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Additional Information

Constraints on Persistent Activity in a Biologically Detailed Network Model of the Prefrontal Cortex with Heterogeneities

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Abstract

Persistent activity, the maintenance of neural activation over short periods of time in cortical networks, is widely thought to underlie the cognitive function of working memory. A large body of modeling studies has reproduced this kind of activity using cell assemblies with strengthened synaptic connections. However, almost all of these studies have considered persistent activity within networks with homogeneous neurons and synapses, making it difficult to judge the validity of such model results for cortical dynamics, which is based on highly heterogeneous neurons. Here, we consider persistent activity in a detailed, strongly data-driven network model of the prefrontal cortex with heterogeneous neuron and synapse parameters. Surprisingly, persistent activity could not be reproduced in this model without incorporating further constraints. We identified three factors that prevent successful persistent activity: heterogeneity in the cell parameters of interneurons, heterogeneity in the parameters of short-term synaptic plasticity and heterogeneity in the synaptic weights. Our model predicts that persistent activity is recovered if the heterogeneity in the activity of individual interneurons is diminished, which could be achieved by a homeostatic plasticity mechanism. Such a plasticity scheme could also compensate the heterogeneities in the synaptic weights and short-term plasticity when applied to the inhibitory synapses. Cell assemblies shaped in this way may be potentially targeted by distinct inputs or become more responsive to specific tuning or spectral properties. Furthermore, the model predicts that a network that exhibits persistent activity is not able to dynamically produce intrinsic *in vivo*-like irregular activity at the same time, because heterogeneous synaptic connections are required for these dynamics. Thus, the background noise in such a network must either be produced by external input or constitutes an entirely different state of the network, which is brought about, e.g., by neuromodulation.

Author summary

To operate effectively in a constantly changing world, it is crucial to keep relevant information in mind for short periods of time. This ability, called working memory, is commonly assumed to rest on reverberating activity among members of cell assemblies. While effective in reproducing key results of working memory, most cell assembly models rest on major simplifications such as using the same parameters for all neurons and synapses, i.e., assuming homogeneity in these parameters. Here, we show that this homogeneity assumption is necessary for persistent activity to arise, specifically for inhibitory interneurons and synapses. Using a strongly data-driven network model of the prefrontal cortex, we show that the heterogeneities in the above parameters that are implied by *in vitro* studies prevent persistent activity. When homogeneity is imposed on inhibitory neurons and synapses, persistent activity is recovered. We propose that the homogeneity constraints can be implemented in the brain by means of homeostatic plasticity, a form of learning that keeps the activity of a network in a constant, homeostatic state. The model makes a number of predictions for biological networks, including a structural separation of networks responsible for generating persistent activity and spontaneous, noise-like activity.

Introduction

Working memory can be defined as the ability to maintain a representation of a sensory stimulus, a thought or a memory retrieved from long-term storage over a period of time, even after the source of this representation is gone [1]. Working memory is a crucial prerequisite for many cognitive functions such as planning, reasoning, goal-directed behavior or language comprehension. It is also among the most commonly distorted cognitive functions in neurological and psychiatric disorders such as schizophrenia [2,3]. A neural basis of working memory is widely believed to be persistent activity in cortical neurons, i.e., elevated firing rates that emerge during synaptic stimulation that persist after the input is removed [4–6]. This type of activity has been consistently found during working memory tasks in electrophysiological recordings [4,7–9] as well as imaging recordings from humans [5,10], most prominently in the prefrontal cortex (PFC). Furthermore, the stability of persistent activity is often strongly correlated with behavioral performance, i.e., if activity does not persist, animals are much more likely to make an error [4,7].

The prominent role of persistent activity in working memory and cognition has inspired a large number of modeling studies that aim to understand the neural mechanisms underlying persistent activity [4,6,11]. The most widely studied of these mechanisms is reverberating synaptic activity in so-called attractor networks or cell assemblies, sets of neurons that are more strongly interconnected to each other compared to the average connectivity. This mechanism has been implemented in models of different levels of biological abstraction [4,11–14]. In this study, we focus on attractor network models that rely on anatomically and physiologically stable assemblies, which may either be shaped by Hebbian learning or by proximity in space (either physical or defined by stimulus features), as opposed to more recent proposals of transient activations of cellular subsets [11], e.g., based on short-term synaptic modifications [15].

An important simplification in most of the existing models of persistent activity is disregard of the substantial heterogeneity of functional properties of neurons and synapses observed in the living brain, i.e., using the same set of parameters for all the simulated entities of the same type. Few studies have investigated the impact of incorporating heterogeneities into working memory models. The existing ones have focused on the ring model [16], for spatial working memory and heterogeneities in the parameters of pyramidal cells and their synaptic connections. Results suggest that random heterogeneities in the model cause the representation to drift to other spatial locations [17–19], while spatially structured heterogeneities [20], homeostatic 10

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regulation of pyramidal cell activity [17] and short-term synaptic plasticity [18, 19] can prevent this drift.

In this work, we investigate persistent activity of single cell assemblies within a biophysically detailed spiking network model of the PFC that is strongly constrained by in vitro data, including cellular and synaptic heterogeneities [21]. We find that these naturally occurring heterogeneities abolish bistability that is necessary for stimulus-dependent persistent activity. Thus, persistent activity itself is lost, instead of drifting away from the encoded location as in the ring model. Three aspects of heterogeneity are found to be harmful for persistent activity: Heterogeneous excitability of the interneurons (but not of the pyramidal cells, as in previous studies), heavy tails in the distribution of synaptic inputs [22] and heterogeneous short-term plasticity, i.e. different types of plasticity (facilitating, depressing or both) in synapses between neurons of the same type. We also show that persistent activity is recovered when the heterogeneities in the interneurons are compensated, e.g., by homeostatic plasticity [23, 24]. The same plasticity scheme could also compensate the heterogeneities in the synaptic weights and short-term plasticity when applied to the inhibitory synapses. However, as an important consequence of the homogeneity requirement for persistent activity, we show that cell assembly neurons cannot dynamically produce the often observed irregular ground state on their own: The dynamics that stabilize persistent activity are mutually exclusive with those dynamics necessary for the generation of irregular behavior. Instead, the assemblies may inherit these noise-like properties from input from other subnetworks or change its dynamic mode, e.g., using neuromodulation.

Results

No persistent activity in the detailed network model

Stimulus-dependent persistent activity is routinely seen in a wide variety of computational models that implement cell assemblies, sets of neurons that either innervate each other with a more dense connectivity or stronger synaptic weights. Mathematically, these assemblies show bistable behavior: In the absence of a stimulus, the firing rates of their member neurons are low, and there is no difference from the neurons outside the assembly (also called the spontaneous activity, e.g., [25]). When a stimulus is given to the neurons of the assembly, however, the strong activation triggers a positive feedback loop among the assembly neurons, which extends a high-rate persistent activity over time due to the long time constant of the NMDA currents. We attempted to generate persistent activity in the same way in a biologically validated model we recently proposed [21], which comprises broad, multivariate cell parameter distributions derived from *in vitro* recordings, laminar anatomy and experimentally constrained synaptic dynamics (see Methods for details). Surprisingly, we did not find the bistable behavior that is necessary for persistent activity as observed in vivo. Depending on the parameters of the model, either the spontaneous or the persistent activity state is stable, while the other one is not. Thus, the network either relaxes back to spontaneous activity after a transient reaction to the stimulus (Figure 1A) or persistent activity is present over the entire simulation time, only mildly modulated by the stimulus (Figure 1B). We varied a number of model parameters that are likely to affect persistent activity, such as the baseline synaptic weights between pyramidal cells and interneurons, synaptic weights and connectivity within the assembly (using scaling factors s_{CA} for weights and p_{CA} for connection probabilities), background currents into the pyramidal cells (I_{ex}) and interneurons (I_{inh}) , as well as the size of the assembly $N_{\rm CA}$ and the ratio between the peak conductance of NMDA and AMPA

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currents, which is originally set to 1.09 [26,27] (see Methods for a full overview over all parameter variations). Almost none of these manipulations led to persistent activity (with a single exception, which requires unphysiological levels of NMDA currents and puts the entire network in an epileptiform state with extremely high firing rates). Figure 1C shows the normalized activity in the cell assembly (see Methods for details) before the stimulus presentation (blue) and for the last 500 ms of the simulation (red). The results are sorted for ascending values of the activity before the stimulus. One can see that all possible activity values are covered, and the activity values at the end of the simulation hover around those before stimulation, i.e. activity that is low before stimulation stays low afterwards (Figure 1A), while high activity levels at the end of the simulation are already generated before stimulus presentation (Figure 1B), reflecting a continuum of monostable states.

Fig 1. Failed persistent activity in the original network. A: Raster plot (upper panel) and instantaneous firing rate (lower panel) of layer 2/3 of the network, with pyramidal cells in red (cell assembly members) and black (neurons outside the assembly) and interneurons in blue. The red bar denotes a stimulus to the assembly neurons. Activity in the cell assembly quickly decays back to baseline after a few hundred ms. B: Raster plot as in A. Activity in the assembly persists, but arises spontaneously, i.e., not in reaction to the external stimulus. C: Summary of normalized activity in the cell assembly (see Methods for details) before stimulus presentation (blue) and at the end of the simulation (red) for 300 simulations with different parameters (assembly size between 30 and 80 cells). Simulations are sorted by ascending values of activity before the stimulus.

The remainder of the paper aims to identify how bistability can be introduced in a physiologically realistic network configuration. To differentiate bistability-induced persistent activity from the monostable states described above (Figure 1A and B), we introduce the measure d_{PA} , which is defined as the difference between the normalized activity in the cell assembly after and before the stimulus, ranging between zero and one (d_{PA} is set to zero for negative differences, i.e. higher activity before the stimulus, see Methods for details). d_{PA} is close to zero in all of the monostable cases (largely constant firing rates over time) and increases only if the stimulus induces a persistent high-activity state, i.e., in the bistable case.

Constraints for persistent activity

The above results (Figure 1) show that none of the simulations exploring a wide range 108 of parameter combinations for the biologically validated network model was able to 109 generate realistic persistent activity. This is a surprising result, as simpler models 110 easily incorporated persistent activity in cell assemblies. Thus, we hypothesized that 111 some of the biological complexity that is mimicked in the present model, but was 112 missing in others, prevents persistent activity. To test this, we simplified the model to 113 see which of the added features cause the problem. Most notably, we reduced the 114 network to contain pyramidal cells and fast-spiking interneurons in layer 2/3 only (i.e., 115 lacking layer 5 and four other types of interneurons, see Methods for details), used a 116 uniform connection probability of 10%, removed short-term plasticity and reduced the 117 input currents to the mean rheobase of each neuron type, which strongly reduced 118 spontaneous firing. Furthermore, we made the neuron parameters of a given cell type 119 much more uniform, reducing its standard deviation to 5% of the original value. These 120 simplifications made the model more similar to previously studied models of persistent 121 activity, in particular the Brunel and Wang model [25] (we also tested a configuration 122 that completely mimicked that model, but results did not qualitatively change). 123

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Most of these manipulations did not improve the ability of the network to produce 124 persistent activity, with three exceptions: The reduction of the standard deviation of 125 the neuron parameters, the removal of short-term plasticity and the use of a uniform 126 connectivity across cell types. Each of these manipulations represents aspects of 127 heterogeneity and variability of the network: Heterogeneity in the neuron parameters, 128 the synaptic connectivity and the dynamics of synaptic response. In the following, we 129 will investigate how these three aspects of heterogeneity relate to persistent activity 130 and how they can be compensated in a biologically meaningful way. 131

Low-rheobase interneurons suppress persistent activity First, we varied the amount of heterogeneity in the neuron parameters by jointly decreasing the standard deviations of the parameter distributions obtained from *in vitro* recordings to a given fraction of their original values for all parameters. Figure 2A shows the persistent activity measure d_{PA} as a function of cell parameter variability. Persistent activity emerges when variability is reduced to about 25% or less of the original value. This is indicated by values of d_{PA} above 0.3 (dotted line in Figure 2A), which has proven a good demarcation between successful and failed persistent activity using visual inspection of the raster plots, and will be used in this way throughout the paper. The exact choice of this demarcation is not critical for the results. To understand why the

Fig 2. Breaking persistent activity by interneuron heterogeneity. A: $d_{\rm PA}$ as a function of the variability in neuron parameters in pyramidal cells only (red), interneurons only (blue) or all neurons (black). B: Persistent activity measure $d_{\rm PA}$ as individual parameters are increased back to 100% of their original variability, with all other parameters at 5%. The leftmost bar shows baseline $d_{\rm PA}$ if all parameters' variability is left at 5%. The dashed line demarks the boundary between successful ($d_{\rm PA} \ge 0.3$) and failed ($d_{\rm PA} < 0.3$) persistent activity. C: $d_{\rm PA}$ in simulations with interneuron variability set to 5% as increasing number of interneurons are replaced by neurons at the original variability, but only with rheobases above (blue) or below (red) the mean rheobase. Background currents are adjusted accordingly (see text for details). The dashed line demarks $d_{\rm PA} = 0.3$ as in A.

141 naturally occurring heterogeneity in the neuron parameters [21,28] prevents persistent 142 activity, we first analyzed the effect of individual cell types and neuron parameters, in 143 particular the rheobase, the minimal amount of input current that elicits an action 144 potential. Figure 2A shows the persistent activity measure $d_{\rm PA}$ as the cellular 145 variability of pyramidal cells or interneurons were varied, respectively, while the other 146 cells were left at their original parameter distribution. It is apparent that only the 147 heterogeneity in the interneurons affects persistent activity (blue curve), while 148 pyramidal cell variability does not have a noticeable effect, either when varied alone 149 (red curve) or in concert with interneuron variability (black vs. blue curve). 150

To differentiate the effects of the individual neuron parameters of the interneurons, 151 we started from a simulation where the variability of all parameters was set to 5% of 152 their original values (Figure 2B, left bar, $d_{\rm PA} = 0.60 \pm 0.02$; mean \pm SEM). Then, we 153 replaced the reduced distribution of each of the nine independent model parameters of 154 the neuron model (the simplified exponential integrate and fire neuron, or 155 simpAdEx [28]) by the full distribution, one at a time. d_{PA} significantly decreased 156 relative to baseline for each of these manipulations, but mostly staved above 0.3, 157 indicating successful persistent activity (dotted line in Figure 2B). However, $d_{\rm PA}$ 158 dropped to values of almost zero when the original heterogeneity was used in the 159 parameters $V_{\rm th}$ (onset of exponential upswing in the membrane potential, $d_{\rm PA} = 0.0$ 160 for all simulations) and E_L (reversal potential of the leak current, $d_{\rm PA} = 0.01 \pm 0.01$; 161 mean \pm SEM). 162

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The parameters $V_{\rm th}$ and E_L that proved most important in the above analysis are 163 the major determinants of the rheobase current in the simpAdEx [28]. Thus, we 164 hypothesized that interneurons with low rheobase and high spontaneous firing rate 165 will prevent the initiation of persistent activity. To test this hypothesis, we set up a 166 simulation with two populations of interneurons, one using the original distribution 167 and another with the variability of all parameters reduced to 5%. We first drew all 168 interneurons from the reduced distribution and then replaced increasing numbers of 169 interneurons by those drawn from the original distribution and computed the $d_{\rm PA}$ 170 measure in each of these cases. Importantly, we replaced only neurons with rheobases 171 above the average in one set of simulations (blue curve in Figure 2C) and only those 172 with rheobases below the average in another one (red curve in Figure 2C). Background 173 currents were adjusted such that the input relative to the mean rheobase was kept 174 constant as neurons were replaced, so the effective mean input into the interneurons 175 are not changed. While the replacement of neurons above rheobase does not affect 176 persistent activity, d_{PA} drops as the number of replaced neurons below rheobase is 177 increased. This implies that interneurons with rheobases below the population average 178 dominate inhibition and prevent the initiation of persistent activity. Importantly, the 179 adjustment of the (constant) background current was not sufficient to compensate the 180 increased inhibitory activity. This suggests that the low-rheobase interneurons may 181 induce a skew in the distribution of inhibitory activity rather than just shifting its 182 mean. 183

Homeostatic compensation for rheobase heterogeneity recovers persistent **activity** Persistent activity occurs in prefrontal cortex where cells are highly heterogeneous, so it is important to explain how persistent activity and cellular heterogeneity can be reconciled in a model of the prefrontal cortex. As we identified high spontaneous firing rates in low-rheobase interneurons as the main limiting factor for the initiation of persistent activity, the effect of these interneurons on total inhibition needs to be limited in a biologically realistic way. A potential mechanism limiting the maximal inhibition is homeostatic scaling of synaptic weights that aims to keep roughly uniform firing rates. This kind of homeostatic plasticity has been observed in both in vivo and in vitro studies and has been shown to regulate both excitatory and inhibitory synapses [24]. We emulate this homeostatic regulation by adjusting the excitatory background inputs (which are meant to represent the average synaptic input from outside the simulated network) individually for each interneuron such that the effective drive (input current minus rheobase) is the same for each interneuron. External input constitutes the bulk of the excitatory input to each cell, so extending the homeostasis to all excitatory input does not significantly change the results. With this adjustment, the distribution of firing rates in the network peaks at the mean rheobase, while previously, it was strongly skewed towards low-rheobase interneurons at high rates (Figure 3A). Furthermore, persistent activity emerges as the background currents are slightly decreased compared to the original value of 37pA (blue curve in Figure 3B). Importantly, the same manipulation lacks any effect without rheobase compensation (red curve). Thus, homeostatic plasticity is a possible way to compensate rheobase heterogeneity and allow for persistent activity.

The role of short-term plasticity The simulations shown in Figure 2 and 3 were conducted without short-term synaptic plasticity (STP). Here, we investigate the how the incorporation of STP affects persistent activity. STP refers to the dynamic change of synaptic efficacies in response to incoming spikes, which relaxes back to a baseline within several hundred milliseconds [29] (see Methods for details). This short time scale sets STP apart from long-term plasticity, which induces lasting changes in the

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Fig 3. Compensation of rheobases restores persistent activity. A: Average interneuron firing rate as a function of rheobase with (blue) and without (red) background currents adapted to rheobases (mean \pm SEM over all neurons). B: Persistent activity measure d_{PA} as a function of the background current into interneurons with (blue) and without (red) compensation of rheobases by adapting the background currents such that the input relative to each individual rheobase is constant (mean \pm SEM over 12 simulations).

synaptic weights. It has also been suggested that STP can be used as a dynamic basis 213 for persistent activity [15], a proposal we do not consider in this study. Instead, we 214 investigate the effect of STP heterogeneity on persistent activity generated by 215 reverberation in a cell assembly. Our original model [21] contained three types of STP, 216 which are suggested by electrophysiological data: Predominantly facilitating plasticity, 217 predominantly depressing plasticity, and a mixed plasticity type [30, 31]. These studies 218 also show that the cell types of the pre- and postsynaptic neuron determines the 219 plasticity type of a given synapse. Connections among pyramidal cells, among 220 fast-spiking interneurons and between fast-spiking interneurons and pyramidal cells, 221 however, can exhibit all three types of plasticity with different probabilities (see 222 Methods for details), introducing another factor of heterogeneity. We refer to 223 simulations with this full distribution of STP types as "heterogeneous STP" and to 224 those with only one STP type for a given pair of cell types as "homogeneous STP". In 225 the homogeneous case, only the most frequently occurring STP is being used for each 226 pair of cell types (facilitating STP for pyramidal-pyramidal connections and 227 depressing STP for fast-spiking interneurons inhibiting pyramidal cells as well as for 228 interneuron-interneuron connections, c.f. Figure 2B in [21]). 229

The different curves in Figure 4 represent these different parameter configurations of STP: Heterogeneous STP (red curve), no STP at all (black curve) and homogeneous STP (blue curve). With heterogeneous STP (red curve), d_{PA} is close to zero for all values of cell parameter variability, while in the simulations without STP (black curve), persistent activity emerges as cellular heterogeneity is reduced. However, removing STP is not a necessary condition for persistent activity. Homogeneous STP (blue curve) also leads to persistent activity for low cell parameter heterogeneity. Thus, it is the heterogeneity of STP that prevents persistent activity to emerge, not STP per se.

Fig 4. Interaction of cellular heterogeneity and short-term plasticity.

Persistent activity measure $d_{\rm PA}$ as a function of the variability in neuron parameters with original STP (red), homogeneous STP (blue) and no STP (black). Mean $d_{\rm PA} \pm$ SEM over 24 simulations is shown.

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Dynamic impact of heterogeneous synaptic input The above results show that 238 persistent activity can be achieved when heterogeneities in interneuron cell parameters 239 and short-term plasticity are removed or compensated. While these conditions are 240 necessary for persistent activity, they are not sufficient: All of the simulations so far 241 used a uniform connectivity of 10% across all cell types, while persistent activity could 242 not be generated in the full network with all layers and cell types included and the 243 original connectivity pattern obtained from *in vitro* data [21]. In this original pattern, 244 connection probabilities are higher among interneurons (25%) and between 245 interneurons and pyramidal cells (15% to 70%) compared to the connectivity among 246 pyramidal cells (about 10%). Fast-spiking interneurons and Martinotti cells project 247 particularly frequently onto pyramidal cells (about 50% and 70%, respectively). In the 248 following, we investigate how this pattern of heterogeneous connectivities between 249 different cell types shapes the dynamics of the full network (including homogeneous short-term plasticity) and why it prevents persistent activity from emerging.

First, we found that the most notable difference between the heterogeneous 252 connectivity patterns and the homogeneous one (10% connectivity among all cell 253 types) lies in the distribution of synaptic weights: In the heterogeneous connection 254 pattern, pyramidal cells receive many more connections from fast-spiking interneurons 255 and Martinotti cells in layer 2/3 compared to other cells types and cells from layer 5. 256 Furthermore, synaptic weights for these densely connected cells type combinations are 257 much stronger compared to others (2.30 mV and 1.91 mV, for fast-spiking 258 interneurons and Martinotti cells, respectively, compared to an average of 0.10 mV for 259 all other types of interneurons). These two sources of inhibition result in a bimodal 260 distribution of synaptic weights (Figure 5A, red curve). When all connectivities are set 261 to a uniform 10%, far fewer of the strong weights from fast-spiking interneurons and 262 Martinotti cells are being drawn, while the distribution of the weaker weights is much 263 less affected (Figure 5A, blue curve). This leads to greatly decreased numbers of 264 strong inhibitory synapses onto L2/3 pyramidal cells in the uniform network compared 265 to the heterogeneous one.

Fig 5. Dynamic difference between homogeneous and heterogeneous

synaptic connections. A: Histogram of synaptic weights in networks with homogeneous connectivity (10% for all cell types, blue curve) and heterogeneous connectivity (connectivity constrained by data, red curve). B: Histogram of the total strength of each synapse in the networks with homogeneous and heterogeneous connectivity, computed as the product of the synaptic weight, the variables of short-term synaptic plasticity, averaged over all synaptic events, and the number of synaptic events. The inset shows the distributions on a log-log scale to demonstrate the heavy tail of the distributions. The black bar shows a $1/x^2$ distribution for comparison.

Second, we investigated the impact of the weight distribution on the total synaptic input from a given interneuron to a given cell assembly member. The total synaptic input is defined as the sum of synaptic conductance changes in the target neuron by spikes from the presynaptic interneuron, summed over the entire spike train of that interneuron. These total inputs follow of a heavy-tailed distribution (Figure 5B), where the tail sits on top of barely visible bimodal distribution at lower values. While the two modes reflect the distribution of the weights, the heavy tail occurs from the fact that large values of the total input require the rare coincidence of both large weights and high firing rates. Furthermore, inputs from interneurons with high rates are often strongly attenuated by short-term synaptic plasticity, as the predominant plasticity type from interneurons to pyramidal cells is synaptic depression. Thus, a high total input also requires suitable parameters of short-term plasticity, most importantly, a short time constant of synaptic depression. All these conditions only rarely coincide, but when they do, very large total synaptic inputs result, constituting the heavy tail of the distribution. Importantly, coincidence is much more likely if there is a large fraction of strong synapses, as in the noise network. Thus, the tail is heavier for heterogeneous connectivity (Figure 5B, red curve) compared to uniform connectivity (Figure 5B, blue curve).

Cell assembly neurons that receive even a few of those extreme inputs well show inhibitory conductances which are much higher than those from neurons which inhibitory inputs are all close to the mean, being effectively dominated by those few inputs. Thus, because of the much higher probability of extreme inputs under heterogeneous connectivity, we expected to see an increase both in the mean and in the standard deviations of the distribution of inhibitory conductances compared to

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uniform connectivity. Indeed, inhibitory conductances did show a higher mean and standard deviation across assembly neurons for the heterogeneous connectivity $(54.30 \pm 12.66 \text{ for heterogeneous connectivity compared to } 14.25 \pm 7.91 \text{ for uniform connectivity; mean } \pm \text{SD}; p=1.4 \cdot 10^{-54}, t(158)=24.01, two-sided t-test), while excitatory conductances only slightly (but significantly) differ in both configurations <math>(1.05 \pm 0.41 \text{ for heterogeneous connectivity compared to } 1.32 \pm 0.43 \text{ for uniform connectivity; mean } \pm \text{SD}; p=-4.8 \cdot 10^{-5}, t(158)=-4.18, two-sided t-test).$

When we compared the homogeneous and the heterogeneous network, we actually did two comparisions at once: One between homogeneous and heterogeneous connectivities (same or different numbers of connections between different cell types) and another one between a high and low overall number of connections (10% in the homogeneous network vs. about 20% in the heterogeneous network constrained by data). Thus, we need to check which of the two differences sets apart the two types of networks. To this end, we conducted one set of simulations where connectivities to 20%. In both of these cases, persistent activity could be achieved. Thus, we conclude that an increased number of connections is only harmful for persistent activity if they increase the overall number of inhibitory connections.

Persistent vs. spontaneous activity

In the previous section, we reported conditions under which a biologically tightly constrained network model [21] can exhibit persistent activity. An important goal of the study that introduced this model [21] was to mimic the asynchronous-irregular (AI) background activity [32, 33], which has been observed both in models [32–34] and experiments [33,35]. Thus, it needs to be tested whether AI activity can be generated in a network that fulfills the constraints for persistent activity (note that we concentrate on the irregularity in the following). We implemented all three conditions (homogeneous short-term plasticity, interneuron parameter variability at 5% of its original value and uniform connectivity at 10% for all cell types) in the original network and assessed both persistent activity and measured the coefficient of variation (C_V) , a measure of inter-spike time variability. As expected, persistent activity robustly emerged in this network (Figure 6A) after decreasing the background current from 500 to 120 pA. However, activity was also much more regular compared to the original, heterogeneous network (mean $C_V = 0.46$ before and mean $C_V = 0.14$ after the stimulus in Figure 6A compared to 1.04 ± 0.33 in the original network [21], mean \pm SD). While removing the constraints on variability of cellular and short-term plasticity parameters did not change the regularity of the activity, using the original, heterogeneous connectivity did: The mean C_V reached 0.64, which is above the lower bound of the values found in the original network. However, persistent activity was not possible in this configuration: Activity levels quickly decay back to baseline once the stimulus is switched off (Figure 6B). This could not be changed by using a range of different background currents and synaptic weight strengths (see below). There are at least two possible explanations for this observation: Persistent activity could either be destabilized by noise, or the dynamic regimes that bring about persistent activity and which generates noisy, irregular behavior may be mutually exclusive. In the following to sections, we present tests of both hypotheses. We found that persistent activity is robust to noise, but that noise indeed needs to be generated at a different synaptic configuration. Finally, we tested to which degree a "noise network" generating irregular activity could overlap with the "signal network" generating persistent activity, both in terms of pyramidal cells and interneurons.

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Fig 6. Persistent and spontaneous activity. A: The full network with interneuron cell parameter, connectivity and short-term plasticity heterogeneity removed shows persistent activity, but at low variability. B: When the original, heterogeneous connectivity pattern is used, persistent activity breaks down, but irregular activity emerges. Both panels show raster plots and instantaneous firing rates as in Figure 1A.

Robustness against external noise To investigate the robustness of persistent 340 activity against noise, we added a second set of 1000 neurons to the network and 341 configured the first subnetwork to generate persistent activity ("signal network", 342 uniform connectivity at 10%) and the second one to generate spontaneous activity 343 ("noise network", original connectivity from [21], i.e., many fewer connections among 344 pyramidal cells compared to interneurons and compared to pyramidal-interneuron 345 connections). Pyramidal cells from the noise network project onto the pyramidal cells 346 in the signal network with the same synapse parameters as within the noise network, 347 but with uniform connection probabilities c between zero and one (which constitutes a 348 minimal perturbation of the signal network). Thus, the two networks simulate 349 spatially separate columns that communicate via pyramidal cells only (see sections 350 below for more interleaved networks). Figure 7A shows the $d_{\rm PA}$ values for different 351 values of the coupling constant c (only for those simulations which show successful 352 persistent activity, i.e., $d_{\rm PA} > 0.3$). These values do not systematically change as c is 353 increased, which shows that persistent activity can survive considerable levels of 354 externally generated noise, compatible with previous results [25]. C_V s within the cell 355 assemblies are consistently above one, largely independent of the values of c (1.16 \pm 356 0.06, mean \pm SEM over all c values). Outside the cell assembly, however, C_V s are 357 strongly modulated by the connection strength to the noise network. Figure 7B shows 358 the C_V both before and after the stimulus presentation as a function of c (same 359 simulations as in panel A). C_V increases with c in both cases. However, only C_V s 360 measured after the stimulus presentation, i.e., during persistent activity itself, reach 361 values that are within the range of experimentally measured C_{VS} in vivo (shaded 362 region in Figure 7A, spanning one standard deviation around the mean C_V s measured 363 in a multi-item working memory task in rats [21, 36]). Activity before stimulus 364 presentation is much more regular: C_V values do not reach the *in vivo* range for any of 365 the c values used (stronger connections with c > 0.25 ignite persistent activity 366 independent of the stimulus). Apparently the additional (noisy) drive from persistent 367 activity is needed to fuel sufficient activity in the neurons outside the cell assembly, so 368 the noisy activity is a joint product of the noisy dynamics of persistent activity and the synaptic bombardment from outside the network.

Fig 7. Generation of spontaneous and persistent activity in two coupled **networks.** A: Persistent activity measure d_{PA} as a function of the connectivity c (see text for details). B: C_V measured during 500 ms before (spontaneous activity, blue) and after (persistent activity, red) stimulus presentation as a function of the connectivity c of the signal network to the noise network (same simulations as in Panel A). The shaded area denotes the range of one standard deviation around the mean (dotted line) of C_V measured experimentally in vivo [21].

370 In summary, input from the noise network is effective for increasing the C_V of the signal network without harming its ability to generate persistent activity for moderate connectivity between the two networks. 373

Dynamic generation of persistent and spontaneous activity To answer the 374 question of whether noise and persistent activity can be dynamically generated within 375

the same network (potentially at different times and states), we tried to transform the 376 signal network into a noise network (i.e., a network exhibiting C_{VS} near one outside 377 the cell assembly) while keeping its ability to generate persistent activity. If this is not 378 possible, the noise within a network exhibiting persistent activity may either have to 379 be generated by external input or by modulation of its properties. As the key 380 difference between the signal and the noise network is in the connection probabilities, 381 we changed these probabilities in the signal network from the uniform 10% back to the 382 original values of the noise network (see previous section) and tried to retrieve 383 persistent activity by compensating for these changes using the synaptic weights: 384 Whenever a connections probability was decreased, we increased the weight of the 385 remaining synapses, and vice versa. Note that this procedure is not intended to mimic 386 a real biological process, but rather to test whether it is possible to construct a 387 network which is capable of both persistent and noise-like activity (see section 388 "Working memory and irregular activity" for a possible way to switch between 389 persistent and irregular activity using neuromodulation). Indeed, we obtained 390 persistent activity with the original connectivity in this way (Figure 8A), but only 391 with fine-tuning of each synaptic weight up to the tenth of a percent. More 392 importantly, this fine-tuning did not yield realistic, noisy activity in the neurons 393 outside the cell assembly: C_V s are very low (mean $C_V = 0.27$), so the ability to 394 generate noisy activity could not be recovered in a network with persistent activity.

Fig 8. Failure to generate spontaneous and persistent activity in the same network. A: Example of persistent activity with original connectivity values and fine-tuned synaptic weights, but regular activity ($C_V = 0.27$, raster plot and instantaneous firing rate as in Figure 1A). B: Persistent activity measure d_{PA} as a function of the C_V for all attempts to combine spontaneous and persistent activity generation in the same network. The shaded area denotes the range of one standard deviation around the mean (dotted line) of C_V measured experimentally in vivo.

We summarize all attempts to combine persistent and spontaneous activity by plotting the mean C_V and d_{PA} values in Figure 8B, excluding those configurations which either put the network in an epileptiform state (mean firing rates ≥ 20 Hz for neurons outside the cell assemblies) or yield almost no firing at all (mean firing rates ≤ 0.1 Hz). One clearly sees that successful persistent activity ($d_{PA} \geq 0.3$, dotted red line) can only be found at low C_V values between 0.5 and 0.6, which are significantly below the Poisson-like value of one usually reported *in vivo*. In particular, all these values are outside the range of C_V s obtained from our reference data set (shaded region in Figure 7B and Figure 8B [21,36]).

Admissible overlap between signal and noise network As subnetworks in the cortex do not exist in isolation, we next studied to which degree the signal network (generating persistent activity) and the noise network (generating noisy, spontaneous activity) can be mutually interconnected without harming persistent activity, as opposed to the one-directional projection from noise to signal network we described above. We considered two different types of connections (Figure 9A): First, synapses between the two networks, but within the same cell type, i.e. from pyramidal cells of the noise network to pyramidal cells of the signal network, and the same for interneurons (with a connectivity of a fraction c_P of the connectivity within the noise network). And second, synapses between the pyramidal cells of one network and the interneurons of the other (connectivity varied as a fraction c_C of the noise network connectivity for arbitrary values of c_P , while even small non-zero values of c_C completely abolished persistent activity (Figure 9B).

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Fig 9. Admissible overlap between signal and noise network. A: Illustration of the network connectivity. A signal network (uniform connectivity of 10%) and a noise network (original connectivity) are connected at variable fractions of the original connectivities. Pyramidal cells and interneurons in both networks are mutually connected at a fraction c_P , and pyramidal cells in one network and interneurons in the other are connected at a fraction c_C . B: Persistent activity measure d_{PA} as a function of the connection strengths c_C (black curve) and c_P (blue curve). As one of the strengths is varied, the other one is kept to zero.

From the previous results of this paper, we hypothesized that persistent activity is 419 destroyed by the variability of the inhibitory input from the noise network onto the 420 cell assembly members. To test this hypothesis, we varied the strength of the synaptic 421 input from the interneurons of the noise network to the pyramidal cells of the signal 422 network and compared the statistics of the resulting synaptic conductances (at 423 $c_P = 0.3$) onto the members of the cell assembly with those conductances emerging 424 without crosstalk between the two networks $(c_P = 0)$. We found that the inhibitory 425 currents at $c_P = 0.3$ and full synaptic strength of the corresponding synapses are 426 higher (average conductance 18.24 ± 1.07 nS for $c_P = 0.3$ compared to 12.27 ± 0.64 427 nS for $c_P = 0.0$; mean \pm SEM; $p = 2.0 \cdot 10^{-45}$, t(78)=30.28, one-sided t-test) and 428 more variable (standard deviation 9.23 \pm 0.52 nS for $c_P = 0.3$ compared to 5.90 \pm 429 0.34 nS for $c_P = 0.0$; mean \pm SEM; p = $1.1 \cdot 10^{-48}$, t(78)=33.90, one-sided t-test) 430 compared to $c_P = 0$. Only when the weights of the cross-network synapses are 431 decreased as much as 90% of the original value, average strength and variability of the 432 inhibitory inputs are reduced close to the values at $c_P = 0$ (average conductance 13.76 433 \pm 0.92 nS, standard deviation 6.82 \pm 0.50 nS), also restoring persistent activity. Thus, 434 although only 30% of the connections of the noise network project onto the cell 435 assembly members, the corresponding weights must be very strongly decreased to 436 compensate for the effects of the heavy tail in the inhibitory inputs. 437

Dynamic mechanism of breaking persistent activity

In this final part of the results section, we investigate the dynamic mechanism of how heterogeneity in the interneuron rheobases breaks persistent activity. This effect seems counterintuitive at first glance, as the stronger input of low-rheobase interneurons should be compensated by the lower input form high-rheobase ones, given a symmetric rheobase distribution. However, inhibitory input to pyramdial cells exhibits two important nonlinearities: If the inhibitory input is too strong, the pyramidal cell ceases firing at all. On the other hand, if the input to interneurons falls below the rheobase, there is no inhibitory input at all, leaving the pyramidal cell constantly active. As heterogeneity of interneuron rheobase increases, more and more pyramidal cells in the cell assembly are being driven into one of these two extremes, taking them out of the bistable regime that is necessary for stimulus-dependent persistent activity.

To show that this mechanism does not depend on the specific configuration of the 450 present model, we use a minimal model of persistent activity [37], comprising only a 451 single pyramidal cell of the simplified Hodgkin-Huxley-type [38] equipped with an 452 GABA and NMDA autapse, which are modelled in the same way as in the present 453 model (conductance-based synapses, NMDA including a magnesium block for low 454 voltages of the postsynaptic cell, see Methods for details). As shown in [37], the 455 positive feedback from the NMDA self-connections enables persistent activity, or more 456 precisely, bistablity between an active state (moderate to high firing rate) and an 457 inactive state (zero firing rate), while the inhibitory GABA self-connection contributes 458 to the stability of the inactive state. We extend this model by filtering the GABA 459

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conductances by a number of rectified linear units (ReLU), each mimicking an 460 interneuron with a different rheobase. The slope and the mean rheobase of the ReLU 461 units are chosen to fit the f-I curve of the interneurons in the full model (see Methods 462 for details). Furthermore, we decrease the synaptic condutances to 10% of the values 463 reported in [37] to keep the firing rate persistent activity in a regime similar to the 464 present simulations (up to 80 Hz). This system constitutes a minimal model of a 465 pyramidal cell activating a number interneurons with heterogeneous rheobases, which 466 in turn inhibit that cell, as it is the case of each member of the cell assemblies of the 467 full model considered before.

Fig 10. Nullclines of a minimal model of persistent activity. A: Example of nullclines with the three fixed points necessary for bistability. B: Mean (thick red curve), mean plus and minus standard deviation (thin red curves), minimum and maximum (green curves) of the $I_{\rm syn}$ nullcline for a large standard deviation of the rheobase of the interneurons (std_rheo=120 pA) together with the firing rate nullcline (blue curve).

To analyze the dynamic behavior of the minimal model, we construct a two-dimensional phasespace by computing the instantaneous firing rate FR (inverse of the interspike interval) and the mean input current $I_{\rm syn}$ between each two spikes (Figure 10A, following the approach in [39]). In particular, we compute the nullclines of the two variables, curves in phasespace at which the flow in the direction of one of the two variables changes sign, i.e. that variable does not change when starting anywhere the nullcline if the other variable is held constant. The FR nullcline is computed by varying the input current into the cell and compute the average firing rate for each current without synaptic feedback (blue curve in Figure 10A) and the $I_{\rm syn}$ nullcline is the mean synaptic current produced at each of these rates (red curve in Figure 10A). The FR nullcline is identical to the f-I curve of the pyramidal cell modelled here and the I_{syn} nullcline is the sum of the average NMDA and GABA currents generated at a given firing rate, plus the background current. The geometric shape of the $I_{\rm syn}$ nullcline determines the number and stability of fixed points, and thus the ability of the system to exhibit bistability and stimulus-specific persistent activity: Each crossing of the FR and $I_{\rm syn}$ nullclines constitutes a fixed point and bistability requires two stable fixed points separated by an unstable one (black dots in Figure 10A). Geometrically, this implies that the slope of the $I_{\rm syn}$ nullcline needs to change sign at least once. As the inhibitory input is varied (directly or via rheobase variation), the inhibitory component of the $I_{\rm syn}$ nullcline is shifted to the left or to the right. When the inhibitory input becomes too strong, the I_{syn} bends more and more to the left, eventually losing the active state fixed point at non-zero firing rates (left green curve in Figure 10B). When the input to the interneurons drops below the rheobase, on the other hand, the inhibitory component of the $I_{\rm syn}$ becomes zero and the $I_{\rm syn}$ is identical to its excitatory component, which lacks a change in the slope sign and does not cross the FR firing rate at zero firing rate (right green curve in Figure 10B), losing the inactive stater fixed point. In both cases, the system becomes monostable, either in the active or the inactive state.

To study the effects of heterogeneity in the interneurons, we model a population of 1000 pyramidal cells, each of which receiving a different amount of inhibition and thus being governed by a different $I_{\rm syn}$ nullcline (Figure 10B, see Methods for details). As heterogeneity increases, more and more of these cells will be driven out of the bistable range, into one of the two monostable regimes. As a result, the number of bistable neurons, i.e., exhibiting three fixed points, drops with increasing rheobase heterogeneity, which is precisely the effect we observed in the full model using the measure $d_{\rm PA}$ (Figure 11A, blue curve). Importantly, changing the (mean) background

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Fig 11. Number of bistable neurons drop as rheobase variability increases. A: Number of bistable neurons, i.e. $I_{\rm syn}$ nullclines with three intersections with the firing rate nullcline, as a function of the standard deviation of the rheobase of the ReLU units providing inhibitory input (blue curve) or excitatory input (black and red curves, using two different slopes for the ReLU units). B: Number of bistable (three intersections), active (one intersection at non-zero firing rates) and inactive (one intersection at zero firing rate) neurons as a function of the background current. The standard deviation of the inhibitory ReLU units is set to 60 pA.

current does not help to restore bistability, as it simply replaces monostable active by monostable inactive cells or vice versa Figure 11B, compare the red curve with Figure 1C).

The amount of rheobase heterogeneity that is admissible for persistent activity 508 depends on the curvature of the $I_{\rm syn}$ nullcline, or more precisely, on the range between 509 the current at zero firing rate and the one at the first local maximum where the 510 nullcline changes its slope (e.g. -6 and 15 nA in Figure 10B). This range is largely 511 determined by the strength of the two synaptic inputs. Thus, it is possible to restore 512 persistent activity at a given level of interneuron heterogeneity, e.g., by choosing larger 513 peak conductances. However, such a scaling also increases the firing rate of the active 514 state, illustrated by comparing the (mean) $I_{\rm syn}$ nullclines in Figure 10A and B. Indeed, 515 as we systematically vary the scaling of the peak conductances of the NMDA and 516 GABA currents (Figure 12A), we observe a linear increase in both the maximal 517 admissable rheobase variability (blue curve) and the firing rate (orange curve) as the 518 scaling, and thus, the range of the $I_{\rm syn}$ nullcline, is increased.

Fig 12. Restoring bistability by increasing peak conductances or number of interneurons A: Maximum of the rheobase standard deviation without a drop in the number of bistable neurons (blue curve) and firing rate in the active state (orange curve) as both NMDA and GABA peak conductances are being scaled by the same factor. As the weights are scaled up, the range of the (mean) I_{syn} nullcline also increases (written above the graph for each value of the scaling factor). B: Number of bistable neurons as the number of interneurons (ReLU units with different rheobase values filtering the inhibitory conductance) is being increased. The standard deviation of the rheobase was set to 60 pA for all simulations.

519 Next, we compare the effect of interneuron heterogeneity with the effect of 520 pyramidal cell heterogeneity. To that end, we filter the NMDA conductances instead 521 of the GABA conductances by a number of ReLUs, effectively modelling a number of 522 pyramidal cells that are excited by the modelled cell and excite that cell in turn. If the 523 same slope is being used for the pyramdial ReLUs as for the interneuron ReLUs 524 (s=0.3), the number of bistable neurons drop with rheobase heterogeneity in the 525 pyramidal cell in the same way as it did with interneuron heterogeneity (Figure 11A, 526 black curve). However, when mimicking the much shallower slope of the pyramidal 527 cells from the simulations (s=0.05), increasing rheobase heterogeneity to the same 528 extend as in the interneurons does not have a significant effect on the number of 529 bistable neurons, as in the full PFC network (Figure 11A, red curve, cf. (Figure 2A, 530 red curve)). Note that in the full model, pyramidal cell heterogeneity was varied while 531 interneurons exhibited their full heterogeneity, while they are all identical here, 532 explaining the different levels of the curves in the two figures. 533

Finally, we assess the effect of the number of interneurons projecting on a single pyramidal cell. In all previous simulations, we used 200 ReLU units with different rheobases to compute the inhibitory component for each of the 1000 I_{syn} nullclines,

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each representing a single pyramidal cell. As this number is increased, more and more of the variability in the inhibitory input is averaged out, which leads to a lower standard deviation in the total input to each pyramidal cell. Consequently, the number of bistable neurons increases as the number of simulated interneurons increase (Figure 12B). As apparent in the figure, about 1000 interneurons are sufficient to keep all pyramidal cells in the bistable regime for a rheobase variability of 60 pA.

As the above results suggest that increasing the number of neurons could be a possible way to produce persistent activity in the presence of heterogeneities, we also conducted a number of simulations of the full model with larger networks. As expected, homogeneous networks showed persistent activity, while heterogeneous networks (using the original connectivity) did not using 3000, 5000 or 10000 neurons. Importantly, the 3000 neuron network also showed persistent activity when the rheobase heterogeneity was increased to 10% of the original values, while the 5000 neuron network could even tolerate 50% heterogeneity. However, even the 10000 neuron network was not able to produce peristent activity at full rheobase heterogeneity. So, increasing the network size can in fact be used to extend the range of admissible heterogeneity, but the full range obtained from *in vitro* data can not be averaged out even in a network that is larger than a cortical column (containing about 7500 neurons [40]).

Discussion

We presented a working memory model based on persistent activity within a biologically validated network model of the prefrontal cortex. We found that the network shows the bistability necessary for stimulus-dependent persistent activity only under the condition that the excitability of the interneurons that inhibit the neurons of the cell assembly is largely homogeneous. The same is true for the strengths of the inhibitory inputs onto the assembly members - a heavy tail in the distribution of these inputs also destroys persistent activity. Furthermore, pairs of neurons need to exhibit the same type of short-term plasticity (if present) for a given combination of cell types. None of these homogeneity criteria is fulfilled in the full network model, nor in the electrophysiological experiments from which the neuron and synapse parameter distributions of the model are taken from [21]. Thus, we argue that persistent activity may require that heterogeneities are compensated by appropriate learning rules such as homeostatic plasticity, at least locally for a given cell assembly.

Breaking persistent activity by interneuron heterogeneity

We have identified the dynamic mechanism of breaking persistent activity by 571 heterogeneity in the rheobase of the interneurons, namely a spread in the inhibitory 572 input that drives part of pyramidal cells into inactivity and another part into 573 hyperactivity. Using a minimal model of persistent activity, we have shown that this 574 mechanism does not depend on the exact implementation details of the model. Rather, 575 using the language of dynamical systems theory, we have shown that this mechanism 576 is a general dynamic phenomenon that can occur in any system where at least one of 577 the nullclines exhibits enough variability to lead to different number of fixed points for 578 the different components (e.g. pyramidal cells). In this sense, the interneuron 579 rheobase can be seen as a bifurcation parameter that is varied above and below the 580 two bifurcation points between bistability and monostability of the active and inactive 581 state, respectively. In particular, any source of heterogeneity in the inhibitory part of 582 the $I_{\rm syn}$ nullcline will have a similar effect, potentially explaining the deleterious effect 583 of heterogeneity in the synaptic weights and in the parameters of short-term synaptic 584

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plasticity for persistent activity as well. Also in line with the results in the full network, the minimal model predicts a very limited effect of pyramidal cell heterogeneity (Figure 11A, red curve), which can be explained by the fact that the f-I curve of the pyramidal cells is much shallower compared to those of interneurons (compare red and black curve in Figure 11A).

Using the minimal model, we have identified two potential mechanisms to compensate the effects of heterogeneity in the interneurons: Increasing the synaptic peak conductances and increasing the number of interneurons that innervate a given pyramidal cell (Figure 12). The former is accompanied by an increase in the firing rate (Figure 12A, orange curve). Thus, a low-rate active state is unlikely to be stabilized against interneuron heterogeneity in this way, especially when starting from the even lower weights that are required rates as low as 20 Hz (about 0.5 to 1% of the original weights). In line with these observations, the only simulations of the full model showing peristent activity at full heterogeneity used strongly increased NMDA-to-AMPA ratios and a large cell assembly (e.g. many strong excitatory inputs and lead to epileptiform firing of the entire network at a very high rate.

The latter analysis allows for an estimate of the number of interneurons that are necessary to sufficiently average out their heterogeneity. From Figure 12B, full bistability is recovered starting from about 1000 interneurons (at a rheobase variability of 60 pA, which is close to the value in the full model). Given the number of 7500 neurons in a cortical column [40] and the cell type distributions used in the full model [21], one arrives at 230 local interneurons in $L_2/3$ and and only 40 local interneurons in L5 in a single column, which is clearly not enough to average out rheobase heterogeneities. Taking together all interneuron types, these numbers increase to 780 and 230 in $L_{2/3}$ and L5, respectively, so the total effect of interneuron heterogeneities may depend on the degree to with these other types of interneurons (e.g. VIP and SOM positive cells) affect persistent activity. A recent study [41] shows that VIP and SOM positive interneurons improves persistent activity rather than hindering it. Finally, we have only considered interneuron heterogeneity in the minimal model, while heterogeneity from other sources will add onto the overall variability of the inhibitory input. In particular, we have seen that the interplay between the (log-normal) distribution of the synaptic weights, the firing rate of interneurons and short-term synaptic plasticity creates a heavy tail of inhibitory inputs (Figure 5). The very strong inputs from that tail often dominate the entire inhibitory input of a neuron, largely irrespective of the number of smaller inputs.

Working memory and irregular activity

To our knowledge, this study is one of very few to consider the relation between the dynamic generation of persistent activity and irregular activity within the same network (e.g. [34, 42]). Previous working memory models were typically provided with Poisson background spike trains [16, 25, 43], which mimicked the synaptic bombardment with random input from outside the memory network, characterized by coefficients of variation near one and spike-time correlations between input neurons near zero. The generation of noisy activity alone has been subject to many studies (e.g. [32, 44] based on the notion of balanced excitation and inhibition.

The biologically constrained model we use here [21] provides a slightly different mechanism for generating irregular activity, namely a balance between few very active neurons (mostly interneurons) and a vast majority of almost silent neurons (mostly pyramidal cells). While a full analysis of this mechanism is out of the scope of this paper, Figure 5B suggests that irregular activity is being driven by a heavy tail of inhibitory synaptic inputs. Previous studies [42] already suggested that self-sustained irregular activity requires strong synaptic weights. The present study shows that only

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a few strong inhibitory inputs together with a constant excitatory input also generate irregular activity. In contrast to previous proposals, this form of noise generation is compatible with the frequent observation of a large fraction of neurons that are mostly silent when using recording techniques that are not biased towards spiking such as calcium imaging or *in vivo* patch-clamp ("dark matter theory" of neuroscience, [45–47]).

Importantly, the heavy tail of the inhibitory inputs [22] does not result from the log-normal distribution of the synaptic weights alone, but in concert with heterogeneous firing rates, i.e. strong weights and high firing rates need to coincide to from the tail of the distribution. On the other hand, we have shown that persistent activity breaks down in the present of heterogeneous inhibitory inputs. Thus, at least within our modeling framework, the dynamic basis of irregular activity seems to be incompatible with the more regular, homogeneous synaptic configuration that is needed for persistent activity: A cell assembly needs relatively homogeneous distributions of synaptic inputs to exhibit persistent activity. The heavy tail explains why the increased inhibitory connectivity in the original network cannot be simply compensated with a decrease in the mean synaptic weights. Such a decrease would only shift the entire input distribution to lower values, but does not abolish the heavy tail that is responsible for most of the increase in the inhibitory conductance.

As a consequence, noisy background activity must be implanted into a cell assembly by its surroundings to show the routinely observed asynchronous-irregular activity [33, 35], while it remains to be investigated whether asynchronicity is also being affected by heterogeneities. Alternatively, the network may be capable of exhibiting both kinds of activity and switch between the signal and noise state by neuromodulation. Dopamine, for instance, is known to stabilize working memory due to a number of modulatory effects on cellular and synaptic properties [25, 48, 49]. Thus, activation of dopamine receptors may compensate the effects of existing heterogeneous inhibitory input. This could enable persistent activity in a noise network, temporarily turning it into a signal network. We can use the minimal model to illustrate how such a modulation could work. In the study [37] from which the model is taken, dopaminergic modulation is modelled by scaling both NMDA and GABA condutances by the same factor, i.e., higher dopamine levels lead to stronger feedback via NMDA and GABA synapses. Thus, we can study the possible effect of dopamine in Figure 12A: As dopamine levels increase, so does the range of the $I_{\rm syn}$ nullclines, the maximal admissible standard deviation, and the firing rate. This raises the possibility that high dopamine levels facilitate persistent activity by increasing its firing rates making it more robust against higher levels of interneuron heterogeneity.

When noise is provided from outside the network, as in previous models [16,25,43], persistent activity is not impaired (Figure 2B), but shows the irregularity that is seen *in vivo*. Interestingly, C_V s are higher during persistent activity compared to spontaneous activity (Figure 7A), likely due to the additional (noisy) drive provided by the cell assembly neurons to those outside the assembly, which is consistent with observations *in vivo* [50].

Regarding interactions of signal and noise networks, we have shown that inhibitory input from a noise network disrupts persistent activity in a signal network, while excitatory input does not. Interestingly, a recent *in vivo* study has shown that optogenetic activation of somatostatin (SOM)- and parvalbumin (PV)-positive interneurons disrupt persistent activity during the delay period of a memory-guided behavioral task, while vasoactive intestinal peptide (VIP)-positive interneuron activation enhanced task performance [41]. PV interneurons are commonly associated with fast-spiking interneurons, SOM interneurons with Martinotti cells and VIP cells

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with bitufted and other cross-layer projecting interneurons. The first two types may project beyond a single cortical column, while the latter is largely confined within its own column. The current study may explain the differential effect of VIP and PV/SOM cells on persistent activity under the assumption that signal and noise networks are spatially separated in different cortical columns: Activating PV and SOM cells may elicit inhibitory effects that travels across networks, contaminating the signal networks supporting the delay phase activity. VIP cells, on the other hand, mostly inhibit cells within one column, so there is much less inhibitory crosstalk between different networks. On the other hand, a recent study [51] suggested that cell assembly formation may not be limited to pyramidal cells, but local interneurons can be recruited into the assembly, while far-reaching interneurons such as SOM cells decorrelate from the assembly. Thus, it seems possible that a cell assembly may learn to separate itself from direct inhibitory input from the outside that is harmful to persistent activity, even in the absence of spatial separation.

From in vitro constraints to in vivo activity

This study demonstrates a number of constraints on persistent activity within our model, namely homogeneity of interneuron excitability, connection probability and synaptic dynamics within a cell assembly, which are unlikely to be fulfilled if the neurons within the assembly are randomly selected. Thus, we must discuss whether these constraints are relevant for persistent activity in the real brain, and if they are, how they can be fulfilled by biologically plausible mechanisms.

Regarding the biological relevance, we emphasize that all heterogeneities in this model are directly derived from electrophysiological experiments [21]. Nevertheless, we cannot exclude that the variability of cellular and synaptic parameters may be artificially inflated, e.g., by combining data from different experimental conditions. So in theory, if this artificial variability is larger than the variability due to biological diversity, the breakdown of persistent activity could be an artifact of the inflated variability. On the other hand, our results show that even moderate heterogeneity in the interneuron excitability breaks persistent activity (20-25%) of the original variability, c.f. Figure 2). Thus, potential artificial variability would have to be inflated at least four-fold over biological variability to break persistent activity on its own, which is highly unlikely. Furthermore, we have drawn all neuron parameters from a multivariate distribution which respects the correlations between these parameters (see Methods for details), so we can exclude that the effect of heterogeneity in one parameter e.g. on firing rate output is compensated by the heterogeneity in another one by mean of correlations between the two [52]. However, we cannot exclude that such compensatory mechanisms exist between neuron and synapse parameters, as we have obtained the latter from the experimental literature. In fact, such correlations could arise the result of homeostatic mechanisms, e.g., compensating the effect of strongly firing interneurons with lower synaptic weights [52] (see below).

For a biologically plausible mechanism to fulfill the homogeneity constraints, we 728 consider that cell assemblies are assumed to be formed by long-term neural plasticity 729 as a result of repeated common input to a set of neurons. This Hebbian plasticity may 730 be complemented by homeostatic plasticity rules that are known to scale synaptic 731 efficacies such that a given average firing rate in the postsynaptic network is 732 maintained [23, 24]. Here, we have considered a (simplified) scaling of the excitatory 733 input into interneurons, such that the differences in excitability are compensated by 734 opposing differences in the input. Another way to limit the effect of the skewed 735 excitability distribution would be to leave the firing rates of the interneurons 736 unaffected, but scale the inhibitory synapses onto pyramidal cells. Recent studies have 737 suggested that both types of homeostatic plasticity may work together to maintain a 738

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balance of excitation and inhibition [24], at a spatial resolution that allows fine-tuning of individual synapses [53]. While such a homeostatic mechanism is unlikely to eliminate all heterogeneity in effective excitability, our results suggest that a decrease to a maximum of approximately 20% of the original heterogeneity will be sufficient to enable persistent activity.

Using homeostatic plasticity on the inhibitory synapses could also help to attenuate heterogeneities in the synaptic input: Pyramidal cells that are subject to very high total inputs from interneurons will fire much less than others, leading to a downregulation of its inhibitory weights. Similarly, combined homeostatic plasticity on excitatory and inhibitory synapses could assimilate inputs from synapses with different types of short-term plasticity: On average, facilitating synapses have a stronger total input on the postsynaptic neuron compared to depressing or mixed-type synapses. As an effect, neurons will exhibit higher or lower firing rates, depending on which type of plasticity dominates, and thus, the respective weights will be adjusted to compensate existing heterogeneities.

Functional benefits from cellular heterogeneity

At first glance, the constraints outlined above appear to limit the utility of cell assemblies as the basis of working memory. However, they could also provide a specific benefit: If each cell assembly is controlled by inhibition of a well-defined strength and timing, the heterogeneity in these parameters between cell assemblies could make each assembly most reactive to a stimulus with specific tuning properties. More specifically, interneuron excitability is an important determinant of the specific frequency of PING-type rhythms (pyramidal-interneuron network gamma [54]) that are generated within a given subnetwork. A cell assembly with a given, mostly homogenous set of interneurons will likely exhibit a rhythm at a narrowly defined frequency and also resonate to input of that particular frequency. If different assemblies exhibit different resonance frequencies, this could help multiplexing multiple memory items at a time, avoiding interference between the assemblies. This proposal is supported by a recent study in monkeys [55], reporting brief gamma bursts during the delay period. Each of these bursts has a well-defined frequency, which varies considerably among different bursts. As cortical oscillations are strongly shaped by interneuron properties, these results are compatible with the existence of a spectrum of cell assemblies, each of which controlled by a relatively homogeneous interneuron population.

Model predictions

The current model makes a number of concrete predictions which could be tested experimentally. Here, we make them explicit and briefly discuss methodological issues of this test.

- 1. Interneurons within a single cell assembly exhibit similar firing rates.
- 2. Isolated networks that show persistent activity have more regular spontaneous activity (lower coefficient of variation) before stimulus onset compared to those in which activity does not persist.
- 3. Synaptic connections between the same cell types within a cell assembly have the same type of short-term synaptic plasticity (namely, facilitating for connections among pyramidal, depressing for interneuron-to-pyramidal connections or those among interneurons).

All three predictions require an experimental identification of cell assemblies and a subsequent analysis of cellular or synaptic properties. A number of computational 785

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methods are now available for cell assembly detection in electrophysiological data, the most recent one even spanning a range of lag times [56]. The non-trivial process of detecting these assemblies may be one of the reasons why the above properties have not been identified before: Assemblies are not necessarily spatially localized, thus anatomical and physiological recordings that are focused in a small spatial volume may fail to see the organization of cellular and synaptic properties within a cell assembly. If these recordings are combined with a detection of the cell assembly, though, one could test, e.g., the first prediction by comparing the firing rates of interneuron units. This could be done both *in vivo* and *in vitro*. Prediction 2 requires the comparison of activity in the same units before and after a stimulus and the isolation of the memory network from most of its environment, both of which should be possible *in vitro*. Finally, prediction three concerns the dynamic properties of synapses within an assembly. Organotypic slices provide access to these properties, with a good chance of intact assembly structures, although it is not always clear to what extent *in vitro* results can be transferred to *in vivo* networks.

Naturally, the predictions rest on the assumption that the network model [21] provides a reasonably valid image of the prefrontal cortex, despite the necessary simplifications, such as a specific selection of cell types and cortex layers, a phenomenological model of the neurons [28] and a constant background current for all pyramidal cells and interneurons of a given layer. We have previously shown that the neuron model provides an accurate description of neuron's responses to fluctuating currents [28] and the network model veridically reproduces many key features of baseline *in vivo* activity [21]. Nevertheless, the model needs to be further tested in terms of its ability to reproduce a wider range of dynamic regimes and functions of the prefrontal cortex.

Heterogeneity in previous working memory models

While there is a wide range of models of working memory based on persistent activity, most of them use homogeneous parameters and few have investigated the effects of heterogeneities on working memory performance: to our knowledge, there are four studies on variants of the ring model [17–20] and another one on a related parametric memory model [57], investigating the effect of random and spatially structures heterogeneities on working memory. In the ring model [16], neurons are spatially organized on a ring and form the strongest connections to their nearest neighbors, effectively forming a continuum of cell assemblies along the ring. Thus, input at any point of the ring can cause a persistent bump of activity in the neurons nearby. Random heterogeneities, e.g., in neural excitability, cause these bumps to drift towards specific points on the ring, compromising their ability to encode the spatial position of the input over time [17-20]. The reason for this drift is the fact that the energy landscape along the ring is basically flat (neutral stability on the ring), such that any point of the line can be represented. Random heterogeneities create hills and valleys in this landscape, so activity drifts towards its local minimum. Homeostatic plasticity has shown to effectively flatten out these local minima such that the representation becomes stable again. In contrast to random heterogeneities, spatially structured heterogeneities are deliberately put into the networks to fulfill a specific function [20, 57], in one case even to counteract the harmful effects of the random heterogeneities [20].

In the present model, we focus on random heterogeneities in the cell parameters, which destroy working memory when present in the interneurons. The cause of this problem is very different from the one in the ring model: Instead of shifting persistent activity into some uninformative region, it is either completely lost or permanently activated without a stimulus (c.f. Figure 1) - in short, the system loses its bistability.

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In the ring model, the instability was compensated using either homeostatic plasticity 837 targeting the activity of pyramidal cells [17], short-term plasticity [18,19] or a 838 systematic structure of the energy landscape, effectively segmenting the line attractor 839 into a discrete set of attractors [20]. In principle, however, this instability could be 840 removed by considering a ring with a sufficiently large number of neurons, which 841 would statistically equalize populations representing different spatial positions. The 842 loss of bistability due to the skewed distribution of interneuron excitability, on the 843 other hand, must be compensated for persistent activity to work at all. These 844 considerations emphasize the need to work with parameter distributions that are 845 derived from experimental studies. 846

The current state of persistence-based working memory models

Persistent activity in cell assemblies in the prefrontal cortex has long been the 848 dominant model for the mechanism of working memory. Recently, this dominance is 849 challenged by a a number of alternative model frameworks which are based, e.g., on 850 short-term synaptic modifications [15], dynamic activity patterns [11], or short-lived 851 attractor states [58], potentially paced by beta, gamma and theta oscillations [59] 852 The theoretical proposals are accompanied by experimental observations that delay 853 activity may be more complex than simply maintaining neurons at elevated firing 854 rates [11,55]. Supporters of persistence-based models counter these arguments showing 855 that the alternative models "could mediate only a limited range of memory-dependent 856 behaviors" and are not mutually exclusive with stimulus-dependent persistent 857 activity [6]. 858

We argue that tests of the current model's predictions would also contribute to the evidence in favor or against the general concept of persistence-based working memory models. As the role of oscillations in working memory has been addressed in a number of recent studies [55, 60, 61], we note that we also see slow, quasi-periodic interruptions of persistent activity in our model (e.g. Figure 1B, frequencies in the delta range, 0.6-2 Hz), and faster (beta/gamma) oscillations can by induced by strongly decreasing the background current into the interneurons while increasing the pyramidal-to-interneuron connections, leaving interneuron activity to be more controlled by pyramidal cell activity and thus enabling a PING-type rhythm (pyramidal-interneuron network gamma [54]). As outlined above, this would lead to a range of assemblies, each of them associated with its own, relatively well defined frequency signature, as seen in [55]. As proposed in [59], increased working memory load would then be associated to more gamma-synchronized assemblies being active at the same time. This would not only result in increased gamma power (e.g. [55, 62]), but also predict that the overall gamma band would be more smeared out, because it would result from several assemblies oscillating at different frequencies. All these proposals can now be tested in a biologically validated modeling framework, contributing to our understanding of the neural underpinnings of working memory.

Methods

Model description

The data-driven model of the prefrontal cortex used here is described in detail in [21]. Briefly, the model consists of 1000 neurons modeled by a simplified version (simpAdEx [28]) of the adaptive exponential integrate-and-fire neuron (AdEx [63]) 847

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which was optimized for high-throughput fitting to *in vitro* electrophysiological data:

$$C \cdot \frac{dV}{dt} = -g_L \cdot (V - E_L) + g_L \cdot \Delta_T \cdot e^{(\frac{V - V_T}{\Delta_T})} + I - w =: w_V - w \tag{1}$$

$$\frac{dw}{dt} = \begin{cases} 0 & \text{for } |w - w_V| > \frac{\tau_m}{\tau_w} w_V \\ \Theta(V_T - V) \cdot \left[1 - \frac{\tau_m}{\tau_w}\right] \frac{dw_V}{dV} \frac{dV}{dt} & \text{otherwise} \end{cases}$$
(2)
if $V > V_{\text{up}}$ then $V \to V_r$ and $w \to w_r = w + b$

if
$$w = \left(1 + \frac{\tau_m}{\tau_w}\right) w_V$$
 then $w \to \left(1 - \frac{\tau_m}{\tau_w}\right) w_V$

where C is the membrane capacitance, g_L a leak conductance (with reversal potential E_L), τ_m and τ_w are the membrane and adaptation time constants, respectively, Θ denotes the heavy-side function, and w_V is the V-nullcline of the system as defined in Equation 1. Like the full AdEx [64], this model consists of one differential equation for the membrane potential V (including an exponential term with slope parameter Δ_T , which causes a strong upswing of the membrane potential once it exceeds V_T), and one for an adaptation variable w, and can reproduce a whole variety of different spiking patterns [28]. A spike is recorded whenever V crosses V_{up} , at which point the voltage is reset to V_r and spike-triggered adaptation is simulated by increasing w by a fixed amount b.

Based on about 200 neurons from rodent PFC, we generated multivariate parameter distributions for five different electrophysiological neuron types [21], namely pyramidal cells in layer 2/3 and 5, fast-spiking, bitufted and Martinotti interneurons. These distributions respect the full covariance structure between the eight parameters of model, which was estimated from the joint data set of all fits of the model to each individual neuron. The simulated neurons are embedded in a laminar and columnar network structure that distinguishes pyramidal cells from superficial (L2/3) and deep layers (L5) as well as four subsets of interneurons which either project within the same layer and column (fast-spiking interneurons), or across layers (bitufted interneurons), columns (large basket cells, with the same electrophysiological properties as the pyramidal cells of the respective layer [65]) or both across layers and columns (Martinotti cells). Neurons from each of these subclasses are randomly connected with different connection probabilities. These values, as well as the statistics on synaptic weights (following a log-normal distribution [66]) and connection delays are directly obtained from the experimental literature. Neurons are connected through conductance-based AMPA-, GABA_A-, and NMDA-type synapses, with kinetics modeled by double exponential functions [39], which time constants again taken from the literature (see Figure SS1 Fig for a schematic of the network architecture and Tables SS1 Table to SS4 Table for a list of the network parameters).

Synapses are equipped with short-term plasticity dynamics implemented by the corrected version [67] of the Tsodyks and Markram model [29]

$$u_k = u_k \cdot R_k \tag{3}$$

$$u_k = U + u_{k-1}(1 - U) \exp(-\Delta_{k-1}/\tau_{\text{fac}})$$
(4)

$$R_k = 1 + (R_{k-1} - u_{k-1}R_{k-1} - 1)\exp(-\Delta_{k-1}/\tau_{\rm rec}).$$
(5)

These recursive equations describe the dynamics of the relative efficiency $a(t_{sp_k})$ across series of spikes, with initial conditions $u_1 = U$ and $R_1 = 1$, where t_{sp_k} is the interval between the (k - 1)th and the kth spike. Model parameters U, τ_{rec} and τ_{fac} were specified according to [30] and [31] who differentiated between facilitating, depressing or combined short-term dynamics, for both excitatory and inhibitory connections. The cell types of the pre- and postsynaptic neurons determine which of these classes is

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used for each individual combination. In particular, connections among pyramidal 914 cells, among interneurons and between fast-spiking interneurons and pyramidal exhibit 915 all three types of connections to varying degrees (pyramidal-pyramidal: 45% E1, 38%916 E2, 17% E3 [31]; interneuron-interneuron: 29% E1, 58% E2, 13% E3 [30], fast-spiking 917 interneuron-pyramidal: 25% E1, 50% E2, 25% E3 [30]). All neurons are driven by 918 constant background currents. The values of these currents are the only parameters 919 which are not directly obtained from the *in vitro* literature, but are estimated in a 920 self-consistent way from the network activity itself (see [21] for details). 921

Importantly, the model has been validated with neuronal data on two levels. On the single-cell level, the fits obtained from the simpAdEx model were tested with *in vitro* data which are fundamentally different from those used for the fit [28], namely fluctuating currents generated by filtering a Poisson spike train with the synapse model described above, as opposed to standard constant DC currents which were used for fitting. The responses of the model neurons to the fluctuating currents matched those of the real neurons within the bounds of their own reliability [28]. On the network level, we compared the activity of the model network which the stationary extracellular recordings from rats *in vivo* during a multi-item working memory task, and intracellular recordings from anesthetized rats. We assessed spike train statistics such as mean, C_V , autocorrelation of interspike intervals and cross-correlations between interspike intervals from pairs of neurons, power spectra of local field potentials and the distribution of membrane potential fluctuations. In almost all of these measures, the distributions from the experimental recordings and the model network were statistically indistinguishable.

Implementation of cell assemblies

For a given simulation, a random population of $N_{\rm CA}^{\rm ex}$ pyramidal cells and $N_{\rm CA}^{\rm inh}$ interneurons was defined as a cell assembly. Synaptic connections within an assembly were strengthened by either increasing all peak conductances (synaptic weights) of existing connections by a factor $s_{\rm CA}$ or by rewiring the neurons with connection probabilities increased by a factor $p_{\rm CA}$. Unless otherwise stated, $N_{\rm CA}^{\rm ex} = 80$ and $N_{\rm CA}^{\rm inh} = 0$ was used. For graphical representations in raster plots, all members of a cell assembly are grouped together in the middle of the respective neuron pool.

Stimulus presentation

External stimuli were simulated by Poisson spike train with firing rate $f_{\rm inp}$ for period 946 of $T_{\rm inp}$ ms in a number of $N_{\rm inp}$ excitatory input neurons. Two cases are considered: In 947 the first case, a single input neuron $(N_{inp}=1)$ is randomly connected to L2/3 cells, 948 using the connection probability for pyramidal cells within that layer. In the second 949 case, there is one input neuron for each pyramidal cell in the assembly $(N_{\rm inp} = N_{\rm CA}^{\rm ex})$, 950 with connections from each input neuron to one of the assembly neurons. In both cases, 951 all synaptic peak conductances of the input are set to s_{inp} times the peak conductance 952 of excitatory synapses within the network. Unless otherwise stated, the following 953 values are used for these parameters: $f_{inp} = 1000 \text{ Hz}$, $T_{inp} = 50 \text{ ms}$ and $s_{inp} = 1$. 954

Measures of persistent activity

To quantify to which degree a cell assembly exhibits persistent activity, we define the measure d_{PA} as follows

$$d_{\rm PA} = \Theta(a_{\rm end} - a_{\rm start}) \left(a_{\rm end} - a_{\rm start} \right), \tag{6}$$

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 a_{end} and a_{start} are normalized measures of the activity during 500 ms before the stimulus and the last 500 ms of the simulation, respectively. Θ is the heavy-side function, setting d_{PA} to zero if activity is higher in the beginning (i.e. $a_{\text{end}} - a_{\text{start}} < 0$). The activity measures are computed by comparing the firing rate in the cells of the assembly with those outside the assembly. An assembly cell is regarded as activated if its average firing rate during the observed time interval exceeds those of the average firing rate of all non-assembly (pyramidal, layer 2/3) cells. Normalized activity a is then defined as the fraction of activated cells in the assembly. Thus, by definition, d_{PA} ranges between zero and one. The maximum of one is reached when all cells in assembly are activated at the end of the simulation and none of them before the stimulus, which reflects the desired case of stimulus-induced persistent activity. On the other hand, d_{PA} is zero if either none of the assembly cells is activated in the end (decayed activity) or more cells are activated before the stimulus $(a_{\text{end}} - a_{\text{start}} < 0$, spontaneous persistent activity).

Variation of model parameters

We systematically varied a number of parameters of the model in order to find configurations allowing for persistent activity. In particular, we varied the size of the cell assembly between 30, 50, 80 and 470 neurons (the latter being the number of neurons in $L^{2}(3)$, the ratio between NMDA and AMPA conductances between 1, 2, 3, 4, 6 and 8 times the original value, the peak conductance among pyramdial cells between 1, 5, 10, 15 and 20 times the original value, the scaling factor s_{CA} of the synaptic weights within the cell assembly between 1, 5, 15 and 20, connectivity within the cell assembly between 10 and 40% and both background currents I_{ex} into pyramidal cells and $I_{\rm inh}$ into interneurons between 0 and 1000 pA. We conducted all of these changes in isolation (using a default of 80 cell assembly neurons, the original ratio of NMDA and AMPA conductances as well as the original value of peak conductances among pyramidal cells, a scaling factor for the weights within the cell assembly of 5 and a connectivity of 10% as well as $I_{\rm ex}$ and $I_{\rm inh}$ values of 300 and 200 pA, respectively) and also assessed a considerable number of these modifications together. Overall, we performed 720 simulations with the full model, summarized in Figure 1C (except for those where the full $L_2/3$ was used as a single assembly, as they exhibit epiletiform activity following stimulus presentation).

Minimal model of persistent activity

To assess the dynamic mechanisms of persistent activity and the effect of rheobase 991 heterogeneitities in more detail, we employ a minimal model of persistent activity [37] 992 comprising a single pyramidal cell modelled by a simplified version of the 993 Hodgkin-Huxley equations [38] that excites itself by an NMDA autapse and also 994 inhibits itself by a GABA autapse. We modify this model in three ways: First, we 995 decrease the strengths of the synaptic weights to 10% of the original values. This 996 allows for firing rates (80 Hz) that are closer to the low rates (about 20 Hz) seen in 997 our full simulations (compared to over 200 Hz at the original weights, c.f. Figure 12A). 998 Further decrease of the weights would have required too much fine-tuning to show the 999 principle results, but we also discuss the effect of the scaling in the main text. Second, 1000 we filter the inhibitory conductances by a function G_I that mimicks interneuron 1001 heterogeneity. This function is described in details below. In some simulations 1002 (indicated at the text), we apply a function G_E of the same form to the excitatory 1003 conductances, but with different parameters. Finally, we simulate a population of 1000 1004 pyramidal cells instead of one, which only differ in their particular realization of G_I or 1005

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 G_E . This population of models (c.f. [52]) allows to assess the impact of heterogeneity on the dynamics of the pyramidal cell.

The equations governing the dynamics of one of the pyramidal cells are given by:

$$-C_{m}\dot{V} = I_{L} + I_{Na} + I_{K} + sI_{NMDA} + sI_{GABA} + I_{ext}$$
(7)

$$I_{L} = g_{L}(V - E_{L})$$
(7)

$$I_{Na} = g_{Na}m_{\infty}(V - E_{Na}),$$
(7)

$$m_{\infty} = [1 + \exp((V_{hNa} - V)/k_{Na})]^{-1}$$
(7)

$$I_{K} = g_{K}n(V - E_{K}),$$
(7)

$$I_{NMDA} = g_{NMDA}(s_{NMDA} + s_{base}) [1 + 0.33 \exp(-0.0625V)]^{-1} (V - E_{NMDA})$$
(7)

$$I_{GABA} = (G_{I} (g_{GABA}(s_{GABA} + s_{base})) (V - E_{GABA})$$
(8)

$$\dot{n} = \frac{n_{\infty} - n}{\tau_{n}}, \qquad n_{\infty} = [1 + \exp((V_{hK} - V)/k_{K})]^{-1}$$
(8)

$$s_{N\dot{M}DA} = H(V) (1 - s_{NMDA}) - s_{NMDA}/\tau_{NMDA}$$
(9)

$$H(V) = [1 + \exp(10(V_{spike} - V))]^{-1}$$

$$s_{\text{GABA}} = H(V) \left(1 - s_{\text{GABA}}\right) - s_{\text{GABA}} / \tau_{\text{GABA}}$$
(10)

By applying the function G_I , we interpret the inhibitory conductance as an input 1009 to a population of interneurons, which are modelled as rectified linear units (ReLU) 1010 representing the f-I curve of the interneurons, each with a different rheobase drawn 1011 from an normal distribution. The conductance is first converted into an input current, 1012 second into a firing rate and third back into a conductance. In this way, one inhibitory 1013 conductance is being generated for each interneuron and all these conductances are 1014 averaged over all interneurons projecting to the pyramidal cell, which is the output of 1015 G_I . The first two steps - conversion of the conductance into a current and into a firing 1016 rate - are being conducted in a self-consistent way by finding the intersection between 1017 the f-I curve of the current interneuron and a curve describing the current that is 1018 being generated at a given firing rate, given the input conductance. That second curve 1019 is computed from the numerically obtained relation between firing rate and average 1020 membrane potential during the interspike interval of the simulated neuron and the 1021 relation between current I and conductance g, I = g(V - E). Once the firing rate is 1022 obtained, it is multiplied with the GABA peak conductance g_{GABA} , which is 1023 equivalent to the computation of the average over a long random spike train at that 1024 rate, convoluted with the response function of the synapse, given a normalization of 1025 the area under that function to one. We tested different variants of implementing G_I 1026 and found that results did not qualitatively depend on these details. 1027

The different pyramidal cells in the population differ only in the inhibitory input they receive via G_I . In particular, we randomly pick 20% of the simulated interneurons (200 by default) and construct the synaptic input in G_I only from these 20%.

The phase space of the model is being constructed by considering one pair of 1031 subsequent spikes at a time and computing both the instantaneous firing rate FR 1032 (inverse of the interspike interval) and the average of the input currents 1033 $(I_{\rm syn} = I_{\rm NMDA} + I_{\rm GABA} + I_{\rm ext})$ over that interval. The time series of these two 1034 variables over all pairs of spikes constitutes the trajectory of the system in the 1035 phasespace. The nullclines were being computed by varying the input current into the 1036 cell and compute the average firing rate for each current without synaptic feedback 1037 (FR nullcline) and the mean synaptic current produced at each of these rates (I_{syn}) . 1038 Intersections of the nullclines constitute fixed points of the system and we verified that 1039 the trajectories converged to one of these fixed points for different parameter sets. 1040

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To classify neurons as bistable, active or inactive, we compute the intersections 1041 between FR and $I_{\rm syn}$ nullclines for each of the simulated pyramidal cells. Cells with 1042 three intersections are labeled as bistable, cells with only one intersection at zero firing 1043 rate as labeled as inactive and cells with one intersection at non-zero firing rates are 1044 termed active. Importantly, in real, connected network, the number of bistable cells 1045 also affects the shape of the $I_{\rm syn}$ nullclines: Inactive pyramidal cells in the cell 1046 assembly do not contribute to the excitatory feedback and constantly active cells also 1047 need to be excluded from the assembly to avoid premature activation before a 1048 stimulus arrives. We mimick these constraints by scaling the excitatory peak 1049 conductance by the fraction of bistable neurons. After that, the numbers of each types 1050 of cells is counted again and the procedure is repeated until convergence to a fixed 1051 number of each type (always the case after at most five iterations). 1052

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Supporting information

S1 Fig. Schematic of the network architecture. (A) Laminar structure of a single network column (only one column is simulated in this study). Arrow widths represent relative strength of connections (black: excitatory, gray: inhibitory), i.e. the product of connection probability and synaptic peak conductance. (B) Left panel: Distribution of three different short-term plasticity types over different combinations of pre- and postsynaptic neuron types. Arrows from or to one of the shaded blocks (rather than from or to a single neuron type) denote connection types that are identical for all excitatory (PC) or inhibitory (IN) neurons. Where all three types are drawn, they are randomly distributed over all synapses between these two neuron types according to the probabilities given in the figure (termed "heterogeneous STP" in the the text). Right panel: Illustration of the postsynaptic potential in response to a series of presynaptic spikes for three types of short-term synaptic plasticity for excitatory (E1 to E3) and inhibitory synapses (I1 to I3). The figure is taken from [21].

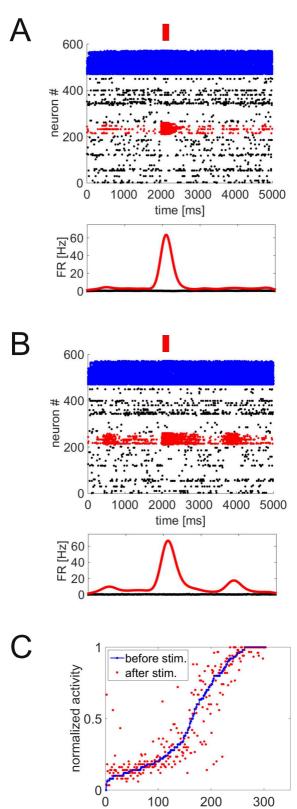
S1 Table. Neuron parameters. Mean and standard deviation of the parameters of the simpAdEx model for the five different neuron types used in the network (PC: Pyramidal cell, FS: Fast-spiking interneuron, BT: Bitufted interneuron, MC: Martinotti cell). The table is taken from [21].

S2 Table. Cell numbers. Relative numbers of cells for each type. PC: pyramidal cell, IN: interneuron, see Materials and Methods section in [21] for interneuron subtypes. The table is taken from [21].

S3 Table. Synaptic parameters. Mean \pm standard deviation of the parameters of the synapses connecting the different pre- and postsynaptic neuron types (p_{con} : connection probability, g_{max} : peak conductance, τ_D : transmission delay). The table is taken from [21].

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S4 Table. Short-term synaptic plasticity. Mean (standard deviation) of the parameters of the six types of short-term synaptic plasticity. The table is taken from [21].



simulation #

