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**PHYSICOCHEMICAL PROPERTIES AND STRUCTURAL CHARACTERISTICS OF
WHOLE GRAIN *Oryza sativa* L. WITH DIFFERENT TREATMENTS**

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Abstract

Physicochemical properties and structural characteristics of whole rice flours with different treatments (soaking, germination and extrusion cooking) were studied. Water solubility (WS), water absorption (WA), crystallinity, adsorption isotherms (BET and GAB models) and glass transition temperature (T_g) of the samples were determined. WS and WA were enhanced by extrusion cooking process (3.17-4.98 vs. 24.1-53.76 g/100g and 2.77-3.05 vs. 4.46-7.04 ml/g, respectively), but crystallinity was decreased (30-33 vs. 4-16 %). Adsorption isotherms showed that extruded samples exhibited higher equilibrium moisture content as compared with their corresponding non extruded samples (5.0-19.2 vs. 4.0-16.1g water/g solids). There were no changes in T_g values in the studied moisture range (3.8 - 16 g/100g). These results allow the correctly use of whole rice flours with different treatments in foods and also contributed to the knowledge of stabilization of the products.

Keywords: whole rice, soaking, germination, extrusion cooking, adsorption isotherms, glass transition temperature.

1. Introduction

Rice (*Oryza sativa L.*) is one of the major cereal crops and a staple food for the world's population. Compared to white rice, brown rice (WG's: whole grains) is nutritionally superior, rich in dietary fiber, antioxidants, protein, minerals, and vitamins. All of these compounds are present in the bran layers and germ (Jones & Engleson, 2010). Even though WG's consumption is recommended, the incorporation into the diet is low because of the lack of habits and difficulty to incorporate them into processed foods. Thus, research on food formulations is needed. Several traditional food-processing and preparation methods can be used to enhance the bio-accessibility of nutrients in plant-based diets. These include thermal processing, mechanical processing, soaking, fermentation, and germination/malting. These strategies aim to decrease the content of non nutritional components, such as phytates, or increase the bio-accessibility of micronutrients. Regarding that, Albarracín et al. (2015) showed 50% and 77% reduction of phytates after soaking and soaking-extrusion, respectively. Also, a 50% reduction of phytates was observed after germination-extrusion process (unpublished results). Extrusion cooking is a multi-stage unit operation that combines transport, mixing, working and forming. It has diverse applications, from de-polymerization of starch for ethyl alcohol fermentation to pet food production. It is a high-temperature short-time (HTST) process in combination with high mechanical shear, which makes it a good 'model' to analyze processing effect on product properties (González et al., 2002). The high shear forces and temperature during extrusion cooking severely affect bio-macromolecules by breaking covalent bonds. Additionally, Maillard reactions may occur between reducing sugars and amino acids of proteins (Mercier, 1993).

To explain the behavior of food components during processing and storage, food structure should be considered as a natural system plasticized by water (Matveev et al., 2000). This interpretation provides a way to physically characterize the system and to determine safe storage conditions for each food. The physical, chemical and microbial changes in foods have conventionally been described using the concept of water activity (a_w). It has provided a reliable

estimation of microbial growth, lipid oxidation, non enzymatic and enzymatic activities, and the texture/mouthfeel of foods.

BET (Brunauer, Emmett and Teller) and GAB (Guggenheim, Andersen and De Boer) are typical water sorption models used for foods. The classical BET multilayer sorption equation is used to calculate monolayer values in very different physicochemical fields because of the simplicity of its application and due to it has the approval of the International Union of Pure and Applied Chemistry (IUPAC) (Timmermann, 2003). According to the GAB model, the sorption state of the sorbate molecules in the layers beyond the first one (monolayer) is the same but different to that of the pure liquid. The extra assumption of the GAB model over the BET formulation demands the introduction of the additional constant **k**. This constant is just the measure of the difference of free enthalpy of the sorbate molecules in these two states, the pure liquid and this second sorption stage, the layers above the monolayer (Timmermann, 2003).

Moisture content changes during processing affect the thermal properties of a food system. Water is a very effective plasticizer and will reduce T_g . It is also known that when food structure is in the glassy state, shelf life is prolonged. Glass transition concept appeared as a powerful tool for understanding mechanisms of the processes controlling shelf life of food. The glass transition temperature (T_g) could be taken as a reference temperature, which is important when predicting the optimal conditions for drying, agglomeration, and storage (Roos & Karel, 1991). At storage temperature (T_s) lower than T_g , the molecular mobility is restricted and the food is stable, but at T_s higher than T_g , instability processes (physical, chemical and biological) would be controlled by the difference between T_s and T_g .

The aim of this work was to study the effect of soaking, germination and extrusion cooking on water absorption, water solubility, crystallinity, adsorption isotherm and glass transition temperature of whole grain rice flours in order to analyze the changes on hydration properties and structural characteristics caused by the above processes.

2. Materials and methods

2.1 Raw samples

Rough and brown long rice Fortuna Type (*Oryza sativa*) were provided by Los Cerrillos S.A (Santa Fe, Argentina).

2.2 Processed samples

A total of six processed samples were studied: brown rice (BR), extruded brown rice (EBR), soaked rice (SR), extruded soaked rice (ESR), germinated rice (GR) and extruded germinated rice (EGR).

2.2.1 *Brown rice (BR)*: Brown long rice was provided dehulled by industry. Rice grains were grinded according to a diagram developed in our laboratory. It was done with a roller mill (Bühler-Miag, Braunschweig, Germany) in such a manner to avoid the production of too many fine particles (less 250 μm). Particle smaller than 420 μm represented less than 3% but were included in the whole sample for extrusion cooking. A progressive and successive reduction in distance between rollers from 1 to 0.5 and 0.25 mm was used. After each grinding step, the flour was screened through 1.190 mm sieve. The thicker particles (> 1.190 mm) were re-ground until the whole sample was obtained.

2.2.2 *Soaked rice (SR)*: An amount of 3 kg of rough rice was previously washed in a 0.2 % NaClO solution during 15 minutes and then soaked in a 5.5 g/L lactic acid solution at 45 °C for 24 h (Albarracín et al., 2015). After that, the grains were washed with tap water, dried to almost 13 g/100 g moisture content using a tray dryer at 40 °C, dehulled in a laboratory rice mill (Galicet, San Salvador, Entre Ríos, Argentina), and grinded to grits (1190 - 420 μm particle size), as mentioned in 2.2.1.

2.2.3 *Germinated rice (GR)*: An amount of 3 kg of rough rice was steeped in distilled water at 20°C during 24h. Then, the water was drained, and the moistened rice grains were germinated during 24 h by putting the rice in an oven at 35 °C and 98% relatively humidity, according to the previous conditions studied. Finally, germinated rice was dried 24 h at 40 °C, dehulled and grinded to grits (1190 - 420 μm particle size), as mentioned in 2.2.1.

2.2.4. Extruded brown rice (EBR), extruded soaked rice (ESR), and extruded germinated rice (EGR). Rice grits obtained in 2.2.1, 2.2.2 and 2.2.3 were conditioned to the adequate moisture (M) 1 h before each run and extruded with a Brabender 20DN extruder, using a 4:1 compression ratio screw, a 3/20-mm (diameter/length) die, and a screw speed of 150 rpm. The feeding rate of the extruder was at full capacity. While the extruder feeding section was maintained cool by circulating water through the jacketed device, the metering and die sections were both kept at the temperature corresponding to each run by using the heat control device of the extruder. The extrusion cooking conditions: 160 °C -14 %M for EBR; 160 °C 16.5 %M for ESR; 175 °C 14 % M for EGR were selected according to preliminary studies (González et al., 2013a) taking into account good expansion, sensorial hardness and flavor.

2.3 Analytical Methods

All samples were ground using a Sample Mill Ciclotec (USA), with a 1 mm sieve.

2.3.1 Composition

Total nitrogen was determined by the semi micro Kjeldahl method (AOAC 920.53, 1999) using the value of 5.95 as nitrogen/protein conversion factor, according to Juliano (1985). Lipids (petroleum ether extract), moisture and ash contents were determined using AOAC (1999) approved methods. Total amount of fiber was determined by AOAC 985.29 method adopted by Megazyme ® commercial kit. Total starch (%) was quantified according to Tovar et al. (1990). Glucose oxidase/peroxidase assay kit (Wiener Lab, Rosario, Argentina) was used for colorimetric determination of glucose, and concentration was determined at 505 nm using a spectrophotometer (*Milton Roy Genesys 5*, Ivyland, USA). Calculation of sample glucose concentration was done using a glucose standard. Available carbohydrates (d.b) were calculated by difference.

Free fatty acids (FFA) were analyzed according to Lowry and Tinsley (1976), previous extraction of the lipids with the Folch procedure. The results were expressed as mg lauric acid/g sample using a Lauric Acid standard calibration curve.

Thiobarbituric acid reactive substances (TBARS) assay according to Siu and Draper (1978) was performed. Results were expressed as μmol of MDA/g sample using the molar absorption coefficient of the malondialdehyde ($\text{MDA} = 0.156 \text{ mM}\cdot\text{cm}^{-1}$).

2.3.2 Water absorption and water solubility

Water absorption (WA), as spontaneous uptake of water, was determined using Baumman method according to González et al. (2002). About 50 mg of sample was spread on a filter paper, and the volume of absorbed water was measured after the equilibrium was reached (approximately 20 minutes). The result was expressed as mL water/g sample d.b.

Water solubility (WS) was evaluated by the method of Gonzalez et al. (2002). An aliquot of 1.25 g of ground sample was dispersed in 25 mL of water, stirred for 30 minutes and centrifuged at 2000xg for 30 min at 25 °C. The supernatant was dried in an oven at 105 °C and the soluble solids determined by weight. The solubility was calculated as: $\text{WS}\% = \text{g soluble solids}/100\text{g sample d.b.}$

2.3.3 Crystallinity

Starch structure was evaluated using X-Ray diffraction techniques. X-Ray diffraction patterns were recorded using a Shimadzu DX-1 X-Ray diffraction system (Shimadzu, Tokyo, Japan). All samples were analyzed between $2\theta=10^\circ$ to $2\theta=30^\circ$ using $\text{K}\alpha\text{Cu}$ radiation, 30 Kv and 40 mA with a step size of $0.5^\circ/\text{min}$. Crystallinity percent (%) was determined by deconvolution of the identified peaks (Colonna et al., 1987) using Origin 7.5 and expressed as the ratio of the sum of the areas corresponding to those peaks and the total area.

2.3.4 Adsorption isotherms

To obtain the adsorption isotherms, samples were dried in an oven (Selecta, Barcelona, Spain) at 40 °C under vacuum. The samples were introduced in desiccators (at 24 °C) using oversaturated solutions to equilibrate the water activity (a_w) to their corresponding generated relative humidity/100. The salts and the respective a_w used were LiCl: 0.113, CH_3COOK : 0.230, MgCl_2 : 0.330, K_2CO_3 : 0.430, $\text{Mg}(\text{NO}_3)_2$: 0.520, CuCl_2 : 0.680, NaCl: 0.755. The sample weights were

controlled till a constant value ($\Delta m < \pm 0.0005$ g) was reached, where the equilibrium was assumed. The equilibrium moisture content was determined in an oven at $105 \text{ }^\circ\text{C} \pm 1 \text{ }^\circ\text{C}$ during 24 h. Experimental adsorption isotherms were fitted to Brunauer–Emmett–Teller (BET) (Eq. 1) (Brunauer et al., 1940) and Guggenheim–Andersen–de Boer (GAB) (Eq. 2) (Van den Berg et al., 1981) linearized models.

$$w_e = \frac{w_o \cdot C \cdot a_w}{(1 - a_w) \cdot (1 + (C - 1) \cdot a_w)} \quad (1)$$

$$w_e = \frac{w_o \cdot C \cdot K \cdot a_w}{(1 - K \cdot a_w) \cdot (1 + (C - 1) \cdot K \cdot a_w)} \quad (2)$$

Where: w_e , water content at equilibrium (g water/g dry solids); a_w , water activity; w_o , monolayer value (g water/g dry solids); C , constant related to monolayer sorption heat and K , constant related to multilayer sorption heat.

2.3.5 Glass transitions temperatures (T_g)

To determine the glass transition temperature of the samples, a differential scanning calorimeter (DSC1, Mettler Toledo, Switzerland) was used. About 10-20 mg of each equilibrated sample was placed into DSC aluminum pans (ME-000267631, Mettler Toledo, Switzerland), sealed and analyzed. Each sample was analyzed in triplicate. The heating rate was $20 \text{ }^\circ\text{C}/\text{min}$ and the temperature range varied between 0 to $95 \text{ }^\circ\text{C}$.

2.3.6 Statistical Analysis

Each assay was performed by triplicate. Analysis of variance was carried out using Statgraphics Plus 5.1 software (Statistical Graphics Corporation). The statistical differences among samples were determined using LSD test (Least Significant Difference). The accepted level of significance was $p < 0.05$.

3. Results and discussion

Table 1 shows the composition of the samples in dry basis. Moisture content ranged between 9.9 and 12.7 g water/100 g of samples for extruded and non extruded samples, respectively. Crude protein and ash content did not change by different treatments. Lipid content (ether extract) of SR was lower than other non extruded samples, probably due to some germ losses during dehulling of soaked grains. Extrusion cooking process reduced ether extract respect to non extruded samples as it was shown by Percibaldi (2003). Moreover, some triglyceride hydrolysis was produced during extrusion cooking and fatty acids in the material could form complexes with amylose, making it more difficult to quantify crude fat (Ruiz-Ruiz et al., 2008).

Table 1 shows also FFA and TBARS of the samples. FFA decreased by 80% due to extrusion cooking in each whole grain rice product as compared with the brown rice without treatment (255.27 vs. 36.89; 28.84; 31.65 mg lauric ac./g for EBR, ESR and EGR, respectively). This can be due to the complex chemical reactions involving lipids in the extrusion cooking as it was shown by Percibaldi (2003). Also, 42 % reduction of FFA was obtained after soaking process and 47% after germination process (147.51 and 135.77 mg lauric ac./g, respectively), probably due to the losses of FFA into the soaking water.

The TBARS assay indicated that the results of processed samples by extrusion cooking were 2-folds higher than non-extruded ones and the GR sample had higher values than the BR and SR. Nevertheless this increase in the TBARS assay would not be significant for the stability of the samples. Also, the development of new whole cereals based ingredients always implies an antioxidant agent addition.

Total fibre did not change by soaking or germination, but extrusion cooking decrease fibre content. Mercier (1993) found that the effect of cooking on the dietary fiber extrusion cooking and physiological properties depends on the extrusion cooking conditions. Generally, under high temperature and low screw speed and moisture content, a solubilization of insoluble dietary fiber is produced by increasing its breakdown. In soft or moderate conditions, fiber content does not

change, but some components are solubilized. The shear forces can break the glycosidic linkages of the cellulose, degrading other components of the fiber, and increasing the soluble fiber at the expense of the insoluble one (Björck & Asp, 1983). Drago et al. (2010) also observed a decrease of total dietary fibre in extruded whole grain maize.

Total starch content in the samples did not change by the treatments; the range was between 73.53-80.19 g/100 g d.b., similar to that reported by Juliano (1985). Significant differences among BR and the other raw samples (SR and GR) were due to BR was dehulled at the industry and SR and GR were dehulled at pilot plant. Available carbohydrates were higher in extruded samples because of macromolecular degradation, which depends on extrusion cooking parameters such as temperature, moisture and screw speed (Mercier, 1993).

Table 2 shows water solubility (WS) and water absorption (WA) of extruded and non extruded samples. WS and WA of non extruded samples were in the expected range, and there were no significant differences among them, indicating that soaking and germination did not modify these physical properties. However, extruded samples showed significant differences for both, WS and WA. WS is an indicator of starch granule disruption degree and was used to evaluate the degree of cooking (DC). Results showed that soaking and germination significantly affected DC attained during extrusion cooking of whole rice (EBR: 24.1 vs. ESR: 53.76 and EGR: 42.83 g/100g). The highest DC (highest WS) showed by ESR sample could be due to the acid treatment during soaking, which would allow starch granules have a higher disruption during extrusion cooking in comparison with extruded brown rice. On the other hand, germination process activates enzymatic systems, particularly proteases, which could disrupt protein network and favors thermo-mechanical effect on starch granules, giving higher values of WS. These results suggest that as pre-treatment, acid soaking allows obtaining a higher DC in extruded whole grain samples than germination.

Values of WA (**Table 2**) are in the range of those obtained for brown rice at similar extrusion cooking conditions in a previous work (González et al., 2013a). WA is directly related with WS

until the maximum WA is reached. This maximum depends on the material and extrusion cooking conditions, but indicates a point from which the proportion of disrupt starch granules predominate in extruded sample, so that a further increment of DC would produce a decrease in WA (González et al., 2013b). This effect is shown in **Figure 1**. It was observed that the lowest value corresponded to ESR (5.81 ml/g of sample), indicating higher DC.

Table 2 shows the angles and intensity of crystalline peaks associated to starch crystal forms and crystallinity percentage in the studied samples, and **Figure 2** shows the diffractogram patterns of non extruded and extruded samples corresponding to each treatment. Extrusion cooking destroys the organized crystalline structure either partially or completely, depending on the extrusion cooking variables such as moisture and shear (Colona et al., 1989). Native starch has a semi-crystalline structure, since amylopectin in rice starch is organized in a monoclinic crystal lattice called A-type (Hoseney, 1986). In BR, SR, and GR the typical structure of native starch with the three picks around at 15.2°, 17.4° (with another at 18.3°) and 23° were observed. The slightly higher value of crystallinity for SR in comparison with the other two non-extruded samples (BR and GR) would be attributed to an annealing effect caused by the heat treatment during soaking process (45 °C during 24 hours) resulting in a more ordered crystalline structure of starch (Shandera & Jackson, 1996). Extruded samples showed the disappearance of native crystalline structure and the formation of a new crystal with two picks (2 θ : 12° and 18°) called “E-type” crystal. According to Colonna et al. (1987), this structure corresponds to the typical amylose-lipid complex. EBR showed a higher value of crystallinity than the other extruded samples, probably due to native starch remaining, which is in agreement with the lowest WS value in comparison with the other two extruded samples (ESR and EGR). Gonzalez et al (2013b) showed in their work that brown rice is not completely cooked at similar extrusion cooking conditions. Moreover, soaking and germination would induce structural starch changes which imply different behavior during extrusion cooking.

Figure 3 shows water adsorption isotherms at 24 °C of all the samples. Adsorption data showed the typical sigmoid shape type II isotherms, frequently found in cereal foods. The adsorption region corresponding to the lowest a_w represents monolayer adsorption, where most of the sites are charged or rich in polar groups, such as OH groups. In this region, water molecules are strongly bound on polar sites via hydrogen bonds (Van den Berg, 1986). As expected from the analysis of WS, WA and x-ray diffraction, extruded samples showed higher equilibrium moisture content than non extruded ones. These could be as a consequence of the disruption of the structure of components during extrusion cooking, mainly those of starch (amylose and amylopectin), which are broken to dextrin and also proteins (Donkor et al., 2012). The total effect of mechanical energy input is complex because it has at least two different roles. First, it promotes the disintegration of hydrated starch granules, protein bodies and lipid components, and mechanically mixes these fragments. Second, it raises the rate of chemical reactions, particularly the breaking of carbohydrate and protein chains (Kokini et al., 1992). The effect of extrusion cooking on water adsorption was enhanced by a previous grain germination process which would produce an increase in free amino and carboxylic groups probably because of the biosynthesis of new compounds (Kim et al., 2012).

The values of the BET and GAB parameters are shown in **Table 3**. In **Figure 3** GAB model appear fitted to the experimental points of the isotherms. The monolayer water content (w_0) estimated by GAB was higher than that obtained by BET. Timmermann (2003) attributed inequality in monolayer water values to the physical and mathematical nature of the models. w_0 represents the moisture content at which the water is attached to each polar and ionic groups. BET monolayer water content value is more widely accepted, as the BET model has a true physical sense in the range of low a_w , related to the adsorption theory, while the GAB model is a more empirical one that provides accurate prediction over the water activity range up to 0.75. Theoretically, both methods should provide similar monolayer value. However, as BET isotherm

fitted well in the range of water activities of 0 to 0.43, adequate range for the calculation of parameters w_0 and C for extruded products, this monolayer water content would be preferred.

According to BET, extruded samples showed higher w_0 than the corresponding sample without extrusion cooking, and GR and EGR have higher w_0 and C values, meaning that these samples would have higher values of heat of water interaction than the other samples. For GAB, SR showed the lowest value and GR the highest one for w_0 in extruded and non-extruded samples.

In both models, C values were higher in extruded samples because they adsorbed more water, increasing heat of sorption. As it was mentioned above, this effect could be due to the extrusion cooking process that breaks some linkages and generates larger available surface area and polar groups for adsorption.

Gonzalez et al. (2004) showed that expanded product based on corn flours would lose their crispness when a_w exceed 0.33 (around 0.065 moisture content dry basis). Although the moisture content of corn extruded product were lower than that of extruded whole rice product (**Figure 3**), our results would suggests that in the case of extruded whole rice products stored at the same moisture content (around 5%), those samples with higher w_0 will have lower a_w , thus EGR will keep the crispness properties in a wider range of moisture.

In spite of the structural changes in starch fraction caused by the different treatments discussed above, no changes on Tg values were observed, neither by the treatment nor by the change of water content on samples. **Figure 4** shows the obtained thermograms of the different samples at the same water content. As can be seen, this figure allows evaluating the onset, midpoint, endpoint of Tg and ΔC_p . **Table 4** shows midpoint Tg and ΔC_p for BR and EBR samples at different water contents.

Despite no differences in Tg were observed, a difference in ΔC_p between BR and EBR can be observed. These would indicated that water-solid energy of interaction is higher in extruded sample (EBR) in comparison with non-extruded one (BR), being in agreement with the results of C values discussed above.

Recent available information on the evaluation of Tg of whole grain flour is poor (Herawat et al., 2014). Perdon et al. (2000) studied the glass transition of brown rice kernels of different type of rice using three methods, DSC, TMA (thermal mechanical analysis) and DMA (dynamic mechanical analysis). These authors showed that Tg was not affected by the changes in moisture content in the range used in our study (around 4 to 17 g of water/100 g of sample). Previous studies showed that DSC was not sensitive enough to detect thermal transition in rice samples. Sun et al. (2002) reported that DSC was not sufficiently sensitive to accurately measure rice kernel Tg due to rice kernel being a partially crystalline/partially amorphous. The composition of the samples would contribute to the poor sensitivity for Tg detection. If most of the carbohydrates, proteins and fiber are non-soluble, especially in the case of non-extruded samples (with a very low WS) or non-compatible with water (may be in the case of the extruded samples), and appear in the rice defining its own phase, a low amount of amorphous phase is expected to be formed. These, together with the low quantity of sample that can be analyzed by DSC, justify the low sensitivity observed.

The fact that no changes on Tg were observed, neither by the change of moisture for BR and EBR nor by any of the treatment used in this study, prevent us to make any interpretation about phase transition behavior and structural changes. On the other hand, breakage of starch and proteins during extrusion, especially in grains previously germinated, increases the hygroscopicity of the samples. The newly formed compounds would make all the samples equally sensitive to the glass transition. From this point of view, the evolution of all of them during storage will be of the same order; suggesting that the structure of extruded whole rice would be stable in a wider range of moisture content. Moreover the glass transition temperature is high enough (about 50 °C) to ensure the stability of the product.

4. Conclusions

Soaking and germination treatments applied to rough rice increase the degree of cooking attained during extrusion cooking respect to the extruded brown rice. This effect was higher for soaked

samples than for germinated ones. Soaking of rough rice also produced an annealing effect in starch structure. Extruded samples showed higher equilibrium moisture content as a consequence of structure components disruption (starch and proteins) during extrusion cooking. The effect of extrusion cooking on increasing equilibrium moisture would be enhanced by a previous grain germination process. The Tg of the samples was not affected neither by the water content nor by the process.

Soaking, germination and extrusion processes could be used at industrial scale because they are low cost, simple and efficient methods for producing physicochemical changes in cereals. Modified WG flours could be used in the production of low phytate content expanded products (snacks) or precooked flours for development of WG foods.

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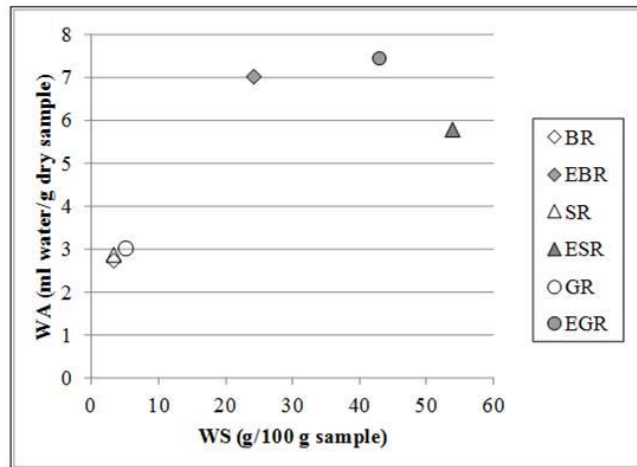


Figure 1: WA and WS relationship of the non extruded and extruded samples. BR: brown rice; EBR: extruded brown rice; SR: soaked rice; ESR: extruded soaked rice; GR: germinated rice; EGR: extruded germinated rice.

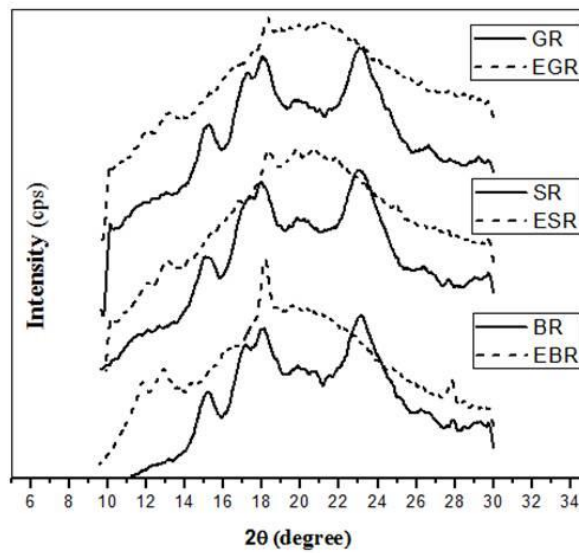


Figure 2: X-ray diffraction patterns of Brown Rice (BR), Extruded Brown Rice (EBR) Soaked Rice (SR), Extruded Soaked Rice (ESR), Germinated Rice (GR) and Extruded Germinated Rice (EGR) samples.

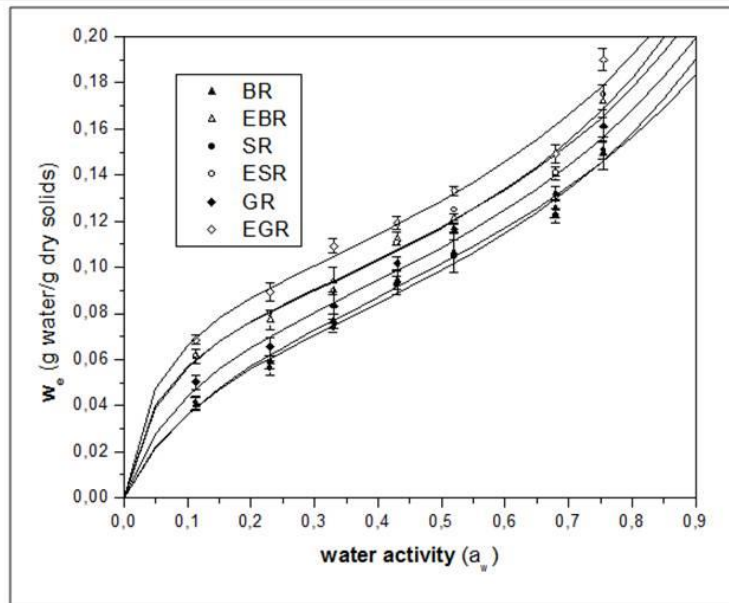


Figure 3: Adsorption isotherms of Brown Rice (BR), Extruded Brown Rice (EBR), Soaked Rice (SR), Extruded Soaked Rice (ESR), Germinated Rice (GR) and Extruded Germinated Rice (EGR) at 24 °C. [Experimental points and GAB fitted model (lines).]

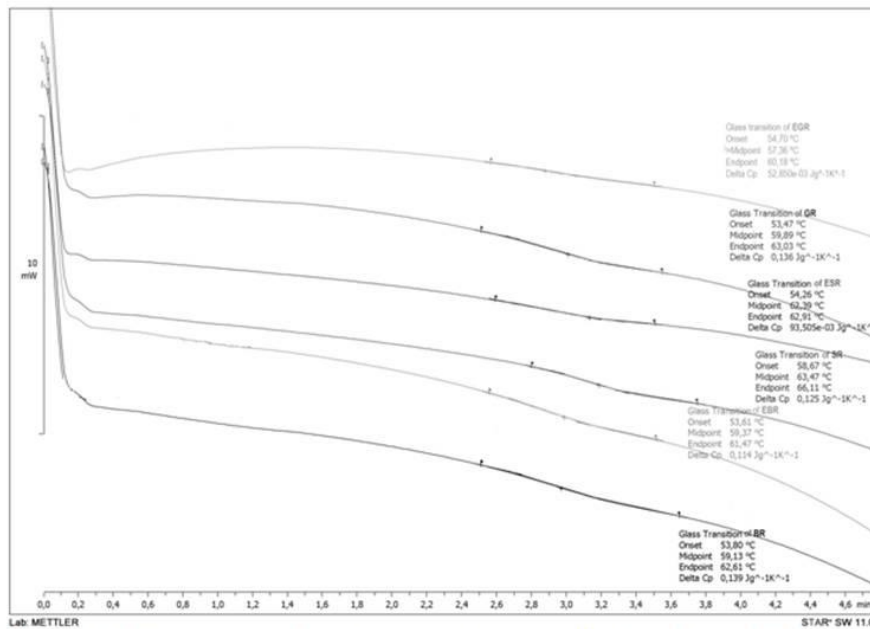


Figure 4: DSC thermograms of the different samples Brown Rice (BR), Extruded Brown Rice (EBR), Soaked Rice (SR), Extruded Soaked Rice (ESR), Germinated Rice (GR) and Extruded Germinated Rice (EGR) at the same moisture content (6 g water/100 g dry matter).]

Table 3: Model parameters of water adsorption isotherms (BET and GAB) of Brown Rice (BR), Extruded Brown Rice (EBR), Soaked Rice (SR), Extruded Soaked Rice (ESR), Germinated Rice (GR) and Extruded Germinated Rice (EGR).

Models	BR	EBR	SR	ESR	GR	EGR
BET						
w_0 (g water/g dry solids)	0.0601	0.0666	0.0578	0.0642	0.0620	0.0714
C	11.238	32.729	12.047	47.436	17.721	69.681
R ²	0.9987	0.9995	0.9947	0.9974	0.9976	0.9901
GAB						
w_0 (g water/g dry solids)	0.084	0.084	0.076	0.081	0.082	0.091
C	9.802	23.042	10.712	25.154	13.872	29.156
K	0.637	0.680	0.693	0.710	0.673	0.671
R ²	0.9686	0.9756	0.9406	0.9700	0.9572	0.9654

1 **Table 4:** Glass transition temperature (midpoint TG and ΔC_p) of BR and EBR samples
 2 equilibrated at different water activities (0.113-0.755).

Water activity	BR Samples			EBR Samples		
	Water content (g water /100 g sample)	Tg (°C)	ΔC_p (J/g.K)	Water content (g water /100 g sample)	Tg (°C)	ΔC_p (J/g.K)
0.113	4.06	58.05	0.098	6.15	59.65	0.130
0.230	5.89	56.86	0.131	7.71	57.82	0.126
0.330	7.66	60.94	0.154	9.40	58.16	0.203
0.430	9.44	62.15	0.165	11.24	59.32	0.166
0.520	10.65	59.77	0.180	12.14	59.74	0.199
0.680	12.57	60.01	0.210	14.06	59.70	0.263
0.755	14.95	59.16	0.182	17.20	59.13	0.412

3

4

Table 1. Composition, Free Fatty Acids (FFA) content and TBA reactive substances (TBARS) of Brown Rice (BR), Extruded Brown Rice (EBR), Soaked Rice (SR), Extruded Soaked Rice (ESR), Germinated Rice (GR) and Extruded Germinated Rice (EGR).

Samples	Crude Proteins (g/100 g d.b)	Lipids (ether extract) (g/100g d.b)	Total Fiber (g/100g d.b)	Ashes (g/100g d.b)	Total Starch (g/100 g d.b)	Available Carbohydrates* (g/100 g d.b)	FFA (mg Lauric acid/g)	TBA reactive substances (μ mol/g)
BR	6.97 \pm 0.15 ^a	1.88 \pm 0.01 ^a	5.5 \pm 0.6 ^a	1.11 \pm 0.01 ^a	77.74 \pm 1.13 ^b	84.54	255.27 \pm 12.24 ^a	2.77 \pm 0.46 ^d
EBR	6.63 \pm 0.20 ^a	0.46 \pm 0.19 ^c	3.9 \pm 0.3 ^b	0.99 \pm 0.01 ^a	76.94 \pm 0.30 ^b	86.60	36.89 \pm 2.30 ^c	7.83 \pm 0.20 ^c
SR	6.48 \pm 0.10 ^a	1.09 \pm 0.02 ^b	5.7 \pm 0.7 ^a	0.98 \pm 0.11 ^a	80.19 \pm 0.92 ^a	85.75	147.51 \pm 7.99 ^b	3.74 \pm 0.13 ^d
ESR	6.37 \pm 0.12 ^a	0.73 \pm 0.04 ^c	3.3 \pm 0.3 ^b	1.03 \pm 0.07 ^a	76.15 \pm 0.26 ^b	88.21	28.84 \pm 2.01 ^c	7.07 \pm 0.52 ^c
GR	6.65 \pm 0.10 ^a	1.63 \pm 0.20 ^a	5.7 \pm 0.6 ^a	1.20 \pm 0.30 ^a	79.37 \pm 0.31 ^a	84.82	135.77 \pm 4.06 ^b	9.89 \pm 0.32 ^b
EGR	6.70 \pm 0.40 ^a	0.47 \pm 0.15 ^c	3.2 \pm 0.4 ^b	1.23 \pm 0.06 ^a	73.53 \pm 0.77 ^c	87.24	31.65 \pm 2.00 ^c	13.11 \pm 0.1 ^a

Mean values \pm standard deviations (n = 3); Different letters in the same column mean significant differences between samples (p < 0.05)

*Available carbohydrate content was calculated by difference.

1 **Table 2.** Water absorption (WA), Water solubility (WS), angles and intensity (cps 10^{-3}) of
 2 crystalline peaks (2θ) and crystallinity value (%) of Brown Rice (BR), Extruded Brown Rice
 3 (EBR), Soaked Rice (SR), Extruded Soaked Rice (ESR), Germinated Rice (GR) and Extruded
 4 Germinated Rice (EGR).

Parameter/ Samples	WS (g sample/100 g sample)	WA (ml of water/ g dry sample)	Angles (2θ) and intensity of Peaks (cps 10^{-3})	Crystallinity Value (%)
BR	3.18±0.05 ^a	2.77±0.15 ^a	15.17° (3.18) 17.65° (9.88) 23.26° (17.13)	30±2 ^a
EBR	24.1±0.9 ^b	7.04±0.06 ^c	12.30° (14.38) 18.17° (2.11)	16±2 ^b
SR	3.17±0.03 ^a	2.90±0.13 ^a	15.06° (3.03) 17.70° (13.88) 23.17° (15.66)	33±2 ^c
ESR	53.76±2.0 ^d	5.81± 0.21 ^b	12.71° (3.10) 18.33° (0.84)	4±2 ^d
GR	4.98±0.19 ^a	3.05±0.20 ^a	15.01° (2.32) 17.73° (9.08) 23.17° (15.99)	30±2 ^a
EGR	42.83±0.20 ^c	7.47±0.11 ^d	12.81° (3.59) 18.29° (0.82)	4.4±1.5 ^d

5 Mean values ± standard deviations (n = 3).

6 Different letters in the same column mean significant differences between samples (p < 0.05)