

Article

Controlled Synthesis of Up-Conversion NaYF₄:Yb,Tm Nanoparticles for Drug Release under Near IR-Light Therapy

Edelweiss Moyano Rodríguez ^{1,2}, Miguel Gomez-Mendoza ², Raúl Pérez-Ruiz ^{2,3}, Beatriz Peñín ⁴,
Diego Sampedro ⁴, Antonio Caamaño ^{1,*} and Víctor A. de la Peña O'Shea ^{2,*}

- ¹ Signal Theory and Communications and Telematic System and Computing, Rey Juan Carlos University, Cam/del Molino 5, 28942 Madrid, Spain; edelweiss.moyano@urjc.es
² Photoactivated Processes Unit, IMDEA Energy Institute, C/Ramón de la Sagra, 3, 28935 Madrid, Spain; miguel.gomez@imdea.org (M.G.-M.); raupreru@qim.upv.es (R.P.-R.)
³ Departamento de Química, Universitat Politècnica de València, Camino de Vera S/N, 46022 Valencia, Spain
⁴ Departamento de Química, Centro de Investigación en Síntesis Química (CISQ), Universidad de La Rioja, Madre de Dios 53, 26006 Logroño, Spain; beatriz.penin@unirioja.es (B.P.); diego.sampedro@unirioja.es (D.S.)
* Correspondence: antonio.caamano@urjc.es (A.C.); victor.delapenya@imdea.org (V.A.d.l.P.O.)

Abstract: Up-Conversion materials have received great attention in drug delivery applications in recent years. A specifically emerging field includes the development of strategies focusing on photon processes that promote the development of novel platforms for the efficient transport and the controlled release of drug molecules in the harsh microenvironment. Here, modified reaction time, thermal treatment, and pH conditions were controlled in the synthesis of NaYF₄:Yb,Tm up-converted (UC) material to improve its photoluminescence properties. The best blue-emission performance was achieved for the UC3 sample prepared through 24 h-synthesis without thermal treatment at a pH of 5, which promotes the presence of the β-phase and smaller particle size. NaYF₄:Yb,Tm has resulted in a highly efficient blue emitter material for light-driven drug release under near-IR wavelength. Thus, NaYF₄:Yb,Tm up-converted material promotes the N-O bond cleavage of the oxime ester of Ciprofloxacin (prodrug) as a highly efficient photosensitized drug delivery process. HPLC chromatography and transient absorption spectroscopy measurements were performed to evaluate the drug release conversion rate. UC3 has resulted in a very stable and easily recovered material that can be used in several reaction cycles. This straightforward methodology can be extended to other drugs containing photoactive chromophores and is present as an alternative for drug release systems.



Citation: Moyano Rodríguez, E.; Gomez-Mendoza, M.; Pérez-Ruiz, R.; Peñín, B.; Sampedro, D.; Caamaño, A.; de la Peña O'Shea, V.A. Controlled Synthesis of Up-Conversion NaYF₄:Yb,Tm Nanoparticles for Drug Release under Near IR-Light Therapy. *Biomedicines* **2021**, *9*, 1953. <https://doi.org/10.3390/biomedicines9121953>

Academic Editor: José García Solé

Received: 14 November 2021

Accepted: 7 December 2021

Published: 20 December 2021

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

Keywords: Up-Conversion; drug-delivery; Near IR-Light Therapy

1. Introduction

The release of drugs through the blood to a target is a field widely investigated in recent decades [1,2]. The use of drug delivery systems by means of liposomes [3,4], mixed micelles [5–7], niosomes [8], micelles [9], bile salts aggregates [10–12], nanoparticles [13,14], nanocapsules, gold nanoparticles, microspheres [15], microcapsules, nanobubbles [16], microbubbles, and dendrimers [17,18] are being investigated for diagnosis and therapy [19,20]. Carriers based on micelles (liposomes, mixed micelles, and niosomes, among others) are made of bile salts, phospholipids, and cholesterol (Ch). They are among the most important biological entities in mammals, exhibiting an outstanding capability for solubilizing lipophilic molecules [21,22]. The study of nanoparticles for drug delivery applications allows both the development of novel platforms for the efficient transport and the controlled release of drug molecules in the harsh microenvironment (changes of pH and/or temperature) of diseased tissues in living systems [23]. Thus, these systems are being exploited for the development of pharmaceutical formulations with improved delivery to

the target [24–26]. Lipophilicity and membrane partition properties of potential drug candidates are key to the recognition of their target in cells [27]. Despite the advantages of using nanosized drug delivery systems, some drawbacks can be found from this methodology. For example, delivering a sufficient amount of these external agents is required in order to provide the best possible efficacy. Moreover, since these nanocarriers could be affected by changes in pH or temperature in different organs, tissues, and subcellular compartments, undesired degradation of these agents could be produced with the corresponding toxic exposure, including swelling of surrounding tissues [23,28].

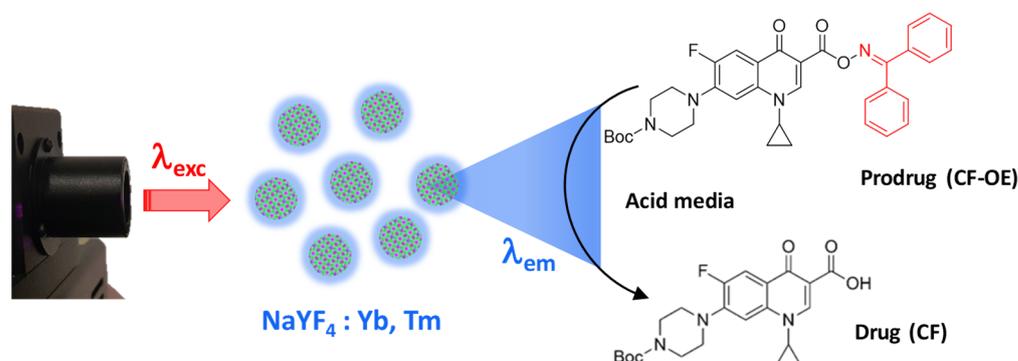
In previous years, the development of strategies focusing on photon Up-Conversion (UC) processes for photothermal therapy [29,30], cell imaging [31,32] and pH-responsive [33,34] treatments have received great attention [35]. These UC processes are based on the absorption of photons leading to the emission of another photon whose energy is higher than the energy of the absorbed photons. The system involves a light absorber named sensitizer or donor (such as Yb) and other species which collects the absorbed energy from the sensitizer, named acceptor (such as Er/Tm/Eu/Ho) embedded on a transparent matrix (such as NaYF₄, BaYF₅, NaGdF₄, or LiYF₅) [36]. In a UC system, initial irradiation with photons of low energy results in the emission of higher-energetic photons [37–39]. These materials can be tuned by modifying their photophysical properties and are able to convert intense light from the near-infrared region to high energy light in the visible or UV region [40–42]. It is important to note the strong stability of these UC particles after receiving NIR radiation, which allows for combining this process with others. For all these reasons, they can be used as drug delivery systems, upon light absorption, which requires the controlled release of active compounds at the specific targets responsible for the pharmacological action [43,44].

A key aspect that directly affects both the physical-chemical and photoluminescence properties of the resulting UC is the synthesis method. State of the art processes mainly consider one of the following options: coprecipitation [45], sol-gel process [46], thermal decomposition [45], or solvothermal process [46]. Coprecipitation and the sol-gel process are quite simple methods but do not allow control of the obtained crystal phase ratios, which is one of the aims to optimize the luminescence of the material. Thermal decomposition produces high-quality UCs but requires high temperatures and contaminant solvents, which are not suitable for biomedical applications. Finally, the solvothermal process is the most adequate for this study because it allows for the fine control of the structural, morphological, and optical properties of materials [47–49]. In addition, unlike other methods, it does not use volatile solvents that require critical synthesis temperatures and pressures conditions which increase costs, as well as decrease the sustainability of the process.

Ciprofloxacin (CF) is an antibiotic that belongs to the quinolone family and has been widely employed in the last thirty years to treat a number of bacterial infections [50] as well as prevent the formation of prostate or lung cancer cells by the inhibition of topoisomerase II [51,52]. Moreover, it is also commonly used for the elimination or reduction of bacteria from the tumor environment that consumes gemcitabine, a very common drug in chemotherapeutic treatments for breast, ovarian, or metastatic cancer [53,54]. The CF is slightly water-soluble and it can occasionally be detected in urine and blood. To achieve a specific delivery of this drug to the target, the formation of a prodrug with better water solubilization is warranted which may release the corresponding drug after an external input such as light; however, the prodrug only absorbs UV-blue light which is a biologically incompatible wavelength.

To overcome this drawback, an interesting strategy is the use of UC materials that can be excited with near-infrared (biologically compatible) light, leading to specific UV-blue emissions which would induce a specific bond cleavage of the prodrug and, therefore, efficient delivery of the active drug (CF). To demonstrate this hypothesis, we effectively illustrate the use of stable NaYF₄:Yb,Tm as a photon UC material with optimized blue luminescence that efficiently induces CF delivery from its prodrug (an oxime-ester derivative, CF-OE) in acid media (Scheme 1). The optimization of the blue emission of NaYF₄:Yb,Tm upconverter materials has been achieved by the modification of synthesis conditions (re-

action time, stabilization temperature, and pH). Obtained systems were characterized by photoluminescence (PL), X-ray diffraction (XRD), and scanning and transmission electron microscopy (SEM and TEM). In addition, to determine their photophysical behavior, the best sample was characterized by time-resolved fluorescence and transient absorption spectroscopy (TAS).



Scheme 1. Representation of the methodology for photosensitized drug delivery based on cleaving the N-O covalent bond by means $\text{NaYF}_4:\text{Yb, Tm}$ UC nanoparticles.

2. Materials and Methods

2.1. Synthesis of Np-UCs

All $\text{NaYF}_4:\text{Yb/Tm}$ samples were prepared by solvothermal synthesis. $\text{Ln}(\text{NO}_3)_3$ was used as stock solution as it contains the trivalent ions of lanthanides for the designed UC. First, the corresponding metal oxide (Ln_2O_3) is dissolved in nitric acid at 40–50 °C while undergoing vigorous stirring. The resultant solution is mixed at room temperature for 15 min with oleic acid and ethanol, used as growth controller and solvent, respectively. Next, fluorine and sodium (NH_4F , NaOH) sources are added to the mixture drop by drop while stirring and the resulting solution stirred for 30 minutes. Next, the solution is introduced into an autoclave (reaction tank) into a conventional oven and then submitted to the solvothermal process at 230 °C, in order to ensure a minimum of 190 °C in the reaction tank. Finally, the UCs are washed multiple times using a mixture of ethanol/water in a centrifuge at 6000 rpm. Occasionally, additional thermal treatment where the UCs are calcined at 400 °C for 4 h is performed.

2.2. Synthesis of 7-(4-(tert-butoxycarbonyl)piperazin-1-yl)-1-cyclopropyl-6-fluoro-4-oxo-1,4-dihydroquinoline-3-carboxylic Acid (CF)

A modification of a reported method was used [55]. Ciprofloxacin (2 g, 6.0 mmol) and di-tert-butyl dicarbonate (1.4 g, 6.6 mmol) was dissolved in THF (60 mL). Next, a solution of sodium hydroxide (0.48 g, 12.0 mmol) in water (12 mL) was added, and the mixture stirred overnight at room temperature. After that time, the solvent was removed in vacuo, and the residue was taken into aqueous saturated ammonium chloride (200 mL). The aqueous phase was extracted with DCM (200 mL) three times. The organic phase was washed with brine (400 mL), dried over anhydrous MgSO_4 , and filtered (see Scheme S1). The solvent was evaporated in vacuo to obtain 2.51 g (97%) of white solid. $^1\text{H-NMR}$ (300 MHz, CDCl_3): δ 14.94 (s, 1H), 8.71 (s, 1H), 7.96 (d, $J = 12.9$ Hz, 1H), 7.35 (d, $J = 7.1$ Hz, 1H), 3.72–3.61 (m, 4H), 3.59–3.50 (m, 1H), 3.34–3.21 (m, 4H), 1.49 (s, 9H), 1.41–1.35 (m, 2H), and 1.26–1.15 (m, 2H).

Benzophenone oxime: Sodium acetate (0.50 g, 6.1 mmol) was dissolved in water (4 mL) and added over a solution of hydroxylamine hydrochloride (0.42 g, 6.1 mmol) in water (4 mL). The resulting mixture was added over benzophenone (1 g, 5.5 mmol) dissolved in ethanol (10 mL), and heated at 35 °C. Next, the sample was stirred for 8 h at 80 °C. After that time, the reaction was cooled at room temperature, and cold water was added to allow the precipitation of 1.04 g (96%) of the oxime as a white solid. $^1\text{H-NMR}$ (300 MHz, CDCl_3):

δ 7.5–7.43 (m, 7H) and 7.37–7.30 (m, 3H). $^1\text{H-NMR}$ spectrum in agreement with published data [56].

2.3. *Tert-butyl 4-(1-cyclopropyl-3-(((diphenylmethylene)amino)oxy)carbonyl)-6-fluoro-4-oxo-1,4-dihydroquinolin-7-yl)piperazine-1-carboxylate (CF-OE)*

Tert-butyl 4-(1-cyclopropyl-3-(((diphenylmethylene)amino)oxy)carbonyl)-6-fluoro-4-oxo-1,4-dihydroquinolin-7-yl)piperazine-1-carboxylate (CF-OE): To a solution of compound 1 (0.2 g, 0.46 mmol) in dry DCM (5 mL) under argon atmosphere, oxalyl chloride (0.17 g, 1.38 mmol) and one drop of DMF were added. The reaction was stirred for 2 h at 40 °C. After that time, the solvent was removed in vacuo to give 0.21 g (100%) of an orange solid that was immediately used in the next step. Benzophenone oxime (0.09 g, 0.46 mmol) and pyridine (74 μL , 0.92 mmol) were added to a solution of the acyl chloride prepared in the previous step (0.21 g, 0.46 mmol) in dry DCM (5 mL) under argon atmosphere. The reaction was stirred at room temperature for 4 h (see Scheme S1). Next, the solvent was removed in vacuum, and the resulting residue was purified by column chromatography (Hexane:THF 1:1), to give 0.17 g (59%) of white solid. $^1\text{H-NMR}$ (400 MHz, CDCl_3): δ 7.96 (s, 1H), 7.83 (d, $J = 13.2$ Hz, 1H), 7.62–7.58 (m, 2H), 7.53–7.45 (m, 5H), 7.44–7.39 (m, 1H), 7.37–7.31 (m, 2H), 7.16 (d, $J = 7.1$ Hz, 1H), 3.64–3.57 (m, 4H), 3.37–3.30 (m, 1H), 3.20–3.14 (m, 4H), 1.49 (s, 9H), 1.20–1.13 (m, 2H), and 0.94–0.87 (m, 2H). $^{13}\text{C-NMR}$ (100 MHz, CDCl_3): δ 172.6, 164.5, 161.3, 154.7, 151.7, 147.7, 144.3, 137.8, 135.0, 133.2, 130.8, 129.6, 129.2, 129.1, 128.4, 128.4, 122.8, 113.0, 108.2, 105.3, 80.2, 50.0, 43.6, 34.7, 28.5, and 8.1 (Figure S1). HR-MS ($[\text{M} + \text{H}]^+$): Calcd. for $\text{C}_{35}\text{H}_{35}\text{FN}_4\text{O}_5 + \text{H}$: 611.2664; Found: 611.2659 (Figure S2)

2.4. Characterization Methods

All prepared UCs were characterized by powder X-ray Diffraction (XRD) by Philips PW 3040/00 X'pert MPD/MRD, using $\text{CuK}\alpha$ radiation ($\lambda = 1.54178$ Å) at a scanning rate of f 0.2/s. SEM micrographs were obtained using a field emission scanning electron microscope (Hitachi TM-1000) and a high-resolution scanning electron microscope (Nova Nano SEM230). Transmission electron microscopy (Philips Tecnai 20 T) to characterize the absorption spectra of UCs a Perkin Elmer Lambda 1050 UV equipment was used. To characterize the emission spectra of the UCs, the compound was irradiated with a laser of 980 nm (Power 1 kW, model LD-WL206, input 85–264 V, 47–63 Hz 0.4 A) and its response was measured with a Fluorescence Spectrometer Perkin Elmer LS 55.

2.5. Photophysical Measurements

The solvents HPLC grades were purchased from Scharlab. Milli-Q[®] water was used for sample preparation. The stock solutions of CF and CF-OE were adjusted to 300 $\mu\text{g}/\text{mL}$ in CH_2Cl_2 . In the case of CF-OE, solutions containing 10% (v/v) of formic acid (FA) were added to protonate all the oxime. At this concentration of FA, the absorbance spectrum CF-OE suffers a hypsochromic shift.

UV-visible spectra for CF or CF-OE liquid samples were registered at room temperature using 10×10 mm² quartz cells with a spectrophotometer model Perkin Elmer Lambda 1050 UV/Vis/NIR. Steady-state fluorescence experiments for CF or CF-OE liquid samples were carried out with a Fluorescence Spectrometer Perkin Elmer LS 55, with an excitation wavelength of 330 nm and using a cut-off filter at 350 nm. Time-resolved (TR) fluorescence for CF or CF-OE liquid measurements were recorded in a Mini Tau system provided with a bandpass filter of 450 nm using an EPL-375 picosecond pulsed diode laser with emission at 372 nm as excitation source with a laser pulse width of 61 ps (both from Edinburgh Instruments, Livingston, UK). For monitoring the TR fluorescence of the solid powdered Np-UC, the powder sample was irradiated with a laser of 980 nm (Power 1 kW) in front-face mode, detecting the kinetic traces with a bandpass filter of 450 or 650 nm.

Transient Absorption Spectroscopy (TAS), i.e., laser flash photolysis measurements, were carried out with LP980 equipment from Edinburgh Instruments (LP980), based on an optical parametric oscillator (OPO) pumped by the third harmonic of an Nd:YAG laser (EKSPLA, Vilnius, Lithuania). The selected excitation wavelength for the measurements

was 355 nm with single low energy pulses of 1 mJ/pulse of ca. 5 ns duration, while a pulsed xenon flash lamp (150 W) was employed as a detecting light source. The probe light is dispersed through a monochromator (TMS302-A, grating 150 lines/mm) after it has passed the sample and then reaches a PMT detector (Hamamatsu Photonics, Hamamatsu Japan) to obtain the temporal profile. The absorbance of the CF of CF-OE liquid samples was kept at ~ 0.3 at $\lambda_{\text{exc}} = 355$ nm in dichloromethane. All transient spectra were recorded at room temperature using 10×10 mm² quartz cells, which were bubbled for 15 min with N₂ before acquisition. TAS signals for Np-UCF were monitored at $\lambda_{\text{exc}} = 980$ nm in the optical parametric oscillator mode in purged organic dispersed solutions.

2.6. Light-Induced Drug Delivered Experiments

A CH₂Cl₂ solution of CF-OE (0.5 mM, 300 $\mu\text{g}/\text{mL}$) containing 10% of FA was prepared. The pH of the solution was ca. 4. Simultaneously, 10 mg of NaYF₄:Yb,Tm (UC3) was deposited in an HPLC vial and 1 mL of the CF-OE solution was added. The resulted suspension was purged under N₂ for 15 min. Then, the solution was irradiated ($\lambda_{\text{exc}} = 980$ nm) at a distance of 5 cm for increasing periods (0–100 min). The solution was maintained by continuous stirring at room temperature. Then, aliquots of 30 μL of the suspended solution were diluted until 1 mL in acetonitrile and filtered before HPLC analysis. The amount of CF and CF-OE, as well as the potential release of the corresponding free CF upon the irradiation reaction, were determined using a reversed-phase HPLC Jasco LC-4000 series system, equipped with a PDA detector MD-4015 and a multisampler AS-4150 controlled by ChromNav software (Jasco Inc., Ashikaga, Japan). A Purple ODS reverse-phase column (5 μm , 4.6 \times 150 mm, Análisis Vínicos SL, Ciudad Real, Spain) was employed. For the quantification of all the chemical species, isocratic conditions were used. The flow rate was 1 mL/min, and the column temperature was fixed at 25 °C. In all cases, the injection volume was 10 μL . The mobile phase was based on a mixture of 70:50 H₂O:CH₃CN, containing 0.1% of FA. The retention times obtained from the analysis of CF or CF-OE were 4.9 and 10 min, respectively. The calibration curve was reached at the concentration of solutions standard of CF or CF-OE in the range of 0.5–30 $\mu\text{g}/\text{mL}$. In parallel, the reaction was monitored in-situ by TAS, in order to check the presence of free CF and Benzophenone as products formed.

3. Results and Discussion

3.1. Optimization of the Synthesis of NaYF₄:Yb/Tm UC Materials

In the first stage, optimization of the synthesis conditions such as reaction time, thermal treatment, and pH of the reaction (Table 1) was performed to determine optimal parameters to prepare NaYF₄:Yb/Tm UC materials with an improved emission on the blue-light range of the spectrum. These modifications lead to changes in structural and morphology properties that have an intrinsic relationship with luminescence properties.

Table 1. Synthesis conditions and crystal β phase proportion of the NaYF₄:Yb,Tm materials prepared in this study.

UC _i (i)	pH	Time (h)	Thermal Treatment (400 °C)	Proportion of β (%)
1	5	12	YES	56.9
2	5	24	YES	100
3	5	24	NO	100
4	6	24	NO	100
5	7	24	NO	100
6	8	24	NO	100
7	10	24	NO	100

Modifications in the synthesis time (UC1 & UC2) lead to minor differences in the visible luminescence spectra (Figure 1A). All samples show three main contributions at 480, 652, and 700 nm previously assigned in the literature to $^1\text{G}^4 \rightarrow ^3\text{H}^6$, $^1\text{G}^4 \rightarrow ^3\text{H}^4$, and

$^3F^3 \rightarrow ^3H^6$ transitions, respectively (see Figure 1A inset) [57], where the IR contribution show a decrease for short reaction times (UC1). On the other hand, the absence of thermal treatment (UC3) leads to a dramatic effect in emission spectrum (Figure 1A) with a huge increase of six times in the luminescence intensity for the 480 nm emission and the presence of two new bands at 360 nm and 456 nm attributed to $^1D^2 \rightarrow ^3H^6$ and $^1D^2 \rightarrow ^3H^4$ transitions, respectively (Figure 1A inset).

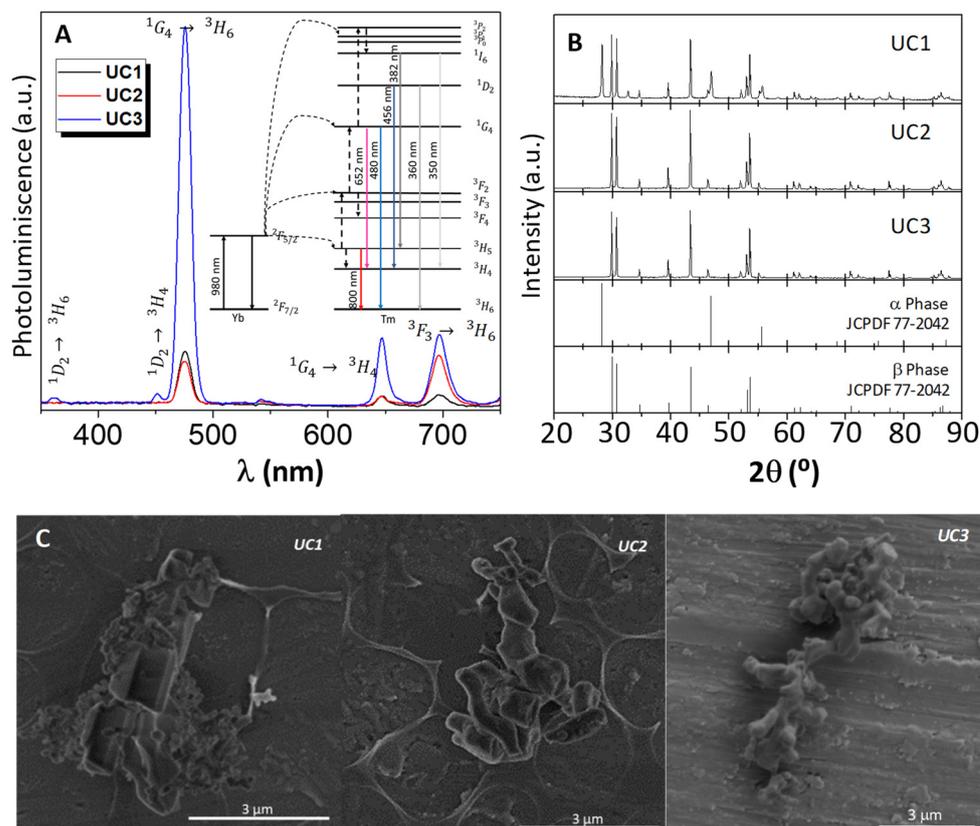


Figure 1. (A) Photoluminescence. Energy transitions scheme for $\text{NaYF}_4:\text{Yb,Tm}$ materials (Inset) and (B) X-ray diffraction, and (C) SEM photographs of UC1, UC2, and UC3 samples.

Regarding structural properties, lower reaction times (UC1) lead to a mixture of two crystal phases known as α or cubic-phase (43.1%) and β or hexagonal-phase (56.9%). Conversely, higher reaction times (UC2 & UC3) exhibit the formation of hexagonal one as a single phase (Figure 1B). This behavior is in concordance with previous studies that show that the β -phase is energetically more favorable [58]. Thus, for a shorter reaction time, the synthesis is kinetically controlled while a thermodynamic control is most favorable for a higher one [59]. Similar results were obtained in absence of thermal treatment also corroborating this fact. On the other hand, morphological properties studied by SEM (Figure 1C) and TEM images (Figure S3), exhibit that in the UC1 sample, two well-differentiated morphologies are observed: faceted crystals with the hexagonal habit (1.5–3 μm) corresponding to the β -phase and smaller irregular crystals with a globular shape (0.2–0.3 μm) assigned to the α -phase. An increase in the particle size of the β -phase (2–3.5 μm) is detected for a longer synthesis time (Figure 1B). In addition, the absence of thermal treatment leads to a decrease in the particle size of (0.8–1.2 μm) the β -phase which maintains the globular shape. These results indicate that the presence of a hexagonal phase with smaller particle size gives rise to an improvement in the luminescence yields in blue emissions and favors emission at lower wavelengths [60].

The pH modulation from 5 to 10 (UC3–UC7) leads to critical changes in the UC properties. Thus, for a pH higher than 6, the luminescence intensity of all emissions

dramatically decrease, being negligible for $\text{pH} \geq 7$ (Figure 2A). These changes in the synthesis media lead also to modifications in the structural and morphological properties. Although, XRD patterns show the presence of a β -phase in all cases (Figure 2B), SEM and TEM studies (Figures 2C and S4) show significant changes in the shape and size of UC materials with the pH increase. Thus, in the case of a basic environment (UC6–UC7), the materials show well-defined prismatic hexagonal single crystals with sizes between 1–2 μm long and 0.2–0.5 μm wide. On the other hand, for acidic and neutral media (UC3–UC5), the particles exhibit a globular shape with smaller sizes between 1–1.5 μm long and 0.1–0.3 μm wide. These changes in morphologic properties are observed with changes in the pH and are also attributed to the presence of NaOH. Previous studies proposed that Na^+ ions react with the oleic acid-forming oleates, which are miscible with the reaction medium favoring the precipitation of particle seeds in preferred orientation forming well-defined hexagonal crystals [61,62].

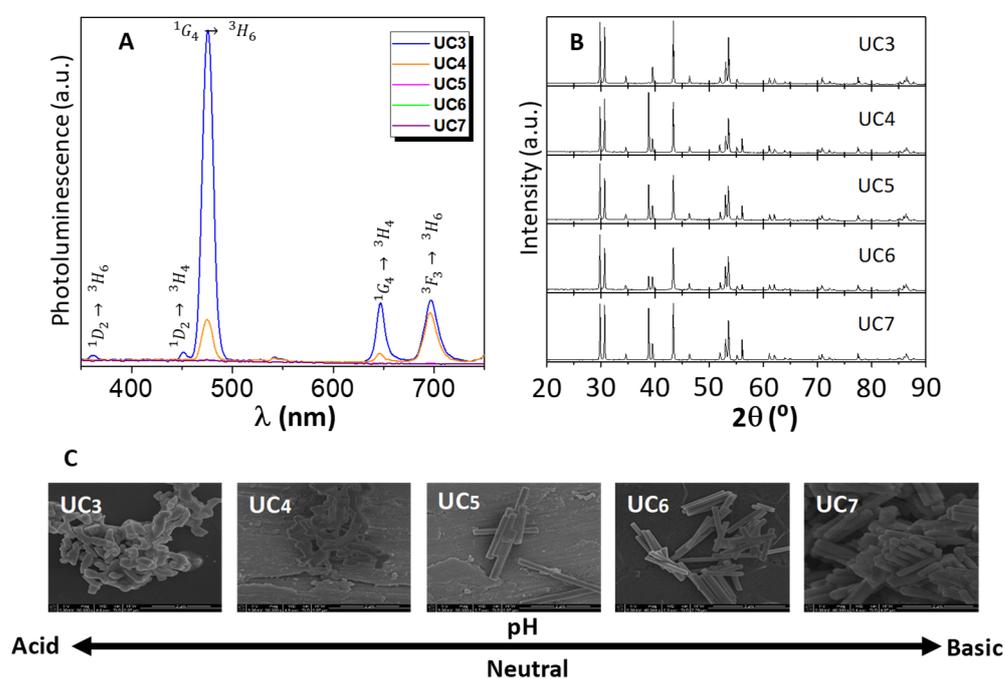


Figure 2. (A) Photoluminescence, (B) X-ray diffraction XRD patterns match closely with that of hexagonal β - NaYF_4 phase (JCPDF 28–1192), and (C) SEM photographs of UC3–UC7 samples under different pH synthesis conditions.

Therefore, changes in luminescence could be attributed to both structural and morphological properties. Thus, higher crystal sizes and crystallinity lead to a decrease in the emission properties of $\text{NaYF}_4:\text{Yb/Tm}$ [63].

3.2. Light-Driven Drug Delivery

The performance of $\text{NaYF}_4:\text{Yb}$ as photoactive materials for drug delivered therapies in Ciprofloxacin photosensitization reactions was explored. As abovementioned, the development of a better water-soluble derivative of CF is needed in order to successfully perform its delivery. In addition, this candidate should contain an adequate leaving group that could be easily removed after absorbing light. Moreover, in drug delivery experiments, it is crucial that the drug is efficiently released in the target to allow for drug absorption where its pharmacological effect is required. The absorption of CF progressively decreases from the stomach (100%) to the descending colon (5%) [64,65].

To fulfill these requirements, the oxime ester CF-OE (see chemical structure in Scheme 1) was chosen as a model compound that was obtained following the corresponding synthetic strategy (see details in Scheme S1). This prodrug CF-OE was found to be soluble in water

with a maximum absorption band at around 425 nm observed in acidic media due to its protonation (Figures 3A and S5). Thus, activation of this molecule by blue emission would afford the drug CF and the leaving group through the N–O bond cleavage. Interestingly, the emission spectrum of NaYF₄:Yb,Tm (UC3) showed a clear superposed signal with the protonated CF-OE absorption spectrum in the broad range of 460–500 nm (Figure 3A). This indicated that UC3 was a suitable system for photoinduced drug delivery.

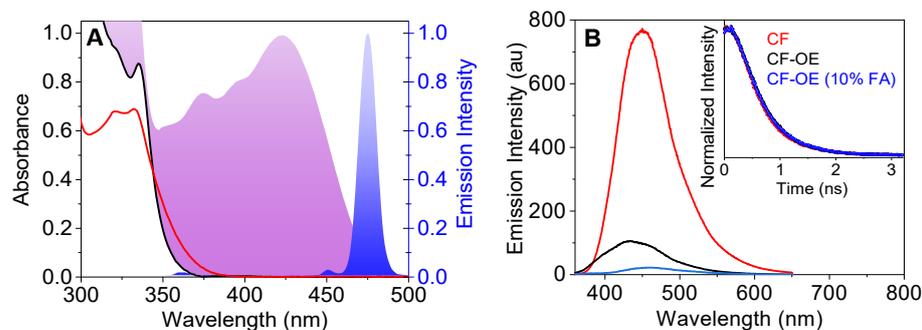


Figure 3. (A) Normalized absorption spectra of ciprofloxacin derivatives (CF or CF-OE, 20 μ M) in CH₂Cl₂ (black and red traces for CF or CF-OE, respectively) and in acidic media (purple shadow for CF-OE). The normalized photoluminescence spectrum of UC3 in solid (blue trace) is included for comparison. (B) Steady-state fluorescence for CF (red), CF-OE (black), and CF-OE in acidic media (blue) in dichloromethane. Inset: normalized decay traces.

To corroborate the suitability of this reaction, we also examined its photoluminescence properties. While CF exhibited a high fluorescence with the maximum at ca. 450 nm, the corresponding oxime-ester resulted in a poor intensity, accompanied by a blue-shift of the maximum (Figure 3B). For the protonated CF-OE (10% FA), the fluorescence was still lower, accompanied by a red-shift of the maximum up to 460 nm, as reported in the literature for commercial ciprofloxacin [66]. On the other hand, time-resolved emission (Figure 3B inset) shows identical kinetic traces ($\tau_F = 575$ ps) for both CF and CF-OE (including in acidic media). The transient absorption spectrum of CF (Figure S6A) showed the main contribution at 430 nm due to the fluorescence signal that avoids the accurate determination of the transient for CF. For this reason, TAS experiments for CF, CF-OE, and CF-OE in acidic media were monitored 100 ns after laser pulse (Figure S6B). These transients showed an identical absorption in the overall spectrum window, with a slight growth in the signal from 600 nm. The kinetic trace ($\lambda_{\text{mon}} = 650$ nm) showed a large lifetime (5.8 μ s) which could be assigned with a radical of CF formed after laser pulse (Figure S6B inset).

The controlled drug release process through photosensitized N–O bond cleavage in CF-OE was investigated upon selective irradiation of Np-UCs (UC3, $\lambda_{\text{exc}} = 980$ nm) in the presence of CF-OE in organic acidic media (Figure 4). First, to ensure accurate HPLC monitoring, the starting CF-OE and the corresponding free CF were tested and the calibration curve was reached at the concentration of solutions standard of CF or CF-OE in the range of 0.5–30 μ g/mL (Figure S7). In the light-driven drug delivery reaction, a remarkable increase in the formation of CF was observed at the first stages of the reaction accompanied by the corresponding decrease in the CF-OE chromatogram signal reaching a practically quantitative conversion (Figures 4A and S8). This behavior could be attributed to the N–O bond splitting, affording the free CF and the imine compound that immediately hydrolyzes to give the corresponding benzophenone (Figure S9). As expected, no changes were observed in absence of Np-UCs at optimal conditions.

Furthermore, the stability of the UC3 sample was also investigated by reusability experiments. After each experiment, the supernatant of the mixture solution was removed, and the recovered UC NPs were washed with CH₂Cl₂ several times. After that, an aliquot of the last supernatant was analyzed by UV-vis spectroscopy to check the presence of the

drugs. Figure 4B and Figure S10 show the UC3 activity during five consecutive cycles, with only a 3% loss of efficiency.

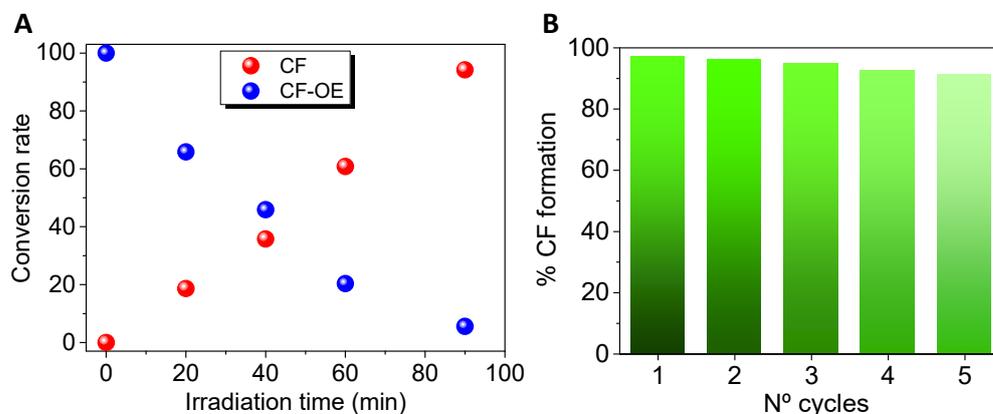


Figure 4. (A) Conversion rate of CF (red) or CF-OE (blue) using UC3 upon increasing irradiation times ($\lambda_{\text{exc}} = 980 \text{ nm}$), at 0.5 M of CF-OE in deaerated CH_2Cl_2 (10% FA *v/v*); (B) Reusability of UC3 by monitoring the conversion rate of the formation of CF upon increasing irradiation times ($\lambda_{\text{exc}} = 980 \text{ nm}$) during five consecutive cycles. The graph shows the maximum percentage of CF formation after 90 min under irradiation.

In addition, to gain further insight into the reaction mechanism, in-situ TAS experiments were performed in the presence of UC3 and released CF under continuous 980 nm irradiation. To monitor the performance of UC3 and drugs, TAS was conducted using two excitation wavelengths. At $\lambda_{\text{exc}} = 980 \text{ nm}$, a transient band at ca. 356 nm was detected (Figure 5A) which was safely ascribed to fundamental absorption from $^3\text{H}^6$ to $^1\text{D}^2$ excited states of Tm^{3+} [67]. This transient completely disappears upon 10 ns after the laser pulse and appears again when the experiments are repeated. On the other hand, at $\lambda_{\text{exc}} = 355 \text{ nm}$, the starting oxime and the released products were monitored. Since the identical TA signal was registered for free CF and CF-OE (Figure S6B), the reaction was followed by the formation of benzophenone (BP). The photolysis of BP generates the corresponding triplet excited state ($^3\text{BP}^*$) with a characteristic band centered at 520 nm which increased with the irradiation times (Figure 5B).

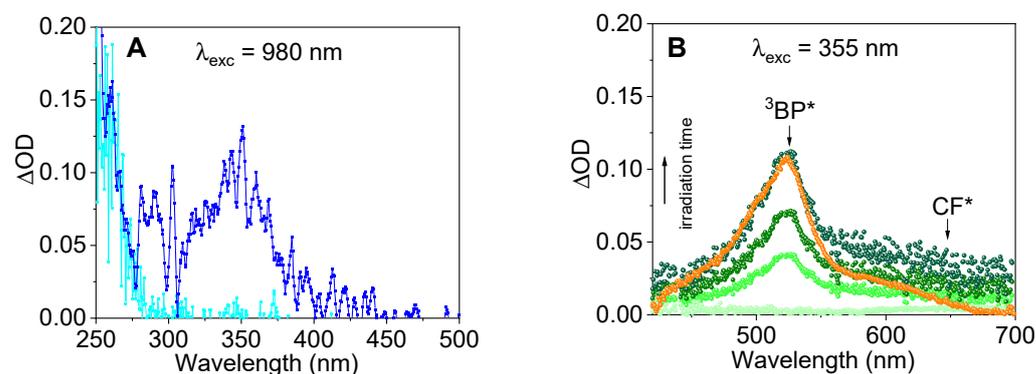


Figure 5. (A) Transient absorption spectra (TAS, $\lambda_{\text{exc}} = 980 \text{ nm}$) for UC3 in deaerated dichloromethane suspension solution immediately (blue trace) and after 10 ns (cyan trace) after the laser pulse. (B) Transient absorption spectra ($\lambda_{\text{exc}} = 355 \text{ nm}$) for the mixture reaction in deaerated methylene chloride (10% FA *v/v*) upon irradiation times. The orange trace corresponds to the commercial BP reference.

4. Conclusions

Here, we have demonstrated that $\text{NaYF}_4:\text{Yb},\text{Tm}$ up-converted material is an excellent candidate for blue light-driven drug release under near-IR wavelength. The emission

properties of these materials were improved through an optimization of the synthesis procedure (reaction time, thermal treatment, and pH conditions). The best blue-emission performance was achieved for the UC3 sample prepared through 24 h-synthesis without thermal treatment at a pH of 5. The enhancement of the blue-emission is attributed to the presence of the β -phase and smaller particle size.

Drug delivery experiments were performed through the highly effective photosensitization, by the means of NaYF₄:Yb,Tm up-converted material, of the N-O bond cleavage of the oxime ester of Ciprofloxacin (prodrug). Stability studies show that UC3 is a very stable and easily recovered material and can be used in several reaction cycles. This methodology can be extended to other drugs containing photoactive chromophores and is available as an alternative for drug delivery systems that require a controlled release of the active compounds at specific targets responsible for pharmacological action.

Supplementary Materials: The following are available online at <https://www.mdpi.com/article/10.3390/biomedicines9121953/s1>, Scheme S1: Synthetic route for the dyads CF (1) and CF-OE (2), Figure S1: ¹H (up) and ¹³C (down) NMR of CF-OE 2, Figure S2: Mass spectrometer characterization data of CF-OE 2, Figure S3: TEM images of (A) UC1; (B) UC2 and; (C) UC3, Figure S4: TEM images of (A) UC4; (B) UC5, (C) UC6 and; (D) UC7. Inset in Figure S2C correspond to electron diffraction of red squared area, Figure S5: Absorption spectra of CF-OE (0.5 mM) in the presence of increasing amounts of formic acid (% *v/v*). Inset: zoom image, Figure S6: (A) Transient absorption spectrum (TAS, $\lambda_{exc} = 355$ nm) for CF in deaerated dichloromethane solution immediately after the laser pulse and (B) upon 100 ns for CF (red), CF-OE (black) or CF-OE un acidic media (blue) in deaerated dichloromethane solutions. Inset: decay trace monitored at 650 nm for CF-OE as model, Figure S7: (A) and (B) HPLC chromatograms for CF (A) and CF-OE (B) upon increasing concentration in aerated 70:50 H₂O:CH₃CN, containing 0.1% of FA. Inset: corresponding linear calibration curve. (C) and (D) Calibration curve for CF (C) and CF-OE (D). The measurements were performed by triplicated for each compound, Figure S8: HPLC chromatograms of the evolution formation of free CF (red) and disappearance of CF-OE compound (blue) upon increasing irradiation times ($\lambda_{exc} = 980$ nm), at 0.5 M of CF-OE in deaerated methylene chloride (10% FA *v/v*). The mobile phase used in HPLC analysis was 70:50 H₂O:CH₃CN, containing 0.1% of FA, Figure S9: HPLC chromatograms of the formation of the benzophenone (BP) as a second product upon increasing irradiation times ($\lambda_{exc} = 980$ nm), at 0.5 mM of CF-OE in deaerated methylene chloride (10% FA *v/v*). The mobile phase used in HPLC analysis was 70:50 H₂O:CH₃CN, containing 0.1% of FA. The orange trace corresponds to the commercial BP reference, Figure S10: Reusability of UC3 by monitoring the conversion rate of the formation of CF upon increasing irradiation times ($\lambda_{exc} = 980$ nm) during five consecutive cycles. Cycle 1 to 5: black, red, blue, green, and purple, respectively.

Author Contributions: E.M.R.: Formal analysis, Investigation, and Writing—Original Draft; M.G.-M.: Formal analysis, Investigation, Writing—Original Draft, Review, and Editing; R.P.-R.: Conceptualizations and Review; B.P.: Formal analysis and Investigation; D.S.: Conceptualization, Writing, Review; A.C.: Conceptualization, Resources, and Supervision; V.A.d.I.P.O.: Conceptualization, Resources, Writing—Review and Editing, and Supervision. All authors have read and agreed to the published version of the manuscript.

Funding: This research was funded by the European Union's Horizon 2020 research and innovation program under European Research Council (ERC) through the HyMAP project, grant agreement No. 648319. Financial support was received from AEI-MICINN/FEDER, UE through the Nympha Project (PID2019-106315RB-I00). R.P.-R. thanks to the Generalitat Valenciana for financial support (CIDEGENT/2018/044).

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: The data presented in this study are available in the article.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Tiwari, G.; Tiwari, R.; Sriwastawa, B.; Bhati, L.; Pandey, S.; Pandey, P.; Bannerjee, S.K. Drug Delivery Systems: An Updated Review. *Int. J. Pharm. Investig.* **2012**, *2*, 2–11. [[CrossRef](#)]
2. Manzari, M.T.; Shamay, Y.; Kiguchi, H.; Rosen, N.; Scaltriti, M.; Heller, D.A. Targeted Drug Delivery Strategies for Precision Medicines. *Nat. Rev. Mater.* **2021**, *6*, 351–370. [[CrossRef](#)]
3. He, H.; Lu, Y.; Qi, J.; Zhu, Q.; Chen, Z.; Wu, W. Adapting Liposomes for Oral Drug Delivery. *Acta Pharm. Sin. B* **2019**, *9*, 36–48. [[CrossRef](#)]
4. Antimisiaris, S.G.; Marazioti, A.; Kannavou, M.; Natsaridis, E.; Gkartziou, F.; Kogkos, G.; Mourtas, S. Overcoming Barriers by Local Drug Delivery with Liposomes. *Adv. Drug Deliv. Rev.* **2021**, *174*, 53–86. [[CrossRef](#)] [[PubMed](#)]
5. Nuin, E.; Gómez-Mendoza, M.; Andreu, I.; Marin, M.L.; Miranda, M.A. New Photoactive Compounds to Probe Cholic Acid and Cholesterol inside Mixed Micelles. *Org. Lett.* **2013**, *15*, 298–301. [[CrossRef](#)]
6. Nuin, E.; Gomez-Mendoza, M.; Marin, M.L.; Andreu, I.; Miranda, M.A. Influence of Drug Encapsulation within Mixed Micelles on the Excited State Dynamics and Accessibility to Ionic Quenchers. *J. Phys. Chem. B* **2013**, *117*, 9327–9332. [[CrossRef](#)]
7. Rodriguez-Muñiz, G.M.; Gomez-Mendoza, M.; Nuin, E.; Andreu, I.; Marin, M.L.; Miranda, M.A. “Snorkelling” vs. “Diving” in Mixed Micelles Probed by Means of a Molecular Bathymeter. *Org. Biomol. Chem.* **2017**, *15*, 10281–10288. [[CrossRef](#)] [[PubMed](#)]
8. Bartelds, R.; Nematollahi, M.H.; Pols, T.; Stuart, M.C.A.; Pardakhty, A.; Asadikaram, G.; Poolman, B. Niosomes, an Alternative for Liposomal Delivery. *PLoS ONE* **2018**, *13*, e0194179. [[CrossRef](#)]
9. Gomez-Mendoza, M.; Nuin, E.; Andreu, I.; Marin, M.L.; Miranda, M.A. Photophysical Probes to Assess the Potential of Cholic Acid Aggregates as Drug Carriers. *J. Phys. Chem. B* **2012**, *116*, 10213–10218. [[CrossRef](#)]
10. Gomez-Mendoza, M.; Marin, M.L.; Miranda, M.A. Photoactive Bile Salts with Critical Micellar Concentration in the Micromolar Range. *Phys. Chem. Chem. Phys.* **2016**, *18*, 12976–12982. [[CrossRef](#)] [[PubMed](#)]
11. Gomez-Mendoza, M.; Marin, M.L.; Miranda, M.A. Dansyl Derivatives of Cholic Acid as Tools to Build Speciation Diagrams for Sodium Cholate Aggregation. *J. Phys. Chem. Lett.* **2011**, *2*, 782–785. [[CrossRef](#)]
12. Gomez-Mendoza, M.; Marin, M.L.; Miranda, M.A. Dansyl-Labeled Cholic Acid as a Tool to Build Speciation Diagrams for the Aggregation of Bile Acids. *J. Phys. Chem. B* **2012**, *116*, 14776–14780. [[CrossRef](#)] [[PubMed](#)]
13. Patra, J.K.; Das, G.; Fraceto, L.F.; Campos, E.V.R.; del Pilar Rodriguez-Torres, M.; Acosta-Torres, L.S.; Diaz-Torres, L.A.; Grillo, R.; Swamy, M.K.; Sharma, S.; et al. Nano Based Drug Delivery Systems: Recent Developments and Future Prospects. *J. Nanobiotechnology* **2018**, *16*, 71. [[CrossRef](#)] [[PubMed](#)]
14. Mitchell, M.J.; Billingsley, M.M.; Haley, R.M.; Wechsler, M.E.; Peppas, N.A.; Langer, R. Engineering Precision Nanoparticles for Drug Delivery. *Nat. Rev. Drug Discov.* **2021**, *20*, 101–124. [[CrossRef](#)]
15. Del Gaudio, C.; Crognale, V.; Serino, G.; Galloni, P.; Audenino, A.; Ribatti, D.; Morbiducci, U. Natural Polymeric Microspheres for Modulated Drug Delivery. *Mater. Sci. Eng. C* **2017**, *75*, 408–417. [[CrossRef](#)]
16. Chandan, R.; Banerjee, R. Pro-Apoptotic Liposomes-Nanobubble Conjugate Synergistic with Paclitaxel: A Platform for Ultrasound Responsive Image-Guided Drug Delivery. *Sci. Rep.* **2018**, *8*, 2624. [[CrossRef](#)]
17. Sherje, A.P.; Jadhav, M.; Dravyakar, B.R.; Kadam, D. Dendrimers: A Versatile Nanocarrier for Drug Delivery and Targeting. *Int. J. Pharm.* **2018**, *548*, 707–720. [[CrossRef](#)]
18. Patel, V.; Rajani, C.; Paul, D.; Borisa, P.; Rajpoot, K.; Youngren-Ortiz, S.R.; Tekade, R.K. Chapter 8—Dendrimers as Novel Drug-Delivery System and Its Applications. In *Advances in Pharmaceutical Product Development and Research*; Tekade, R.K., Ed.; Elsevier Inc.: Amsterdam, The Netherlands; Academic Press: Cambridge, MA, USA, 2020; pp. 333–392. ISBN 978-0-12-814487-9. [[CrossRef](#)]
19. Silindir Gunay, M.; Ozer, Y.; Chalon, S. Drug Delivery Systems for Imaging and Therapy of Parkinson’s Disease. *Curr. Neuropharmacol.* **2015**, *14*, 376–391. [[CrossRef](#)]
20. Zhai, B.; Zeng, Y.; Zeng, Z.; Zhang, N.; Li, C.; Zeng, Y.; You, Y.; Wang, S.; Chen, X.; Sui, X.; et al. Drug Delivery Systems for Elemene, Its Main Active Ingredient β -Elemene, and Its Derivatives in Cancer Therapy. *Int. J. Nanomed.* **2018**, *13*, 6279–6296. [[CrossRef](#)]
21. Porter, C.J.H.; Trevaskis, N.L.; Charman, W.N. Lipids and Lipid-Based Formulations: Optimizing the Oral Delivery of Lipophilic Drugs. *Nat. Rev. Drug Discov.* **2007**, *6*, 231–248. [[CrossRef](#)]
22. Hammad, M.A.; Müller, B.W. Increasing Drug Solubility by Means of Bile Salt–Phosphatidylcholine-Based Mixed Micelles. *Eur. J. Pharm. Biopharm.* **1998**, *46*, 361–367. [[CrossRef](#)]
23. Gao, W.; Chan, J.M.; Farokhzad, O.C. PH-Responsive Nanoparticles for Drug Delivery. *Mol. Pharm.* **2010**, *7*, 1913–1920. [[CrossRef](#)]
24. Ding, J.; Sun, Y.; Li, J.; Wang, H.; Mao, S. Enhanced Blood–Brain Barrier Transport of Vinpocetine by Oral Delivery of Mixed Micelles in Combination with a Message Guider. *J. Drug Target.* **2017**, *25*, 532–540. [[CrossRef](#)]
25. Lasic, D.D. Mixed Micelles in Drug Delivery. *Nature* **1992**, *355*, 279–280. [[CrossRef](#)]
26. Cheng, L.; Kamkaew, A.; Sun, H.; Jiang, D.; Valdovinos, H.F.; Gong, H.; England, C.G.; Goel, S.; Barnhart, T.E.; Cai, W. Dual-Modality Positron Emission Tomography/Optical Image-Guided Photodynamic Cancer Therapy with Chlorin E6-Containing Nanomicelles. *ACS Nano* **2016**, *10*, 7721–7730. [[CrossRef](#)] [[PubMed](#)]
27. Lipinski, C.A.; Lombardo, F.; Dominy, B.W.; Feeney, P.J. Experimental and Computational Approaches to Estimate Solubility and Permeability in Drug Discovery and Development Settings. *Adv. Drug Deliv. Rev.* **1997**, *23*, 3–25. [[CrossRef](#)]

28. Dikmen, G.; Genç, L.; Güney Eskiler, G. Advantage and Disadvantage in Drug Delivery Systems. *J. Mater. Sci. Eng.* **2011**, *5*, 468–472. [[CrossRef](#)]
29. Zhang, Y.; Chen, B.; Xu, S.; Li, X.; Zhang, J.; Sun, J.; Zheng, H.; Tong, L.; Sui, G.; Zhong, H.; et al. Dually Functioned Core-Shell NaYF₄:Er³⁺/Yb³⁺@NaYF₄:Tm³⁺/Yb³⁺ Nanoparticles as Nano-Calorifiers and Nano-Thermometers for Advanced Photothermal Therapy. *Sci. Rep.* **2017**, *7*, 11849. [[CrossRef](#)]
30. Dai, Y.; Ma, P.; Cheng, Z.; Kang, X.; Zhang, X.; Hou, Z.; Li, C.; Yang, D.; Zhai, X.; Lin, J. Up-Conversion Cell Imaging and PH-Induced Thermally Controlled Drug Release from NaYF₄:Yb³⁺/Er³⁺@Hydrogel Core-Shell Hybrid Microspheres. *ACS Nano* **2012**, *6*, 3327–3338. [[CrossRef](#)]
31. Hou, Z.; Li, C.; Ma, P.; Cheng, Z.; Li, X.; Zhang, X.; Dai, Y.; Yang, D.; Lian, H.; Lin, J. Up-Conversion Luminescent and Porous NaYF₄:Yb³⁺, Er³⁺@SiO₂ Nanocomposite Fibers for Anti-Cancer Drug Delivery and Cell Imaging. *Adv. Funct. Mater.* **2012**, *22*, 2713–2722. [[CrossRef](#)]
32. Wang, C.; Cheng, L.; Liu, Z. Drug Delivery with Upconversion Nanoparticles for Multi-Functional Targeted Cancer Cell Imaging and Therapy. *Biomaterials* **2011**, *32*, 1110–1120. [[CrossRef](#)] [[PubMed](#)]
33. Liu, Y.; Zhang, C.; Liu, H.; Li, Y.; Xu, Z.; Li, L.; Whittaker, A. Controllable Synthesis of Up-Conversion Nanoparticles UCNPs@MIL-PEG for PH-Responsive Drug Delivery and Potential Up-Conversion Luminescence/Magnetic Resonance Dual-Mode Imaging. *J. Alloys Compd.* **2018**, *749*, 939–947. [[CrossRef](#)]
34. Liu, Y.; Zhang, C.; Xu, C.; Lin, C.; Sun, K.; Wang, J.; Chen, X.; Li, L.; Whittaker, A.K.; Xu, H.-B. Controlled Synthesis of Up-Conversion Luminescent Gd/Tm-MOFs for PH-Responsive Drug Delivery and UCL/MRI Dual-Modal Imaging. *Dalton Trans.* **2018**, *47*, 11253–11263. [[CrossRef](#)]
35. Lim, S. Phototherapy and the Benefits of LEDs. *J. Soc. Inf. Disp.* **2011**, *19*, 882–887. [[CrossRef](#)]
36. Liu, Y.; Tu, D.; Zhu, H.; Chen, X. Lanthanide-Doped Luminescent Nanoprobes: Controlled Synthesis, Optical Spectroscopy, and Bioapplications. *Chem. Soc. Rev.* **2013**, *42*, 6924–6958. [[CrossRef](#)]
37. Gray, V.; Dzebo, D.; Abrahamsson, M.; Albinsson, B.; Moth-Poulsen, K. Triplet–Triplet Annihilation Photon-Upconversion: Towards Solar Energy Applications. *Phys. Chem. Chem. Phys.* **2014**, *16*, 10345–10352. [[CrossRef](#)]
38. Kerzig, C.; Wenger, O.S. Sensitized Triplet–Triplet Annihilation Upconversion in Water and Its Application to Photochemical Transformations. *Chem. Sci.* **2018**, *9*, 6670–6678. [[CrossRef](#)]
39. Zhou, Y.; Castellano, F.N.; Schmidt, T.W.; Hanson, K. On the Quantum Yield of Photon Upconversion via Triplet–Triplet Annihilation. *ACS Energy Lett.* **2020**, *5*, 2322–2326. [[CrossRef](#)]
40. Li, Z.; Wang, L.; Wang, Z.; Liu, X.; Xiong, Y. Modification of NaYF₄:Yb,Er@SiO₂ Nanoparticles with Gold Nanocrystals for Tunable Green-to-Red Upconversion Emissions. *J. Phys. Chem. C* **2011**, *115*, 3291–3296. [[CrossRef](#)]
41. Wang, L.; Qin, W.; Liu, Z.; Zhao, D.; Qin, G.; Di, W.; He, C. Improved 800 Nm Emission of Tm³⁺ Sensitized by Yb³⁺ and Ho³⁺ in β-NaYF₄ Nanocrystals under 980 Nm Excitation. *Opt. Express* **2012**, *20*, 7602–7607. [[CrossRef](#)]
42. Homann, C.; Krukewitt, L.; Frenzel, F.; Grauel, B.; Würth, C.; Resch-Genger, U.; Haase, M. NaYF₄:Yb,Er/NaYF₄ Core/Shell Nanocrystals with High Upconversion Luminescence Quantum Yield. *Angew. Chem. Int. Ed.* **2018**, *57*, 8765–8769. [[CrossRef](#)] [[PubMed](#)]
43. Ren, W.; Tian, G.; Jian, S.; Gu, Z.; Zhou, L.; Yan, L.; Jin, S.; Yin, W.; Zhao, Y. TWEEN Coated NaYF₄:Yb,Er/NaYF₄ Core/Shell Upconversion Nanoparticles for Bioimaging and Drug Delivery. *RSC Adv.* **2012**, *2*, 7037–7041. [[CrossRef](#)]
44. Tian, G.; Gu, Z.; Zhou, L.; Yin, W.; Liu, X.; Yan, L.; Jin, S.; Ren, W.; Xing, G.; Li, S.; et al. Mn²⁺ Dopant-Controlled Synthesis of NaYF₄:Yb/Er Upconversion Nanoparticles for in Vivo Imaging and Drug Delivery. *Adv. Mater.* **2012**, *24*, 1226–1231. [[CrossRef](#)]
45. Tu, D.; Liu, Y.; Zhu, H.; Li, R.; Liu, L.; Chen, X. Breakdown of Crystallographic Site Symmetry in Lanthanide-Doped NaYF₄ Crystals. *Angew. Chem. Int. Ed.* **2013**, *52*, 1128–1133. [[CrossRef](#)]
46. Shan, S.-N.; Wang, X.-Y.; Jia, N.-Q. Synthesis of NaYF₄:Yb³⁺, Er³⁺ Upconversion Nanoparticles in Normal Microemulsions. *Nanoscale Res. Lett.* **2011**, *6*, 539. [[CrossRef](#)]
47. D’Vries, R.F.; de la Peña-O’Shea, V.A.; Snejko, N.; Iglesias, M.; Gutiérrez-Puebla, E.; Monge, M.A. Insight into the Correlation between Net Topology and Ligand Coordination Mode in New Lanthanide MOFs Heterogeneous Catalysts: A Theoretical and Experimental Approach. *Cryst. Growth Des.* **2012**, *12*, 5535–5545. [[CrossRef](#)]
48. D’Vries, R.F.; de la Peña-O’Shea, V.A.; Snejko, N.; Iglesias, M.; Gutiérrez-Puebla, E.; Monge, M.A. H₃O₂ Bridging Ligand in a Metal–Organic Framework. Insight into the Aqua-Hydroxo↔Hydroxyl Equilibrium: A Combined Experimental and Theoretical Study. *J. Am. Chem. Soc.* **2013**, *135*, 5782–5792. [[CrossRef](#)] [[PubMed](#)]
49. Quesada, M.; Monrabal, M.; Aromí, G.; de la Peña-O’Shea, V.A.; Gich, M.; Molins, E.; Roubeau, O.; Teat, S.J.; MacLean, E.J.; Gamez, P.; et al. Spin transition in a triazine-based Fe(II) complex: Variable-temperature structural, thermal, magnetic and spectroscopic studies. *J. Mater. Chem.* **2006**, *16*, 2669–2676. [[CrossRef](#)]
50. Jain, D.; Banerjee, R. Comparison of Ciprofloxacin Hydrochloride-Loaded Protein, Lipid, and Chitosan Nanoparticles for Drug Delivery. *J. Biomed. Mater. Res. Part B Appl. Biomater.* **2008**, *86B*, 105–112. [[CrossRef](#)] [[PubMed](#)]
51. Kloskowski, T.; Gurtowska, N.; Bajek, A.; Drewa, T. Ciprofloxacin as a Prophylactic Agent against Prostate Cancer: A “Two Hit” Hypothesis. *Med. Hypotheses* **2012**, *78*, 235–238. [[CrossRef](#)] [[PubMed](#)]
52. Kloskowski, T.; Gurtowska, N.; Olkowska, J.; Nowak Marcin, J.; Adamowicz, J.; Tworkiewicz, J.; Dębski, R.; Grzanka, A.; Drewa, T. Ciprofloxacin Is a Potential Topoisomerase II Inhibitor for the Treatment of NSCLC. *Int. J. Oncol.* **2012**, *41*, 1943–1949. [[CrossRef](#)] [[PubMed](#)]

53. Geller, L.T.; Barzily-Rokni, M.; Danino, T.; Jonas, O.H.; Shental, N.; Nejman, D.; Gavert, N.; Zwang, Y.; Cooper, Z.A.; Shee, K.; et al. Potential Role of Intratumor Bacteria in Mediating Tumor Resistance to the Chemotherapeutic Drug Gemcitabine. *Science* **2017**, *357*, 1156–1160. [[CrossRef](#)]
54. Straussman, R.; Morikawa, T.; Shee, K.; Barzily-Rokni, M.; Qian, Z.R.; Du, J.; Davis, A.; Mongare, M.M.; Gould, J.; Frederick, D.T.; et al. Tumour Micro-Environment Elicits Innate Resistance to RAF Inhibitors through HGF Secretion. *Nature* **2012**, *487*, 500–504. [[CrossRef](#)]
55. Ji, C.; Miller, P.A.; Miller, M.J. Syntheses and Antibacterial Activity of N-Acylated Ciprofloxacin Derivatives Based on the Trimethyl Lock. *ACS Med. Chem. Lett.* **2015**, *6*, 707–710. [[CrossRef](#)] [[PubMed](#)]
56. Liu, S.; Yu, Y.; Liebeskind, L.S. N-Substituted Imines by the Copper-Catalyzed N-Imination of Boronic Acids and Organostannanes with O-Acyl Ketoximes. *Org. Lett.* **2007**, *9*, 1947–1950. [[CrossRef](#)]
57. Kannan, P.; Rahim, F.A.; Teng, X.; Chen, R.; Sun, H.; Huang, L.; Kim, D.-H. Enhanced Emission of NaYF₄:Yb,Er/Tm Nanoparticles by Selective Growth of Au and Ag Nanoshells. *RSC Adv.* **2013**, *3*, 7718–7721. [[CrossRef](#)]
58. Zeng, S.; Ren, G.; Xu, C.; Yang, Q. High Uniformity and Monodispersity of Sodium Rare-Earth Fluoride Nanocrystals: Controllable Synthesis, Shape Evolution and Optical Properties. *Cryst. Eng. Comm.* **2011**, *13*, 1384–1390. [[CrossRef](#)]
59. Li, C.; Quan, Z.; Yang, J.; Yang, P.; Lin, J. Highly Uniform and Monodisperse Beta-NaYF₄:Ln(3+) (Ln = Eu, Tb, Yb/Er, and Yb/Tm) Hexagonal Microprism Crystals: Hydrothermal Synthesis and Luminescent Properties. *Inorg. Chem.* **2007**, *46*, 6329–6337. [[CrossRef](#)] [[PubMed](#)]
60. Liu, C.; Wang, H.; Li, X.; Chen, D. Monodisperse, Size-Tunable and Highly Efficient β -NaYF₄:Yb,Er(Tm) up-Conversion Luminescent Nanospheres: Controllable Synthesis and Their Surface Modifications. *J. Mater. Chem.* **2009**, *19*, 3546–3553. [[CrossRef](#)]
61. Li, C.; Yang, J.; Quan, Z.; Yang, P.; Kong, D.; Lin, J. Different Microstructures of β -NaYF₄ Fabricated by Hydrothermal Process: Effects of PH Values and Fluoride Sources. *Chem. Mater.* **2007**, *19*, 4933–4942. [[CrossRef](#)]
62. Lin, M.; Zhao, Y.; Liu, M.; Qiu, M.; Dong, Y.; Duan, Z.; Li, Y.H.; Pingguan-Murphy, B.; Lu, T.J.; Xu, F. Synthesis of Upconversion NaYF₄:Yb³⁺,Er³⁺ Particles with Enhanced Luminescent Intensity through Control of Morphology and Phase. *J. Mater. Chem. C* **2014**, *2*, 3671–3676. [[CrossRef](#)]
63. Lutz, T.; Veissier, L.; Thiel, C.; Cone, R.; Barclay, P.; Tittel, W. Modification of Phonon Processes in Nano-Structured Rare-Earth-Ion-Doped Crystals. *Phys. Rev. A* **2015**, *94*, 013801. [[CrossRef](#)]
64. Harder, S.; Fuhr, U.; Beermann, D.; Staib, A.H. Ciprofloxacin absorption in different regions on the human gastrointestinal tract. Investigations with the hf-capsule. *Br. J. Clin. Pharmacol.* **1990**, *30*, 35–39. [[CrossRef](#)] [[PubMed](#)]
65. Staib, A.H.; Beermann, D.; Harder, S.; Fuhr, U.; Liermann, D. Absorption differences of ciprofloxacin along the human gastrointestinal tract determined using a remote-control drug delivery device (HF-capsule). *Am. J. Med.* **1989**, *87*, 566–569.
66. Yang, R.; Fu, Y.; Li, L.-D.; Liu, J.-M. Medium Effects on Fluorescence of Ciprofloxacin Hydrochloride. *Spectrochim. Acta Part A Mol. Biomol. Spectrosc.* **2003**, *59*, 2723–2732. [[CrossRef](#)]
67. Halubek-Gluchowska, K.; Szymański, D.; Tran, T.N.; Ferrari, M.; Lukowiak, A. Upconversion Luminescence of Silica–Calcium Nanoparticles Co-Doped with Tm³⁺ and Yb³⁺ Ions. *Materials* **2021**, *14*, 937. [[CrossRef](#)]