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Additional Information

12 **Abstract**

13 The effect of surfactant addition on structural, mechanical, optical and barrier properties of
14 corn starch-glycerol based films was studied. Sorbitan monopalmitate, monostearate or
15 monooleate were incorporated into starch-glycerol (1:0.25) at a surfactant:starch ratio of
16 0.15:1. The film forming dispersions (FFD) were characterized as to rheology, ζ -potential,
17 particle size distribution and contact angle. Film characterization was carried out at 1 and 5
18 storage weeks (at 25°C and 53 % relative humidity). Surfactants led to different particle
19 size distribution, zeta potential and viscosity in FFD, and film extensibility, depending on
20 their hydrophobicity and melting properties. Their incorporation to the corn starch-glycerol
21 films produced a coarser film microstructure due to the appearance of free surfactant
22 aggregates or V-amylose inclusion complexes which produce discontinuities in the
23 amorphous continuous matrix. The size of these crystalline complexes was smaller for the
24 surfactant with the lowest hydrophobicity with saturated fatty acid (span 40). This
25 contributed to decrease the WVP values with respect to surfactant-free film. Films
26 containing surfactants were less hard, resistant and extensible, and more permeable to
27 oxygen, than surfactant-free films, but they did not notably affect the film gloss and
28 transparency. Saturated fatty acid compounds with higher melting temperature are
29 recommended to ensure a finer microstructure in the final film which favours water barrier
30 efficiency.

31

32 Key words: starch, surfactants, physical properties, microstructure, ageing.

33 **1. Introduction**

34 The environmental conservation policies, the integral use of natural resources and the
35 reduction of hydrocarbon reserves have generated particular interest in developing
36 alternatives to petroleum synthetic polymers for different industrial uses, particularly for
37 foodstuffs. Several studies focused on the development of biodegradable materials to
38 replace, at least partially, conventional plastics. The materials obtained from natural
39 biopolymers, such as polysaccharides, are an interesting alternative, of which, starch is one
40 of the most promising materials for the manufacture of biodegradable plastics (Ma *et al.*,
41 2009). This polymer is a renewable, low cost resource, readily available and has
42 thermoplastic characteristics which permit it to be processed easily by using conventional
43 synthetic polymer methods (Shah *et al.*, 1995). However, starch presents worse physical
44 characteristics than synthetic polymers. Its main weaknesses are its highly hydrophilic
45 nature, which makes it a poor water vapour barrier, and the fact that it undergoes
46 retrogradation processes, which implies that its mechanical properties vary over time.
47 Nevertheless, their properties can be modified by adding small quantities of chemical
48 compounds (García *et al.*, 2000, Ma *et al.*, 2009).

49 Plasticizers act by increasing the molecular mobility in the polymeric network, thus
50 improving mechanical properties, but reducing the water vapour barrier properties (Rosen,
51 1993). Glycerol can be added as a plasticizer to improve the mechanical properties of the
52 film, increasing the flexibility and tensile strength by lowering the glass transition
53 temperature (Vieira *et al.*, 2011).

54 Other components with potential capacity to improve some properties of the starch-based
55 films are surfactants. Some studies have found that surfactants may enhance the wettability
56 and stability of the dispersions (Ghebremeskel *et al.*, 2007; Chen *et al.*, 2009), reduce the

57 starch retrogradation (Jovanovich and Añón, 1999) and improve the water vapour barrier
58 properties (Villalobos *et al.*, 2006). Vieira *et al.*, 2011 have observed that if the surfactant
59 was added without glycerol, it had a significant effect on mechanical properties but did not
60 significantly modify the water vapour barrier properties. Nevertheless, if the surfactant was
61 added with glycerol, it provoked a reduction in the tensile strength and an increase in both
62 elongation and water vapour permeability. Surfactants have been added to different
63 polysaccharide matrices, such as tapioca starch/decolorized hsian-tsau leaf gum films
64 (Chen *et al.*, 2009). When concentration and HLB of surfactant increased, there was an
65 observed improvement in the water vapour barrier properties, although films showed a loss
66 of mechanical resistance. Nevertheless, in matrices of corn starch reinforced with
67 microfibrils of cellulose, the mechanical properties were improved with the addition of
68 glyceryl monostearate by the formation of complexes which increase the V-type
69 crystallinity (Mondragón *et al.*, 2008).

70 Working on films of potato starch and glycerol, Rodríguez *et al.* (2006) observed that the
71 presence of surfactants increased the wettability of film forming dispersions, with a
72 synergistic effect between glycerol and surfactants. Similarly, in chitosan films, surfactants
73 showed a synergistic effect with the glycerol on the water vapour permeability (Ziani *et al.*,
74 2008). Zhong and Li (2011) also observed that the addition of surfactants and citric acid
75 diminished the surface tension in the film forming dispersions and the formation of
76 crystalline forms in kudzu starch films.

77 Despite several works have report the effect of different surfactants on biopolymer films,
78 no previous studies have been published in corn starch films containing this kind of
79 compounds, except that reported by Jimenez *et al.* 2012 where the effect of different fatty
80 acids, with very low (between 1-2) hydrophilic-lipophilic balance (HLB), was analysed. A

81 low HLB value indicates the predominant lipophilic balance in the molecular structure
82 which can contribute to limit the water vapour permeability of the films, although slightly
83 higher values could improve the better integration of these compounds in the hydrophilic
84 matrix. Likewise, the presence of saturated and unsaturated hydrocarbon chains could also
85 play a relevant role in the component interactions in the matrix. In this sense, the study of
86 the effect a family of surfactants with intermediate HLB values containing saturated or
87 unsaturated chains could give interesting information about the different effects on the
88 properties of the film forming dispersions and films, which are relevant for both coating
89 applications on a determined product and film formation for packaging ends. Surfactants
90 incorporated to film aqueous forming dispersions can act as carrier vehicles of non-polar
91 bioactive compounds (such as antimicrobials or antioxidants), making their dispersion easy
92 in the non-polar core of the formed micellar structures.

93 The aim of this work was to study the effect of addition of sorbitan esters of different fatty
94 acids (saturated and unsaturated), with low-intermediate HLB values (sorbitan
95 monopalmitate: 6.7, sorbitan monostearate: 4.7 and sorbitan monooleate: 4.3) on physical
96 properties of film forming dispersions (particle size distribution, ζ -potential, contact angle)
97 and films (mechanical, optical, structural and barrier properties) of corn starch-glycerol
98 based blends. The effect of storage time on film properties was also analyzed.

99

100 **2. Material and Methods**

101 **2.1 Materials**

102 Corn starch was obtained from Roquette (Roquette Laisa España, Benifaió, Spain). The
103 glycerol and surfactants (sorbitan monopalmitate: span 40, sorbitan monostearate: Span 60

104 and sorbitan monooleate: Span 80) were provided by Panreac Química, S.A. (Castellar del
105 Vallès, Barcelona, Spain).

106

107 **2.2 Preparation of film-forming dispersions (FFD)**

108 Four different dispersions based on corn starch, glycerol and surfactant were prepared by
109 using starch:glycerol:surfactant ratios of 1:0.25:0.15. The starch:surfactant ratio was chosen
110 on the basis of previous studies (Jimenez *et al.*, 2012). Corn starch (1% w/w in distilled
111 water) was gelatinized in a thermostat-controlled bath at 95 °C for 30 min. Then, the
112 glycerol and the surfactants (previously melted) were added and the mixtures were
113 homogenized. The homogenization was carried out under vacuum, to avoid bubble
114 formation, by using a venturi vacuum pump connected to the homogenization chamber.
115 This step was performed at 95 °C using a rotor-stator homogenizer (Ultraturrax T25, Janke
116 and Kunkel, Germany) for 1 min at 13,500 rpm and for 3 min at 20,500 rpm. Melting
117 temperature of surfactants are 46-47, 54-57 and 0.98 °C, respectively for Span 40, Span 60
118 and Span 80. Therefore, they remain in liquid state during the homogenization step, thus
119 making their dispersion easy. Prior to the characterization, FFD were cooled to 25 °C.
120 Sample control (FFD-C) was considered as a dispersion containing only starch and
121 glycerol. FFD with surfactants span 40, span 60 and span 80 were designated FFD-S40,
122 FFD -S60 and FFD-S80, respectively.

123

124 **2.3 Characterization of the film-forming dispersions**

125 **2.3.1 Rheological behaviour**

126 The rheological behaviour of FFD was analyzed in triplicate at 25 °C by means of a
127 rotational rheometer (Haake Rheostress1, Thermo Electric Corporation, Germany), by

128 using a coaxial cylinder sensor system (Z34DIN Ti). Rheological curves were obtained
129 after a sample resting time in the sensor of 5 min at 25 °C. The shear stress (σ) was
130 measured as a function of shear rate ($\dot{\gamma}$) from 0 to 512 s⁻¹, obtaining the up and down
131 curves taking 5 min for each one.

132

133 **2.3.2 Particle size, ζ -potential and surface tension measurements**

134 The droplet size distribution, volume-length mean diameter ($D_{4.3}$) and volume-surface
135 mean diameter ($D_{3.2}$) were determined in FFD, in triplicate, with a laser light scattering
136 instrument (Malvern Mastersizer, Malvern Instruments, Worcestershire, U.K.). To obtain ζ -
137 potential values, all FFD were diluted to a droplet concentration of 0.02 % using deionised
138 water. ζ -Potential was determined at 25 °C by mean of Zetasizer nano-Z (Malvern
139 Instruments, Worcestershire, UK). The Smoluchowsky mathematical model was used to
140 convert the electrophoretic mobility into ζ -potential values.

141 The surface tension was determined at 25 °C by means of the ring method using a ring
142 tensiometer (Mobil-K9 Tensiometer, Kruss GmbH, Hamburg, Germany) and a platinum-
143 iridium ring of 19.09 mm in diameter. Measurements were performed in triplicate in each
144 FFD.

145

146 **2.3.3 Contact angle**

147 Due to the potential application in multilayer packaging of these dispersions with synthetic
148 polymers, contact angle (θ) was measured on the surface of polyethylene terephthalate
149 (PET) (Cubil S. L. Barcelona, Spain) copolymer polypropylene (PP) (Cubil S. L.
150 Barcelona, Spain) and polystyrene (PS) (Cubil S. L. Barcelona, Spain). The shape of a

151 sessile drop (0.01 ml) was studied after 10 s by means of a Video-Based Contact Angle
152 Meter model OCA 20 (DataPhysics Instruments GmbH, Filderstadt, Germany). Image
153 analyses were carried out using SCA20 software. Ten replicates were made per
154 formulation.

155

156 **2.4 Preparation and characterization of films**

157 Films were obtained by casting method. The mass of the FFD containing 1 g of total solids
158 was spread evenly over a teflon casting plate of 15 cm diameter resting on a level surface.
159 Films were formed by drying for approximately 48h at 45% RH and 20 °C. Dry films could
160 be peeled intact from the casting surface and they were conditioned for 1 and 5 weeks at
161 53% RH and 25°C depending on the analysis. RH in the storage chamber was controlled by
162 using an oversaturated solution of Magnesium nitrate-6-hydrat (Panreac Quimica, S.A.).
163 Films without surfactant (control) are denominated F-C and films containing Span 40, Span
164 60 and Span 80, FS-40, FS-60 and F-S80, respectively. Film thickness was measured with a
165 Palmer digital micrometer to the nearest 0.0025 mm at 4-6 random position, depending on
166 the analysis.

167

168 **2.4.1 Scanning electron microscopy (SEM)**

169 Microstructural analysis of the films was carried out by SEM using a scanning electron
170 microscope JEOL JSM-5410 (Japan). Film samples were maintained in a desiccator with
171 P₂O₅ (Panreac Quimica, S.A.) for two weeks to ensure that water was not present in the
172 sample (theoretical relative humidity in desiccator 0). Afterwards, films were frozen in
173 liquid nitrogen and cryofractured to observe the cross-section and surface of the samples.

174 Films were fixed on copper stubs, gold coated, and observed using an accelerating voltage
175 of 10 kV.

176

177 **2.4.2 X-ray diffraction**

178 X-ray diffraction patterns were recorded using an X-ray diffractometer (XRD, Bruker
179 AXS/D8 Advance). All the aged samples (equilibrated for five weeks at 25 °C and 53%
180 RH) were analyzed between 2θ : 5° and 50°, using $K\alpha$ Cu radiation (λ : 1.542 Å), 40 kV and
181 40 mA with a step size of 0.05°. For this analysis, samples were cut into 4 cm squares, prior
182 to storage, in order to avoid breakage during handling. Span 40 and Span 60 reagents were
183 also analyzed in order to identify their characteristic peaks.

184

185 **2.4.3 Atomic force microscopy (AFM)**

186 The surface morphology of dried aged films (equilibrated in a desiccator with P₂O₅ for 1
187 week) was analyzed using an atomic force microscope (Multimode 8, Bruker AXS, Santa
188 Barbara, USA) with a NanoScope[®] V controller electronics. Measurements were taken
189 from several areas of the film surface (50 x 50 μm and 5 x 5 μm) using the Tapping mode.
190 According to method ASME B46.1 (ASME, 1995), the following statistical parameters
191 related with sample surface roughness were calculated: average roughness (Ra: average of
192 the absolute value of the height deviations from a mean surface), root-mean-square
193 roughness (Rq: root-mean-square average of height deviations taken from the mean data
194 plane), and factor of roughness (r: ratio between the three-dimensional surface and two-
195 dimensional area projected onto the threshold plane). Three replicates were considered to
196 obtain these parameters.

197 Phase Imaging mode derived from Tapping Mode, that goes beyond topographical data,
198 was also applied. Phase imaging is the mapping of the phase lag between the periodic
199 signal that drives the cantilever and the oscillations of the cantilever. Changes in the phase
200 lag indicate changes in the properties of the sample surface in composition, adhesion,
201 friction, viscoelasticity, and other properties, including electric and magnetic.

202

203 **2.4.4 Optical properties**

204 The film transparency was determined by applying the Kubelka-Munk theory for multiple
205 scattering to the reflection spectra (Hutchings, 1999). The surface reflectance spectra of the
206 films were determined from 400 to 700 nm with a spectrophotometer CM- 3600d (Minolta
207 Co., Tokyo, Japan) on both a white and a black background. As the light passes through the
208 film, it is partially absorbed and scattered, which is quantified by the absorption (K) and the
209 scattering (S) coefficients. Internal transmittance (Ti) of the films was quantified using eq.
210 (1). In this equation R_0 is the reflectance of the film on an ideal black background.
211 Parameters a and b were calculated by eqs. (2) and (3), where R is the reflectance of the
212 sample layer backed by a known reflectance R_g . Measurements were taken in triplicate for
213 each sample on the free film surface. Wavelength of 450 nm was considered for analysis.

214

$$215 \quad T_i = \sqrt{(a - R_0)^2 - b^2} \quad (1)$$

216

$$217 \quad a = \frac{1}{2} \left(R + \frac{R_0 - R + R_g}{R_0 R_g} \right) \quad (2)$$

218

219
$$b = \sqrt{a^2 - 1} \quad (3)$$

220

221 The gloss was measured on the free film surface, at 60° incidence angle, according to the
222 ASTM standard D523 method (ASTM, 1999), by means of a flat surface gloss meter (Multi
223 Gloss 268, Minolta, Germany). Measurements were taken in triplicate for each sample and
224 three films of each formulation were considered. All results were expressed as gloss units,
225 relative to a highly polished surface of black glass standard with a value near to 100.
226 Transparency and gloss measurements were carried out with films conditioned for 1 week
227 (initial time) and 5 weeks (final time) in hermetic desiccators at 25 °C and 53% RH.

228

229 **2.4.5 Moisture content**

230 Films previously conditioned at 53% RH were dried for 24h at 60 °C in a natural
231 convection stove and subsequently placed in a desiccator with P₂O₅ at 25 °C for 2 weeks.
232 The reported results represent the mean values of at least three samples.

233

234 **2.4.6 Water vapour permeability (WVP)**

235 WVP measurements were carried out with films conditioned for 1 and 5 weeks in hermetic
236 desiccators at 25 °C and 53% RH. WVP of films was determined by using the ASTM E96-
237 95 (ASTM, 1995) gravimetric method, taking into account the modification proposed by
238 McHugh et al., 1993. Films were selected for WVP tests based on the lack of physical
239 defects. Distilled water was placed in Payne permeability cups (3.5 cm diameter, Elcometer
240 SPRL, Hermelle/s Argenteau, Belgium) to expose the film to 100% RH on one side. Once
241 the films were secured, each cup was placed in a relative humidity equilibrated cabinet at

242 25 °C, with a fan placed on the top of the cup in order to reduce resistance to water vapour
243 transport, thus avoiding the stagnant layer effect in this exposed side of the film. RH of the
244 cabinets (53%) was held constant using oversaturated solutions of magnesium nitrate-6-
245 hydrate. The free film surface during film formation was exposed to the lowest relative
246 humidity to simulate the actual application of the films in high water activity products when
247 stored at intermediate relative humidity. The cups were weighed periodically (0.0001 g)
248 and water vapour transmission (WVTR) was determined from the slope obtained from the
249 regression analysis of weight loss data versus time, once the steady state had been reached,
250 divided by the film area. From WVTR data, the vapour pressure on the film's inner surface
251 (p_2) was obtained with eq. (4), proposed by McHugh et al. (1993), to correct the effect of
252 concentration gradients established in the stagnant air gap inside the cup.

253

$$254 \quad WVTR = \frac{PDL_n \left[\frac{P - p_2}{P - p_1} \right]}{RT\Delta z} \quad (4)$$

255

256 where P, total pressure (atm); D, diffusivity of water through air at 25 °C (m^2 /s); R, gas
257 law constant ($82.057 \times 10^{-3} m^3 atm kmol^{-1} K^{-1}$); T, absolute temperature (K); Δz , mean
258 stagnant air gap height (m), considering the initial and final z value; p_1 , water vapour
259 pressure on the solution surface (atm); and p_2 , corrected water vapour pressure on the film's
260 inner surface (atm). Water vapour permeance was calculated using eq. (5) as a function of
261 p_2 and p_3 (pressure on the film's outer surface in the cabinet).

262

263
$$permeance = \frac{WVTR}{P_2 - P_3} \quad (5)$$

264

265 Permeability was obtained by multiplying the permeance by the average film thickness.

266

267 **2.4.7 Oxygen permeability (OP)**

268 The oxygen permeation rate of the corn starch films was determined at 53% RH and 25 °C
269 using an OX-TRAN Model 2/21 ML Mocon (Lippke, Neuwied, Germany). Film samples
270 were previously conditioned for 1 or 5 weeks at 25 °C and 53% RH by using magnesium
271 nitrate-6-hydrate saturated solutions. Two samples were placed in the equipment for
272 analysis and they were conditioned in the cells for 6 h, then the transmission values were
273 determined every 20 min until the equilibrium was reached. The exposure area during the
274 tests was 50 cm² for each formulation. In order to obtain the oxygen permeability, film
275 thickness was considered in all cases. Measurements were carried out in triplicate.

276

277 **2.4.8 Tensile properties**

278 The mechanical behaviour of films was tested by using conditioned samples (1 or 5 weeks
279 at 25 °C and 53 % RH). A universal test Machine (TA.XTplus model, Stable Micro
280 Systems, Haslemere, England) was used to determine the tensile strength (TS), elastic
281 modulus (EM), and elongation (E) of the films, according to ASTM standard method D882
282 (ASTM, 2001) EM, TS, and E were determined from the stress-strain curves, estimated
283 from force-distance data obtained for the different films (2.5 cm wide and 10 cm long).
284 Equilibrated samples were mounted in the film-extension grips of the testing machine and
285 stretched at 50 mm min⁻¹ until breaking. The relative humidity of the environment was held

286 at nearly 53% during the tests, which were performed at 25 °C. At least seven replicates
287 were obtained from each sample.

288

289 **2.5 Statistical analysis**

290 Statgraphics Plus for Windows 5.1 (Manugistics Corp., Rockville, MD) was used to carry
291 out statistical analyses of data through analysis of variance (ANOVA). Fisher's least
292 significant difference (LSD) was used at the 95% confidence level.

293

294 **3. Results and discussion**

295 **3.1 Properties of film forming dispersions**

296 [Figure 1](#) shows the particle size distribution for the film forming dispersions containing
297 corn starch and surfactants (Span 40, Span 60 and Span 80). The values of mean diameters
298 $D_{3,2}$ and $D_{4,3}$ are shown in [Table 1](#). A similar distribution is observed in all cases. Most of
299 the volume fraction of the dispersed particles is under 1 μm in size, although there are large
300 particles (over 10 μm) in all cases that greatly contribute to the average size. $D_{4,3}$ values
301 indicate that the particle size of FFD-S80 was significantly smaller ($p < 0.05$) than those of
302 FFD-S40 and FFD-S60 whereas no significant differences ($p < 0.05$) were observed
303 between the particle sizes of these last FFD.

304 Surfactants, as polar lipids, form different kind of molecular association (association
305 colloids) in aqueous systems, where the hydrocarbon chains are oriented to minimize the
306 contact with water molecules. These aggregates (micellar structures) have different size and
307 shape depending on their relative water affinity (HLB) and the overall balance of molecular
308 interactions which minimize the total free energy of the system (Dickinson, 1992). Usually,
309 the lower the water affinity (HLB) the greater the aggregation number and so, the greater

310 the particle size. In this sense, it is remarkable that S80, with the lowest HLB, forms the
311 smallest particles which could be related with the double bond in the oleic acid chain,
312 which make the packing of molecules more difficult. This is also related with the liquid
313 state of this compound under room conditions, whereas S40 and S60 are solids under the
314 same conditions. There are a greater proportion of large particles in FFD-S60 and FFD-
315 S40, which may be associated with the formation of big aggregates of the surfactant
316 molecules (Villalobos et al., 2005) or even combined aggregates of surfactants and starch
317 polymer chains. Associations of helical amylose which give rise to gel zones in the FFD
318 could also be detected as particles in the measurements. Surfactant molecules can also
319 participate in these associations through the formation of complexes with helical
320 conformation of amylose (Wokadala et al., 2012).

321 Table 1 shows the rheological behaviour, ζ -potential, surface tension and contact angle with
322 different polymer films (PET: Polyethylene terephthalate, PP: polypropylene copolymer,
323 PS: polystyrene) of FFDs containing corn starch and surfactants. All film-forming
324 dispersions showed similar rheological behaviour. At low shear rates, Newtonian behaviour
325 was observed, but from a determined shear rate, FFDs exhibited shear thickening
326 behaviour. No differences were observed between up and down which means that no
327 tixotropic effects occur during the fluid flow.

328 Table 1 shows the Newtonian viscosity values in the first step and the shear rate values
329 where the change in behaviour occurs, which range between 130 and 173 s^{-1} . The shear
330 thickening behaviour from a determined shear rate can be explained by the induced
331 orthokinetic flocculation of particles. Particle aggregates retain a part of the continuous
332 phase and this, together with the greater flow resistance of the bigger aggregates, defines
333 the higher shear stress-shear rate relationship or flow resistance (Peker and Helvacı, 2007).

334 Non-Newtonian behaviour has previously been observed by other authors in starch
335 dispersions and pastes (Zhong *et al*, 2009) where the viscosity varied according to the shear
336 rate applied. Table 1 shows that the presence of surfactants caused a significant increase
337 ($p < 0.05$) in the Newtonian viscosity in all cases; the FFD-S40 dispersion exhibiting the
338 highest values. No significant differences were observed between the FFD-S60 and FFD-
339 S80 formulations despite their different particle sizes. This viscosity increase provoked by
340 the addition of surfactants is explained by the greater concentration of dispersed/dissolved
341 species in the aqueous media.

342 All dispersions had negative ζ -potential values with significant differences ($p < 0.05$) among
343 formulations (Table 1). All FFDs containing surfactants showed higher values of ζ -
344 potential than that containing only starch. This can be due to the promotion of the
345 adsorption of negative ions on the polar heads of the surfactant associations. Some authors
346 (Ho and Ahmad, 1999; Hsu and Nacu, 2003) have reported that at pH near 7, non-ionic
347 surfactants attract OH^- ions of the aqueous medium, thus increasing the surface charge of
348 the lipid associations while a pH decrease is promoted.

349 In Table 1, the lower pH of FFD-S60 and FFD-S80 can be observed, while these FFDs also
350 showed the highest surface charge. Variations in ζ -potential could also be due to the
351 different degree of hydrolysis in sorbitan esters, giving rise to free fatty acids with ionisable
352 groups. The ζ -potential is defined as the electrostatic potential in the layer of the particle
353 electrophoretic mobility and greatly affects the stability of the colloidal systems by
354 electrostatic forces (Ravina and Moramarco, 1993). Accordingly, the most stable FFD
355 would be FFD-S80 and FFD-S60 because the contribution of repulsive forces between
356 particles to the stability would be greater. However, it should be noted that colloidal
357 stabilization is the result of the interaction of numerous factors. In the case of FFD, the

358 particle charge may not be enough to ensure stability of the dispersion because it is a
359 complex system in which the hydrocolloid interactions (amylose and amylopectin) also
360 play an important role. In particular, the viscosity of the continuous phase, which depends
361 on the molecular interactions, greatly affects the system stability. So, as FFD-S80 and FFD-
362 S60 are less viscous, destabilization phenomena during film drying could take place more
363 easily, promoting coalescence and creaming, despite their greater surface charge. Other
364 factors, such as the physical state of the lipid, may affect the coalescence phenomena, this
365 being inhibited in solid lipids.

366 Surface tension values showed significant differences ($p < 0.05$) among formulations. The
367 control dispersion (FFD-C) exhibited similar values to pure water (72 mN/m at 25 °C), but,
368 as expected, the presence of surfactants reduced the surface tension of the dispersions to
369 values below 47.6 mN/m at 25 °C. The surface tension reduction in the FFDs is interesting
370 from the point of view of their application on a determined surface as coatings, since this
371 favours their extensibility during the product coating (Fernandez et al., 2006) and the
372 possible formation of multi-layer materials with other polymers (Chen *et. al.*, 2010)

373 The contact angle values of FFD on the surface of different polymer films (PET:
374 Polyethylene terephthalate, PP: polypropylene copolymer, PS: polystyrene) are shown in
375 Table 1. Values lower than 90° indicate surface wettability and therefore greater
376 extensibility of the dispersion on the polymer. The polymer which was least wettable with
377 the studied dispersions was PP, since the greatest contact angles were obtained in this case
378 ($p < 0.05$). PET had the lowest contact angles and therefore it offers the best possibilities to
379 be coated with the studied FFDs, for instance as vehicles of active compounds, thus
380 obtaining multi-layer films. Nevertheless, the obtained values were dependent on the

381 composition of the FFDs and it was remarkable that the formulations without surfactant
382 were those which showed the greatest extensibility despite their higher surface tension.

383

384 **3.2 Microstructure of the films**

385 Films obtained had $50 \pm 5 \mu\text{m}$ in thickness, regardless of the FFD composition. The final
386 structure of the film depends on both the interactions of film components and the drying
387 conditions of the film-forming dispersion and has a great impact on the different film
388 properties (Villalobos et al., 2005). The microstructural analysis of the films gives relevant
389 information about the arrangement of the components, which, in turn, allows us to explain
390 the differences in the barrier, mechanical or optical properties of the films.

391 Figure 2 shows the SEM micrographs of the surface (left) and the cross-section (right) of
392 starch based films with and without surfactants, conditioned for one week. Control films
393 (F-C) showed an homogeneous structure, coherent with the formation of a compact
394 arrangement of polymer chains. The addition of surfactants resulted in less homogeneous
395 films whose differences can be observed in the cross-section images. Films with surfactants
396 showed a coarser cross section than the control films, which suggests that polymer chain
397 packaging was interrupted by other structural elements present in the matrix. These
398 elements may be free surfactant particles, resulting from the micellar structure in the
399 aqueous FFD and surfactat particles of V-amylose inclusion complexes which produce
400 discontinuities of different magnitude in the amorphous continuous matrix, thus
401 contributing to weaken the bond strength between the polymer chains in the amorphous
402 matrix. Interactions of polar lipids with V-amylose leads to complex formation, where the
403 helical conformation of amylose traps the hydrocarbon chains of lipids, as has been

404 previously described by other authors (Singh et al., 2002). V-amylose forms small particles
405 with size in the nano or micro scale, depending on medium conditions (Lesmes et al.,
406 2009).

407 In F-S80, micro-droplets of span 80 (liquid at room temperature) can be seen in the matrix,
408 whereas no droplets can be observed for F-S40 and F-S60, but finer no spherical
409 formations. Lipid droplets also appear on the F-S80 film surface, even larger than in the
410 cross-section due to the coalescence and creaming phenomena occurring in the drying step.
411 Similar phenomena were observed by other authors (Zhong and Li, 2011) in kudzu starch
412 films. The presence of droplets in F-S80 films is related with the liquid state of span 80 at
413 room temperature and reveals the no total integration of the lipid in the starch matrix, and
414 its separation in a dispersed liquid phase. Span 40 and span 60 seem also separated
415 (discontinuities in the matrix) but their solid state at room temperature did not allow
416 droplets formation, but crystalline or amorphous solid particles.

417 Figure 3 shows the phase imaging analysis obtained from the Tapping Mode AFM for films
418 conditioned for five weeks. The phase image is providing material property contrast on the
419 film surface since this technique is sensitive to variations in composition, adhesion, friction,
420 viscoelasticity, and other surface properties (Baker et al., 2001). In control film (F-C), the
421 image reveals the presence of lighter zones which can be attributed to crystalline aggregates
422 of helical amylose (Baker et al., 2001). In films containing span 40 the presence of lighter
423 spots reveals the presence of lipid particles on the film surface due to lipid coalescence and
424 creaming during film drying and subsequent solidification. The excess of lipid contributed
425 to its separation in the matrix and its migration to the film surface where solidifies,
426 according to its high melting temperature (46-47 °C). In the case of span 60, no big lipid
427 formations were observed on the film surface probably due to its higher melting

428 temperature (54-57°C), which contribute to inhibit coalescence phenomena during the film
429 drying step, which also limits creaming, due to the restrictions in the particle size growth.
430 For the film containing span 80 (liquid at room temperature, melting point: 0.98 °C) a great
431 proportion of lipid formations can be observed on the film surface, coherent with a high
432 progress of the coalescence and creaming phenomena during the film drying. The irregular
433 shape of lipid droplets reveals that they are droplet aggregates, whose surface is covered by
434 a thin polymer layer, which entraps them.

435 The melting properties and molecular characteristics of the surfactant had an impact on the
436 film structure and surface morphology which will affect the film properties, mainly its
437 mechanical and optical parameters.

438 Figure 4a shows X-ray diffraction (XR) spectra of corn starch films with and without
439 surfactants, equilibrated at 25°C and 53% RH for 5 weeks. Generally, the native starch
440 crystallinity is types A and B, type C being an intermediate form (Jiménez et al., 2013);
441 crystalline V type could also be observed, corresponding to the amylose helical
442 associations, which are the result of the amylose complexation with other substances, such
443 as lipids (Jiménez et al., 2013). The addition of surfactants affected the crystallization
444 pattern of the films. Control film showed the typical peaks of amylose type B, with a peak
445 of maximum intensity at 2θ 17° and other peaks at 2θ about 15°, 20° and 22 ° (Lesmes et al.,
446 2009). When surfactants were added, two main peaks are shown at 2θ about 13° and 20°, as
447 has been observed for V-type amylose hosting complexes with long chain fatty acids
448 (Lesmes et al., 2009). So, the XR spectra reveal that surfactants in the film induce the
449 formation of complexes with amylose, as observed for other lipids. Wokadala et al., (2012)
450 reported peaks at 2θ 7.5°, 13° and 20°, attributable to V-type crystallinity as a result of the

451 presence of amylose-lipid complexes in hydrolyzed maize starch in the presence of stearic
452 acid.

453 By considering the XR diffraction pattern of the pure surfactants (Figure 4b), where a very
454 sharp peak appears at 2θ 22° , in both cases, the non-crystalline forms of the non-complexed
455 (free) lipid in the films must be deduced, since at this angle no sharp peak was observed in
456 the film RX diffraction spectra.

457 These results indicate that in the structure of surfactant-starch films, V-type amylose
458 complexes are formed with surfactant molecules giving rise to crystalline aggregates,
459 whereas the excess of lipid is not crystallized. These crystalline aggregates interrupt the
460 starch matrix producing discontinuities in the same way as the free surfactant molecules. In
461 the case of span 80, free lipids are clearly observable in the SEM micrographs as liquid
462 droplets dispersed in the starch matrix, with a high degree of lipid creaming on the film
463 surface. Film F-S40 showed the lowest peak intensity in the XR spectra, which suggests
464 that V-type crystalline forms were the smallest in size, which could have a positive effect
465 on the film properties.

466

467 **3.3 Physical properties of films**

468 Table 2 shows the optical properties (gloss at incidence angle of 60° and internal
469 transmittance at 450 nm) of corn starch films with and without surfactants, conditioned for
470 one and five weeks. The gloss values obtained were less than 35 units and, therefore, films
471 can be considered of low gloss (Villalobos et al., 2005). The addition of surfactants did not
472 have a significant effect ($p > 0.05$) on the gloss values as compared with the control
473 formulation. In the case of F-S80, the gloss significantly increased during the storage time,
474 which did not occur in the other cases. This increase can be attributed to a progressive

475 migration of the liquid droplets to the film surface, in line with the progressive chain
476 aggregations and the reduction of the matrix volume. The fluid lipid acted as a filler of the
477 micro-pores in the surface, thus reducing the apparent roughness and increasing the gloss.
478 Table 2 shows the values of roughness parameters (R_a , R_b and r) analyzed through AFM in
479 films stored for five weeks. No significant differences in the film roughness were detected,
480 except for F-S40 where higher values were obtained. Nevertheless, this difference was not
481 coherent with the gloss values, unlike that reported by other authors (Villalobos et al.,
482 2005). This can be due to the liquid state of span 80 which interferes with the cantilever
483 signal in terms of the height of peaks on the surface.

484 All films were highly transparent, as deduced by the values of the internal transmittance
485 (T_i) in Table 2. This is associated with a greater homogeneity in the film structure. The
486 addition of surfactants resulted in a slight decrease of the transparency with respect to the
487 control film, in line with a more anisotropic structure due to the formation of crystalline
488 forms and the presence of dispersed surfactants aggregates, with a different refractive
489 index, which enhance light scattering, as observed by Jimenez et al. (2012) in corn starch
490 films containing fatty acids. The F-S80 films showed a significantly ($p < 0.05$) lower T_i ,
491 which can be attributed to the liquid droplet dispersion observed in the film microstructure.
492 During film storage, no notable changes in T_i could be observed, although control films
493 showed a slight decrease which could be attributed to the greater progress of amylose
494 crystallization, thus inducing a greater degree of anisotropy (Jimenez et al., 2012).

495 Table 3 shows the moisture content, water vapour permeability (WVP), oxygen
496 permeability (OP) and mechanical properties of corn starch films, with and without
497 surfactants. The incorporation of surfactants greatly decreased the equilibrium water
498 content of the films, even if this is considered in the lipid free basis (4.3, 4.9 and 5.4,

499 respectively for the films with span 40, span 60 and span 80). This agrees with the changes
500 in the packing of a part of the polymer chains involved in the V-type crystalline forms
501 complexing the lipids, which reduces the water binding capacity of these chains.

502 The WVP was measured at a gradient of 53-100% RH. At one week of storage, the
503 addition of span 40 implied a significant ($p < 0.05$) reduction of WVP, although this was
504 not observed for films containing span 60 (without significant differences with respect to
505 the control) and span 80, where a significant increase was observed, more markedly at 5
506 weeks' storage. The improvement of the water vapour barrier properties due to the addition
507 of surfactants has been observed in different matrices, such as tapioca starch with sucrose
508 esters (Chen et al., 2009), hydroxypropyl methylcellulose with span 60 and sucrose esters
509 (Villalobos et al., 2006) and kudzu starch and ascorbic acid based films with Tween 20
510 (Zhong and Li, 2011). The scarce effect of surfactants on WVP observed in these films,
511 regardless of their hydrophobicity, could be explained by the formation of crystalline
512 structures in all cases (even in control films) which could be a strong limitation factor for
513 transfer of water molecules.

514 In terms of WVP, the different behaviour induced by each of the three studied surfactants
515 must be related with the structural differences reached in the matrix for each case. F-S40
516 showed the smallest V-type crystalline forms, as deduced from RX spectra, which could
517 imply a great increase in the tortuosity factor for mass transfer in the matrix, thus reducing
518 WVP. On the contrary, the increase in WVP provoked by the addition of span 80 could be
519 related with the greater size of crystalline complexes. Lesmes et al. (2009) demonstrated
520 that increased fatty acid unsaturation affects the microscopic and nanoscopic arrangement
521 of V-amylose, and fatty acid unsaturation leads to bigger and more disperse populations.
522 This can suppose a more open structure where water molecules could diffuse more easily.

523 No significant differences ($p < 0.05$) between WVP of the films stored for one and five
524 weeks were observed, except for F-S80 where WVP increased over time. This could be
525 explained by the above mentioned migration of the free liquid droplets of span 80 to the
526 film surface, leaving fewer hydrophobic areas or void nanopores in the matrix, which
527 facilitate transfer of water molecules.

528 Oxygen permeability values (Table 3) significantly ($p < 0.05$) increased when surfactants
529 were incorporated into the films, as usually happens when hydrophobic components are
530 added, since these substances are more permeable to gases due to their greater chemical
531 affinity and solubility (Miller and Krochta, 1997). The oxygen permeability did not vary
532 with the storage time, except in the control film, where it decreased significantly ($p < 0.05$).
533 This can be explained by the progressive increase in the film crystallinity, as observed in
534 previous works (Donhowe and Fennema, 1993; Jimenez et al. 2012).

535 Table 3 shows the values of the parameters used to describe the mechanical properties of
536 starch based films at 25 °C and 53% RH: elastic modulus (EM), tensile strength (TS) and
537 elongation at break (E). In general, the addition of surfactants caused the decrease of EM,
538 TS and E, in agreement with the formation of a more anisotropic structure with reduced
539 cohesion forces. Similar results were observed by Jimenez et al. (2012) in corn starch films
540 when fatty acids were added. Rodriguez, et al. (2006) also observed that the addition of
541 surfactants in the presence of glycerol causes a significant reduction in tensile strength and
542 elongation at break with respect to the control formulation. TS and EM initial values for F-
543 S80 were significantly higher ($p < 0.05$) than for the other films with surfactants, which
544 showed very similar values. This indicates that bonding forces in the matrix were more
545 intense in the F-S80 films. Nevertheless, all films were very short, especially F-S60 and F-
546 S80, which showed the lowest deformation at break, thus indicating the low capacity of the

547 structural elements to slippage during tensile test without structural failure. This can be
548 attributed, in part, to the formation of crystalline zones with low deformability.

549

550 **4. Conclusions**

551 Incorporation of span 40, 60 or 80 into the corn starch-glycerol film forming dispersions
552 led to different particle size distribution, zeta potential, viscosity and extensibility on other
553 films, depending on their hydrophobicity and melting properties (related with the
554 unsaturation in the fatty acid chain). These aspects affected the final film microstructure
555 and its surface morphology, since the growing of the surfactant molecule aggregates during
556 the film drying (loss of water availability) occurred to a different extent, depending on the
557 dispersion stability. When the surfactant melting temperature was the highest (span 60),
558 creaming of surfactants in the film surface was not observed and the size of the final lipid
559 aggregates in the film were lower. Likewise, the size of the type-V crystalline amylose
560 complexes was smaller for the surfactant with the highest HLB value with saturated fatty
561 acid (span 40). This contributed to decrease the WVP values with respect to surfactant-free
562 film. Nevertheless, films containing surfactants were less hard, resistant and extensible than
563 surfactant-free films, but they did not notably affect the film gloss and transparency. The
564 use of surfactants in film formulation can be useful to incorporate non-polar bioactive
565 compounds in the aqueous film forming dispersions. In this sense, the obtained results
566 allow us to recommend saturated fatty acid compounds with higher melting temperature to
567 ensure a finer microstructure in the final film which favour water barrier efficiency.

568

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574 **References**

575 ASME. (1995). Surface texture: Surface roughness, waviness and lay. Standard
576 Designation: ASME B46.1. New York, USA: An American National Standard.

577 ASTM. (1995). Standard test methods for water vapor transmission of materials. Standards
578 Designations: E96-95. In Annual book of ASTM standards. Philadelphia, PA:
579 American Society for Testing and Materials., 406–413.

580 ASTM. (1999). Standard test method for specular gloss. Standard designation: D523. In
581 ASTM, annual book of ASTM, 06.01. Philadelphia: ASTM.

582 ASTM. (2001). Standard test method for tensile properties of thin plastic sheeting. Standard
583 D882. Annual book of American standard testing methods. Philadelphia, PA:
584 American Society for Testing and Materials, pp. 162-170.

585 Baker, A. A., Miles, M. J., & Helbert, W. (2001). Internal structure of the starch granule
586 revealed by AFM. *Carbohydrate Research*, 330, 249–256.

587 Chen, C.H., Kuo, W.S., & Lai, L.S. (2009). Effect of surfactants on water barrier and
588 physical properties of tapioca starch/decolorized hsian-tsao leaf gum films. *Food*
589 *Hydrocolloids*, 23, 714–721.

590 Chen, C.H., Kuo, W.S., & Lai, L.S. (2010). Water barrier and physical properties of
591 starch/decolorized hsian-tsao leaf gum films: Impact of surfactant lamination. *Food*
592 *Hydrocolloids*, 24, 200–207

593 Dickinson, E. (1992). An introduction to Food Colloids. Oxford Univesity Press, Oxford.

594 Donhowe, LG. & Fennema, O. (1993). The effects of solution composition and drying
595 temperature on crystallinity, permeability and mechanical properties of methylcellulose
596 films. *Journal of Food Processing and Preservation.*, 17 (4), 231-246.

597 Fernández, L., Díaz de Apodaca, E., Cebrián, M., Villarán, M.C. & Maté, J. I (2006). Effect
598 of the unsaturation degree and concentration of fatty acids on the properties of WPI-
599 based edible films. *European Food Research and Technology*, 224 (4), 415-420.

600 García M., Martino, M., & Zaritzky, N. (2000). Lipid addition to improve barrier properties
601 of edible starch-based films and coatings. *Food Chemistry and Toxicology*, 65 (6), 941-
602 946.

603 Ghebremeskel, A. N., Vemavarapu, C., & Lodaya, M. (2007). Use of surfactants as
604 plasticizers in preparing solid dispersions of poorly soluble API: Selection of polymer-
605 surfactant combinations using solubility parameters and testing the processability.
606 *International Journal of Pharmaceutics*, 328, 119-129.

607 Ho, C.C., & Ahmad, K. (1999). Electrokinetic Behavior of Palm Oil Emulsions in Dilute
608 Electrolyte Solutions. *Journal of Colloid and Interface Science*, 216, 25-33.

609 Hsu, J. P., & Nacu, A. (2003). Behavior of soybean oil-in-water emulsion stabilized by
610 nonionic surfactant. *Journal of Colloid and Interface Science*, 259, 374-381.

611 Hutchings, J. B. (1999). Food color and appearance (2nd ed.). Gaithersburg, Maryland,
612 USA: Aspen Publishers, Inc.

613 Jiménez, A., Fabra, M. J., Talens, P., & Chiralt, A. (2012). Effect of re-crystallization on
614 tensile, optical and water vapour barrier properties of corn starch films containing fatty
615 acids. *Food Hydrocolloids*, 26, 302-310.

616 Jiménez, A., Fabra, M. J., Talens, P., & Chiralt, A. (2013). Phase transitions in starch based
617 films containing fatty acids. Effect on wáter sorption and mechanical behaviour. *Food*
618 *Hydrocolloids*, 30, 408-418.

619 Jovanovich, G., & Añón, M. (1999). Amylose–lipid complex dissociation. A study of the
620 kinetic parameters. *Biopolymers*, 49 (1), 81–89.

621 Lesmes, U., Cohen, S. H., Shener, Y., & Shimoni, E. (2009). Effects of long chain fatty
622 acid unsaturation on the structure and controlled release properties of amylose
623 complexes. *Food Hydrocolloids*, 23, 667–675.

624 Ma, X., Chang, P., Yu, J., & Stumborg, M. (2009). Properties of biodegradable citric acid-
625 modified granular starch/thermoplastic pea starch composites, *Carbohydrate*
626 *Polymers.*, 75 (1), 1–8.

627 McHugh, T. H., Avena-Bustillos, R., & Krochta, J. M. (1993). Hydrophobic edible films:
628 Modified procedure for water vapour permeability and explanation of thickness effects.
629 *Journal of Food Science*, 58(4), 899–903.

630 Miller, K.S., & Krochta, J.M. (1997). Oxygen and aroma barrier properties of edible films:
631 A review. *Trends in Food Science and Technology*, 81, 228-237.

632 Mondragón, M., Arroyo, K., & Romero-García, J. (2008). Biocomposites of thermoplastic
633 starch with surfactant. *Carbohydrate Polymers*, 74, 201–208.

634 Peker, S.M., & Helvacı, S.S. (2007). Solid-liquid two phase flow. Chapter 3, Concentrated
635 Suspensions. Elsevier.

636 Ravina, L., & Moramarco, N. (1993). Everything you want to know about coagulation &
637 flocculation. Zeta-meter, Inc., Virginia.

638 Rodríguez, M., Osés, J., Ziani, K., & Maté, J. I. (2006). Combined effect of plasticizers and
639 surfactants on the physical properties of starch based edible films. *Food Research*
640 *International*, 39, 840–846.

641 Rosen, S.L. (1993). *Fundamental principles of polymeric materials*: 2nd edition; Wiley
642 Interscience, New York.

643 Shah, P., Bandopadhyay, S., & Bellare, J. (1995). Environmentally degradable starch filled
644 low density polyethylene. *Polymer Degradation and Stability*, 47 (2), 165–173.

645 Singh, J., Singh, N., & Saxena, S.K. (2002). Effect of fatty acids on the rheological
646 properties of corn and potato starch. *Journal of Food Engineering*, 52 (1), 9-16.

647 Vieira, M., Altenhofen, M., Oliveira L., & Masumi, M. (2011). Natural-based plasticizers
648 and biopolymer films: A review. *European Polymer Journal*, 47, 254–263.

649 Villalobos, R., Chanona, J., Hernández, P., Gutiérrez, G., & Chiralt, A. (2005). Gloss and
650 transparency of hydroxypropyl methylcellulose films containing surfactants as affected
651 by their microstructure. *Food Hydrocolloids*, 19, 53–61.

652 Villalobos, R., Hernández-Muñoz, P., & Chiralt, A. (2006). Effect of surfactants on water
653 sorption and barrier properties of hydroxypropyl methylcellulose films. *Food*
654 *Hydrocolloids*, 20, 502–509.

655 Wokadala, O. C., Ray, S. S. & Emmambux, M. N. (2012). Occurrence of amylose–lipid
656 complexes in teff and maize starch biphasic pastes. *Carbohydrate Polymers*, 90, 616–
657 622.

658 Zhong, F., Li, Y., Ibañes, A. M., Oh, M. H., McKenzie, K.S., & Shoemaker, C. (2009). The
659 effect of rice variety and starch isolation method on the pasting and rheological
660 properties of rice starch pastes. *Food Hydrocolloids*, 23, 406–414.

661 Zhong, Y., Li, Y. (2011). Effects of surfactants on the functional and structural properties
662 of kudzu (*Pueraria lobata*) starch/ascorbic acid films. *Carbohydrate Polymers*, 85,
663 622–628.

664 Ziani, K., Osés, J., Coma, V., Maté, J. I. (2008). Effect of the presence of glycerol and
665 Tween 20 on the chemical and physical properties of films based on chitosan with
666 different degree of deacetylation. *LWT - Food Science and Technology*, 41, 2159-2165.