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Additional Information

1 **Influence of air temperature on drying kinetics and antioxidant potential of**
2 **olive pomace**

3
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26 **ABSTRACT**

27

28 This work aims to evaluate the influence of olive pomace drying (a solid by-
29 product of the olive oil industry) on both antioxidant potential and drying
30 kinetics. The two main fractions of olive pomace (pits, PI and pulps+peels, P+P)
31 were characterized by image analysis and density measurement. The drying
32 process was analyzed in experiments carried out at different temperatures (from
33 50 to 150 °C) and mathematically described from the diffusion and Weibull
34 models. The antioxidant potential of the extracts (ethanol-water 80:20 v/v, 22 ±
35 1 °C, 170 rpm for 24 h) obtained from the dry product was analyzed by
36 measuring the total phenolic content and antioxidant capacity and the main
37 polyphenols were quantified by HPLC-DAD/MS-MS.

38 The drying behavior of olive pomace was well described by considering the
39 diffusion in the PI and P+P fractions separately and the influence of
40 temperature on effective moisture diffusivities was quantified by an Arrhenius
41 type equation. The antioxidant potential was only mildly influenced by the drying
42 temperature. However, long drying times at the highest temperature tested (150
43 °C) significantly ($p < 0.05$) increased the antioxidant potential.

44

45 *Key words:* phenolic content; antioxidant capacity; drying kinetics; diffusion;
46 olive pomace

47

48

49

50 **1. Introduction**

51

52 The olive (*Olea europea*) is an evergreen tree traditionally cultivated for
53 the production of oil and table olives. As regards both wealth and tradition, the
54 olive oil industry is a relevant one, especially in the Mediterranean countries
55 where 97 % of the world's olive production is harvested. Spain is the leading
56 country in terms of the total crop surface and the number of productive trees
57 (Niaounakis & Halvadakis, 2004).

58 Nowadays, the olive oil industry generates a great environmental impact
59 due to the production of high polluting residues (Baeta-Hall et al., 2005).
60 Several studies have stated the negative effects of these forms of waste on
61 soil's microbial populations (Paredes et al., 1987), aquatic ecosystems
62 (DellaGreca et al., 2001) and even on the air (Rana et al., 2003). However, olive
63 polyphenols, such as oleuropein, verbascoside or hydroxytyrosol, are present
64 not only in olive oil but also in oil waste products, exhibiting among other things,
65 antiviral, antitumoral and antioxidant activities (Della Ragione et al., 2000; Liu et
66 al., 2003; Micol et al., 2005). One of the most problematic olive oil waste
67 products is pomace (the solid byproduct made up from pieces of pit, skin and
68 pulp), also known as cake. Actually, it is used for animal feed, residual oil
69 extraction, energy recovery, soil amendment or the extraction of valuable
70 polyphenols (Roig et al., 2006). A previous dehydration stage reduces the
71 pomace water content to 5-6 % (wet basis), aiming to stabilize the byproduct
72 and so avoiding undesirable degradation during storage. Moreover, in the
73 particular case of bioactive compound extraction, drying avoids the interference
74 of water in the polyphenol release (Soysal & Öztekin, 2001), improving the

75 extraction yield. For industrial purposes, hot air drying is the most widely used
76 method, since it allows an accurate control of the process variables.
77 Traditionally, low air temperatures are used as a means of better protecting the
78 bioactive compounds from degradation during drying. However, drying at low
79 temperatures constitutes a slow process in which metabolic reactions may be
80 long lasting, leading to quality loss (Fennell et al., 2004). Thereby, certain
81 studies also suggest the use of high temperatures for the industrial drying of
82 olive pomace (Göğüs & Maskan, 2006). High temperatures speed up the drying
83 kinetics, which could be interesting for the purposes of increasing productivity
84 on an industrial scale (Ahmad-Qasem et al., 2013a), but at the same time it
85 could promote the oxidative degradation of polyphenols (Gomes & Caponio,
86 2001) and requires the use of a great amount of energy. For this reason, the
87 main aim of this work was to assess the influence of the air temperature on the
88 drying kinetics and antioxidant potential of olive pomace, two aspects which
89 have not previously been considered together.

90

91 **2. Materials and methods**

92

93 *2.1 Raw material*

94 The raw material used in this work was olive pomace from a traditional
95 pressing system for obtaining olive oil, provided by an oil factory located in
96 Altura (Castellón, Spain) The pomace was collected just after the pressing
97 operation and immediately vacuum packaged and stored at 4 °C. The initial
98 moisture content was determined by drying in a vacuum chamber at 70 °C until
99 reaching constant weight (AOAC method nº 934.06, AOAC, 1997).

100 It could be considered that olive pomace is mainly composed of two main
101 fractions: pits (PI) and pulps+peels (P+P). Homogeneous samples of olive
102 pomace were taken, both fractions were separated by hand and their
103 corresponding mass fraction (X) calculated and characterized by image analysis
104 (Table 1). RGB images were taken (Figures 1a and 1c) and processed using
105 Image J software (Research Service Branch, National Institute of Mental Health,
106 US, available as freeware from <http://rsbweb.nih.gov/ij/>). Images were
107 converted to the binary system (Figures 1b and 1d) using an automatic
108 threshold. Finally, the particles were counted and their surface (S, mm²)
109 calculated considering the scale reference. From another set of experiments,
110 the initial moisture content of both fractions was also determined, as already
111 explained for the olive pomace.

112 The bulk density (ρ) of both the PI and P+P fractions, as well as that of
113 the fresh olive pomace, was determined at 20 °C by liquid displacement using
114 water, a volumetric standard picnometer (48.89 mL) and an analytical balance
115 (PB 303-S, Mettler Toledo).

116

117 *2.2 Drying experiments*

118 Drying experiments were conducted in a forced air laboratory drier (FD,
119 Binder, Tuttlingen, Germany), using a horizontal air flow of 0.094 m³/s and an
120 air velocity of 0.683 m/s. Each run was carried out with an initial mass load of
121 40 g of olive pomace, uniformly distributed in a monolayer (4 ± 1 mm thick,
122 0.083 g/cm²).

123 Two different sets of experiments were designed. In the first one, the
124 variable to be considered was that of the air temperature in order to determine
125 its influence on both the drying kinetics and antioxidant potential of the extracts
126 obtained from the dried product. For this purpose, the drying experiments were
127 carried out at different air drying temperatures: 50, 70, 90, 120 and 150 °C.
128 During the process, the samples were weighed (XS204, Mettler Toledo,
129 Barcelona, Spain) at pre-set times. The drying experiments finalized when the
130 sample weight loss reached 30 ± 1 %. This fact was established by previous
131 experiments, ensuring that the water activity was below 0.4 and the obtained
132 product was stable.

133 In the second set of experiments, the variable to be studied was the
134 drying time. For that purpose, drying experiments were carried out at 150 °C
135 and for different drying times: 5, 10, 20, 30 and 60 min. It should be highlighted
136 that, in this set of experiments the effective drying period took place between 5
137 and 10 min, from which mass transfer could be considered negligible.
138 Therefore, this involves overexposing the olive pomace to a high temperature
139 (150 °C).

140 The drying experiments for each experimental condition tested were
141 carried out three times, at least.

142

143 *2.3 Modeling of hot air drying kinetics*

144 The experimental drying kinetics were determined from the initial sample
145 mass and the weight loss measured during drying. Previous approaches to the
146 modeling of the drying kinetics of olive pomace have been addressed through
147 the use of deep beds, assuming in the modeling that the sample is as thick as

148 the bed is high and that it behaves like an infinite slab (Göğüs & Maskan, 2006).
149 In addition, Vega-Gálvez et al. (2010) molded olive cake into a rectangular form
150 and conducted drying experiments in monolayer at different temperatures in
151 order to identify an effective moisture diffusivity in this particular body. However,
152 the drying of the individual particles of olive pomace has not previously been
153 addressed. This could be considered a complicated issue, since olive pomace
154 is a heterogeneous material made up mainly of pits, peels and pulp pieces,
155 which represents a handicap when using a diffusion model where the samples
156 are assumed to be homogeneous. In this work, therefore, the monolayer drying
157 of this byproduct has been studied in order to identify the drying behavior of
158 olive pomace at particle level, and further studies should be performed to
159 address the drying of the bulk of the olive pomace. For that purpose, two
160 different approaches were considered.

161 On the one hand, a diffusion model for the olive pomace was used by
162 considering the diffusion in both fractions of the olive pomace to be different:
163 Pits (PI) and pulps+peels (P+P). It was assumed that pits could be considered
164 as geometrically spherical particles, while peels+pulps could behave like infinite
165 slabs. Eqs. 1 and 2 show the solution of diffusion models for spheres and
166 infinite slabs, respectively, considering:

- 167 - Homogenous and isotropic solids.
- 168 - Constant effective diffusivity.
- 169 - Negligible shrinkage.
- 170 - Uniform initial moisture and temperature.
- 171 - The solid surface at equilibrium with the drying air.
- 172 - Solid symmetry.

173
$$W_{PI} = W_e + (W_c - W_e) \left[\sum_{n=1}^{\infty} \frac{6}{n^2 \pi^2} \exp\left(-\frac{D_e^{PI}}{R^2} n^2 \pi^2 t\right) \right] \quad (1)$$

174
$$W_{P+P}(t) = W_e + (W_c - W_e) \left[\sum_{n=0}^{\infty} \frac{8}{(2n+1)^2 \pi^2} \exp\left(-\frac{D_e^{P+P} (2n+1)^2 \pi^2 t}{4L^2}\right) \right] \quad (2)$$

175 where W is the average moisture content (dry basis), subscripts c and e refer to
 176 the critical and equilibrium states, t (min) the drying time, D_e is the effective
 177 moisture diffusivity (m^2/s), which was considered to be different in both PI and
 178 P+P fractions. The characteristic diffusion paths, radius (R) and thickness (L),
 179 were experimentally determined. The average radius of pit pieces was obtained
 180 from image analysis. For that purpose, the radius of individual particles, which
 181 was calculated from the measurement of the surface ($S_p = \pi R^2$), was computed
 182 and weighed to obtain an average value. Whereas the average particle
 183 thickness of the pulp+peel fraction was calculated from the measurement of the
 184 total particles surface (S_t), also obtained by image analysis, the mass (M) and
 185 the density (ρ) (Eq. 3):

186
$$L = \frac{M}{\rho S_t} \quad (3)$$

187 Considering the diffusion in PI and P+P fractions to be different, the
 188 moisture content of olive pomace could be calculated from Eqs. 1 and 2 by
 189 using a compositional model (Eq. 4). Similar diffusion models have been used
 190 for analyzing mass transfer phenomena in geometrically complex plant tissues,
 191 such as grape stalk (García-Pérez et al., 2006) and broccoli (Sanjuán et al.,
 192 2001).

193
$$W = X_{PI} W_{PI} + X_{P+P} W_{P+P} \quad (4)$$

194 where X is the mass fraction of PI and P+P fractions ($X_{PI} + X_{P+P} = 1$).

195 On the other hand, the Weibull empirical model (Cunha et al., 1998;
196 Simal et al., 2005) was also used for the mathematical description of the drying
197 kinetics of olive pomace. The Weibull model (Eq. 5) is a probability function
198 used to explain the behavior of changeable complex systems (Cunha et al.,
199 1998). Initially, it was used to predict material failures caused by fatigue. In food
200 technology, it has been used for the description of degrading processes, since
201 the degradation of food can be considered as a system fault under certain
202 stress conditions (Blasco, 2003), such as when exposed to hot air. Thus, the
203 Weibull model adapted to a drying process is presented in Eq. 5:

$$204 \quad \Psi(t) = \frac{W(t) - W_e}{W_c - W_e} = \exp\left(-\left(\frac{t}{\beta}\right)^\alpha\right) \quad (5)$$

205 where Ψ represents the dimensionless moisture content, α the dimensionless
206 parameter related to the shape, assimilated to the behavior index of the product
207 during drying, and β (min^{-1}) is the kinetic parameter inversely related ($1/\beta$) with
208 the process rate.

209 The identification of the model parameters (α and β in the Weibull model,
210 and D_e for PI and P+P fractions in the diffusion model) was carried out by
211 minimizing the sum of the squared differences between the experimental and
212 calculated moisture content of olive pomace samples using the Solver tool from
213 Excel™ (Microsoft Corporation, Seattle, USA). For each drying condition tested,
214 the parameter identification was simultaneously carried out with all the
215 replicates. The explained variance (VAR) was computed (Eq. 6) to determine
216 the goodness of the model's fit to experimental data:

217
$$\text{VAR} = 1 - \frac{S_{xy}^2}{S_y^2} \quad (6)$$

218 where S_{xy}^2 is the variance of the estimation and S_y^2 the variance of the sample.
 219 In addition, the mean relative error (MRE) was calculated (Eq. 7) to establish
 220 the difference between the experimental (W_{EXPi}) and calculated (W_{CALi}) data:

221
$$\text{MRE} = \frac{100}{N} \left(\sum_{i=1}^N \frac{W_{EXPi} - W_{CALi}}{W_{EXPi}} \right) \quad (7)$$

222 where N is the number of experimental data.

223 Moreover, in order to evaluate the influence of temperature on the kinetic
 224 parameters, an Arrhenius type equation (Meziane, 2011) was used (Eqs. 8 and
 225 9):

226
$$\frac{1}{\beta} = \frac{1}{\beta_0} \cdot \exp\left(-\frac{E_a}{R \cdot T}\right) \quad (8)$$

227
$$D_e = D_0 \cdot \exp\left(-\frac{E_a}{R \cdot T}\right) \quad (9)$$

228
 229 where $1/\beta_0$ (min^{-1}) and D_0 (m^2/s) are the pre-exponential factors, E_a (kJ/mol) the
 230 activation energy, R (kJ/mol·K) the universal gas constant and T (K) the drying
 231 temperature.

232

233 *2.4 Extraction experiments*

234 Dried olive pomace was milled (Blixer 2, Robot Coupe USA, Inc.,
 235 Jackson, MS, USA) and sieved (Metallic mesh 0.05 mm, Filtra Vibración,
 236 Barcelona, Spain) to obtain particles with a diameter of under 0.05 ± 0.01 mm.
 237 The extraction was carried out in sealed containers protected from light and
 238 immersed in a thermostatic shaking water bath (SBS40, Stuart, Staffordshire,

239 UK) working at 170 rpm. The solvent used was a solution of ethanol-water
240 (80:20, v/v) and the ratio between the weight of the olive pomace and the
241 solvent volume, 20 g/30 mL. The extraction process was carried out at 22 ± 1
242 °C for 24 h, which was based on previous works (Ahmad-Qasem et al., 2013a
243 and 2013b). In addition, another set of extraction experiments was carried out
244 by varying the extraction times from 5 to 48 hours to monitor the extraction
245 process, which is relevant in the case of industrial operations where a high level
246 of productivity needs to be attained. In this case, extraction experiments were
247 conducted with fresh and dried olive pomace at 50 and 150 °C.

248 Every extract was centrifuged for 10 min at 5000 rpm (Medifriger BL-S,
249 J.P. Selecta, Barcelona, Spain), filtered (nylon filters of 0.45 µm) and stored in
250 opaque vials at 4 °C until analysis. At least 3 replicates were made for each
251 different condition tested.

252

253 *2.5 Total phenolic content (TPC) and antioxidant capacity (AC) measurement*

254 The phenolic content and antioxidant capacity of the extracts obtained
255 from the dried olive pomace were determined by the Folin-Ciocalteu (Singleton
256 et al., 1999) and Ferric-Reducing Ability Power (FRAP) methods, respectively
257 (Benzie & Strain, 1996). These methods are exhaustively described by Ahmad-
258 Qasem et al. (2013a and 2013b).

259 The TPC was expressed as mg of gallic acid (GAE) per g of dry weight of
260 olive pomace (g d.w.), while the AC was expressed as mg of Trolox per g of dry
261 weight of olive pomace (g d.w.).

262

263 *2.6 Identification and quantification of polyphenols by HPLC-DAD/MS-MS*

264 In order to identify and quantify the main polyphenols, the olive pomace
265 extracts were analyzed using an HPLC instrument (Agilent LC 1100 series;
266 Agilent Technologies, Inc., Palo Alto, CA, USA) controlled by the Chemstation
267 software. The HPLC instrument was coupled to an Esquire 3000+ (Bruker
268 Daltonics, GmbH, Germany) mass spectrometer equipped with an ESI source
269 and ion-trap mass analyzer, and controlled by Esquire control and data analysis
270 software. A Merck Lichrospher 100RP-18 (5 μ m, 250 x 4 mm) column was used
271 for analytical purposes.

272 Separation was carried out through a linear gradient method using 2.5 %
273 acetic acid (A) and acetonitrile (B), starting the sequence with 10 % B and
274 programming the gradient to obtain 20 % B at 10 min, 40 % B at 35 min, 100 %
275 B at 40 min, 100 % B at 45 min, 10 % B at 46 min and 10 % B at 50 min. For
276 the LC-MS pump to perform accurately, 10 % of organic solvent was pre-mixed
277 in the water phase. The flow-rate was 1 mL/min and the chromatograms
278 monitored at 240, 280 and 330 nm. Mass spectrometry operating conditions
279 were optimized in order to achieve maximum sensitivity values. The ESI source
280 was operated in negative mode to generate $[M-H]^-$ ions under the following
281 conditions: desolvation temperature at 365 °C and vaporizer temperature at 400
282 °C; dry gas (nitrogen) and nebulizer were set at 12 L/min and 4.83 bar,
283 respectively. The MS data were acquired as full scan mass spectra at 50–1100
284 m/z by using 200 ms for the collection of the ions in the trap.

285 The olive pomace compounds were identified by HPLC-DAD analysis,
286 comparing the retention time, the UV spectra and the MS/MS data of the peaks.
287 Only the luteoilin-7-O-glucoside was quantified using a commercial standard
288 (Phytolab, Vestenbergsgreuth, Germany). The quantitative evaluation was

289 performed with a calibration curve using methanol solutions of known
290 concentrations. The polyphenol concentration was expressed as mg luteolin-7-
291 O-glucoside per g of dry weight of olive pomace (g d.w.).

292

293 **3. Results and discussion**

294

295 *3.1 Hot air drying kinetics at different temperatures*

296 In order to evaluate the influence of air temperature on drying kinetics,
297 experiments were carried out at temperatures ranging from 50 to 150 °C. In
298 these experiments, the initial moisture content of olive pomace was reduced
299 from 0.33 to 0.05 kg water/kg fresh olive pomace (30 % of the initial weight
300 loss). Despite the fact that olive pomace is a heterogeneous material, an
301 adequate repeatability and a small experimental variability was found in the
302 experimental drying kinetics (Figure 2).

303 As can be observed in Figure 2, the air temperature significantly affected
304 ($p < 0.05$) the drying rate: the higher the temperature, the shorter the processing
305 time. Thus, the drying times needed to achieve a 30 % loss of the initial weight
306 ranged from 40 min at 50 °C to 10 min at 150 °C and increasing the drying
307 temperature from 90 to 150 °C shortened the drying time by 50 %. These
308 results agreed with those reported by other authors who studied olive pomace
309 drying by means of different techniques. Thus, Ruiz-Celma et al. (2008), who
310 studied the infrared drying of wet olive husk (pomace), found that a rise in
311 temperature from 80 to 140 °C reduced the drying time by a third.

312 The experimental data showed that, for the air temperatures tested, the
313 drying only took place during the falling rate period. As an example, the drying

314 rate for two particular temperatures tested (50 and 120 °C) is shown in Figure 3.
315 Therefore, the initial moisture content was assumed to be equal to the critical
316 one. These results agreed with previous works, like those published by Gögüs
317 & Maskan (2006) who studied olive pomace behavior during hot air drying at
318 temperatures ranging between 60 and 80 °C or Ruiz-Celma et al. (2008)
319 working on infrared drying at temperatures from 80 to 140 °C. However, Kadi &
320 Hamlat (2002) found a constant drying period during the hot air drying of olive
321 pomace. These contradictory results could be linked to the sample layer
322 thickness used in each experimental design. Thus, the constant rate period
323 cited by Kadi & Hamlat (2002) could be attributed to a thick sample layer, which
324 leads to air saturation, but not to the behavior of the particle.

325 Modeling of drying kinetics was addressed from the diffusion and Weibull
326 (Cunha et al., 1998) models, as explained in section 2.3. The modeling not
327 only aimed to quantify the influence of the temperature on the drying kinetics
328 but also to better characterize the olive pomace drying at particle level. The
329 characterization of PI and P+P fractions is shown in Table 1, while the results of
330 drying kinetics modeling are included in Table 2.

331 A much higher effective moisture diffusivity (D_e) was found in the PI
332 fraction than in the P+P one (Table 2). Thus, D_e ranged from $1.17 \cdot 10^{-7}$ to
333 $2.92 \cdot 10^{-7}$ m²/s for the PI fraction and between $3.58 \cdot 10^{-11}$ and $1.60 \cdot 10^{-10}$ m²/s for
334 the P+P fraction. The temperature was found to have a significant influence on
335 D_e values: the higher the temperature, the higher the D_e . The fact that the D_e
336 figures found in the PI fraction are higher suggests that its structure has a low
337 water retention capacity, which leads to a higher water removal rate than in the
338 P+P fraction. This fact could be explained by taking the lower density of the PI

339 fraction into account (Table 1) while the low D_e values of the P+P fraction could
340 be ascribed to the water proof capacity of peels, which constitutes a natural
341 protection of olive fruit from dehydration . To our knowledge, there are no
342 references in the literature to the D_e in the PI and P+P fractions since this issue
343 has not been previously addressed. However, the values identified in this work
344 for both fractions are similar to others reported in literature for olive pomace
345 drying in deep beds, thin layers or regular-shaped bodies. Thus, Meziane
346 (2011) in working on fluidized bed drying (thickness 41-33 mm) at 50-80 °C,
347 reported D_e figures from $0.68 \cdot 10^{-7}$ to $2.15 \cdot 10^{-7}$ m²/s, which were similar to those
348 reported by Göğüs & Maskan (2006) for tray drying (thickness 6-12 mm) at 60-
349 80 °C. However, lower D_e figures have also been reported. Thus, in the case of
350 tray drying (thickness 4-12 mm) at 80-110 °C, Doymaz et al. (2006) found
351 values ranging from $4.89 \cdot 10^{-10}$ to $9.89 \cdot 10^{-10}$ m²/s while Montero et al. (2011)
352 identified values ranging from $9.1 \cdot 10^{-11}$ m²/s to $1.4 \cdot 10^{-10}$ m²/s for solar drying at
353 20-50 °C (thickness 20-40 mm). The great differences found in the literature for
354 the D_e figures of olive pomace could be ascribed not only to the effect of the
355 thickness of the sample being dried but also to the highly heterogeneous nature
356 of this product.

357 The diffusion model proposed in this work fitted the experimental data
358 closely, providing similar explained variance (VAR) and mean relative errors
359 (MRE) to the Weibull model (Table 2). In overall terms, the VAR and MRE were
360 close to 99 % and 10 %, respectively. The only significant difference between
361 both models was found at 150 °C (Figure 4), at which temperature the Weibull
362 model fitted the drying kinetic much better than the diffusion (Table 2); the VAR
363 was found to fall from 99.7 to 96.9 %. This could suggest that, at high

364 temperatures, diffusion was less important and other significant mass transport
365 phenomena appeared. In this sense, it was found that the α Weibull parameter,
366 related to product behavior, was not significantly ($p<0.05$) affected by
367 temperature in the range of 50 to 120 °C, reaching an average value of $0.87 \pm$
368 0.02. However, the value identified at 150 °C (0.99) was significantly ($p<0.05$)
369 higher. Therefore, although it could be stated that the behavior of olive pomace
370 remained stable during drying over the temperature range of 50 to 120 °C,
371 water removal phenomena seemed to change at 150 °C.

372 The drying temperature also affected the identified kinetic parameter (β)
373 of the Weibull model. Thus, $1/\beta$ increased when the air drying temperature rose,
374 showing that, over the range studied, the higher the temperature applied, the
375 faster the drying. The influence of temperature on D_e of the PI and P+P
376 fractions and the $1/\beta$ parameter was well described from an Arrhenius-type
377 relationship (Figure 5) over the range of 50 to 120 °C. In every case, the value
378 identified at 150 °C departed from the trend observed at the other temperatures,
379 as can be seen in Figure 5. So, excluding the kinetic data at 150 °C, the
380 identified activation energies (E_a) were 20.3 kJ/mol for olive pomace from the
381 $1/\beta$ Weibull parameter, 21.9 kJ/mol for the P+P fraction and 14.6 kJ/mol for the
382 PI fraction. The E_a figure reported for olive pomace was in the same order of
383 magnitude as that obtained by other authors. Thus, Meziane (2011) reported a
384 value of 36.8 kJ/mol (50-80 °C), Göğüs & Maskan (2006) 25.7 kJ/mol (60-80 °C)
385 and Doymaz et al. (2006) 26.71 kJ/mol (80-110 °C).

386

387 *3.2 Antioxidant potential affected by drying temperature*

388 Olive pomace is susceptible to spoilage due to the fact that it presents a
389 high level of enzymatic and microbial activity. This must be considered when it
390 is used as a potential source of bioactive compounds. Drying stabilizes the raw
391 material during storage and limits some degradative reactions but, in a certain
392 way, it can influence the bioactive potential of olive pomace. Hence, the total
393 phenolic content (TPC) and the antioxidant capacity (AC) were assessed in the
394 extracts obtained from olive pomace dried at temperatures ranging between 50
395 and 150 °C and were compared with those obtained from fresh pomace.

396 As can be observed in Figure 6, once the drying temperature exceeded
397 70 °C, it was noticeable that there was a slight tendency of the TPC to increase
398 as the temperature rose. This could be attributed to the formation of new
399 phenolic compounds at high temperatures (90-150 °C), due to the fact that non-
400 enzymatic interconversion leads to the availability of precursors of phenolic
401 molecules (Que et al., 2008). However, the statistical analysis highlighted the
402 fact that the influence of temperature on TPC was not significant ($p < 0.05$).
403 Moreover, the dried material exhibited a similar TPC to that shown by fresh
404 pomace. Different results have been found in literature when using similar
405 biomaterials. Thus, Khanal et al. (2010) reported that drying temperatures over
406 60 °C had a negative effect on the phenolic content of grape and blueberry
407 pomace.

408 As regards the AC of extracts, the drying temperature of pomace had a
409 significant ($p < 0.05$) effect on the antioxidant potential. Olive pomace dried at
410 150 °C provided the extracts with the highest AC (Figure 6), it being 15.5 %
411 higher than the one obtained from pomace dried at 50 °C. Furthermore,
412 compared with fresh pomace, drying at 150 °C increased the AC of extracts by

413 12.8 %. Samples dried over the range of 50 to 120 °C did not exhibit significant
414 ($p < 0.05$) differences compared with the fresh product. Studying the effect of the
415 drying air temperature on the antioxidant capacity of polyphenolic compounds in
416 mulberry leaves, Katsube et al. (2009) also observed an increase in the
417 antioxidant capacity when the air temperature rose from 70 to 110 °C.

418 The effect of the drying conditions on the antioxidant properties of
419 different byproducts and materials has been evaluated in several research
420 studies. In overall terms, it can be stated that there is great controversy over the
421 most suitable drying conditions. Thus, the use of mild drying temperatures (60
422 °C) and intermediate drying times is reported as the most suitable for orange
423 peel (Garau et al., 2007) or mulberry (Katsube et al., 2009). On the contrary,
424 Harbourne et al. (2009) found that, over the range of 30-70 °C, the drying air
425 temperature did not influence the phenolic constituents of meadowsweet and
426 willow. Other authors state that the use of high temperatures (90 °C) allows
427 extracts to be obtained with a high antioxidant potential (Vega-Gálvez et al.,
428 2009). These different conclusions concerning the effect of the drying
429 temperature on bioactive properties could probably be ascribed to the different
430 nature of the raw material processed.

431 In the case of the present study, the highest drying temperature tested,
432 150 °C, seemed to be the most suitable drying conditions under which to obtain
433 the highest AC of the extracts. It should be remarked that it is not only the
434 temperature but also the length of exposure to heat which can influence the
435 extract properties (Erbay & Icier, 2009), since the short treatments at high
436 temperatures may promote the presence of bioactive compounds in the extracts
437 (García-Pérez et al., 2010). However, since in a certain way drying also

438 involves thermal treatment, the impact of the heating time during drying at high
439 temperatures was further studied and the results are presented in the following
440 section.

441

442 *3.3 Antioxidant potential affected by drying time at high temperatures*

443 A new set of drying experiments at 150 °C was carried out varying the
444 drying time of olive pomace from 5 to 60 min. It should be noticed that after
445 approximately 8 min of drying, see section 3.1, samples could be considered
446 dried (water activity less than 0.4). Thus, longer processing times are
447 unnecessary for water removal and represent an additional overheating due to
448 the product being overexposed to high air temperatures. Once processing
449 finalized, the TPC and AC of the extracts obtained from these dried samples
450 were measured.

451 Although phenolic compounds are considered as heat sensitive
452 antioxidants (Erbay & Icier, 2009), the results showed that there was a
453 significant ($p < 0.05$) increase in the TPC as the drying time lengthened . As can
454 be observed in Figure 7, at a drying time of over 10 min the samples exhibited a
455 significantly ($p < 0.05$) higher TPC than the fresh material, and the longer the
456 drying time, the higher the TPC of the extracts. In such a way, the highest TPC
457 was obtained after a drying time of 30 min, representing a 39.9 % increase
458 compared to what was observed in the fresh olive pomace extracts. The
459 increase in the drying time, from 30 to 60 min, did not significantly ($p < 0.05$)
460 affect the TPC. Thereby, it is not advisable to heat olive pomace longer than 30
461 min, since this would reduce the productivity and increase both the processing
462 costs and the energy consumption. As far as this aspect is concerned, the

463 literature throws up contradictory results. It is widely recognized that
464 polyphenols are heat labile, thus, it is reported that heat treatments cause
465 irreversible chemical changes (Mejía-Meza et al., 2008). In this way, Kyi et al.
466 (2005) highlighted the fact that, when cocoa beans were dried at temperatures
467 over the 40-60 °C range, the concentration of total polyphenols declined
468 drastically when the drying time was extended, additionally observing that the
469 higher the temperature, the lower the residual amount of polyphenols. On the
470 contrary, the positive influence that heating has on the antioxidant capacity has
471 also been observed in microwave treatments. Hence, Hayat et al. (2010) stated
472 that, in mandarin pomace, the sum of the content of the individual phenolic
473 acids in the free fraction significantly increased as the drying time lengthened.

474 As regards the AC measurements, overheating the olive pomace also
475 significantly ($p < 0.05$) increased the AC of extracts (Figure 7). For drying times
476 longer than 5 min, the extracts exhibited a significantly ($p < 0.05$) higher AC than
477 the extracts obtained from fresh olive pomace; from 5 min of drying onwards,
478 the longer the overheating, the higher the AC. However, it is important to
479 highlight that, as in TPC measurement, no significant ($p < 0.05$) differences were
480 found between extracts obtained from pomace treated for 30 and 60 min.
481 Therefore, drying at 150 °C for 30 min seemed to be the best processing
482 conditions under which to obtain the highest AC. It should be remarked upon
483 that, under these conditions (150 °C and 30 min) and compared with the fresh
484 product, AC increased almost twice as much (78 %) as TPC (40 %).

485 Vashisth et al. (2011) observed that the drying time had no influence on
486 the antioxidant capacity of muscadine pomace at 70 and 80 °C. Considering the
487 fact that long drying times at low air temperatures (30-40 °C) promote a

488 decrease in the antioxidant capacity (Garau et al., 2007) and in view of the
489 results obtained in this work, it could be reasonable to consider that there is a
490 temperature threshold from which point onwards the drying time increases the
491 content of antioxidant compounds. This behavior could be explained by
492 considering that high temperatures promote the inactivation of oxidative
493 enzymes (Sanjuán et al., 2000), avoiding the degradation of antioxidants for
494 later processing, which includes the extraction stage. Furthermore, at high
495 temperatures, the generation and accumulation of Maillard-derived melanoidins
496 with a varying degree of antioxidant activity could also enhance the antioxidant
497 properties of extracts (Que et al., 2008).

498 In order to clarify the effect the overheating had on the increase in
499 antioxidant phenolic compounds, the composition of the extracts was analyzed
500 by HPLC-DAD/MS-MS. The HPLC-DAD profile of the samples was quite
501 complex and a large variety of peaks were detected at UV wavelengths.
502 Nevertheless, the fact that ionization occurred in only a few of them was
503 probably due to the presence of organic polymers in pomace samples, which
504 complicated the identification of the polyphenolic profile of the samples. Among
505 the phenolic compounds identified, minor quantities of secoiridoids, such as
506 oleuropein and ligstroside, were detected. The main polyphenol to be identified
507 and quantified was luteolin, which was selected as a marker to be quantified in
508 the different extracts obtained from samples subjected to different drying times.
509 The drying treatment at 150 °C led to an increase in the luteolin concentration
510 as compared to fresh material (Figure 8); additionally, the longer the drying
511 time, the higher the luteolin content. Thus, as shown in Figure 8, increasing the
512 drying time from 10 to 60 min led to a rise in the luteolin content of

513 approximately 100 %. Therefore, these results highlighted the relationship
514 between the previously observed enhancement of antioxidant potential and the
515 increase in the content of some individual polyphenols, such as the flavone
516 luteolin.

517

518 *3.4 Monitoring of the extraction process*

519 In experiments carried out to assess how the drying temperature of olive
520 pomace or the drying time at high temperatures affected the antioxidant
521 potential of extracts, an extraction time of 24 h was considered enough to reach
522 equilibrium conditions according to previous studies (Ahmad-Qasem et al.,
523 2013a and 2013b). In order to test the feasibility of using shorter extraction
524 times as a means of improving productivity, which could be relevant for
525 industrial purposes, another set of experiments was performed using the fresh
526 and dried pomace at the lowest (50 °C) and the highest (150 °C) temperatures
527 tested. Thus, separate extraction experiments were conducted, varying the
528 extraction time from 5 to 48 hours, and replicated at least three times.

529 As observed in Figure 9, most of the phenolic compounds were extracted
530 from the solid matrix during the first 5 h of contact with the solvent. Afterwards,
531 some slight variation of TPC was found, which was especially noticeable in the
532 case of fresh material and olive pomace dried at 50 °C. Thus, increasing the
533 extraction time from 5 to 48 h led to an observed rise in the TPC of fresh and
534 dried pomace of 23.4 and 24.8 % at 50 °C, respectively. However, for the
535 material dried at 150 °C, the difference in the TPC at these extraction times was
536 almost negligible. This fact highlights another noticeable benefit of drying at
537 high temperatures (150 °C), which is the degradation of the raw structure which

538 promotes a sharp rise in the TPC of the solvent. From experimental results, it is
539 also evident that 24 h is a reasonable extraction time after which to evaluate the
540 TPC of olive pomace, since equilibrium is reached. Nevertheless, if further
541 industrial applications are considered, it seems reasonable to choose a short
542 extraction time, like 5 h, in order to increase productivity, thereby generating
543 large volumes of extracts with a high TPC after short treatment times.

544

545 **4. Conclusions**

546 A compositional diffusion model considering a different effective
547 diffusivity in pit and pulp+peel fractions provided a good description of the
548 drying behavior of olive pomace. Effective diffusivity for the pit fraction was
549 higher than in that of the pulps+peels and increased as the air temperature
550 rose. Although the influence of the drying temperature on the antioxidant
551 potential was only mild, long drying times at the highest temperature tested (150
552 °C) significantly increased the antioxidant potential.

553 Further studies should analyze the deep bed drying of olive pomace in
554 order to validate the developed model and confirm the effect of temperature on
555 the antioxidant potential of olive pomace.

556

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564

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714

Figure captions

715

716

717 Figure 1. RGB images of pulp+peel (P+P, a) and pit (PI, c) fractions of olive
718 pomace and binary conversion (P+P, b and PI, d) using Image J.

719 Figure 2. Experimental (average \pm standard deviation) drying kinetics of olive
720 pomace at different temperatures.

721 Figure 3. Evolution of drying rate (average \pm standard deviation) during drying
722 of olive pomace at 50 and 120 °C.

723 Figure 4. Experimental and calculated dimensionless moisture content using
724 Weibull and diffusion models. Experiments carried out at 50 (a) and 150 °C (b).

725 Figure 5. Fit of an Arrhenius type equation to the kinetic parameter of the
726 Weibull model ($1/\beta$, a) and effective moisture diffusivities for pit (D_e^{PI} , b) and
727 pulp+peel (D_e^{P+P} , c) fractions.

728 Figure 6. Experimental TPC and AC (average \pm standard deviation) of extracts
729 of olive pomace dried at different temperatures. Superscript letters show
730 homogeneous groups established from Least Significance Difference (LSD)
731 intervals ($p < 0.05$).

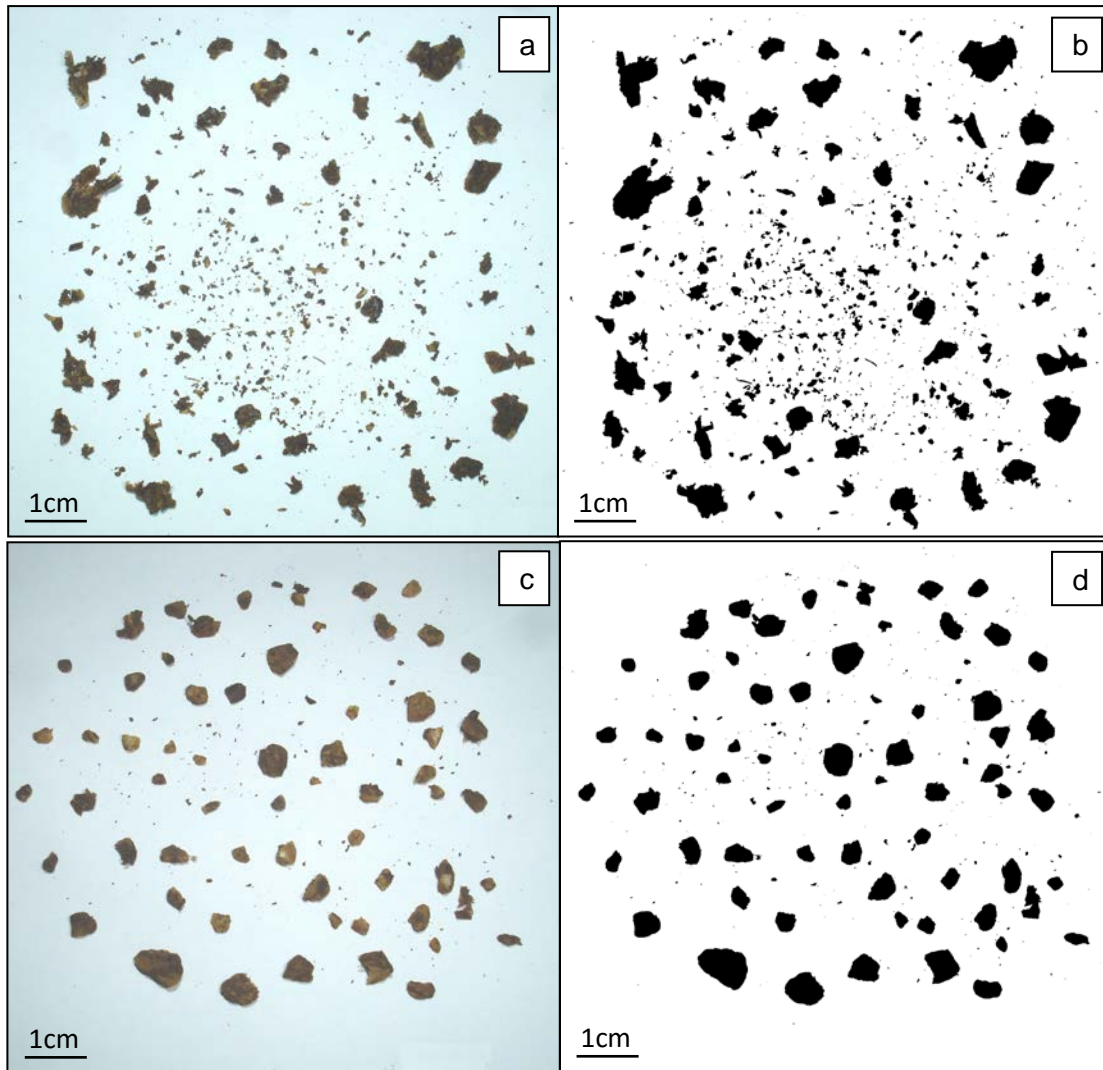
732 Figure 7. Experimental TPC and AC (average \pm standard deviation) of extracts
733 of olive pomace dried at 150 °C for different times. Superscript letters show
734 homogeneous groups established from Least Significance Difference (LSD)
735 intervals ($p < 0.05$).

736 Figure 8. Luteolin-7-O-glucoside content (average \pm standard deviation) of
737 extracts of olive pomace dried at 150 °C for different times. Superscript letters
738 show homogeneous groups established from Least Significance Difference
739 (LSD) intervals ($p < 0.05$).

740 Figure 9. TPC of extracts obtained from fresh and dried olive pomace (at 50 and
741 150 °C) at different extraction times. Average \pm standard deviation.

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743

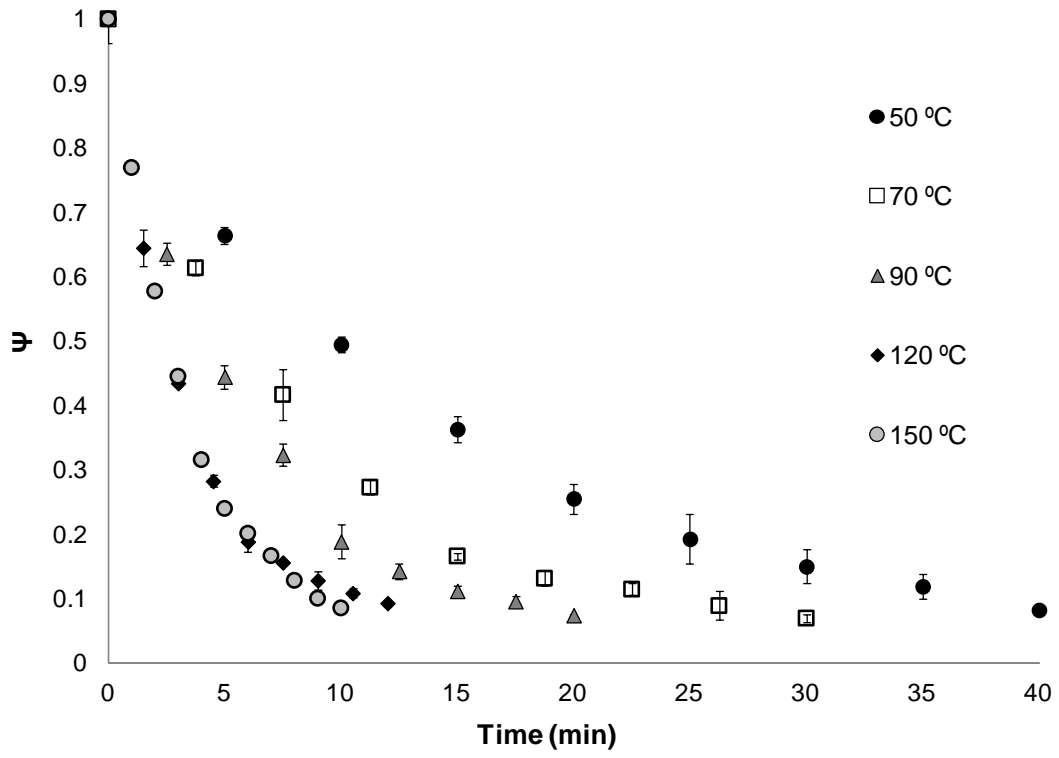


744

745 Figure 1

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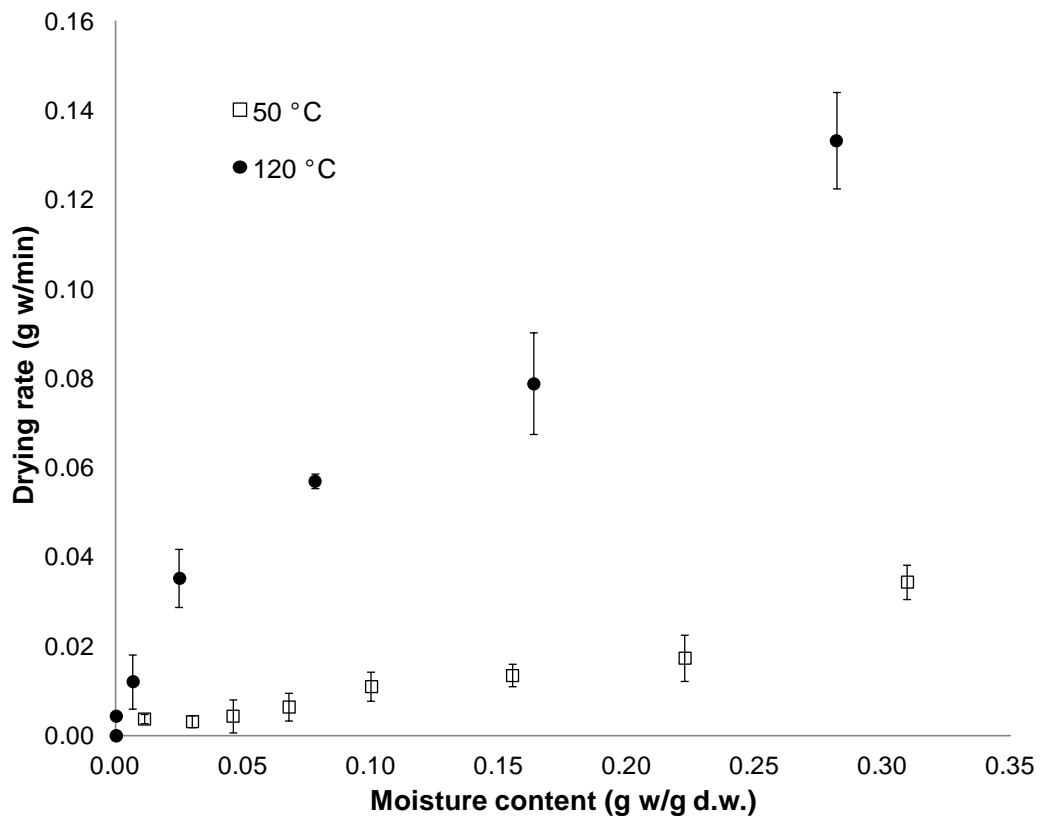
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749 Figure 2

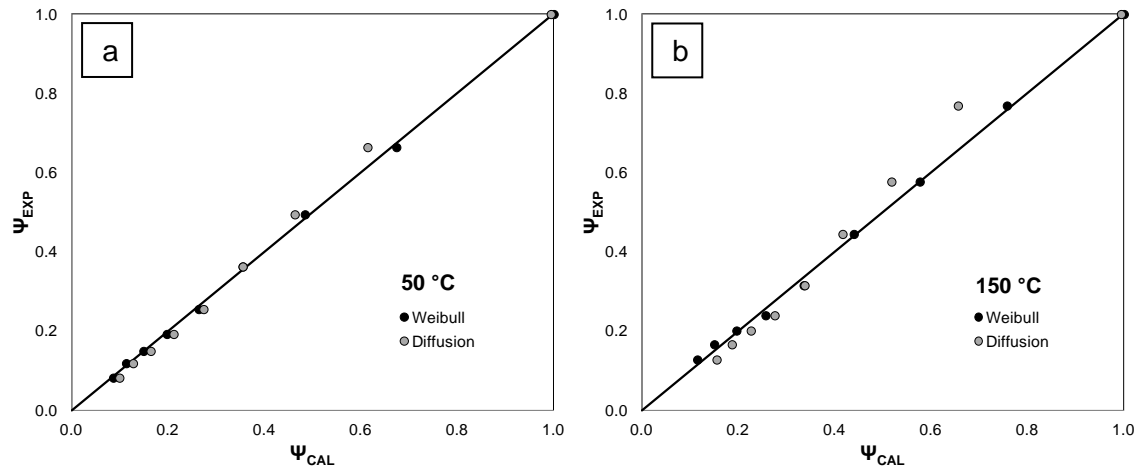
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752 Figure 3

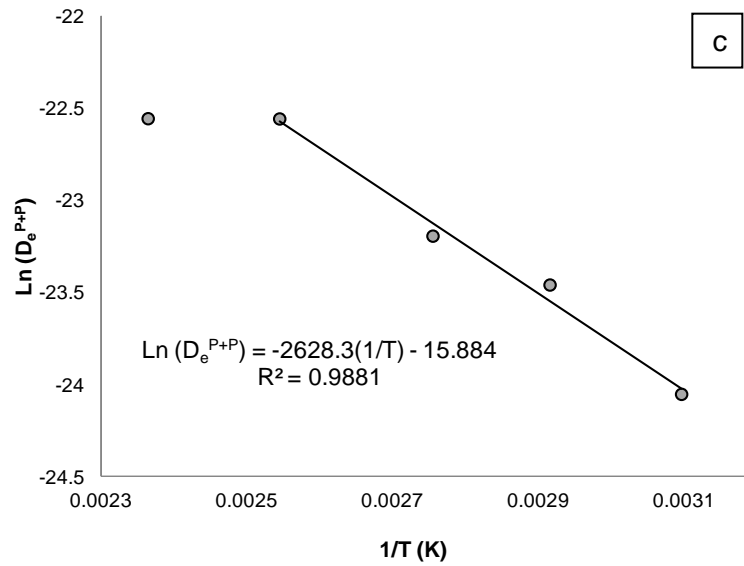
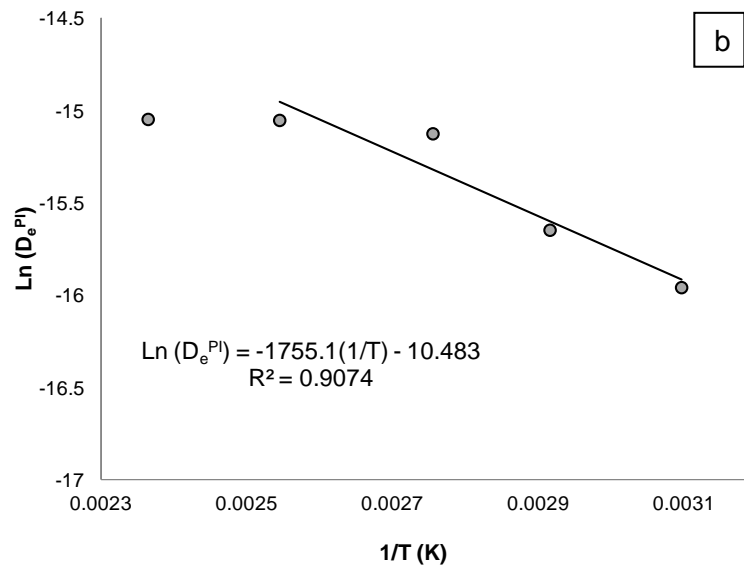
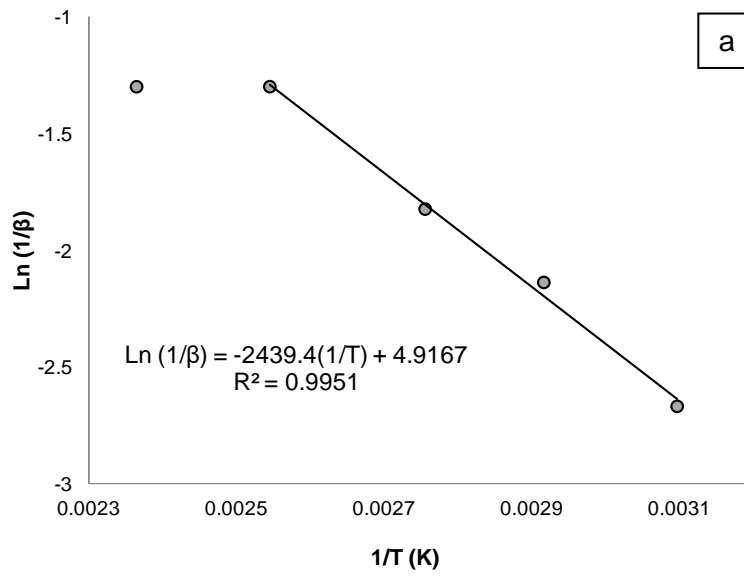
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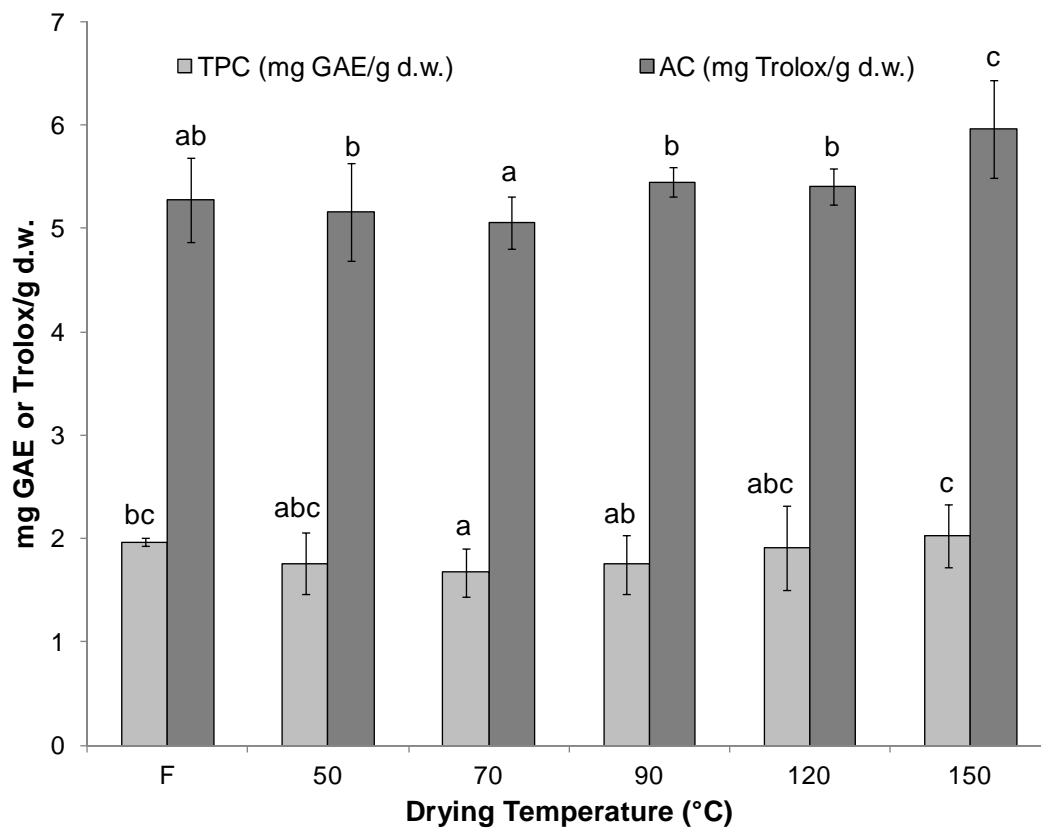
755 Figure 4

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757 Figure 5

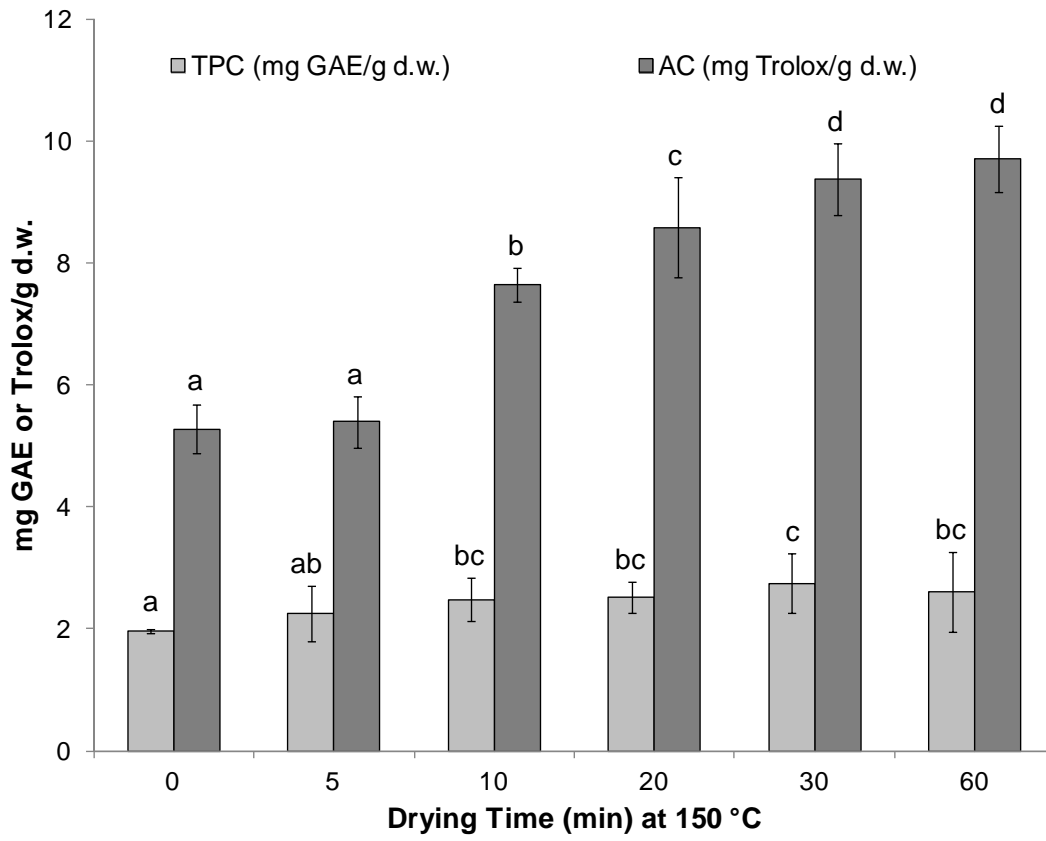
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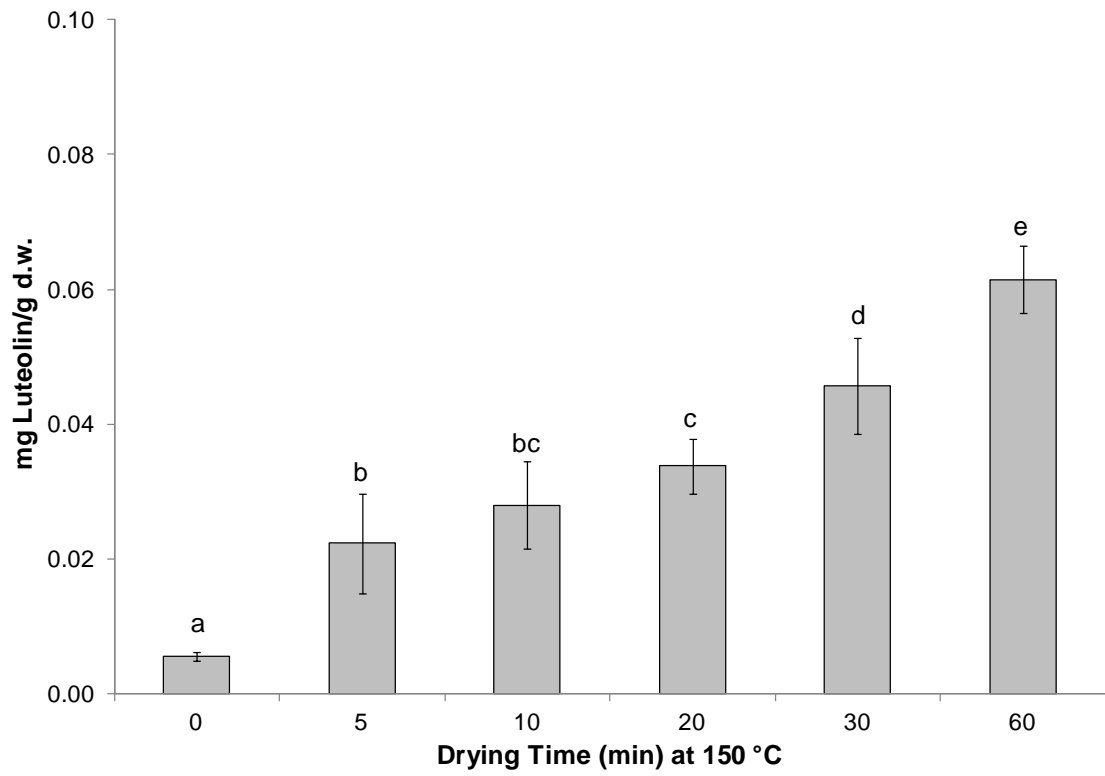
760 Figure 6

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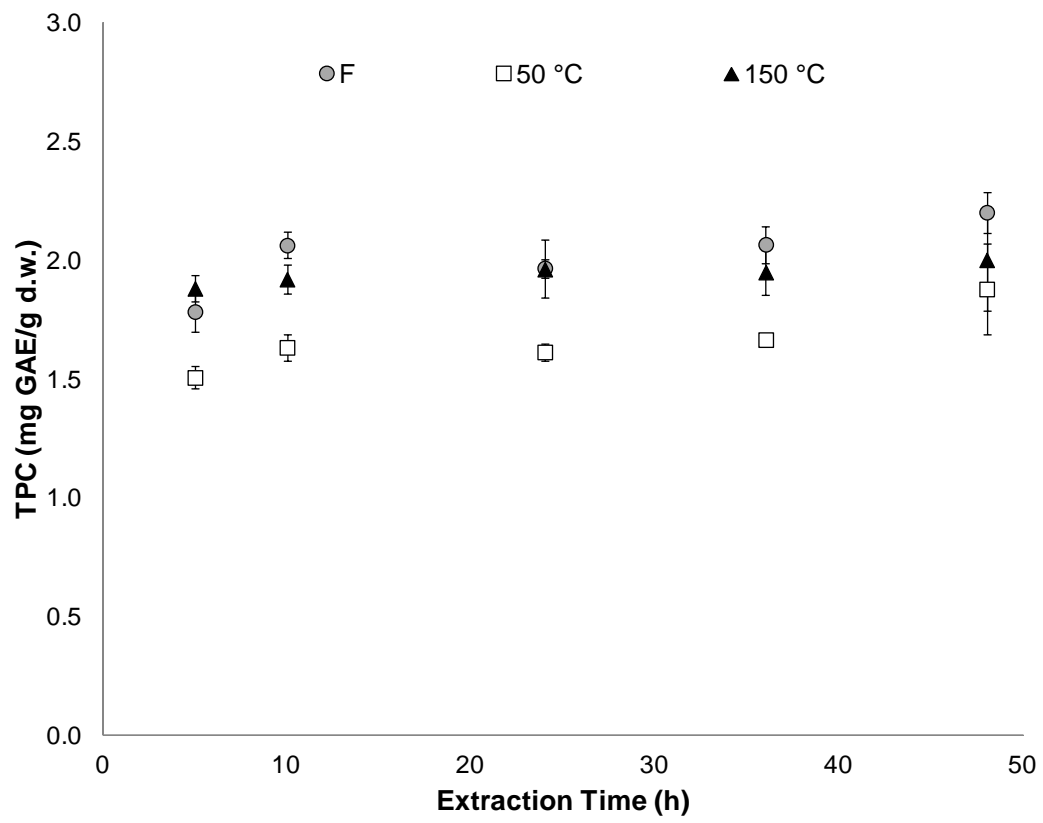
762 Figure 7

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764
765 Figure 8

766



767 Figure 9

768

769 **Table 1.** Characterization of pit (PI) and pulp+peel (P+P) fractions of olive
 770 pomace.
 771

	PI	P+P
ρ (kg/L)	1.30 \pm 0.05	1.5 \pm 0.2
W_0 (g w/g d.w)	0.234 \pm 0.004	0.66 \pm 0.05
X	0.424 \pm 0.005	0.576 \pm 0.005
r_m (mm)	1.80 \pm 0.02	0.311 \pm 0.015
Y_1 ($S_p > 10$ mm ²)	0.584	0.502
Y_2 ($1 < S_p < 10$ mm ²)	0.381	0.384
Y_3 ($0.25 < S_p < 1$ mm ²)	0.021	0.063
Y_4 ($S_p < 0.25$ mm ²)	0.015	0.051

772 ρ = density, W_0 =initial moisture content, X = mass fraction, r_m = characteristic dimension
 773 (thickness in PI and radius in P+P fraction), Y = sub-fraction of particles with a specific surface
 774 (S_p), $Y_1+Y_2+Y_3+Y_4 = 1$)

775

776

777 **Table 2.** Modelling of drying kinetics of olive pomace carried out at different
 778 temperatures and identified parameters of Weibull and Diffusion models.
 779

		Drying temperature (°C)				
		50	70	90	120	150
Weibull	α	0.88	0.85	0.88	0.85	0.99
	β (s)	14.4	8.5	6.2	3.7	3.6
	% VAR	99.4	99.5	99.6	99.4	99.7
	% MRE	9.1	11.0	8.4	10.1	4.6
Diffusion	D_e^{PI} (m ² /s)	$1.17 \cdot 10^{-7}$	$1.60 \cdot 10^{-7}$	$2.69 \cdot 10^{-7}$	$2.90 \cdot 10^{-7}$	$2.92 \cdot 10^{-7}$
	D_e^{P+P} (m ² /s)	$3.58 \cdot 10^{-11}$	$6.48 \cdot 10^{-11}$	$8.45 \cdot 10^{-11}$	$1.59 \cdot 10^{-10}$	$1.60 \cdot 10^{-10}$
	% VAR	98.8	99.1	99.03	98.9	96.9
	% MRE	11.9	10.9	9.5	9.2	11.3

780

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782