also observed a low influence of free radicals in the ultrasonication 421 422 process and the predominance of hydro-mechanical shear forces in the sludge 423 disintegration. The results of SMPp and SMPc for concentrated mixed liquor (Fig.9) corroborate that. At the optimum conditions for the mixed liquor (50 W; 8 min 45 sec), 424 425 the concentrated mixed liquor suffered a low disintegration. However, when 90W;60min 426 was applied, an increment of more than 450% and 390% of SMPp and SMPc were achieved, respectively. According to these results and the ones regarding the recovery of 427 428 MPs, the degree of MPs recovery seems to be related to the extent of floc disintegration, 429 as it was also concluded in the section 3.2.1 for the mixed liquor. Nevertheless, an intense 430 sludge fragmentation can be negative for the recovery of MPs. After 90 W; 60 min of 431 ultrasonication a reduction in the recovery of MPs compared to 30 min was observed. Our hypothesis is that, under extreme sludge disintegration conditions, the sample are not 432 433 able to decant, and the consequence is that the sample is more homogeneous with a more 434 intense mixture of sludge and MPs. Therefore, without a separation of water from the sludge, the buoyancy of MPs is seriously compromised. 435





438Fig.9. Released SMPp and SMPc after ultrasonication of concentrated mixed liquor

This is the first investigation into how the US can accomplish the extraction of MPs from activated sludge, and therefore the results obtained in this study cannot be compared with previous ones. However, this research provides important information about the viability of treating the activated sludge for the removal PE microspheres. Several microplastics can be found in activated sludge (microfibers, fragments and spheres), of different sizes and compositions, thus further research must be carried out to evaluate the potential for removing these MPs through US.

447

```
448 3.3. Particle size distribution of PE microspheres after ultrasonication
```

According to the results of the particle size distribution in the supernatant for the optimum
point (80 kHz; 50 W; 8 min 45 sec) applied to the mixed liquor, and for the best conditions
applied to the concentrated sludge (80 kHz; 90 W; 30 min), the fragmentation of the PE

microspheres into smaller fragments was not observed. This result would demonstrate
that PE microspheres fragmentation should be caused by factors like biodegradation, UV
irradiation, and not just mechanical processes such as US [41].

455

456 **4.** Conclusions

457 The main conclusion of this work is that ultrasonication can become an excellent tool to 458 assist in the separation of PE microspheres, a type of MPs, from the activated sludge. It was possible to remove $48 \pm 4\%$ of MPs that were retained in the mixed liquor by applying 459 460 of ultrasonication optimal conditions at a frequency of 80 kHz (power input 50 W and time 8 min 45 sec). After the US, the PE microspheres were transferred to the water 461 462 column phase due to the disruption of the sludge floc by cavitation process, which allowed its extraction and therefore, the reduction of the concentration of MPs in the sludge. In 463 464 addition, the quality of the final effluent was not compromised as showed the SCOD and 465 SMP measured values.

466 Regarding the concentrated mixed liquor, when these optimal operational conditions were applied, only $12 \pm 4\%$ of MPs could be recovered from the activated sludge. The higher 467 total solid concentration compromised the cavitation process, leading to a poor sludge 468 469 disintegration compared to that of the mixed liquor. Since the sludge was insufficiently disintegrated, only a small amount of MPs were recovered. However, when the 470 concentrated mixed liquor was exposed to a higher power and a longer US time, (80 kHz; 471 472 90 W; 30 min) $38 \pm 4\%$ of MPs were detached from the sludge floc and moved to the sample surface. Moreover, the fragmentation of the MPs into NPs after the US for both 473 474 mixed liquor and concentrated mixed liquor was not observed. Based on these results, the US process has an interesting potential for the extraction of MPs from a biomass matrix. 475 Further studies should be developed to better understand the process of extracting MPs 476

via US and to study the feasibility of its application on an industrial scale. The reduction
of the concentration of MPs in the activated sludge deserves attention, since mostly MPs
that enter the secondary treatment are retained in the sludge, and this material is
commonly applied as fertilizers in agricultural lands leading to their contamination by
these synthetic polymers.

482

483 ACKNOWLEDGMENTS

Authors thank the Spanish Ministry of Science, Innovation and Universities for the
financial support (Reference of the project: RTI2018-096916-B-I00)

486

487 **REFERENCES**

M.A. Abu-hassan, D. Mantzavinos, I.S. Metcalfe, Wet air oxidation and
ultrasound for the removal of linear alkylbenzene sulfonates from wastewater: the
beneficial role of catalysis, 33 (2005). https://doi.org/10.1007/s11244-005-2520-6.

491 [2] N. Ghasemi, G. Gbeddy, P.Egodawatta, F. Zare, A. Goonetilleke, Removal of

492 polycyclic aromatic hydrocarbons from wastewater using dual-mode ultrasound system,

493 Water and Environment Journal (2017) 1–10. https://doi.org/10.1111/wej.12540.

494 [3] P. Chowdhury, T. Viraraghavan, Sonochemical degradation of chlorinated

495 organic compounds, phenolic compounds and organic dyes – A review, Sci. Total

496 Environ. 407 (2009) 2474–2492. https://doi.org/10.1016/j.scitotenv.2008.12.031.

497 [4] M. Zhang, Z. Zhang, S. Liu, Y. Peng, J. Chen, S. Yoo Ki, Ultrasound-assisted
498 electrochemical treatment for phenolic wastewater, Ultrason. Sonochem. 65 (2020)

499 105058. https://doi.org/10.1016/j.ultsonch.2020.105058.

- 500 [5] S. Chandak, P.K. Ghosh, P.R. Gogate, Treatment of real pharmaceutical
- 501 wastewater using different processes based on ultrasound in combination with oxidants,
- 502 Process Saf. Environ. Prot. 137 (2020) 149–157.
- 503 https://doi.org/10.1016/j.psep.2020.02.025.
- 504 [6] M. Vázquez-López, L.E. Amabilis-Sosa, G.E. Moeller-Chávez, A. Roé-Sosa, P.
- 505 Neumann, G. Vidal, Evaluation of the ultrasound effect on treated municipal
- 506 wastewater, Environ. Technol. 40 (2019) 3568–3577.
- 507 https://doi.org/10.1080/09593330.2018.1481889.
- 508 [7] Blume, T., Leis, U. Improving chlorine disinfection of wastewater by ultrasound
- application. Water science and technology 52 (10-11) (2005), 139-144.
- 510 [8] X. Zhou, J. Zhao, Z. Li, J. Song, X. Li, X. Yang, D. Wang, Enhancement effects
- of ultrasound on secondary wastewater effluent disinfection by sodium hypochlorite and
- disinfection by-products analysis, Ultrason. Sonochem. 29 (2016) 60–66.
- 513 https://doi.org/10.1016/j.ultsonch.2015.09.001.
- 514 [9] J. Lin, Y. Hu, G. Wang, W. Lan, Sludge reduction in an activated sludge sewage
- treatment process by lysis-cryptic growth using ClO2-ultrasonication disruption,
- 516 Biochem. Eng. J. 68 (2012) 54–60. https://doi.org/10.1016/j.bej.2012.07
- 517 [10] P.M. Romero-Pareja, C.A. Aragon, J.M. Quiroga, M.D. Coello, Evaluation of a
- 518 biological wastewater treatment system combining an OSA process with ultrasound for
- sludge reduction, Ultrason. Sonochem. 36 (2017) 336–342.
- 520 https://doi.org/10.1016/j.ultsonch.2016.12.006.
- 521 [11] J. Quarmby, J.R. Scott, A.K. Mason, G. Davies, S.A. Parsons, The Application
- of Ultrasound as a Pre-Treatment for Anaerobic Digestion, Environ. Technol. 20 (1999)
- 523 1155–1161. https://doi.org/10.1080/09593332008616913.

- 524 [12] X. Li, S. Guo, Y. Peng, Y. He, S. Wang, L. Li, M. Zhao, Anaerobic digestion
- 525 using ultrasound as pretreatment approach: Changes in waste activated sludge,
- 526 anaerobic digestion performances and digestive microbial populations, Biochem. Eng. J.
- 527 139 (2018) 139–145. https://doi.org/10.1016/j.bej.2017.11.009.
- 528 [13] H.S. Auta, C.U. Emenike, S.H. Fauziah, Distribution and importance of
- 529 microplastics in the marine environment A review of the sources, fate, effects, and
- 530 potential solutions, Environ. Int. 102 (2017) 165–176.
- 531 https://doi.org/10.1016/j.envint.2017.02.013.
- 532 [14] S.A. Carr, J. Liu, A.G. Tesoro, Transport and fate of microplastic particles in
- 533 wastewater treatment plants., Water Res. 91 (2016) 174–82.
- 534 https://doi.org/10.1016/j.watres.2016.01.002.
- 535 [15] M. Cole, H. Webb, P.K. Lindeque, E.S. Fileman, C. Halsband, T.S. Galloway,
- 536 Isolation of microplastics in biota-rich seawater samples and marine organisms, Sci.
- 537 Rep. 4 (2014) 1–8. https://doi.org/10.1038/srep04528.
- 538 [16] H. Bouwmeester, P.C.H. Hollman, R.J.B. Peters, Potential Health Impact of
- 539 Environmentally Released Micro- and Nanoplastics in the Human Food Production
- 540 Chain: Experiences from Nanotoxicology, Environ. Sci. Technol. 49 (2015) 8932–8947.
- 541 https://doi.org/10.1021/acs.est.5b01090.
- 542 [17] B. Hu, Y. Li, L. Jiang, X. Chen, L. Wang, S. An, F. Zhang, Influence of
- 543 microplastics occurrence on the adsorption of 17β -estradiol in soil, J. Hazard. Mater.
- 544 400 (2020) 123325. https://doi.org/10.1016/j.jhazmat.2020.123325.
- 545 [18] M.D. Sharma, A.I. Elanjickal, J.S. Mankar, R.J. Krupadam, Assessment of
- 546 cancer risk of microplastics enriched with polycyclic aromatic hydrocarbons, J. Hazard.
- 547 Mater. 398 (2020) 122994. https://doi.org/10.1016/j.jhazmat.2020.122994.

- 548 [19] G. Kalčíková, B. Alič, T. Skalar, M. Bundschuh, A.Ž. Gotvajn, Wastewater
- treatment plant effluents as source of cosmetic polyethylene microbeads to freshwater,

550 Chemosphere. 188 (2017) 25–31. https://doi.org/10.1016/j.chemosphere.2017.08.131.

- 551 [20] C. Edo, M. González-Pleiter, F. Leganés, F. Fernández-Piñas, R. Rosal, Fate of
- 552 microplastics in wastewater treatment plants and their environmental dispersion with
- effluent and sludge, Environ. Pollut. 259 (2020).
- 554 https://doi.org/10.1016/j.envpol.2019.113837.
- 555 [21] J. Talvitie, A. Mikola, O. Setälä, M. Heinonen, A. Koistinen, How well is
- 556 microlitter purified from wastewater? A detailed study on the stepwise removal of
- 557 microlitter in a tertiary level wastewater treatment plant, Water Res. 109 (2017) 164–
- 558 172. https://doi.org/10.1016/j.watres.2016.11.046.
- 559 [22] D. Lobelle, M. Cunliffe, Early microbial biofilm formation on marine plastic
- 560 debris, Mar. Pollut. Bull. 62 (2011) 197–200.
- 561 https://doi.org/10.1016/j.marpolbul.2010.10.013.
- 562 [23] E.R. Zettler, T.J. Mincer, L.A. Amaral-Zettler, Life in the "plastisphere":
- 563 Microbial communities on plastic marine debris, Environ. Sci. Technol. 47 (2013)
- 564 7137–7146. https://doi.org/10.1021/es401288x.
- 565 [24] C. Bretas Alvim, S. Castelluccio, E. Ferrer-Polonio, M.A. Bes-Piá, J.A.
- 566 Mendoza-Roca, J. Fernández-Navarro, J.L. Alonso, I. Amorós, Effect of polyethylene
- 567 microplastics on activated sludge process Accumulation in the sludge and influence on
- the process and on biomass characteristics, Process Saf. Environ. Prot. 148 (2021) 536–
- 569 547. https://doi.org/10.1016/j.psep.2020.10.014.

- 570 [25] M. Wu, W. Tang, S. Wu, H. Liu, C. Yang, Fate and effects of microplastics in
- wastewater treatment processes, Sci. Total Environ. 757 (2021) 143902.
- 572 https://doi.org/10.1016/j.scitotenv.2020.143902.
- 573 [26] F. Corradini, P. Meza, R. Eguiluz, F. Casado, E. Huerta-Lwanga, V. Geissen,
- 574 Evidence of microplastic accumulation in agricultural soils from sewage sludge
- 575 disposal, Sci. Total Environ. 671 (2019) 411–420.
- 576 https://doi.org/10.1016/j.scitotenv.2019.03.368.
- 577 [27] P. van den Berg, E. Huerta-Lwanga, F. Corradini, V. Geissen, Sewage sludge
- 578 application as a vehicle for microplastics in eastern Spanish agricultural soils, Environ.
- 579 Pollut. 261 (2020) 114198. https://doi.org/10.1016/j.envpol.2020.114198.
- 580 [28] L. Nizzetto, M. Futter, S. Langaas, Are Agricultural Soils Dumps for
- 581 Microplastics of Urban Origin?, Environ. Sci. Technol. 50 (2016) 10777–10779.
- 582 https://doi.org/10.1021/acs.est.6b04140.
- 583 [29] E.L. Ng, E. Huerta Lwanga, S.M. Eldridge, P. Johnston, H.W. Hu, V. Geissen,
- 584 D. Chen, An overview of microplastic and nanoplastic pollution in agroecosystems, Sci.
- 585 Total Environ. 627 (2018) 1377–1388. https://doi.org/10.1016/j.scitotenv.2018.01.341.
- 586 [30] T. Bosker, L.J. Bouwman, N.R. Brun, P. Behrens, M.G. Vijver, Microplastics
- accumulate on pores in seed capsule and delay germination and root growth of the
- terrestrial vascular plant Lepidium sativum, Chemosphere. 226 (2019) 774–781.
- 589 https://doi.org/10.1016/j.chemosphere.2019.03.163.
- 590 [31] K. Dietz, S. Herth, Plant nanotoxicology, Trends Plant Sci. 16 (2011) 582–589.
- 591 https://doi.org/10.1016/j.tplants.2011.08.003.

- 592 [32] G. Oliveri Conti, M. Ferrante, M. Banni, C. Favara, I. Nicolosi, A. Cristaldi, M.
- 593 Fiore, P. Zuccarello, Micro- and nano-plastics in edible fruit and vegetables. The first
- diet risks assessment for the general population, Environ. Res. 187 (2020) 109677.
- 595 https://doi.org/10.1016/j.envres.2020.109677.
- 596 org/10.1016/j.chemosphere.2019.03.163.
- 597 [33] W. Wei, Q.S. Huang, J. Sun, J.Y. Wang, S.L. Wu, B.J. Ni, Polyvinyl Chloride
- 598 Microplastics Affect Methane Production from the Anaerobic Digestion of Waste
- 599 Activated Sludge through Leaching Toxic Bisphenol-A, Environ. Sci. Technol. 53
- 600 (2019) 2509–2517. https://doi.org/10.1021/acs.est.8b07069.
- 601 [34] E. Ferrer-Polonio, J. Fernández-Navarro, J.L. Alonso-Molina, J.A. Mendoza-
- 602 Roca, A. Bes-Piá, I. Amorós, Towards a cleaner wastewater treatment: Influence of
- folic acid addition on sludge reduction and biomass characteristics, J. Clean. Prod. 232
- 604 (2019) 858–866. https://doi.org/10.1016/j.jclepro.2019.06.021.
- 605 [35] C. Bretas Alvim, M.A. Bes-Piá, J.A. Mendoza-Roca, Separation and
- 606 identification of microplastics from primary and secondary effluents and activated
- sludge from wastewater treatment plants, Chem. Eng. J. 402 (2020) 126293.
- 608 https://doi.org/10.1016/j.cej.2020.126293.
- 609 [36] Z. Meng, Z. Zhou, D. Zheng, L. Liu, J. Dong, Y. Yang, X. Li, T. Zhang,
- 610 Optimizing dewaterability of drinking water treatment sludge by ultrasound treatment:
- 611 Correlations to sludge physicochemical properties, Ultrason. Sonochem. 45 (2018) 95–
- 612 105. https://doi.org/10.1016/j.ultsonch.2018.02.027.
- 613 [37] X. Feng, H. Lei, J. Deng, Q. Yu, H. Li, Physical and chemical characteristics of
- waste activated sludge treated ultrasonically, Chem. Eng. Process. Process Intensif. 48
- 615 (2009) 187–194. https://doi.org/10.1016/j.cep.2008.03.012.

- 616 [38] E. Zuriaga-Agustí, A. Bes-Piá, J.A. Mendoza-Roca, J.L. Alonso-Molina,
- 617 Influence of extraction methods on proteins and carbohydrates analysis from MBR
- 618 activated sludge flocs in view of improving EPS determination, Sep. Purif. Technol.
- 619 112 (2013) 1–10. https://doi.org/10.1016/j.seppur.2013.03.048.
- 620 [39] B. Frølund, R. Palmgren, K. Keiding, P.H. Nielsen, Extraction of extracellular
- 621 polymers from activated sludge using a cation exchange resin, Water Res. 30 (1996)
- 622 1749–1758. doi:10.1016/0043-1354(95)00323-1.
- 623 [40] P. Wang, C. Cheng, Y. Ma, M. Jia, Degradation behavior of polyphenols in
- 624 model aqueous extraction system based on mechanical and sonochemical effects
- 625 induced by ultrasound, Sep. Purif. Technol. 247 (2020) 116967.
- 626 https://doi.org/10.1016/j.seppur.2020.116967.
- 627 [41] M. Enfrin, J. Lee, Y. Gibert, F. Basheer, L. Kong, L.F. Dumée, Release of
- 628 hazardous nanoplastic contaminants due to microplastics fragmentation under shear
- 629 stress forces, J. Hazard. Mater. 384 (2020) 121393.
- 630 https://doi.org/10.1016/j.jhazmat.2019.121393.
- 631 [42] R. Guan, X. Yuan, Z. Wu, L. Jiang, Y. Li, Principle and application of hydrogen
- 632 peroxide based advanced oxidation processes in activated sludge treatment: A review,
- 633 Chem. Eng. J. 339 (2018) 519–530. https://doi.org/10.1016/j.cej.2018.01.153.
- 634 [43] G.J. Xie, B.F. Liu, Q. Wang, J. Ding, N.Q. Ren, Ultrasonic waste activated
- 635 sludge disintegration for recovering multiple nutrients for biofuel production, Water
- 636 Res. 93 (2016) 56–64. https://doi.org/10.1016/j.watres.2016.02.012.
- 637 [44] K.R. Morison, C.A. Hutchinson, Limitations of the Weissler reaction as a model
- reaction for measuring the efficiency of hydrodynamic cavitation, Ultrason. Sonochem.
- 639 16 (2009) 176–183. https://doi.org/10.1016/j.ultsonch.2008.07.001.

- 640 [45] N.N. Mahamuni, Y.G. Adewuyi, Ultrasonics Sonochemistry Advanced
- 641 oxidation processes (AOPs) involving ultrasound for waste water treatment : A review
- 642 with emphasis on cost estimation, Ultrason. Sonochemistry. 17 (2010) 990–1003.
- 643 https://doi.org/10.1016/j.ultsonch.2009.09.005.
- 644 [45] F. Wang, Y. Wang, M. Ji, Mechanisms and kinetics models for ultrasonic waste
- activated sludge disintegration, J. Hazard. Mater. 123 (2005) 145–150.
- 646 https://doi.org/10.1016/j.jhazmat.2005.03.033.
- 647 [46] R.A. Al-Juboori, T. Yusaf, V. Aravinthan, L. Bowtell, Investigating natural
- organic carbon removal and structural alteration induced by pulsed ultrasound, Sci.
- 649 Total Environ. 541 (2016) 1019–1030. https://doi.org/10.1016/j.scitotenv.2015.09.143.
- 650 [47] P. Kanthale, M. Ashokkumar, F. Grieser, Sonoluminescence, sonochemistry
- (H2O2 yield) and bubble dynamics: Frequency and power effects, Ultrason. Sonochem.
- 652 15 (2008) 143–150. https://doi.org/10.1016/j.ultsonch.2007.03.003.
- 653 [48] V.K. Tyagi, S.L. Lo, L. Appels, R. Dewil, Ultrasonic treatment of waste sludge:
- A review on mechanisms and applications, Crit. Rev. Environ. Sci. Technol. 44 (2014)
- 655 1220–1288. https://doi.org/10.1080/10643389.2013.763587.
- 656 [49] X. Tian, W.J. Ng, A.P. Trzcinski, Optimizing the synergistic effect of sodium
- hydroxide/ultrasound pre-treatment of sludge, Ultrason. Sonochem. 48 (2018) 432–440.
- 658 https://doi.org/10.1016/j.ultsonch.2018.07.005.