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Additional Information

1	Effect of the citric acid addition on the properties and ageing time of starch-
2	polycaprolactone based films
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11	ABSTRACT
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13	
14	1. INTRODUCTION
15	The packaging films, made of synthetic polymers, cause serious ecological problems
16	due to their non-biodegradability. The increase of the consumers' conscience for the
17	environmental protection is causing the intensification in research about biodegradable
18	packaging materials (Fajardo, Martins, Fuciños, Pastrana, Teixeira, & Vicente, 2010).
19	The development of biodegradable materials designed to maintain or improve the shelf
20	life of food is very important today (García, Pinotti, Martino, & Zaritzky, 2004). In last
21	decade, many research about materials for food packaging has been brought out, these
22	research have had in common the work with economically viable natural sources.
23	Among these materials, the starch is known for its low cost, its renewability and its
24	processability by mean of traditional techniques (Yoon, Chough & Park, 2006;

25 Ghanbarzadeh, Almasi & Entezami, 2011).

Starch represents more than 60% of cereal grains produced in the world and it is 26 27 relatively easy separate it from other chemicals compounds (Lourdin, Della Valle & Colonna, 1995). It has great importance in the packaging industry compared to other 28 biopolymers by its technical characteristics. The starch based films are continuous, 29 homogeneous, odourless and colourless and its oxygen permeability is very low (Dole, 30 Joly, Espuche, Alric, & Gontard, 2004; Liu, 2005; Ortega-Toro, Jiménez, Talens & 31 Chiralt, 2014a). However, starch films have some disadvantages such as high 32 hydrophilicity (water sensitivity) and poor mechanical properties compared to 33 conventional synthetic polymers (Averous & Boquillon, 2004). Furthermore, the 34 temperature and relative humidity (RH) during storage, have an effect on the 35 crystallinity of the films and therefore on its physical properties (Rindlava, Hulleman, & 36 Gatenholma, 1997; Bergo, Sobral & Prison, 2010). For improve or maintain their 37 properties, is necessary to add plasticizers, fillers and cross-linking agents. The most 38 common plasticizer in the starch based films is the glycerol. This polyol reduces the 39 intermolecular attractive forces in the native starch and increases the flexibility and the 40 resistance to fracture (Mali, Grossmann, García, Martino & Zaritzky, 2005). 41

On the other hand, the polycaprolactone (PCL) is a biodegradable thermoplastic 42 polymer obtained by chemical synthesis from crude oil (Flieger, Kantorová, Prell, 43 Rezanka & Votruba, 2003). It has some interesting characteristics like it is waterproof 44 and has great flexibility (Averous, Moro, Dole, & Fringant, 2000). Several authors have 45 shown that blends of polycaprolactone and starch are readily biodegradable and 46 processible by conventional methods (Matzinos, Tserki, Gianikouris, Pavlidou, & 47 Panayiotou, 2002; Rosa, Lopes & Calil, 2005; Calil, Gaboardi, Bardi, Rezende & Rosa, 48 2007). However these mixtures are chemically incompatible and therefore have a phase 49

50 separation as occurs with other polymers (Annable, Fitton, Harris, Philips, & Williams,

51 1994; Mathew & Abraham, 2008; Ortega-Toro, Talens & Chiralt, 2015)

In previous works, Ortega-Toro et al. (2015) have observed the separation between 52 these two polymers by mean of Scanning Electron Microscopy and Atomic Force 53 Microscopy, concluding that it is necessary the incorporating of a compatibilizer 54 compound to improve the blends. One way to increase the number of interactions 55 between polymers is the addition of cross-linking agents. An appropriate compatibilizer 56 could be the citric acid (CA) because is a cheap organic acid and widely used in the 57 food industry as a cross-linking agent (Ghanbarzadeh, Almasi, & Entezami, 2010; 58 59 Ortega-Toro, Jiménez, Talens & Chiralt, 2014b). The CA use is interesting due to the three carboxyl groups in its structure which can interact with the hydroxyl groups of the 60 starch molecules through the formation of esters. According to other works, the 61 62 esterification reaction using polycarboxylic acids occur with the polymer's hydroxyl groups, at high temperatures (about 160 °C), through the formation of anhydride groups 63 (Reddy & Yang, 2010; Ortega-Toro et al., 2014b). This blend can improve the water 64 vapour barrier properties, due to the reduction of available hydroxyl groups (Borredon, 65 Bikiaris, Prinos & Panayiotou, 1997) through the formation of strong hydrogen bonds 66 67 (carboxyl- hydroxyl) in order to decrease the crystallization and retrogradation (Shi, Zhang, Liu, Han, Zhang, Chen, & Tian, 2007). However, the action mechanisms of the 68 CA are not clear and it can to act like compatibilizer, plasticizer and depolymerization 69 agent over different polymers according to processing conditions (Chabrat, Abdillahi, 70 Rouilly, & Rigal, 2012). 71

The aim of this work was to study the influence of the citric acid addition, and the effect of the storage time,on the structural, thermal and physicochemical properties of starchpolycaprolactone based films obtained by compression molding.

## 76 2. MATERIALS AND METHODS

#### 77 **2.1. Materials**

Corn starch was purchased from Roquette (Roquette Laisa España, Benifaió, Spain).
Glycerol was obtained from Panreac Química, S.A. (Castellar del Vallès, Barcelona,
Spain). Polycaprolactone (pellets ~3 mm, average M<sub>n</sub> 80.000, impurities <1.0% water)</li>
was provided by Fluka (Sigma–Aldrich Chemie, Steinheim, Germany). Citric acid was
provided by Fisher (Scientific Afora, Valencia, Spain). Phosphorus pentoxide (P<sub>2</sub>O<sub>5</sub>)
and Magnesium nitrate-6-hydrate (Mg(NO<sub>3</sub>)<sub>2</sub>) were obtained from Panreac Química,
S.A. (Castellar Vallés, Barcelona).

NaOH and phenolphthalein was provided by ???

## 86 2.2. Film preparation

Eleven formulations based on starch and PCL, with and without CA, were prepared. Starch:PCL ratios were 100:0, 90:10, 80:20, 70:30, 60:40 and 0:100, and starch:CA ratio was 1:0.01. A 30% of glycerol was added respect to the starch content. Firstly, native starch and glycerol were dispersed in water. Afterwards, PCL was added to the aqueous mixture of starch-glycerol. The blends were named as: S, S-CA, S90, S90-CA, S80, S80-CA, S70, S70-CA, S60, S60-CA and PCL. The controls used were starchglycerol (S) and pure PCL (PCL).

The formulations were gradually mixed on a two-roll mill (Model LRM-M-100, Labtech Engineering, Thailand) at 160 °C and 8 rpm for 30 min until a homogeneous paste sheet was obtained. The CA was added to the formulations 10 min before the homogenization. Paste sheet formed was conditioned at 25 °C and 53% RH using  $Mg(NO_3)_2$  oversaturated solutions for 48 h. The films were obtained by compression molding (Model LP20, Labtech Engineering, Thailand). Four grams of the paste were put onto steel sheets and preheated on the heating unit for 5 min. The films were performed at 160 °C for 2 min at 30 bars, followed by 6 min at 130 bars; thereafter a cooling cycle was applied for 3 min. The films obtained were conditioned at 25 °C and 53% RH for 1 and 5 weeks for the initial and final time characterization.

105

## 106 **2.3. Film characterization**

# 107 <u>2.3.1. Film thickness and extensibility</u>

The film thickness was measured with a Palmer digital micrometer (Palmer–Comecta, Spain, +/- 0.001 mm) at six random positions around the film. The extensibility was calculated by the relation between the surface and the weight of films with known dimensions. This parameter was expressed in  $cm^2/g$  of film.

112

## 113 <u>2.3.2. Structural properties</u>

114 The surface morphology of the samples, conditioned for 5 weeks at 25 °C and 53% RH , was analyzed using an Atomic Force Microscope (AFM) (Multimode 8, Bruker AXS, 115 Santa Barbara, USA) with a NanoScope<sup>®</sup> V controller electronics. Measurements were 116 taken using the PeakForce ONM<sup>®</sup> mode (Quantitative Nanomechanical Mapping). The 117 statistical parameters: average roughness (Ra: average of the absolute value of the 118 height deviations from a mean surface), root-mean-square roughness (Rq: root-mean-119 square average of height deviations taken from the mean data plane), and roughness 120 factor (r: ratio between the three-dimensional surface and two-dimensional area 121 projected onto the threshold plane), were calculated according to the method ASME 122

B46.1 (ASME, 1995). The DMT Modulus mode, derived from PeakForce QNM, was 123 124 considered. Three replicates for each formulation were taken to obtain these parameters. The microstructural analysis of cross-sections of films was carried out by using a 125 Scanning Electron Microscope (SEM) (JEOL JSM-5410, Japan). The film samples were 126 maintained in desiccators with P2O5 for 2 weeks at 25 °C and the measurements were 127 taken in duplicate for each formulation. Pieces about 0.5 cm<sup>2</sup> were cut from films and 128 mounted on copper stubs perpendicularly to their surface. Samples were gold coated 129 130 and observed, using an accelerating voltage of 10 kV.

131 A diffractometer (XRD, Bruker AXS/D8 Advance) was used for obtained X-ray 132 diffraction patterns. All samples (equilibrated for 1 and 5 weeks) were analyzed at 25 133 °C and 53% RH, between  $2\theta$ : 5° and 30° using K $\alpha$  Cu radiation ( $\lambda$ : 1.542 Å), 40 kV and 134 40 mA with a step size of 0.05°. For this analysis, the samples were cut into 4 cm<sup>2</sup>.

135

## 136 <u>2.3.3. Thermal properties</u>

A Differential Scanning Calorimeter DSC 1 Star<sup>e</sup> System (Mettler-Toledo Inc., 137 Switzerland) was used to analyze the thermal properties. Weighted amounts of samples 138 were placed into aluminum pans, sealed and drilled to promote moisture loss during the 139 heating. The thermograms were obtained using a triple scan. First, a scan from -80 °C to 140 160 °C at a rate of 50 °C/min was used for obtaining the PCL melting temperature ( $T_m$ ) 141 and to eliminate the bonded water in the film. The initial and final weights of the pans 142 were registered to assess water loss during the first heating Then, the temperature was 143 lowered to -80 °C at a rate of 50 °C/min. In this cooling step the PCL crystallization 144 temperature (T<sub>c</sub>) was obtained. Finally, samples were heated to 160 °C at a rate of 20 145 °C/min in order to analyze the starch glass transition and the second melting 146 temperature of PCL. 147

# 149 <u>2.3.4.</u> Physicochemical properties

150

The film water content was determined conditioning the samples at 53% RH and dried for 24 h at 60 °C using a convection oven (J.P. Selecta, S.A. Barcelona, España). Afterwards, the samples wereplaced in a desiccator at 25 °C with  $P_2O_5$  for 2 weeks. This assay was realized for triplicate.

The film solubility was determined holding the sample in bidestilled water for 48 h. The 155 film:water ratio was 1:10. These samples were transferred to a convection oven (J.P. 156 157 Selecta, S.A., Barcelona, Spain) for 24 h at 60 °C to remove free water and then were transferred to a dessicator with P<sub>2</sub>O<sub>5</sub> at 25 °C for 2 weeks to remove the linked water. 158 The initial and final weights were registered for calculating the solubility in water. 159 160 Three replicates were analyzed from each formulation Bonded citric acid in the film, was determined using the water solution of the solubility assay.. Free CA was assumed 161 to dissolve in the aqueous phase and was determined by titration of an aliquot of the 162 solution with NaOH (0.1 N), using phenolphthalein as indicator. Three replicates were 163 considered for each formulation. 164

An universal test machine (TA.XTplus model, Stable Micro Systems, Haslemere, England) was used to determine the tensile strength (TS), the elastic modulus (EM), and the elongation (E) of the films, according to the ASTM standard method D882 (ASTM, 2001). 2.5 cm wide and 5 cm long equilibrated samples were mounted in the filmextension grips of the testing machine and stretched at 50 mm min<sup>-1</sup> until breaking. Ten replicates were analyzed from each formulation.

The ASTM E96-95 (ASTM, 1995) gravimetric method was used for determining the
Water Vapour Permeability (WVP) of the films, considering the modification proposed

by McHugh, Avena-Bustillos & Krochta (1993).. Distilled water was placed in Payne 173 174 permeability cups (3.5 cm diameter, Elcometer SPRL, Hermelle/s Argenteau, Belgium) to expose the film to 100% RH on one side. Each cup was placed in a cabinet 175 equilibrated at 25 °C and 53% RH, with a fan placed on the top of the cup in order to 176 reduce the resistance to water vapor transport, thus avoiding the stagnant layer effect in 177 this exposed side of the film. The relative humidity of the cabinet (53%) was held 178 179 constant using  $Mg(NO_3)_2$  oversaturated solutions. The cups were weighed periodically (0.0001 g) and the water vapor transmission (WVTR) was determined from the slope 180 obtained from the regression analysis of weight loss data versus time. From WVTR 181 182 data, the vapor pressure on the film's inner surface  $(p_2)$  was obtained with eq. (1), proposed by McHugh et al. (1993) to correct the effect of concentration gradients. 183

- 184
- 185

$$WVTR = \frac{P \cdot D \cdot L_n \left[ P - p_2 \setminus P - p_1 \right]}{R \cdot T \cdot \Delta z} \tag{1}$$

186

where P, total pressure (atm); D, diffusivity of water through air at 25 °C (m<sup>2</sup>/s); R, gas law constant (82.057 x  $10^{-3}$  m<sup>3</sup> atm kmol<sup>-1</sup> K<sup>-1</sup>); T, absolute temperature (K);  $\Delta z$ , mean stagnant air gap height (m), considering the initial and final z value; p<sub>1</sub>, water vapor pressure on the solution surface (atm); and p<sub>2</sub>, corrected water vapor pressure on the film's inner surface (atm). Water vapor permeance was calculated using eq. (2) as a function of p<sub>2</sub> and p<sub>3</sub> (pressure on the film's outer surface in the cabinet).

193

$$permeance = \frac{WVTR}{p_2 - p_3} \tag{2}$$

195

194

The permeability was calculated by multiplying the permeance by film thickness. Thisstudy was carried out for triplicate.

The Oxygen Permeability (OP) of the films was determined using an OX-TRAN Model 2/21 ML (Mocon Lippke, Neuwied, Germany) according to the ASTM Standard Method D3985–05 (2010). Three samples conditioned at 53% RH and 25 °C for each formulation were studied. The transmission values were determined every 20 min until to reach the equilibrium. The area used for the tests was 50 cm<sup>2</sup>. The film thickness was considered in all cases for obtaining the OP.

The transparency, using the Kubelka-Munk theory for multiple scattering (Hutchings, 204 1999) and the gloss, at 85° angle from normal to the surface, according to the ASTM 205 standard D523 method (ASTM, 1999) were determined on the films. The surface 206 reflectance spectrum was determined from 400 to 700 nm with a spectro-colorimeter 207 CM- 3600d (Minolta Co., Tokyo, Japan) on both a white and a black background. As 208 light passes through the film, it is partially absorbed and scattered, which is quantified 209 210 by the absorption (K) and the scattering (S) coefficients. Internal transmittance (Ti) of films was determined using eq. (3). 211

- 212
- $T_i = \sqrt{(a-R_0)^2 b^2}$

(3)

214

213

where  $R_0$  is the reflectance of the film on an ideal black background. The parameters *a* and *b* were calculated by eqs. (4) and (5).

217

218 
$$a = \frac{1}{2} \left( R + \frac{R_0 - R + R_g}{R_0 R_g} \right)$$
(4)

219

 $b = \sqrt{a^2 - 1} \tag{5}$ 

where R is the reflectance of the sample layer backed by a known reflectance  $R_g$ . Three replicates were used for each formulation. For the analysis was considered a wavelength of 650 nm.

The gloss was determinate using a flat surface gloss meter (Multi.Gloss 268, Minolta, Germany). Three films of each formulation were considered taken three measurements in each sample. All results are expressed as gloss units (GU), relative to a highly polished surface of black glass standard with a value near to 100 GU.

229

#### 230 <u>2.3.5. Statistical analysis</u>

Statgraphics Plus for Windows 5.1 (Manugistics Corp., Rockville, MD) was used for carrying out statistical analyses of data through analysis of variance (ANOVA). Fisher's least significant difference (LSD) was used at the 95% confidence level.

234

#### 235 **3. RESULTS**

## 236 **3.1. Thickness and extensibility**

According to Chabrat *et al.*, 2012, in blends of starch and PCL processed at high temperature, the use of citric acid could act like a compatibilizer (esterification, hydrogen bonds), as a starch plasticizer and/or as a depolymerization agent for starch and PCL.

Table 1 shows the thickness and extensibility of the studied films. The PCL films had lower thickness, and greater extensibility (area per mass unit in the compression molding) than the other formulations. By contrast, the thickness tends to increase when increase the amount of starch in the matrix, due to their reduced ability to flow. On the other hand, the plasticizer effect of citric acid was masked with the high plasticity of PCL.

## 248 **3.2. Structural analysis**

Atomic Force Microscopy on mode PeakForce QNM was used to acquire the images of 249 the Figure 1. For the study of the nano-mechanical properties of the material the DMT 250 modulus was considered. The control formulations (S and PCL) did not presented 251 marked changes in its structure, showing more homogeneous colors than the starch-PCL 252 253 blends. When the PCL was added to the starch matrix, the heterogeneity of the material was higher and the DMT modulus values were greater than control formulations. The 254 blends exhibit dark and light zones corresponding to amorphous and crystalline 255 256 fragments (Baker, Miles & Helbert, 2001; Ortega-Toro et al., 2014b). Regarding the effect of citric acid, in the S formulation, a hardening on the surface of films was 257 observed when citric acid was added. The same effect was observed for blends that had 258 259 high content of starch. This phenomenon suggests an effect of cross-linking or an increasing in the hydrogen bonds in the structure of polymeric matrix (Ortega-Toro et 260 al., 2014b), which is masked when the content of PCL is higher. 261

Figure 2 shows SEM micrographs of the cross-section of the studied films, with or 262 without citric acid, conditioned at 25 °C and 53% RH for one week. S and PCL 263 formulations have a continuous and homogeneous structure; the PCL showed 264 deformations caused by the cryo-fracture with liquid nitrogen. When the PCL was 265 added to the starch matrix, the films were more heterogeneous, manifesting the 266 immiscibility of the polymers. In the formulations with less PCL (S90-S80) the 267 distribution of PCL was more effective. When the proportion of PCL was increased 268 (S70 and S60) zones rich in PCL and zones rich in starch were observed. 269

The addition of citric acid provoked changes in the polymers interaction's. The S formulation has a continuous matrix with some cracks when the CA was added (S-CA).

This suggests the formation of a matrix with less deformation capacity. However, the 272 273 S-CA formulation was more homogeneous than the S formulation, caused by citric acid which can promotes the fragmentation and dissolution of the native starch granules (Yu, 274 Wang, & Ma, 2005; Ma, Chang, Yu, & Stumborg, 2009). On the other hand, in the 275 formulations with less PCL (S80-CA and S90-CA), the CA promotes a change in the 276 PCL structure doing the PCL more thin and with better distribution. Similar results were 277 reported by Chabrat et al., 2012, in blends of starch and PCL in presence of citric acid. 278 In the formulations S70-CA and S60-CA the changes were less marked because the 279 high amount of PCL could mask the citric acid effect. These phenomena may be due to 280 281 the depolymerization effect on the PCL; this could induce to better affinity between the polymers (Reddy & Yang, 2010; Chabrat et al., 2012). 282

Figure 3 shows the X-ray diffraction patterns of the films stored for 1 and 5 weeks at 283 284 controlled conditions. In all formulations which contain starch, the typical crystalline forms of amylose V type at 20: 7°, 14° and 20° were observed (Lesmes, Cohen, Shener, 285 & Shimoni, 2009). When the amount of PCL increased the peaks were less marked and 286 the peak at 20: 14° was the only observed. Thinnest peaks and more intense were 287 observed in the PCL diffractogram due to crystalline zones biggest at 20: 22° and 24° 288 289 (Mark, 1999). On the other hand, the amorphous zones were less pronounced with the PCL increasing. In the formulations containing PCL, the peaks intensity were smaller 290 with the storage time because of the reduction of the polymeric matrix crystallinity. 291 Regarding the citric acid effect, the displacement of peaks were observed, which 292 suggest polymorphisms in the polymeric matrix (Reddy & Yang, 2010), and a decrease 293 in the peaks intensity with the citric acid presence was observed on both the initial and 294 the final time of storage. These phenomena are interesting because this could suggest 295 the partial inhibition of starch retrogradation. These results are in accordance with other 296

authors demonstrating that the CA can protect the thermoplastic starch from
retrogradation (Yu *et al.*, 2005; Ortega-Toro *et al.*, 2014b). During the process, the CA
penetrates the starch granule and replaces partially the intermolecular hydrogen bonds
of starch. This causes the crystallinity reduction of the thermoplastic starch (Xie, Liu, &
Cui 2006; Ma *et al.*, 2009)

302

#### 303 **3.3. Thermal analysis**

Table 2 shows the thermal properties of the studied films conditioned for 1 and 5 weeks. 304 Two heating scans and one cooling scan were performed. In the first heating scan the 305 melting temperature (Tm) of PCL was determined as the peak temperature of the 306 endotherm. In the first scan, the Tm and  $\Delta$ Hm of the initial and the final storage time 307 were considered. During cooling, the crystallization temperature (Tc) of PCL was 308 309 determined as the peak temperature of the exotherm. Finally, during the second heating scan another fusion temperature value was taken (PCL crystallized during cooling) and 310 311 the glass transition temperature (Tg) of starch at the midpoint was determined. The 312 values of enthalpy of fusion ( $\Delta$ Hm) and crystallization ( $\Delta$ Hc) are also shown in Table 2. The Tg of starch decreased significantly (p < 0.05) with the PCL incorporation (S90). 313 This decrease only was promoted with the addition of 10% of PCL. When the amount 314 was bigger than 10%, the effect was not progressive and the values were almost 315 constants. On the other hand, the  $\Delta Cp$  increased with the PCL addition in concordance 316 with a decrease in average molecular weight or a higher molecular mobility in the 317 polymeric matrix. 318

Moreover, the Tc values of the PCL was about 12 °C with  $\Delta$ Hc about 52 J/g PCL, and among the formulations were not observed significant differences. Besides, the Tm value of PCL in the first scan was approximately 63°C and  $\Delta$ Hm was observed about

72 J/g of PCL. Similar results have been reported by other authors (Averous et al., 322 323 2000; Matzinos, et al., 2002; Kweon, Kawasaki, Nakayama & Aiba, 2003). With the storage time, a slight increase in Tm and  $\Delta$ Hm were observed, which suggests that it is 324 necessary an amount bigger of energy for melt the crystals formed during the storage 325 and the interactions among the PCL molecules could be stronger. Additionally, the Tm 326 and  $\Delta$ Hm on the second scan were lower than the values obtained in the first scan, 327 according with other authors for pure PCL (Koenig & Huang, 1995); the  $\Delta$ Hm values 328 on the second scan were very close to  $\Delta$ Hc values, this suggests that the crystals formed 329 during the cooling cycle can melt completely during the second heating cycle (Campos, 330 331 Marconcini, Martins-Franchetti & Mattoso, 2012).

Regarding the citric acid, the principal effects were on the Tg and  $\Delta$ Cp of starch. The Tg 332 value of starch was decreased significantly (p < 0.05) when the CA was added in the S 333 334 formulation; and also decreased the Tg values in the films containing PCL until about 101 °C, remaining constant among these formulations. Meanwhile, the  $\Delta$ Cp values were 335 lower with the CA addition in every case; perhaps the interactions among starch and 336 PCL were more effective with the citric acid at least when the PCL was in less amount, 337 as was observed in the SEM micrographs. Furthermore it is possible to observe that the 338 Tm and  $\Delta$ Hc decreased slightly when the CA was added, this may be caused by the 339 decrease in the interactions among the starch molecules, suggesting a possible 340 hydrolysis or a plasticizer effect. 341

342

## 343 **3.4. Physicochemical properties**

Figure 4 shows the tensile properties of studied films stored under controlled conditions for 1 and 5 weeks. When the starch is blend with PCL, EM is bigger than the control formulations (S and PCL) except for the S90 formulation, where EM is significantly smaller (p < 0.05). The blend between starch and PCL could improve the elastic modulus of the resulting material (Avella, Errico, Laurienzo, Martuscelli, Raimo & Rimedio, 2000).In general, TS decreased when the PCL was added, and the elongation at break point was smaller than the S formulation except in the S90. These results, in accordance with the thermal properties and the structural properties, suggest that the interaction between starch and PCL is better when the PCL is presented in low proportion in the polymer blend. .

Figure 5 shows the typical stress-strain curves of the PCL films at initial storage time. Pure PCL films presented permanent deformation after the first break. The tensile strength (18.17  $\pm$  1.06 MPa) and deformation (13  $\pm$  4%) at the yield point were determined. Pure PCL is a ductile polymer so it can be deformed until 1100% in agreement with other authors (Ishiaku, Pang, Lee, & Mohd, 2002). In the present work, the universal test machine used was unable to register the maximum deformation (only 150% was possible).

The CA effect caused in elongation was especially remarkable; when the citric acid was added, the elongation had a significant increase (p < 0.05) for S90-CA and S80-CA but decreased for the S formulation, which suggested a plasticizer effect of the citric acid unbound (Ghanbarzadeh *et al.*, 2011). Regarding EM and TS, the citric acid promoted the increased in these parameters for the S formulation, especially with the storage time, and a decreased in formulations containing less PCL, although in most cases the changes were slight.

On the other hand, the formulations showed two trends with the storage time, when the PCL was in low proportion, the EM and TS parameters increased and the elongation decreased. The changes were not marked when the amount of PCL was high (S70, S70371 CA, S60 and S60-CA); perhaps, high amount of PCL limited the molecular
372 reorganization and therefore the retrogradation during storage.

Table 3 shows the values and standard deviation of water content, film solubility and 373 bonded citric acid of studied films conditioned under controlled conditions at initial and 374 final storage time. The water content (Xw) decreased with the increasing of PCL due to 375 its hydrophobic character. In general, the formulations with citric acid presented 376 significantly less (p < 0.05) water content than the CA-free films. This phenomenon 377 indicates a possible esterification of the hydroxyl groups of starch; during the process of 378 films formation (blend in roll mills and compression molding), the temperature used 379 380 was high (160 °C) and therefore the matrix was in absence of water and, the citric acid had the adequate conditions for take the anhydrous form, it could lead to esterification 381 reactions described by Reddy & Yang (2010). Cross-linking of starch reinforces the 382 383 intermolecular binding by introducing covalent bonds, which strengthen natural intermolecular hydrogen bonds, so as to improve the water uptake resistance (Yu et al., 384 2005; Krumova, Lopez, Benavente, Mijangos & Perena, 2000; Ghanbarzadeh et al., 385 2011). Besides, a significant increased (p < 0.05) in Xw during the storage time was 386 observed in every case, except in S70 and pure PCL. 387

Regarding the film solubility, a significant decrease (p < 0.05) with the PCL addition 388 was observed compared with the S formulation; however, were not observed differences 389 among the formulations containing PCL. On the other hand, the film solubility 390 increased when CA was added, except for the formulations with higher PCL content 391 (S60 and S60-CA). The increase in this parameter suggests partial hydrolysis of starch 392 molecules caused by the presence of citric acid, resulting in the increase of linear and 393 smaller fragments of starch compared to the native starch (Carvalho, Zambon, da Silva 394 Curvelo, & Gandini 2005; Wang, Yu, Chang, & Ma, 2007; Ortega-Toro et al., 2014b). 395

Other works reported similar results for starch-polyvinyl based films in presence of citric acid (Yoon *et al.*, 2006), as also in modified starch-based films in combination with polyvinyl alcohol and citric acid like plasticizer (Lee, Youn, Yun & Yoon, 2007; Yun, Wee, Byun & Yoon, 2008). Other authors reported similar results for thermoplastic starch co-plasticized with citric acid/glycerol (Shi *et al.*, 2007).

The values of bonded CA ratio with respect to the amount in the film did not present marked differences among the formulations. Considering that the proportions between the starch and CA were equal, maybe the CA does not act directly with the PCL and/or the PCL presence does not affect the reactions between the CA and the starch molecules.

Table 4 presents the water vapor permeability (WVP) and oxygen permeability (OP) of 406 studied films conditioned at 53% RH and 25 °C for 1 and 5 weeks. In food packaging 407 applications, the WVP should be as low as possible. The addition of PCL promoted a 408 significant decreased (p < 0.05) in the WVP of studied films due its hydrophobic 409 410 character. The effect of PCL was more market than the citric acid effect, however, the 411 addition of CA promotes a slight decreased in the values of this parameter. The CA trend has been observed for other authors (Ma et al., 2009; Reddy & Yang, 2010; 412 Ghanbarzadeh et al., 2011; Olivato, Grossmann, Yamashita, Eiras, & Pessan, 2012), 413 which could attribute to the hydrophilic OH groups substitution with hydrophobic ester 414 groups, and probably introduced a tortuous path for water molecules to pass through 415 with hydrophobic ester groups. Significant differences were not observed through the 416 417 storage time.

The OP values for S and PCL formulations were not presented because the S formulation had very low OP and the pure PCL presented high OP and it was out of range of the equipment used (OX-TRAN Model 2/21 ML Mocon). The values of OP 421 increased significantly (p < 0.05) with the PCL, and the CA addition did not present a 422 considerable effect. Regarding the storage time, the changes were very slight.

Table 5 presents the values and standard deviation of roughness parameters, internal 423 transmittance at 650 nm and gloss at 85° of studied films conditioned under controlled 424 conditions for 1 and 5 weeks. In general, the roughness parameters did not present 425 marked differences, and showed a slight trend to decrease with the PCL addition. As 426 expected, the pure PCL showed the less roughness (Ra and Rq) than the other 427 formulations. On the other hand, a slight trend to decrease with the CA addition was 428 observed because the acidity of citric acid promotes the fragmentation and dissolution 429 of starch granules (Yu et al., 2005; Wang, Zhang, Han, & Bai, 2009; Chabrat et al., 430 2012). In the analysis of this parameter it is necessary to take into account that the 431 compression molding could affect the surface roughness and mask the effect caused by 432 433 the components of the polymeric matrix.

The gloss of the films was related with the roughness (Sánchez-González, Cháfer, Chiralt, González-Martínez, 2010) and therefore the obtaining process could have an influence in it, in general, the gloss is proportional to the smoothness of the material. The pure PCL and S90-CA have showed the higher values of gloss and the formulation S80-CA has showed the least value of gloss, according to Ra and Rq parameters obtained by AFM.

440 On the other hand, the internal transmittance has trended to decrease with the PCL 441 incorporation, which indicated higher heterogeneity and therefore with a higher opacity. 442 Also is possible to observe that the incorporation of CA to S90 promoted a higher 443 transmittance and a contrary effect was presented for the other blends. This suggests a 444 highest effect of CA when the PCL is less in the formulation, such as occur in the tensil 445 properties and thermal properties. Is possible to deduce that the interaction between the starch and PCL is stronger when the PCL is least in the formulation, and the CA could interact with these polymers and improve the blends although its effect could be masked when the PCL have high proportion.

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## 451 **4. CONCLUSIONS**

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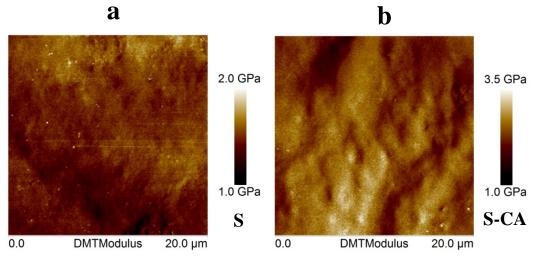
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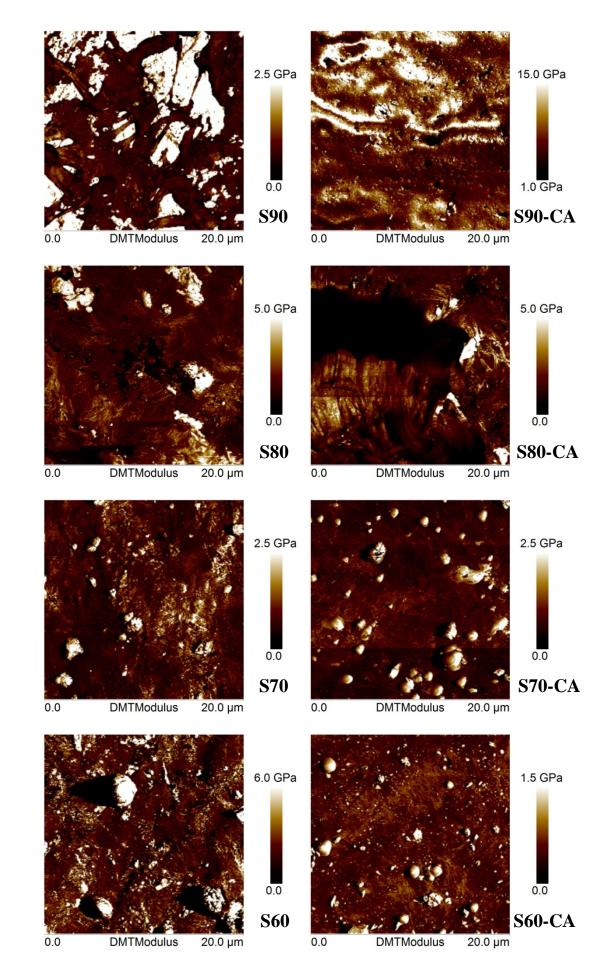
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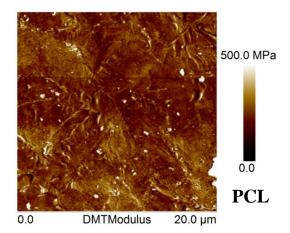
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609	Figure captions
610	Figure 1 AFM images of starch:PCL films without (a) and with (b) citric acid at initial time of storage
611	at 53% relative humidity and 25 °C.
612	Figure 2SEM micrographs of starch:PCL films without (a) and with (b) citric acid at initial time of
613	storage at 53% relative humidity and 25 °C.
614	Figure 3X-Ray diffraction patterns of starch:PCL films without (a) and with (b) citric acid at initial and
615	final time of storage at 53% relative humidity and 25 °C.
616	Figure 4 Mean values of elastic modulus and stress and strain at break for the different films stored for 1
617	(black bars) and 5 (grey bars) weeks at 53% relative humidity and 25 °C. LSD intervals (95% confidence
618	level) for each mean value are shown.
619	Figure 5Typical stress-strain curves of PCL at initial time.

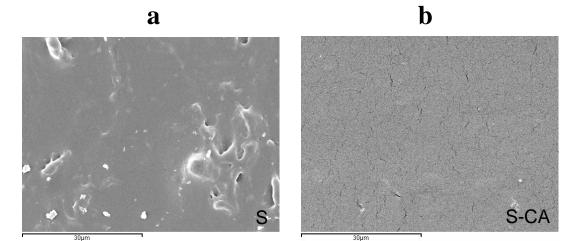


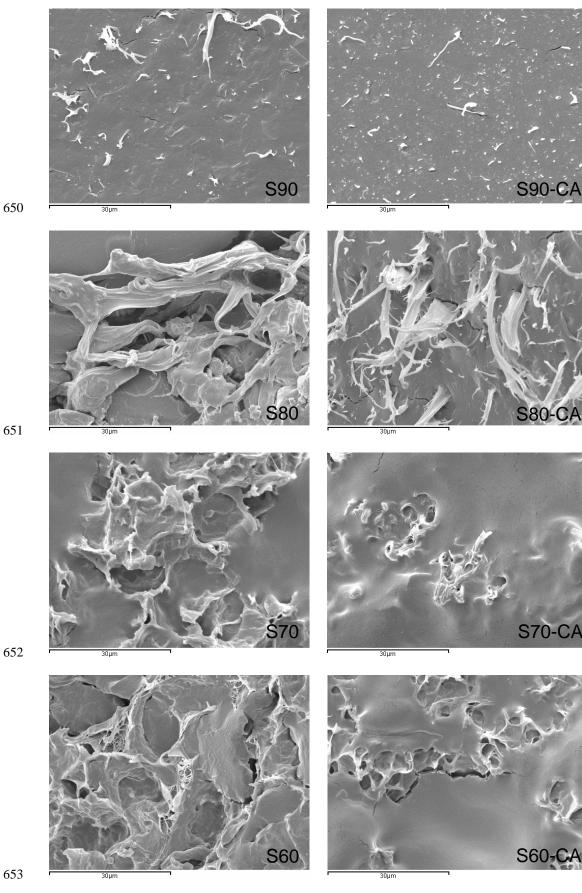


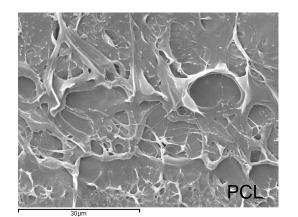




- 640 Figure 1. AFM images of starch:PCL films without (a) and with (b) citric acid at initial
- time of storage at 53% relative humidity and 25 °C.







- Figure 2. -SEM micrographs of of starch:PCL films without (a) and with (b) citric acid
- at initial time of storage at 53% relative humidity and 25 °C.

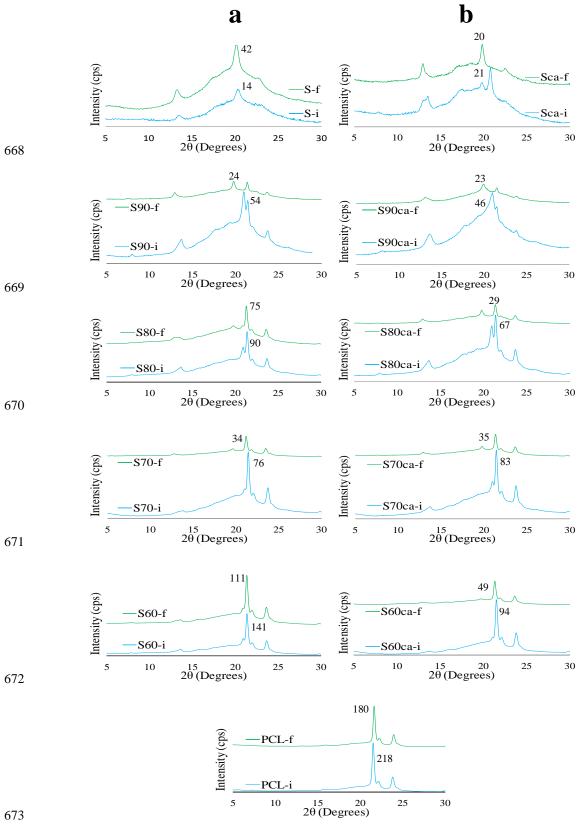
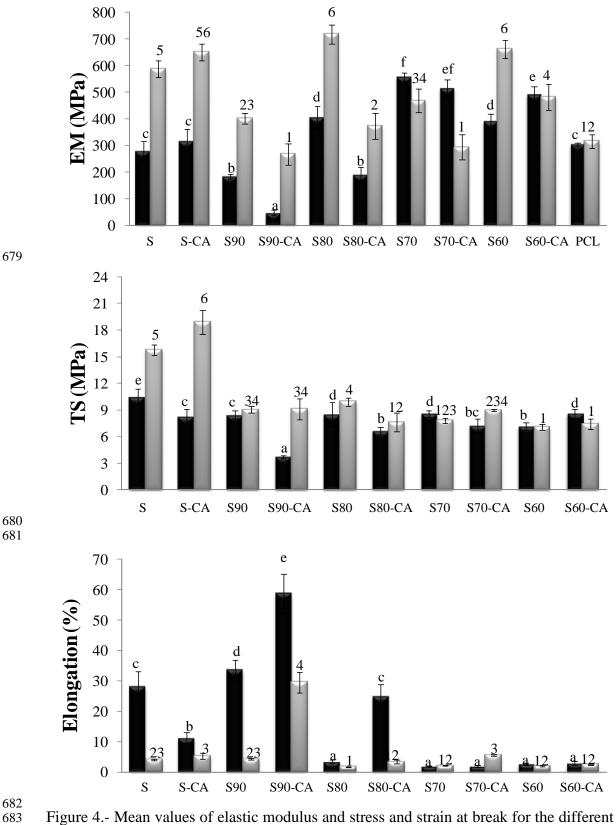
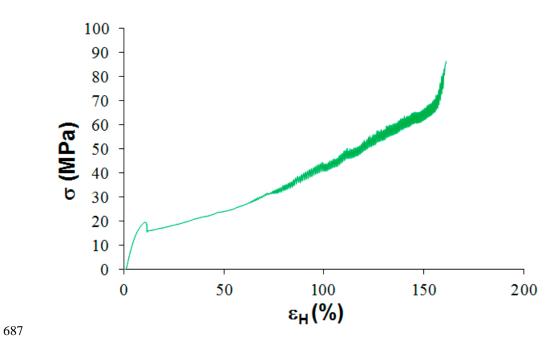


Figure 3. -X-Ray diffraction patterns of starch:PCL films without (a) and with (b) citric 

acid at initial and final time of storage at 53% relative humidity and 25 °C. 



films stored for 1 (black bars) and 5 (grey bars) weeks at 53% relative humidity and 25 °C. LSD intervals (95% confidence level) for each mean value are shown. 



688 Figure 5.-Typical stress-strain curves of PCL at initial time.

689	Table 1. Mean values and standard deviation of thickness and extensibility of the different films stored at 53% relative humidity and 25 °C.
690	

Films	Thickness (µm)	Extensibility (cm <sup>2</sup> /g)		
S	268 (22) <sup>e</sup>	23.9 (1.1) <sup>a</sup>		
S-CA	217 (22) <sup>c</sup>	32 (4) <sup>bc</sup>		
S90	232 (17) <sup>cd</sup>	28 (2) <sup>abc</sup>		
S90-CA	293 (27) <sup>f</sup>	23.7 (0.7) <sup>a</sup>		
<b>S</b> 80	221 (9) <sup>cd</sup>	32 (2) <sup>c</sup>		
S80-CA	239 (17) <sup>d</sup>	27 (2) <sup>ab</sup>		
<b>S</b> 70	224 (9) <sup>cd</sup>	31.1 (1.1) <sup>bc</sup>		
S70-CA	227 (11) <sup>cd</sup>	29.8 (0.6) <sup>bc</sup>		
S60	168 (13) <sup>b</sup>	45 (6) <sup>d</sup>		
S60-CA	172 (14) <sup>b</sup>	41 (2) <sup>d</sup>		
PCL	149 (17) <sup>a</sup>	57 (9) <sup>e</sup>		

692 Different superscript letters within the same column indicate significant differences among formulations (p < 0.05).

706	Table 2. Mean values and standard deviation of thermal properties of the starch: PCL films equilibrated at 53% relative humidity and 25 °C	
707		

	Glass transition		Crystallization		Fusion on the first scan				Fusion on the second scan	
Films					Initial		Final		Fusion on the second scan	
1 111115	Tg midpoint	ΔCp	Tc peak	ΔHc (J/g PCL)	Tm peak	ΔHm (J/g PCL)	Tm peak	ΔHm (J/g PCL)	Tm peak	ΔHm (J/g PCL)
S	126.0 (0.3) <sup>d</sup>	0.112 (0.002) <sup>b</sup>								
S-CA	113.0 (0.7) <sup>b</sup>	0.054 (0.009) <sup>a</sup>								
S90	114.5 (0.7) <sup>c</sup>	0.17 (0.02) <sup>d</sup>	12.0 (0.2) <sup>a</sup>	51.4 (0.9) <sup>bc</sup>	62.4 (0.7) <sup>bc</sup>	72.4 (0.4) <sup>a</sup>	63.9 (0.6) <sup>b</sup>	81.9 (1.0) <sup>ab</sup>	55.18 (0.05) <sup>bcd</sup>	51.8 (1.3) <sup>a</sup>
S90-CA	101.6 (0.2) <sup>a</sup>	0.112 (0.008) <sup>b</sup>	12.10 (0.14) <sup>a</sup>	49.2 (0.9)a	61.1 (0.9) <sup>a</sup>	72.1 (0.5) <sup>a</sup>	63.84 (0.08) <sup>b</sup>	79.7 (0.5) <sup>a</sup>	54.0 (0.2) <sup>a</sup>	50 (2) <sup>a</sup>
<b>S80</b>	114.31 (0.05) <sup>c</sup>	0.1280 (0.0014) <sup>bc</sup>	12.5 (0.6) <sup>ab</sup>	52.3 (0.3) <sup>c</sup>	62.9 (0.6) <sup>c</sup>	72.3 (1.3) <sup>a</sup>	63.29 (0.11) <sup>ab</sup>	82.3 (0.7) <sup>b</sup>	55.4 (0.3) <sup>d</sup>	51.2 (1.3) <sup>a</sup>
S80-CA	101.2 (0.2) <sup>a</sup>	0.131 (0.010) <sup>bc</sup>	12.6 (0.4) <sup>ab</sup>	49.3 (0.9) <sup>a</sup>	60.97 (0.12) <sup>a</sup>	72.2 (0.4) <sup>a</sup>	63.92 (0.11) <sup>b</sup>	81.0 (1.5) <sup>ab</sup>	54.11 (0.05) <sup>a</sup>	50.5 (0.9) <sup>a</sup>
<b>S</b> 70	114.35 (0.13) <sup>c</sup>	0.139 (0.013) <sup>c</sup>	12.3 (0.3) <sup>ab</sup>	52.0 (0.6) <sup>c</sup>	62.32 (0.62) <sup>bc</sup>	72.1 (0.3) <sup>a</sup>	63.32 (0.04) <sup>ab</sup>	81.7 (0.9) <sup>ab</sup>	54.9 (0.2) <sup>bc</sup>	51.4 (1.4) <sup>a</sup>
S70-CA	101.53 (0.13) <sup>a</sup>	0.052 (0.014) <sup>a</sup>	12.95 (0.07) <sup>b</sup>	50.7 (1.1) <sup>abc</sup>	61.55 (0.03) <sup>ab</sup>	72.4 (0.4) <sup>a</sup>	63.39 (0.11) <sup>ab</sup>	79.9 (0.6) <sup>a</sup>	54.8 (0.4) <sup>b</sup>	49.9 (1.1) <sup>a</sup>
S60	114.8 (0.2) <sup>c</sup>	0.126 (0.004) <sup>bc</sup>	12.33 (0.07) <sup>ab</sup>	52.2 (0.8) <sup>c</sup>	63.0 (0.2) <sup>c</sup>	72.5 (0.6) <sup>a</sup>	63.3 (0.2) <sup>ab</sup>	81.9 (1.0) <sup>ab</sup>	55.13 (0.04) <sup>bcd</sup>	51.0 (1.3) <sup>a</sup>
S60-CA	100.9 (0.5) <sup>a</sup>	0.041 (0.004) <sup>a</sup>	12.3 (0.3) <sup>ab</sup>	50.3 (0.2) <sup>ab</sup>	61.51 (0.02) <sup>ab</sup>	72.2 (0.4) <sup>a</sup>	63.07 (0.04) <sup>a</sup>	79.8 (0.4) <sup>a</sup>	54.88 (0.14) <sup>b</sup>	49.7 (1.0) <sup>a</sup>
PCL			12.6 (0.6) <sup>ab</sup>	52.4 (0.3) <sup>c</sup>	63.5 (0.6) <sup>a</sup>	72.8 (1.4) <sup>a</sup>	63.7 (0.6) <sup>a</sup>	82.2 (1.5) <sup>b</sup>	55.4 (0.4) <sup>cd</sup>	51.0 (1.2) <sup>a</sup>

Different superscript letters within the same column indicate significant differences among formulations (p < 0.05). 710 711 712 713 714

Table 3. Mean values and standard deviation of water content (g water/g dried film), film solubility (g solubilised film/g initial dried film) and bonded citric acid (ratio with respect to amount in the film) of the different films stored at 53% relative humidity and 25 °C.

Eilma	Х	W	Film solu	Bonded	
Films	Initial	Final	Initial	Final	citric acid
S	0.0613 (0.0006) <sup>h1</sup>	0.079 (0.004) <sup>g2</sup>	0.19 (0.07) <sup>d1</sup>	0.24 (0.05) <sup>d1</sup>	
S-CA	0.059 (0.002) <sup>gh1</sup>	0.076 (0.002) <sup>fg2</sup>	0.35 (0.03) <sup>f1</sup>	0.371 (0.012) <sup>f1</sup>	0.80 (0.03) <sup>a</sup>
S90	0.0664 (0.0004) <sup>i1</sup>	0.074 (0.003) <sup>ef2</sup>	$0.120 (0.009)^{bc1}$	0.138 (0.002) <sup>b2</sup>	
S90-CA	0.0543 (0.0007) <sup>ef1</sup>	0.0713 (0.0005) <sup>e2</sup>	$0.328 \ (0.005)^{f1}$	0.331 (0.005) <sup>e1</sup>	0.84(0.03) <sup>a</sup>
S80	0.0535 (0.0008) <sup>e1</sup>	0.061 (0.002) <sup>d2</sup>	0.132 (0.003) <sup>bc1</sup>	0.137 (0.013) <sup>b1</sup>	
S80-CA	0.056 (0.003) <sup>ef1</sup>	0.06161 (0.00015) <sup>d2</sup>	0.362 (0.013) <sup>f1</sup>	$0.32 (0.02)^{e^2}$	0.86(0.03) <sup>a</sup>
S70	0.057 (0.003) <sup>fg1</sup>	0.063 (0.004) <sup>d1</sup>	0.099 (0.003) <sup>b1</sup>	0.106 (0.006) <sup>b1</sup>	
S70-CA	0.047 (0.003) <sup>d1</sup>	$0.0578 (0.005)^{c^2}$	$0.24 (0.02)^{e_1}$	$0.30(0.02)^{e^2}$	0.81(0.04) <sup>a</sup>
S60	$0.0409 (0.0006)^{c1}$	0.046 (0.002) <sup>b2</sup>	0.13 (0.03) <sup>bc1</sup>	0.13 (0.02) <sup>b1</sup>	
S60-CA	0.03331 (0.00014) <sup>b1</sup>	0.0458 (0.0014) <sup>b2</sup>	0.16 (0.02) <sup>cd1</sup>	0.19 (0.02) <sup>c1</sup>	$0.83(0.04)^{a}$
PCL	$0.003 (0.002)^{a1}$	$0.0021 (0.0005)^{a1}$	0.00022 (0.00011) <sup>a1</sup>	0.0004 (0.0002) <sup>a1</sup>	

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729 Different superscript numbers within the same row indicate significant differences due to storage time (p < 0.05).

Table 4. Mean values and standard deviation of water vapour permeability (WVP) and oxygen permeability (OP) of the different films at 1 (Initial time) and 5 (Final time) weeks of storage at 53% relative humidity and 25 °C.

Films	WVP (g∙mm∙	kPa⁻¹⋅h⁻¹⋅m⁻²)	OP · 10 <sup>14</sup> (cm <sup>3</sup> ·m <sup>-1</sup> ·s <sup>-1</sup> ·Pa <sup>-1</sup> )				
FIIIIS	Initial	Final	Initial	Final			
S	18.1 (1.4) <sup>h1</sup>	16 (2) <sup>e1</sup>	< D.L.	< D.L.			
S-CA	12 (2) <sup>de1</sup>	15.6 (1.2) <sup>e1</sup>	0.26 (0.05) <sup>a1</sup>	0.281 (0.004) <sup>a1</sup>			
S90	$14.5 (0.7)^{f1}$	15.8 (1.2) <sup>e1</sup>	0.22 (0.09) <sup>a1</sup>	0.23 (0.15) <sup>a1</sup>			
S90-CA	13.0 (1.4) <sup>ef1</sup>	15 (2) <sup>e1</sup>	0.4 (0.2) <sup>a1</sup>	0.54 (0.06) <sup>a1</sup>			
S80	11 (2) <sup>de1</sup>	9.3 (0.6) <sup>cd1</sup>	6.5 (0.3) <sup>ab1</sup>	6 (3) <sup>b1</sup>			
S80-CA	10.0 (0.6) <sup>d1</sup>	10.8 (1.0) <sup>d1</sup>	2.4 (0.3) <sup>a1</sup>	3.9 (0.2) <sup>ab2</sup>			
S70	7.5 (0.7) <sup>c1</sup>	10.4 (0.7) <sup>cd2</sup>	17.1 (0.9) <sup>cd1</sup>	14.6 (1.1) <sup>c1</sup>			
S70-CA	$6.3 (0.7)^{c1}$	8.5 (0.3) <sup>c1</sup>	11.6 (1.2) <sup>bc1</sup>	$12 (2)^{c1}$			
S60	8 (2) <sup>c1</sup>	5.4 (0.4) <sup>b1</sup>	33 (12) <sup>e1</sup>	26 (3) <sup>d1</sup>			
S60-CA	2.9 (0.8) <sup>b1</sup>	3.55 (0.11) <sup>b1</sup>	24 (2) <sup>d1</sup>	27.1 (0.8) <sup>d1</sup>			
PCL	0.120 (0.04) <sup>a1</sup>	0.117 (0.011) <sup>a1</sup>	> D.L.	> D.L.			

744 D.L.: 0.1-200 cc/(m<sup>2</sup>.day)

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748 Different superscript numbers within the same row indicate significant differences due to storage time (p < 0.05).

Table 5. Mean values and standard deviation of surface roughness parameters and optical properties of the different films at 1 (Initial time) and 5
 (Final time) weeks of storage at 53% relative humidity and 25 °C.

Films	Roughnes		ness parameters		Gloss (85°)		Ti (650nm)	
FIIIIS	r (%)	Ra (nm)	Rq (nm)	Initial	Final	Initial	Final	
S	6.1 (0.9) <sup>abc</sup>	215 (61) <sup>a</sup>	263 (63) <sup>a</sup>	40 (5) <sup>c1</sup>	37.2 (1.5) <sup>bcd1</sup>	85.1 (0.3) <sup>h1</sup>	84.2 (0.2) <sup>fg2</sup>	
S-CA	2.5 (0.2) <sup>a</sup>	165 (59) <sup>a</sup>	206 (72) <sup>a</sup>	39 (5) <sup>c1</sup>	32 (2) <sup>b1</sup>	85.5 (0.6) <sup>h1</sup>	85.9 (0.8) <sup>g1</sup>	
S90	13 (5) <sup>d</sup>	198 (60) <sup>a</sup>	247 (65) <sup>a</sup>	37 (9) <sup>bc1</sup>	35 (6) <sup>bc1</sup>	77.3 (0.6) <sup>bc1</sup>	79.8 (0.6) <sup>cd2</sup>	
S90-CA	3.2 (1.2) <sup>a</sup>	148 (30) <sup>a</sup>	200 (38) <sup>a</sup>	52 (3) <sup>d1</sup>	43 (17) <sup>de2</sup>	79.4 (0.3) <sup>def1</sup>	81 (0.4) <sup>de2</sup>	
S80	8 (3) <sup>bc</sup>	202 (26) <sup>a</sup>	255 (26) <sup>a</sup>	29 (9) <sup>c1</sup>	31 (9) <sup>b1</sup>	82.4 (0.6) <sup>g1</sup>	82.5 (1.1) <sup>ef1</sup>	
S80-CA	14 (4) <sup>d</sup>	390 (216) <sup>b</sup>	497 (35) <sup>b</sup>	20 (13) <sup>a1</sup>	14 (2) <sup>a1</sup>	73.6 (0.6) <sup>a1</sup>	76 (0.7) <sup>a1</sup>	
S70	8 (3) <sup>bc</sup>	220 (57) <sup>a</sup>	271 (68) <sup>a</sup>	34 (10) <sup>bc1</sup>	46 (4) <sup>e2</sup>	80.9 (0.14)efg1	80.9 (0.4) <sup>de1</sup>	
S70-CA	11 (3) <sup>cd</sup>	199 (58) <sup>a</sup>	265 (72) <sup>a</sup>	31(9) <sup>bc1</sup>	35 (10) <sup>bcd1</sup>	76.2 (0.7) <sup>b1</sup>	77.1 (0.6) <sup>ab1</sup>	
S60	5.6 (1.4) <sup>ab</sup>	163 (60) <sup>a</sup>	210 (79) <sup>a</sup>	38 (6) <sup>c1</sup>	37 (4) <sup>bcd1</sup>	81.2 (1.3) <sup>fg1</sup>	81.6 (0.7) <sup>de1</sup>	
S60-CA	9.2 (1.3) <sup>bcd</sup>	163 (26) <sup>a</sup>	214 (26) <sup>a</sup>	50 (7) <sup>d1</sup>	41 (10) <sup>cde2</sup>	78.8 (0.8) <sup>cde1</sup>	79.4 (0.8) <sup>cd1</sup>	
PCL	6.0 (0.9) <sup>ab</sup>	154 (16) <sup>a</sup>	195 (25) <sup>a</sup>	59 (16) <sup>e1</sup>	57 (9) <sup>f1</sup>	78 (2) <sup>cd1</sup>	78 (2) <sup>bc1</sup>	

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764 Different superscript numbers within the same row indicate significant differences due to storage time (p < 0.05).